

Versatile “click” synthesis of 1-hydroxy-1,1-methylenebisphosphonic acids with thioalkoxy substituents for the preparation of stable gold nanoparticles

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Experimental

All water was distilled and subsequently purified to Millipore Milli-Q quality. For Au NP synthesis, all glassware used was cleaned with concentrated nitric acid (60%), then rinsed thoroughly with water before use. UV-Visible Absorption spectroscopy of Au NPs dispersions was recorded on a Jasco V630 spectrophotometer at neutral pH. All experiments under microwave (MW) irradiations were run in a 30 mL MW tube, using an Anton Paar monowave 300 microwave synthesis reactor. ^1H NMR spectra (400 MHz), proton-decoupled ^{13}C NMR spectra (100.6 MHz) and proton decoupled ^{31}P spectra (162.0 MHz) were recorded on a Bruker Avance 400 spectrometer and chemical shifts are reported in parts per million (ppm) on the δ scale. High Resolution Mass Spectrometry (HR-MS) experiments were realized on a LTQ Orbitrap Velos (Thermo Scientific) in positive mode using an ESI source or by MALDITOF/MS (Ultraflex TOF/TOF, Bruker Daltonique) in a DHB matrix in positive mode. MS spectra recorded in the Orbitrap mass analyzer allowing a mass accuracy around 1–2 ppm. DLS and ζ potential analyses were performed on a nano ZS (red badge) ZEN3600 Zetasizer at neutral pH. TEM images were taken on a FEI CM10 electron microscope. Samples were prepared by dropping of the colloidal solution onto holey carbon-coated Cu grid 10 times. FTIR absorption spectra were recorded on a Nicolet 380 Thermo Scientific FTIR spectrophotometer. Concentrated Au NPs colloidal solution (neutral pH, pre-washed by ultrafiltration) was added to analytical grade KBr and left dried at 80°C overnight. The pellet was prepared with this crude powder. The fluorescence of free Rhodamine 123 and coupled Au NPs was determined by using a spectrofluorimeter (Jobin-Yvon FluoroMax+) with excitation at 500 nm and emission at 522 nm.

1 and **2** were synthesized as previously described.^{24, 39}

General procedure for obtaining of compound 1a-c and 2a-c (all solvents are freshly degassed prior use): 1.1 mmole (for **1a-c**) or 0.55 mmole (for **2a-c**) of mercapto propionic acid (116 mg for **1a** 0.58 mg for **2a**) or ω -HS-PEG-carboxylic acids (488 mg for **1b**, 244 mg for **2b**, 2.35 g for **1c** and 1.18 g for **2c**) were dissolved in 3 mL of DMF and added in a 20 mL tube to 0.5 mmole of the compound **1** (147.5 mg) or **2** (134 mg) in 3 mL of distilled water. A small amount of 1-hydroxycyclohexylphenyl ketone is then added and the tube is sealed and stirred under a UV lamp at 360 nm during 3 hours. 8 mL of water are then added to the mixture and the resulting solution is washed with 3 times 10 mL of dichloromethane and 3 times 10 mL of diethyl ether. The aqueous solution is then lyophilized and a white powder is obtained. This protocol yield from 80 to 90 % of pure product (products are obtained as monosodium salts) except for compound **1c** were a mixture of the mono and difunctionalized compound was obtained. **1a**: ^{31}P NMR (162 MHz, D_2O) δ 18.46. ^1H NMR (400 MHz, D_2O) δ 2.86 (m, 1H, CH-S), 2.76 (d, $J = 6.3$ Hz, 2H, $\text{CH}_2\text{-S}$), 2.72 (m, 4H, $2\times\text{CH}_2\text{-S}$), 2.54 (m, 4H, $2\times\text{CH}_2\text{CO}_2\text{H}$), 1.82 (m, 2H), 1.63 (m, 1H), 1.46 (m, 3H), 1.33 (m, 2H). ^{13}C NMR (101 MHz, D_2O) δ 177.3 (s, COOH), 74.1 (t, $J = 134.8$ Hz, P-C-P), 45.3 (s, CH-S-), 37.0 (s, $\text{CH}_2\text{-COOH}$), 34.8 (s, $\text{CH}_2\text{-S-}$), 34.4, 33.5, 33.0 (s, $\text{CH}_2\text{-S-}$), 27.3, 26.9, 25.1, 23.3 (t, $J = 6.6$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$). HR-MS (ESI⁺ Q ToF) $\text{C}_{13}\text{H}_{26}\text{NaO}_{11}\text{P}_2\text{S}_2$: m/z (M+H)⁺: 507.02918 ; calc: 507.0289. **1b**: ^{31}P NMR (162 MHz, D_2O) δ 18.53. ^1H NMR (400 MHz, D_2O) δ 3.68 (m, 4H, $2\times\text{CH}_2\text{CO}_2\text{H}$), 3.52 (m, 60H, CH_2O), 2.78 (m, 1H, CH-S), 2.70 (d, $J = 6.3$ Hz, 2H, $\text{CH}_2\text{-S}$), 2.52 (m, 4H, $2\times\text{CH}_2\text{-S}$), 1.80 (m, 2H), 1.64 (m, 1H), 1.48 (m, 3H), 1.33 (m, 2H). ^{13}C NMR (101 MHz, D_2O) δ 176.5 (s, COOH), 74.1 (t, $J = 135.0$ Hz, P-C-P), 69.9 (CH_2O), 69.6 (CH_2O), 69.5 (CH_2O), 69.3 (CH_2O), 66.3 (CH_2O), 45.6 (s, CH-S-), 37.2 (s, $\text{CH}_2\text{-COOH}$), 34.6 (s, $\text{CH}_2\text{-S-}$), 33.5, 33.3, 31.4 (s, $\text{CH}_2\text{-S-}$), 29.2, 27.0, 23.5 (t, $J = 6.3$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$). HR-MS (ESI⁺ Q ToF) $\text{C}_{43}\text{H}_{84}\text{Na}_2\text{O}_{27}\text{P}_2\text{S}_2$: m/z (M+H)⁺: 603.7501 ; calc: 603.5919.

