

Supplementary Information

Synthesis, Characterization and *in vitro* screening of nano-Hydroxyapatite/Chitosan/Euryale ferox nanoensemble- An inimitable approach for Bone Tissue Engineering

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1. Experimental

1.1. Synthesis of nano-hydroxyapatite

Wet chemical synthesis was performed based on the use of $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ and $(\text{NH}_4)_2\text{HPO}_4$ (DAHP) precursors for the preparation of n-HA nanocrystals where 0.6mol of $\text{Ca}(\text{NO}_3)_2 \cdot 4\text{H}_2\text{O}$ was added drop-wise to 0.4mol of DAHP at constant stirring in order to obtain Ca/P ratio of 1.67. The pH value of the mixture was maintained above 10 throughout the precipitation process by the addition of ammonia (25%) solution. The resultant precipitate obtained by vacuum filtration was rinsed with deionised water and oven dried at 85 °C.^{1,2}

2. Characterizations

2.1 Agar Diffusion Method

The inoculum was prepared by diluting the overnight cultures with sterile normal saline to a 0.5 McFarland standard. The agar petri plates were prepared by spreading the 1×10^8 CFU per 50 μL of mature broth culture of specific bacterial strains with a sterile L-shaped glass rod. Now, the stock solution of formulations of n-HA/CS/EF nanocomposite suspended in sterile PBS was used to evaluate the antibacterial activity. The experimental procedure was performed under sterile conditions using bio-safety level 2 (BSL-2) hoods. The petri plates were incubated at 37 °C overnight. The susceptibility of our formulation was determined on the basis of zone of clearance developed against human pathogenic strains *E.coli* and *S.aureus*. The inhibition zone was analysed thrice and the average was calculated for the antibacterial activity and compared with the reference drug vancomycin as a control.³

2.2. RBC Lysis test

In vitro erythrocyte lysis test was carried out as a primary toxicity test of the as-synthesized nanocomposite formulations, which is evaluated by quantifying the haemoglobin released as a result of membrane leakage or disruption caused by incorporation of low doses of the n-HA/CS/EF nanocomposite. In brief, centrifugation of the fresh blood procured from a healthy rabbit collected in anticoagulant solution (EDTA) was performed at 1000g for 10 min at 4°C. Both buffy coat and plasma were discarded. Washed erythrocytes were diluted with isotonic buffer (20 mM PBS) to prepare 50% haematocrit. n-HA/CS/EF nanocomposite was incubated in RBC suspension with different concentrations at 37 °C for 1 h to determine the degree of haemolysis.⁴ After the centrifugation of incubated solutions at 1500g for 15min, the supernatant obtained was characterized by UV-visible spectroscopy ($\lambda_{\text{max}} = 576\text{nm}$). The haemolysis percent was measured by the following equation:

$$\% \text{ Haemolysis} = \{(\text{Abs(T)} - \text{Abs(C)} / \text{Abs(100\%)} - \text{Abs (C)})\} * 100$$

where, Abs(T) = Absorbance of the supernatant from samples incubated with n-HA/CS/EF.

Abs(c) = Absorbance of the supernatant from controls (normal saline).

Abs(100%) = Absorbance of the supernatant of controls incubated in the presence of 1% Triton®

X-100, which causes complete lysis of RBCs (total lysis).

