Electronic Supplementary Material (ESI) for New Journal of Chemistry. This journal is © The Royal Society of Chemistry and the Centre National de la Recherche Scientifique 2018

Anticarcinogenic and metal chelation properties of novel hydroxybenzylidene-1-indanone derivatives in the U-251 glioblastoma cell line

Mariana Lozano-González,*a María Teresa Ramírez-Apan,^b Antonio Nieto,^b R. Alfredo Toscano^b and Cecilio Álvarez-Toledano*^b

SUPPORTING INFORMATION

Contents

General Information	2S-4S
General experimental procedure	5 S
Characterization Data for the Products	6S-9S
Copies of spectra of products	10S-41S

Experimental section

General materials and methods

All reagents and solvents were obtained from commercial suppliers (Sigma Aldrich México, Fluka México) and used without further purification. Fe₂(CO)₉ was synthesized from Fe(CO)₅ according to the literature.¹ All compounds were characterized by IR spectra, recorded on a Perkin-Elmer 283B or 1420 spectrophotometer, by the KBr technique, and all data are expressed in wavenumbers (cm⁻¹). Melting points were obtained on a Melt-Temp II apparatus and are uncorrected. Nuclear magnetic resonance spectra were recorded with a Bruker AV 400 or JEOL Eclipse +300 spectrometer. Chemical shifts for the ¹H NMR spectra were recorded in parts per million from tetramethylsilane with the solvent resonance as the internal standard (chloroform, δ = 7.25 ppm). Chemical shifts for the ¹³C NMR spectra were recorded in parts per million from tetramethylsilane using the central peak of CDCl₃ (δ = 77.1 ppm) as the internal standard. Mass spectra were recorded with a JEOL JMSAX 505 HA spectrometer at 70 eV using the electronic impact (EI), (DART) and fast atom bombardment (FAB⁺) technique.

Crystallographic structure determination

A suitable X-ray-quality crystal for **6** and **8** were grown by slow evaporation of an *n*-hexane-CH₂Cl₂ mixture at room temperature. The crystal was mounted on a glass fiber at room temperature, and then placed on a Bruker Smart Apex CCD diffractometer, equipped with Mo K α radiation; decay was negligible. Structure solutions and refinements were performed using SHELXTL V6.10.^{2,3}

Cytotoxicity Assay.

The compounds were screened *in vitro* against six human cancer cell lines: U251 (human glioblastoma), HCT-15 (human colorectal adenocarcinoma), and SKLU-1 (human lung adenocarcinoma), PC-3 (human prostatic adenocarcinoma), K562 (human chronic myelogenous leukemia), MCF-7 (human mammary adenocarcinoma), cell lines were supplied by National Cancer Insitute (USA). The human tumor cytotoxicity was determined using the protein-binding dye sulforhodamine B (SRB) in microculture assay to measure cell growth, as described in the protocols established by the NCI. The cell lines were cultured in RPMI-1640 medium supplemented with 10% fetal bovine serum, 2 mM L-glutamine, 10000 units/mL penicillin G sodium, 10000 μg/mL streptomycin sulfate, and 25 μg/mL amphotericin B (Gibco) and 1% nonessential aminoacids (Gibco). They were maintained at 37° in humidified atmosphere with 5% CO₂. The viability of the cells used in the experiments exceeds 95% as determined with trypan blue. The cells were removed from the tissue culture flask and diluted with fresh media. Of this cell suspension, 100 μL containing 5000 or 10000 cells per well, were pipetted into 96-well microtiter plates (Costar) and the material was incubated at 37 °C for 24 h in a 5% CO₂ atmosphere. Subsequently, 100 μ L of a solution of the test compounds obtained by diluting the stocks were added to each well. The cultures were exposed for 48 h to the drug at concentrations ranging from 1.0 to 50 μ M. After the incubation period, cells were fixed to the plastic substratum by the addition of 50 μ L of cold 50% aqueous trichloroacetic acid. The plates were incubated at 4 °C for 1 h. Washed with tap H₂O, and air-dried. The trichloroacetic-acid-fixed cells were stained by the addition of 0.4% SRB. Free SRB solution was removed by washing with 1% aqueous acetic acid. The plates were airdried, and the bound dye was solubilized by the addition of 10 μ M unbuffered tris based (100 μ L). The plates were placed on a shacked for 5 min, and the absorption was determinate at 515 nm using an ELISA plates reader (Bio-Tex Instruments).

Polymophonuclear (PMN) assay

For the test of PMN the protocol reported by Ivan. J. Fuss et al.⁴ was followed.

Statistical analysis

All biological experiments were performed at least twice with triplicates in each experiment. Representative results were depicted in this report. Data were presented as means + standard deviations.

