

Supporting Information

**Broad Scope Gold(I)-Catalysed Polyenyne Cyclisations for the Formation of up
to Four Carbon-Carbon Bonds**

Zhouting Rong and Antonio M. Echavarren*

*Institute of Chemical Research of Catalonia (ICIQ), Barcelona Institute of Science
and Technology, Av. Països Catalans 16, 43007 Tarragona (Spain)*

*Departament de Química Orgànica i Analítica, Universitat Rovira y Virgili, C/
Marcel·lí Domingo s/n, 43007 Tarragona (Spain).*

Table of Contents

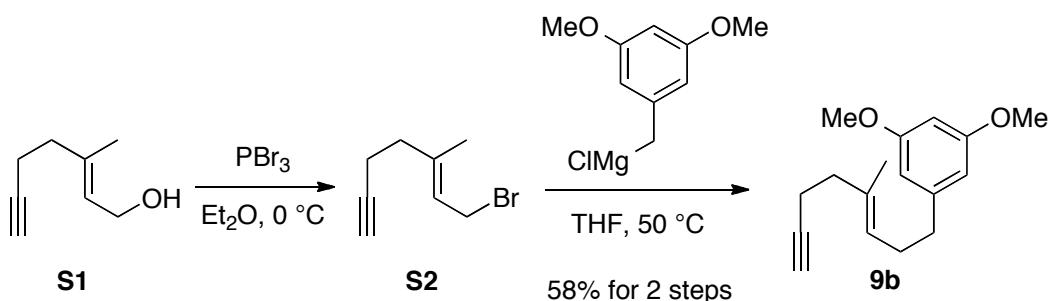
1. General Methods.....	S2
2. Procedures for the Preparation of Substrates for Polycyclisations.....	S2
3. Procedures for Gold(I)-Catalyzed Polycyclisations.....	S20
4. Procedures for Enantioselective Cyclisations.....	S28
5. Crystal Data.....	S33
6. References.....	S44
7. ^1H NMR and ^{13}C NMR Spectra.....	S45

1. General Methods.

All reactions were carried out in solvents dried using a Solvent Purification System (SPS). Thin layer chromatography was carried out using TLC aluminum sheets coated with 0.2 mm of silica gel (Merck Gf234). Chromatographic purifications were carried out using flash grade silica gel (SDS Chromatogel 60 ACC, 40-60 µm). NMR spectra were recorded at 23 °C on Bruker Avance 300, 400 or 500 Ultrashield apparatus. Mass spectra were recorded on a Waters LCT Premier Spectrometer (ESI and APCI) or on an Autoflex Broker Daltonics (MALDI and LDI). Melting points were determined using a Büchi melting point apparatus. Crystal structure determinations were carried out using a Bruker-Nonius diffractometer equipped with an APPEX 2 4K CCD area detector, a FR591 rotating anode with MoKa radiation, Montel mirrors as monochromator and a Kryoflex low temperature device ($T = -173$ °C). Full-sphere data collection was used with w and j scans. Programs used: Data collection APEX-2, data reduction Bruker Saint V./60A and absorption correction SADABS. Structure Solution and Refinement: Crystal structure solution was achieved using direct methods as implement in SHELXTL and visualized using the program XP. Missing atoms were subsequently located from difference Fourier synthesis and added to the atom list. Least-squares refinement on F2 using all measured intensities was carried out using the program SHELXTL. All non-hydrogen atoms were refined including anisotropic displacement parameters. HPLC analysis was carried out on an Agilent Tehcnologies instrument HPLC 1100 series with VWD detector or HPLC 1200 series with DAD detector.

2. Procedures for the Preparation of Substrates for Polycyclisations.

(E)-1,3-dimethoxy-5-(4-methyloct-3-en-7-yn-1-yl)benzene (9b)



Phosphorus tribromide (33 µL, 0.35 mmol) was added dropwise to a solution of **S1**¹ in Et₂O (7 mL) at 0 °C and the mixture was stirred at this temperature for 30 min before 10 mL of water was added. The aqueous layer was extracted with Et₂O (5 mL) and the combined organic layer was washed sequentially with water (10 mL), saturated aqueous NaHCO₃ (10 mL) and brine (10 mL), dried over anhydrous

Na_2SO_4 . The solvent was evaporated to give **S2** as colorless oil which was used without further purification.

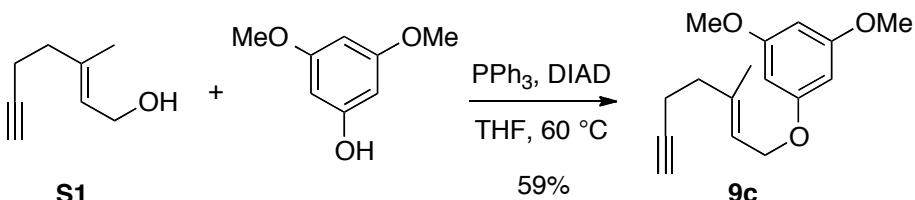
To **S2** (prepared in the last step) was added (3,5-dimethoxybenzyl)magnesium chloride (5.2 mL, 0.2 M in THF, 1.04 mmol) at 0 °C and the mixture was stirred at 50 °C for 5 h before it was cooled to 0 °C and quenched with saturated aqueous NH_4Cl (10 mL). The aqueous layer was extracted with Et_2O (5 mL) and the combined organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na_2SO_4 . The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/ EtOAc 30:1) to give **9b** (103.2 mg, 58% for two steps) as colorless oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 6.39 (d, $J = 2.3$ Hz, 2H), 6.33 (t, $J = 2.3$ Hz, 1H), 5.28 (ddq, $J = 8.4, 7.1, 1.3$ Hz, 1H), 3.81 (s, 6H), 2.62 (dd, $J = 8.9, 6.7$ Hz, 2H), 2.39 - 2.27 (m, 4H), 2.27 - 2.20 (m, 2H), 1.98 (t, $J = 2.5$ Hz, 1H), 1.62(s, 3H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 160.67, 144.7, 134.0, 124.9, 106.6, 97.7, 84.4, 68.4, 55.2, 38.4, 36.3, 29.7, 17.6, 15.8.

HRMS-ESI calculated for $\text{C}_{17}\text{H}_{23}\text{O}_2$ [$\text{M}+\text{H}]^+$: 259.1693; found: 259.1697.

(E)-1,3-dimethoxy-5-((3-methylhept-2-en-6-yn-1-yl)oxy)benzene (**9c**)



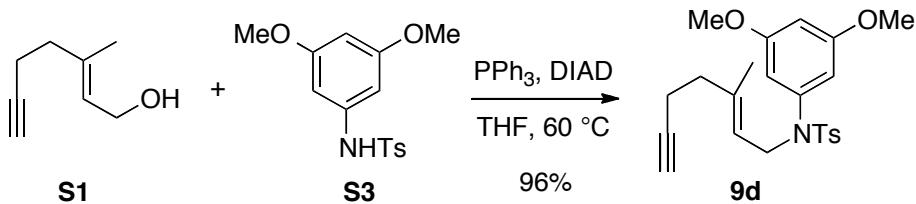
Diisopropyl azodicarboxylate (0.13 mL, 0.66 mmol) was added dropwise to a solution of **S1** (54 mg, 0.44 mmol), 3,5-dimethoxyphenol (101 mg, 0.66 mmol) and triphenylphosphine (171 mg, 0.66 mmol) in THF (5 mL) at 0 °C and the mixture was stirred at 60 °C for 3 h. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/ CH_2Cl_2 2:1) to give **9c** (66.8 mg, 59%) as colorless oil.

$^1\text{H NMR}$ (500 MHz, CDCl_3) δ 6.13 (d, $J = 2.1$ Hz, 2H), 6.12 - 6.10 (m, 1H), 5.57 (ddt, $J = 7.8, 5.2, 1.3$ Hz, 1H), 4.62 - 4.44 (m, 2H), 3.79 (s, 6H), 2.42 - 2.35 (m, 2H), 2.35 - 2.30 (m, 2H), 1.99 (t, $J = 2.5$ Hz, 1H), 1.77 (s, 3H).

$^{13}\text{C NMR}$ (126 MHz, CDCl_3) δ 161.5, 160.7, 139.1, 120.7, 93.6, 93.0, 83.8, 68.8, 64.7, 55.3, 38.1, 17.2, 16.5.

HRMS-ESI calculated for $\text{C}_{16}\text{H}_{20}\text{NaO}_3$ [$\text{M}+\text{Na}]^+$: 283.1305; found: 283.1292.

(E)-N-(3,5-dimethoxyphenyl)-4-methyl-N-(3-methylhept-2-en-6-yn-1-yl)benzenesulfonamide (9d)



9d was prepared by the same method as **9c** from **S1** and **S3**.

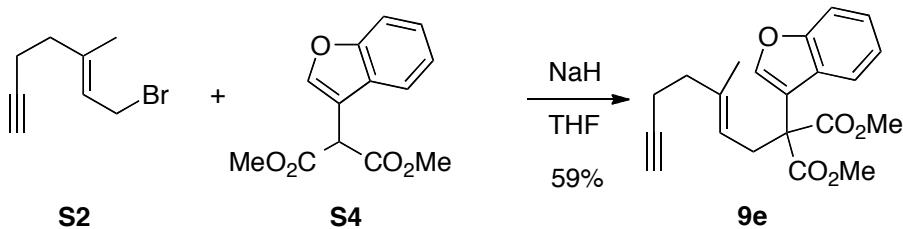
M.p.: 92-94 °C.

¹H NMR (500 MHz, CDCl₃) δ 7.58 (d, *J* = 8.2 Hz, 2H), 7.35 - 7.15 (m, 2H), 6.37 (d, *J* = 2.3 Hz, 1H), 6.22 (d, *J* = 2.3 Hz, 2H), 5.19 (ddt, *J* = 6.9, 5.6, 1.3 Hz, 1H), 4.26 - 4.03 (m, 2H), 3.71 (s, 6H), 2.44 (s, 3H), 2.22 - 2.07 (m, 4H), 1.88 (t, *J* = 2.5 Hz, 1H), 1.55 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 160.5, 143.4, 141.1, 138.4, 135.7, 129.4, 127.8, 120.1, 107.0, 100.0, 83.6, 68.7, 55.4, 48.5, 38.0, 21.5, 17.2, 16.0.

HRMS-ESI calculated for C₂₃H₂₇NNaO₄S [M+Na]⁺: 436.1553; found: 436.1536.

(E)-dimethyl 2-(benzofuran-3-yl)-2-(3-methylhept-2-en-6-yn-1-yl)malonate (9e)



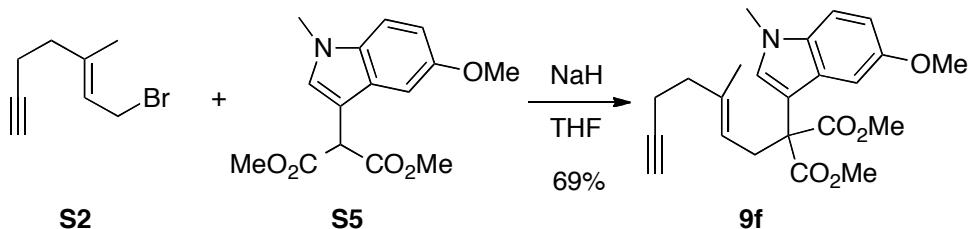
NaH (10.6 mg, 60% dispersion in mineral oil, 0.26 mmol) was added to a solution of **S4**² (54.6 mg, 0.22 mmol) in THF (3 mL) at 0 °C and the mixture was stirred at this temperature for 30 min. A solution of **S2** (49.4 mg, 0.26 mmol) in THF (3 mL) was added and the reaction mixture was stirred at 23 °C for 24 h before it was quenched with water (10 mL). The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 7:1) to give **9e** (45.9 mg, 59%) as colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.56 - 7.49 (m, 2H), 7.31 (ddd, *J* = 8.4, 7.2, 1.3 Hz, 1H), 7.23 (ddd, *J* = 8.1, 7.2, 1.0 Hz, 1H), 5.08 (tq, *J* = 7.4, 1.3 Hz, 1H), 3.76 (s, 6H), 3.17 - 3.12 (m, 2H), 2.23 - 2.11 (m, 4H), 1.93 (t, *J* = 2.5 Hz, 1H), 1.52 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 170.0, 155.0, 144.2, 137.8, 126.0, 124.3, 122.6, 120.6, 119.0, 116.1, 111.6, 83.9, 68.6, 57.1, 52.8, 38.4, 33.5, 17.5, 15.9.

HRMS-ESI calculated for C₂₁H₂₂NaO₅ [M+Na]⁺: 377.1359; found: 377.1372.

(E)-dimethyl 2-(5-methoxy-1-methyl-1*H*-indol-3-yl)-2-(3-methylhept-2-en-6-yn-1-yl)malonate (9f)



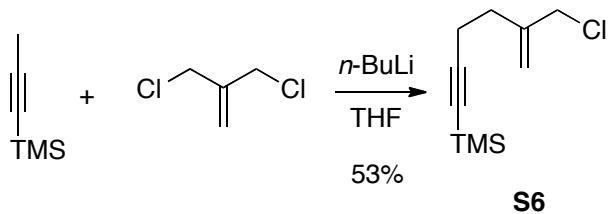
9f was prepared by the same method as 9e from S2 and S5³.

¹H NMR (500 MHz, CDCl₃) δ 7.43 (s, 1H), 7.20 (dd, J = 8.9, 0.5 Hz, 1H), 7.03 (d, J = 2.4 Hz, 1H), 6.90 (dd, J = 8.8, 2.4 Hz, 1H), 5.16 (tdd, J = 7.2, 2.7, 1.4 Hz, 1H), 3.85 (s, 3H), 3.77 (s, 3H), 3.74 (s, 6H), 3.17 - 3.14 (m, 2H), 2.24 - 2.14 (m, 4H), 1.94 (t, J = 2.4 Hz, 1H), 1.57 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 171.1, 153.9, 136.9, 132.4, 129.1, 126.6, 120.1, 111.8, 110.1, 109.3, 102.3, 84.1, 68.5, 57.8, 55.9, 52.6, 38.5, 34.1, 33.1, 17.7, 16.0.

HRMS-ESI calculated for C₂₃H₂₇NNaO₅ [M+Na]⁺: 420.1781; found: 420.1796.

(5-(chloromethyl)hex-5-en-1-yn-1-yl)trimethylsilane (S6)



n-BuLi (12 mL, 2.5 M in hexanes, 30 mmol) was added dropwise to a solution of 1-(trimethylsilyl)propane (4.4 mL, 30 mmol) in THF (300 mL) at -40 °C and the mixture was kept at this temperature for 45 min before it was cooled to -78 °C and 3-chloro-2-(chloromethyl)prop-1-ene (3.47 mL, 30 mmol) was added and the reaction mixture was allowed to warm up to -20 °C during 3 h before it was quenched with saturated aqueous NH₄Cl (300 mL). The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (400 mL) and brine (400 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue

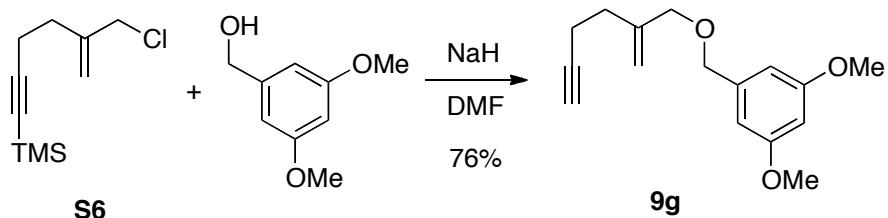
was purified by flash column chromatography (cyclohexane) to give **S6** (3.20 g, 53%) as colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 5.20 (d, J = 1.0 Hz, 1H), 5.04 (d, J = 1.1 Hz, 1H), 4.10 (d, J = 1.0 Hz, 2H), 2.43 (m, 4H), 0.16 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 143.5, 115.5, 106.1, 85.4, 48.0, 32.0, 18.7, 0.1.

HRMS-APCI calculated for C₁₀H₁₈ClSi [M+H]⁺: 201.0861; found: 201.0858.

1,3-dimethoxy-5-((2-methylenehex-5-yn-1-yl)oxy)methyl)benzene (**9g**)



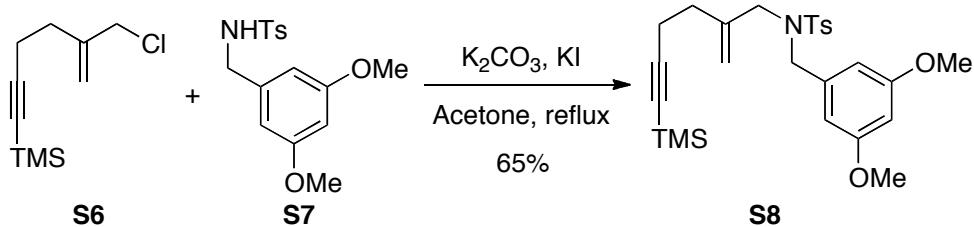
NaH (13.8 mg, 60% dispersion in mineral oil, 0.35 mmol) was added to a solution of 3,5-dimethoxybenzyl alcohol (58.0 mg, 0.35 mmol) in DMF (2 mL) at 0 °C and the mixture was stirred at this temperature for 20 min. A solution of **S6** (63 mg, 0.31 mmol) in DMF (1 mL) was added and the reaction mixture was stirred at 23 °C for 1.5 h before it was quenched with water (5 mL). The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 20:1) to give **9g** (58.0 mg, 76%) as colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 6.53 (d, J = 2.3 Hz, 2H), 6.41 (t, J = 2.3 Hz, 1H), 5.14 (t, J = 1.3 Hz, 1H), 5.04 (d, J = 1.8 Hz, 1H), 4.46 (s, 2H), 4.00 (d, J = 1.2 Hz, 2H), 3.81 (s, 6H), 2.43 - 2.35 (m, 4H), 1.98 (t, J = 2.4 Hz, 1H).

¹³C NMR (75 MHz, CDCl₃) δ 160.9, 144.1, 140.7, 113.2, 105.3, 99.7, 83.9, 72.9, 71.9, 68.6, 55.3, 32.1, 17.1.

HRMS-ESI calculated for C₁₆H₂₀NaO₃ [M+Na]⁺: 283.1305; found: 283.1311.

N-(3,5-dimethoxybenzyl)-4-methyl-N-(2-methylene-6-(trimethylsilyl)hex-5-yn-1-yl)benzenesulfonamide (**S8**)



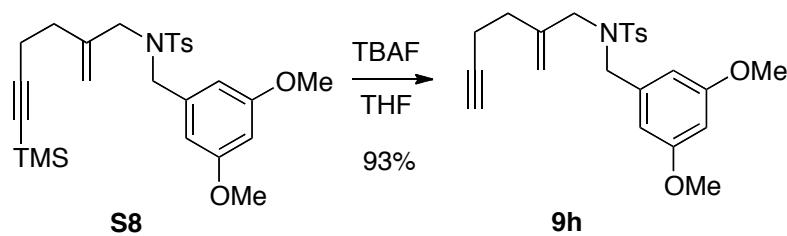
To a solution of **S6** (360 mg, 1.79 mmol) and **S7** (632 mg, 1.97 mmol) in acetone (20 mL) was added K_2CO_3 (272 mg, 1.97 mmol) and KI (29.7 mg, 0.18 mmol) at 23 °C. The mixture was set to reflux for 24 h before the solvent was evaporated. The residue was taken up in Et_2O (20 mL) and washed sequentially with water (20 mL) and brine (20 mL), dried over anhydrous Na_2SO_4 . The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/ EtOAc 5:1) to give **S8** (566 mg, 65%) as yellow oil.

$^1\text{H NMR}$ (400 MHz, CDCl_3) δ 7.75 (d, J = 8.2 Hz, 2H), 7.35 - 7.30 (m, 2H), 6.34 (t, J = 2.3 Hz, 1H), 6.29 (d, J = 2.3 Hz, 2H), 4.94 (d, J = 1.7 Hz, 1H), 4.87 (d, J = 1.1 Hz, 1H), 4.29 (s, 2H), 3.74 (s, 2H), 3.71 (s, 6H), 2.45 (s, 3H), 2.28 - 2.23 (m, 2H), 2.13 (t, J = 7.3 Hz, 2H), 0.15 (s, 9H).

$^{13}\text{C NMR}$ (101 MHz, CDCl_3) δ 160.7, 143.3, 141.8, 138.2, 137.5, 129.7, 127.2, 115.1, 106.5, 106.4, 99.9, 85.1, 55.3, 52.0, 50.8, 31.9, 21.5, 18.3, 0.1.

HRMS-ESI calculated for $\text{C}_{26}\text{H}_{35}\text{NNaO}_4\text{SSI} [\text{M}+\text{Na}]^+$: 508.1948; found: 508.1948.

N-(3,5-dimethoxybenzyl)-4-methyl-N-(2-methylenehex-5-yn-1-yl)benzenesulfonamide (9h)



Tetrabutylammonium fluoride (1.3 mL, 1.0 M in THF, 1.3 mmol) was added dropwise to a solution of **S8** (530 mg, 1.09 mmol) in THF (11 mL) at 0 °C. The reaction mixture was then stirred at 23 °C for 30 min before it was quenched with saturated aqueous NH_4Cl (20 mL). The aqueous layer was extracted with Et_2O (10 mL) and the combined organic layer was washed sequentially with water (20 mL) and brine (20 mL), dried over anhydrous Na_2SO_4 . The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/ EtOAc 5:1) to give **9h** (419 mg, 93%) as a white solid.

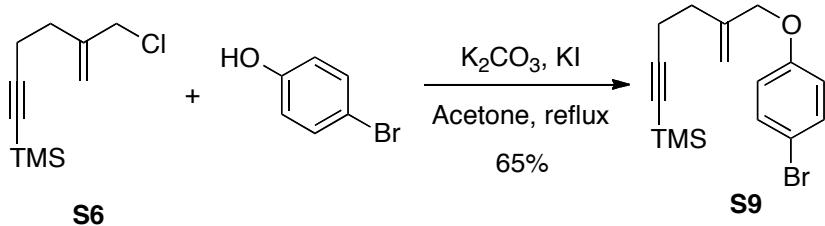
M.p.: 76-77 °C.

¹H NMR (400 MHz, CDCl₃) δ 7.75 (d, J = 8.2 Hz, 2H), 7.35 - 7.31 (m, 2H), 6.34 (t, J = 2.3 Hz, 1H), 6.29 (d, J = 2.3 Hz, 2H), 4.97 - 4.94 (m, 1H), 4.90 - 4.87 (m, 1H), 4.29 (s, 2H), 3.75 (s, 2H), 3.71 (s, 6H), 2.45 (s, 3H), 2.27 - 2.21 (m, 2H), 2.17 - 2.10 (m, 2H), 1.94 (t, J = 2.6 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 160.7, 143.3, 141.8, 138.2, 137.4, 129.7, 127.2, 115.1, 106.4, 99.9, 83.6, 68.7, 55.3, 52.1, 50.9, 31.7, 21.5, 16.8.

HRMS-ESI calculated for C₂₃H₂₇NNaO₄S [M+Na]⁺: 436.1553; found: 436.1549.

(5-((4-bromophenoxy)methyl)hex-5-en-1-yn-1-yl)trimethylsilane (**S9**)



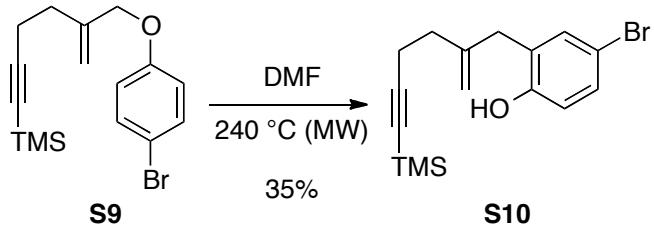
S9 was prepared by the same method as **S8** from **S6** and 4-bromophenol.

¹H NMR (500 MHz, CDCl₃) δ 7.40 - 7.37 (m, 2H), 6.85 - 6.81 (m, 2H), 5.20 (dd, J = 1.3, 0.7 Hz, 1H), 5.09 (q, J = 1.1 Hz, 1H), 4.49 (t, J = 1.1 Hz, 2H), 2.49 - 2.43 (m, 2H), 2.42 - 2.36 (m, 2H), 0.16 (s, 9H).

¹³C NMR (126 MHz, CDCl₃) δ 157.77, 142.80, 132.22, 116.60, 113.60, 113.02, 106.40, 85.33, 70.92, 32.11, 18.87, 0.11.

HRMS-APCI calculated for C₁₆H₂₂BrOSi [M+H]⁺: 337.0618; found: 337.0615.

4-bromo-2-(2-methylene-6-(trimethylsilyl)hex-5-yn-1-yl)phenol (**S10**)



A 5 ml microwave vial was charged with a solution of **S9** (200 mg, 0.59 mmol) in DMF (2 mL). The vial was purged with Argon and then fitted with a septa and vial seal. The reaction was then heated to 240 °C under microwave conditions for 2 hours.

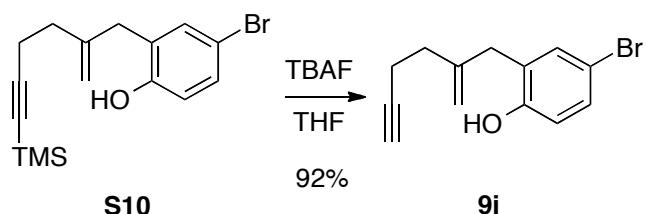
The reaction mixture was poured into water (10 mL) and extracted with Et₂O. The combined organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 10:1) to give **S10** (70 mg, 35%) as colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 7.30 - 7.22 (m, 2H), 6.76 (d, J = 8.5 Hz, 1H), 5.27 (s, 1H), 5.01 (q, J = 1.2 Hz, 1H), 4.87 (t, J = 1.5 Hz, 1H), 3.39 (s, 2H), 2.50 - 2.43 (m, 2H), 2.29 (m, 2H), 0.18 (s, 9H).

¹³C NMR (75 MHz, CDCl₃) δ 153.8, 145.4, 133.5, 130.9, 126.9, 118.0, 113.3, 112.7, 106.3, 86.1, 37.5, 34.3, 18.9, 0.1.

HRMS-ESI calculated for C₁₆H₂₀BrOSi [M-H]⁻: 335.0472; found: 335.0461.

4-bromo-2-(2-methylenehex-5-yn-1-yl)phenol (9i**)**



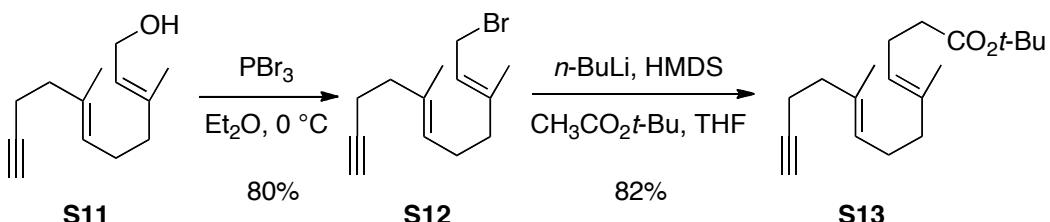
9i was prepared by the same method as **9h** from **S10**.

¹H NMR (500 MHz, CDCl₃) δ 7.29 - 7.24 (m, 2H), 6.75 (d, J = 8.4 Hz, 1H), 5.19 (s, 1H), 5.04 (q, J = 1.2 Hz, 1H), 4.92 (q, J = 1.4 Hz, 1H), 3.39 (s, 2H), 2.43 (tdd, J = 7.1, 2.6, 0.8 Hz, 2H), 2.30 (t, J = 7.0 Hz, 2H), 2.05 (t, J = 2.6 Hz, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 153.6, 145.3, 133.5, 130.9, 126.9, 117.9, 113.3, 112.8, 83.6, 69.3, 37.5, 34.1, 17.2.

