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Dibromination of Alkenes with LiBr and H₂O₂ Under Mild Conditions

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Supporting Information

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Procedures and Characterization Data for Starting Materials



To a three-neck round-bottom flask under argon and at 0 °C (ice bath) NaH (60% dispersion in mineral oil, 0.8 g, 20 mmol) was added to a solution of cinnamyl alcohol (98% purity, 2.74 g, 20 mmol) dissolved in dry THF (20 mL). After ten minutes, benzyl chloride (97% purity, 2.4 mL, 20 mmol) was added and the reaction mixture stirred overnight at room temperature. Then, MeOH (20 mL) was slowly added followed by distilled water (100 mL). The product was extracted with AcOEt (3 x 15 mL) and the combined organic phases were dried under MgSO₄, filtered and solvents removed under reduced pressure. Purification was performed by flash chromatography (hexanes/AcOEt = 98/2). Further purification (to remove remaining benzyl chloride) was accomplished by stirring the product at 60 °C under high vacuum during 3 hours. Product **1n** was obtained as a pale yellow oil (2.13 g, 48% yield);^{[1] 1}H NMR (CDCl₃, 400 MHz) δ = 7.38-7.20 (m, 10H), 6.62 (d, *J* = 15.9 Hz, 1H), 6.35-6.28 (m, 1H), 4.55 (s, 2H), 4.18 (d, *J* = 6.0 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ = 138.6, 137.0, 132.8, 128.8, 128.7, 128.1, 127.95, 127.92, 126.8, 126.4, 72.5, 71.0.



To an one-neck round-bottom flask at 0 °C (ice bath) benzoyl chloride (3.2 mL, 27.5 mmol) was dropwise added to a solution of cinnamyl alcohol (98% purity, 3.2 mL, 25 mmol) dissolved in DCM (80 mL). After one hour of reaction at room temperature, a solution of Et₃N (3.5 mL, 25 mmol) dissolved in DCM (20 mL) was added and the reaction stirred at room temperature overnight. The reaction mixture was washed with water (2 x 50 mL) and then with NaOH 0.2 M (2 x 50 mL). The organic phase was dried under MgSO₄, filtered and the solvent removed under reduced pressure. Purification by flash chromatography (hexanes/AcOEt = 95/5) gave product **1m** as a colorless oil (3.77 g, 63% yield);^{[2] 1}H NMR (CDCl₃, 400 MHz) δ = 8.09-8.07 (m, 2H), 7.56-7.52 (m, 1H), 7.45-7.40 (m, 4H), 7.33-7.23 (m, 3H), 6.73 (d, *J* = 15.9 Hz, 1H), 6.43-6.36 (m, 1H), 4.98

(d, J = 6.4 Hz, 2H); ¹³C NMR (CDCl₃, 100 MHz) $\delta = 166.6$, 136.5, 134.5, 133.2, 129.9, 128.9, 128.6, 128.3, 126.9, 123.5, 65.8.

Characterization of Brominated Products



The general procedure was followed using **1a** (102 μ L, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 μ L, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a pale yellow oil. **Condition A**: 1 hour of reaction, 193 mg, 80% yield; **Condition B**: 2 hours of reaction, 135 mg, 56% yield.^{[3] 1}H NMR (CDCl₃, 400 MHz) δ = 4.46 (br, 2H), 2.49-2.43 (m, 2H), 1.91-1.79 (m, 4H), 1.53-1.50 (m, 2H); ¹³C NMR (CDCl₃, 400 MHz) δ = 55.1, 31.9, 22.3.



The general procedure was followed using **1b** (95% purity, 137 µL, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 µL, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a pale yellow oil. **Condition A**: 1 hour of reaction, 234 mg, 87% yield; **Condition B**: 2 hours of reaction, 144 mg, 53% yield.^{[4] 1}H NMR (CDCl₃, 400 MHz) δ = 4.59-4.58 (m, 2H), 2.44-2.38 (m,

2H), 2.12-2.11 (m, 2H), 2.07-1.88 (m, 2H), 1.70-1.47 (m, 6H); ¹³C NMR (CDCl₃, 400 MHz) δ = 61.8, 33.5, 26.2, 25.7.



