

Pyridoxal conjugated gold nanoparticles for distinct colorimetric detection of chromium(III) and iodide ions in biological and environmental fluids

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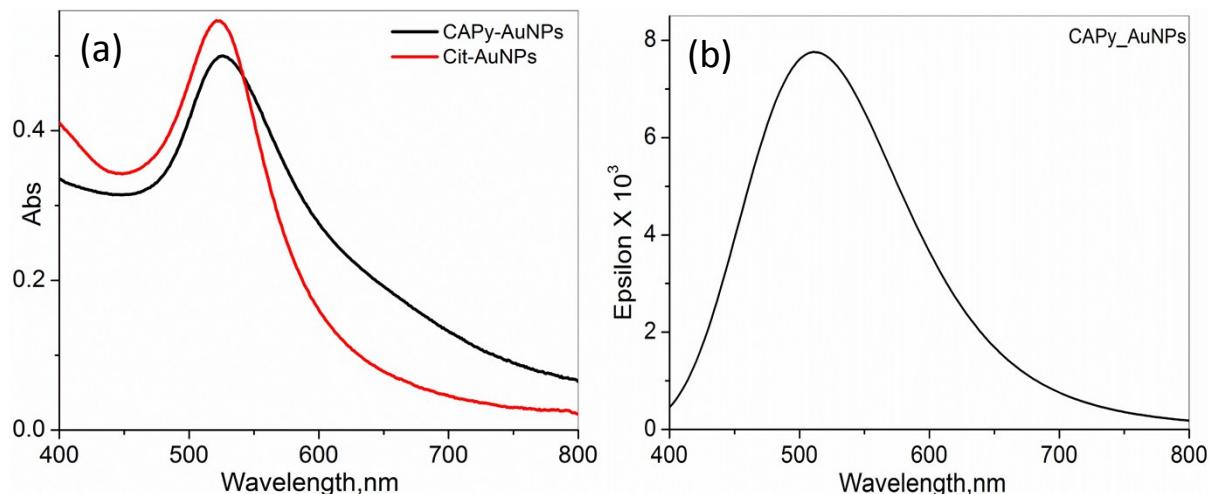


Fig. S1. (a) UV-Vis spectra of the citrate capped AuNPs and CAPy-AuNPs and (b) TD-DFT calculated UV-Vis spectra of CAPy-Au₆.

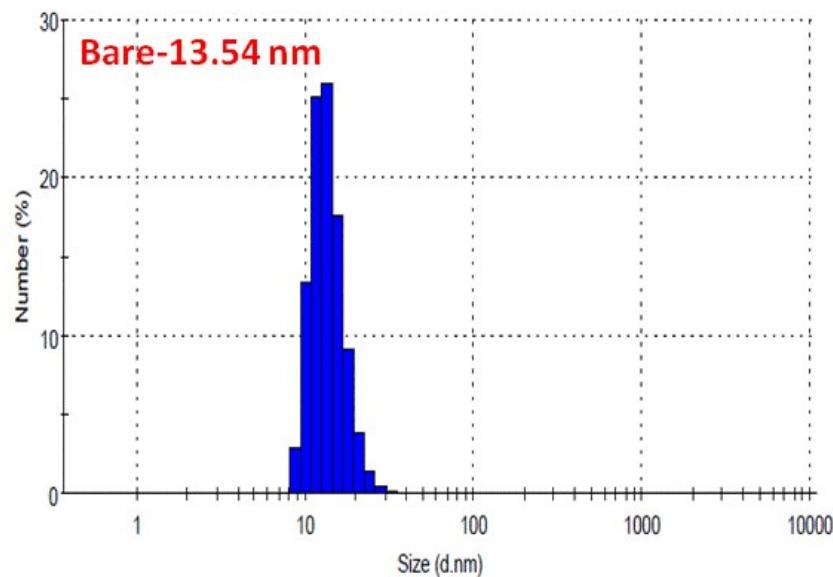


Fig. S2. DLS data of citrate capped AuNPs.