HRMS-ESI calculated for C₁₃H₁₂BrO [M-H]⁻: 263.0077; found: 263.0080.

(4E,8E)-tert-butyl 5,9-dimethyltrideca-4,8-dien-12-yneate (S13**)**



S12 was prepared by the same method as **S2** from **S11**⁴.

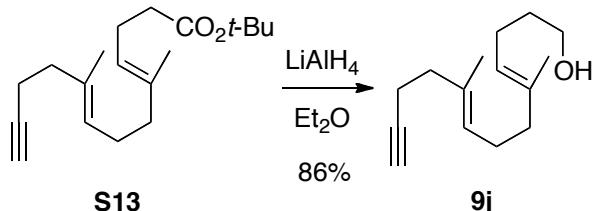
n-BuLi (2.5 mL, 2.5 M in hexanes, 6.3 mmol) was added dropwise to a solution of hexamethyldisilazane (1.37 mL, 6.6 mmol) in THF (15 mL) at -78 °C and the mixture was kept at 0 °C for 30 min before it was re-cooled to -78 °C and *tert*-butyl acetate (0.44 mL, 3.3 mmol) was added. After 1 h, a solution of **S12** (762 mg, 2.99 mmol) in THF (5 mL) was added and the reaction mixture was stirred at 0 °C for 30 min before it was quenched with saturated aqueous NH₄Cl (20 mL). The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (20 mL) and brine (20 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 20:1) to give **S13** (711 mg, 82%) as colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 5.19 (tq, J = 7.1, 1.4 Hz, 1H), 5.12 (ddt, J = 6.9, 5.5, 1.3 Hz, 1H), 2.32 - 2.19 (m, 8H), 2.10 (m, 2H), 2.01 (m, 2H), 1.96 (t, J = 2.6 Hz, 1H), 1.68 - 1.59 (m, 6H), 1.46 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 160.7, 144.7, 134.0, 124.9, 106.6, 97.7, 84.4, 68.4, 55.2, 38.4, 36.3, 29.7, 17.6, 15.8.

HRMS-ESI calculated for C₁₉H₃₀NaO₂ [M+Na]⁺: 313.2138; found: 313.2136.

(4*E*,8*E*)-5,9-dimethyltrideca-4,8-dien-12-yn-1-ol (**9j**)



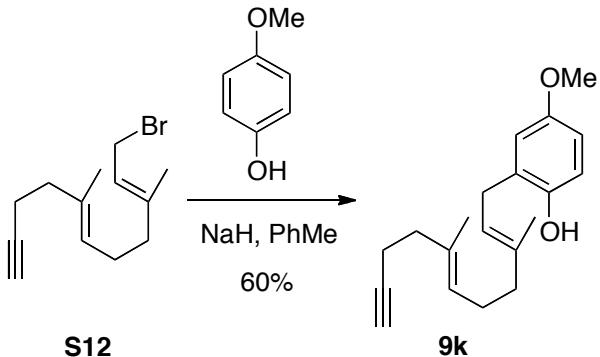
A solution of **S13** (100 mg, 0.34 mmol) in Et₂O (4 mL) was added dropwise to a solution of LiAlH₄ (26.2 mg, 0.69 mmol) in Et₂O (2 mL) at 0 °C and the mixture was kept at this temperature for 1 h before it was quenched with 10% aqueous NaOH (10 mL) at 0 °C. The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (20 mL) and brine (20 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 8:1) to give **9j** (65 mg, 86%) as colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 5.18 (m, 2H), 3.67 (t, J = 6.5 Hz, 2H), 2.33 - 2.26 (m, 2H), 2.25 - 2.20 (m, 2H), 2.15 - 2.07 (m, 4H), 2.03 (m, 2H), 1.97 (t, J = 2.6 Hz, 1H), 1.68 - 1.61 (m, 8H), 1.40 (br m, 1H).

¹³C NMR (75 MHz, CDCl₃) δ 135.6, 133.3, 125.5, 123.9, 84.4, 68.3, 62.7, 39.5, 38.4, 32.7, 26.5, 24.3, 17.6, 16.0, 15.8.

HRMS-APCI calculated for C₁₅H₂₅O [M+H]⁺: 221.1900; found: 221.1896.

2-((2*E*,6*E*)-3,7-dimethylundeca-2,6-dien-10-yn-1-yl)-4-methoxyphenol (9k)



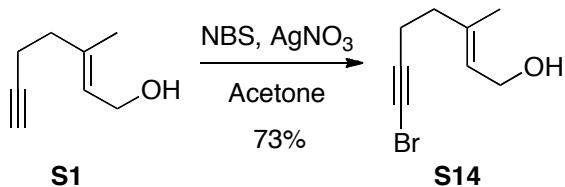
NaH (10.8 mg, 60% dispersion in mineral oil, 0.27 mmol) was added to a solution of 4-methoxyphenol (33.5 mg, 0.27 mmol) in PhMe (2 mL) at 23 °C and the mixture was stirred at this temperature for 15 min. A solution of **S12** (64 mg, 0.25 mmol) in PhMe (1 mL) was added and the reaction mixture was stirred at 23 °C for 15 h before it was quenched with water (5 mL). The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 7:1) to give **9k** (45 mg, 60%) as colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 6.76 (dd, J = 8.5, 0.6 Hz, 1H), 6.72 - 6.66 (m, 2H), 5.33 (tq, J = 7.2, 1.3 Hz, 1H), 5.18 (dddd, J = 6.8, 5.4, 2.7, 1.2 Hz, 1H), 4.79 (s, 1H), 3.78 (s, 3H), 3.35 (d, J = 7.2 Hz, 2H), 2.32 - 2.25 (m, 2H), 2.25 - 2.08 (m, 6H), 1.96 (t, J = 2.5 Hz, 1H), 1.79 (s, 3H), 1.63 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 153.6, 148.2, 138.3, 133.8, 128.1, 125.0, 121.6, 116.3, 115.7, 112.0, 84.5, 68.3, 55.7, 39.5, 38.3, 29.9, 26.3, 17.5, 16.2, 15.9.

HRMS-ESI calculated for C₂₀H₂₆NaO₂ [M+Na]⁺: 321.1830; found: 321.1821.

(E)-7-bromo-3-methylhept-2-en-6-yn-1-ol (S14)



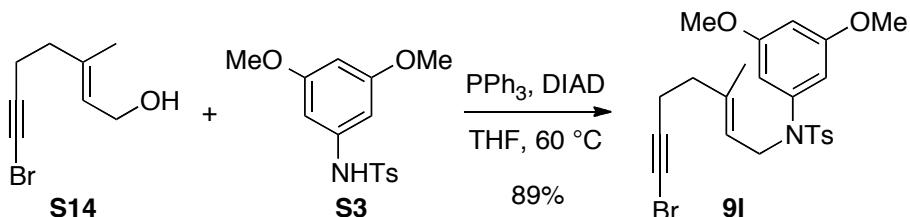
N-bromosuccinimide (117 mg, 0.66 mmol) and silver nitrate (11.2 mg, 66 μ mol) were added sequentially to a solution of **S1** (74.2 mg, 0.6 mmol) in acetone (2 mL) at 23 °C and the resulting mixture was stirred at this temperature for 1 h. The solvent was evaporated and the residue was taken up in Et₂O (10 mL) and washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 4:1) to give **S14** (88.7 mg, 73%) as colorless oil.

¹H NMR (300 MHz, CDCl₃) δ 5.47 (tdd, J = 6.9, 2.7, 1.3 Hz, 1H), 4.18 (d, J = 6.9 Hz, 2H), 2.40 - 2.30 (m, 2H), 2.25 (m, 2H), 1.69 (s, 3H), 1.31 (br s, 1H).

¹³C NMR (75 MHz, CDCl₃) δ 137.5, 124.8, 79.6, 59.3, 38.4, 37.9, 18.5, 16.1.

HRMS-ESI calculated for C₈H₁₁BrNaO [M+Na]⁺: 224.9885; found: 224.9888.

(E)-N-(7-bromo-3-methylhept-2-en-6-yn-1-yl)-N-(3,5-dimethoxyphenyl)-4-methylbenzenesulfonamide (9l)



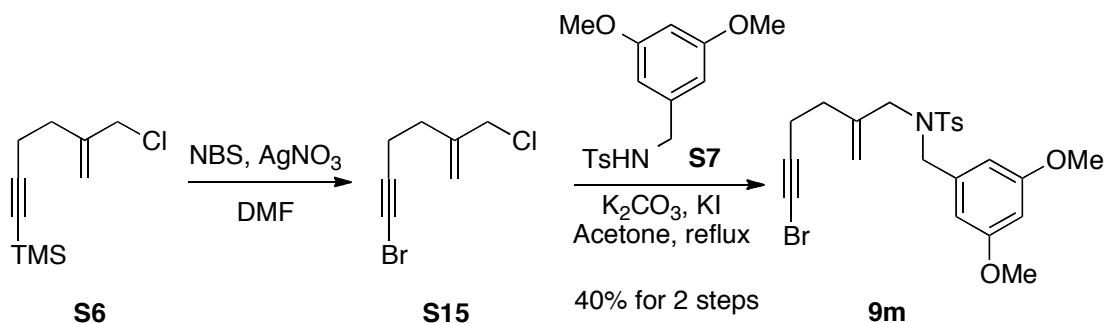
9l was prepared by the same method as **9c** from **S14** and **S3**.

¹H NMR (400 MHz, CDCl₃) δ 7.61 - 7.55 (m, 2H), 7.32 - 7.23 (m, 2H), 6.39 (t, J = 2.3 Hz, 1H), 6.22 (d, J = 2.3 Hz, 2H), 5.18 (tq, J = 6.9, 1.2 Hz, 1H), 4.20 - 4.10 (m, 2H), 3.73 (s, 6H), 2.45 (s, 3H), 2.22 - 2.16 (m, 2H), 2.12 (dd, J = 8.6, 5.2 Hz, 2H), 1.56 - 1.53 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 160.5, 143.4, 141.2, 138.4, 135.7, 129.4, 127.8, 120.1, 107.0, 100.1, 79.5, 55.4, 48.5, 38.3, 37.8, 21.6, 18.6, 16.2.

HRMS-ESI calculated for C₂₃H₂₆BrNNaO₄S [M+Na]⁺: 514.0653; found: 514.0658.

N-(6-bromo-2-methylenehex-5-yn-1-yl)-N-(3,5-dimethoxybenzyl)-4-methylbenzenesulfonamide (9m)



N-bromosuccinimide (100.7 mg, 0.56 mmol) and silver nitrate (8.7 mg, 0.05 mmol) were added sequentially to a solution of **S6** (103.4 mg, 0.51 mmol) in DMF (2 mL) at 23 °C and the resulting mixture was stirred at this temperature for 1 h. The mixture was diluted with water (10 mL) and extracted with Et₂O (10 mL). The organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated to give **S15** as colorless oil which was used without further purification.

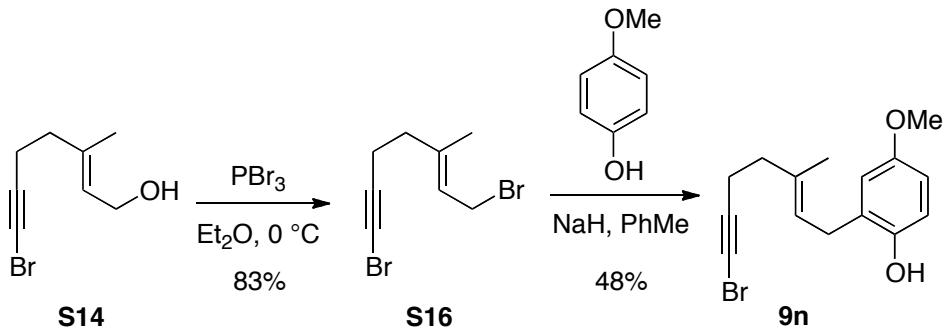
To a solution of **S15** (62.3 mg, 0.3 mmol) and **S7** (115.6 mg, 0.36 mmol) in acetone (5 mL) was added K₂CO₃ (49.7 mg, 0.36 mmol) and KI (5.0 mg, 0.03 mmol) at 23 °C. The mixture was set to reflux for 24 h before the solvent was evaporated. The residue was taken up in Et₂O (10 mL) and washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 5:1) to give **9m** (100 mg, 40% from) as colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 7.77 - 7.72 (m, 2H), 7.35 - 7.31 (m, 2H), 6.35 (t, J = 2.3 Hz, 1H), 6.29 (d, J = 2.3 Hz, 2H), 4.92 (d, J = 1.5 Hz, 1H), 4.87 (d, J = 1.4 Hz, 1H), 4.28 (s, 2H), 3.73 (s, 2H), 3.72 (s, 6H), 2.45 (s, 3H), 2.24 (td, J = 7.2, 1.0 Hz, 2H), 2.10 (t, J = 7.3 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 160.8, 143.4, 141.7, 138.2, 137.4, 129.7, 127.2, 115.3, 106.4, 99.9, 79.4, 55.3, 52.1, 51.0, 38.4, 31.4, 21.5, 18.0.

HRMS-ESI calculated for C₂₃H₂₆BrNNaO₄S [M+Na]⁺: 514.0658; found: 514.0652.

(E)-2-(7-bromo-3-methylhept-2-en-6-yn-1-yl)-4-methoxyphenol (**9n**)



S16 was prepared by the same method as **S2** from **S14**.

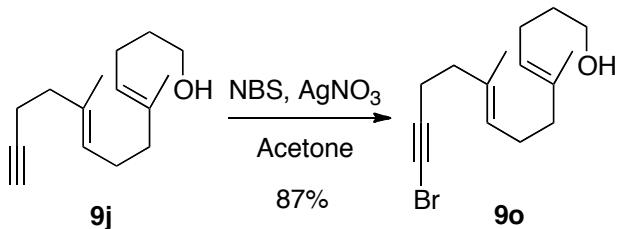
9n was prepared by the same method as **9k** from **S16** and 4-methoxyphenol.

¹H NMR (500 MHz, CDCl₃) δ 6.77 (d, J = 8.6 Hz, 1H), 6.71 (d, J = 3.2 Hz, 1H), 6.68 (dd, J = 8.6, 3.1 Hz, 1H), 5.40 (tq, J = 7.2, 1.3 Hz, 1H), 4.68 (s, 1H), 3.78 (s, 3H), 3.37 (d, J = 7.1 Hz, 2H), 2.40 - 2.35 (m, 2H), 2.29 (m, 2H), 1.79 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 153.7, 148.0, 136.1, 128.0, 123.1, 116.4, 115.7, 112.1, 79.6, 55.7, 38.6, 38.1, 29.7, 18.7, 16.0.

HRMS-ESI calculated for C₁₅H₁₇BrNaO₂ [M+Na]⁺: 331.0311; found: 331.0317.

(4*E*,8*E*)-13-bromo-5,9-dimethyltrideca-4,8-dien-12-yn-1-ol (**9o**)



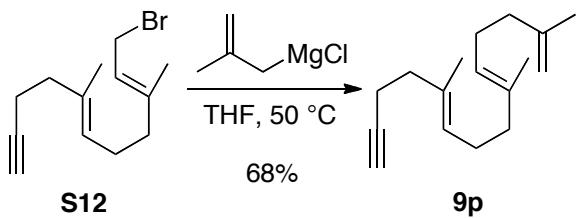
9o was prepared by the same method as **S14** from **9j**.

¹H NMR (500 MHz, CDCl₃) δ 5.18 (m, 2H), 3.67 (t, J = 6.5 Hz, 2H), 2.34 - 2.27 (m, 2H), 2.24 - 2.18 (m, 2H), 2.15 - 2.06 (m, 4H), 2.06 - 2.00 (m, 2H), 1.72 - 1.63 (m, 2H), 1.64 (s, 3H), 1.62 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 135.6, 133.1, 125.7, 123.9, 80.1, 62.8, 39.5, 38.2, 37.9, 32.8, 26.5, 24.3, 18.9, 16.0, 15.8.

HRMS-ESI calculated for C₁₅H₂₃BrNaO [M+Na]⁺: 321.0832; found: 321.0836.

(5*E*,9*E*)-2,6,10-trimethyltetradeca-1,5,9-trien-13-yne (**9p**)



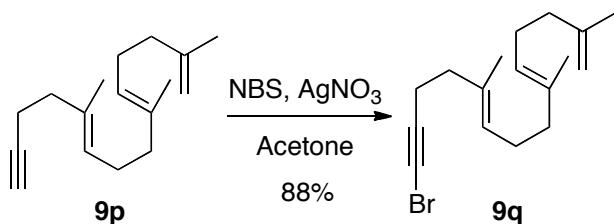
To a solution of **S12** (368 mg, 1.44 mmol) in THF (10 mL) was added (2-methylallyl)magnesium chloride (4.3 mL, 0.5 M in THF, 2.16 mmol) at 0 °C and the mixture was stirred at 50 °C for 8 h before it was cooled to 0 °C and quenched with saturated aqueous NH₄Cl (10 mL). The aqueous layer was extracted with Et₂O (5 mL) and the combined organic layer was washed sequentially with water (10 mL) and brine (10 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane) to give **9p** (225 mg, 68%) as colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 5.20 (ddq, J = 7.0, 5.6, 1.3 Hz, 1H), 5.15 (tq, J = 6.9, 1.3 Hz, 1H), 4.74 (ddq, J = 2.8, 2.0, 0.9 Hz, 1H), 4.71 (dq, J = 2.1, 1.1 Hz, 1H), 2.29 (tdd, J = 6.9, 2.5, 1.1 Hz, 2H), 2.23 (ddt, J = 8.3, 7.0, 1.3 Hz, 2H), 2.19 - 2.09 (m, 4H), 2.09 - 2.00 (m, 4H), 1.97 (t, J = 2.5 Hz, 1H), 1.76 - 1.74 (s, 3H), 1.65 - 1.62 (s, 6H).

¹³C NMR (126 MHz, CDCl₃) δ 145.9, 135.0, 133.1, 125.6, 124.2, 109.8, 84.5, 68.3, 39.5, 38.4, 37.8, 26.5, 26.2, 22.5, 17.6, 16.0, 15.8.

HRMS-APCI calculated for C₁₇H₂₇ [M+H]⁺: 231.2107; found: 231.2100.

(5E,9E)-14-bromo-2,6,10-trimethyltetradeca-1,5,9-trien-13-yne (**9q**)



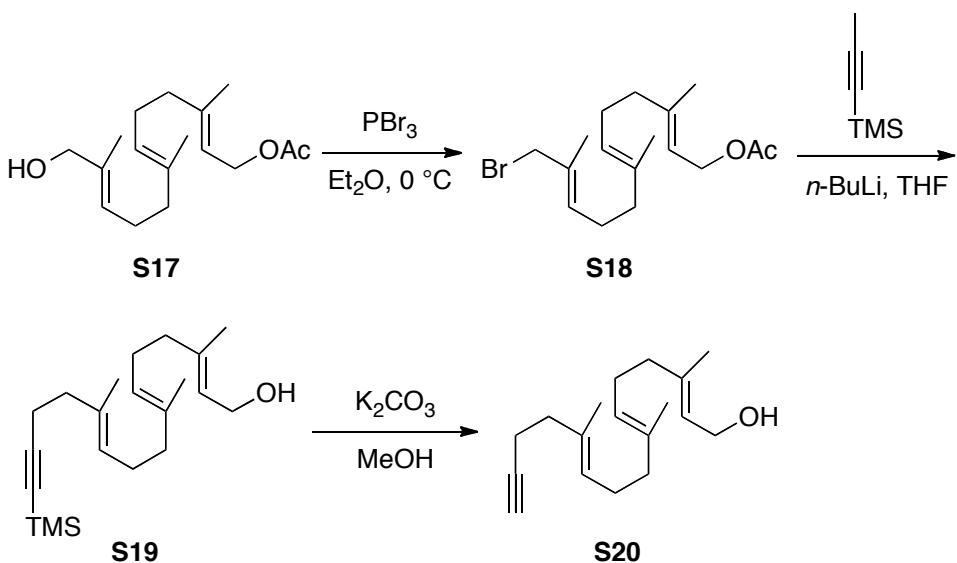
9q was prepared by the same method as **S14** from **9p**.

¹H NMR (400 MHz, CDCl₃) δ 5.24 - 5.09 (m, 2H), 4.72 (ddd, J = 11.6, 2.3, 1.2 Hz, 2H), 2.30 (m, 2H), 2.23 - 1.99 (m, 10H), 1.75 (s, 3H), 1.63 (s, 3H), 1.62 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 145.9, 135.0, 132.9, 125.8, 124.2, 109.8, 80.1, 39.5, 38.2, 37.9, 37.8, 26.5, 26.2, 22.5, 18.9, 16.0, 15.8.

HRMS-APCI calculated for C₁₇H₂₆Br [M+H]⁺: 309.1212; found: 309.1216.

(2E,6E,10E)-3,7,11-trimethylpentadeca-2,6,10-trien-14-yn-1-ol (S20)



S18 was prepared by the same method as **S2** from **S17**⁵ (2.04 g, 7.3 mmol).

n-BuLi (11.7 mL, 2.5 M in hexanes, 29.2 mmol) was added dropwise to a solution of 1-(trimethylsilyl)propane (4.3 mL, 29.2 mmol) in THF (80 mL) at -40 °C and the mixture was kept at this temperature for 45 min before it was cooled to -60 °C and a solution of **S18** (prepared in the last step) in THF (20 mL) was added and the reaction mixture was stirred at -60 °C for 1 h before it was quenched with saturated aqueous NH₄Cl (100 mL). The aqueous layer was extracted with Et₂O and the combined organic layer was washed sequentially with water (100 mL) and brine (100 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was subjected to the next step without further purification.

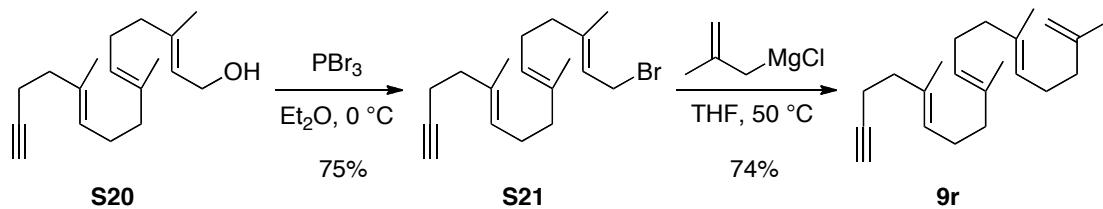
K₂CO₃ (4.03 g, 29.2 mmol) was added to a solution of the residue above in MeOH (40 mL) at 23 °C and the mixture was stirred at this temperature for 12 h. The solvent was evaporated and the residue was taken up in Et₂O (20 mL) and washed sequentially with water (20 mL) and brine (20 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 7:1) to give **S20** (930 mg, 49% for three steps) as colorless oil.

¹H NMR (500 MHz, CDCl₃) δ 5.44 (m, 1H), 5.20 (m, 1H), 5.14 (m, 1H), 4.18 (d, J = 6.9 Hz, 2H), 2.29 (m, 2H), 2.26 - 2.20 (m, 2H), 2.18 - 1.99 (m, 8H), 1.97 (t, J = 2.5 Hz, 1H), 1.71 (s, 3H), 1.63 (s, 3H), 1.62 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 139.8, 135.2, 133.2, 125.5, 123.9, 123.4, 84.4, 77.2, 68.3, 59.4, 39.5, 38.4, 26.6, 26.3, 17.6, 16.3, 16.0, 15.8.

HRMS-APCI calculated for C₁₈H₂₉O [M+H]⁺: 261.2218; found: 261.2215.

(5E,9E,13E)-2,6,10,14-tetramethyloctadeca-1,5,9,13-tetraen-17-yne (9r)



S21 was prepared by the same method as **S2** from **S20**.

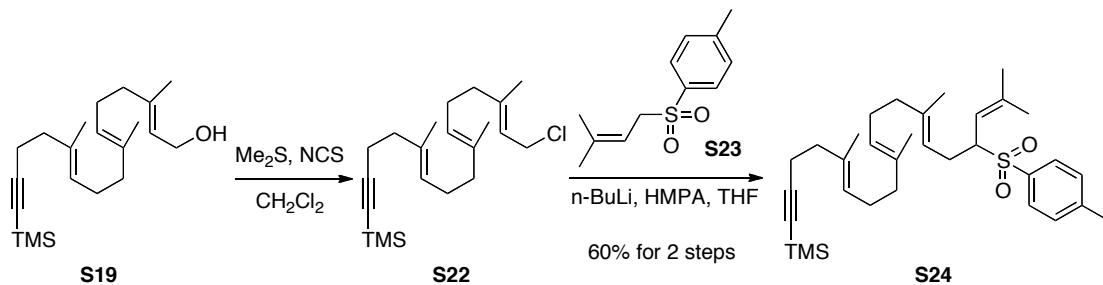
9r was prepared by the same method as **9p** from **S21**.

¹H NMR (400 MHz, CDCl₃) δ 5.23 - 5.11 (m, 3H), 4.74 - 4.69 (m, 2H), 2.33 - 2.26 (m, 2H), 2.25 - 2.21 (m, 2H), 2.17 - 1.98 (m, 12H), 1.96 (t, J = 2.5 Hz, 1H), 1.75 (s, 3H), 1.63 (s, 6H), 1.62 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 145.9, 135.1, 134.7, 133.1, 125.6, 124.4, 124.1, 109.7, 84.4, 68.3, 39.7, 39.6, 38.4, 37.8, 26.6, 26.2, 22.5, 17.6, 16.0, 15.8.

HRMS-APCI calculated for C₂₂H₃₅ [M+H]⁺: 299.2733; found: 299.2745.

Trimethyl((5E,9E,13E)-5,9,13,18-tetramethyl-16-tosylnonadeca-5,9,13,17-tetraen-1-yn-1-yl)silane (S24)



Dimethyl sulfide (0.28 mL, 3.75 mmol) was added to a solution of *N*-chlorosuccinimide (459 mg, 3.44 mmol) in CH₂Cl₂ (10 mL) at -30 °C and the reaction mixture was stirred at this temperature for 15 min. A solution of **S19** (1.04 g, 3.13 mmol) in CH₂Cl₂ (5 mL) was added slowly and the reaction mixture was allowed to warm to room temperature and stirred for another 2 h. the reaction was quenched by the addition of water (20 mL) and the organic layer was washed sequentially with water (20×3 mL) and brine (20 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was subjected to the next step without further purification.

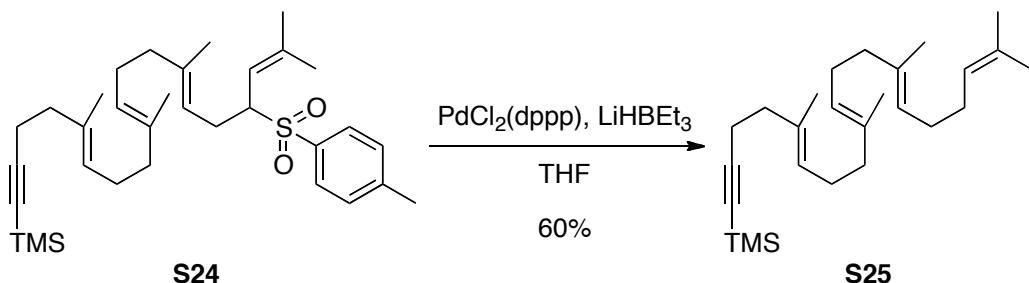
n-BuLi (1.5 mL, 2.5 M in hexanes, 3.76 mmol) and hexamethylphosphoramide (0.71 mL, 4.07 mmol) was added sequentially to a solution of **S23**⁶ (701 mg, 3.13 mmol) in THF (8 mL) at -20 °C and the mixture was kept at this temperature for 30 min before it was cooled to -78 °C and a solution of **S22** (prepared in the last step) in THF (5 mL) was added and the reaction mixture was stirred at -78 °C for 1 h. The cooling bath was removed and the mixture was stirred at room temperature for another 1 h before it was quenched with saturated aqueous NH₄Cl (40 mL). The aqueous layer was extracted with Et₂O (30 mL) and the combined organic layer was washed sequentially with water (40 mL) and brine (40 mL), dried over anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 10:1) to give **S24** (1.01 g, 60% for 2 steps) as colorless oil.

