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A facile route by using FeCl₃ to prepare dimeric BODIPY based porous organic polymers

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1. Materials and methods

1.1 Materials.

All chemical reagents were commercially available and used as received unless otherwise stated. Dichloromethane was dried by standard methods using CaCl₂ before distillation using distilling apparatus. All reactions were performed under an inert atmosphere of nitrogen. Analytical thin-layer chromatography (TLC) was performed using TLC plates pre-coated with silica gel (TLC: 10-40 μ m, 0.2±0.03 mm). Flash column chromatography was performed using 40-63 μ m (230-400 mesh) silica gels as the stationary phase.

1.2 Structure Characterization and Analysis.

The ¹H and ¹³C NMR spectra were obtained from a Bruker Avance spectrometer at 500 and 125 MHz respectively. All chemical shifts were reported in δ units relative to tetramethylsilane (TMS). A Hitachi U-5100 was used to measure UV-vis absorption spectra. A Hitachi F-2700 was used to perform fluorescence measurements. A high resolution mass spectrum was obtained by using a Tsq quantum access max from Thermo. Fourier transform infrared (FTIR) spectra were obtained on a Nicolet 6700 and reported in terms of the frequency of absorption (cm⁻¹). Samples were prepared by dispersing in anhydrous KBr. Solid-state cross-polarization magic angle spinning (CP/MAS) NMR spectra were recorded from a AVANCE III 400 MHz produced by Bruker. Thermogravimetric analyses were conducted in an N₂ stream with an SDT Q600 V8.0 Build 95 analyzer. The samples were heated from room temperature to 800 °C with a heating rate of 10 °C min⁻¹ under N₂ atmosphere. Scanning electron microscopy (SEM) images and Energy-dispersive X-ray spectroscopy (EDS) measurements of POPs were made in a FEI SIRION200 microscopy with an accelerating voltage of 10 and 20 kV respectively. All the samples were coated with gold before test. High-resolution transmission electron microscopy (HR-TEM) images of BDPs were performed on a JEOL JEM-2100F microscopy with an accelerating voltage of 200 kV. Prior to TEM measurements, samples were ultrasonically dispersed in ethanol and dropped to a copper grid with a diameter of 3 mm and coated with carbon film. Nitrogen sorption isotherms were measured at 77 K with a Micromeritics ASAP 2020 sorption analyzer. According to the absorption-desorption isotherms, BET (Brunauer-Emmett-Teller) specific surface area, pore size distribution and pore volume could be evaluated. Before each measurement, the samples were degassed at 120 °C for 6 h.

2. Synthesis

2.1 Synthesis of BDP-0

BDP-0 was prepared according to the literature.^[1] Crush product was purified via chromatography silica gel column with an eluting solvent petroleum ether (PE- DCM (v/v = 2:1) and afford the product **BDP-0** as red power (50 mg). ¹H NMR (500 MHz, CDCl₃) δ : 6.02 (s, 4H, pyrrole-H), 2.57 (s, 12H, CH₃), 1.90 (s, 12H, CH₃).

2.2 Synthesis of BDP-1

BDP-1 was prepared according to the literature.^[2] Crush product was purified by chromatography silica gel column with an eluting solvent PE-DCM (v/v = 1:1) and afford the product **BDP-1** as red power (180 mg). ¹H NMR (500 MHz, CDCl₃) δ : 7.52 (s, 4H, Ar-H), 6.01 (s, 4H, pyrrole-H), 2.57 (s, 12H, CH₃), 1.52 (s, 12H, CH₃).

2.3 Synthesis of BDP-2

4,4'-Biphenyldicarboxaldehyde (631 mg, 3 mmol) was dissolved in dry DCM (100 ml), followed by dropwise addition of 2,4-dimethylpyrrole (1142 mg, 12 mmol). After that, catalytic amount of TFA was added to the mixture. The reaction system was degassed for 2 minutes and protected by a N₂ atmosphere. After oxidation by 2,3-Dichloro-5,6-dicyano-1,4-benzoquinone (DDQ, 1430 mg, 6.3 mmol), the product was complexed with boron trifluoride etherate (BF₃·OEt₂) without further purification. After a 16 h, the solvent was evaporated under vacuum and the dark residue was purified by chromatography silica gel column with an eluting solvent PE-DCM (v/v = 1:1) and afforded product **BDP-2** as a red power (181 mg). Due to the bad solubility, no satisfied ¹³C NMR spectrum of **BDP-2** could be obtained. ¹H NMR (500 MHz, CDCl₃): δ = 7.82 (d, 4H, Ar-H), 7.41 (d, 4H, Ar-H), 6.01 (s, 4H, pyrrole-H) 2.57 (s, 12H, CH₃), 1.48 (s, 12H, CH₃). HRMS-EI calcd for C₃₈H₃₇B₂F₄N₄ 646.3062, found [M+H]⁺ 647.2916; [M+Na]⁺ 669.2738; [M+K]⁺ 685.4121.