Lipid peroxidation inhibition

Animals. Adult male Wistar rat (200-250 g) was provided by the Instituto de Fisiología Celular, Universidad Nacional Autónoma de México (UNAM). Procedures and care of animals were conducted in conformity with Mexican Official Norm for Animal Care and Handling (NOM-062-ZOO-1999). They were maintained at 23±2 °C on a 12/12 h light-dark cycle with free access to food and water.

Rat Brain Homogenate Preparation. Animal sacrifice was carried out avoiding unnecessary pain. Ten rats were sacrificed with CO_2 to carry out all experiments. The cerebral tissue (whole brain), was rapidly dissected and homogenized in phosphate buffered saline (PBS) solution (0.2 g of KCl, 0.2 g of KH₂PO₄, 8 g of NaCl, and 2.16 g of NaHPO₄ ·7 H₂O/l, pH adjusted to 7.4) as reported elsewhere ^{5,6} to produce a 1/10 (w/v) homogenate. Then, the homogenate was centrifuged for 10 min at 800 rcf (relative centrifugal field) to yield a pellet that was discarded. The supernatant protein content was measured using the Folin and Ciocalteu's phenol reagent ⁷ and adjusted with PBS at 2.666 mg of protein/ml.

Induction of lipid peroxidation and Thiobarbituric Acid Reactive Substances (TBARS) Quantification.

As an index of lipid peroxidation, TBARS levels were measured using rat brain homogenates according to the method described by Ng et al.⁸, with some modifications. Supernatant (375 μ l) was added with 50 μ l of 20 μ M EDTA and 50 μ l of each sample concentration solved in DMSO (50 μ l of DMSO for control group) and incubated at 37 °C for 30 min. Lipid peroxidation was started adding 50 μ L of freshly prepared 100 μ M FeSO₄ solution (final concentration 10 μ M), or AAPH and incubated at 37 °C for 1 hour. The TBARS content was determined as described by Ohkawa et al.⁹ with some modifications. 500 μ l of TBA reagent (1% 2-thiobarbituric acid in 0.05 N NaOH and 30% trichloroacetic acid, in 1:1 proportion) was added at each tube and the final suspension was cooled on ice for 10 min, centrifugated at 13400 rcf for 5 min and heated at 80°C in a water bath for 30 min. After cooling at room temperature, the absorbance of 200 μ L of supernatant was measured at λ =540 nm in a Bio-Tek Microplate Reader Synergy HT. Concentration of TBARS was calculated by interpolation in a standard curve of tetra-methoxypropane (TMP) as a precursor of MDA. Results were expressed as nmoles of TBARS per mg of protein. The inhibition ratio (I_R[%]) was calculated using following formula I_R=(*C*-*E*)*100/*C*, where *C* is the absorbance of control and *E* is the absorbance of the test sample. Butylated hydroxytoluene (BHT) and α -tocopherol were used as positive standards.

All data were represented as mean \pm standard error (SEM). Data were analyzed by one-way ANOVA followed by Dunnett's test for comparison against control. Values of $p \le 0.05$ (*) and $p \le 0.01$ (**) were considered statistically significant. The inhibitory concentration 50 (IC₅₀), was estimated by means of a linear regression.

Synthesis and characterizations

General procedure for the synthesis of 2-benzyliden-1-indanones derivatives.

The reaction between *o*-phthalaldehyde and acetophenone in a basic medium was performed to produce the 2-benzyliden-1-indanone type compound under simpler and faster conditions. *O*-phthalaldehyde was added slowly to a cool sodium hydroxide (1.5 eq.) ethanolic solution with the appropriate amount of acetophenone. The reaction mixture was stirred at room temperature for approximately 3 h, and then poured into a mixture of ice and commercial hydrochloric acid (pH was adjusted to about 7). The resulting solid was filtered and in some cases purified by column chromatography using hexane/ethyl acetate. All compounds are stable to aire and temperature, they are soluble in ether, CH₂Cl₃, CHCl₃, DMSO, ethyl acetate and partially soluble in ethanol.

General procedure for the synthesis of iron (III) complex of (Z)-2-(1-arylhydroxiliden)-1-indanone

a) A solution of (Z)-2-(1-arylhydroxiliden)-1-indanone in anhydrous ethyl ether (20 mL) was treated with $Fe_2(CO)_9$ or $Fe(CO)_5$ at room temperature for 2-3 h, under an inert atmosphere. After the reaction was complete, the crude product was filtered off through a Celite column, washed with ethyl ether and dissolved in dichloromethane. The solvent was evaporated to dryness. The iron complex (III) was obtained pure.

b) A solution of (Z)-2-(1-arylhydroxiliden)-1-indanone in anhydrous ethyl ether (10mL) was treated with FeCl₃ or FeCl₂ at room temperature for 3h, under an inert atmosphere. After the reaction was complete, the crude product was filtered off through a Celite column, washed with ethyl ether and crystallized in dichloromethane/hexane. All compounds are stable to aire and temperature, they are soluble in CH₂Cl₃, CHCl₃, DMSO and ethyl acetate.