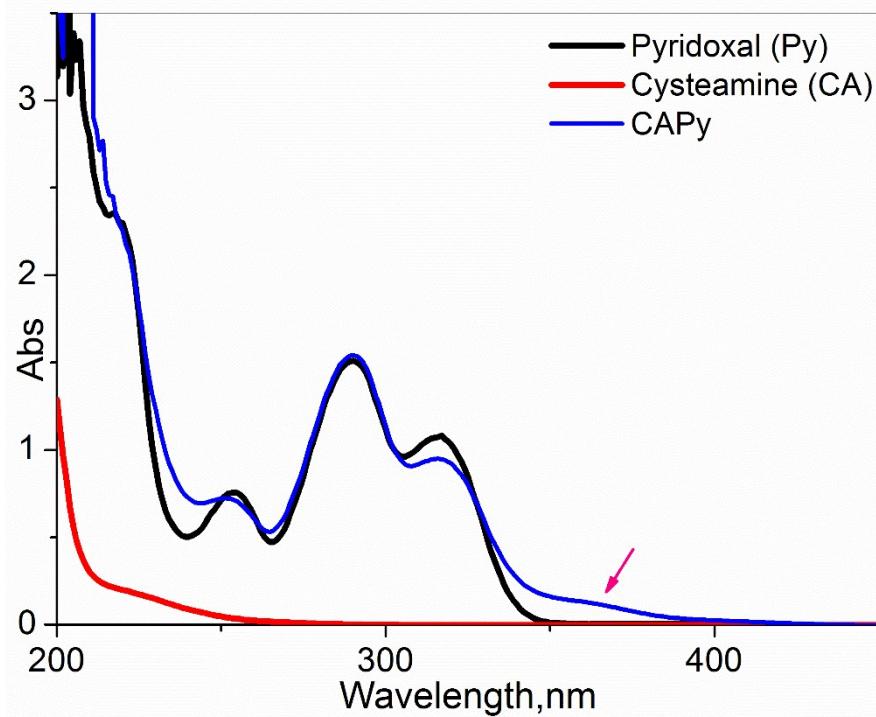


Fig. S3. UV-Visible analysis for the interaction between CA and Py.

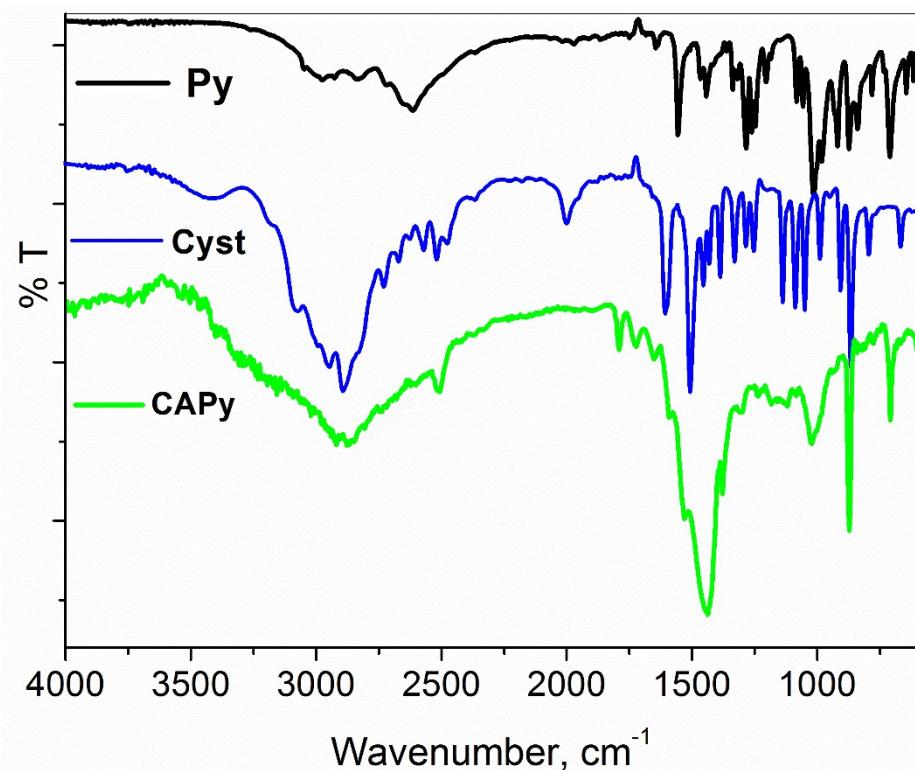


Fig. S4. FTIR analysis for the interaction between CA and Py.

