Supporting Information

Alkyl phosphonic acid-based ligands as tools for converting hydrophobic iron nanoparticles into water soluble iron-iron oxide core-shell nanoparticles.

Kais Gharbi,^{a,b} Florent Salles,^a Paul Mathieu,^a Catherine Amiens,^a Vincent Collière,^a Yannick Coppel,^a Karine Philippot,^a Laurent Fontaine,^c Véronique Montembault,^c Leila Samia Smiri^b and Diana Ciuculescu-Pradines^{*a}

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SI1: TEM and size histogram of FeNP:



Figure SI1: Transmission electron micrograph of **FeNP** deposited onto a Cu grid from a solution of nanoparticles in THF (a) and size distribution from TEM images of nanoparticles (b)

SI2: FTIR-ATR of FeNP:



Figure SI2 : FTIR –ATR spectrum of FeNP.

SI3: TEM of FeNP dispersed in water and water-ethanol





Figure SI3: TEM of **FeNP** deposited onto a Cu carbon covered grid from a suspension of **FeNP** in water (a) and water-ethanol mixture (b) respectively, without phosphonic acid protection.

SI4: Snapshots of the transfer of FeNP into water



Figure SI4: Transfer into water of **FeNP** nanoparticles

SI5: HRTEM of Fe@FeoxNP@L1 and Fe@FeoxNP@L2 and size distribution from TEM images of core and shell of Fe@FeoxNP@L1 and Fe@FeoxNP@L2



Figure SI5: HRTEM of **Fe@FeoxNP@L1** (a) and **Fe@FeoxNP@L2** (b), showing a core/shell structure and size distribution from TEM images of core and shell of **Fe@FeoxNP@L1** (c) and **Fe@FeoxNP@L2** (d) respectively





K-a <u>1</u> [Å]	: 1,	540598 🗘 K-	o2 / K-o1 ratio:	0,500000 💲	Mode
K-a <u>2</u> (Å	: 1,	544426 💲 <u>s</u> h	ape factor K:	0,900000 🗘	 Crystalite size
<u>K</u> -a (Å):	1,	541874 🗘 Ca	lculation <u>b</u> ased on:	K-Alpha1 🔹	Lattice strain
No.	B obs. [°20]	B std. [°20]	Peak pos. [°28] B struct. [°20]	Crystallite size [Å]
1	0,901	0,0	79 44,5	13 0,822	104
2	1,150	0,0	79 64,8	63 1,071	. 88
3	1,463	0,0	79 82,1	32 1,384	76



Figure SI6: XRD analysis and the calculation using Hifhscore Plus software of the size of coherent diffraction domains for the α -Fe component for **Fe@FeoxNP@L1** and **Fe@FeoxNP@L2** respectively.

SI7: Stability investigations on Fe@FeoxNP@L2



Figure SI7: HRTEM images of Fe@FeoxNP@L2 (a) after the synthesis ; (b) 3 months later

SI8: Control experiments:

a) First control experiment: In the glove box, 30 mg of **FeNP** were dispersed in dry and O₂ free THF (15ml) together with 65 mg of **L2** ligand. 10 ml of degassed miliQ water were next transferred to the nanopaticle-THF solution using a cannula system in order to avoid air contamination. The obtained solution was of black color and free of any suspension which proves that the nanoparticles are highly soluble in this mixture. After two days of stirring, a TEM grid was realized from this solution (Figure SI8b) revealing the presence of nanoparticles which started to disintegrate. After magnetic precipitation of the nanoparticles and subsequent washing with degassed water the nanoparticles were no more soluble in water. TEM micrographs obtained from a suspension in water of the nanoparticles at the end of the washing process revealed seriously damaged nanoparticles. (Figure SI8c)





Figure SI8a: 1). TEM of **FeNP** from the THF-water degassed solution and 2) after magnetic precipitation and washing with degassed water.

b). Second control experiment: In the glove box, 30 mg of **FeNP** were dispersed in dry and O_2 free CH_2Cl_2 together with 60 mg of **L2**. The solution was then mechanically stirred during 12h after which the nanoparticles were magnetically isolated,



Figure SI8b: IR spectra of nanoparticles from the second control experiment

washed several times with CH_2CI_2 and dried under vacuum. The final powder was then analyzed by IR spectroscopy in order to evaluate the coordination of **L2** to the surface of the nanoparticles.

SI9: Determination of the grafting density of the ligands

Mass concentration of Fe and P determined by ICP-AES after digestion of the sample in aqua regia (mixture of 3:1v/v HCl and HNO₃)

Sample	Iron concentration	Phosphorus concentration (mg L ⁻¹)
Fe@FeoxNP@L1	177.8 ppm	13.66
Fe@FeoxNP@L2	695.3 ppm	3.283

Calculation of P and Fe atoms number considering 1L of solution:

Number (P)= $\frac{ICP(P)*N_A}{M(P)}$,

Number (Fe)= $\frac{ICP(Fe) * N_A}{M(Fe)}$

Where ICP (P) is the phosphorus concentration in the sample, N_A the Avogadro number 6.022*10²³mol⁻¹, M(P) the atomic weight of phosphorus 30.97 g.mol⁻¹, M(Fe) the atomic weight of iron 55.8 g.mol⁻¹.