General procedure for the synthesis of cupper (II) complex of (Z)-2-(1-arylhydroxiliden)-1-indanone

A mixture 2 eq. of (Z)-2-(1-arylhydroxiliden)-1-indanone and 1 eq. of cupper (0) in DMSO (20 mL) at room temperature was agitated during 7 days, under an inert atmosphere. After the reaction was complete, the crude product was filtered, washed with ethyl ether and dissolved in dichloromethane. The solvent was evaporated to dryness. The cupper complex (II) was obtained pure. Compound is stable to aire and temperature, it is soluble in CH₂Cl₃, CHCl₃, DMSO and ethyl acetate.

Compound 1 ($C_{16}H_{12}O_2$, *M*=236 g/mol) was prepared starting from *o*-phthalaldehyde (0.5 g, 3.73 mmol), acetophenone (0.48g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid, **mp**. 90°C, (0.73g, 83%). **IR**: v 1604, 1564 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 15.067(s, 1H, OH), 7.49, 7.5(d, 1H, 1), 7.54 (ddd, 1H, 2), 7.40 (dd,1H,3), 7.85(d, 1H, 4), 7.91(m, 1H,10), 7.48 (m,3H,11,12,13), 7.91(m, 1H,14), 3.86 (sa, 2H, 15). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.54 (C1), 133.27 (C2), 127.40 (C3), 123.36 (C4), 137.85 (C5), 195.70 (C6), 109.41 (C7), 170.79 (C8), 134.79 (C9), 128.07 (C10), 128.55 (C11), 131.21 (C12), 128.55 (C13), 128.07 (C14), 32.19 (C15), 148.51(C16). **MS (EI):** m/z (%) = 236 (1.9%). **HRMS** (FAB⁺): calculated for C₁₆H₁₂O₂: 237.0821. Found: 237.0823.

Compound 2 ($C_{16}H_{11}ClO_2$, *M*=270.5 g/mol) was prepared starting from *o*-phthalaldehyde (0.5 g, 3.73 mmol), 4'-chloroacetophenone (0.69g, 4.48mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale brown solid, **mp**. 162-164°C, (0.86g, 85%). **IR**: v 1605, 1559 cm⁻¹. ¹**H NMR** (300

MHz, CDCl₃): δ = 15.03 (s, 1H, OH), 7.54 (d, 1H9, 1), 7.60 (d, 1H, 2), 7.44 (dd, 1H, 3), 7.86 (dd, 1H, 4), 7.89 (d, 2H, 10, 14), 7.48 (d, 2H, 11, 13), 3.91 (s, 2H, 15). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.45 (C1), 133.35 (C2), 127.35 (C3), 123.19 (C4), 137.44 (C5), 195.63 (C6), 109.29 (C7), 169.14 (C8), 132.99 (C9), 129.22 (C10), 128.70 (C11), 137.22 (C12), 128.70 (C13), 129.22 (C14), 31.95 (C15), 148.19 (C16). **MS (EI)**: m/z (%) = 270 (15%). **HRMS** (FAB⁺): calculated for C₁₆H₁₁ClO₂: 271.0526. Found: 271.0522.

Compound 3 ($C_{17}H_{14}O_2$, *M*=250 g/mol) was prepared starting from *o*-phthalaldehyde (0.5 g, 3.73 mmol), 4'-methylacetophenone (0.60g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid, **mp**. 96-98°C, (0.79g, 85%). **IR:** v 1604, 1540 cm ⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 15.17 (s, 1H, OH), 7.50 (d, 1H, 1), 7.55 (dd, 1H, 2), 7.41 (dd, 1H, 3), 7.85(d, 1H, 4), 7.84 (d, 1H, 10), 7.30 (d, 2H, 11,13), 7.84 (d, 1H, 14), 3.89 (s, 2H, 15), 2.41 (s, 3H, 17). ¹³**C NMR** (75MHz, CDCl₃): δ =125.50 (C1), 133.11 (C2), 127.35 (C3), 123.27 (C4), 137.95 (C5), 195.45 (C6), 109.01 (C7), 171.09 (C8), 131.97 (C9), 128.08 (C10), 129.29 (C11), 141.89 (C12), 129.29 (C13), 128.08 (C14), 32.32 (C15), 148.39 (C16), 21.52 (C17). **MS (EI)**: m/z (%) = 250 (20%). **HRMS** (FAB⁺): calculated for C₁₇H₁₄O₂: 251.1072. Found: 251.1079.