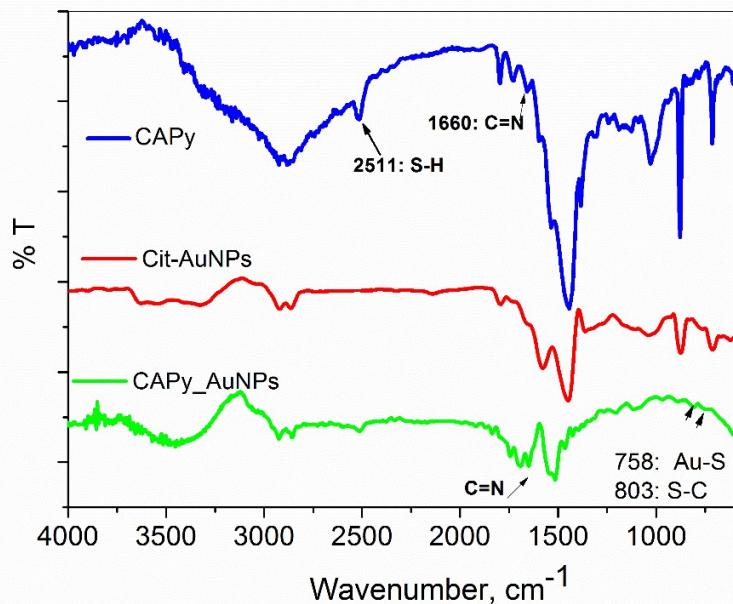


Fig. S5. FTIR spectra of bare AuNPs, capping agent CAPy and CAPy-AuNPs.

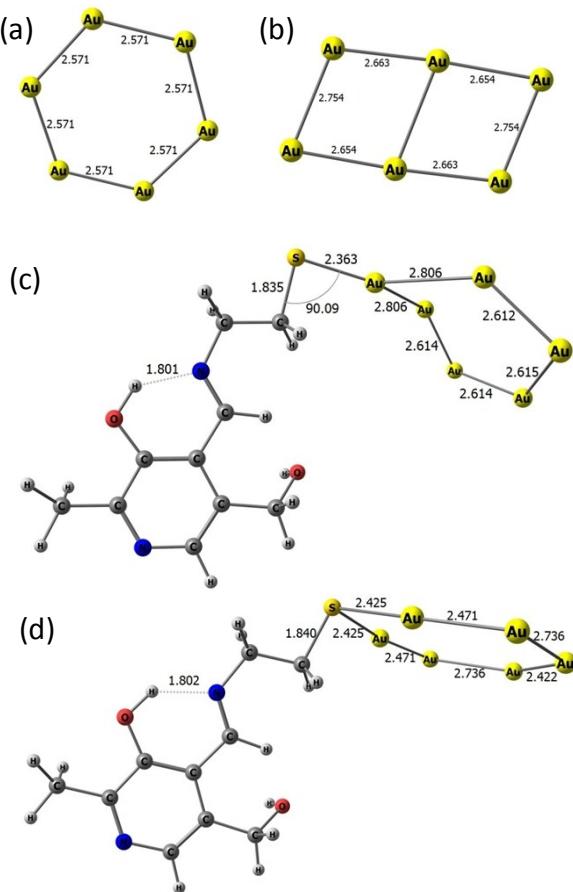


Fig. S6. Optimized structure of Au_6 (**a**) and CAPy- Au_6 (**c, d**) by semi-empirical PM6 method and Au_6 (**b**) by B3LYP/LANL2DZ method in the gas phase. The structure '**d**' is energetically more stable (-22.16 kcal/mol) than the '**c**', where the thiol group of CAPy interacting with the two Au atoms.