1c (compound **1c** was obtained in mixture with the monofunctionalized product as visible on the MS MALDI spectra): ^{31}P NMR (162 MHz, D_2O) δ 18.78. ^1H NMR (400 MHz, D_2O) δ 3.75 (m, 4H, $\text{CH}_2\text{CO}_2\text{H}$) 3.60 (m, 356H) 3.39 (m, 2H), 2.83 (m, 1H, CH-S), 2.78 (m, 2H, $\text{CH}_2\text{-S}$), 2.66 (m, 4H, $2\times\text{CH}_2\text{-S}$), 1.83 (m, 2H), 1.66 (m, 1H), 1.48 (m, 3H), 1.34 (m, 2H). ^{13}C NMR (101 MHz, D_2O) δ 175.4 (s, COOH), 129.1*, 127.1*, 74.0 (t, $J = 135.3$ Hz, P-C-P), 69.9 (CH_2O), 69.8 (CH_2O), 69.6 (CH_2O), 69.4 (CH_2O), 68.4 (CH_2O), 68.3 (CH_2O), 48.8, 45.6 (s, CH-S-), 37.2 (s, $\text{CH}_2\text{-COOH}$), 33.5 (s, $\text{CH}_2\text{-S-}$), 33.3, 31.4 (s, $\text{CH}_2\text{-S-}$), 30.7, 29.2, 27.0, 23.43 (t, $J = 6.1$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$). *vinylic carbon corresponding to mono addition. **2a**: ^{31}P NMR (162 MHz, D_2O) $\delta = 18.46$. ^1H NMR (400 MHz, D_2O) $\delta = 2.79$ (t, $J = 6.8$ Hz, 2H), 2.66 (t, $J = 7.1$ Hz, 2H), 2.60 (t, $J = 7.1$ Hz, 2H), 1.93 (m, 2H), 1.60 (m, 4H). ^{13}C NMR (126 MHz, D_2O) $\delta = 177.1$ (s, COOH), 74.0 (t, $J = 135.3$ Hz, P-C-P), 34.4 (s, $\text{CH}_2\text{-S-}$), 33.2, 30.9 (s, $\text{CH}_2\text{-S-}$), 29.7, 26.1, 23.0 (t, $J = 6.4$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$). HR-MS (ESI⁺ Q Tof) $\text{C}_8\text{H}_{18}\text{NaO}_9\text{P}_2\text{S}$: m/z (M+H)⁺: 375.00414 ; calc: 375.0044. **2b**: ^{31}P NMR (162 MHz, D_2O) $\delta = 18.73$. ^1H NMR (400 MHz, D_2O) δ 3.65 (m, 2H), 3.62 – 3.47 (m, 30H), 2.64 (t, $J = 6.4$ Hz, 2H), 2.57 – 2.45 (m, 4H), 1.81 (m, 2H), 1.50 (m, 4H). ^{13}C NMR (101 MHz, D_2O) δ 176.2 (s, COOH), 73.9 (t, $J = 135.8$ Hz, P-C-P), 69.5 (CH_2O), 69.2 (CH_2O), 66.1 (CH_2O), 34.3 (s, $\text{CH}_2\text{-S-}$), 33.1, 31.0 (s, $\text{CH}_2\text{-S-}$), 30.3, 29.8, 23.02 (t, $J = 6.4$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$). **2c**: ^{31}P NMR (162 MHz, D_2O) $\delta = 18.39$. ^1H NMR (400 MHz, D_2O) $\delta = 3.70$ (m, 16H), 2.76 (t, $J = 6.4$ Hz, 2H), 2.60 (t, $J = 7.1$ Hz, 2H), 2.54 (t, $J = 6.4$ Hz, 2H), 1.93 (m, 2H), 1.61 (m, 2H). ^{13}C NMR (126 MHz, D_2O) $\delta = 178.0$ (s, COOH), 74.1 (t, $J = 135.0$ Hz, P-C-P), 69.52 (m), 69.4 (CH_2O), 69.4 (CH_2O), 69.3 (CH_2O), 66.9 (CH_2O), 35.8 (s, $\text{CH}_2\text{-S-}$), 33.2, 31.1 (s, $\text{CH}_2\text{-S-}$), 30.4, 29.1, 23.11 (t, $J = 6.3$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$).