¹H NMR (400 MHz, CDCl₃) δ 7.74 - 7.70 (m, 2H), 7.34 - 7.30 (m, 2H), 5.19 - 5.13 (m, 1H), 5.07 (tt, J = 5.6, 3.0 Hz, 1H), 4.98 (dddd, J = 7.5, 4.7, 3.0, 1.5 Hz, 2H), 3.70 (td, J = 10.6, 3.4 Hz, 1H), 2.89 - 2.79 (m, 1H), 2.45 (s, 3H), 2.39 - 2.25 (m, 3H), 2.19 (dd, J = 8.3, 6.7 Hz, 2H), 2.12 - 1.93 (m, 8H), 1.69 (d, J = 1.5 Hz, 3H), 1.63 - 1.57 (m, 9H), 1.23 (d, J = 1.4 Hz, 3H), 0.15 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 144.2, 141.5, 138.5, 135.2, 135.0, 133.4, 129.3, 129.1, 125.5, 124.0, 118.7, 117.3, 107.4, 84.5, 65.0, 39.7, 39.6, 38.7, 26.6, 26.6, 26.6, 25.8, 21.6, 19.2, 18.1, 16.4, 16.0, 15.8, 0.2.

HRMS-APCI calculated for C₃₃H₅₁O₂SSi [M+H]⁺: 539.3379; found: 539.3376.

Trimethyl((5E,9E,13E)-5,9,13,18-tetramethylnonadeca-5,9,13,17-tetraen-1-yn-1-yl)silane (S25**)**



To a solution of **S24** (812 mg, 1.51 mmol) in THF (15 mL) was added PdCl₂(dppp) (89 mg, 0.151 mmol) at 0 °C. Lithium triethylborohydride (LiHBET₃, 1.0 M solution in THF, 5 mL, 5.0 mmol) was then added slowly to the solution over a 1 min period. The reaction mixture was stirred for an additional 4 h at 0 °C and then diluted with Et₂O (20 mL), followed by the addition of saturated NH₄Cl (40 mL).. The organic layer was washed sequentially with water (40 mL×2), and brine (40 mL), dried over

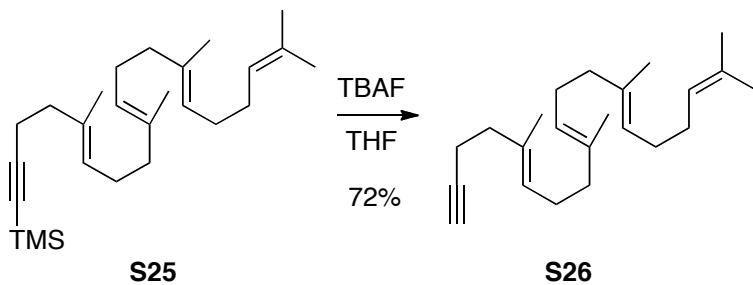
anhydrous Na₂SO₄. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane) to give **S25** (349 mg, 60%).

¹H NMR (400 MHz, CDCl₃) δ 5.23 - 5.10 (m, 4H), 2.32 (ddd, J = 7.8, 6.9, 1.2 Hz, 2H), 2.24 - 2.18 (m, 2H), 2.14 - 2.06 (m, 4H), 2.06 - 1.97 (m, 8H), 1.71 (d, J = 1.5 Hz, 3H), 1.64 - 1.61 (m, 12H), 0.17 (s, 9H).

¹³C NMR (101 MHz, CDCl₃) δ 135.1, 134.8, 133.3, 131.4, 125.6, 124.5, 124.4, 124.3, 107.4, 84.5, 39.7, 39.6, 38.7, 28.4, 28.3, 26.6, 25.7, 19.2, 17.7, 16.0, 16.0, 15.8, 0.2.

HRMS-APCI calculated for C₂₆H₄₅Si [M+H]⁺: 385.3290; found: 385.3296.

(5E,9E,13E)-5,9,13,18-tetramethylnonadeca-5,9,13,17-tetraen-1-yne (**S26**)



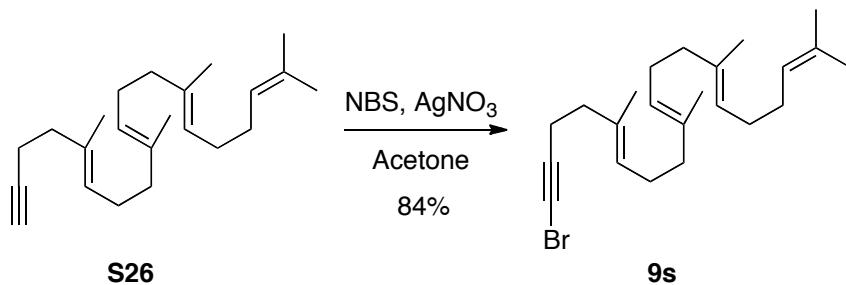
S26 was prepared by the same method as **9h** from **S25**.

¹H NMR (500 MHz, CDCl₃) δ 5.23 - 5.12 (m, 4H), 2.30 (dd, J = 7.9, 6.8, 2.5, 1.6 Hz, 2H), 2.25 - 2.20 (m, 2H), 2.15 - 2.07 (m, 4H), 2.05 - 1.99 (m, 8H), 1.96 (t, J = 2.6 Hz, 1H), 1.71 (d, J = 1.5 Hz, 3H), 1.64 - 1.62 (m, 12H).

¹³C NMR (126 MHz, CDCl₃) δ 135.1, 134.7, 133.1, 131.4, 125.6, 124.5, 124.4, 124.3, 84.4, 68.3, 39.7, 39.6, 38.4, 28.4, 28.3, 26.6, 26.6, 25.7, 17.7, 17.6, 16.0, 16.0, 15.8.

HRMS-APCI calculated for C₂₃H₃₇ [M+H]⁺: 313.2895; found: 313.2899.

(5E,9E,13E)-1-bromo-5,9,13,18-tetramethylnonadeca-5,9,13,17-tetraen-1-yne (**9s**)



9s was prepared by the same method as **S14** from **S26**.

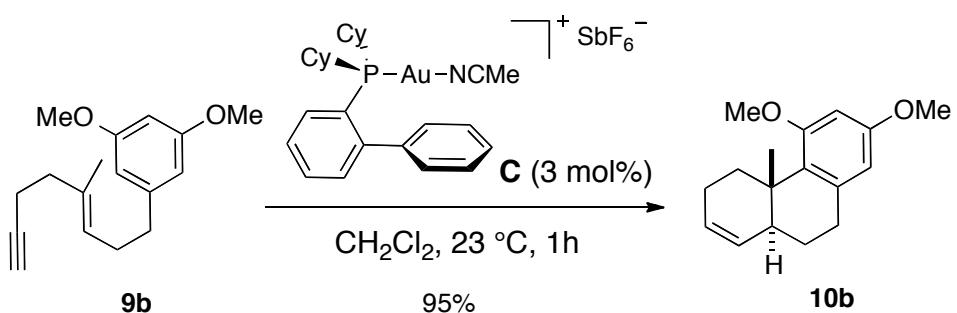
¹H NMR (400 MHz, CDCl₃) δ 5.24 - 5.10 (m, 4H), 2.36 - 2.25 (m, 2H), 2.25 - 2.16 (m, 2H), 2.15 - 2.06 (m, 4H), 2.06 - 1.98 (m, 8H), 1.71 (s, 3H), 1.63 (m, 15H).

¹³C NMR (101 MHz, CDCl₃) δ 135.1, 134.7, 132.9, 131.5, 125.8, 124.5, 124.4, 124.3, 80.1, 39.7, 39.5, 38.2, 37.9, 28.4, 28.3, 26.7, 26.6, 25.7, 18.9, 17.7, 16.0, 16.0, 15.8.

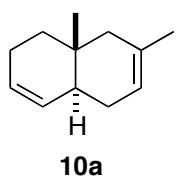
HRMS-ESI calculated for C₂₃H₃₅BrNa [M+Na]⁺: 413.1822; found: 413.1825.

3. Procedures for Gold(I)-Catalyzed Polycyclisations

Representative procedure for gold(I)-catalyzed polycyclisations



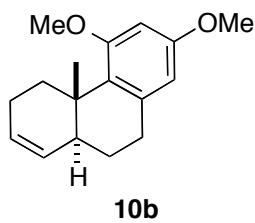
C (4.9 mg, 6 μ mol) was added to a solution of **9b** (51.6 mg, 0.2 mmol) in CH₂Cl₂ (2 mL) and the mixture was stirred at 23 °C for 1 h. The solvent was evaporated and the residue was purified by flash column chromatography (cyclohexane/EtOAc 30:1) to give **10b** (49 mg, 95%) as colorless oil.



¹H NMR (400 MHz, CDCl₃) δ 5.67 - 5.60 (m, 1H), 5.43 - 5.36 (m, 2H), 2.18 - 1.93 (m, 4H), 1.91 - 1.82 (m, 1H), 1.78 - 1.63 (m, 2H), 1.68 (m, 3H), 1.54 - 1.37 (m, 2H), 0.81 - 0.78 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 133.4, 130.2, 126.1, 120.5, 45.9, 39.5, 36.7, 31.0, 28.7, 24.0, 23.2, 15.8.

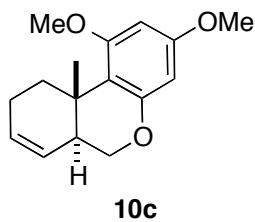
HRMS-ESI calculated for C₁₂H₁₉ [M+H]⁺: 163.1488; found: 163.1494.



¹H NMR (400 MHz, CDCl₃) δ 6.34 (d, J = 2.6 Hz, 1H), 6.27 (dt, J = 2.6, 0.8 Hz, 1H), 5.70 (dq, J = 9.9, 3.7 Hz, 1H), 5.48 (dq, J = 9.7, 2.0 Hz, 1H), 3.82 (s, 3H), 3.80 (s, 3H), 3.16 (ddt, J = 13.2, 6.6, 1.3 Hz, 1H), 3.01 (dddt, J = 17.1, 12.1, 7.1, 1.0 Hz, 1H), 2.87 - 2.77 (m, 1H), 2.41 (dddd, J = 12.3, 6.1, 3.1, 1.9 Hz, 1H), 2.34 - 2.12 (m, 2H), 1.75 (tdd, J = 13.1, 12.1, 5.7 Hz, 1H), 1.64 (ddt, J = 8.7, 4.2, 1.4 Hz, 1H), 1.54 - 1.44 (m, 1H), 1.18 (m, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 160.1, 158.0, 139.0, 131.2, 127.8, 127.0, 105.2, 97.3, 55.1, 55.0, 44.0, 36.5, 32.4, 32.3, 24.6, 24.5, 17.0.

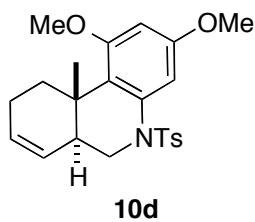
HRMS-ESI calculated for C₁₇H₂₃O₂ [M+H]⁺: 259.1693; found: 259.1687.



¹H NMR (400 MHz, CDCl₃) δ 6.10 - 6.00 (m, 2H), 5.84 - 5.76 (m, 1H), 5.38 (dq, J = 9.8, 2.2 Hz, 1H), 4.10 (dd, J = 10.3, 4.0 Hz, 1H), 3.99 (dd, J = 12.5, 10.3 Hz, 1H), 3.78 (s, 3H), 3.76 (s, 3H), 3.10 - 3.01 (m, 1H), 2.71 (ddqd, J = 12.8, 5.8, 3.4, 1.9 Hz, 1H), 2.29 - 2.18 (m, 2H), 1.56 - 1.49 (m, 1H), 1.18 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 160.1, 159.1, 155.7, 129.2, 123.6, 113.6, 94.0, 92.1, 66.2, 55.2, 55.2, 41.5, 33.2, 31.8, 24.2, 17.9.

HRMS-ESI calculated for C₁₆H₂₁O₃ [M+H]⁺: 261.1485; found: 261.1473.

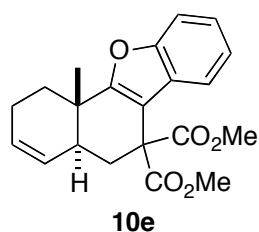


M.p.: 111-112 °C.

¹H NMR (400 MHz, CDCl₃) δ 7.53 (d, J = 8.3 Hz, 2H), 7.24 - 7.18 (m, 3H), 6.24 (d, J = 2.5 Hz, 1H), 5.73 (dq, J = 9.9, 3.3 Hz, 1H), 5.35 (dq, J = 9.8, 2.1 Hz, 1H), 4.02 (dd, J = 11.9, 4.3 Hz, 1H), 3.82 (s, 3H), 3.74 (s, 3H), 3.24 (dd, J = 13.6, 11.9 Hz, 1H), 3.00 (dt, J = 13.4, 4.4 Hz, 1H), 2.38 (s, 3H), 2.35 - 2.31 (m, 1H), 2.09 (m, 2H), 1.29 (m, 1H), 0.67 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 159.2, 158.1, 143.7, 137.8, 135.7, 129.5, 128.6, 127.3, 124.9, 120.3, 100.3, 96.1, 55.3, 55.3, 47.7, 40.9, 35.0, 31.8, 23.7, 21.5, 16.3.

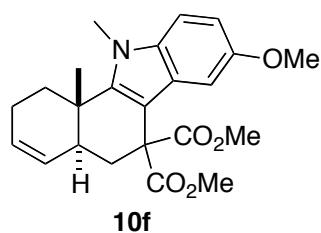
HRMS-ESI calculated for C₂₃H₂₇NNaO₄S [M+Na]⁺: 436.1553; found: 436.1533.



¹H NMR (300 MHz, CDCl₃) δ 7.57 - 7.52 (m, 1H), 7.47 - 7.42 (m, 1H), 7.26 - 7.20 (m, 2H), 5.73 (ddd, J = 10.2, 4.3, 2.7 Hz, 1H), 5.56 (dq, J = 9.9, 2.0 Hz, 1H), 3.78 (s, 3H), 3.76 (s, 3H), 2.76 (dd, J = 14.0, 4.9 Hz, 1H), 2.66 (dd, J = 13.6, 2.6 Hz, 1H), 2.37 - 2.16 (m, 4H), 1.82 - 1.70 (m, 1H), 1.23 (s, 3H).

¹³C NMR (75 MHz, CDCl₃) δ 171.4, 170.8, 163.7, 154.4, 127.6, 127.4, 123.2, 122.6, 121.3, 111.1, 107.7, 55.2, 52.9, 52.7, 40.0, 34.5, 32.6, 30.6, 22.9, 18.3.

HRMS-ESI calculated for C₂₁H₂₂NaO₅ [M+Na]⁺: 377.1359; found: 377.1373.

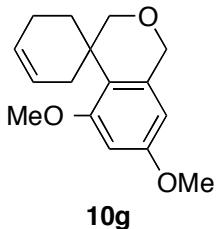


M.p.: 165-168 °C.

¹H NMR (500 MHz, CDCl₃) δ 7.18 (d, J = 8.9 Hz, 1H), 7.06 (d, J = 2.4 Hz, 1H), 6.88 (dd, J = 8.9, 2.5 Hz, 1H), 5.76 (dq, J = 9.9, 3.3 Hz, 1H), 5.56 (dq, J = 9.9, 2.1 Hz, 1H), 3.86 (s, 3H), 3.85 (s, 3H), 3.77 (s, 3H), 3.75 (s, 3H), 2.78 (m, 1H), 2.61 (m, 2H), 2.35 - 2.23 (m, 3H), 1.86 (m, 1H), 1.33 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 172.8, 171.8, 154.2, 145.9, 133.1, 129.1, 126.9, 126.5, 111.1, 109.4, 104.6, 103.2, 56.3, 56.1, 52.7, 52.6, 41.3, 35.3, 33.2, 32.7, 32.1, 23.6, 17.5.

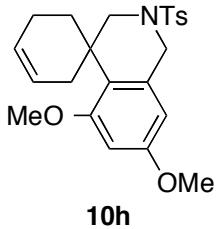
HRMS-ESI calculated for C₂₃H₂₈NO₅ [M+H]⁺: 398.1962; found: 398.1963.



¹H NMR (500 MHz, CDCl₃) δ 6.37 (d, J = 2.5 Hz, 1H), 6.20 - 6.09 (m, 1H), 5.73 (s, 2H), 4.77 - 4.63 (m, 2H), 3.95 (d, J = 11.4 Hz, 1H), 3.82 (s, 3H), 3.80 (s, 3H), 3.62 (dd, J = 11.3, 1.3 Hz, 1H), 2.90 - 2.82 (m, 1H), 2.75 (ddd, J = 13.3, 10.2, 9.1 Hz, 1H), 2.12 - 2.01 (m, 3H), 1.50 - 1.42 (m, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 159.8, 158.6, 137.5, 126.4, 125.4, 122.5, 99.9, 98.1, 73.8, 70.0, 55.2, 55.0, 34.5, 31.5, 27.4, 22.0.

HRMS-ESI calculated for C₁₆H₂₁O₃ [M+H]⁺: 261.1485; found: 261.1475.

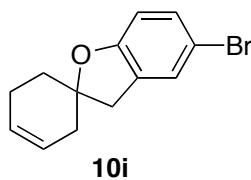


M.p.: 176-178 °C.

¹H NMR (500 MHz, CDCl₃) δ 7.75 (d, J = 8.2 Hz, 2H), 7.39 - 7.35 (m, 2H), 6.35 (d, J = 2.5 Hz, 1H), 6.20 - 6.16 (m, 1H), 5.80 - 5.74 (m, 1H), 5.74 - 5.67 (m, 1H), 4.24 - 4.16 (m, 1H), 4.04 (dd, J = 14.5, 1.1 Hz, 1H), 3.79 (s, 3H), 3.77 (s, 3H), 3.19 (dd, J = 11.5, 1.5 Hz, 1H), 3.04 - 2.96 (m, 2H), 2.70 (ddd, J = 13.3, 11.8, 6.5 Hz, 1H), 2.46 (s, 3H), 2.18 (m, 1H), 2.13 - 2.02 (m, 1H), 1.94 (ddd, J = 18.5, 4.2, 2.2 Hz, 1H), 1.52 - 1.44 (m, 1H).

¹³C NMR (126 MHz, CDCl₃) δ 159.7, 158.6, 143.6, 134.3, 133.0, 129.7, 127.9, 125.9, 125.3, 122.8, 102.1, 98.5, 55.2, 55.1, 52.4, 50.0, 36.9, 31.8, 27.7, 21.9, 21.5.

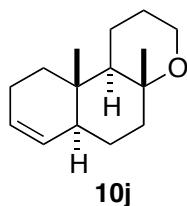
HRMS-ESI calculated for C₂₃H₂₇NNaO₄S [M+Na]⁺: 436.1553; found: 436.1558.



¹H NMR (400 MHz, CDCl₃) δ 7.27 - 7.20 (m, 2H), 6.66 (d, J = 8.4 Hz, 1H), 5.80 (m, 1H), 5.66 (m, 1H), 3.07 - 2.95 (m, 2H), 2.52 - 2.28 (m, 3H), 2.25 - 2.14 (m, 1H), 2.01 (dtd, J = 12.9, 6.3, 1.2 Hz, 1H), 1.81 (dtd, J = 12.9, 6.4, 1.4 Hz, 1H).

¹³C NMR (101 MHz, CDCl₃) δ 158.0, 130.7, 129.1, 128.1, 126.8, 123.8, 111.6, 111.1, 87.6, 40.8, 37.1, 32.4, 23.4.

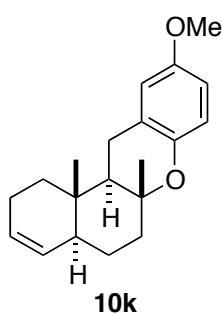
HRMS-APCI calculated for C₁₃H₁₄BrO [M+H]⁺: 265.0223; found: 265.0219.



¹H NMR (500 MHz, CDCl₃) δ 5.55 (dq, J = 9.9, 3.2 Hz, 1H), 5.35 (dq, J = 9.8, 2.1 Hz, 1H), 3.75 (ddd, J = 11.9, 9.6, 7.0 Hz, 1H), 3.70 - 3.63 (m, 1H), 2.09 (dh, J = 6.7, 3.2, 2.3 Hz, 2H), 2.00 (dddd, J = 14.1, 4.8, 3.3, 1.4 Hz, 1H), 1.81 - 1.64 (m, 5H), 1.59 - 1.38 (m, 4H), 1.36 - 1.26 (m, 4H), 1.22 - 1.13 (m, 1H), 0.66 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 130.3, 125.8, 75.0, 61.0, 55.1, 46.2, 41.2, 35.0, 34.4, 27.7, 25.6, 23.2, 20.7, 18.3, 11.6.

HRMS-APCI calculated for C₁₅H₂₅O [M+H]⁺: 221.1900; found: 221.1890.

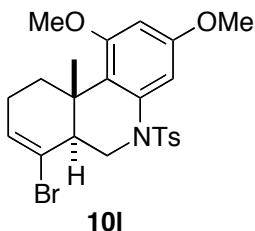


¹H NMR (500 MHz, CDCl₃) δ 6.74 - 6.67 (m, 2H), 6.65 (dd, J = 2.8, 1.0 Hz, 1H), 5.60 (dq, J = 9.9, 3.5 Hz, 1H), 5.42 (dq, J = 9.8, 2.0 Hz, 1H), 3.77 (s, 3H), 2.75 (dd, J = 16.4, 5.2 Hz, 1H), 2.70 - 2.63 (m, 1H), 2.17 - 2.11 (m, 2H), 2.08 (ddt, J = 12.6, 10.3, 3.1 Hz, 2H), 1.86 - 1.79 (m, 1H), 1.79 - 1.71 (m, 2H), 1.63 (ddt, J = 13.9, 4.3,

3.1 Hz, 1H), 1.48 (td, J = 13.4, 3.3 Hz, 1H), 1.34 - 1.30 (m, 1H), 1.29 (d, J = 1.0 Hz, 3H), 0.83 (d, J = 0.9 Hz, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 153.0, 147.4, 130.1, 125.8, 123.0, 117.6, 114.3, 113.1, 55.7, 49.4, 45.9, 40.4, 34.9, 34.8, 25.2, 23.1, 23.0, 21.4, 11.4.

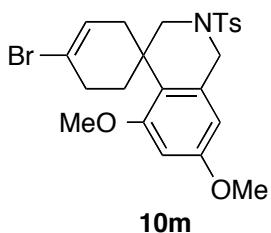
HRMS-ESI calculated for $\text{C}_{20}\text{H}_{26}\text{NaO}_2 [\text{M}+\text{Na}]^+$: 321.1825; found: 321.1810.



^1H NMR (500 MHz, CDCl_3) δ 7.63 - 7.59 (m, 2H), 7.27 - 7.23 (m, 2H), 7.22 (d, J = 2.5 Hz, 1H), 6.28 (d, J = 2.5 Hz, 1H), 6.11 (dt, J = 5.4, 3.0 Hz, 1H), 4.68 (dd, J = 13.0, 3.5 Hz, 1H), 3.83 (s, 3H), 3.76 (s, 3H), 3.27 (t, J = 12.8 Hz, 1H), 3.09 - 3.00 (m, 1H), 2.46 - 2.42 (m, 1H), 2.40 (s, 3H), 2.22 - 2.12 (m, 1H), 2.08 (dtdd, J = 13.8, 7.8, 3.9, 2.3 Hz, 1H), 1.18 (ddd, J = 13.5, 11.6, 6.6 Hz, 1H), 0.96 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 159.1, 158.4, 143.8, 137.8, 136.2, 130.6, 129.6, 127.3, 120.9, 119.0, 100.7, 96.6, 55.4, 55.3, 47.1, 46.2, 38.1, 31.3, 25.8, 21.6, 17.2.

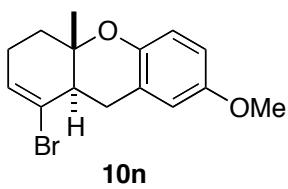
HRMS-ESI calculated for $\text{C}_{23}\text{H}_{26}\text{BrNNaO}_4\text{S} [\text{M}+\text{Na}]^+$: 514.0658; found: 514.0651.



^1H NMR (500 MHz, CDCl_3) δ 7.78 - 7.74 (m, 2H), 7.41 - 7.37 (m, 2H), 6.35 (d, J = 2.5 Hz, 1H), 6.18 (d, J = 2.5 Hz, 1H), 6.13 - 6.09 (m, 1H), 4.18 (d, J = 14.5 Hz, 1H), 4.09 (d, J = 14.4 Hz, 1H), 3.81 (s, 3H), 3.77 (s, 3H), 3.51 (ddt, J = 18.0, 4.6, 2.5 Hz, 1H), 3.23 (d, J = 12.0 Hz, 1H), 2.98 (d, J = 11.9 Hz, 1H), 2.64 - 2.54 (m, 1H), 2.47 (s, 3H), 2.36 - 2.25 (m, 1H), 2.25 - 2.19 (m, 1H), 2.19 - 2.11 (m, 1H), 1.61 - 1.57 (m, 1H).

^{13}C NMR (126 MHz, CDCl_3) δ 159.4, 159.0, 143.8, 134.4, 133.0, 129.8, 127.8, 127.0, 121.0, 120.8, 102.2, 98.4, 55.3, 55.2, 52.2, 49.8, 41.4, 39.6, 26.3, 24.0, 21.6.

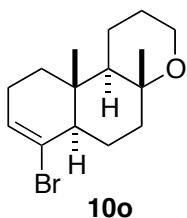
HRMS-ESI calculated for $\text{C}_{23}\text{H}_{26}\text{BrNNaO}_4\text{S} [\text{M}+\text{Na}]^+$: 514.0658; found: 514.0664.



¹H NMR (500 MHz, CDCl₃) δ 6.78 (d, J = 8.8 Hz, 1H), 6.74 (dd, J = 9.0, 3.0 Hz, 1H), 6.71 - 6.69 (m, 1H), 6.12 (dt, J = 5.0, 2.9 Hz, 1H), 3.79 (s, 3H), 3.07 (dd, J = 16.3, 5.3 Hz, 1H), 2.90 - 2.82 (m, 1H), 2.61 (ddt, J = 16.2, 13.6, 1.0 Hz, 1H), 2.37 - 2.18 (m, 2H), 2.01 (m, 1H), 1.93 (m, 1H), 1.17 (s, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 153.3, 147.1, 129.1, 123.7, 122.5, 117.8, 114.2, 114.0, 76.0, 55.7, 45.0, 34.8, 28.9, 25.7, 16.3.

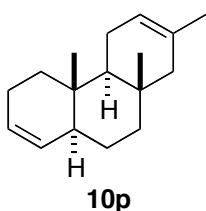
HRMS-ESI calculated for C₁₅H₁₇BrNaO₂ [M+Na]⁺: 331.0311; found: 331.0314.



¹H NMR (500 MHz, CDCl₃) δ 6.03 (dt, J = 4.3, 3.3 Hz, 1H), 3.77 - 3.70 (m, 1H), 3.69 - 3.64 (m, 1H), 2.35 - 2.28 (m, 1H), 2.16 - 2.10 (m, 3H), 1.84 - 1.75 (m, 2H), 1.70 - 1.66 (m, 2H), 1.46 - 1.41 (m, 2H), 1.37 - 1.33 (m, 1H), 1.34 (d, J = 0.9 Hz, 3H), 1.30 - 1.27 (m, 1H), 1.24 - 1.21 (m, 1H), 0.75 (d, J = 0.9 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 128.2, 126.9, 74.5, 60.9, 54.8, 51.2, 41.0, 37.9, 33.8, 27.5, 25.3, 25.1, 20.5, 18.6, 11.9.

