

## Supplementary information

### A highly sensitive and fast responsive naphthalimide-based fluorescent probe for Cu<sup>2+</sup> and its application

Beibei Zhang<sup>a</sup>, Fengyun Qin<sup>a</sup>, Huawei Niu<sup>a</sup>, Yao Liu<sup>a</sup>, Di Zhang,<sup>b\*</sup> Yong Ye<sup>a,c,\*</sup>

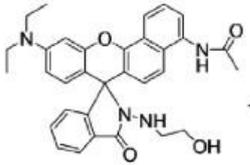
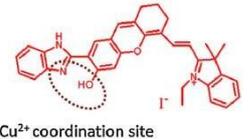
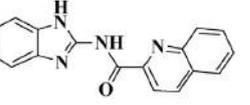
<sup>a</sup> Phosphorus Chemical Engineering Research Center of Henan Province, the College of Chemistry and Molecular Engineering, Zhengzhou University, Zhengzhou, China.

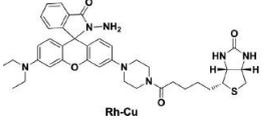
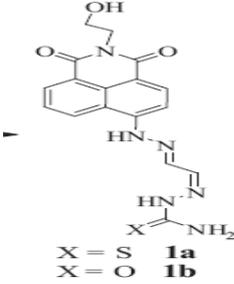
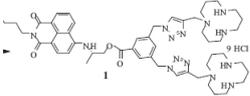
Email address: [yeyong03@tsinghua.org.cn](mailto:yeyong03@tsinghua.org.cn)

<sup>b</sup> Institute of Agricultural Quality Standards and Testing Technology, Henan Academy of Agricultural Sciences, Zhengzhou, China. Email address: [pandy811@163.com](mailto:pandy811@163.com)

<sup>c</sup> Key Laboratory of Bioorganic Phosphorus Chemistry & Chemical Biology (Ministry of Education), Department of Chemistry, Tsinghua University, Beijing, China.

**Table S1.** Comparison of some reported probes for Cu<sup>2+</sup>

Probe	Linear range	Detection limit	Response time	applications	Ref.
 BF-Cu	/	7.8x10 <sup>-8</sup> M	30 min	L929 cells	25
 Cu <sup>2+</sup> coordination site	1 × 10 <sup>-7</sup> to 5 × 10 <sup>-7</sup> M	14 nM	/	Hela cells	21
	0-50 uM	1.35 × 10 <sup>-7</sup> M	4 min	HeLa cells	22

N-doped carbon dots from lemon juice	0 to 15 $\mu\text{M}$ .	0.047 $\mu\text{M}$	few minutes.	river water sample	20
A hydrophobic carbon dots (HCDs) prepared by self-assembly of polymer DSPE-PEG	$2 \times 10^{-7}$ to $1 \times 10^{-6}$ M	36 nM	/	live HL-7702 cells	23
	0–40 $\mu\text{M}$	0.15 $\mu\text{M}$	30 min	HeLa cells 4T1 cells	24
	/	0.0326 $\mu\text{M}$	/	HeLa cells	17
	0–10 $\mu\text{M}$	$1.3 \times 10^{-8}$ M	/	HeLa and A549 cells	28
naphthalimide-based fluorescent probe containing diglycolamine	0-10 $\mu\text{M}$	49 nM	20 s	MCF-7 cells	This work