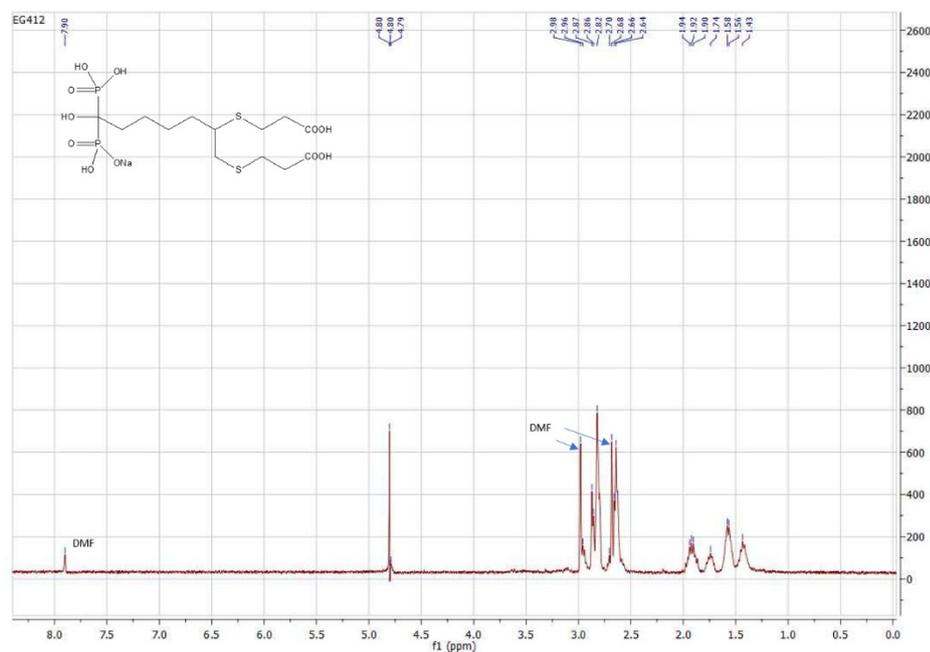
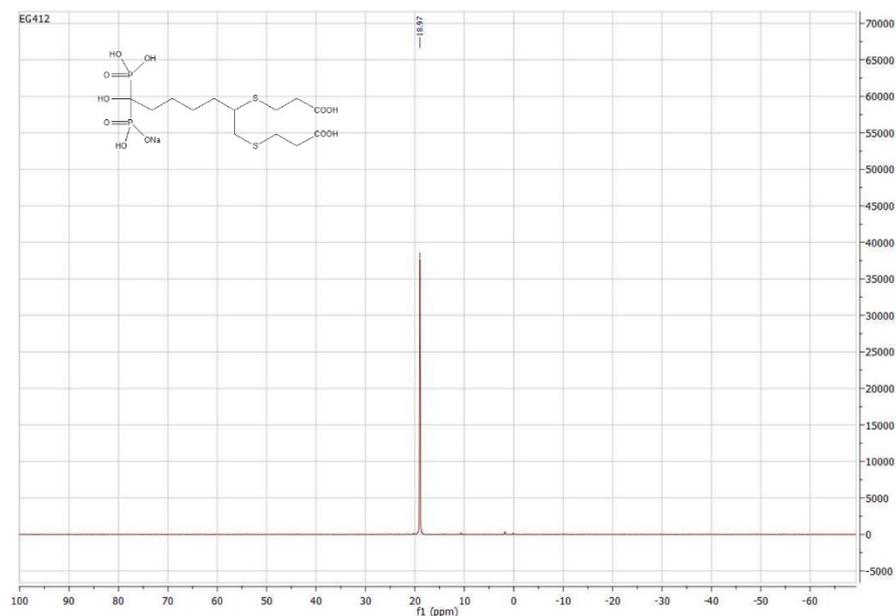
General procedure for oxidation of compound 2a and 2c. To a solution of compound **2a** or **2c** (0.5 mmole, 176mg for **2a**, 1.18 g for **2c**) in 4 mL of water, 282 mg of m-CPBA (1.65 mmol, 284 mg) were added in 4 mL of DMF under vigorous stirring. The solution is stirred for 24h at room temperature and then washed with 3 times 10 mL of diethyl ether. The aqueous solution is then lyophilized and a white powder is obtained. This protocol yield from 70% of pure product (obtained as monosodium salts) **3a**: ^{31}P NMR (162 MHz, D_2O) δ 18.40. ^1H NMR (400 MHz, D_2O) δ 3.84 (t, $J = 7.1$ Hz, 2H), 3.70 – 3.52 (m, 2H), 3.19 (t, $J = 7.1$ Hz, 2H), 2.26 (m, 2H), 2.08 (m, 2H), 1.99 (m, 2H). ^{13}C NMR (101 MHz, D_2O) δ 174.8 (s, COOH), 73.7 (t, $J = 145.4$ Hz, P-C-P), 52.4 (CH_2SO_2), 47.9 (CH_2SO_2), 33.4, 27.2, 22.6, 22.2. HR-MS (ESI⁺ Q Tof) $\text{C}_8\text{H}_{18}\text{NaO}_{11}\text{P}_2\text{S}$: m/z (M+H)⁺: 407.127 ; calc: 406.994. **3c**: ^{31}P NMR (162 MHz, D_2O) $\delta = 18.14$. ^1H NMR (400 MHz, D_2O) $\delta = 3.89$ (t, $J = 5.6$ Hz, 2H), 3.83 (s, 2H), 3.77 (d, $J = 4.2$ Hz, 2H), 3.59 (s, 178H), 3.40 (t, $J = 5.6$ Hz, 2H), 3.27 – 3.13 (m, 2H), 1.83 (m, 2H), 1.72 (m, 2H), 1.64 (m, 2H). ^{13}C NMR (126 MHz, D_2O) $\delta = 177.2$ (s, COOH), 73.9 (t, $J = 137.9$ Hz, P-C-P), 69.6 (CH_2O), 69.4 (CH_2O), 63.4 (CH_2O), 53.5 (CH_2SO_2), 51.8 (CH_2SO_2), 33.1, 22.7 (t, $J = 6.2$ Hz, $\text{CH}_2\text{-C}(\text{PO}_3\text{H}_2)_2$), 21.8.

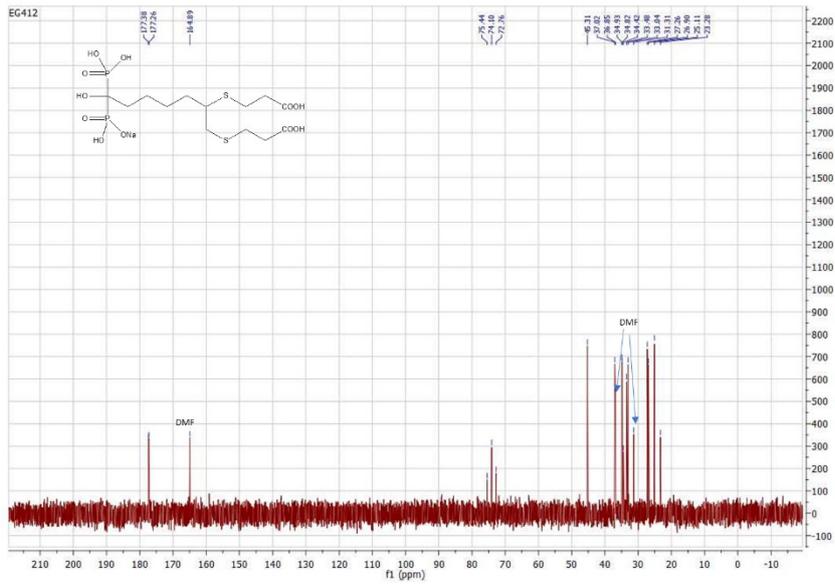
General procedure for the Au NP synthesis under MW irradiation: In a 4 mL vial, 900 μL of water, 125 μL of a 20 mM gold solution ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$, $\geq 99,9\%$, Sigma-Aldrich, St Louis, MO) and 100 μL of a 10 mM solution of compound **2c**, **3c** or the commercial phosphonate-PEG were mixed. The resulting solution was heated under MW (stirring at 1200 rpm) following three stages (1) heat as fast as possible to 100°C (2) hold at 100°C for 10 min (3) cool down.