Compound 4 ($C_{17}H_{14}O_3$, *M*=266 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.73 mmol), 4'-methoxyacetophenone (0.672g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid, **mp**.112-114 °C, (0.79 g, 80%). **IR**: v 1601, 1566 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 15.33(s, 1H, OH), 7.47 (d, 1H, 1), 7.51 (dd, 1H, 2), 7.37 (dd, 1H,3), 7.82 (d, 1H, 4), 7.89 (d, 2H, 10, 14), 6.95 (d, 2H, 11, 13), 3.82 (s, 2H, 15), 3.83 (s, 3H, 17). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.37 (C1), 132.85 (C2), 127.23 (C3), 123.03 (C4), 137.87 (C5), 194.84 (C6), 108.32 (C7), 170.04 (C8), 127.00 (C9), 129.99 (C10), 113.88 (C11), 162.06 (C12), 113.88 (C13), 129.99 (C14), 32.45 (C15), 148.09 (C16), 55.30 (C17). **MS (EI)**: m/z (%) = 266 (15%). **HRMS** (FAB⁺): calculated for C₁₇H₁₄O₃: 267.1021. Found: 267.1023.

Compound 5 ($C_{16}H_{11}O_2Br$, *M*=314 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.73 mmol), 4'-bromoacetophenone (0.89g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid **mp**.160-162 °C, (0.46 g, 40%). **IR**: v 1622, 1555 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 15.03(s, 1H, OH), 7.57 (m, 1H, 1, 2, 4, 11, 13), 7.44 (dd, 1H, 3), 7.81 (d, 1H, 10), 7.89 (d, 1H, 14), 3.90 (s, 3H, 15). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.67 (C1), 133.61 (C2), 127.62 (C3), 123.57 (C4), 137.74 (C5), 194.04 (C6), 109.57 (C7), 169.34 (C8), 130.00 (C9), 131.94 (C10), 129.63 (C11), 135.12 (C12),

129.63 (C13), 131.94 (C14), 32.20 (C15), 148.44 (C16). **MS (EI)**: m/z (%) = 314 (20%). **HRMS** (FAB⁺): calculated for C₁₆H₁₂O₂Br [M⁺]_: 315.0021 Found: 315.0017.

Compound 6 ($C_{14}H_{10}O_2S$, *M*=242 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.73 mmol), 2-thioacetophenone (0.57g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid **mp**.130-132 °C, (0.7 g, 77%). **IR**: v 1709, 1604 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 14.83(s, 1H, OH), 7.56 (m, 2H, 1, 2), 7.44 (dd, 1H, 3), 7.67 (d, 1H, 4), 7.81 (d, 1H, 10), 7.22 (m, 1H, 11), 7.85 (d, 1H, 12), 3.89 (s, 2H, 13). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.68 (C1), 133.20 (C2), 127.54 (C3), 123.21 (C4), 138.31 (C5), 194.12 (C6), 108.07 (C7), 166.09 (C8), 135.12 (C9), 128.29 (C10), 130.79 (C11), 131.62 (C12), 32.18 (C13), 147.83 (C14). **MS (EI)**: m/z (%) = 242 (50%). **HRMS** (FAB⁺): Found: $C_{15}H_{10}O_2S$ [M⁺⁻] 243.0480.

Compound 7 ($C_{15}H_{11}O_2N$, *M*=237 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.7mmol), 2-acetylpyridine (0.54g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid **mp**.136-138 °C, (0.7 g, 70%). **IR**: v 1615, 1559 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 14.75(s, 1H, OH), 7.50 (m, 3H, 10, 11, 12), 7.78 (d, 1H, 1), 8.1 (d, 1H, 2), 7.66 (dd, 1H, 3), 8.44 (d, 1H, 4), 8.65 (d, 1H, 13), 3.87 (s, 2H, 14). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.02 (C1), 130.11 (C2), 126.82 (C3), 127.02 (C4), 133.64 (C5), 183.11 (C6), 114.31 (C7), 168.11 (C8), 153.12 (C9), 122.36 (C10), 139.96 (C11), 124.56 (C12), 149.16 (C13), 35.59 (C14), 144.63 (C15). **MS (EI):** m/z (%) = 237 (100%). **HRMS** (FAB⁺): calculated for C₁₅H₁₂O₂N [M⁺]: 238.0868 Found: 238.0874.

