

## Supporting Information

### Label-free colorimetric detection of tetracycline using analyte-responsive inverse-opal hydrogels based on molecular imprinting technology

Qian Yang,<sup>a,b</sup> Hailong Peng,<sup>a,c</sup> Jinhua Li,<sup>b</sup> Yanbin Li,<sup>c</sup> Hua Xiong,<sup>\*a</sup> and Lingxin Chen<sup>\*b</sup>

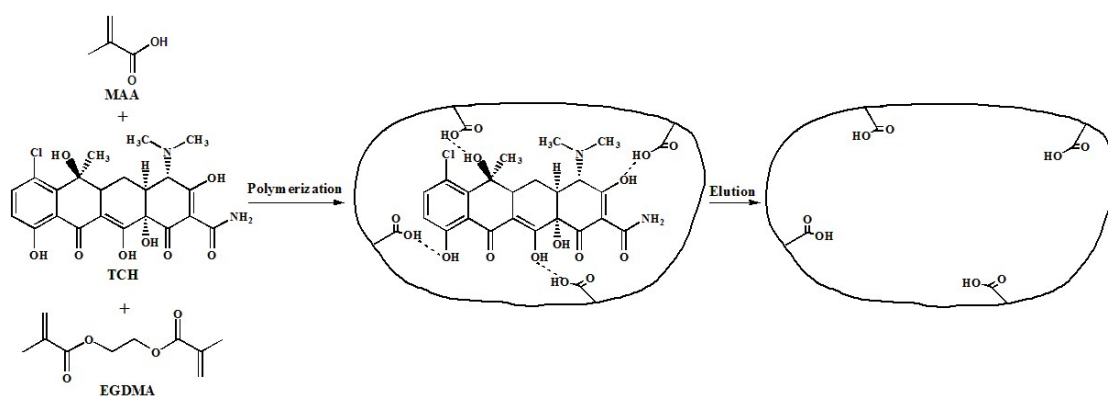
<sup>a</sup> State Key Laboratory of Food Science and Technology, Nanchang University, Nanchang 330047, China

<sup>b</sup> Key Laboratory of Coastal Environmental Processes and Ecological Remediation, Yantai Institute of Coastal Zone Research, Chinese Academy of Sciences, Yantai 264003, China

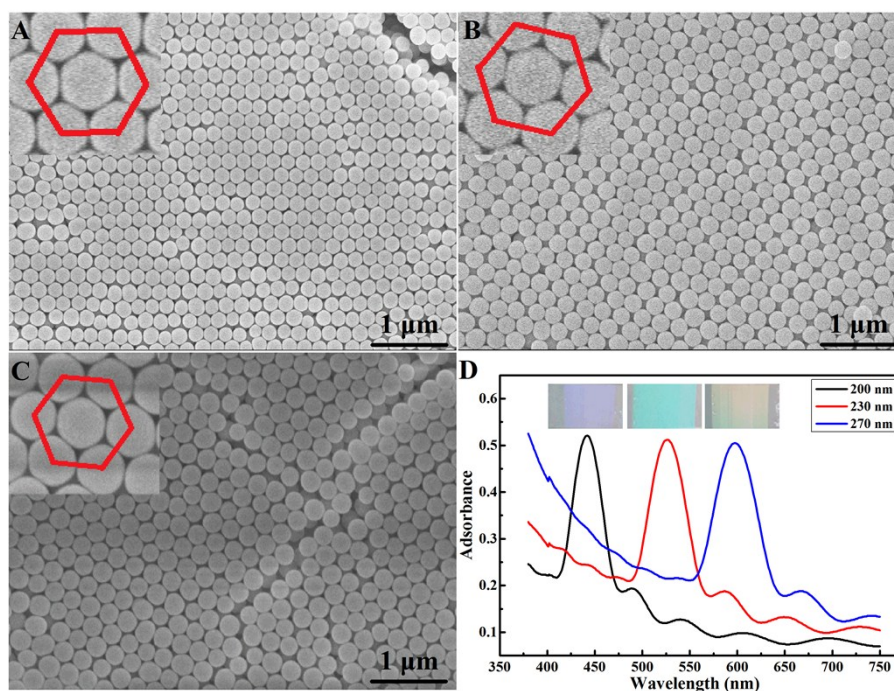
<sup>c</sup> Department of Biological and Agricultural Engineering, University of Arkansas, Fayetteville, AR 72701, USA

\* Corresponding author.

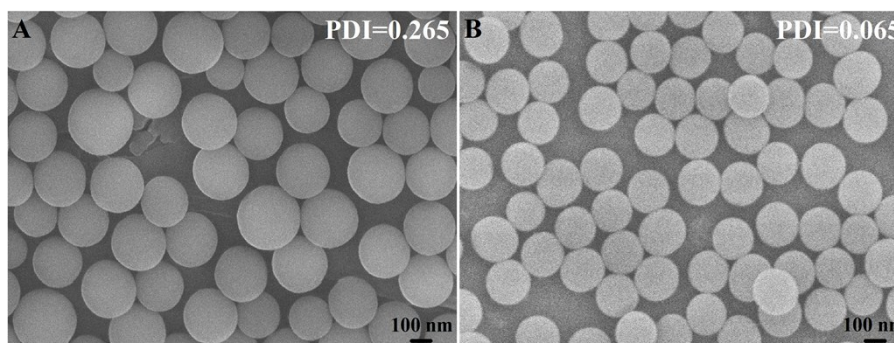
E-mail address: lxchen@yic.ac.cn (L. Chen); huaxiong100@126.com (H. Xiong)