General procedure for the PEGylated Au NP synthesis with sodium ascorbate: in a 20 mL vial, 125 μL of a 20 mM gold solution ($\text{HAuCl}_4 \cdot 3\text{H}_2\text{O}$, $\geq 99,9\%$, Sigma-Aldrich, St Louis, MO), 82 μL of a 10 mM solution of compound **3c** and 164 μL of a 31 mM solution of phosphonate-PEG (PEG PHOSPHONIC CARBOXYLIC ACID, ref. SP-1P-10-002 specific polymers) were mixed in 9.5 mL of water. The pH of the solution was then fixed at 10.5 with a 100 mM NaOH solution. Then, 100 μL of 50 mM solution of sodium ascorbate ($\text{C}_6\text{H}_7\text{NaO}_6$, $\geq 98\%$, Sigma-Aldrich, St Louis, MO) were added rapidly at room temperature. The mixture was stirred for 10 min at room temperature and afforded a pink Au NP solution. The Au NP solution is then washed several times with water and purified using ultrafiltration over a 30kDa membrane (Amicon[®], Millipore).

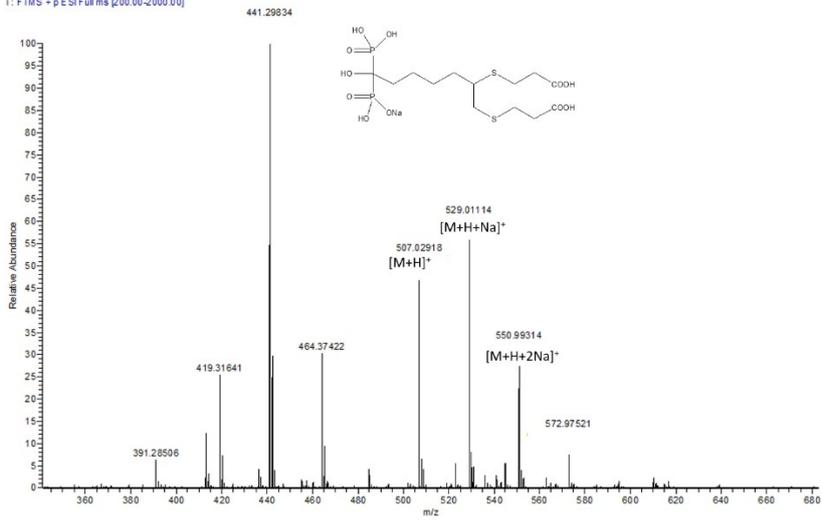
Rhodamine coupling onto the PEGylated Au NP: To 400 μL of PEGylated Au NP solution ($[\text{Au}] = 2,5 \text{ mM}$), 30 μL of a 650 μM EDC solution ($\text{C}_8\text{H}_{17}\text{N}_3$, $\geq 97 \%$, Sigma-Aldrich, St Louis, MO) and 30 μL of a 650 μM NHS solution were added. Then, the pH of the solution was fixed at pH=3. The resulting mixture was stirred for 2.5 h before addition of 30 μL of a 130 μM rhodamine 123 solution ($\text{C}_{21}\text{H}_{17}\text{ClN}_2\text{O}_3$, $\geq 85 \%$, Sigma-Aldrich, St Louis, MO). A triethylamine solution (25 g/L) was added to reach a pH = 10 and the solution was stirred for 16 h. The Au NP solution was washed several times with water and treated with HBSS 30 min to remove unbounded rhodamine before a last filtration.

Compound 1a (monosodium salt) RMN ^{31}P , ^1H , ^{13}C and MS spectra

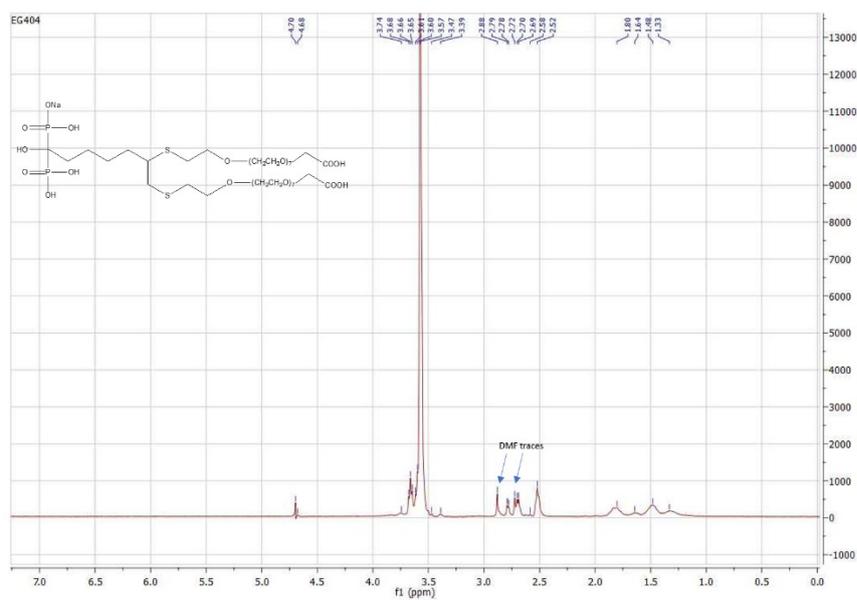
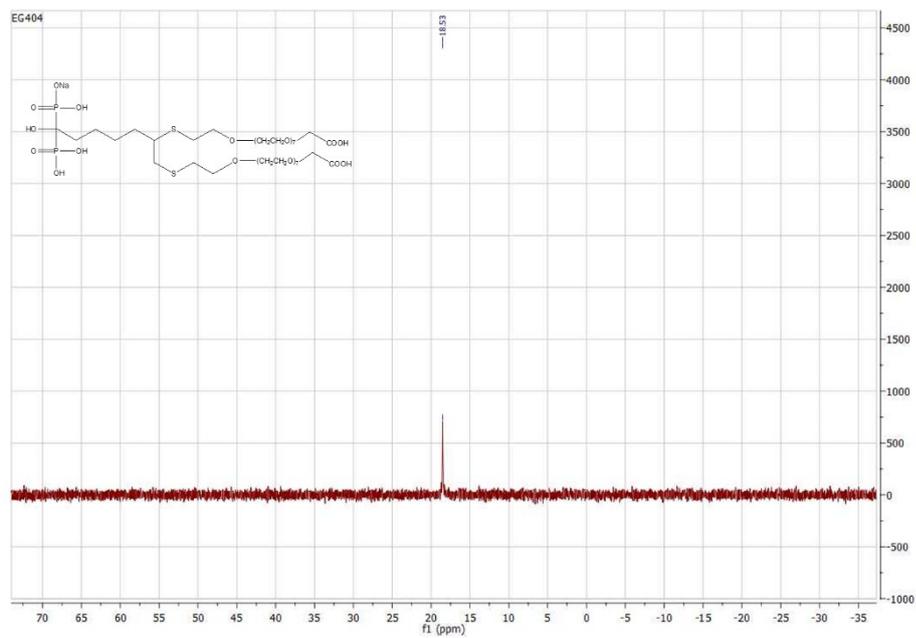