Compound 8 ($C_{20}H_{16}O_2Fe$, *M*=344 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.7mmol), acetylferrocene (1.02g, 4.48 mmol) and *t*BuOK (0.42g, 3.73mmol) and was obtained as a red solid **mp**.140-141 °C, (0.60 g, 45%). **IR**: v 1708, 1603 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 14.98(s, 1H, OH), 7.87 (d, 1H, 4), 7.58 (m, 2H, 1, 2), 7.47 (dd, 1H, 3), 4.97 (s, 2H, 10,13), 4.58 (s, 2H, 11,12), 4.21 (s, 5H, 14), 3.80 (s, 2H, 15). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.59 (C1), 132.5 (C2), 127.43 (C3), 122.97 (C4), 138.62 (C5), 192.33 (C6), 107.94 (C7), 177.57 (C8), 132.5 (C9), 71.91 (C10, 13), 69.27 (C11, 12), 70.27 (C14), 31.90 (C15), 147.43 (C16). **MS (DART):** m/z (%) = 344 (20%). **HRMS** (ESI⁺): calculated for C₂₀H₁₇O₂Fe [M^{+.}]: 345.05780 Found: 345.05734.

Compound 9 ($C_{20}H_{14}O_2$, *M*=286 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.73 mmol), 2-acetylnaphthone (0.76g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid **mp**.102-104 °C, (0.80 g, 75%). **IR**: v 3056, 1674, 1617 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 15.23(s, 1H, OH), 7.94 (m, 7H), 7.55 (m, 4H), 4.00 (s, 2H, H15). ¹³C NMR (75MHz, CDCl₃): δ = 195.8, 170.8, 148.6, 137.9, 133.4, 120.2, 126.8, 125.7, 124.3, 109.8, 32.43. **MS (DART):** m/z (%) = 287 (360%). **HRMS** (ESI⁺): calculated for C₂₀H₁₅O₂ [M⁺⁻]: 287.10720 Found: 287.10694.

Compound 10 ($C_{15}H_{11}O_2N$, *M*=237 g/mol) was prepared starting from *o*-phthalaldehyde (0.5g, 3.7mmol), 4-acetylpyridine (0.54g, 4.48 mmol) and NaOH (0.22 g, 5.5 mmol) and was obtained as a pale yellow solid **mp**.136-138 °C, (0.7 g, 80%). **IR**: v 1615, 1559 cm⁻¹. ¹**H NMR** (300 MHz, CDCl₃): δ = 14.75(s, 1H, OH), 7.50 (m, 3H, 10, 11, 12), 7.78 (d, 1H, 1), 8.1 (d, 1H, 2), 7.66 (dd, 1H, 3), 8.44 (d, 1H, 4), 8.65 (d, 1H, 13), 3.87 (s, 2H, 14). ¹³**C NMR** (75MHz, CDCl₃): δ = 125.02 (C1), 130.11 (C2), 126.82 (C3), 127.02 (C4), 133.64 (C5), 183.11 (C6), 114.31 (C7), 168.11 (C8), 153.12 (C9), 122.36 (C10), 139.96 (C11), 124.56 (C12), 149.16 (C13), 35.59 (C14), 144.63 (C15). **MS (EI):** m/z (%) = 237 (100%). **HRMS** (FAB⁺): calculated for C₁₅H₁₂O₂N [M⁺⁻]. 238.0868 Found: 238.0874.

Compound 1a ($C_{48}H_{33}FeO_6$, *M*=761 g/mol) was prepared starting from **1** (0.5 g, 2.1 mmol), and $Fe_2(CO)_9$ (0.21 g, 0.56 mmol) and was obtained as a purple solid, **mp**. 244-246°C, (0.425 g, 80 %). **MS (FAB⁺):** m/z (%) = 761 (2%). **HRMS** (FAB⁺): calculated for $C_{32}H_{22}O_4Fe$ [M⁺.]: 526.0867. Found: 526.0877. **Compound 2a** ($C_{48}H_{30}Cl_3FeO_6$, *M*=863 g/mol) was prepared starting from **2** (0.5g, 2.1 mmol), and $Fe_2(CO)_9$ (0.18 g, 0.56 mmol) and was obtained as a purple solid, **mp**. 232-234°C, (0.48 g, 90%). **MS (FAB⁺)**: m/z (%) = 864 (2%). **HRMS** (FAB⁺): calculated for $C_{32}H_{20}O_4CIFe$ [M⁺.]: 594.0088. Found: 594.0092.

Compound 3a ($C_{51}H_{39}FeO_6$, *M*=803 g/mol) was prepared starting from **3** (0.5 g, 2.1 mmol), and $Fe_2(CO)_9$ (0.19 g, 0.56 mmol) and was obtained as a purple solid, **mp**. 238-240 °C, (0.33 g, 61%). **MS (FAB+)**: m/z (%) = 803 (5%). **HRMS** (FAB+): calculated for $C_{34}H_{26}O_4Fe$ [M⁺]. 554.1180. Found: 554.1187.