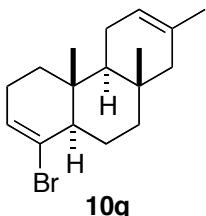
HRMS-ESI calculated for C₁₅H₂₃BrNaO [M+Na]⁺: 321.0832; found: 321.0838.



¹H NMR (500 MHz, CDCl₃) δ 5.55 (ddt, J = 10.0, 4.3, 3.2 Hz, 1H), 5.38 (ddt, J = 9.8, 4.2, 1.8 Hz, 2H), 2.08 (dddd, J = 10.2, 7.1, 4.2, 2.1 Hz, 3H), 1.90 (ddddd, J = 15.3, 13.9, 6.4, 3.5, 2.0 Hz, 3H), 1.78 - 1.71 (m, 1H), 1.67 - 1.60 (m, 4H), 1.58 - 1.54 (m, 1H), 1.51 (dd, J = 13.1, 3.0 Hz, 1H), 1.41 - 1.34 (m, 1H), 1.31 - 1.25 (m, 1H), 1.22 - 1.16 (m, 1H), 1.16 - 1.10 (m, 1H), 0.96 (s, 3H), 0.81 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 132.2, 131.4, 125.4, 120.2, 51.6, 49.9, 46.8, 42.4, 35.3, 35.0, 33.6, 24.7, 23.7, 23.4, 22.9, 20.9, 11.9.

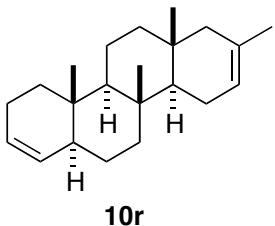
HRMS-APCI calculated for $\text{C}_{17}\text{H}_{27}$ $[\text{M}+\text{H}]^+$: 231.2113; found: 231.2111.



^1H NMR (500 MHz, CDCl_3) δ 6.06 - 6.01 (m, 1H), 5.38 - 5.34 (m, 1H), 2.21 - 2.16 (m, 1H), 2.14 - 2.04 (m, 4H), 2.01 (m, 1H), 1.90 (m, 1H), 1.82 - 1.76 (m, 1H), 1.71 (dt, $J = 13.3, 3.2$ Hz, 1H), 1.64 (s, 3H), 1.56 (m, 1H), 1.47 (dd, $J = 13.1, 3.1$ Hz, 1H), 1.21 (m, 3H), 0.96 (s, 3H), 0.89 (s, 3H).

^{13}C NMR (126 MHz, CDCl_3) δ 132.3, 128.3, 128.0, 119.9, 52.1, 51.4, 49.9, 42.1, 38.1, 34.6, 33.3, 25.2, 24.0, 23.7, 23.2, 20.8, 12.3.

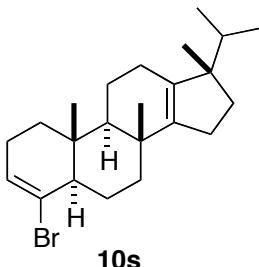
HRMS-APCI calculated for $\text{C}_{17}\text{H}_{26}\text{Br}$ $[\text{M}+\text{H}]^+$: 309.1212; found: 309.1214.



^1H NMR (400 MHz, CDCl_3) δ 5.54 (m, 1H), 5.34 (m, 2H), 2.09 (m, 2H), 2.02 - 1.74 (m, 6H), 1.73 - 1.61 (m, 6H), 1.51 - 1.32 (m, 4H), 1.26 - 1.16 (m, 2H), 1.13 - 1.06 (m, 1H), 1.05 - 0.93 (m, 4H), 0.93 - 0.83 (m, 4H), 0.75 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 131.7, 131.2, 125.5, 119.9, 58.5, 53.5, 51.3, 46.4, 43.5, 40.6, 37.8, 35.5, 35.3, 33.2, 24.3, 23.7, 23.6, 22.7, 20.4, 17.8, 17.0, 12.3.

HRMS-APCI calculated for $\text{C}_{22}\text{H}_{35}$ $[\text{M}+\text{H}]^+$: 299.2733; found: 299.2730.

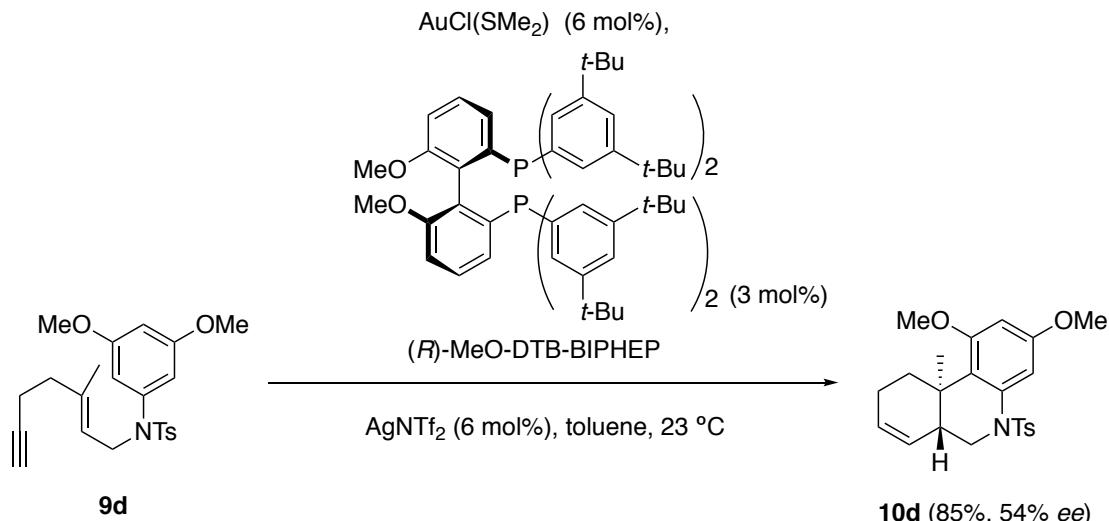


¹H NMR (500 MHz, CDCl₃) δ 6.03 (q, J = 3.6 Hz, 1H), 2.26 - 2.06 (m, 4H), 2.06 - 1.97 (m, 2H), 1.93 - 1.87 (m, 1H), 1.84 - 1.75 (m, 4H), 1.60 (m, 1H), 1.52 - 1.44 (m, 2H), 1.31 - 1.26 (m, 2H), 1.25 - 1.20 (m, 2H), 1.14 (m, 1H), 1.03 (s, 3H), 0.97 (s, 3H), 0.86 (s, 3H), 0.85 (d, J = 7.1 Hz, 3H), 0.73 (d, J = 6.8 Hz, 3H).

¹³C NMR (126 MHz, CDCl₃) δ 143.6, 137.6, 128.0, 127.9, 53.8, 52.3, 51.7, 38.5, 38.0, 36.3, 34.6, 33.5, 30.2, 27.7, 25.3, 25.3, 24.3, 23.4, 20.8, 18.4, 18.2, 17.6, 12.5.

HRMS-ESI calculated for C₂₃H₃₅BrNa [M+Na]⁺: 413.1822; found: 413.1826.

4. Procedures for Enantioselective Cyclisations



Chloro(dimethylsulfide)gold(I) (1.8 mg, 6 μmol) was added to a solution of (R)-MeO-DTBM-BIPHEP (3.0 mg, 3 μmol) in toluene (0.5 mL) and the mixture was stirred at 23 °C for 30 min before silver bis(trifluoromethanesulfonyl)imide (2.3 mg, 6 μmol) was added. The mixture was stirred at this temperature for another 30 min and a solution of **9d** (41 mg, 0.1 mmol) in toluene (0.5 mL) was added. The mixture was stirred at 23 °C for 1 h before it was filtered through a pad of Celite and purified by preparative TLC. The purified product was analyzed on HPLC.

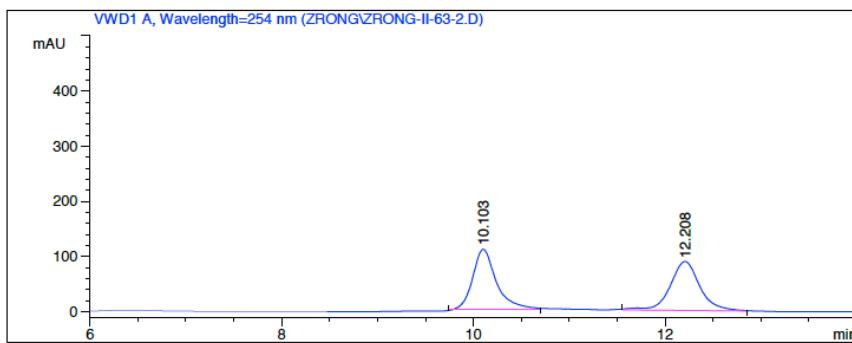
Identical procedure was applied for the reaction of **9f** to form **10f**.

HPLC analysis of racemic 10d.

Data File C:\HPCHEM\1\DATA\ZRONG\ZRONG-II-63-2.D
Sample Name: ZRONG-II-63-2

```
=====
Acq. Operator   : ZHOUTING
Acq. Instrument : HPLC1100                               Location : Vial 71
Injection Date  : 3/5/2015 3:33:19 PM
                                                Inj Volume : 5 μl
Acq. Method     : C:\HPCHEM\1\DATA\KATYA\MASHA.M
Last changed    : 3/5/2015 3:28:03 PM by ZHOUTING
                    (modified after loading)
Analysis Method : C:\HPCHEM\1\DATA\ZRONG\ZRONG-II-63-2.D\DA.M (MASHA.M)
Last changed    : 1/26/2017 4:54:10 PM by MASHA
Method Info     : STANDARD FLAVANONE IB

Sample Info      : chiralpack IA
                    95:5 hex:ipa
                    1.0 ml/min
```



```
=====
Area Percent Report
=====

Sorted By          : Signal
Calib. Data Modified : 10/28/2004 1:11:56 PM
Multiplier        : 1.0000
Dilution          : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: VWD1 A, Wavelength=254 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU	Area *s	Area %	Name
1	10.103	MM	0.2917	1922.37549	50.1159	?	
2	12.208	MM	0.3594	1913.48779	49.8841	?	

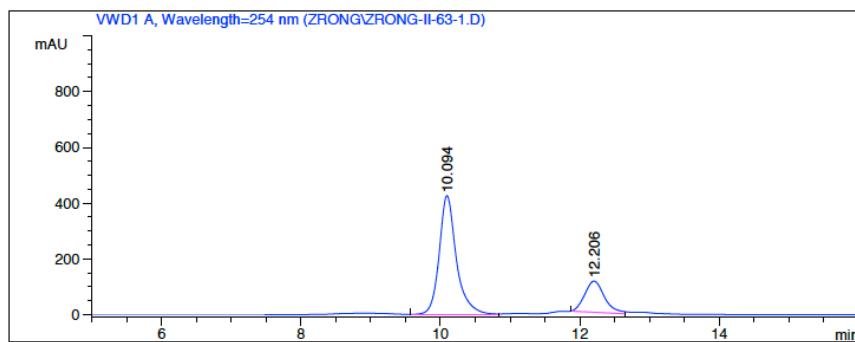
Totals : 3835.86328

HPLC analysis of enantioenriched **10d**.

Data File C:\HPCHEM\1\DATA\ZRONG\ZRONG-II-63-1.D
Sample Name: ZRONG-II-63-1

```
=====
Acq. Operator   : ZHOUTING
Acq. Instrument : HPLC1100                               Location : Vial 81
Injection Date  : 3/5/2015 3:54:37 PM
                                                Inj Volume : 5 µl
Acq. Method     : C:\HPCHEM\1\DATA\KATYA\MASHA.M
Last changed    : 3/5/2015 3:51:54 PM by ZHOUTING
                  (modified after loading)
Analysis Method : C:\HPCHEM\1\DATA\ZRONG\ZRONG-II-63-1.D\DA.M (MASHA.M)
Last changed    : 1/26/2017 4:17:38 PM by MASHA
                  (modified after loading)
Method Info     : STANDARD FLAVANONE IB

Sample Info      : chiralpack IA
                   95:5 hex:ipa
                   1.0 ml/min
```



```
=====
Area Percent Report
=====
```

```
Sorted By       : Signal
Calib. Data Modified : 10/28/2004 1:11:56 PM
Multiplier      : 1.0000
Dilution        : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: VWD1 A, Wavelength=254 nm

Peak #	RetTime [min]	Type	Width [min]	Area mAU	Area *s	Name %
1	10.094	VV	0.2607	7396.45166	76.9948	?
2	12.206	MM	0.3297	2209.97461	23.0052	?

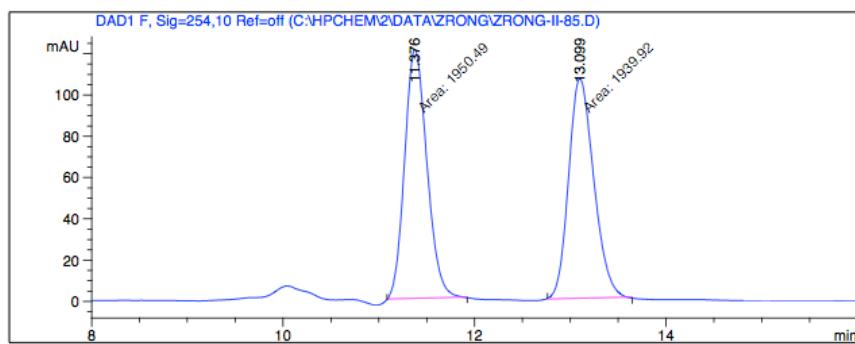
Totals : 9606.42627

HPLC analysis of racemic 10f.

Data File C:\HPCHEM\2\DATA\ZRONG\ZRONG-II-85.D
Sample Name: ZRONG-II-85

```
=====
Acq. Operator   : ZHOUTING
Acq. Instrument : AG1200HPLC          Location : Vial 41
Injection Date  : 4/16/2015 4:21:47 PM
                           Inj Volume : 5 μl
Acq. Method     : C:\HPCHEM\2\METHODS\MICHAEL.M
Last changed    : 4/16/2015 3:55:06 PM by ZHOUTING
                           (modified after loading)
Analysis Method : C:\HPCHEM\2\DATA\ZRONG\ZRONG-II-85.D\DA.M (MICHAEL.M)
Last changed    : 1/28/2017 2:11:19 PM by GZ
                           (modified after loading)
Method Info     : STANDARD FLAVANONE IA

Sample Info      : CHIRALCEL IA
                           90:10 Hex:IPA
                           1 ml/min
```



```
=====
Area Percent Report
=====
```

```
Sorted By           : Signal
Multiplier         : 1.0000
Dilution          : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: DAD1 F, Sig=254,10 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.376	MM	0.2696	1950.49011	120.55746	50.1358
2	13.099	MM	0.3038	1939.92041	106.42841	49.8642

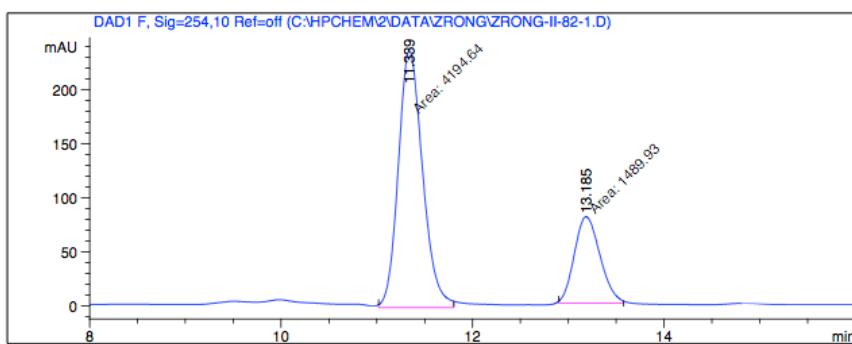
Totals : 3890.41052 226.98586

HPLC analysis of enantioenriched **10f**.

Data File C:\HPCHEM\2\DATA\ZRONG\ZRONG-II-82-1.D
Sample Name: ZRONG-II-82-1

```
=====
Acq. Operator   : Zhouting
Acq. Instrument : AG1200HPLC          Location : Vial 71
Injection Date  : 4/19/2015 5:07:38 PM
                                                Inj Volume : 5 μl
Acq. Method     : C:\HPCHEM\2\METHODS\JAVI.M
Last changed    : 4/19/2015 4:54:16 PM by Javi
                  (modified after loading)
Analysis Method : C:\HPCHEM\2\DATA\ZRONG\ZRONG-II-82-1.D\DA.M (JAVI.M)
Last changed    : 1/28/2017 2:18:42 PM by GZ
                  (modified after loading)
Method Info     : STANDARD FLAVANONE IA

Sample Info      : CHIRALCEL IA
                   90:10 Hex:IPA
                   1 ml/min
```



