

1 **[Supplementary information]**

2 **Aptabody-Aptatope Interactions in Aptablotting Assays**

3 Running head: Aptabody-Aptatope Interactions

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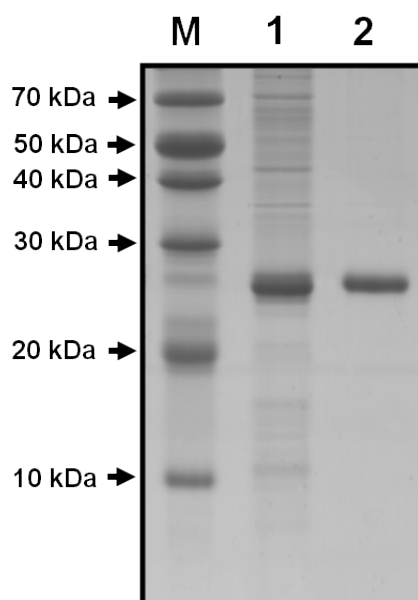
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Figures

Supplementary Figure S1

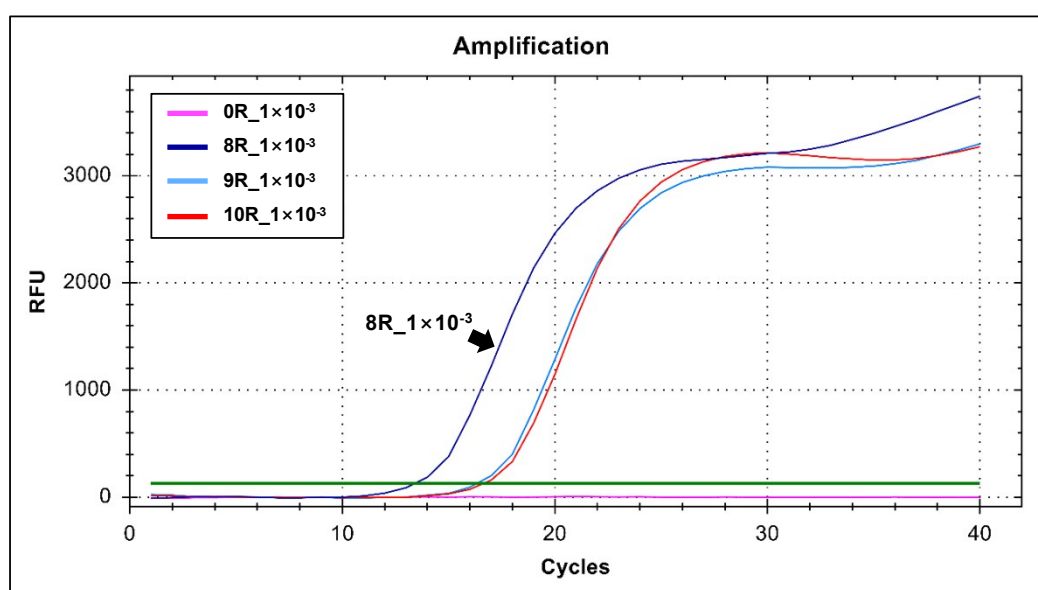


GST overproduction and purification

GST overexpression was successfully accomplished in *E. coli* with the pGEX4T-1 system.

The GST of *E. coli* was stably overexpressed as a soluble protein in *E. coli* BL21 (DE3). The GST protein was purified using glutathione-agarose beads, and the purified GST showed a single protein band with an apparent M_r of approximately 26000 on the SDS-PAGE stained with Coomassie blue. Lane 1, whole-cell lysates of pGETX4T-1 (10 μ g of protein); lane 2, purified GST (1 μ g of protein); M, molecular size markers.