Compound 4a ($C_{51}H_{39}FeO_9$, *M*=851 g/mol) was prepared starting from **4** (0.5 g, 2.1 mmol), and $Fe_2(CO)_9$ (0.18 g, 0.56 mmol) and was obtained as a purple solid, **mp**. 234-236°C, (0.37 g, 70%). **MS (FAB⁺)**: m/z (%) = 851 (2%). **HRMS** (FAB⁺): calculated for $C_{34}H_{26}O_6Fe$ [M⁺⁻]. 586.1079. Found: 554.1082.

Compound 7a ($C_{45}H_{30}FeN_{3}O_{6}$, *M*=764 g/mol) was prepared starting from **8** (0.5 g, 2.1 mmol), and $Fe_{2}(CO)_{9}$ (0.21 g, 0.56 mmol) and was obtained as a dark green solid, **dp**. >280°C, (0.52 g, 97%). **IR**: v 3059, 1590, 1552 cm⁻¹**MS (FAB**⁺): m/z (%) = 765 (5%), 528 (40%).

Compound 8a ($C_{60}H_{45}Fe_4O_6$, *M*=1085 g/mol) was prepared starting from **9** (0.5 g, 2.1 mmol), and $Fe_2(CO)_9$ (0.14g, 0.56 mmol) and was obtained as a deep red solid, **dp**. 220-222°C, (0.49 g, 95%). **IR**: v 3082, 1589, 1551 cm⁻¹. **MS (FAB+)**: m/z (%) = 1085 (5%), 742 (10%). HRMS (FAB+): calculated for $C_{40}H_{30}O_4Fe_3$ [M+.]: 742.0192. Found: 742.0189.

Compound 9a (C₆₀H₃₉FeO₆, *M*=911 g/mol) was prepared starting from **10** (0.5 g, 2.1 mmol), and Fe₂(CO)₉ (0.17 g, 0.56 mmol) and was obtained as a deep red solid, **mp**. 196-198°C, (0.45 g, 85%). **IR**: v 3051, 1583, 1554 cm⁻¹. **MS (FAB+)**: m/z (%) = 911 (5%), 626 (10%). HRMS (FAB+): calculated for C₄₀H₂₆FeO₄ [M^{+.}]: 626.1180. Found: 626.1189.

Compound 1b (C₃₂H₂₂O₄Cu, *M*=533.5 g/mol) was prepared starting from 1 (0.5g, 2.1mmol) and Cu (0.05g, 0.79mmol) in DMSO and was obtained as a green solid, mp. >200 °C **IR**: v 3058, 1593, 1561. **MS (FAB+)**: m/z (%) = 534 (10%). **HRMS** (FAB+): calculated for C₃₂H₂₃O₄Cu [M^{+.}]: 534.0892. Found: 534.0883

REFERENCES

 H. Braye and W. Hu"bel, Inorg. Synth., 1966, 7, 178–181.
 G. Altomare, C. Cascarano and A. Giacovazo, Refinement of Crystal Structures, University of Goettingen, Germany, 1997.
 G. M. Sheldrick, Acta Crystallogr., Sect. A: Found. Crystallogr., 1990, 46, 467–473.
 I. J. Fuss, M. E. Kanof, P. D. Smith and H. Zola, Curr. Protoc. Immunol., 2009, S85, 7.1.1 S.
 M. Domi´nguez, A. Nieto, J. C. Marı´n, A. S. Keck, E. Jeffery and C. L. Ce´spedes, J. Agric. Food Chem., 2005, 53, 5889–5895.
 J. I. Rossato, L. A. Ketzer, F. B. Centuriao, S. J. Silva, G. Lu"eni, A. L. Braga, M. A. Rubin and B. T. Rocha, Neurochem. Res., 2002, 27, 297–303.
 O. H. Lowry, N. J. Rosebrough, A. L. Farr and R. J. Randall, J. Biol. Chem., 1951, 193, 265–275.
 T. B. Ng, F. Liu and Z. T. Wang, Antioxidative activity of natural products from plants. Life Sci., 2000, 66, 709–723. 9) H. Ohkawa, N. Ohishi and K. Yagi, Anal. Biochem., 1979, 95, 351–358.

10) M. Turek, D. Szczesna, M. Koprowski and P. Balczewski, Beilstein J. Org. Chem., 2017, 13, 451–494.