```
=====
Area Percent Report
=====

Sorted By       : Signal
Multiplier      : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
```

Signal 1: DAD1 F, Sig=254,10 Ref=off

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.339	MM	0.2942	4194.64258	237.66092	73.7900
2	13.185	MM	0.3105	1489.92896	79.96814	26.2100

Totals : 5684.57153 317.62906

5. Crystal Data

Compound 10f

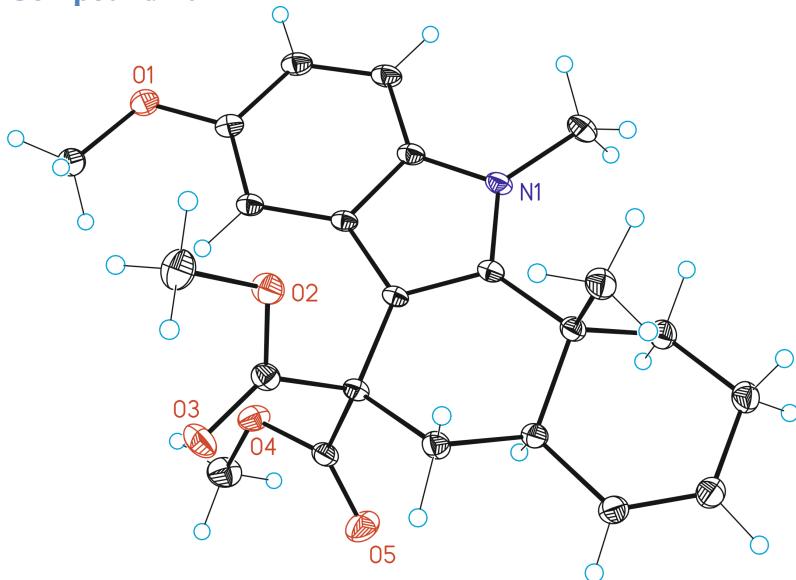


Table 1. Crystal data and structure refinement for mo_zrii7.

Identification code	mo_zrii7		
Empirical formula	C ₂₃ H ₂₇ N O ₅		
Formula weight	397.45		
Temperature	100(2) K		
Wavelength	0.71073 Å		
Crystal system	Monoclinic		
Space group	P2(1)/c		
Unit cell dimensions	a = 9.8767(14)Å	b = 23.638(3)Å	c = 8.6175(12)Å
	a= 90°.	b= 102.363(4)°.	g = 90°.
Volume	1965.2(5) Å ³		
Z	4		
Density (calculated)	1.343 Mg/m ³		
Absorption coefficient	0.094 mm ⁻¹		
F(000)	848		
Crystal size	0.20 x 0.08 x 0.04 mm ³		
Theta range for data collection	1.723 to 33.188°.		
Index ranges	-15<=h<=15,-36<=k<=36,-13<=l<=12		
Reflections collected	59094		
Independent reflections	7505[R(int) = 0.0271]		
Completeness to theta =33.188°	99.8%		
Absorption correction	Empirical		

Max. and min. transmission	0.996 and 0.766
Refinement method	Full-matrix least-squares on F^2
Data / restraints / parameters	7505/ 0/ 267
Goodness-of-fit on F^2	1.127
Final R indices [$I > 2\sigma(I)$]	$R_1 = 0.0482$, $wR_2 = 0.1255$
R indices (all data)	$R_1 = 0.0530$, $wR_2 = 0.1284$
Largest diff. peak and hole	0.629 and -0.338 e. \AA^{-3}

Table 2. Bond lengths [\AA] and angles [$^\circ$] for mo_zrii7.

Bond lengths----	
N1-C1	1.3850(13)
N1-C16	1.3877(12)
N1-C17	1.4571(13)
O1-C4	1.3748(12)
O1-C18	1.4229(13)
O2-C21	1.3338(12)
O2-C22	1.4460(13)
O3-C21	1.2098(12)
O4-C19	1.3390(12)
O4-C20	1.4461(13)
O5-C19	1.2037(12)
C1-C2	1.4012(13)
C1-C6	1.4110(13)
C2-C3	1.3786(15)
C3-C4	1.4110(14)
C4-C5	1.3862(13)
C5-C6	1.4121(13)
C6-C7	1.4361(12)
C7-C16	1.3841(13)
C7-C8	1.5127(12)
C8-C21	1.5279(13)
C8-C19	1.5390(13)
C8-C9	1.5490(13)
C9-C10	1.5252(14)
C10-C11	1.5066(14)
C10-C15	1.5491(13)
C11-C12	1.3343(14)

C12-C13	1.4994(15)
C13-C14	1.5340(15)
C14-C15	1.5469(14)
C15-C16	1.5104(13)
C15-C23	1.5485(14)

Angles-----

C1-N1-C16	108.44(8)
C1-N1-C17	122.56(8)
C16-N1-C17	129.00(9)
C4-O1-C18	116.95(8)
C21-O2-C22	114.88(8)
C19-O4-C20	115.16(8)
N1-C1-C2	129.34(9)
N1-C1-C6	108.72(8)
C2-C1-C6	121.93(9)
C3-C2-C1	117.57(9)
C2-C3-C4	121.34(9)
O1-C4-C5	124.60(9)
O1-C4-C3	113.94(8)
C5-C4-C3	121.46(9)
C4-C5-C6	117.99(8)
C1-C6-C5	119.67(8)
C1-C6-C7	106.16(8)
C5-C6-C7	134.15(8)
C16-C7-C6	107.67(8)
C16-C7-C8	123.98(8)
C6-C7-C8	128.29(8)
C7-C8-C21	113.76(7)
C7-C8-C19	110.59(7)
C21-C8-C19	107.39(7)
C7-C8-C9	110.43(7)
C21-C8-C9	105.34(7)
C19-C8-C9	109.10(7)
C10-C9-C8	110.29(8)
C11-C10-C9	113.31(8)
C11-C10-C15	112.27(8)
C9-C10-C15	112.36(8)

C12-C11-C10	122.24(9)
C11-C12-C13	122.75(9)
C12-C13-C14	113.29(8)
C13-C14-C15	111.47(8)
C16-C15-C14	115.17(8)
C16-C15-C23	107.98(8)
C14-C15-C23	110.55(8)
C16-C15-C10	105.61(7)
C14-C15-C10	105.58(7)
C23-C15-C10	111.89(8)
C7-C16-N1	108.99(8)
C7-C16-C15	124.70(8)
N1-C16-C15	125.98(8)
O5-C19-O4	123.32(9)
O5-C19-C8	124.67(9)
O4-C19-C8	112.00(8)
O3-C21-O2	124.11(9)
O3-C21-C8	123.35(9)
O2-C21-C8	112.39(8)

Table 3. Torsion angles [°] for mo_zrii7.

C16-N1-C1-C2	-179.11(10)
C17-N1-C1-C2	1.55(16)
C16-N1-C1-C6	1.19(10)
C17-N1-C1-C6	-178.15(9)
N1-C1-C2-C3	-178.90(9)
C6-C1-C2-C3	0.77(14)
C1-C2-C3-C4	0.88(15)
C18-O1-C4-C5	0.30(14)
C18-O1-C4-C3	-179.68(9)
C2-C3-C4-O1	178.84(9)
C2-C3-C4-C5	-1.14(15)
O1-C4-C5-C6	179.75(9)
C3-C4-C5-C6	-0.27(14)
N1-C1-C6-C5	177.55(8)
C2-C1-C6-C5	-2.18(14)

N1-C1-C6-C7	-1.04(10)
C2-C1-C6-C7	179.23(9)
C4-C5-C6-C1	1.87(13)
C4-C5-C6-C7	179.99(9)