"/" not mentioned

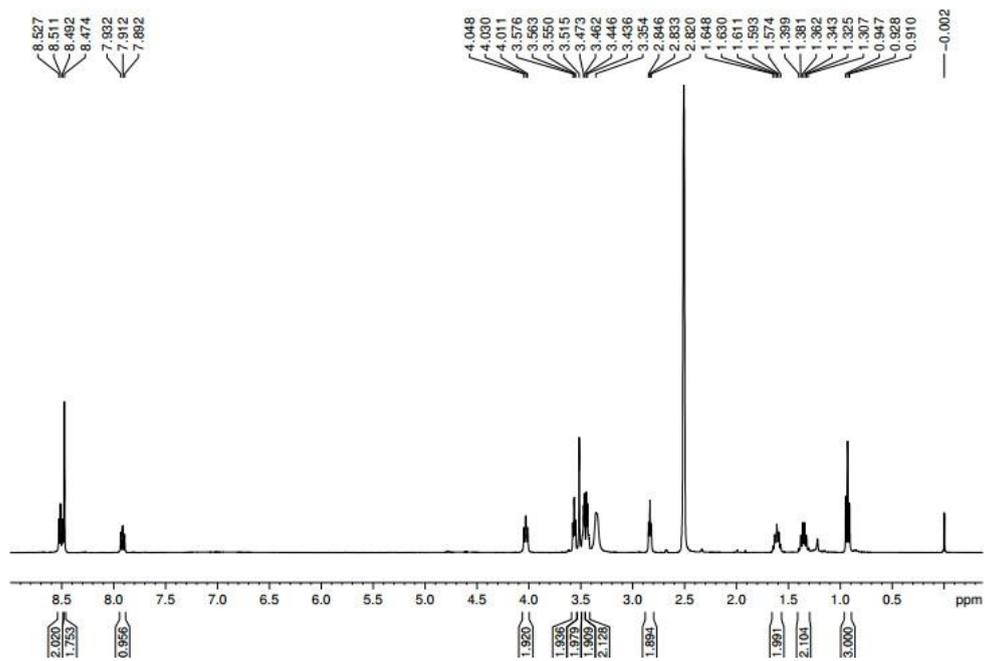


Figure S1.  $^1\text{H}$  NMR spectra of L (400 MHz, DMSO)

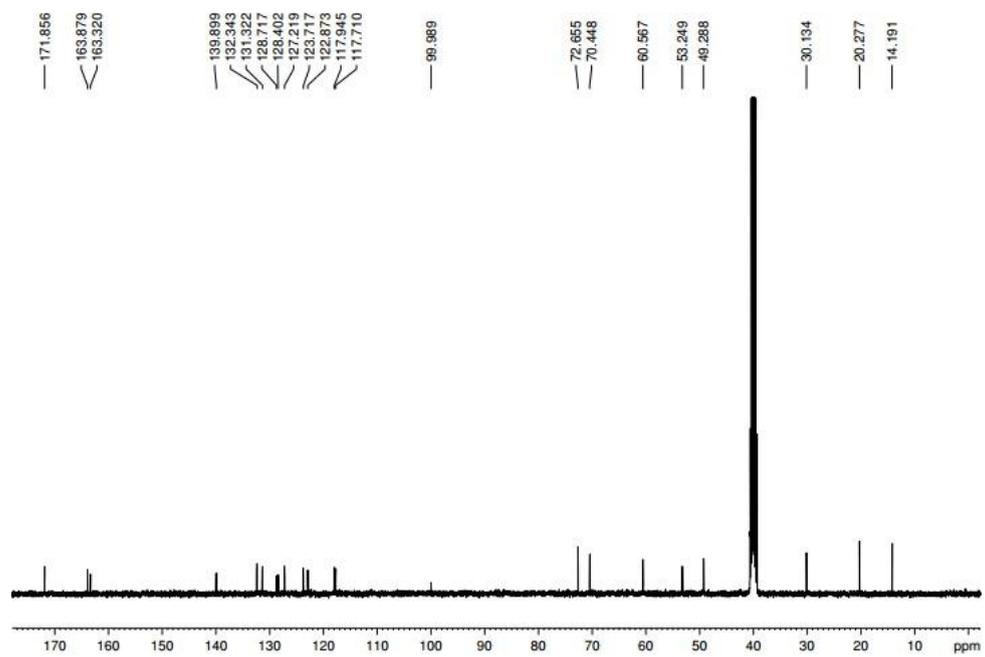


Figure S2.  $^{13}\text{C}$  NMR spectra of L (400 MHz, DMSO)