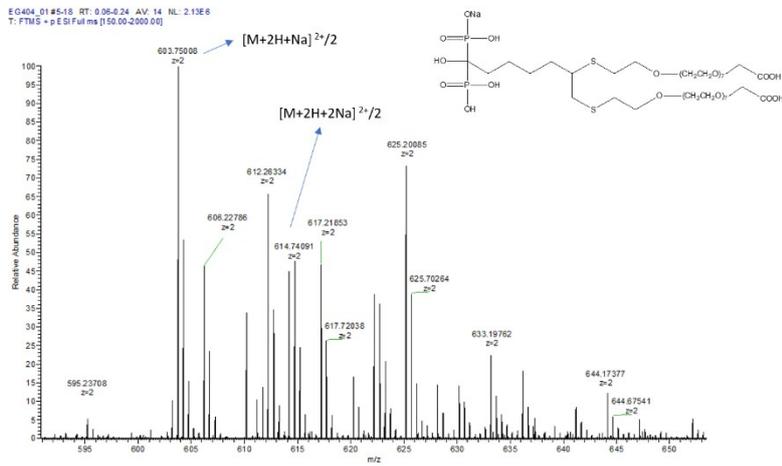
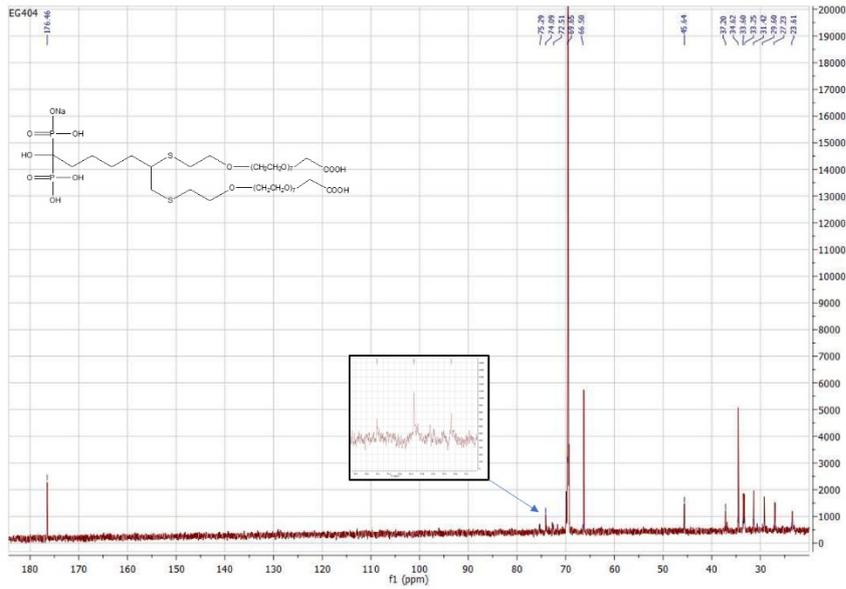


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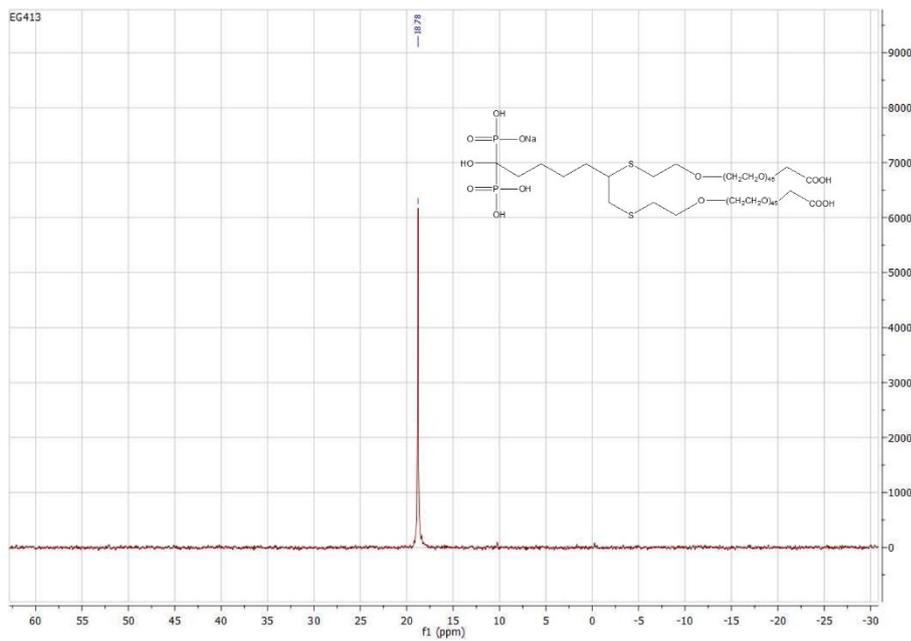