2.4 General synthetic procedure for the BDP-POPs

The preparation and treatment of **BDP-POP-n** (n = 0, 1 and 2) were in a similar way.

A typically procedure for preparation of **BDP-POP-1** was given as follows: **BDP-1** (100 mg, 0.175 mmol) was dissolved in anhydrous 1,2-dichloroethane (8 mL). The mixture was degassed by three freeze-pump-thaw cycles and then FeCl₃ (234 mg, 1.44 mmol) was added into the mixture under N₂ atmosphere. After that, the mixture was degassed by three freeze-pump-thaw cycles and then kept at room temperature with stirring for 48 h. The precipitate was obtained by filtration washed by vast quantities of dichloromethane, tetrahydrofuran and chloroform. To guarantee a total removal of impurities, the precipitate was further purified by Soxhlet extraction with tetrahydrofuran, acetone, chloroform, methanol and water for 24 hours, respectively. Finally, the solid was freeze-dried for 3 days to afford black powders **BDP-POP-1** (70 mg, 70 %). **BDP-POP-0** and **BDP-POP-2** were synthesized by the similar route with yield 65% and 80%, respectively.

References

- [1] L. Wang, J. Cao, J. Wang, Q. Chen, A. Cui and M. He, *RSC Adv.*, 2014, **45**, 14786-14790.
- [2] J. Y. Liu, Y. Huang, R. Menting, B. Röder, E. A. Ermilov and D. K. P. Ng, Chem. Commun., 2013, 49, 2998-3000.

3. FTIR spectra



Fig S1. FT-IR spectra of BODIPY-based monomers and POPs.

4. Solid-state NMR spectra



Fig S2. ¹³C CP-MAS solid-state NMR spectra of **BDP-POP-n** (n = 0, 1 and 2).

5. TGA analysis



6. EDS analysis results



Fig S4. EDS results of (a) BDP-POP-0, (b) BDP-POP-1 and (c) BDP-POP-2.



Fig S5. EDS mapping results of (a) BDP-POP-0, (b) BDP-POP-1 and (c) BDP-POP-2.

7. Time dependent absorption spectra of DPBF in dimethylformamide (DMF) upon irradiation.



Fig S6. Time dependent absorption spectra of DPBF in dimethylformamide (DMF) upon irradiation with a low power green LED lamp: (a) **BDP-0**, (b) **BDP-1**, (c) **BDP-2**. (d) decay kinetics of DPBF with different BDPs. Test conditions: DPBF $(4 \times 10^{-5} \text{ M})$ in DMF (2.5 mL) solution with BDP $(6.4 \times 10^{-6} \text{ M})$ for (a), (b) and (c), respectively.

8. Solid emission spectra of POPs researched in this work.



Fig. S7 Solid emission spectra of (a) BDP-POP-0, (b) BDP-POP-1 and (c) BDP-POP-2.

9. Porous properties comparison of BDP-POP-1 synthesized in CHCl₃ and CH₂ClCH₂Cl.



Fig. S8 (a) Nitrogen adsorption-desorption isotherms, (b) pore size distribution for **BDP-POP-1** synthesized in CHCl₃, (c) pore size distribution for **BDP-POP-1** synthesized in CH₂ClCH₂Cl calculated by NLDFT measured at 77 K.

10. Singlet oxygen generation comparasion of BDP-POP-1 synthesized in CHCl₃ and CH₂ClCH₂Cl.



Fig S9. Time dependent absorption spectra of DPBF in dimethylformamide (DMF) upon irradiation with a low power green LED lamp: (a) **BDP-POP-1** synthesized in CH₂ClCH₂Cl, (b) **BDP-POP-1** synthesized in CH₃Cl. Test conditions: DPBF (4×10^{-5} M) in DMF (2.5 mL) solution with BDP-POP (0.1 mg), respectively.



Fig S10. Decay kinetics of DPBF with BDP-POP-1 synthesized in (a) CH₂ClCH₂Cl, (b) CH₃Cl.

11. ¹H NMR and Mass spectra of dimeric BODIPY monomers



Fig S11. ¹H NMR spectrum of BDP-0.