RMN Spectrum



Exist an equilibrium keto-enolic, enol form is greatly favored (90-95%)









¹³C RMN /Compound **1** (CDCl₃) 100MHz



¹³C RMN /Compound 2 (CDCl₃) 100MHz



Elemental Composition Compound 2

Sample: 2013	LC1-ME						
Note : -luis-ve	elasco						
Inlet : Direct			Io	n Mode	FAB+		
RT : 5.53 min			Sca	an#: 14			
Elements : C 40	0/0, H 58/0,	0 6/1.	C1 3/	0			
Mass Tolerance	: 1000	ppm. 1mr	nu if i	m/z > 1			
Unsaturation (I	J.S.) : 0.0	- 25.0		,			
Observed m/z	Int%						
271.0522	100.0						
Estimated m/z	Error [ppm]	U.S.	С	н	0	CI	
271.0526	-1.2	10.5	16	12	2	1	
			10		4	Т	





Elemental Composition Compound **3**

```
Sample: 1407
                СЗМе
Note : -luis-velasco
                                   Ion Mode : FAB+
Inlet : Direct
                                   Scan#: (14,16)
RT : 5.78 min
Elements : C 40/0, H 49/0, O 9/0
Mass Tolerance : 1000ppm, 1mmu if m/z > 1
Unsaturation (U.S.) : 4.0 - 12.0
Observed m/z
              Int%
  251.1079
              100.0
Estimated m/z Error[ppm] U.S.
                                 C
                                             0
                                       H
                +2.7
                                              2
                          10.5 17
                                       15
  251.1072
```







E	lemental Composition Com	pound 4	~ ~ -	-			~		
	Sample: 1408	C40Me							
	Note : -luis-v	elasco							
	Inlet : Direct			Io	n Mode	: FAB+			
	RT : 2.89 min			SC	an#: 8				
	Elements : C 4	0/0, H 49/0,	0 9/0						
	Mass Tolerance	: 1000	ppm, 1mm	nu if	m/z > 1				
	Unsaturation (U.S.) : 4.0	- 12.0						
	Observed m/z 267.1022 Estimated m/z	Int% 100.0 Error[ppm]	U.S.	С	н	0			
	267.1021	+0.3	10.5	17	15	3			





Elemental Composition Compound 5

Sample: 1409 Note : -luis-ve	C5Br elasco						- 1	
Inlet : Direct RT : 6.88 min Elements : C 4 Mass Tolerance Unsaturation (0/0, H 49/0, : 1000 U.S.) : 4.0	0 9/0, 1 ppm, 1mm - 12.0	Ion Sca Br 3/0 u if n	1 Mode 1n#: (1) n/z > 1	: FA L9,21 L	B+)		
Observed m/z 315.0017 Estimated m/z 315.0021	Int% 25.3 Error[ppm] -1.1	U.S. 10.5	C 16	Н 12	0 2	Br 1		





¹³C RMN /Compound **7** (CDCl₃) 75MHz



Elemental Composition of Compound 7

Instrument : MStation Sample: 229 L7 Note : -Inlet : Direct Ion Mode : FAB+ RT : 0.90 min Scan#: (5,8) Elements : C 24/0, H 49/0, O 3/0, S 2/0 Mass Tolerance : 1000ppm, 2mmu if m/z > 2 Unsaturation (U.S.) : -0.5 - 20.0 Observed m/z Int% Err[ppm / mmu] U.S. Composition 1 243.0480 100.00 +0.1 / +0.0 10.5 C14 H11 O2 S

¹H RMN /Compound 8 (CDCl₃) 300MHz



¹³C RMN /Compound 8 (CDCl₃) 75MHz



Elemental Composition Compound 8

Sample: 2855 L Py Note : -luis-velasco Ion Mode : FAB+ Inlet : Direct Scan#: (10,11) RT : 3.98 min Elements : C 44/0, H 49/0, O 5/0, N 2/1 Mass Tolerance : 20ppm, 1mmu if m/z > 50Unsaturation (U.S.) : 0.0 - 23.0 Observed m/z Int% 100.0 238.0874 Ν С Η 0 Estimated m/z Error[ppm] U.S. 1 15 12 2 10.5 +2.7 238.0868



¹³C RMN /Compound **9** (CDCl₃) 75MHz



Elemental Composition of Compound 9 Sample Name Dr Alvarez Cecilio

Description History:Determine m/z[Peak Detect[Centroid,30,Area];Correct Base[5.0%]];Correct Base[5.0%];Average(MS[1] 0.6... Created by:AccuTOF Ionization Mode ESI+

Mass Calibration data:Cal_Peg_600 Created 10/27/2014 1:57:46 PM

Unsaturation Number:0.0 .. 14.0 (Fraction:Both)