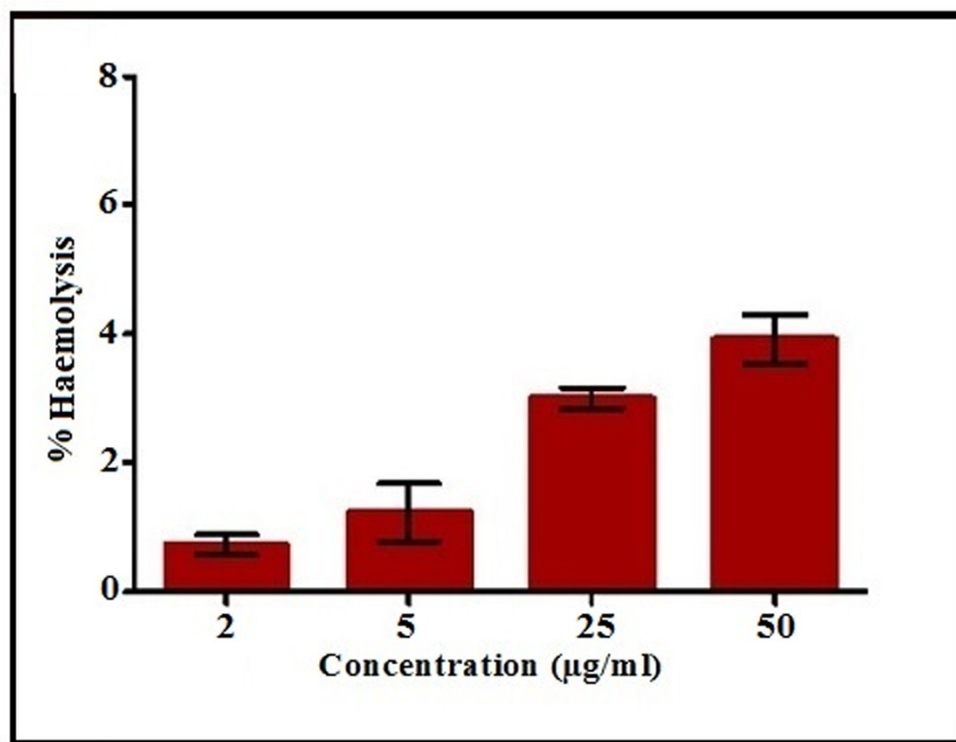


Fig. S1. RBC lysis of n-HA/CS/EF nanocomposite

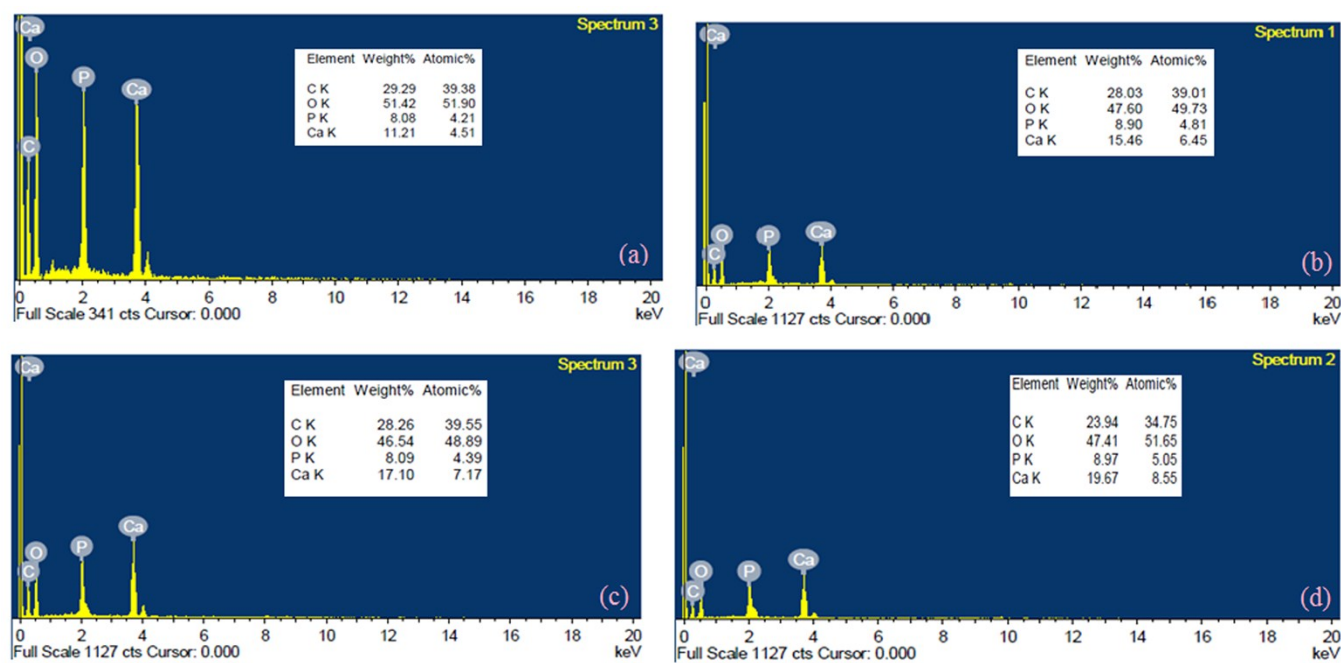


Fig. S2. EDX of (a) n-HA/CS (b) n-HA/CS/EF nanocomposite and their respective SBF study after 30 days (c-d).

References

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