The general procedure was followed using **1c** (95% purity, 121 µL, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 µL, 2.0 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a white solid. **Condition A**: 1 hour of reaction, 183 mg, 75% yield; **Condition B**: 2 hours of reaction, 24 mg, 10% yield, m.p. = 70 °C ¹H NMR (CDCl₃, 400 MHz) δ = 1.95 (s). ¹³C NMR (CDCl₃, 100 MHz) δ = 74.1, 32.1; MS (EI, 70 eV): m/z (%) = 163/165 (100/97) [M - Br]⁺, 83 (64) [C₆H₁₁]⁺, 69 (55) [C₅H₉]⁺, 55 (83) [C₄H₇]⁺. Elem. analysis calculated: C = 29.54, H = 4.96, found: C = 29.39, H = 4.63.



The general procedure was followed using **1d** (98% purity, 160 µL, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 µL, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a pale yellow oil. **Condition A**: 1 hour of reaction, 249 mg, 92% yield; **Condition B**: 2 hours of reaction, 143 mg, 53% yield.^[4] ¹H NMR (CDCl₃, 400 MHz) δ = 4.20-4.14 (m, 1H), 3.85 (dd, J^{1} = 10.2 Hz, J^{2} = 4.4 Hz, 1H), 3.63 (t, J = 9.9 Hz, 1H), 2.18-2.09 (m, 1H), 1.83-1.173 (m, 1H), 1.62-1.51 (m, 1H), 1.51-1.30 (m, 7H), 0.91-0.88 (m, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ = 53.1, 36.3, 36.0, 31.6, 28.5, 26.7, 22.5, 14.0.



The general procedure was followed using **1e** (115 μ L, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 μ L, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by

silica gel cromatography (hexanes/AcOEt = 98/2) to afford a white solid. **Condition A**: 1 hour of reaction, 263 mg, >99% yield; **Condition B**: 2 hours of reaction, 161 mg, 61% yield, m.p. = 72–73 °C (literature = 71–73 °C)^[4]. ¹H NMR (CDCl₃, 400 MHz) δ = 7.31-7.22 (m, 5H), 5.04 (dd, J^1 = 16.1 Hz, J^2 = 10.6 Hz, 1H), 3.99-3.89 (m, 2H); ¹³C NMR (CDCl₃, 100 MHz) δ = 138.5, 129.1, 128.8, 127.6, 50.9, 35.0.



The general procedure was followed using **1f** (95% purity, 193 µL, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 µL, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a white solid. **Condition A**: 1 hour of reaction, 320 mg, >99% yield; **Condition B**: 2 hours of reaction, 296 mg, 93% yield, m.p. = 48–49 °C. ¹H NMR (CDCl₃, 400 MHz) δ = 7.40-7.31 (m, 4H), 5.15 (dd, J^1 = 10.2 Hz, J^2 = 5.8 Hz, 1H), 4.09-4.00 (m, 1H), 1.32 (s, 9H). ¹³C NMR (CDCl₃, 100 MHz) δ = 152.5, 135.8, 127.5, 126.1, 51.5, 35.4, 35.0, 31.5; HRMS (APCI, positive mode) m/z calculated for C₁₂H₁₆⁷⁹Br [M - Br]⁺ 239.0430, found: 239.0434.



The general procedure was followed using **1g** (97% purity, 132 µL, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 µL, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a white wax. **Condition A**: 1 hour of reaction, 228 mg, 77% yield; **Condition B**: 2 hours of reaction, 151 mg, 51% yield.^{[5] 1}H NMR (CDCI₃, 400 MHz) δ = 7.35-7.30 (m, 4H), 5.09 (dd, J^1 = 11.0 Hz, J^2 = 5.1 Hz, 1H), 4.04 (dd, J^1 = 10.3 Hz, J^2 = 5.1 Hz, 1H), 3.94 (t, J = 10.7Hz, 1H); ¹³C NMR (CDCI₃, 100 MHz) δ = 137.4, 135.2, 129.4, 129.3, 49.9, 35.0.