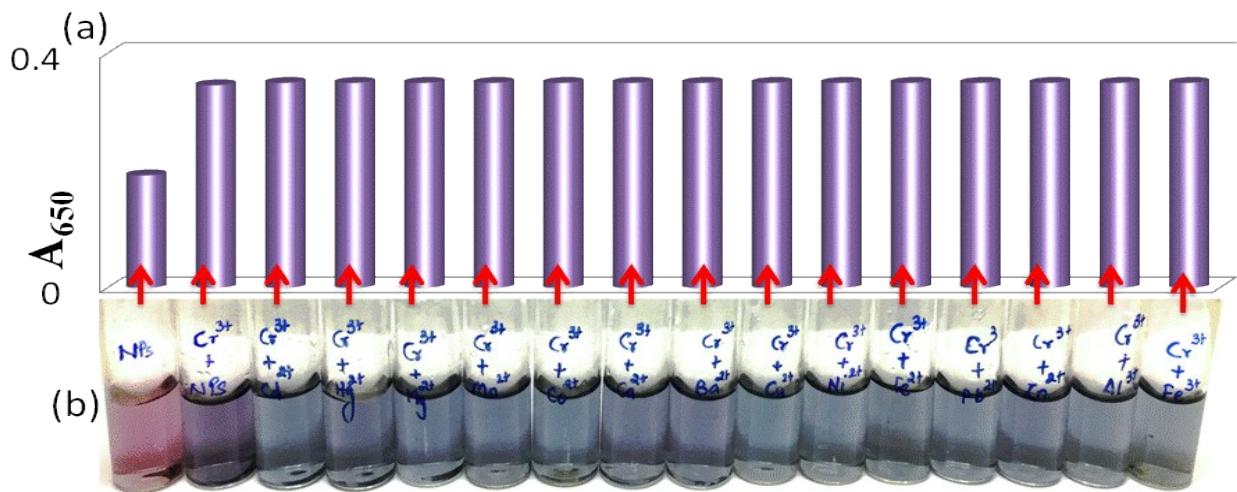


Fig. S7. (a) Bar graph at wavelength 650 nm and (b) naked-eye responses depicting the selectivity of CAPy-AuNPs in presence of equivalent molar concentration of Cr^{3+} ($2.5 \times 10^{-4}\text{M}$) and other metal ions ($2.5 \times 10^{-4}\text{M}$).



Fig. S8. (a) Bar graph at wavelength 700 nm and (b) naked-eye responses depicting the selectivity of CAPy-AuNPs in presence of equivalent molar concentration of I^- ($1 \times 10^{-4}\text{M}$) and other anions ($1 \times 10^{-4}\text{M}$).

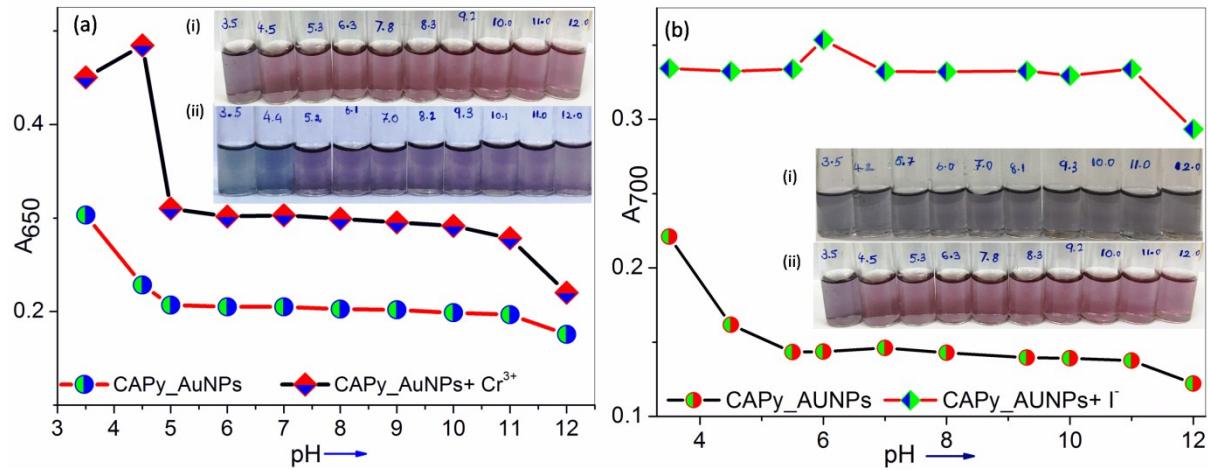


Fig. S9. Effects of change of pH on (a) CAPy-AuNPs solution in the absence and presence of Cr³⁺ and (b) CAPy-AuNPs in the absence and presence of I⁻.

Table S1. Brief account of various colorimetric NPs systems reported for Cr³⁺ and I⁻ detection.

Developed system	Detecting Ions	LOD (M)	Real samples	References
AgNP-I	Cr ³⁺	0.441 x 10 ⁻⁶	-	1
PAH-AuNPs		1.17 x 10 ⁻⁶	Milk powder, milk, lake water	2
PDCA- AuNPs		2.27 x 10 ⁻⁶	-	3
GA-AuNPs		0.05 x 10 ⁻⁶	Lake and tap water	4
CAPy-AuNPs		11.5 x 10⁻⁶	Tap and river water, liquid milk, milk powder	This study
Virgin AgNPs	I ⁻	0.32 x 10 ⁻⁶ Fe ³⁺ 1.32 x 10 ⁻⁶ Cr ³⁺	-	5
Cit-Core/Shell Cu@AuNPs		5 x 10 ⁻⁶	-	6
Au@Ag core–shell NPs with Cu ²⁺		0.5 x 10 ⁻⁶	Drinking water , Dried kelps,Agarose gels	7
ATTP-AuNPs		15 x 10 ⁻⁹	Lake water	8
CAPy-AuNPs		5.89 x 10⁻⁷	Tap and river water, urine	This Study

Table S2. Brief account of various reported official methods for Cr³⁺ and I⁻ detection.