C1-C6-C7-C16	0.51(10)
C5-C6-C7-C16	-177.79(10)
C1-C6-C7-C8	177.88(9)
C5-C6-C7-C8	-0.42(17)
C16-C7-C8-C21	129.70(9)
C6-C7-C8-C21	-47.28(12)
C16-C7-C8-C19	-109.36(10)
C6-C7-C8-C19	73.66(11)
C16-C7-C8-C9	11.50(12)
C6-C7-C8-C9	-165.47(9)
C7-C8-C9-C10	-40.96(10)
C21-C8-C9-C10	-164.20(8)
C19-C8-C9-C10	80.78(9)
C8-C9-C10-C11	-164.69(8)
C8-C9-C10-C15	66.75(10)
C9-C10-C11-C12	-155.31(10)
C15-C10-C11-C12	-26.70(13)
C10-C11-C12-C13	4.21(16)
C11-C12-C13-C14	-12.11(14)
C12-C13-C14-C15	42.71(11)
C13-C14-C15-C16	-179.11(8)
C13-C14-C15-C23	58.16(10)
C13-C14-C15-C10	-63.03(10)
C11-C10-C15-C16	176.39(8)
C9-C10-C15-C16	-54.50(10)
C11-C10-C15-C14	53.95(10)
C9-C10-C15-C14	-176.94(8)
C11-C10-C15-C23	-66.37(10)
C9-C10-C15-C23	62.74(10)
C6-C7-C16-N1	0.21(10)
C8-C7-C16-N1	-177.31(8)
C6-C7-C16-C15	174.01(8)
C8-C7-C16-C15	-3.50(14)
C1-N1-C16-C7	-0.86(11)

C17-N1-C16-C7	178.42(10)
C1-N1-C16-C15	-174.57(8)
C17-N1-C16-C15	4.72(16)
C14-C15-C16-C7	139.83(9)
C23-C15-C16-C7	-96.09(10)
C10-C15-C16-C7	23.75(12)
C14-C15-C16-N1	-47.42(13)
C23-C15-C16-N1	76.66(11)
C10-C15-C16-N1	-163.49(9)
C20-O4-C19-O5	2.32(15)
C20-O4-C19-C8	-178.55(9)
C7-C8-C19-O5	112.58(11)
C21-C8-C19-O5	-122.77(11)
C9-C8-C19-O5	-9.07(14)
C7-C8-C19-O4	-66.53(10)
C21-C8-C19-O4	58.12(10)
C9-C8-C19-O4	171.82(8)
C22-O2-C21-O3	-2.27(14)
C22-O2-C21-C8	-177.90(8)
C7-C8-C21-O3	152.50(9)
C19-C8-C21-O3	29.79(12)
C9-C8-C21-O3	-86.41(11)
C7-C8-C21-O2	-31.83(11)
C19-C8-C21-O2	-154.55(8)
C9-C8-C21-O2	89.26(9)

Compound 10h

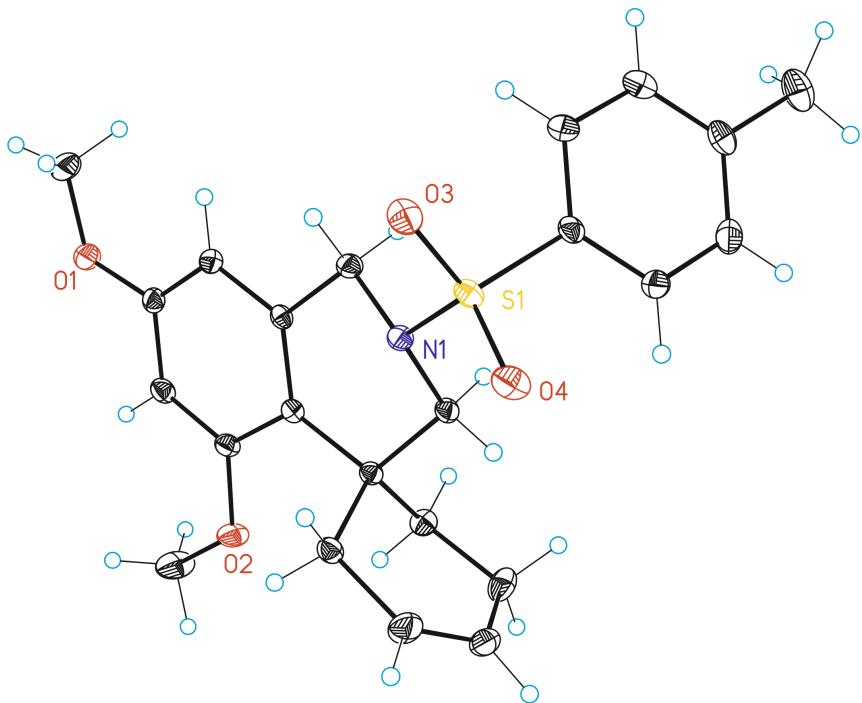


Table 1. Crystal data and structure refinement for mo_zri185_0m.

Identification code	mo_zri185_0m				
Empirical formula	C23 H27 N O4 S				
Formula weight	413.51				
Temperature	100(2) K				
Wavelength	0.71073 Å				
Crystal system	Monoclinic				
Space group	P2(1)/c				
Unit cell dimensions	a = 9.4223(6)Å	$\alpha = 90^\circ$.			
	b = 20.6490(13)Å	$\beta =$			
	105.9641(19)°.	$c = 10.9082(8)$ Å			
		$\gamma = 90^\circ$.			
Volume	2040.5(2) Å ³				
Z	4				
Density (calculated)	1.346 Mg/m ³				
Absorption coefficient	0.189 mm ⁻¹				
F(000)	880				
Crystal size	0.35 x 0.35 x 0.30 mm ³				
Theta range for data collection	1.972 to 32.427°.				
Index ranges	-13<=h<=8, -15<=k<=31, -14<=l<=16				
Reflections collected	14366				
Independent reflections	6481 [R(int) = 0.0237]				

Completeness to theta =32.427°	88.0%
Absorption correction	Empirical
Max. and min. transmission	0.946 and 0.885
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	6481/ 0/ 269
Goodness-of-fit on F ²	1.046
Final R indices [I>2sigma(I)]	R1 = 0.0426, wR2 = 0.1059
R indices (all data)	R1 = 0.0510, wR2 = 0.1111
Largest diff. peak and hole	0.428 and -0.493 e.Å ⁻³

Table 2. Bond lengths [Å] and angles [°] for mo_zri185_0m.

Bond lengths----

S1-O4	1.4318(10)
S1-O3	1.4357(9)
S1-N1	1.6377(10)
S1-C17	1.7598(12)
N1-C8	1.4650(15)
N1-C7	1.4685(14)
O1-C11	1.3588(14)
O1-C15	1.4224(16)
O2-C13	1.3713(14)
O2-C16	1.4245(15)
C1-C14	1.5264(15)
C1-C7	1.5397(15)
C1-C2	1.5453(16)
C1-C6	1.5453(16)
C2-C3	1.5007(18)
C3-C4	1.3664(19)
C4-C5	1.4554(19)
C5-C6	1.5166(17)
C8-C9	1.5117(15)
C9-C14	1.3960(15)
C9-C10	1.3994(16)
C10-C11	1.3832(16)
C11-C12	1.3965(16)
C12-C13	1.3802(16)
C13-C14	1.4213(15)

C17-C18	1.3919(17)
C17-C22	1.3938(16)
C18-C19	1.3875(18)
C19-C20	1.3952(19)
C20-C21	1.3936(19)
C20-C23	1.5065(19)
C21-C22	1.3859(18)

Angles-----

O4-S1-O3	119.61(6)
O4-S1-N1	106.97(5)
O3-S1-N1	106.90(5)
O4-S1-C17	107.64(6)
O3-S1-C17	108.29(6)
N1-S1-C17	106.78(5)
C8-N1-C7	110.10(9)
C8-N1-S1	116.60(7)
C7-N1-S1	116.84(8)
C11-O1-C15	116.82(10)
C13-O2-C16	117.33(10)
C14-C1-C7	109.36(9)
C14-C1-C2	110.41(9)
C7-C1-C2	109.00(9)
C14-C1-C6	112.69(9)
C7-C1-C6	105.86(9)
C2-C1-C6	109.37(9)
C3-C2-C1	112.93(10)
C4-C3-C2	121.84(12)