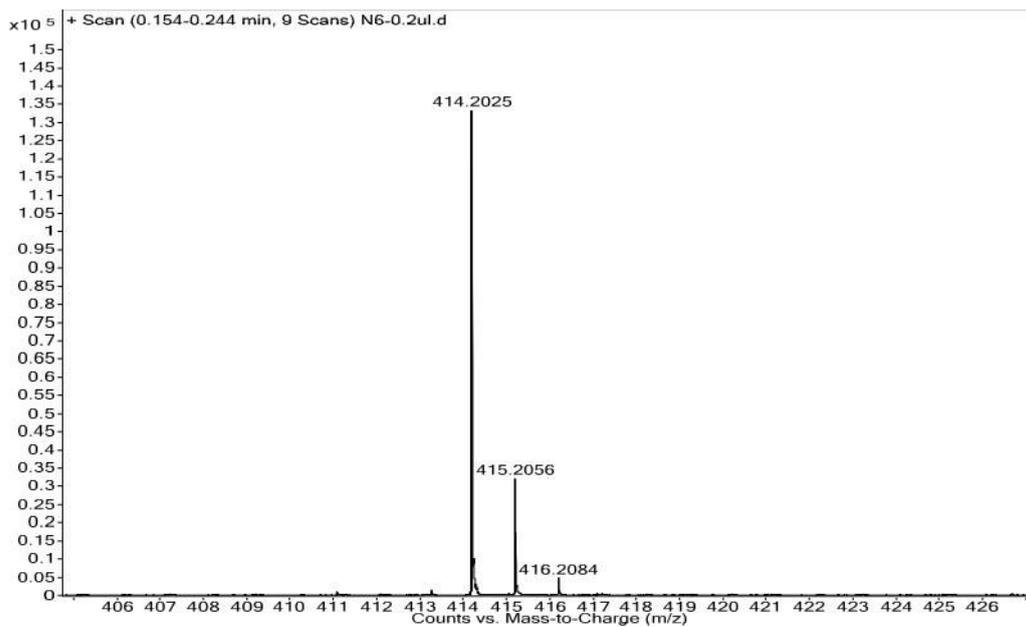


Figure S3. HR-MS of L

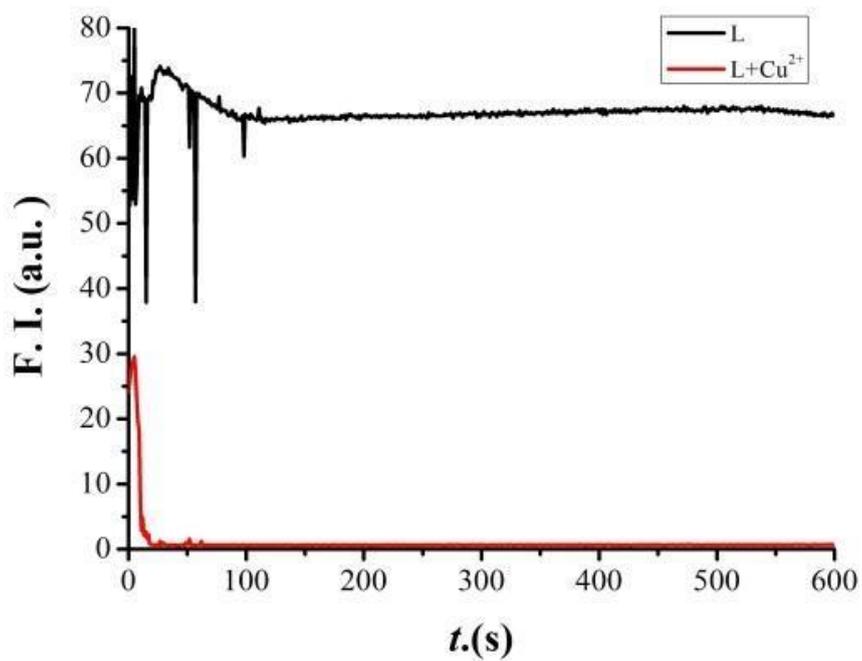


Figure S4. Changes in fluorescence intensity of L (10  $\mu$ M) solution versus time in the presence of 10.0 equiv. of Cu<sup>2+</sup>.