The general procedure was followed using **1h** (98% purity, 157 µL, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 µL, 1.2 mmol). At the end of the reaction the product was extracted with AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 98/2) to afford a pale yellow oil. **Condition A**: 2 hours of reaction, 297 mg, 96% yield; **Condition B**: 2 hours of reaction, 244 mg, 79% yield.^[4] ¹H NMR (CDCl₃, 400 MHz) δ = 7.19 (d, *J* = 8.3 Hz, 2H), 6.86 (d, *J* = 8.3 Hz, 2H), 4.35-4.28 (m, 1H), 3.79 (s, 3H), 3.60 (t, *J* = 9.3 Hz, 1H), 3.40 (dd, *J*¹ = 14.7 Hz, *J*² = 4.7 Hz, 1H), 3.09 (dd, *J*¹ = 14.6 Hz, *J*² = 7.5 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ = 158.7, 130.5, 128.7, 113.9, 55.2, 52.9, 41.0, 35.9.



The general procedure was followed using **1i** (98% purity, 137 mg, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 μ L, 2.0 mmol). At the end of the reaction the product extracted with AcOEt and purified by silica gel cromatography (hexanes/AcOEt = 90/10) affording a white solid. **Condition A**: 6 hours of reaction, 239 mg, 81% yield; **Condition B**: 6 hours of reaction, 253 mg, 86% yield, m.p. = 69 °C (literature = 69 °C).^{[3] 1}H NMR (CDCl₃, 400 MHz) δ = 7.38-7.34 (m, 5H), 5.27 (d, *J* = 11.0 Hz, 1H), 4.71-4.67 (m, 1H), 4.34-4.22 (m, 2H), 2.23 (br, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ = 140.1, 129.2, 129.0, 128.1, 66.1, 59.5, 52.5.



The general procedure was followed using **1j** (139 μ L, 1.0 mmol) as substrate, LiBr (208 mg, 2.4 mmol) and H₂O₂ 8.8 M (136 μ L, 1.2 mmol). At the end of the reaction the product was extracted with a 9/1 solution of hexanes/AcOEt (3 x 10 mL) and purified by

silica gel cromatography (hexanes/AcOEt = 98/2) to afford a white solid. **Condition A**: 6 hours of reaction, 247 mg, 79% yield; **Condition B**: 6 hours of reaction, 275 mg, 88% yield, m.p. = 102-103 °C. The product was obtained as a 17/1 mixture of diastereoisomers (assessed by the examination of the ¹H NMR spectrum) ¹H NMR (CDCl₃, 400 MHz) δ = 7.42-7.35 (m, 5H), 5.30 (d, *J* = 9.9 Hz, 1H), 4.79-4.75 (m, 1H), 4.38-4.34 (m, 1H), 4.09-4.06 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ = 139.0, 129.4, 129.0, 128.4, 55.1, 53.2, 49.5. HRMS (APCI, positive mode) *m/z* calculated for C₉H₉⁻⁷⁹BrCl [M - Br]⁺ 230.9571, found: 230.9572.



The general procedure was followed using **1k** (210 mg, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 μ L, 2.0 mmol). At the end of the reaction the product was extracted with AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 95/5) to afford a white solid. **Condition A**: 6 hours of reaction, 300 mg, 78% yield; **Condition B**: 6 hours of reaction, 265 mg, 69% yield, m.p. = 64-65 °C. ¹H NMR (CDCl₃, 400 MHz) δ = 7.40-7.33 (m, 10H), 5.36 (d, *J* = 10.1 Hz, 1H), 4.67-4.63 (m, 3H), 4.21-4.17 (m, 1H), 4.00-3.97 (m, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ = 139.9, 137.9, 129.1, 128.8, 128.7, 128.3, 128.14, 128.08, 73.8, 72.7, 55.0, 53.0; HRMS (APCI, positive mode) *m*/*z* calculated for C₁₆H₁₆⁷⁹BrO [M - Br]⁺ 303.0379, found: 303.0384.



The general procedure was followed using **1I** (238 mg, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 μ L, 2.0 mmol). At the end of the reaction the product was extracted with AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 95/5) to afford a white solid. **Condition A**: 6 hours of reaction, 225 mg, 57% yield; **Condition B**: 6 hours of reaction, 311 mg, 78% yield, m.p. = 111-112 °C.¹H NMR (CDCl₃, 400 MHz) δ = 8.12 (d, *J* = 7.6 Hz, 2H), 7.63-7.59 (m, 1H), 7.51-7.34 (m, 7H), 5.28 (d, *J* = 10.3 Hz, 1H), 5.04-5.00 (m, 1H), 4.97-4.96 (m, 1H), 4.85-4.82 (m,

1H); ¹³C NMR (CDCI₃, 100 MHz) δ = 166.1, 139.6, 133.6, 130.0, 129.8, 129.3, 129.0 128.8, 128.2, 67.5, 52.9, 52.8. HRMS (APCI, positive mode) *m/z* calculated for C₁₆H₁₄⁷⁹BrO₂ [M - Br]⁺ 317.0172, found: 317.0173.