Supplementary Figure S2

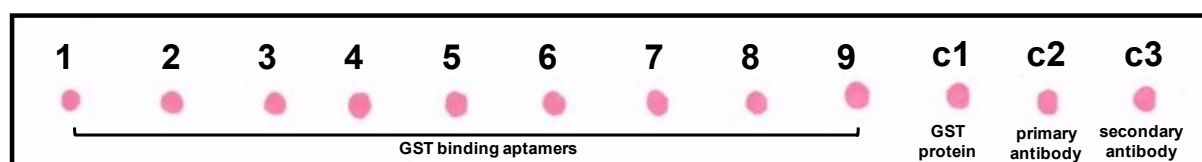


Round	0 Round	8 Round	9 Round	10 Round
Dilution rate	1×10 ⁻³	1×10 ⁻³	1×10 ⁻³	1×10 ⁻³
Ct value	-	13.33	16.49	16.97
Efficiency	90.38	83.90	80.06	74.39

Real-time PCR and round selection

We employed the real-time PCR (RT-PCR) analysis that provided a valuable tool for validating the optimal round of ssDNA aptamer enrichment. The eluted ssDNA bound from 0, 8th, 9th, and 10th rounds were normalized with the same concentration and reintroduced to GST. The recovered DNA samples from 0, 8th, 9th, and 10th round were amplified using an optimized RT-PCR cycle and the fluorescence signal was monitored by MiniOpticon™ Real-time PCR fluorescence signal detection system (Bio-Rad, USA). PCR experiment was independently triplicated and data was analyzed to obtain the average C(t) values. The samples from 8th round SELEX showed a lower C(t), C(t)=13.33, as compared to other samples, C(t)=15.20 for 9th round and 16.97 for 10th round, indicating the presence of higher amounts of DNA aptamer for GST.

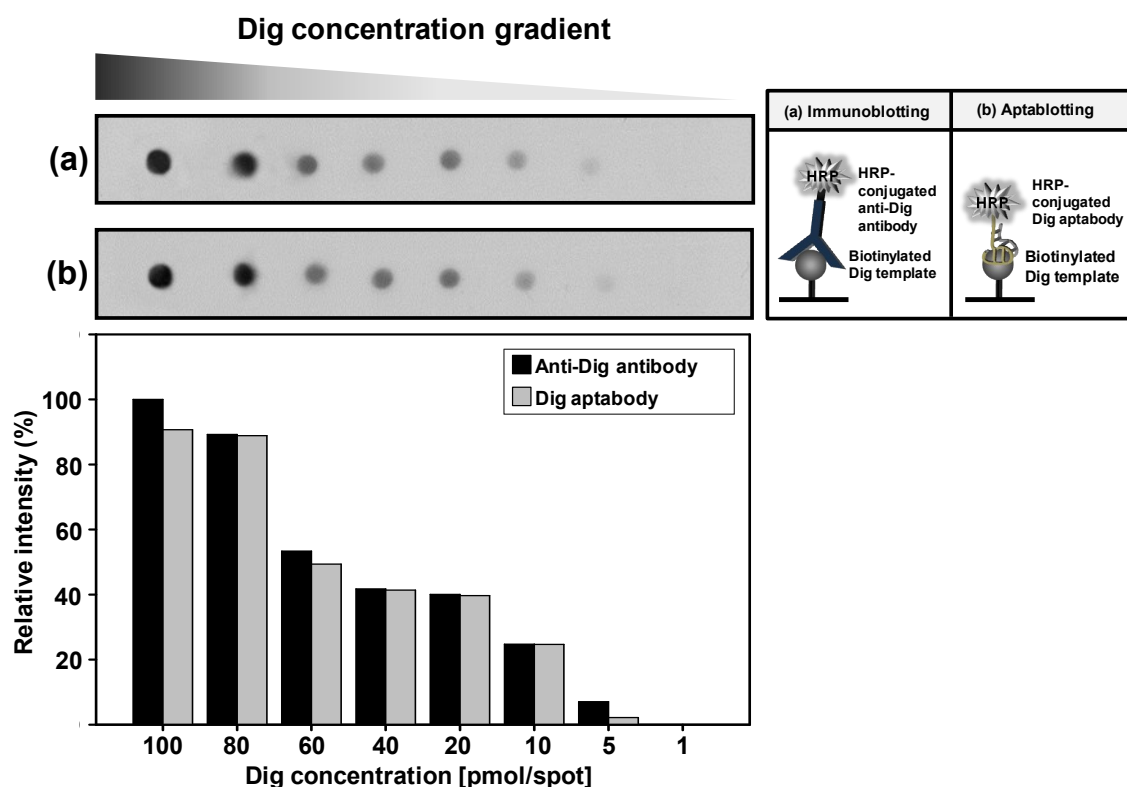
Supplementary Figure S3



Dot-blotting analysis

Serial dilutions of purified GST (1 μ g to 1 ng) were spotted on the methanol-pretreated PVDF membranes. Blotted proteins are stained with Ponceau S solution (Sigma Aldrich, P7170).