Tolerance:3.00(mmu) Charge number:1 Element:12C:0 ... 50, 1H:0 ... 120, 56Fe:0 ... 1, 16O:0 ... 3







¹³C RMN /Compound **9** (CDCl₃) 75MHz



Elemental Composition of Compound 10



Elemental Composition	of fragment C ₃₂ H ₂₂	₂ O ₄ Fe [M ^{+.}] C	ompoun	d 1a			
Sample: 2017	LC8NFe						
Note : Luis-Ve	lasco						
Inlet : Direct			Ior	n Mode	: FAB-	+	
RT : 5.30 min			Sca	an#: (1	L7,31)		
Elements : C 40/0, H 49/0, O 10/1, Fe 5/1							
Mass Tolerance	: 1000	ppm, 1mm	u if n	n/z > 1	L		
Unsaturation (U.S.) : -0.5	- 35.0					
Observed m/z	Int%						
526.0877	100.0						
Estimated m/z	Error[ppm]	U.S.	C	H	0	Fe	
526.0867	+1.8	22.5	32	22	4	1	

Elemental Composition of fragment $C_{32}H_{20}O_4CIFe$ [M^{+.}] Compound **2a**

Sample: 2009	FeLC8C	l-ME							
Note : -luis-v	elasco								
Inlet : Direct			Ioi	n Mode	: FAB	+			
RT : 0.75 min			Sca	Scan#: (3,6)					
Elements : C 4	Elements : C 40/0, H 58/0, O 6/1, Cl 3/0, Fe 3/0								
Mass Tolerance	: 1000	ppm, 1m	mu if 1	m/z > 1					
Unsaturation (U.S.) : 13.0	- 25.0							
Observed m/z	Int%								
594.0092	100.0								
Estimated m/z	Error[ppm]	U.S.	С	Н	0	Cl	Fe		
594.0088	+0.6	22.5	32	20	4	2	1		

Elemental Composition of f	ragment C ₃₄ H ₂₆ O ₄ Fe	[M ^{+.}] Compo	und 3a					
Sample: 2010	FeLC8M	le-ME						
Note : -luis-ve	elasco							
Inlet : Direct			Ioi	n Mode	: FAB	+		
RT : 0.43 min			Sca	an#: 3				
Elements : C 4	0/0, H 58/0,	0 6/1,	Fe 3/0	0				
Mass Tolerance	: 1000	ppm, lm	umu if r	n/z > 3	1			
Unsaturation (1	U.S.) : 13.0	- 25.0)	'				
Observed m/z	Int%							
554.1187	100.0							
Estimated m/z	Error [ppm]	U.S.	С	Н	0	Fe		
554.1180	+1.1	22.5	34	2.6	4	1		
					-	-		

Elemental Composition of fragment $C_{34}H_{26}O_6Fe$ [M^{+.}] Compound 4a

Elemental Composition of fragment C₄₀H₃₀O₄Fe₃ [M^{+.}] Compound **9a**

1

Instrument : MStation Sample: 230 L Fc Fe Note : -Inlet : Direct Ion Mode : FAB+ RT: 0.00 min Scan#: (1,7) Elements : C 50/0, H 49/0, O 5/0, Fe 4/0 Mass Tolerance : 1000ppm, 3mmu if m/z > 3 Unsaturation (U.S.) : 25.0 - 30.0 Observed m/z Int% 742.0189 80.16 Fe Estimated m/z Err[ppm / mmu] U.S. С н 0 -0.4 / -0.3 27.5 30 4 1 742.0192 40 3 Elemental Composition of fragment $C_{40}H_{26}O_4Fe$ [M^{+.}] Compound **10a** Instrument : MStation Sample: 227 LNaphFe Note : -Inlet : Direct Ion Mode : FAB+ RT : 0.35 min Scan#: (4,5) Elements : C 50/0, H 49/0, O 5/0, Fe 2/0 Mass Tolerance : 1000ppm, 2mmu if m/z > 2 Unsaturation (U.S.): 22.0 - 35.0 Observed m/z Int% 626 1189 22 51

020.1105	22.01				
Estimated m/z	Err[ppm / mmu] U.S.	С	н	0	Fe
626.1180	+1.4 / +0.9 28.5	40	26	4	1

Elemental Composition of Compound 1b Note : -luis-velasco Inlet : Direct Ion Mode : FAB+ RT : 6.53 min Scan#: (24,35) Elements : C 44/0, H 49/0, O 7/2, Cu 2/0 Mass Tolerance : 20ppm, 5mmu if m/z > 250Unsaturation (U.S.) : 12.0 - 23.0 Observed m/z Int% 534.0883 8.2 Estimated m/z Error [ppm] U.S. С Η Cu 0 534.0892 -1.7 21.5 23 32 4 1