The general procedure was followed using **1m** (162 mg, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 μ L, 2.0 mmol). At the end of the reaction the product was extracted with AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 90/10) to afford a white solid. **Condition A**: 48 hours of reaction, 269 mg, 84% yield; **Condition B**: 48 hours of reaction, 179 mg, 56% yield (NMR yield), m.p. = 112-113 °C (literature = 115-116 °C °C).^{[6] 1}H NMR (CDCl₃, 400 MHz) δ = 7.39-7.37 (m, 5H), 5.34 (d, *J* = 11.7 Hz, 1H), 4.84 (d, *J* = 11.7 Hz, 1H), 3.90 (s, 3H); ¹³C NMR (CDCl₃, 100 MHz) δ = 168.6, 137.8, 129.6, 129.1, 128.3, 53.6, 50.9, 46.9.



The general procedure was followed using **1n** (215 mg, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 μ L, 2.0 mmol). At the end of the reaction the product was extracted with AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 90/10) to afford a white solid. **Condition A**: 48 hours of reaction, 306 mg, 83% yield; **Condition B**: 48 hours of reaction, 147 mg, 40% yield, m.p. = 156-157 °C (literature = 157 °C).^[3] ¹H NMR (CDCl₃, 400 MHz) δ = 8.10 (d, *J* = 7.3 Hz, 2H), 7.68-7.64 (m, 1H), 7.57-7.52 (m, 4H), 7.44-7.38 (m, 3H), 5.83 (d, *J* = 11.3 Hz, 1H), 5.65 (d, *J* = 11.3 Hz, 1H); ¹³C NMR (CDCl₃, 100 MHz) δ = 191.4, 138.5, 134.7, 134.4, 129.5, 129.2, 129.13, 129.09, 128.6, 50.0, 47.1.



The general procedure was followed using **1o** (313 mg, 1.0 mmol) as substrate, LiBr (350 mg, 4.0 mmol) and H₂O₂ 8.8 M (227 μ L, 2.0 mmol). At the end of the reaction the product was extracted with AcOEt (3 x 10 mL) and purified by silica gel cromatography (hexanes/AcOEt = 90/10) to afford a white solid. **Condition A**: 48 hours of reaction, 325 mg, 69% yield; **Condition B**: 48 hours of reaction, 322 mg, 68% yield, m.p. = 123-124 °C. Recrystallization was performed with DCM and hexanes. ¹H NMR (CDCI₃, 400 MHz) δ = 7.40 (br, 3H), 7.27-7.20 (m, 12H), 5.61 (d, *J* = 11.3 Hz, 1H), 5.06 (d, *J* = 14.2 Hz, 1H), 4.91 (d, *J* = 14.2 Hz, 1H), 4.71 (d, *J* = 11.4 Hz, 1H); ¹³C NMR (CDCI₃, 100 MHz) δ = 167.3, 140.9, 138.4, 136.8, 130.1, 129.3, 129.2, 128.9, 128.7, 128.4, 127.9, 54.1, 52.4, 45.8. HRMS (ESI, positive mode) *m/z* calculated for C₂₂H₁₉⁷⁹Br₂NNaO [M + Na]⁺ 493.9726, found: 493.9710.