C3-C4-C5	123.30(12)
C4-C5-C6	115.43(11)
C5-C6-C1	111.92(10)
N1-C7-C1	110.68(9)
N1-C8-C9	109.63(9)
C14-C9-C10	122.90(10)
C14-C9-C8	121.78(10)
C10-C9-C8	115.32(9)
C11-C10-C9	118.99(10)
O1-C11-C10	125.26(11)

O1-C11-C12	114.52(10)
C10-C11-C12	120.21(11)
C13-C12-C11	119.91(10)
O2-C13-C12	121.53(10)
O2-C13-C14	116.58(10)
C12-C13-C14	121.88(10)
C9-C14-C13	115.99(10)
C9-C14-C1	121.90(10)
C13-C14-C1	122.07(10)
C18-C17-C22	120.40(11)
C18-C17-S1	119.29(9)
C22-C17-S1	120.25(9)
C19-C18-C17	119.39(11)
C18-C19-C20	121.20(12)
C21-C20-C19	118.34(12)
C21-C20-C23	120.33(12)
C19-C20-C23	121.33(13)
C22-C21-C20	121.40(12)
C21-C22-C17	119.26(12)

Table 3. Torsion angles [°] for mo_zri185_0m.

O4-S1-N1-C8	176.01(8)
O3-S1-N1-C8	46.78(10)
C17-S1-N1-C8	-68.96(9)
O4-S1-N1-C7	-50.82(10)
O3-S1-N1-C7	179.95(8)
C17-S1-N1-C7	64.21(9)
C14-C1-C2-C3	171.69(10)
C7-C1-C2-C3	-68.16(12)
C6-C1-C2-C3	47.14(13)
C1-C2-C3-C4	-20.97(18)
C2-C3-C4-C5	2.2(2)
C3-C4-C5-C6	-11.62(18)
C4-C5-C6-C1	39.40(15)
C14-C1-C6-C5	179.88(10)
C7-C1-C6-C5	60.39(12)

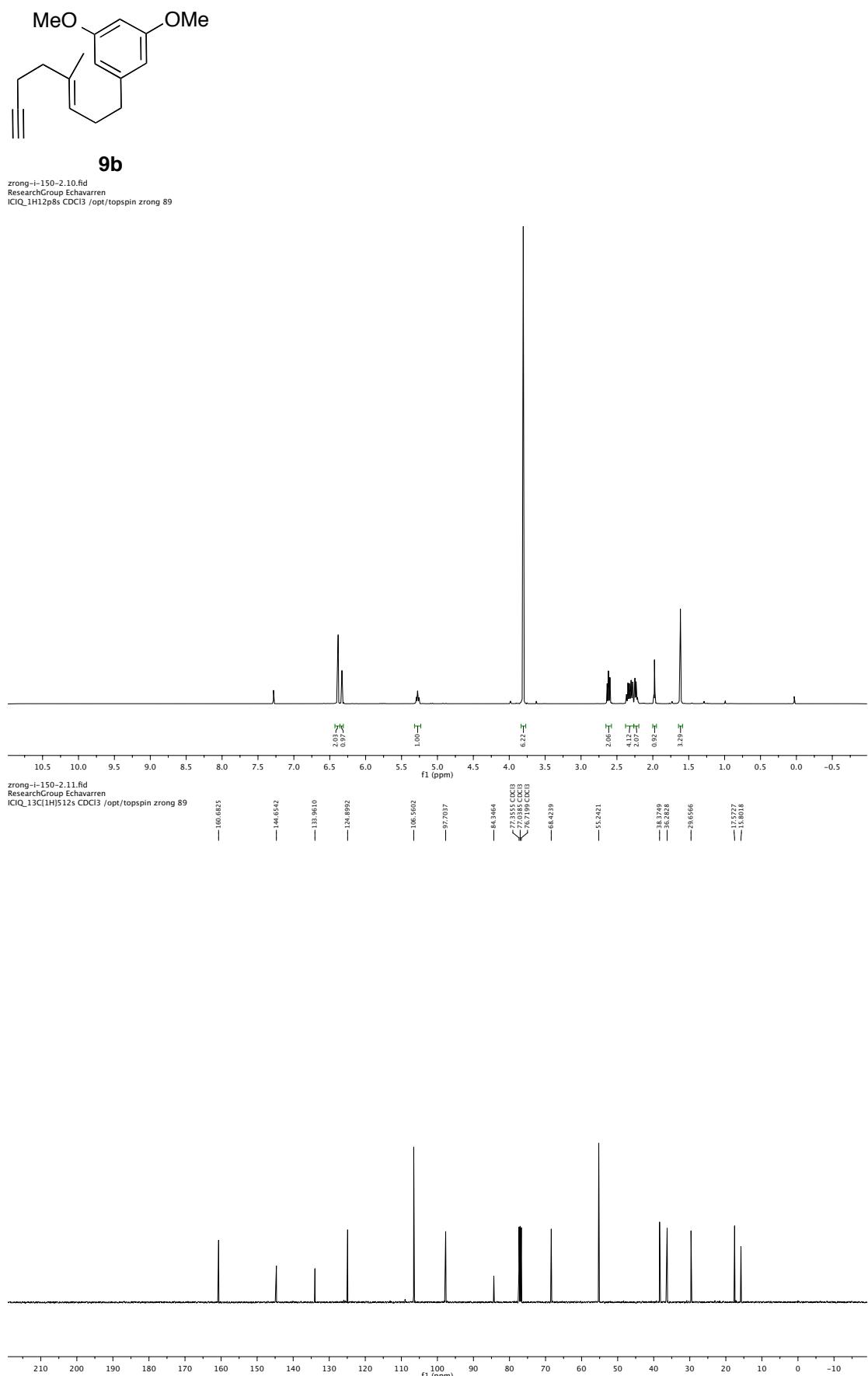
C2-C1-C6-C5	-56.91(13)
C8-N1-C7-C1	-70.83(12)
S1-N1-C7-C1	153.15(8)
C14-C1-C7-N1	45.99(12)
C2-C1-C7-N1	-74.81(11)
C6-C1-C7-N1	167.64(9)
C7-N1-C8-C9	55.82(12)
S1-N1-C8-C9	-168.05(8)
N1-C8-C9-C14	-22.40(15)
N1-C8-C9-C10	158.67(10)
C14-C9-C10-C11	-1.13(17)
C8-C9-C10-C11	177.78(10)
C15-O1-C11-C10	-6.79(18)
C15-O1-C11-C12	171.95(12)
C9-C10-C11-O1	179.97(11)
C9-C10-C11-C12	1.29(17)
O1-C11-C12-C13	-177.64(11)
C10-C11-C12-C13	1.17(18)
C16-O2-C13-C12	5.34(18)
C16-O2-C13-C14	-176.30(12)
C11-C12-C13-O2	174.37(11)
C11-C12-C13-C14	-3.91(18)
C10-C9-C14-C13	-1.41(17)
C8-C9-C14-C13	179.74(10)
C10-C9-C14-C1	-179.23(10)
C8-C9-C14-C1	1.93(17)
O2-C13-C14-C9	-174.41(10)
C12-C13-C14-C9	3.95(17)
O2-C13-C14-C1	3.40(17)
C12-C13-C14-C1	-178.24(11)
C7-C1-C14-C9	-13.22(15)
C2-C1-C14-C9	106.71(12)
C6-C1-C14-C9	-130.66(11)
C7-C1-C14-C13	169.10(10)
C2-C1-C14-C13	-70.97(13)
C6-C1-C14-C13	51.66(14)
O4-S1-C17-C18	28.37(11)
O3-S1-C17-C18	159.00(10)

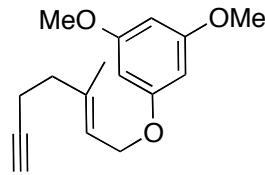
N1-S1-C17-C18	-86.20(10)
O4-S1-C17-C22	-154.36(10)
O3-S1-C17-C22	-23.73(11)
N1-S1-C17-C22	91.07(10)
C22-C17-C18-C19	0.08(18)
S1-C17-C18-C19	177.35(10)
C17-C18-C19-C20	0.46(19)
C18-C19-C20-C21	-0.25(19)
C18-C19-C20-C23	-179.94(12)
C19-C20-C21-C22	-0.51(19)
C23-C20-C21-C22	179.18(12)
C20-C21-C22-C17	1.04(19)
C18-C17-C22-C21	-0.82(18)
S1-C17-C22-C21	-178.06(9)

6. References

- (1) Surendra, K.; Rajendar, G.; Corey, E. J. *J. Am. Chem. Soc.* **2014**, *136*, 642–645.
- (2) Liu, Y.; Ma, S. *Org. Lett.* **2012**, *14*, 720–723.
- (3) Johansen, M. B.; Kerr, M. A. *Org. Lett.* **2010**, *12*, 4956–4959.
- (4) Huang, J.; Wu, C.; Wulff, W. D. *J. Am. Chem. Soc.* **2007**, *129*, 13366–13367.
- (5) Suhara, Y.; Hirota, Y.; Nakagawa, K.; Kamao, M.; Tsugawa, N.; Okano, T. *Bioorg. Med. Chem.* **2008**, *16*, 3108–3117.
- (6) Wu, B.; Woodward, R.; Wen, L.; Wang, X.; Zhao, G.; Wang, P. G. *Eur. J. Org. Chem.* **2013**, 8162–8173.

7. ^1H NMR and ^{13}C NMR Spectra

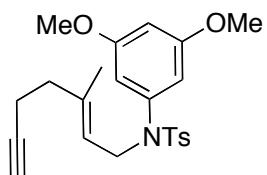




9c

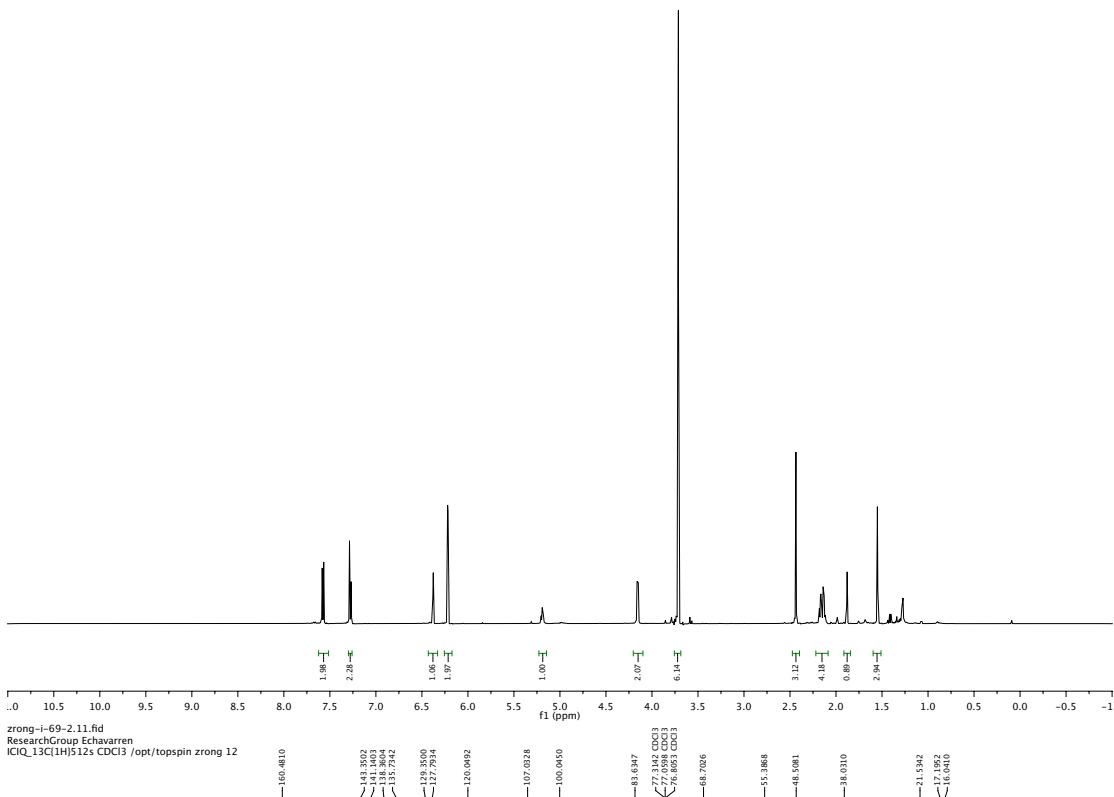
zong-i-69-1.11.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zong 11



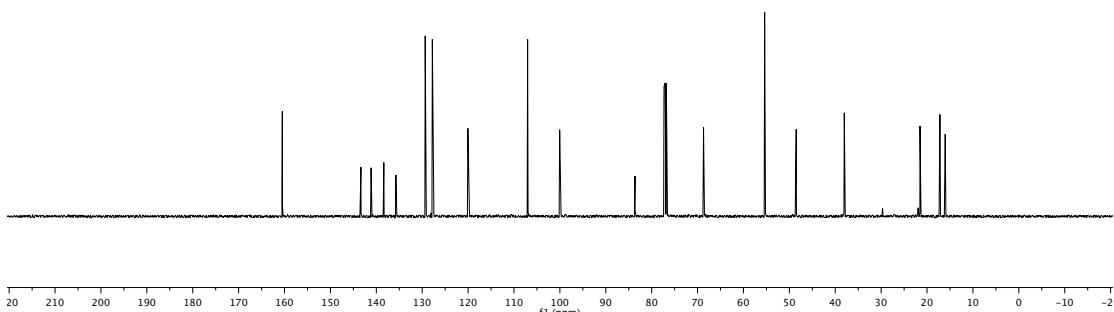


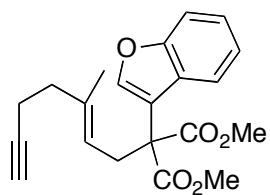
9d

zrong-i-69-2.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 12



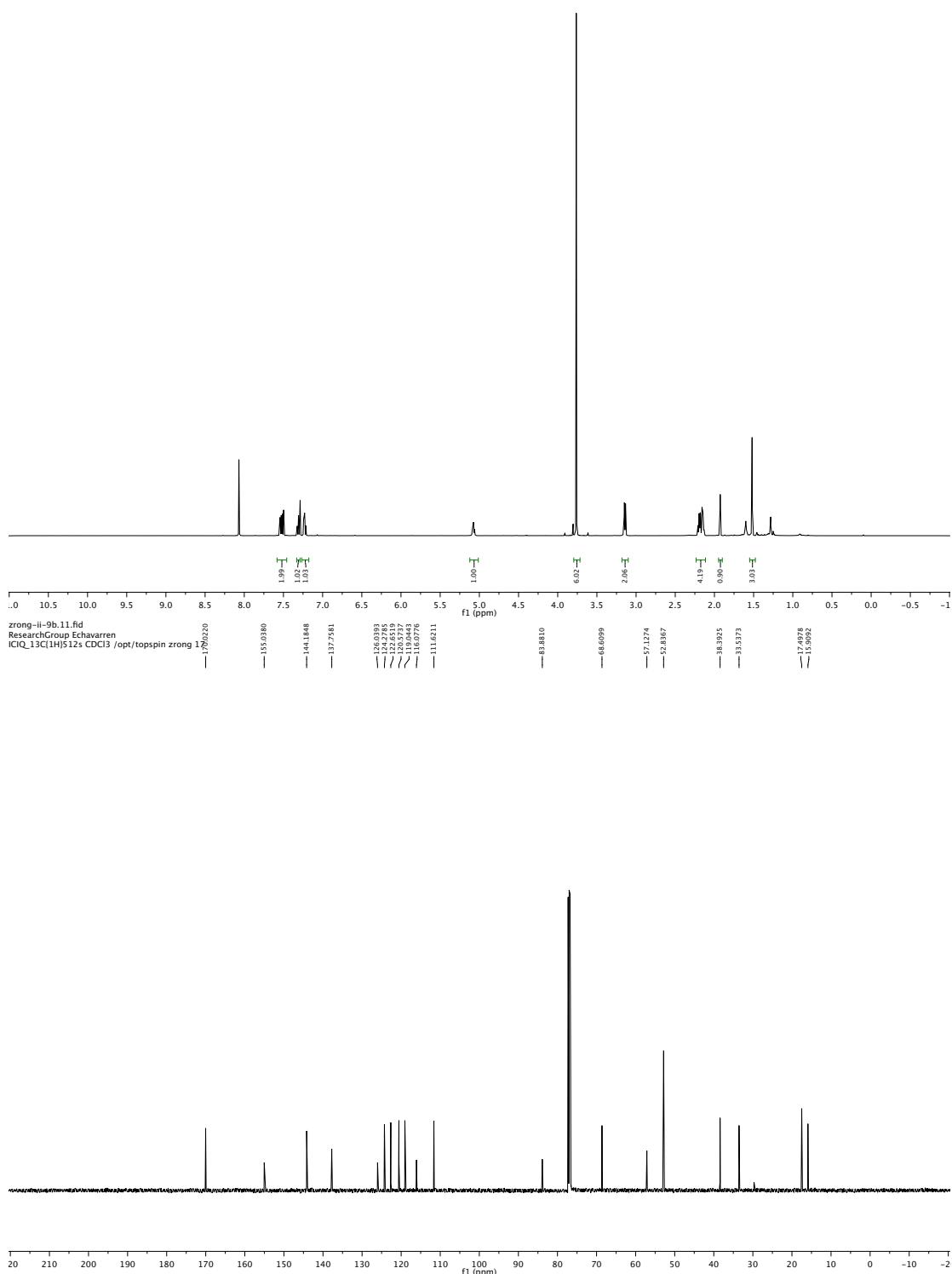
zrong-i-69-2.11.fid
ResearchGroup Echavarren
ICIQ_13C[1H]512s CDCl₃ /opt/topspin zrong 12

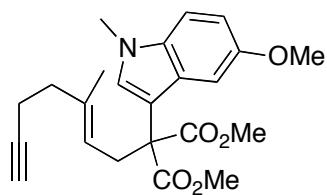




9e

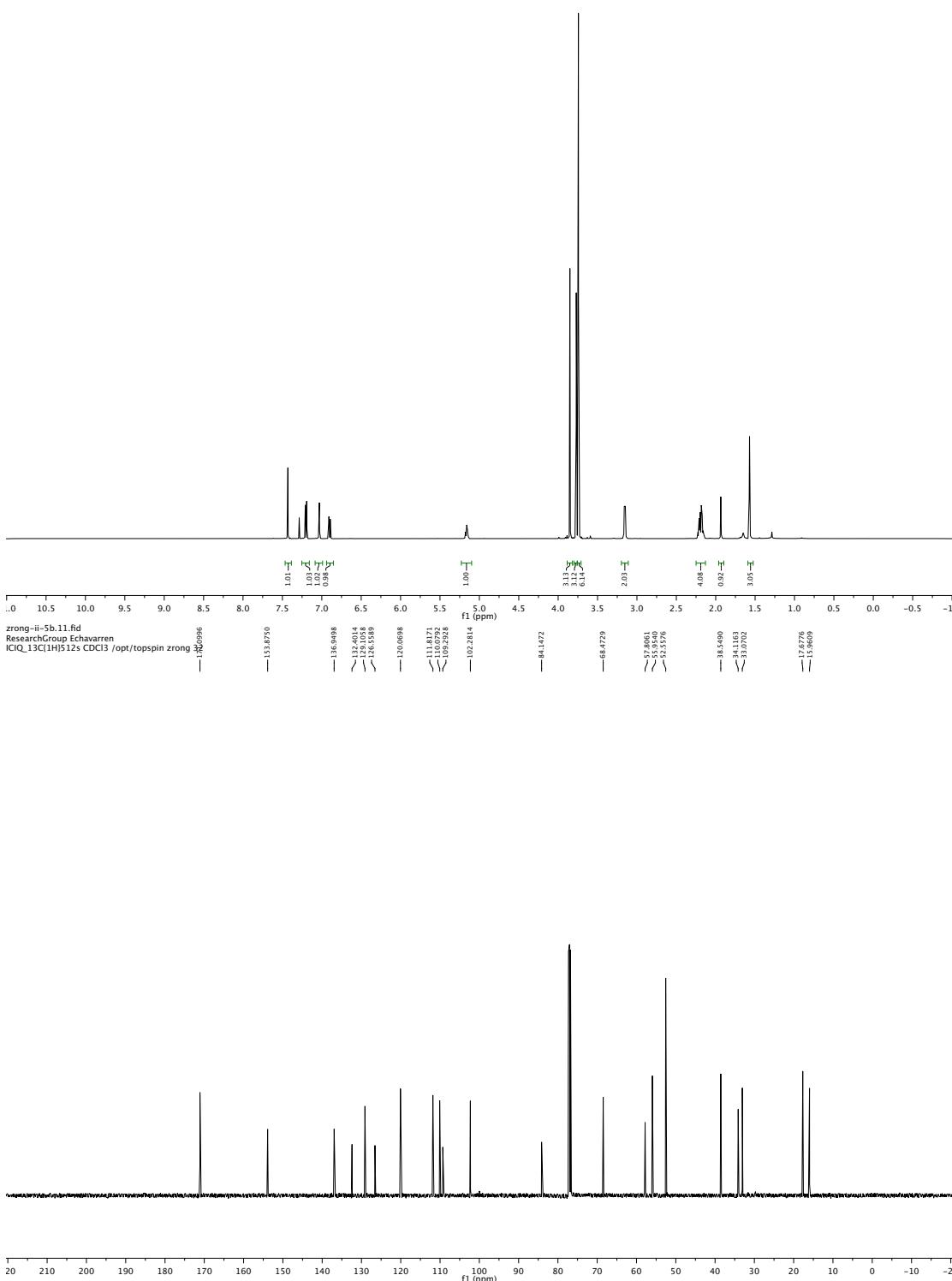
zrong-ii-9b.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 17

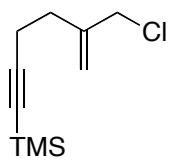




9f

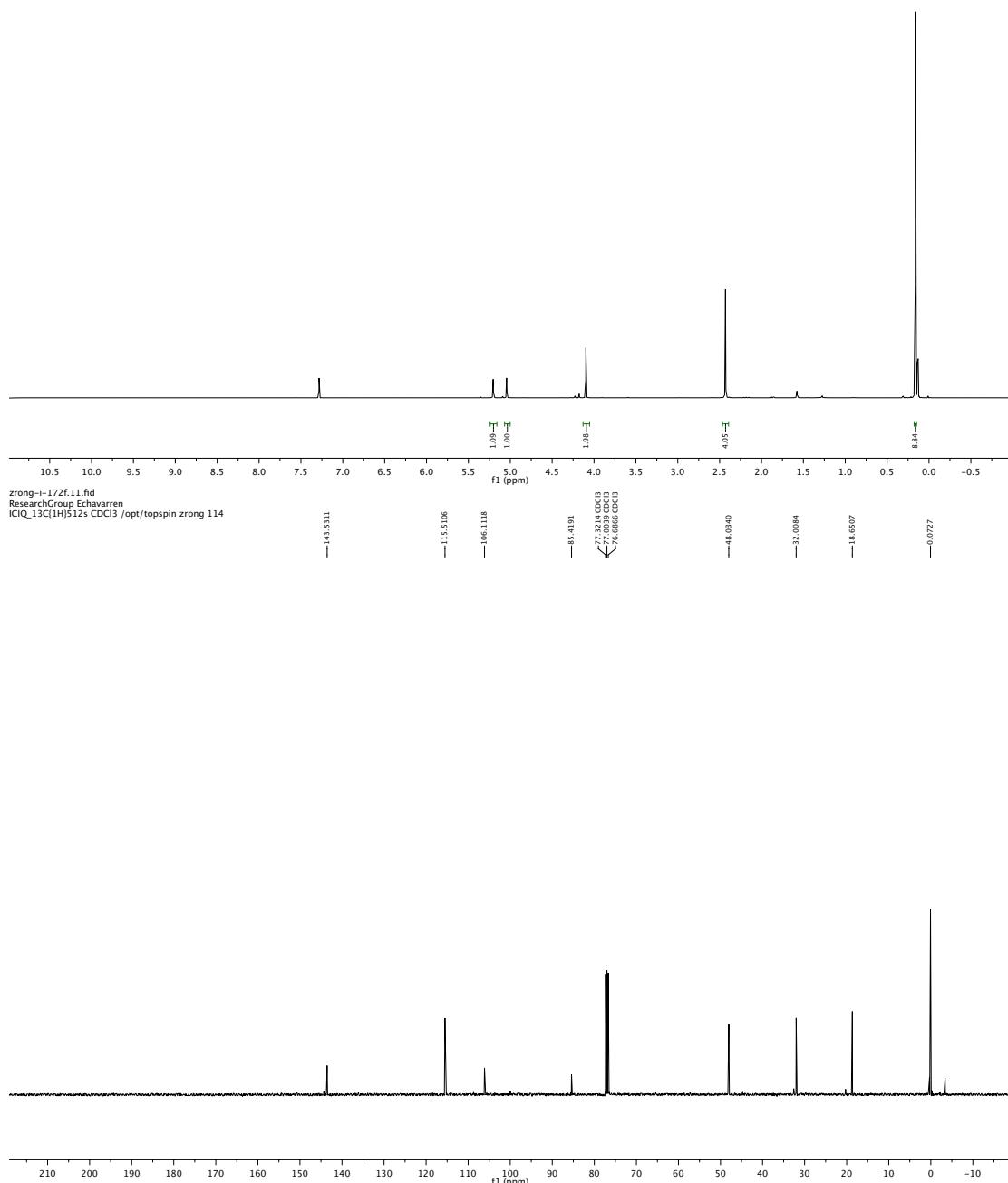