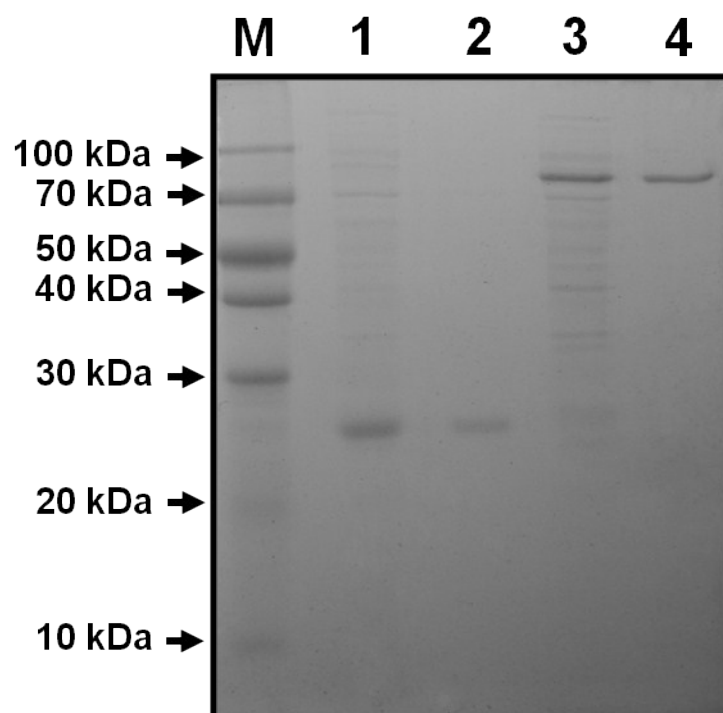
Supplementary Figure S4



DIG aptabody test

The synthesized DIG template (1 μ l/spot) was loaded onto the PVDF membrane in concentrations of 100 pmol, 80 pmol, 60 pmol, 40 pmol, 20 pmol, 10 pmol, 5 pmol, and 1 pmol. The membrane was then dried at 37 $^{\circ}$ C for 1 hr and immersed in 5% nonfat dry milk with PBST blocking buffer for 1 hr to block the non-specific binding sites. After washing three times for 5 min with a PBST washing buffer, the membranes were incubated with 1 μ g/ml diluted DIG-1-HRP and 1:5000 diluted anti-mouse DIG-HRP-conjugated antibodies (Abcam, UK). The binding sensitivity of the DIG aptabody and anti-DIG antibody was assessed via a comparison with the anti-DIG-HRP tests (a) immunoblotting and (b) aptablotting, respectively

Supplementary Figure S5



GST and GST-fused protein overexpression and purification

The overexpression of GST and GST-fused PNBE proteins was successfully accomplished in *E. coli* with the pGEX4T-1 system. The GST and GST-fused PNBE protein of *E. coli* has been stably overexpressed as a soluble protein in *E. coli* BL21(DE3). The recombinants were purified with glutathione-agarose beads, and the purified GST and GST-fused PNBE showed a single protein band with an apparent M_r of approximately 26000 and 75000 via the SDS-PAGE analysis on a 12% gel stained with Coomassie Brilliant Blue. In the panels, lane 1 contains pGEX4T-1 whole-cell lysate (5 ug), lane 2 contains purified GST (0.5 ug), lane 3 contains pGEX4T-1::PNBE cell lysate (5 ug) and lane 4 contains purified GST-tagged PNBE protein (0.5 ug). The molecular size markers are shown in the lane labeled M.

Table S1 Sequence and structure analysis of the GST aptabodies and DIG aptabodies used in this study.