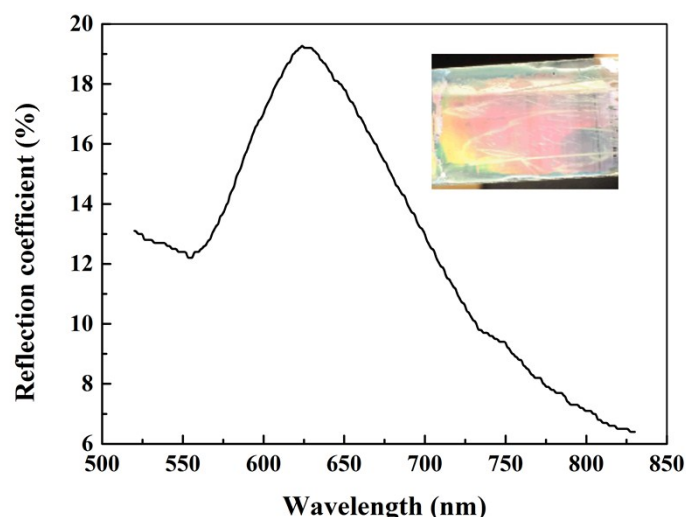
**Fig. S1.** Schematic illustration for the preparation of TCH-responsive IOH.



**Fig. S2.** SEM images of periodic silica-air arrays formed using silica microspheres of different sizes: (A) 200 nm prepared with 4 mL ammonia, (B) 230 nm prepared with 7 mL ammonia, (C) 270 nm prepared with 10 mL ammonia, and (D) UV-vis absorption spectra and photographs of silica-air arrays corresponding to different sizes.



**Fig. S3.** SEM images of silica microspheres prepared (A) before and (B) after optimization.



**Fig. S4.** UV-vis diffuse reflection spectrum of fabricated TC-responsive IOH; the inset shows a photograph of the TC-responsive IOH.

**Table S1** Comparisons of fabricated TC-responsive IOH analytical performance with other reported typical imprinted analyte-responsive IOHs.

Target analyte	Linear range	Diffraction peak shift or wavelength change ( $\Delta\lambda$ )	Responsive time	Reference
Cholesterol	$2.59 \times 10^{-13} - 2.59 \times 10^{-7}$ M	425 nm – 400 nm, $\Delta\lambda = 25$ nm	2 min	25
Bile acid	$10^{-12} - 10^{-6}$ M	425 nm – 395 nm, $\Delta\lambda = 30$ nm	2 min	33
Imidacloprid	$10^{-13} - 10^{-7}$ M	551 nm – 589 nm, $\Delta\lambda = 38$ nm	–	34
Bisphenol A	$10^{-13} - 10^{-3}$ M	$\Delta\lambda = 15$ nm	–	35
Cinchonine	$0 - 10^{-3}$ M	527 nm – 503 nm, $\Delta\lambda = 24$ nm	5 min	36
Ketamine	$0 - 1 \mu\text{g mL}^{-1}$	543.5 nm – 622.5 nm, $\Delta\lambda = 79$ nm	2 min	37
Atrazine	$10^{-16} - 10^{-6}$ M	558 nm – 618 nm, $\Delta\lambda = 60$ nm	2 min	18
L-Tryptophan	$10^{-10} - 10^{-5}$ M	$\Delta\lambda = 126$ nm	–	26
Tetracycline	$1 \times 10^{-10} - 1 \times 10^{-6}$ M	438 nm – 395 nm, $\Delta\lambda = 43$ nm	3 min	This work

#### Reference:

18. Z. Wu, C. Tao, C. Lin, D. Shen and G. Li, *Chem-Eur. J.*, 2008, **14**, 11358.
25. J. Li, Z. Zhang, S. Xu, L. Chen, N. Zhou, H. Xiong and H. Peng, *J. Mater. Chem.*, 2011, **21**, 19267.
26. Z. Yang, D. Shi, M. Chen and S. Liu, *Anal. Methods*, 2015, **7**, 8352.
33. Z. Wu, X. Hu, C. Tao, Y. Li, J. Liu, C. Yang, D. Shen and G. Li, *J. Mater. Chem.*, 2008, **18**, 5452.
34. X. Wang, Z. Mu, R. Liu, Y. Pu and L. Yin, *Food Chem.*, 2013, **141**, 3947.
35. N. Griffete, H. Frederich, A. Maitre, C. Schwob, S. Ravaine, B. Carbonnier, M. M. Chehimi and C. Mangeney, *J. Colloid Interf. Sci.*, 2011, **364**, 18.
36. Y. Zhang, S. Huang, C. Qian, Q. Wu and J. He, *J. Appl. Polym. Sci.*, 2016, **133**, 43191.
37. L. Meng, P. Meng, Q. Zhang and Y. Wang, *Anal. Chim. Acta*, 2013, **771**, 86.