Crystallographic Data and Structure Refinement for Compound 20:



<i>a</i> = 9.8099 (5) Å	Cell parameters from 20953 reflections
<i>b</i> = 14.0372 (7) Å	q = 2.5–26.1°
<i>c</i> = 14.7788 (11) Å	m = 3.99 mm ⁻¹
b = 90.878 (6)°	<i>T</i> = 298 K
V = 2034.9 (2) Å ³	Prism, colorless
<i>Z</i> = 4	0.53 × 0.30 × 0.30 mm

Data collection

Xcalibur, Atlas, Gemini ultra diffractometer	5499 independent reflections
Radiation source: fine-focus sealed X-ray tube, Enhance (Mo) X-ray Source	3667 reflections with $l > 2s(l)$
Graphite monochromator	$R_{\rm int} = 0.042$
Detector resolution: 10.4186 pixels mm ⁻¹	$q_{max} = 29.6^{\circ}, q_{min} = 2.5^{\circ}$
w scans	<i>h</i> = -13→13
Absorption correction: multi-scan <i>CrysAlis</i> <i>PRO</i> 1.171.38.43f (Rigaku Oxford Diffraction, 2015) Empirical absorption correction using spherical harmonics, implemented in SCALE3 ABSPACK scaling algorithm.	<i>k</i> = -19→19
$T_{\min} = 0.597, \ T_{\max} = 1.000$	/=-20→20
70535 measured reflections	
Refinement	
Refinement on F^2	0 restraints
Least-squares matrix: full	Hydrogen site location: inferred from neighbouring sites
$R[F^2 > 2s(F^2)] = 0.043$	H-atom parameters constrained
$wR(F^2) = 0.106$	$w = 1/[s^2(F_o^2) + (0.0351P)^2 + 2.1756P]$ where $P = (F_o^2 + 2F_c^2)/3$
S = 1.02	$(D/s)_{max} = 0.001$
5499 reflections	$D\rho_{max} = 0.83 \text{ e} \text{ Å}^{-3}$
235 parameters	$D\rho_{min} = -0.90 e Å^{-3}$

References

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2b										Curre NAME EXPNO PROCN	ent Da [.]) 10	ta Parameters I-NSM-151 5 1
										F2 - Date_ Time INST PROBH PULPH TD SOLVE NS DS SWH FIDRI AQ RG RG DW DE TE D1 D11 TD0	Acquis - RUM ID 5 ROG :NT ES	sition Parameter 20170626 8.45 spect mm DUL 13C-1 zgpg30 32768 CDC13 82 4 25252.525 Hz 0.770646 Hz 0.6488064 sec 197.86 19.800 usec 10.00 usec 10.00 usec 300.6 K 2.0000000 sec 0.03000000 sec 1
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										SFO2 NUC2 CPDP1 PCPD2 PLW2 PLW12 PLW12	=== CH. RG[2 2 3	ANNEL f2 400.1116004 MHz 1H waltz16 90.00 use(11.19999981 W 0.19090000 W 0.15463001 W
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Current NAME EXPNO PROCNO	Data Parameters I-NSM-121 1 1
F2 - Acq Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH FIDRES AQ RG DW DE TE D1 TD0	uisition Parameter 20170510 8.26 spect 5 mm DUL 13C-1 zg30 65536 CDC13 20 0 8012.820 Hz 0.122266 Hz 4.0894465 sec 63.68 62.400 usec 10.00 usec 299.7 K 1.0000000 sec 1
SF01 NUC1 P1 PLW1	CHANNEL f1 400.1124007 MHz 1H 11.75 usec 11.19999981 W
F2 - Pro SI SF WDW SSB LB GB PC	0 0 0 0 0 0 0 0 0 0 0 0 0 0

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10 9 8 7 6 5 4 3 2 1 ppm

Br Br 2c						74.1				Current NAME EXPNO PROCNO	Data Parameters I-NSM-121 2 1
										F2 - Acc Date_ Time INSTRUM PROBHD PULPROG TD SOLVENT NS DS SWH FIDRES AQ DW DE TE D1 D11 TD0	<pre>guisition Parameter 20170510 8.20 5 mm DUL 13C-1 zgpg30 32768 CDC13 100 4 24038.461 Hz 0.733596 Hz 0.6815744 sec 197.86 20.800 usec 10.00 usec 300.6 K 2.0000000 sec 0.03000000 sec 1</pre>
										SFO1 NUC1 P1 PLW1	CHANNEL f1 100.6177998 MH2 13C 13.86 usec 17.98900032 W
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200	180	160	140	120	100	80	60	40	20	LB GB PC	0 1.00 Hz









