Electronic supplementary information

Efficient intracellular delivery of biomacromolecules employing clusters of zinc oxide nanowires[†]

Prashant Sharma,^{‡a,b} Hyun Ah Cho,^{‡c} Jae-Won Lee,^{a,b} Woo Seung Ham,^c Bum Chul Park,^c Nam-Hyuk Cho^{a,b,d,*} and Young Keun Kim^{c,*}

^aDepartment of Microbiology and Immunology, ^bDepartment of Biomedical Sciences, Seoul National University College of Medicine, Seoul 03080, Republic of Korea

^cDepartment of Materials Science and Engineering, Korea University, Seoul 02841, Republic of Korea

^dInstitute of Endemic Disease, Seoul National University Medical Research Center and Bundang Hospital, Seoul 03080, Republic of Korea

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Fig. S1 – S3.



Fig. S1. XRD patterns of vertical ZnO nanowire (VNW) and fan-shaped ZnO nanowire (FNW) arrays. The characteristic peaks of wurtzite ZnO (JCPDS No. 36-1451) are denoted in the bottom part of the graph.



Fig. S2. Coating of ZnO nanowire arrays with ZBP-FITC. (**A**) Vertical (VNW) or fan-shaped (FNW) nanowires were incubated with indicated concentration of ZnO-binding peptides (ZBP) conjugate with FITC for 1 h at 37°C. ZnO nanowire arrays coated with ZBP-FITC were assessed the relative binding of ZBP-FITC by measuring the fluorescence intensity. (**B**) Representative images of ZnO nanowire arrays coated with ZBP-FITC. DIC, differential interference contrast. White bar, 10 μm.



Fig. S3. Complex formation of ZBP-streptavidin and biotin-labeled DNA. (**A**) Schematic diagram of complex formation strategy for ZBP (purple)-streptavidin (green) and biotin (red)-labeled DNA (oragnge) on ZnO nanowire (blue) array. (**B**) Kinetic binding of ZBP-streptavidin to ZnO nanowire array on a coverslip. Clustered ZnO NW array on a coverslip were incubated with 300 μg purified ZBP-streptavidin at 37°C for indicated time interval and the bound proteins were assessed by BCA protein assay kit (see Methods). (**C** and **D**) The electrophoretic mobility shift assay was performed to confirm the complex formation of ZBP-streptavidin and biotinylated DNA. Indicated amount of ZBP-spreptavidin protein were incubated with biotinylated DNA encoding GFP expression cassette and the resulting complexes were

visualized under UV light after agarose gel electrophoresis (C). Relative binding of biotinylated DNA to ZBP-streptavidin were presented from triplicate experiments. Error bar, \pm SD.