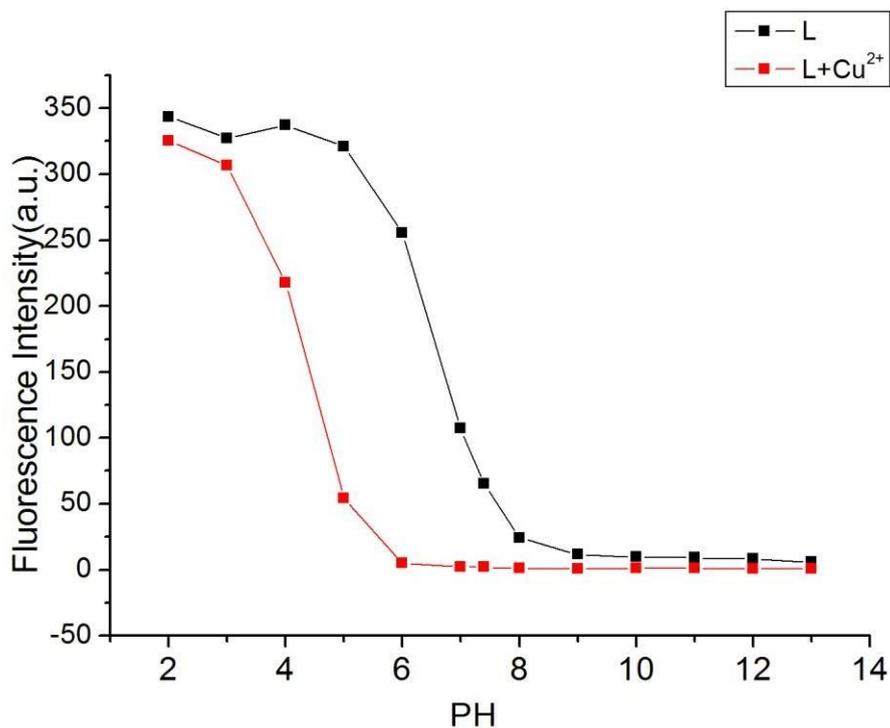


Figure S5. Fluorescence intensity of probe **L** (10  $\mu\text{M}$ ) at 463 nm under different pH values in the absence and the presence of 10 equiv. of  $\text{Cu}^{2+}$  (100  $\mu\text{M}$ ) measured in HEPES buffer (1.0 mM, pH = 7.4, containing 10% DMF)