Detected Ion	Method	LOD	Reference
Cr ³⁺	ICP-MS	0.03 µg/L	9
	AAS	5.0 ng	10
	GFAAS	0.03–0.04 µg/L	11
	HRS	25 ppt	12
	DPSV	0.4 µM	13
	AAS	0.19 ng mL ⁻¹	14
	Chemiluminescence	5 x 10 ⁻¹⁰ M	15
I ⁻	Total reflection X-ray fluorescence	180 µg L ⁻¹	16
	Molecular absorption spectrometry	60 µg L ⁻¹	17
	Flow-injection cold-vapor AAS	3.6 µg L ⁻¹	18
	ISE	1.47 µg L ⁻¹	19
	ICP-OES	4.5 µg L ⁻¹	20
	Capillary Electrophoresis	0.06 µM	21
	ICP-MS	0.19 ng g ⁻¹	22

References

1. M. Elavarasi, A. Rajeshwari, S. A. Alex, D. Nanda Kumar, N. Chandrasekaran and A. Mukherjee, *Analytical Methods*, 2014, **6**, 5161-5167.
2. W. Jin, P. Huang, Y. Chen, F. Wu and Y. Wan, *Journal of Nanoparticle Research*, 2015, **17**, 358.
3. R. Shaikh, N. Memon, A. R. Solangi, H. I. Shaikh, M. H. Agheem, S. A. Ali, M. R. Shah and A. Kandhro, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2017, **173**, 241-250.
4. C. Dong, G. Wu, Z. Wang, W. Ren, Y. Zhang, Z. Shen, T. Li and A. Wu, *Dalton Transactions*, 2016, **45**, 8347-8354.
5. S. Bothra, R. Kumar, R. K. Pati, A. Kuwar, H.-J. Choi and S. K. Sahoo, *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*, 2015, **149**, 122-126.
6. J. Zhang, X. Xu, C. Yang, F. Yang and X. Yang, *Analytical Chemistry*, 2011, **83**, 3911-3917.
7. J. Zeng, Y. Cao, C.-H. Lu, X.-d. Wang, Q. Wang, C.-y. Wen, J.-B. Qu, C. Yuan, Z.-f. Yan and X. Chen, *Analytica Chimica Acta*, 2015, **891**, 269-276.
8. I.-L. Lee, Y.-M. Sung, C.-H. Wu and S.-P. Wu, *Microchimica Acta*, 2014, **181**, 573-579.
9. M. M. L. Guerrero, E. V. Alonso, J. M. C. Pavon, M. T. S. Cordero and A. G. de Torres, *Journal of Analytical Atomic Spectrometry*, 2012, **27**, 682-688.
10. Y. SUZUKI and F. SERITA, *Industrial Health*, 1985, **23**, 207-220.
11. C. Veillon, K. Patterson and N. Bryden, *Analytica Chimica Acta*, 1982, **136**, 233-241.
12. S. I. Hughes, S. S. Dasary, A. K. Singh, Z. Glenn, H. Jamison, P. C. Ray and H. Yu, *Sensors and Actuators B: Chemical*, 2013, **178**, 514-519.
13. S. Wu, N. Chandra Sekar, S. N. Tan, H. Xie and S. H. Ng, *Analytical Methods*, 2016, **8**, 962-967.
14. M. H. Mashhadizadeh and M. Amoli-Diva, *Journal of Analytical Atomic Spectrometry*, 2013, **28**, 251-258.
15. W. R. Seitz, W. W. Suydam and D. M. Hercules, *Analytical Chemistry*, 1972, **44**, 957-963.

16. I. Varga, *Microchemical journal*, 2007, **85**, 127-131.
17. M. D. Huang, H. Becker-Ross, S. Florek, M. Okruss, B. Welz and S. Morés, *Spectrochimica Acta Part B: Atomic Spectroscopy*, 2009, **64**, 697-701.
18. O. Haase and J. Broekaert, *Spectrochimica Acta Part B: Atomic Spectroscopy*, 2002, **57**, 157-165.
19. A. A. Almeida, X. Jun and J. L. Lima, *Microchimica Acta*, 1997, **127**, 55-60.
20. M. R. Cave and K. A. Green, *Journal of Analytical Atomic Spectrometry*, 1989, **4**, 223-225.
21. A. N. d. Macedo, K. Teo, A. Mente, M. J. McQueen, J. Zeidler, P. Poirier, S. A. Lear, A. Wielgosz and P. Britz-McKibbin, *Analytical Chemistry*, 2014, **86**, 10010-10015.
22. T. K. D. Nguyen and R. Ludwig, *Analytical Sciences*, 2014, **30**, 1089-1092.
