

1 **Supporting Information**

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3 **Translocation, distribution and degradation of prochloraz-loaded**
4 **mesoporous silica nanoparticles in cucumber plant**

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20 **Preparation of suspension concentrates**

21 In order to compare the distribution and degradation of p-MSNs and prochloraz,
22 conventional prochloraz SC and p-MSNs SC were prepared by bead milling process,
23 respectively. In the proposed SC, Morwet D425 was used as dispersant. As the active
24 ingredient, prochloraz accounted for 20 % in both p-MSNs and conventional SC. The
25 particle size and distribution of conventional SC were carried out by BT-9300ST laser
26 particle size distribution analyzer (Bettersize Instruments Ltd., Liaoning, China). The
27 D_{50} and D_{90} of the particle size was 1.397 and 3.771 μm for conventional SC,
28 respectively.

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30 **Sample preparation and analytical method**

31 A modified QuEChERS method was carried out to measure the concentration
32 levels of prochloraz and its metabolite 2,4,6-TCP in cucumber plants. The procedure
33 involved miniaturized extraction of 2.0 g homogenized sample with acetonitrile,
34 followed by liquid–liquid partition by adding 3.0 g of sodium chloride. After that, the
35 processes of cleanup and residual water removal were carried out by mixing 1 mL of
36 acetonitrile extract with some loose sorbents. For prochloraz analysis, 150 mg of
37 anhydrous magnesium sulfate and 25 mg of PSA were used as the sorbent; for
38 2,4,6-TCP analysis, 150 mg of anhydrous magnesium sulfate and 5 mg of MWCNTs
39 were used. The samples of treated leaves were diluted 20-fold before injection.
40 HPLC-MS/MS was then operated for confirmatory and quantitative analysis of
41 prochloraz and 2,4,6-TCP.

42 Prochloraz and 2,4,6-TCP determinations were performed on an Agilent 1200
43 HPLC equipped with a reversed-phase column (ZORBAX SB-C18, 3.5 μm , 2.1 mm
44 \times 50 mm, Agilent, USA) at 30 $^{\circ}\text{C}$. The mobile phase was acetonitrile/water (80/20,
45 v/v) with a flow rate of 0.3 mL/min. The injection volume was 5 μL . An Agilent 6410
46 Triple Quad LC/MS system with ESI source was conducted for the mass
47 spectrometric analysis. Nitrogen was introduced as the nebulizer and collision gas.
48 The parameters of operation were as followed: gas temperature, 350 $^{\circ}\text{C}$; gas flow, 8
49 L/min; nebulizer gas, 35 psi; capillary voltage, 4,000 V. The multiple reaction
50 monitoring mode was used to monitor the precursor-product ion transitions. Agilent
51 Mass Hunter Data Acquisition and Qualitative Analysis and Quantitative Analysis
52 software was employed for method development and data acquisition. Table S1
53 showed the multiple reaction monitoring transitions and other HPLC–MS/MS
54 parameters for prochloraz and 2,4,6-TCP analysis.

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56 **Analytical method validation**

57 The matrix-matched calibration solutions were obtained from calibration solutions
58 in extracts of blank samples at five different concentrations for prochloraz and
59 2,4,6-TCP in the range of 0.005 ~ 1 mg/L. To evaluate the precision and accuracy of
60 the sample preparation method, prochloraz and 2,4,6-TCP were spiked at four
61 fortified levels of 0.005, 0.01, 0.1 and 1 mg/kg in blank leaf, root and cucumber
62 samples, respectively. Each spiked level test was repeated five times to verify the
63 repeatability of the proposed method. Recovery rate was the amount determined as a

64 percentage of the amount of prochloraz or 2,4,6-TCP originally spiked into the blank
65 samples. The repeatability of the proposed method was shown as a relative standard
66 deviation (RSD, %, n=5). Limits of quantification (LOQs) were determined as the
67 concentration of the compound giving a signal to noise ratio (S/N) of 10 for the target
68 ion, which was calculated by Agilent Mass Hunter Qualitative Analysis software.

Table S1. MRM transitions and other HPLC–MS/MS parameters for prochloraz and 2,4,6-TCP analysis

Compound	Retention time (min)	Fragmentor (V)	Quantifying ions	Qualifying ions	Collision energy (V)	Ionization mode
prochloraz	1.28	80	376.2/308.2	376.2/266.2	10;10	positive
2,4,6-TCP	1.38	80	195.0/158.8	195.0/93.2	40;10	negative

Table S2. Average recoveries at fortified levels of 0.005, 0.01, 0.1 and 1 mg/kg, RSDs (n=5), LOQs, linear equation, determination coefficients (R^2) of prochloraz and its metabolite 2,4,6-TCP in leaf, root and cucumber.

Compound	Sample	Linear equation	R^2	LOQ (mg/kg)	Average recovery % (RSD %)			
					0.005 mg/kg	0.01 mg/kg	0.1 mg/kg	1 mg/kg
prochloraz	Leaf	$y = 2907302 x - 8257$	0.9997	0.002	71 (6)	80 (5)	91 (4)	92 (6)
	Root	$y = 1966195 x - 1473$	0.9999	0.001	78 (9)	89 (4)	87 (3)	81 (5)
	Cucumber	$y = 2051555 x - 83$	0.9991	0.001	77 (10)	92 (6)	78 (7)	82 (7)
2,4,6-TCP	Leaf	$y = 31071x + 97$	0.9998	0.005	81 (8)	90 (6)	74 (8)	73 (6)
	Root	$y = 43113x - 199$	0.9981	0.002	79 (7)	83 (7)	77 (10)	82 (5)
	Cucumber	$y = 26186 x + 718$	0.9997	0.002	73 (9)	81 (9)	83 (6)	89 (4)