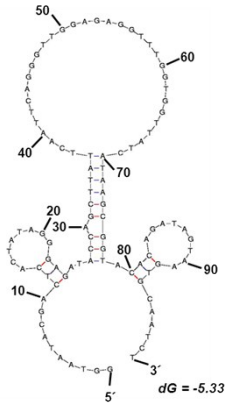
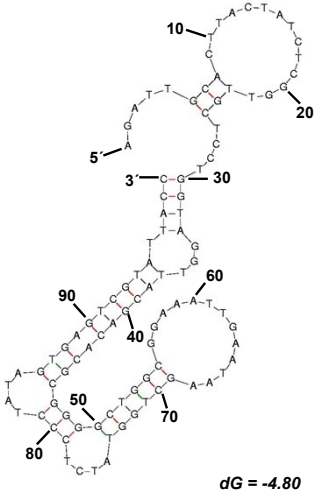
Clone	Selected sequences	K_D (nM)	Structures
GST-2	CAGGGTTGGAGAGGTTTGGT GGTTATCATAAGCGGTACAC	0.002 ± 0.003	
DIG-1	CGGTTGCTCCTGGTAGGTTA CGACACGCGGGGCTGGCGGA	0.007 ± 0.001	

Table S2 List of selected aptamers and target binding affinities of each aptamer as determined from the direct coupling SPR assay.

Clone	Selected sequences	Sequence size (bp)	Identical sequences	K_D (nM)
GST-1	ACCGCTGCAATCGTGTGCCATACACAGATGATCCGCACA	39	3	0.019 ± 0.028
GST-2	CAGGGTTGGAGAGGTTTGGTGGTTATCATAAGCGGTACAC	41	2	0.002 ± 0.003
GST-3	TAGGTTTGGGTAGGTTGGAATGGTGAATAATATGGGACGG	40	6	0.015 ± 0.016
GST-4	CGGACGTTGCACTGGAAAAGGAGGTTTGGTTCGGTTGGTC	40	2	0.119 ± 0.151
GST-5	AGACAGCATAGCACTGTAACGATTGGTTTGGTGTGGTTGG	40	8	2.280 ± 1.250
GST-6	AGATATCAGGAGCGGGGTTTGGGGTGGTTGGCGGTGGACG	40	2	95.110 ± 15.35
GST-7	GCTCTAAGTATACCGGTAGGGTTGGCTGCGGTTTGGCTCG	40	3	0.014 ± 0.011
GST-8	ACTATCGGATTTCTGGGTTTGGATCGGTTGGCAGCGAGTG	40	2	0.340 ± 0.121
GST-9	GCTCTAAGTATTCCGGTAGGGTTGGCTGCGGTTTGGCTCA	40	2	8.37 ± 1.250
DIG-1	CGGTTGCTCCTGGTAGGTTACGACACGCGGGGCTGGCGGA	39	8	0.007 ± 0.001
DIG-2	ACCGTAACAGATAAAAAGATTCTGTGATTGACTGCTTCTGG	41	2	0.742 ± 0.024
DIG-3	CGCCGGCTAGGAAACAATTGGTTGGTGTGTTTCGGCCCCCTT	40	3	0.030 ± 0.001
DIG-4	CAGTGAAACTTTGGTTGGCCGCCTGTAGGGTCACTTTGTG	40	1	9.513 ± 0.413

DIG-5	CCCCGCCAAGGTTTTTGCAGTTGGTCCCTCCGGTGATTCTG	40	2	2.491 ± 0.439
DIG-6	CGCATTCGACATAGCCGGTAGTAAAAGCATGGTCGCTTCA	40	2	1.751 ± 0.112
DIG-7	CCAAAGTGGGATTTTTTAGTGGTTGCATCGCGAGACAGCT	40	2	1.035 ± 0.457
DIG-8	CGAGGTAGCAGGTTGCTTGCGGCTAAAGCATTTTCATGCT	40	2	1.442 ± 0.318
DIG-9	CACAAAGGAATGAACGATGCGATCTGGTAGCTTCTTTGGC	40	3	0.011 ± 0.017
DIG-10	CGTGGAATGTTGTTGCTTCTGCTACGATACGTCAACCCTC	40	1	1.775 ± 0.571
DIG-11	AGGTGTAGATACCCAGGCGTATGGTTGCGCTGTTTCGTGG	40	2	4.001 ± 0.672
DIG-12	CAAGGTGTCGCACTTTTTGACGTTGGACTGCTTCGAACAG	40	2	2.184 ± 0.753