Sample	Fe atoms number (considering 1L)	P atoms number (considering 1L)
Fe@FeoxNP@L1	19.18*10 ²⁰	2.66*10 ²⁰
Fe@FeoxNP@L2	75.03*10 ²⁰	0.63*10 ²⁰

Calculation of Fe atoms number in one zerovalent iron cube of 12.8 nm:

It was considered that the number of iron atoms in the cube (nanoparticle) remains the same after water transfer and oxidation of the surface.

Number (Fe atoms in one cube) = $\frac{m(cube) * N_A}{M(Fe)} = \frac{d * V * N_A}{M(Fe)} = \frac{7.87 * 10^6 * (12.8 * 10^{-9})^3 * 6.022 * 10^{23}}{55.8} = 178120$ iron atoms in a cube

Where d is iron density (7.87 $*10^{6}$ g*m⁻³), V is the volume of the cube in m³ (N_A the Avogadro number 6.022 $*10^{23}$ mol⁻¹ and M(Fe) the atomic weight of iron 55.8 g.mol⁻¹.

Calculation of P atoms coverage on iron cubes:

1. <u>Calculation of number of cubes in the sample:</u>

For Fe@FeoxNP@L1 sample:

Number iron cubes= $\frac{Fe \ atoms \ in \ the \ sample}{Fe \ atoms \ in \ one \ cube} = \frac{19.18 \times 10^{20}}{178120} = 10.7 \times 10^{15} \ iron \ cubes$

For Fe@FeoxNP@L2 sample:

Number iron cubes= $\frac{Fe \ atoms \ in \ the \ sample}{Fe \ atoms \ in \ one \ cube} = \frac{75.03 \times 10^{20}}{178120} = 42.12 \times 10^{15} \ iron \ cubes$

2. Calculation of number of P atoms per iron cube:

For Fe@FeoxNP@L1 sample:

 $P/NP = \frac{number \ of \ P \ atoms}{number \ of \ iron \ cubes} = \frac{2.66*10^{20}}{10.7*10^{15}} = 24859 \ P/NP$

For the Fe@FeoxNP@L2 sample:

 $\mathsf{P/NP} = \frac{number \ of \ P \ atoms}{number \ of \ iron \ cubes} = \frac{0.63 \times 10^{20}}{42.12 \times 10^{15}} = 1495 \ P/NP$

3. <u>Calculation of surface of the iron cube after transfer into water:</u>

For Fe@FeoxNP@L1 sample:

Surface cube (water)=6* (edge)²=6 *(13.8)²=1143 nm²

For the Fe@FeoxNP@L2 sample:

Surface cube (water)= 6^* (edge)²= $6^*(13.5)^2$ =1094 nm²

4. <u>Calculation of P coverage (P/nm²):</u>

For Fe@FeoxNP@L1 sample:

 $P-coverage = \frac{number of P atoms per cube}{surface of the cube} = \frac{24859}{1143} = 22 P/nm^2 = 22 L1/nm^2$

For Fe@FeoxNP@L2 sample:

 $P-coverage = \frac{number of P atoms per cube}{surface of the cube} = \frac{1495}{1094} = 1.4 P/nm^2 = 1.4 L2/nm^2$

If we consider the footprint of phosphonate group to be 0.24 nm² as indicated by Daou et al.¹ the maximum grafting density for L_1 should be $1/0.24=4.2P/nm^2$.

This means that in the case of L1 more than one monolayer of L1 is adsorbed on the nanoparticle surface. In the case of L2 the grafting of 1.4 P/nm² is six times more important than the one obtained when the same ligand was grafted on the iron oxide nanoparticles of the same size, but it correspond to the grafting density of dendritric molecules terminated with a phosphonate group anchored on iron oxide nanoparticles of about 11 nm.²

5. Calculation of the ratio P/Fe surface atoms

a) from the quantities of reactants:

Calculation of Fe atoms number on the surface of one zerovalent iron cube of 12.8 nm:

Iron with a body centered cubic structure has a lattice contact a=0.28 nm.