Compound 1b RMN 31P, 1H, 13C and MS spectra

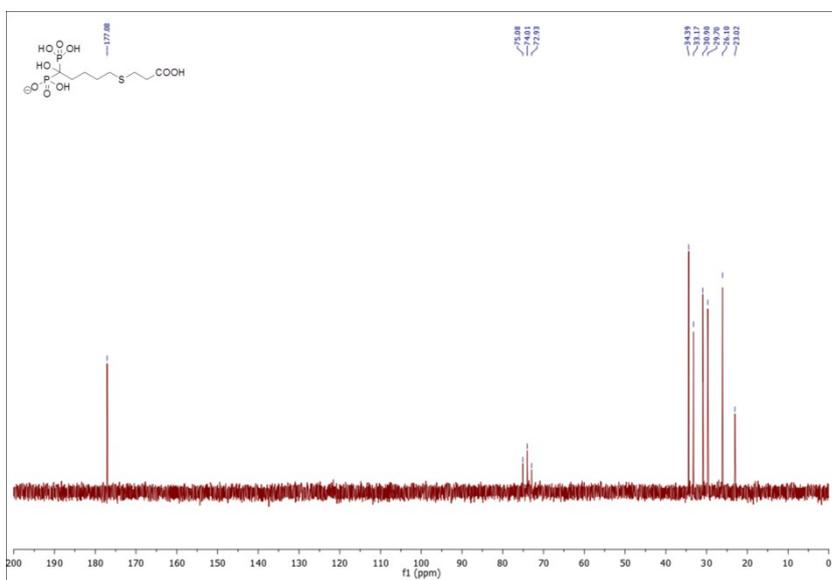
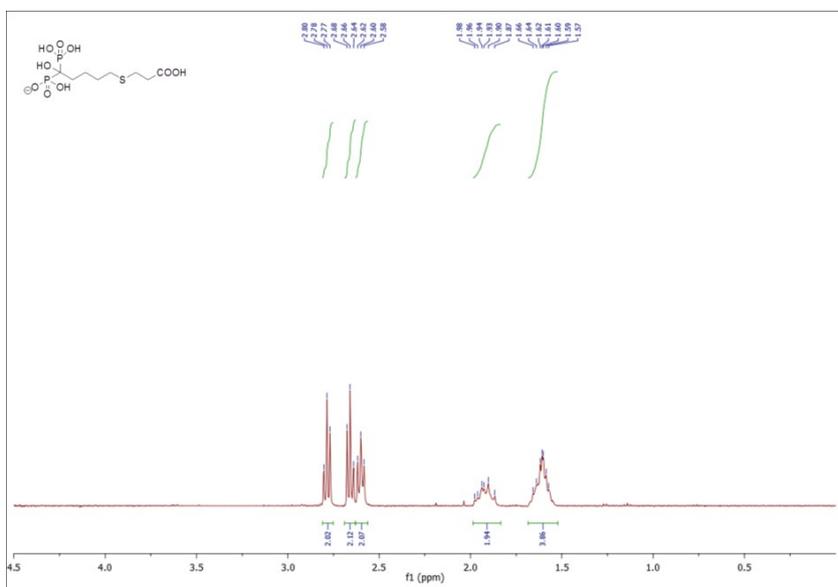
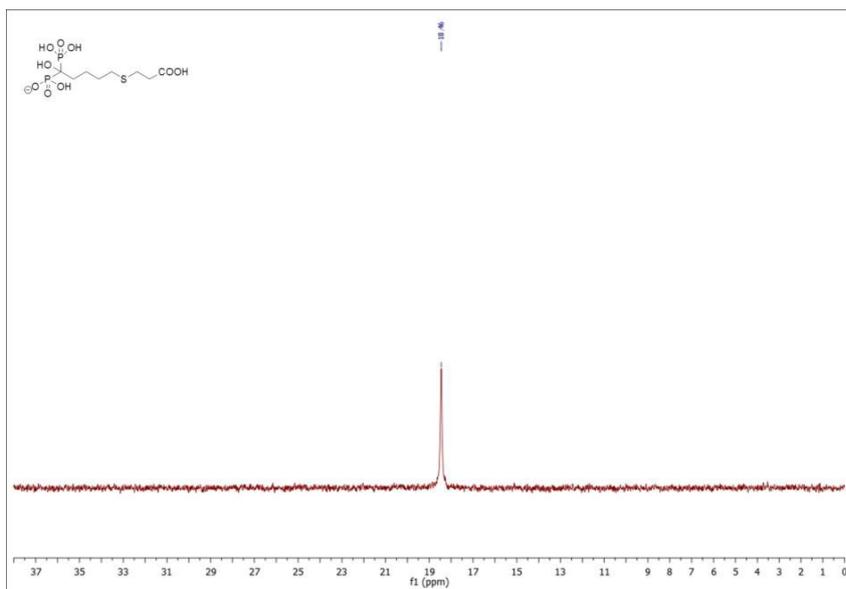




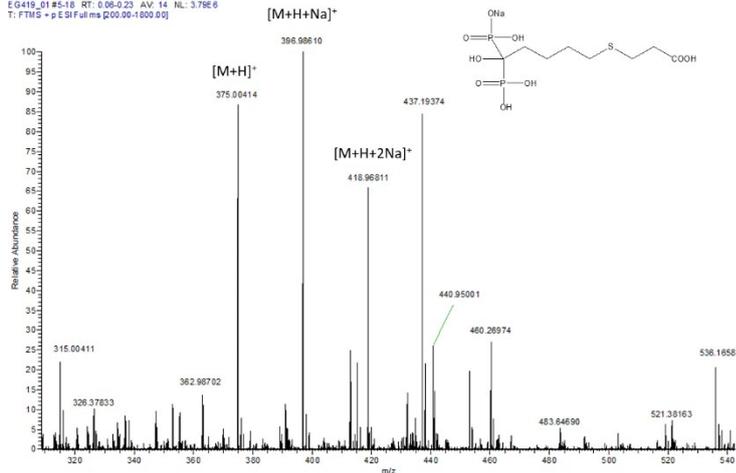
Compound 1c RMN 31P, 1H, 13C and MS spectra