Raw Spectrum 3.317 (scan : 399) Base Peak m/z 163.00 (Inten : 834,743)

m/z	Relative Intensity (%)
53.05	13.36
55.05	82.54
67.05	15.19
69.05	55.54
83.05	64.17
84.05	29.65
163.00	100
165.00	96.55















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Ph $\stackrel{\text{Br}}{{}_{{}}}_{\text{Br}}$	^o∕Ph 2k		133.9 137.9 128.8 128.7 128.7	128.1		72.7				Current D NAME EXPNO PROCNO	Ata Parameters I-NSM-149 2 1
										F2 - Acqui Date	Lsition Parameter 20170914 9.33 spect 5 mm DUL 13C-1 zgpg30 32768 CDC13 500 4 25252.525 Hz 0.770646 Hz 0.6488064 sec 197.86 19.800 usec 10.00 usec 300.2 K 2.00000000 sec 0.03000000 sec 1
										===== C: SF01 NUC1 P1 PLW1	HANNEL f1 100.6176997 MHz 13C 13.86 usec 17.98900032 W
										SF02 NUC2 CPDPRG[2 PCPD2 PLW2 PLW12 PLW13	HANNEL f2 400.1116004 MH2 1H waltz16 90.00 usec 11.19999981 W 0.19090000 W 0.15463001 W
in the	alan yan dalama kan sa	haagaya may haanaa ahaa ahaa ahaa ahaa ahaa ahaa	hannin hannan hann	uulan marinadan y	landa ta kina palanting bary Palating ang palating bary	d water and the second second	and a traditional	and the latest second		F2 - Proce SI SF WDW SSB	essing parameters 32768 100.6077179 MHz EM
Т	180	160	140	120 1	00 80	0 6	0 4	0 2	20 pr	LB TGB 0 PC	1.00 Hz 1.40

Display Report

Analysis Info

Analysis Name Method Sample Name Comment

C:\Data\LabSelen 13.09.17 APCI\I-NSM-149000004.d tune_low_APCI.m I-NSM-149

13/9/2017 11:20:50 Acquisition Date

Operator Instrument Bruker

micrOTOF-Q 10243



Bruker Compass DataAnalysis 4.0 printed:

13/9/2017 11:23:35

Display Report Analysis Info Acquisition Date 13/9/2017 11:20:50 Analysis Name C:\Data\LabSelen 13.09.17 APCI\I-NSM-149000004.d Method tune_low_APCI.m Operator Bruker Sample Name I-NSM-149 Instrument micrOTOF-Q 10243 Comment **Acquisition Parameter** 2.5 Bar 250 °C 1.5 l/min Ion Polarity Source Type APCI Positive Set Nebulizer Set Capillary Set End Plate Offset 2000 V -500 V Set Dry Heater Set Dry Gas Focus Not active Scan Begin 50 m/z Scan End 1000 m/z Set Collision Cell RF 300.0 Vpp Set Divert Valve Source Intens. x10⁵ +MS, 0.2min #(10) 196.9958 3.0 compound 2k 2.5 680.4726 2.0 338.3412 1.5 303.0384 241.1254 1.0 0.5 408.2139 460.1271 552.0510 737.9020 0.0 200 300 400 500 600 700 m/z

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Analysis Info

Analysis Name Method Sample Name Comment

lame C:\Data\LabSelen 13.09.17 APCI\I-NSM-137000003.d tune_low_APCI.m ame I-NSM-137

Acquisition Date 13/9/2017 11:04:44

Operator Instrument Bruker

micrOTOF-Q 10243



Bruker Compass DataAnalysis 4.0 printed: 13/9/2017 11:07:04 Page 1 of 1



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Analysis Info

Analysis Name Method Sample Name Comment

ame C:\Data\LabSelen 01.08.17\I-NSM-142000001.d tune_low_POS_internalcalibration INJEÇÃO.m me I-NSM-142

Acquisition Date 1/8/2017 09:35:32

Operator Instrument Bruker

micrOTOF-Q 10243



Display Report

Analysis Info

Analysis Name Method Sample Name Comment

C:\Data\LabSelen 01.08.17\I-NSM-142000001.d
tune_low_POS_internalcalibration INJEÇÃO.m
I-NSM-142

Acquisition Date 1/8/2017 09:35:32

Operator Instrument Bruker

micrOTOF-Q 10243



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