Number of Fe on the surface of the cube= $6 * (\frac{edge}{a})^2 = 6^* (\frac{12.8}{0.28})^2 = 13696$ Fe atoms

Where edge is the edge of the zerovalent iron cube determined by TEM (12.8 nm)

Quantity of Fe in the sample (mmol)=
$$\frac{mass of FeNP sample}{M(Fe)} * w\% Fe = \frac{30}{55.8} * 0.72 = 0.40 mmol Fe$$

% of iron atoms on the surface= $\frac{number\ of\ iron\ atoms\ at\ the\ surface\ of\ a\ cube}{number\ of\ iron\ atoms\ in\ the\ cube} * 100 = \frac{13696}{178120} * 100 \cong 8\%$

For Fe@FeoxNP@L1 sample:

Quantity of P in the sample (mmol)= $\frac{mass of L1 \ ligand}{M(L1)} = \frac{10}{139.09} = 0.07 \ mmol \ P$ Ratio P/Fe surface atom = $\frac{mmol P}{mmol Fe*\% of iron on the surface} = \frac{0.07}{0.40*0.08} \approx 2$

For Fe@FeoxNP@L2 sample:

Quantity of P in the sample (mmol)= $\frac{mass of L2 \ ligand}{M(L2)} = \frac{10}{2120} = 0.005 \ mmol \ P$ Ratio P/Fe surface atom = $\frac{mmol P}{mmol Fe*\% of iron on the surface} = \frac{0.005}{0.40*0.08} \approx 0.15 \approx \frac{1}{7}$

b) after purification of the sample:

For Fe@FeoxNP@L1 sample:

Ration P/Fe surface atoms =

 $\frac{P \text{ atoms number in the sample}}{Fe \text{ atoms number in the cube}^*} Fe \text{ atoms number on the surface of a cube} = \frac{2.66 \times 10^{20}}{\frac{19.18 \times 10^{20}}{17.8120} \times 13696} = 1.8$

For Fe@FeoxNP@L2 sample:

Ration P/Fe surface atoms =

P atoms number in the sample	$0.63*10^{20}$	_ 0 1
Fe atoms number in the sample Fe atoms number in the cube * Fe atoms number on the surface of a cube	75.03*10 ²⁰ 178120 *13696	- 0.1

SI10: DLS measurements:



DLS particles size distribution for the **Fe@FeoxNP@L1**. The results are presented as number-based distribution.



DLS particles size distribution for the **Fe@FeoxNP@L2**. The results are presented as number-based distribution.

SI11: Measurement of the T2 transverse relaxation time of the water protons as a function of iron concentration .

The measurements were performed with a Bruker Avance 500 MHz spectrometer (11.5T), equipped with a 5mm triple-resonance inverse Z-gradient probe. All chemical shifts were reported relative to tetramethylsilane (TMS). The temperature was set to 25°C.

The measurements were realized using as a solvent a mixture of 0.1% H₂O in D₂O in order to avoid the radiation damping effect.³ In order to avoid the agglomeration of the nanoparticles in the magnetic field of the spectrometer the nanoparticles were fixed in a 4% agar gel. Transverse relaxation times were determined by fitting the H₂O resonance linewidth as the magnetic field inhomogoneity contribution can be considered as negligible in the agar gel. Several samples were prepared as described in the following table and the measurements were realized after three days of storage of the nanoparticles in these conditions.

Sample	Quantity of Agar gel 4% (μL)	Solution of nanoparticles 0.1% H ₂ O/D ₂ O (μL)	0.1% H ₂ O/D ₂ O (μL)
1	750	0	750
2	750	50	700
3	750	100	650
4	750	150	600
5	750	200	550

Determination of transverse relaxivity (r2):

The T_2 times were determined from the inverse of half height linewidth of the water proton signal measured by spectral deconvolution (Figure S11a)



Figure SI11a: ¹H-NMR signal of water protons. The peak widening is observed when the iron concentration increases.

The values obtained are listed in the following table as a function of iron concentration in each sample (determined after the measurement by ICP-AES)

[Fe] mmol L ⁻¹	1/T ₂ (s ⁻¹)
0	14
0.27	80
0.46	140
0.61	233
0.97	325

In order to determine transverse relaxivity (r2) the plot of $1/T_2$ (sample)- $1/T_2$ (control) against iron concentration ([Fe] in mmol L⁻¹) (Figure SI14b) was fitted with a linear function: $1/T_2$ (sample)- $1/T_2$ (control)= r_2x [Fe]; where the r_2 was determined as the slope of the fit. Control is the sample without nanoparticles.



Figure SI11b: Sample plot using T_2 measurements of nanoparticles as varying iron concentrations to determine relaxivity.

- 1. T. J. Daou, S. Begin-Colin, J. M. Greneche, F. Thomas, A. Derory, P. Bernhardt, P. Legare and G. Pourroy, *Chemistry of Materials*, 2007, **19**, 4494-4505.
- B. Basly, G. Popa, S. Fleutot, B. P. Pichon, A. Garofalo, C. Ghobril, C. Billotey, A. Berniard, P. Bonazza, H. Martinez, D. Felder-Flesch and S. Begin-Colin, *Dalton Transactions*, 2013, 42, 2146-2157.
- 3. A. Vlassenbroek, J. Jeener and P. Broekaert, *The Journal of Chemical Physics*, 1995, **103**, 5886-5897.