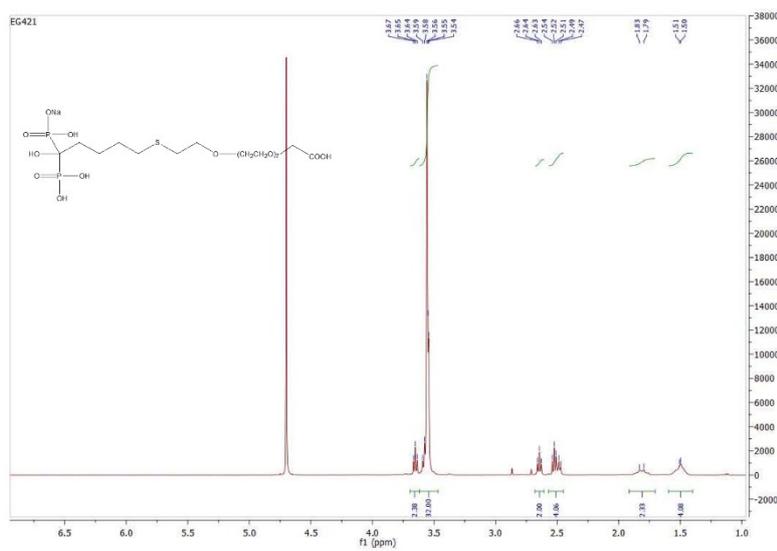
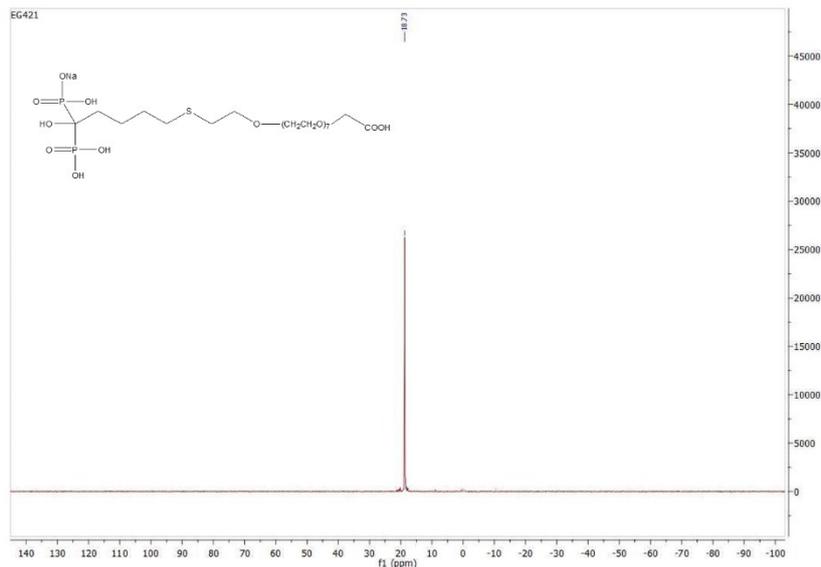
Compound 2a RMN 31P, 1H, 13C and MS spectra



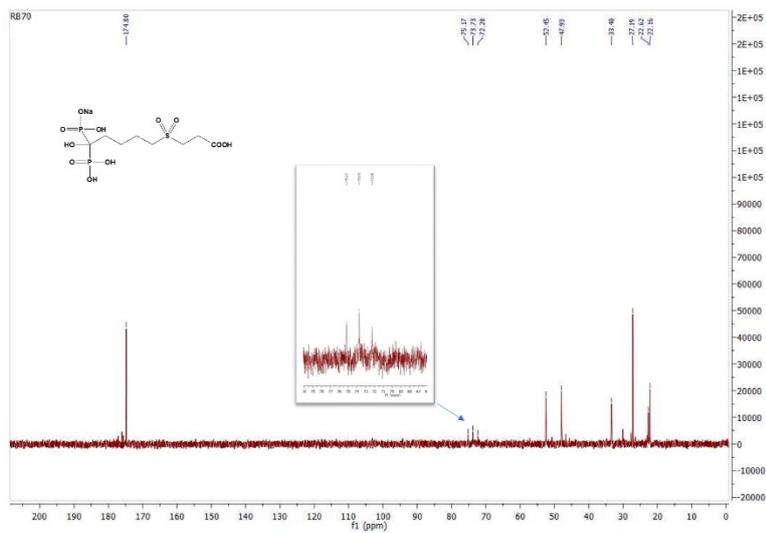
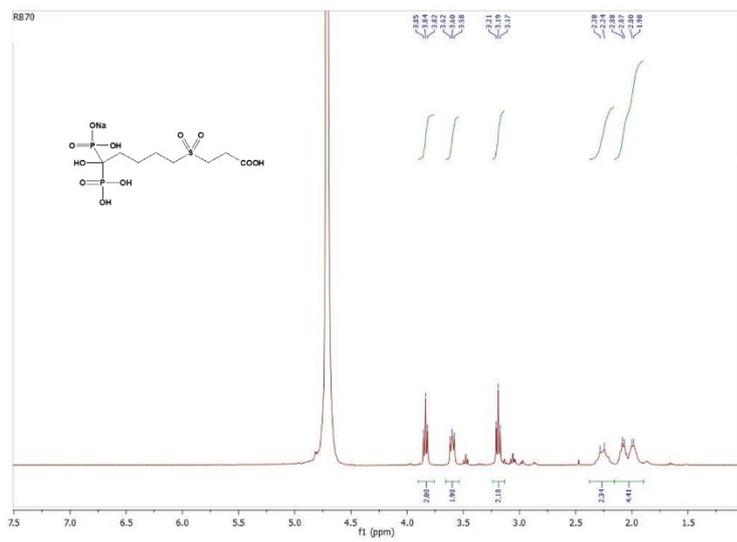
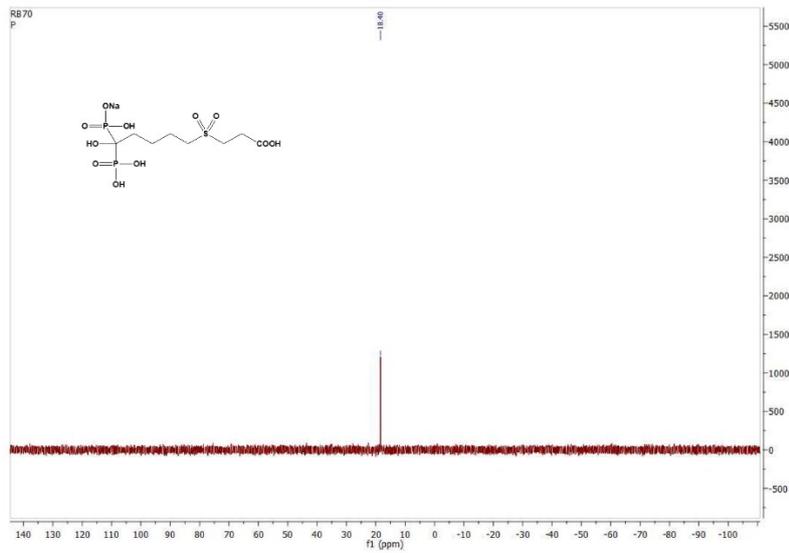
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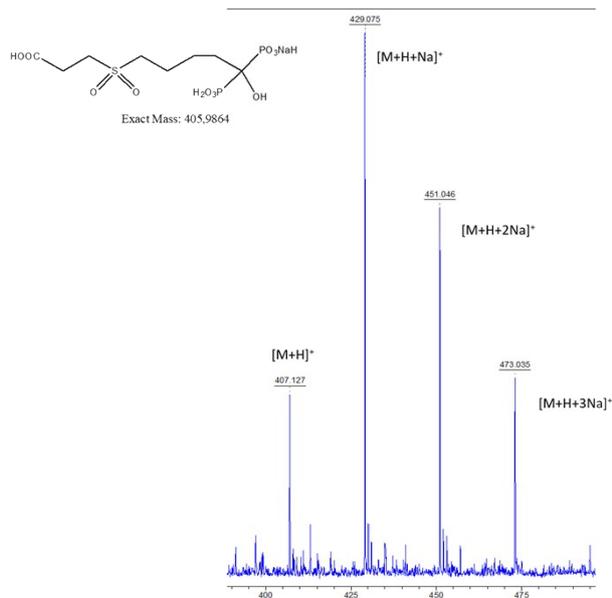