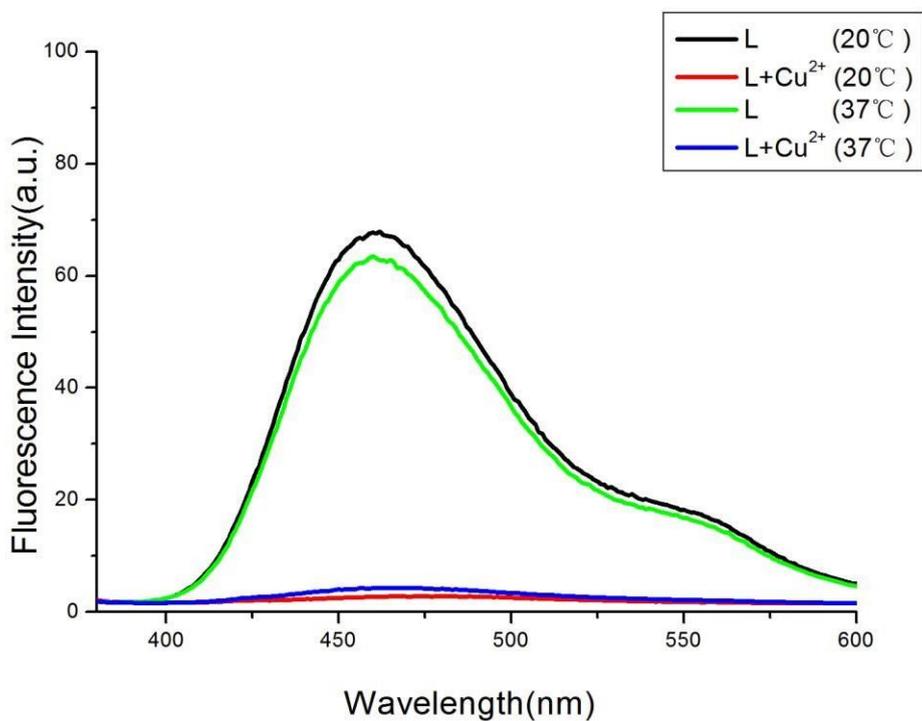
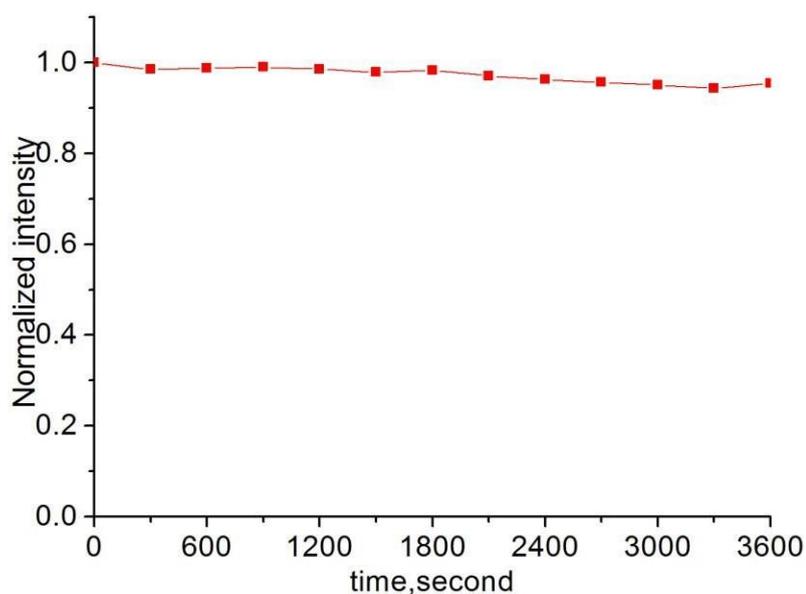
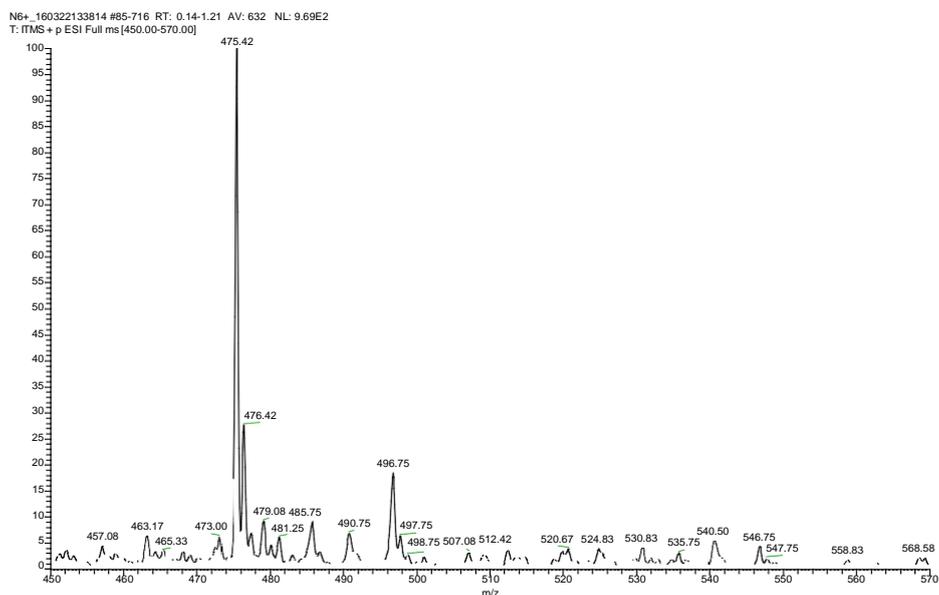


Figure S6. Fluorescence intensity of probe **L** (10  $\mu\text{M}$ ) at 463 nm under different temperature in the absence and the presence of 10 equiv. of  $\text{Cu}^{2+}$  (100  $\mu\text{M}$ ) measured in HEPES buffer (1.0 mM, pH = 7.4, containing 10% DMF).