Compound 2b RMN 31P, 1H and 13C

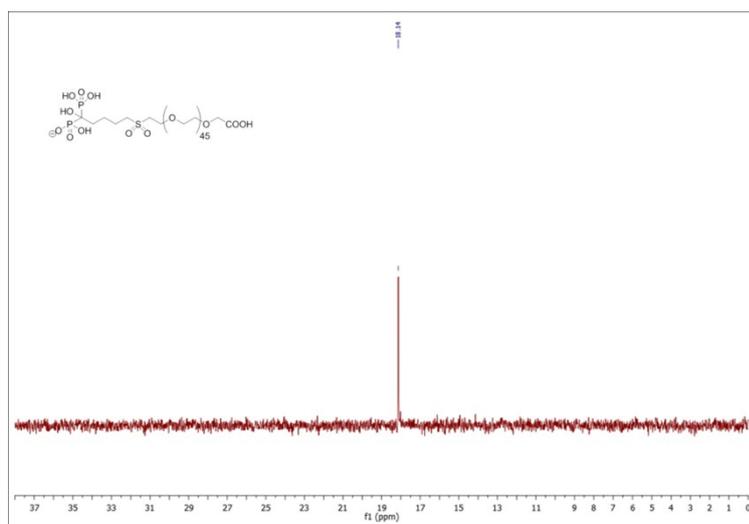


Compound 3a RMN 31P, 1H, 13C and MS spectra

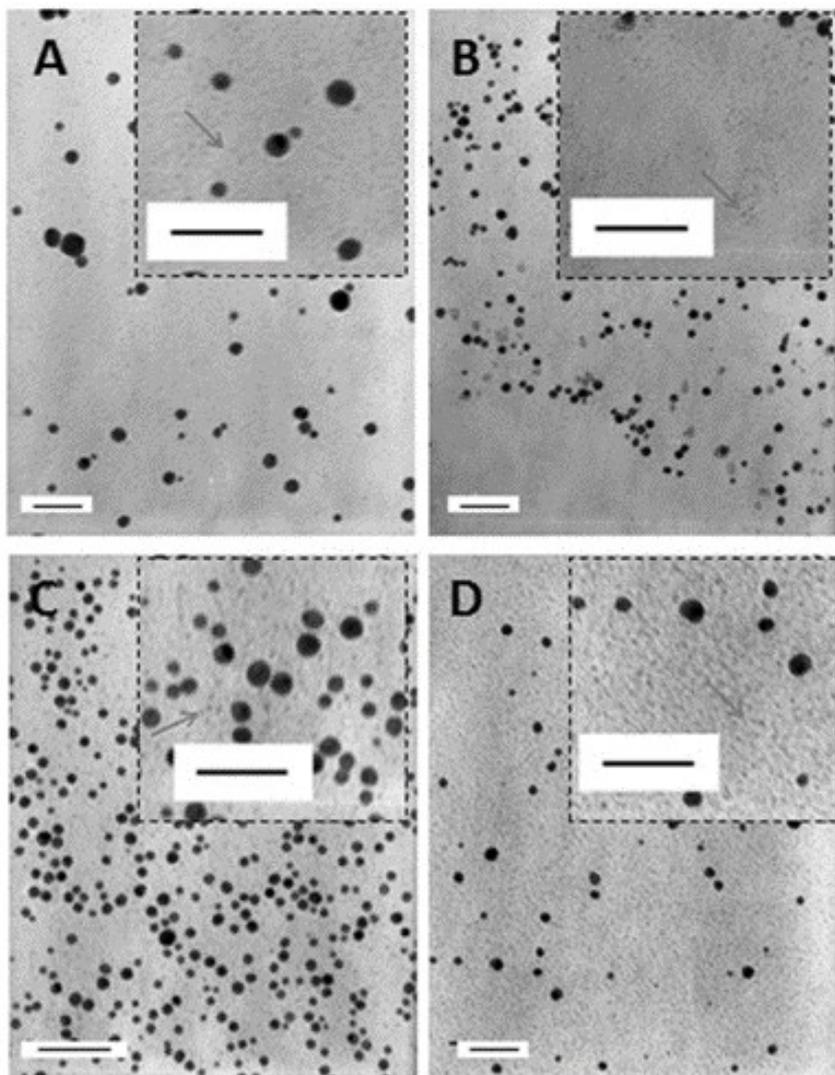




Compound 3c RMN 31P, 1H and 13C



Au NP obtained with compound 2c at: pH = 4.1 (A); pH = 8 (B); pH = 9.6 (C); pH = 11.8 (D) (scale bare is 50 nm). On insert arrows are indicating the small Au seeds.



Au NPs obtained with compound 3c at: pH = 4.6 (A); pH = 6.5 (B); pH = 8.5 (C); pH = 10.4 (D) (scale bare is 500 nm).