**Figure S7.** Photostability of **L** in HEPES buffer (1.0 mM, pH = 7.4, containing 10% DMF). The samples were continuously irradiated by a xenon lamp (150 W) at 5 nm slit width at the maximal absorption wavelength of 370nm.



**Figure S8.** ESI-MS spectrum of **L-Cu<sup>2+</sup>** in **CH<sub>3</sub>OH** solution.

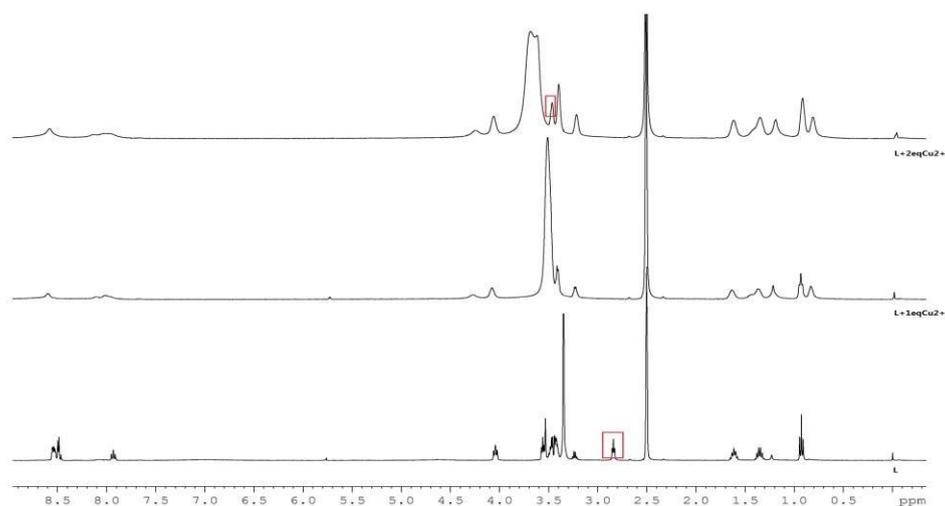


Figure S9.  $^1\text{H}$  NMR (400 MHz) spectra of (down) free L in  $\text{d}_6\text{-DMSO}$ , (middle) L + 1 eq.  $\text{Cu}^{2+}$ , (upper) L + 2 eq.  $\text{Cu}^{2+}$ .

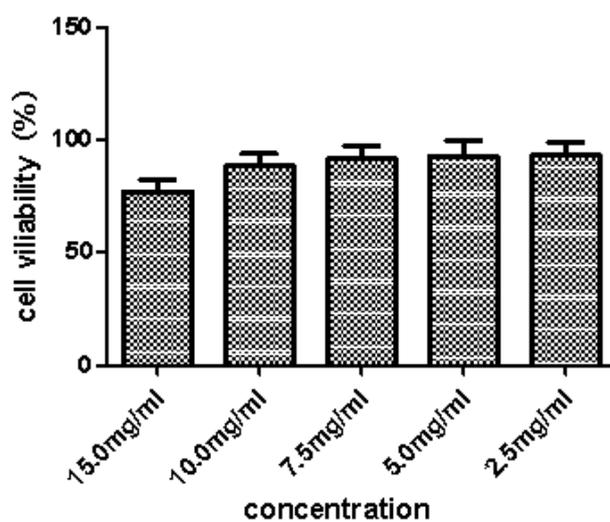


Figure S10. The cell viability of living MCF-7 cell treated with L of various concentrations (2.5 , 5, 7.5 , 10 , or 15  $\mu\text{M}$ ) for 24 hours measured by standard MTT assay at  $37^\circ\text{C}$ .