zrong-ii-5b.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 32

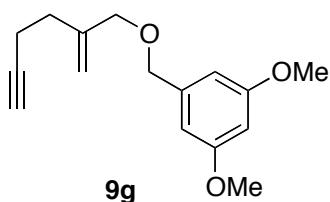




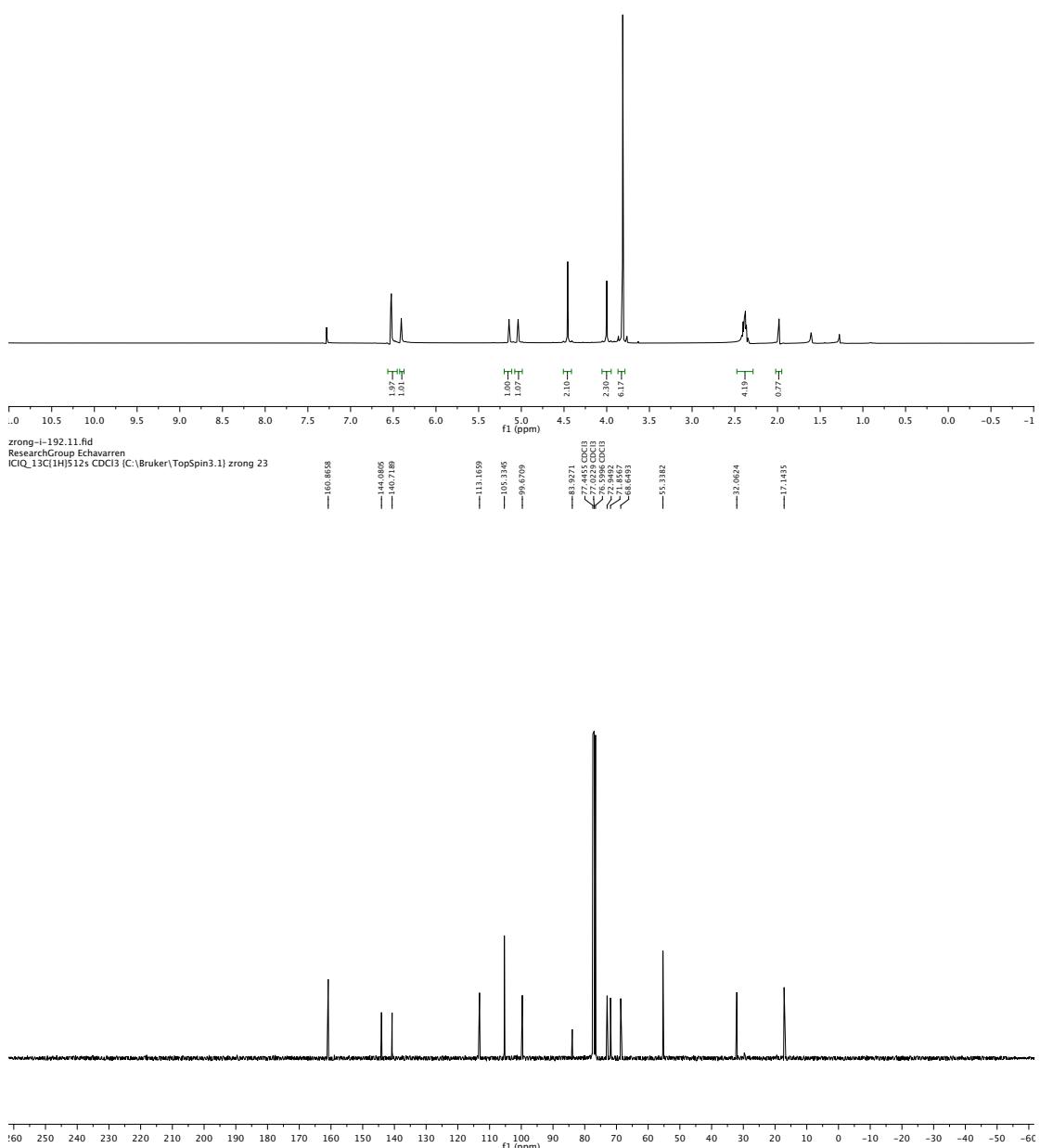
S6

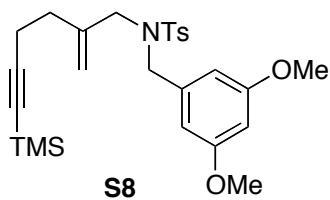
zrong-i-172f.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 114



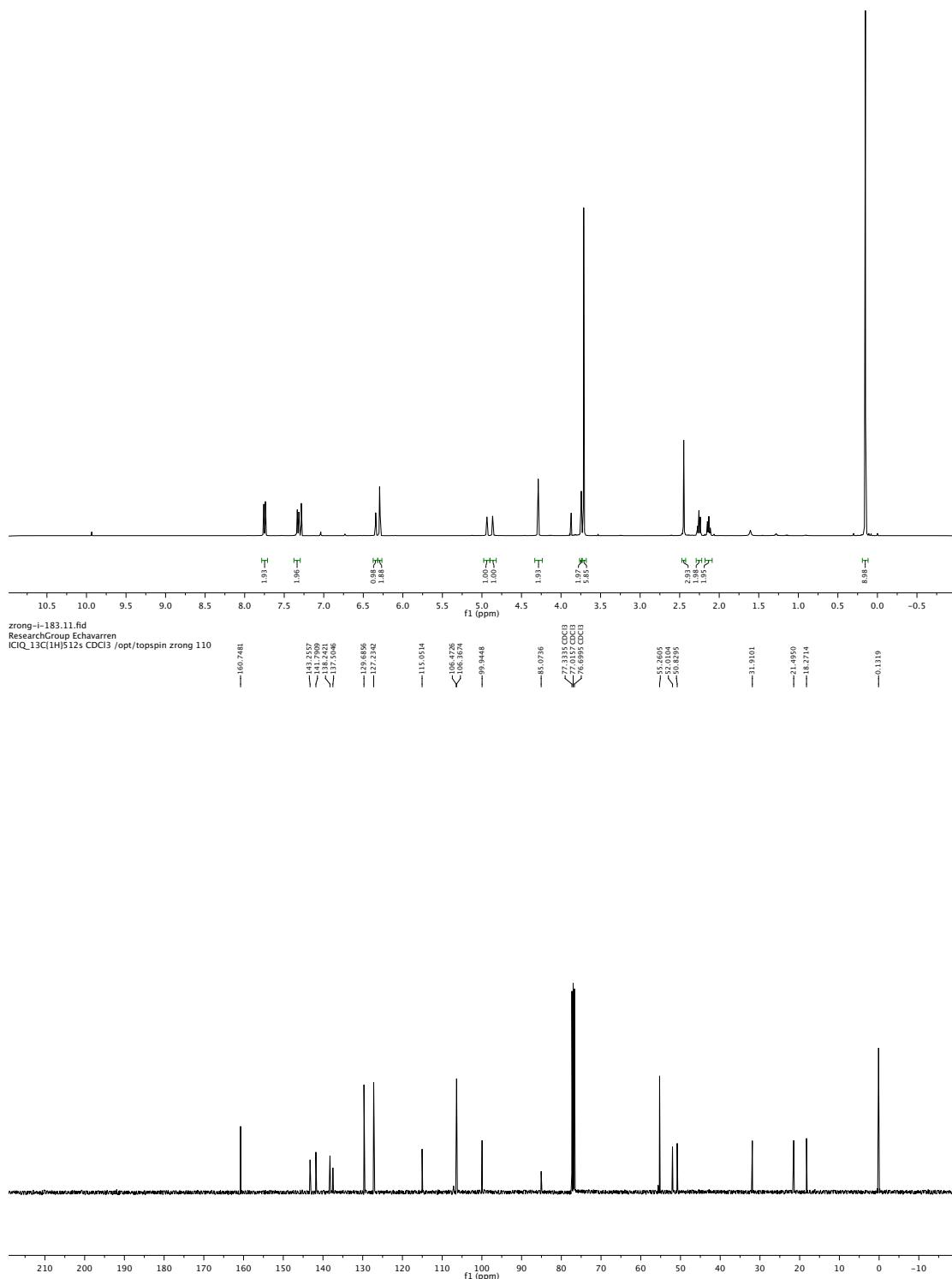


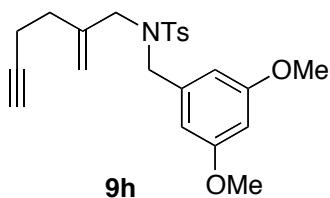
zrong-i-192.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p8s CDCl3 /opt/topspin zrong 23



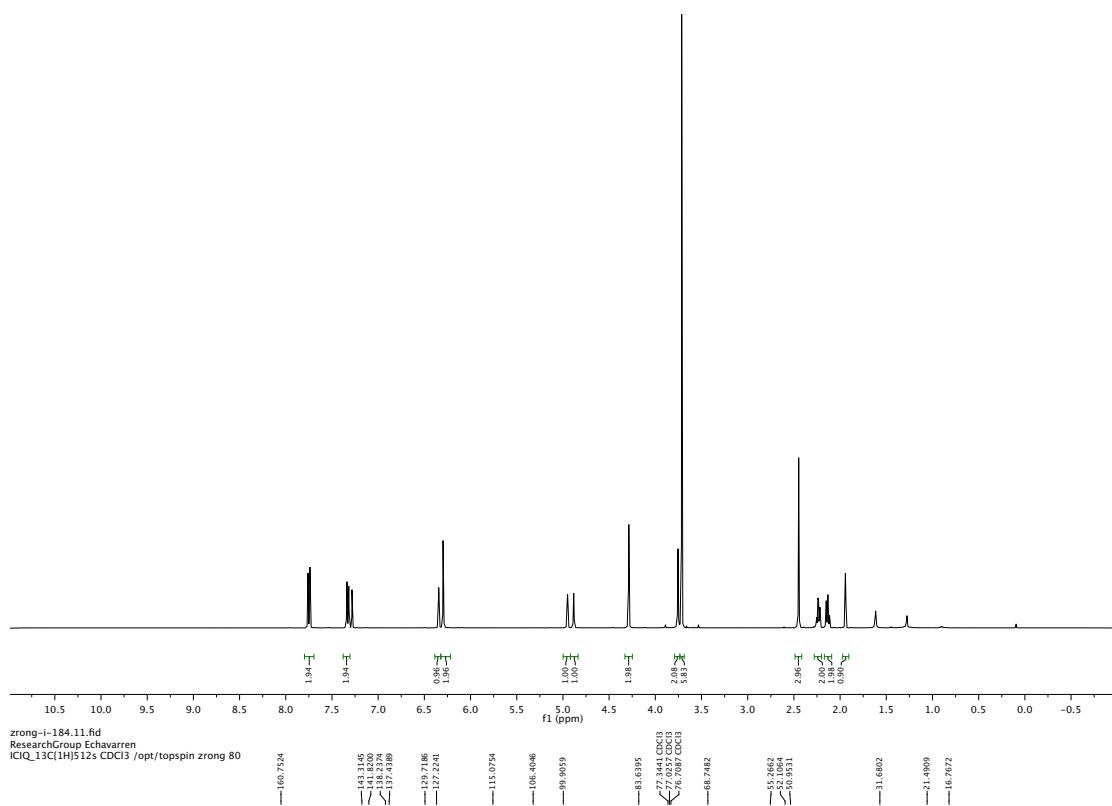


zrong-i-183.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 110

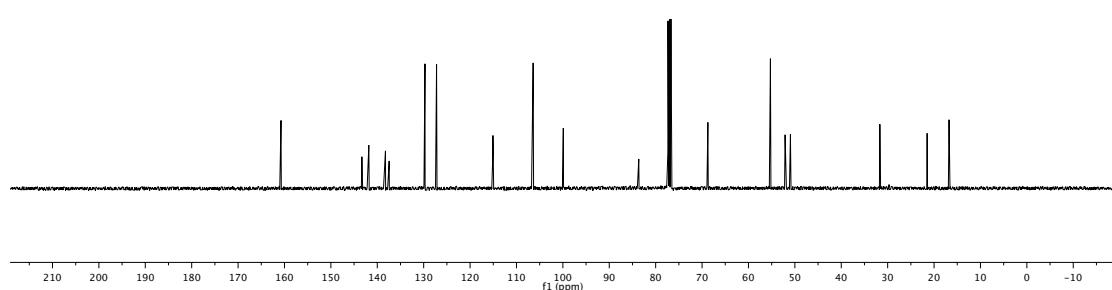


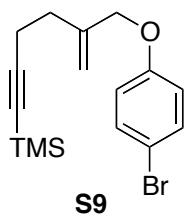


zrong-i-184.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 80

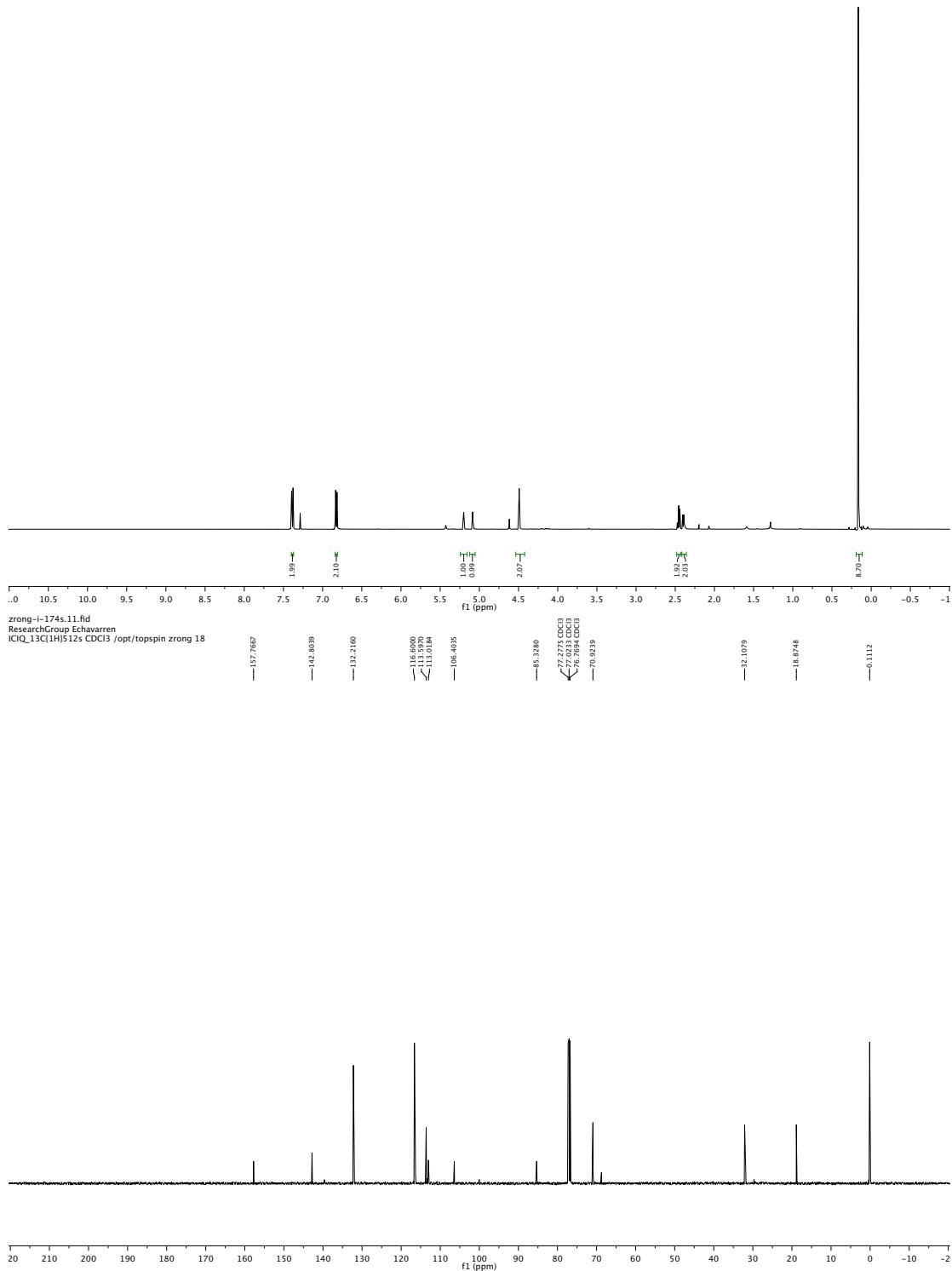


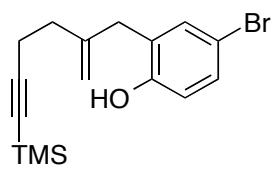
zrong-i-184.11.fid
 ResearchGroup Echavarren
 ICIQ_13C[1H]512s CDCl₃ /opt/topspin zrong 80



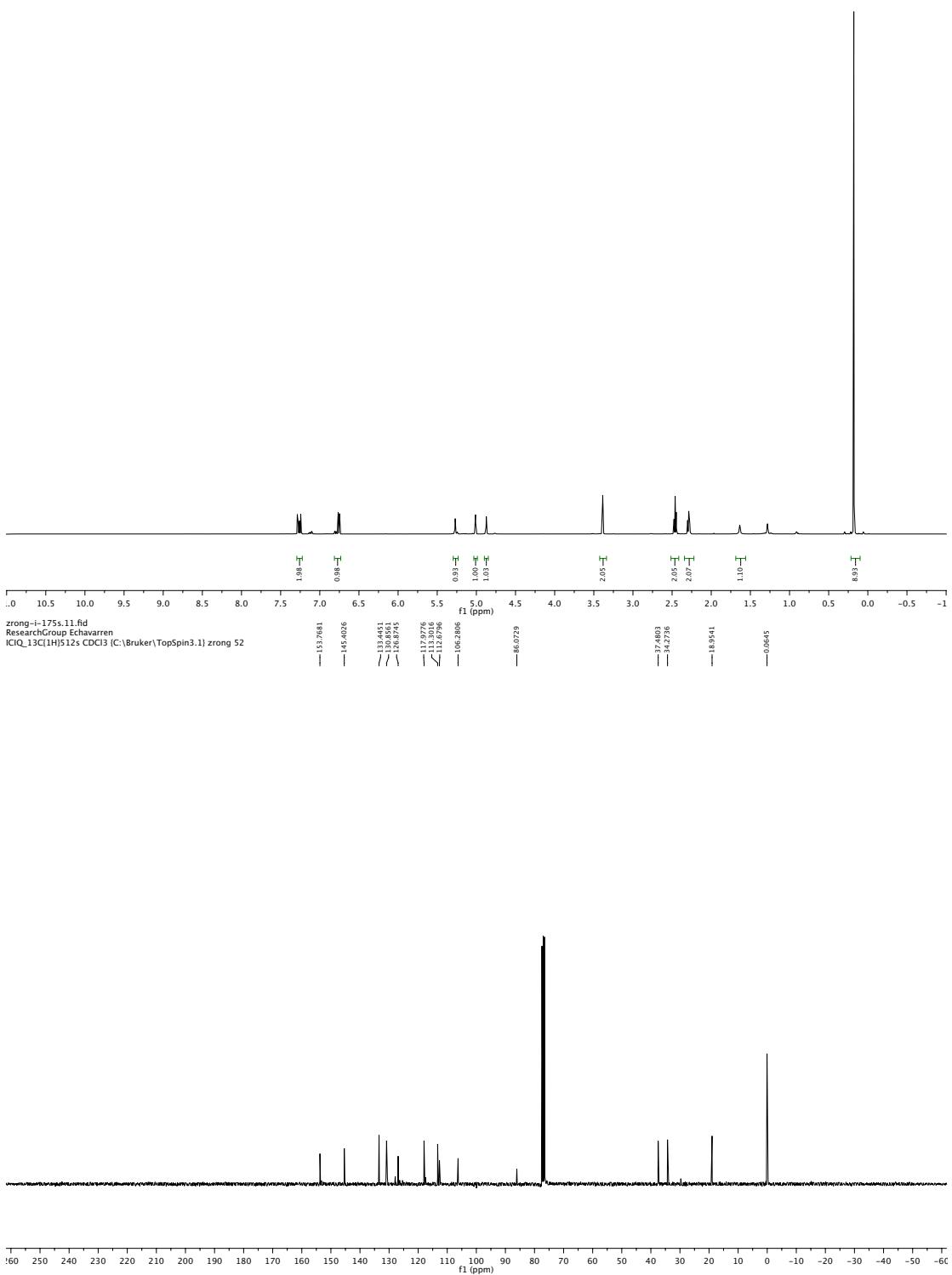


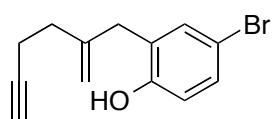
zrong-i-174s.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 18





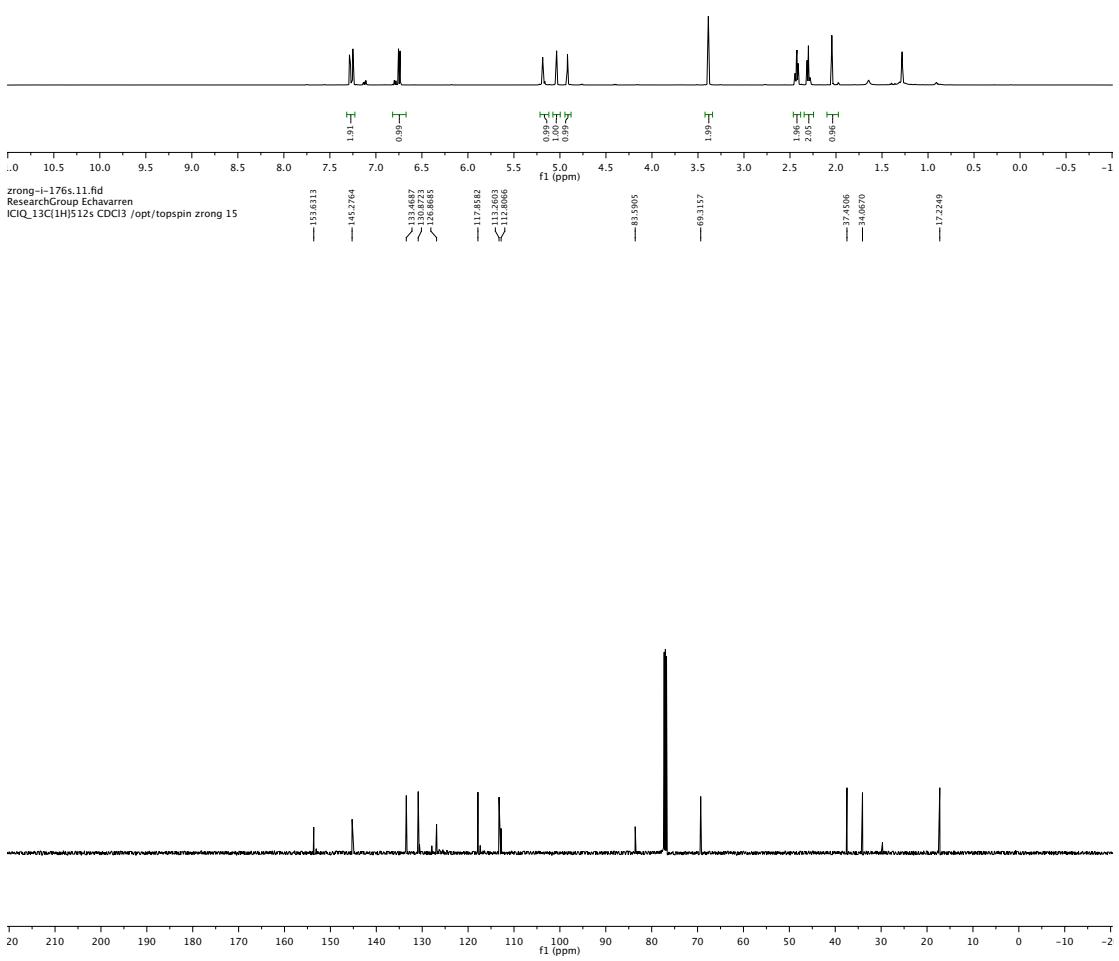
zong-i-175s.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zong 52

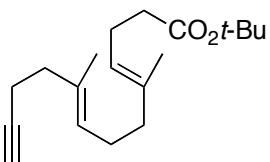




9i

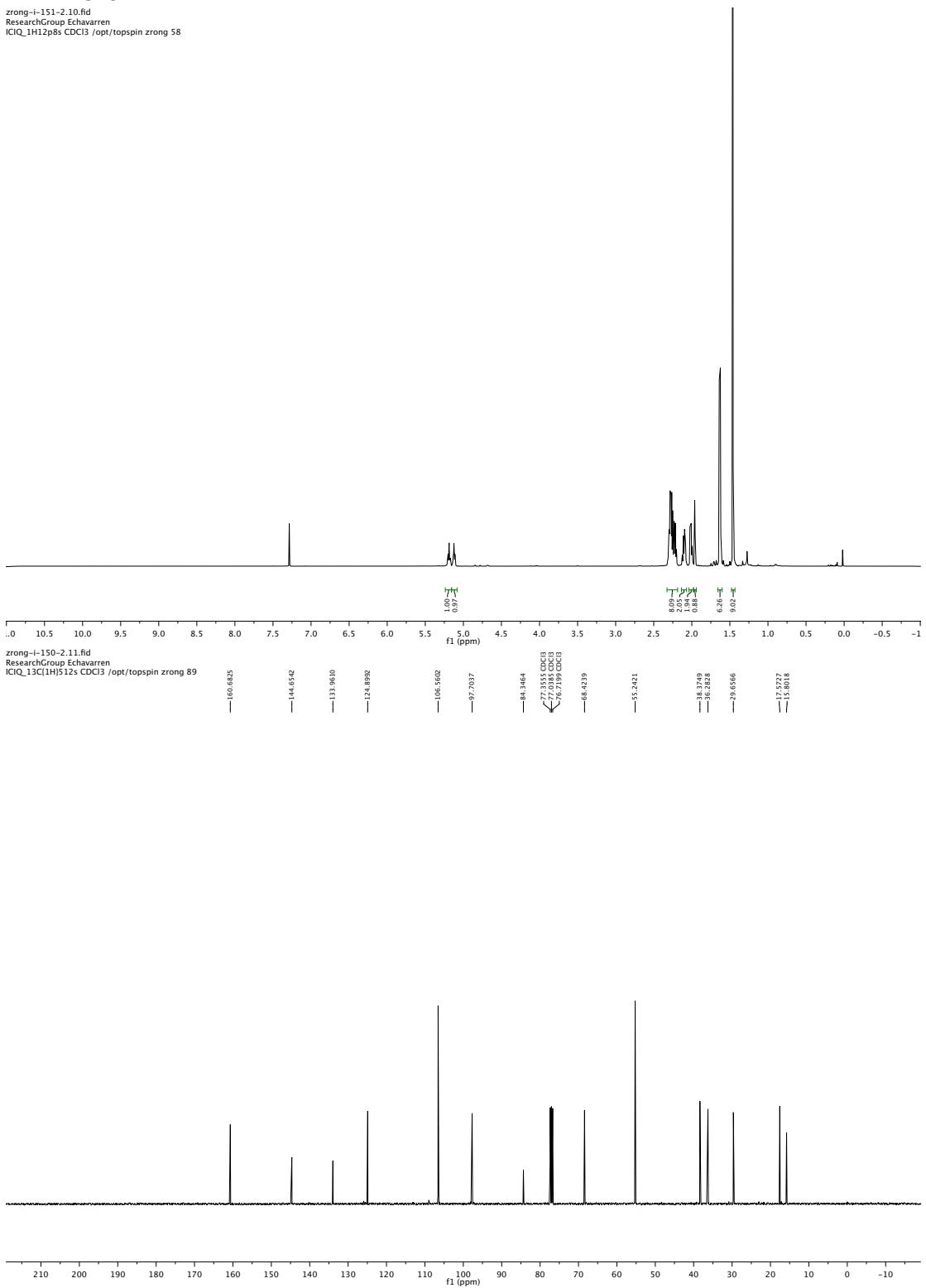
zong-i-176s.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zong 15

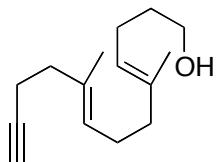




S13

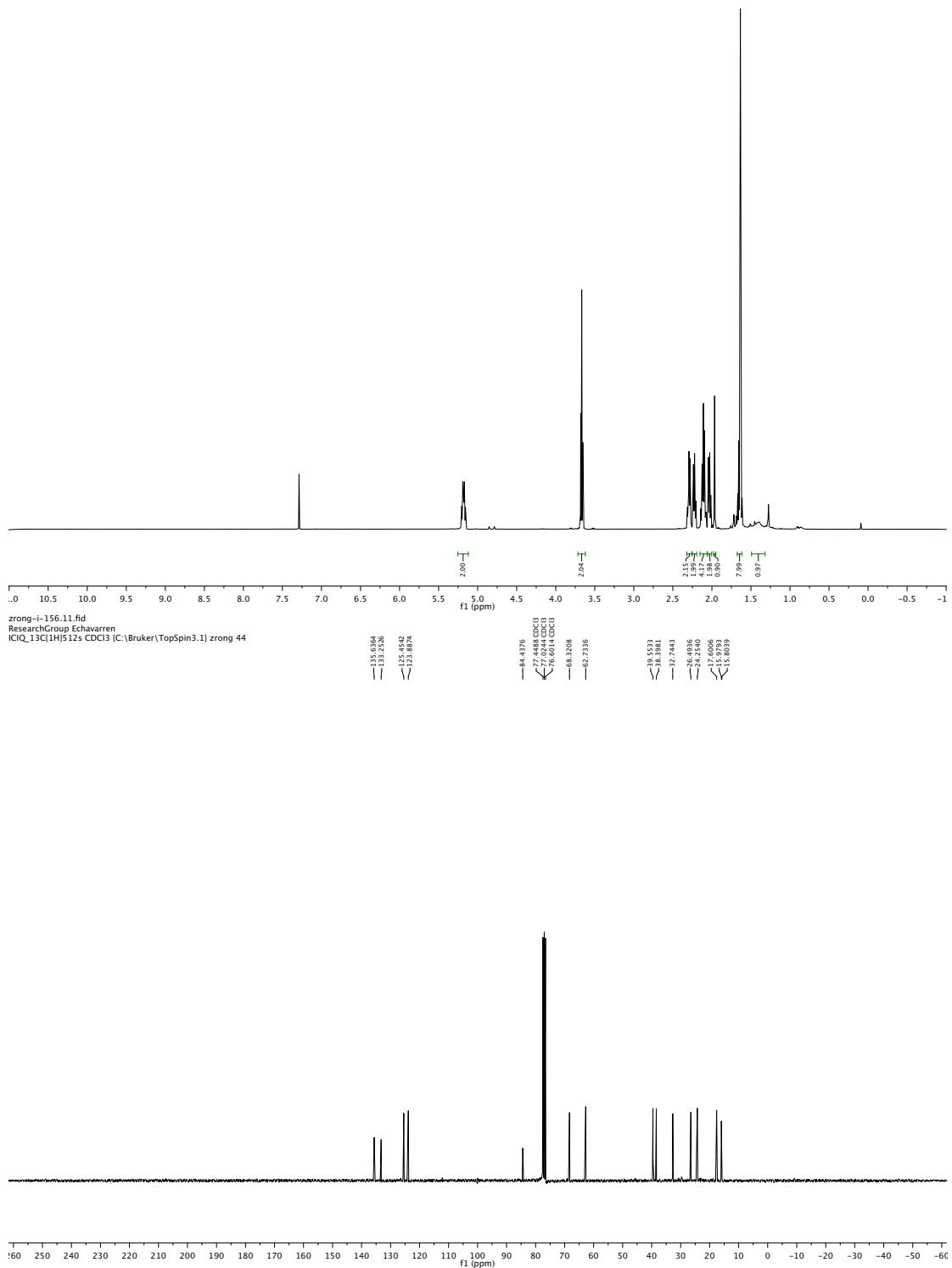
zrong-i-151-2.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCI3 /opt/topspin zrong 58

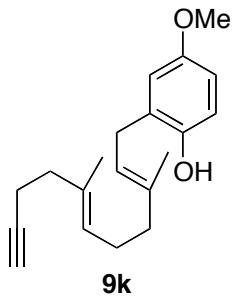




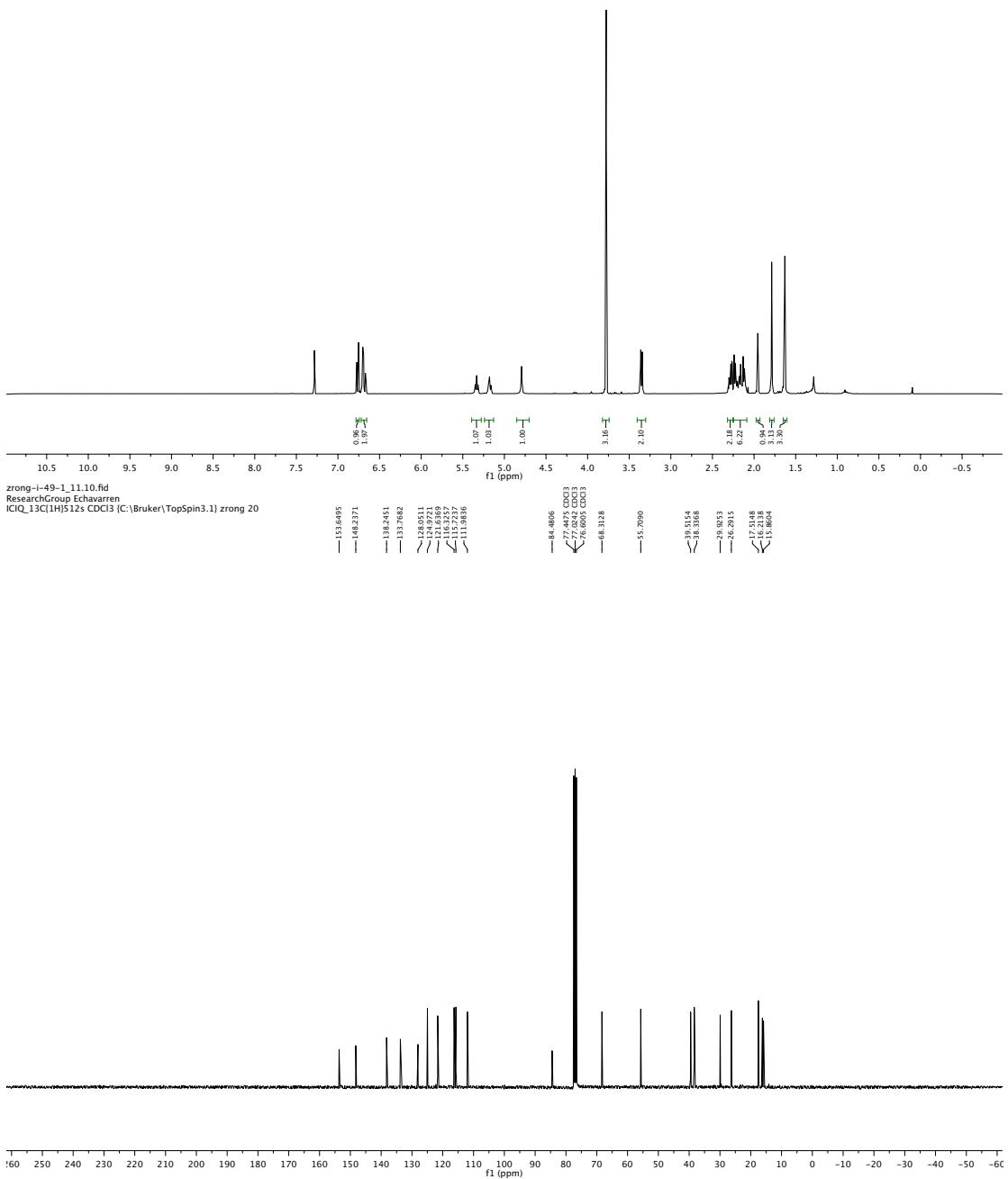
9j

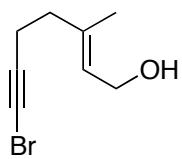
zrong-i-156.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDC13 /opt/topspin zrong 44





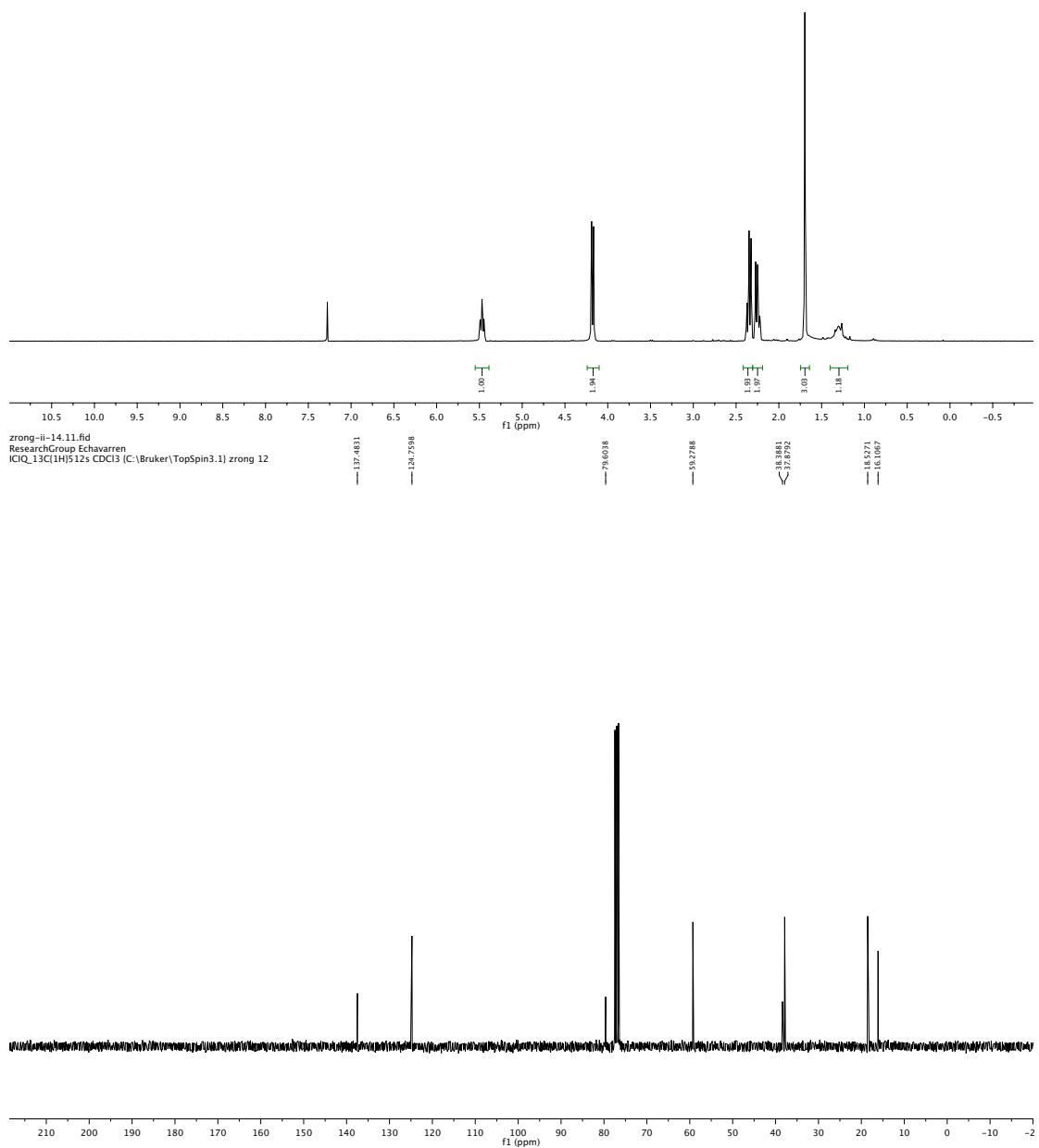
zrong-i-49-1_10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 80

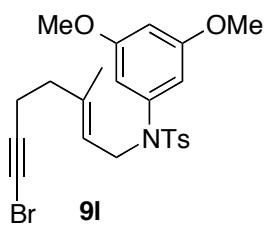




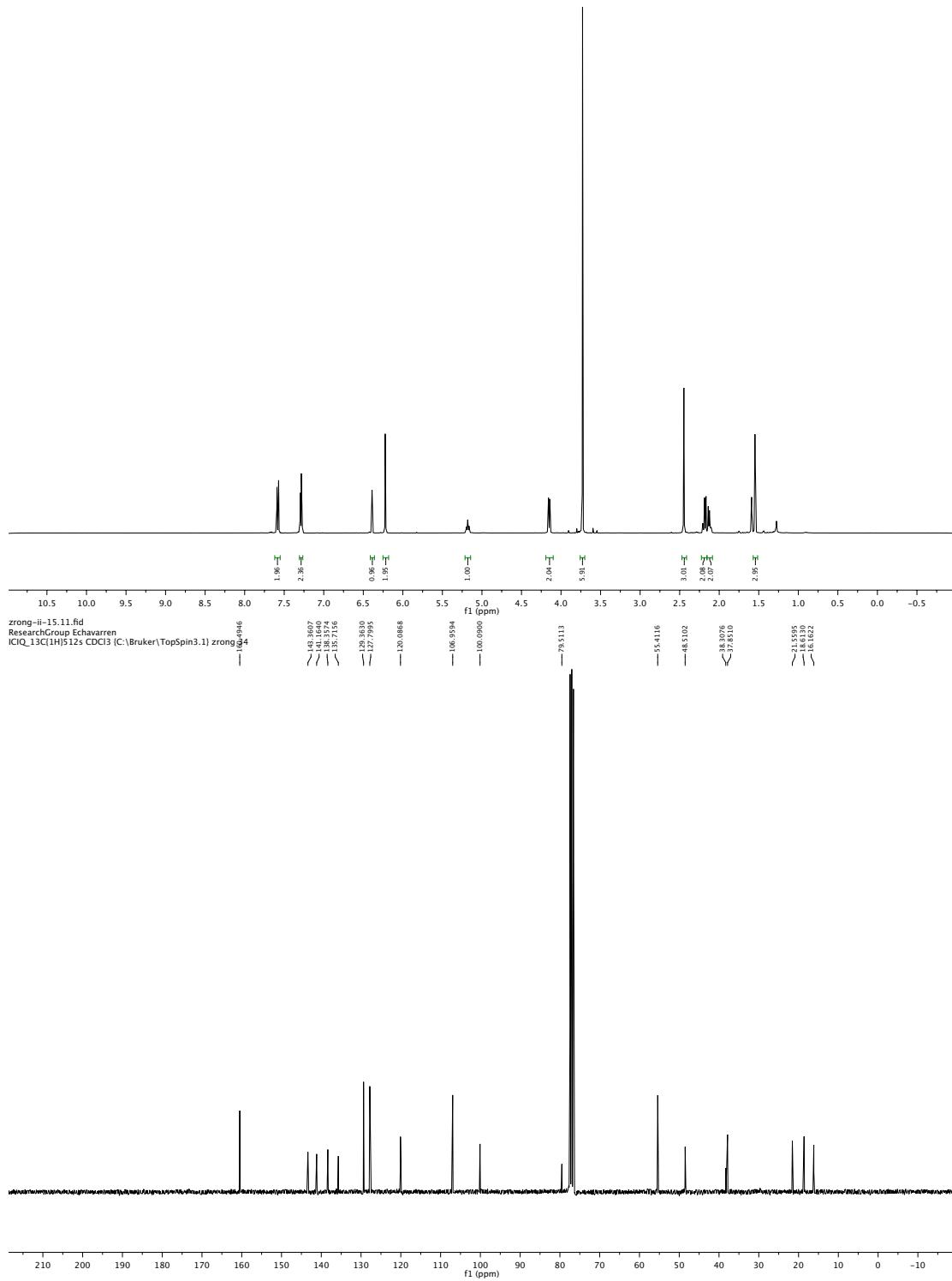
S14

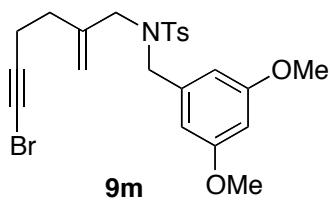
zrong-ii-14.10.fid
ResearchGroup Echavarren
ICIQ_1H20p8s CDCl3 (C:\Bruker\TopSpin3.1) zrong 12



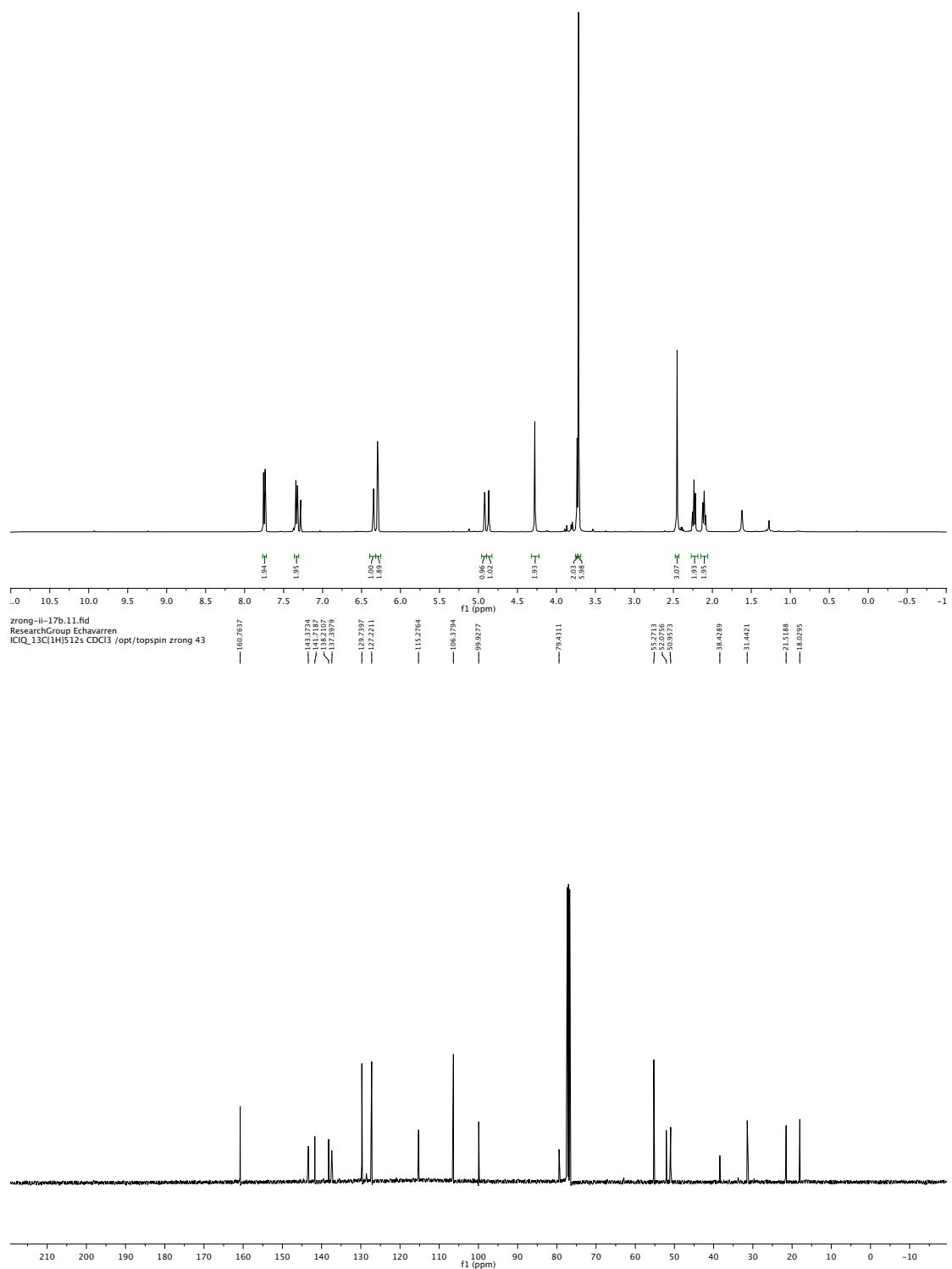


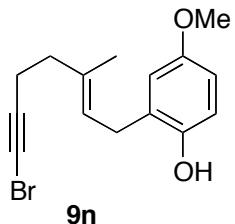
zrong-ii-15.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p8s CDCl3 /opt/topspin zrong 94



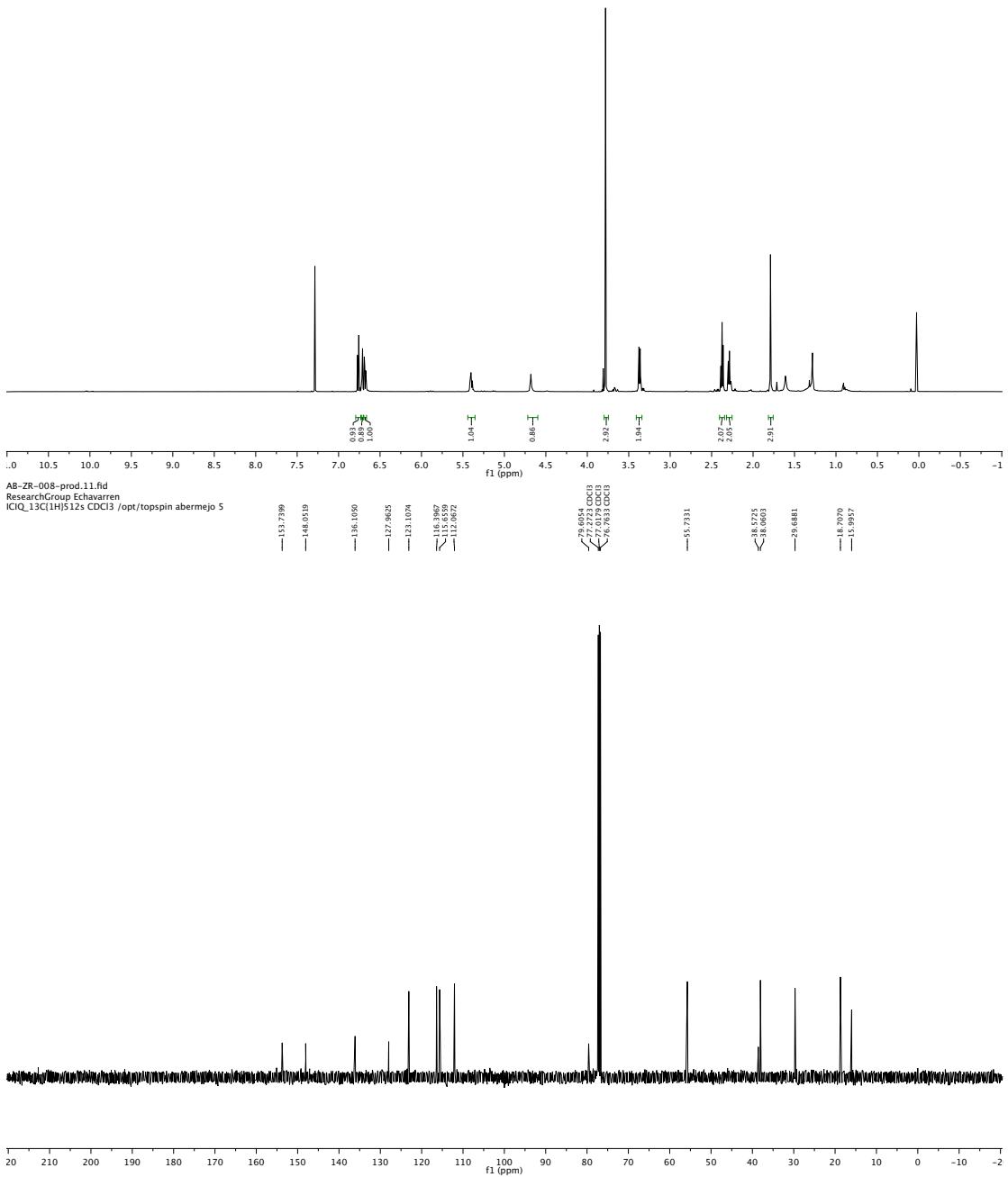


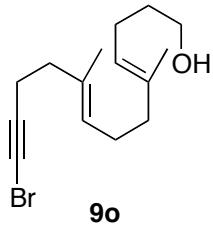
zrong-ii-17b.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 43



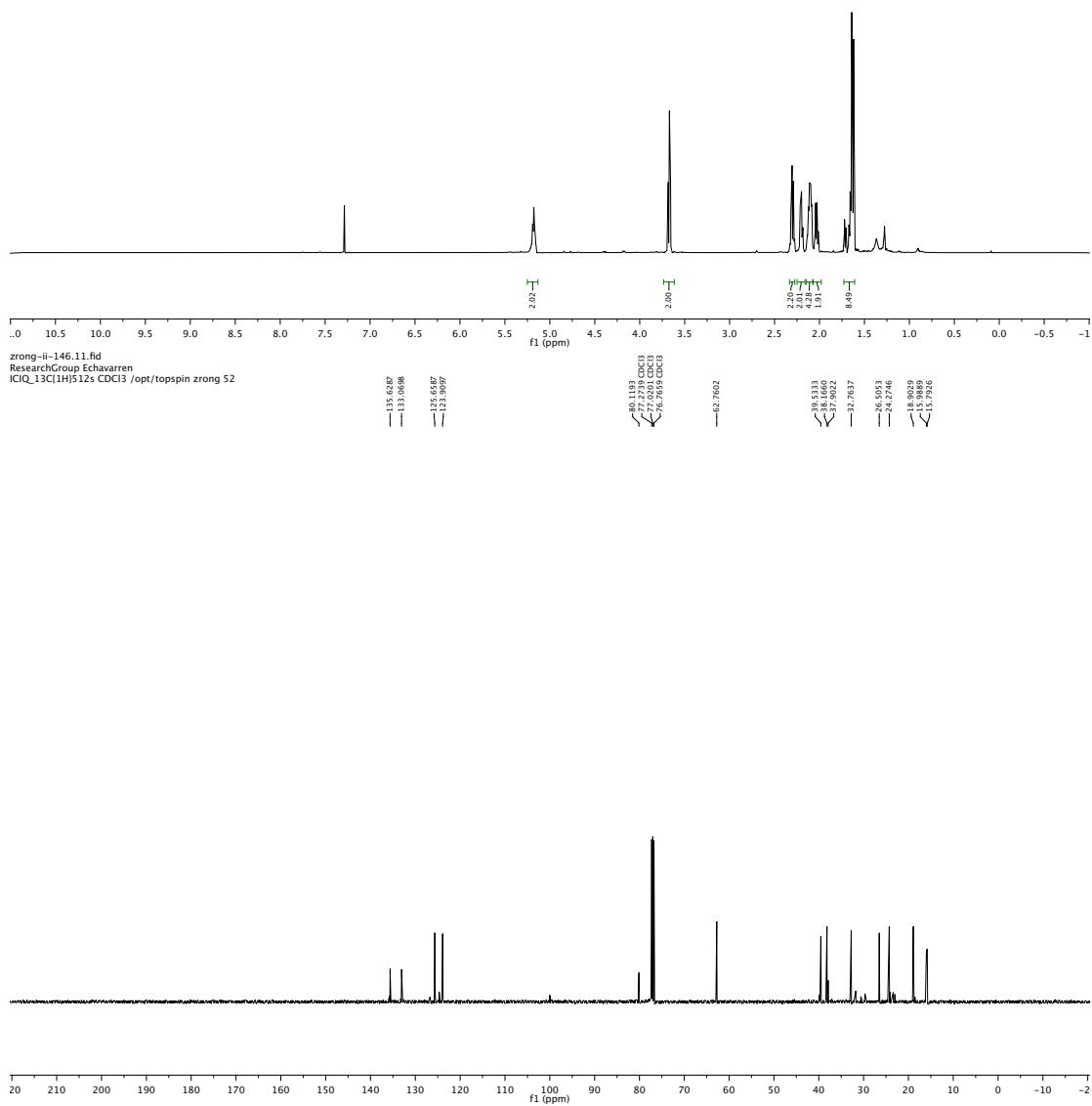


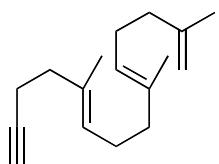
AB-ZR-008-prod.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p8s CDCl₃ /opt/topspin abermejo 5





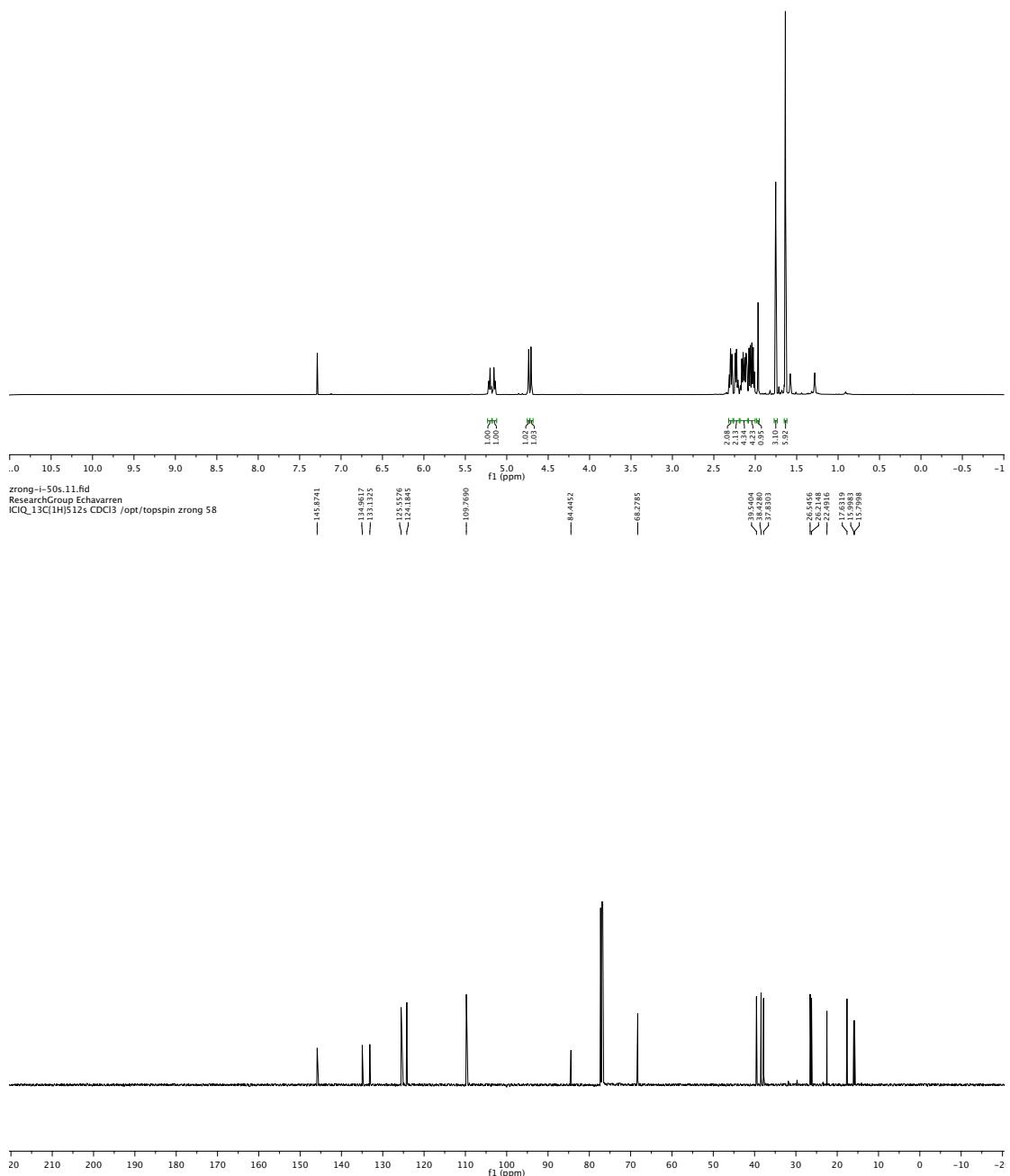
zrong-ii-146.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 52

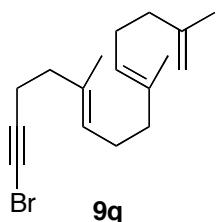




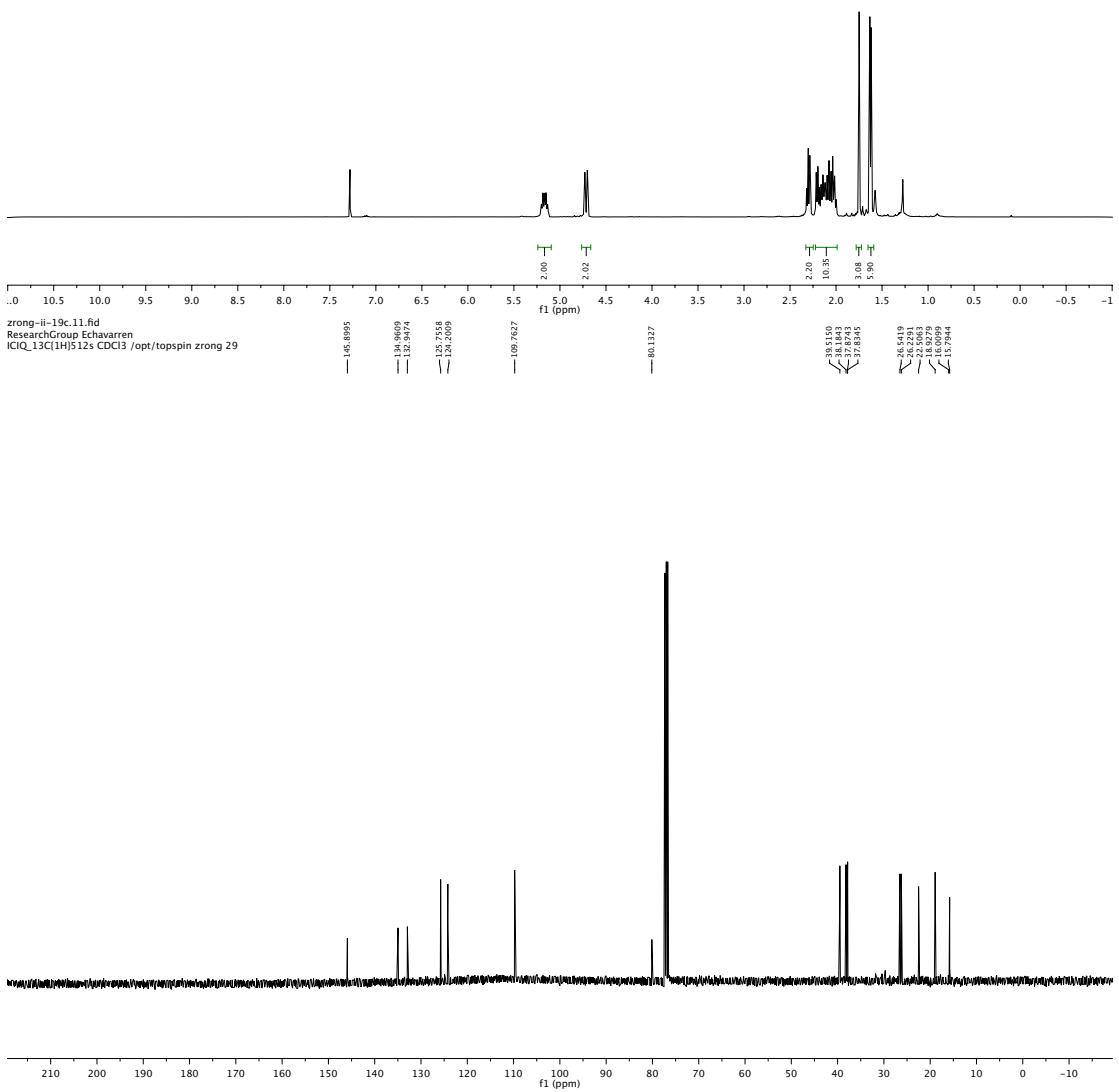
9p

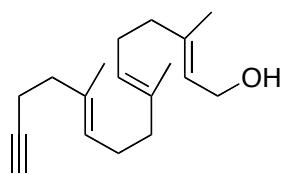
zrong-i-50s.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 58





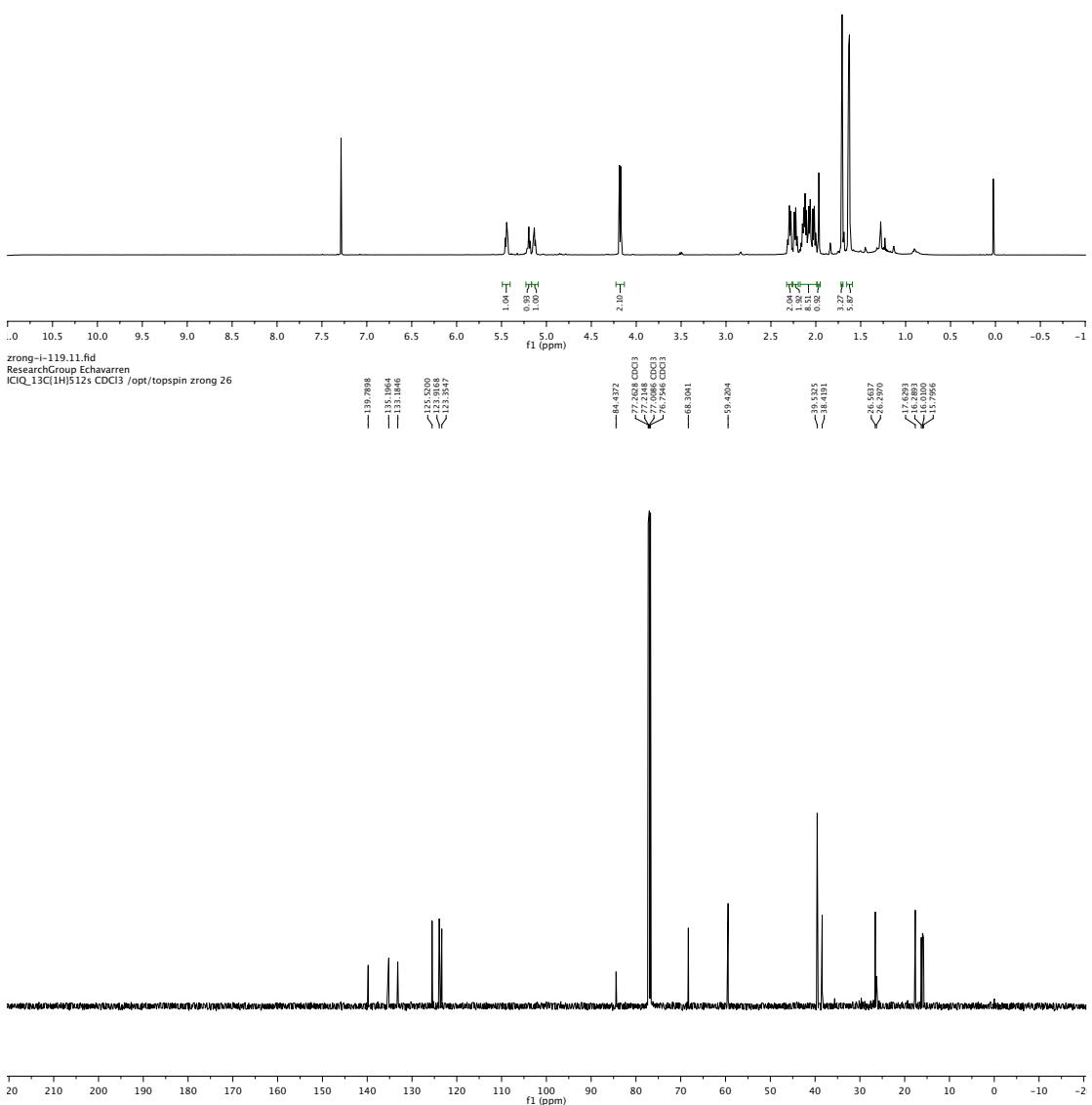
zrong-ii-19c.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 29

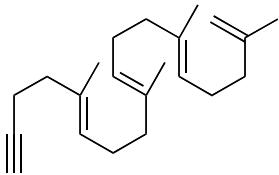




S20

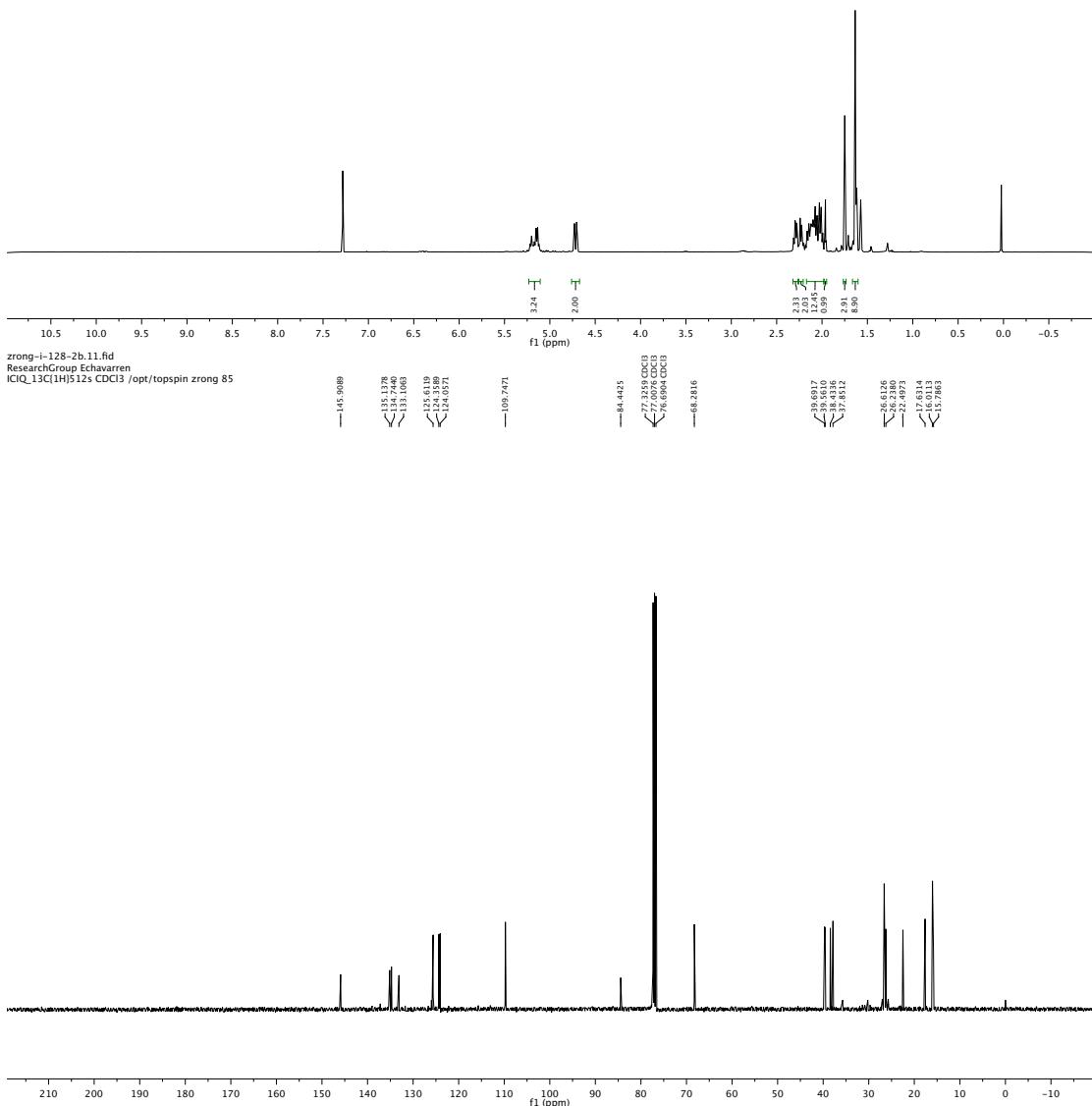
zrong-i-119.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 26

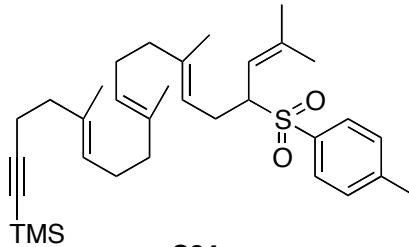




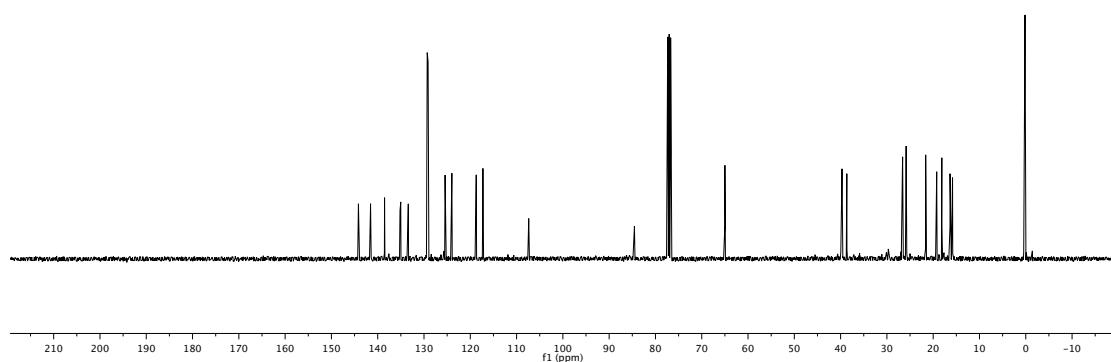
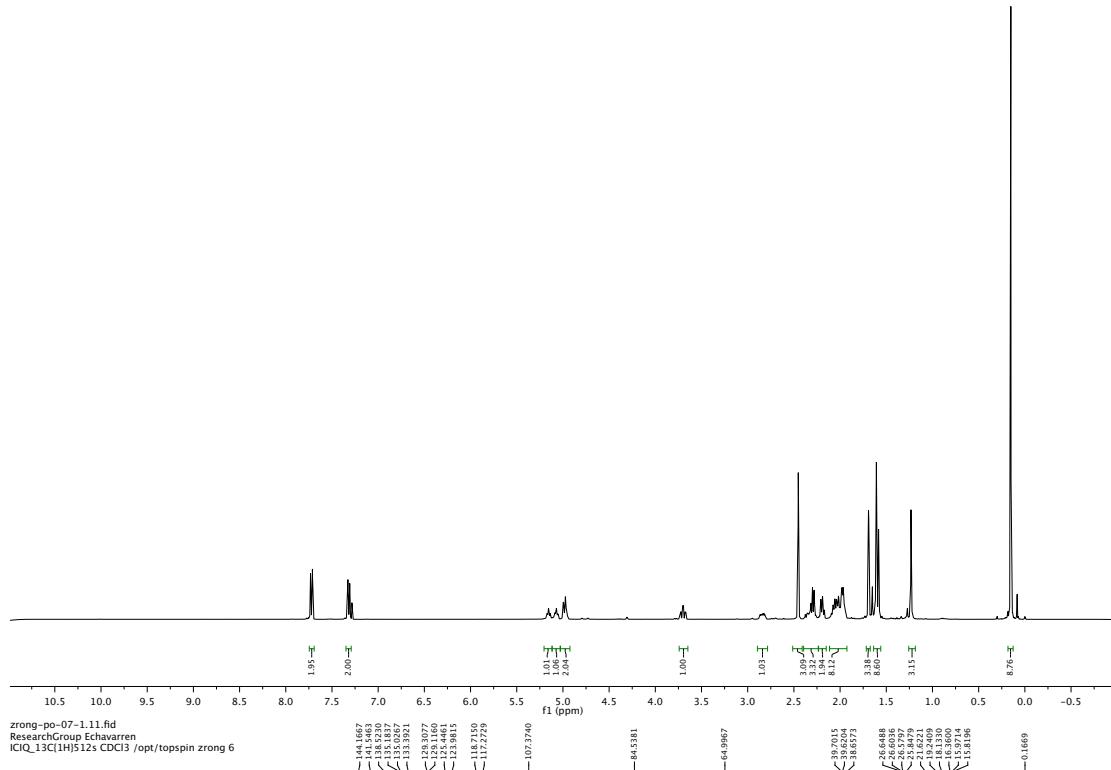
9r

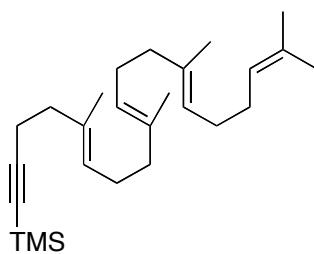
zrgn-i-128-2b.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrgn 85



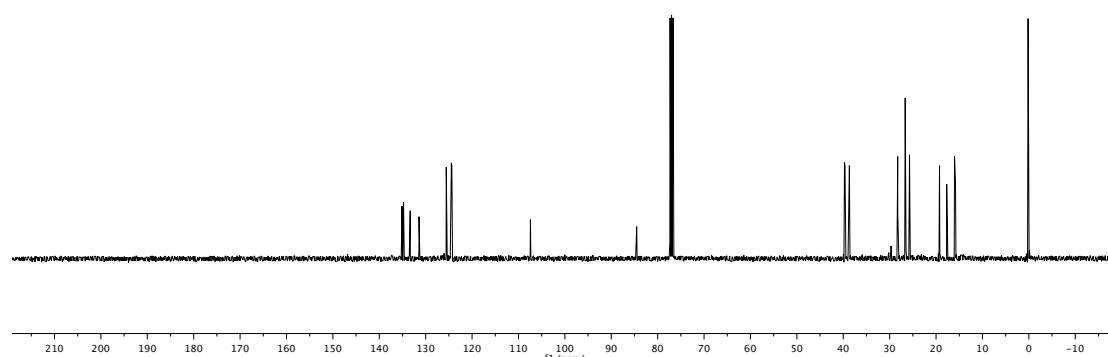
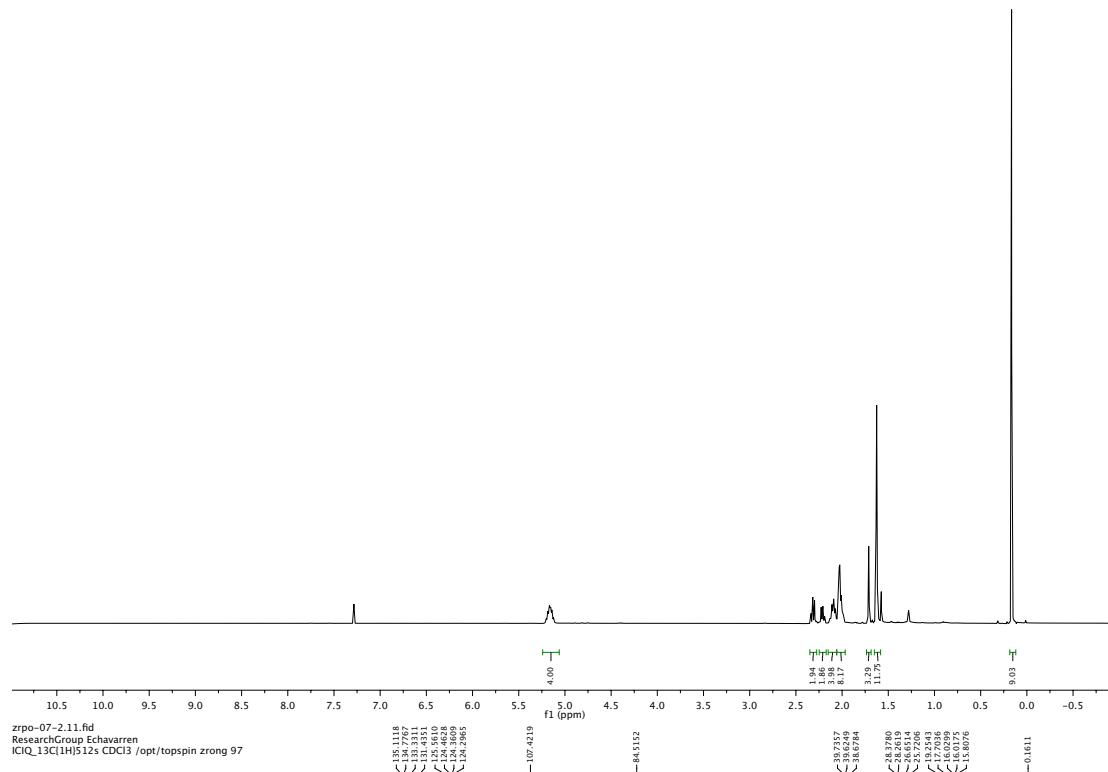


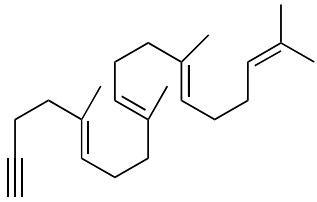
zrong-po-07-1.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 66





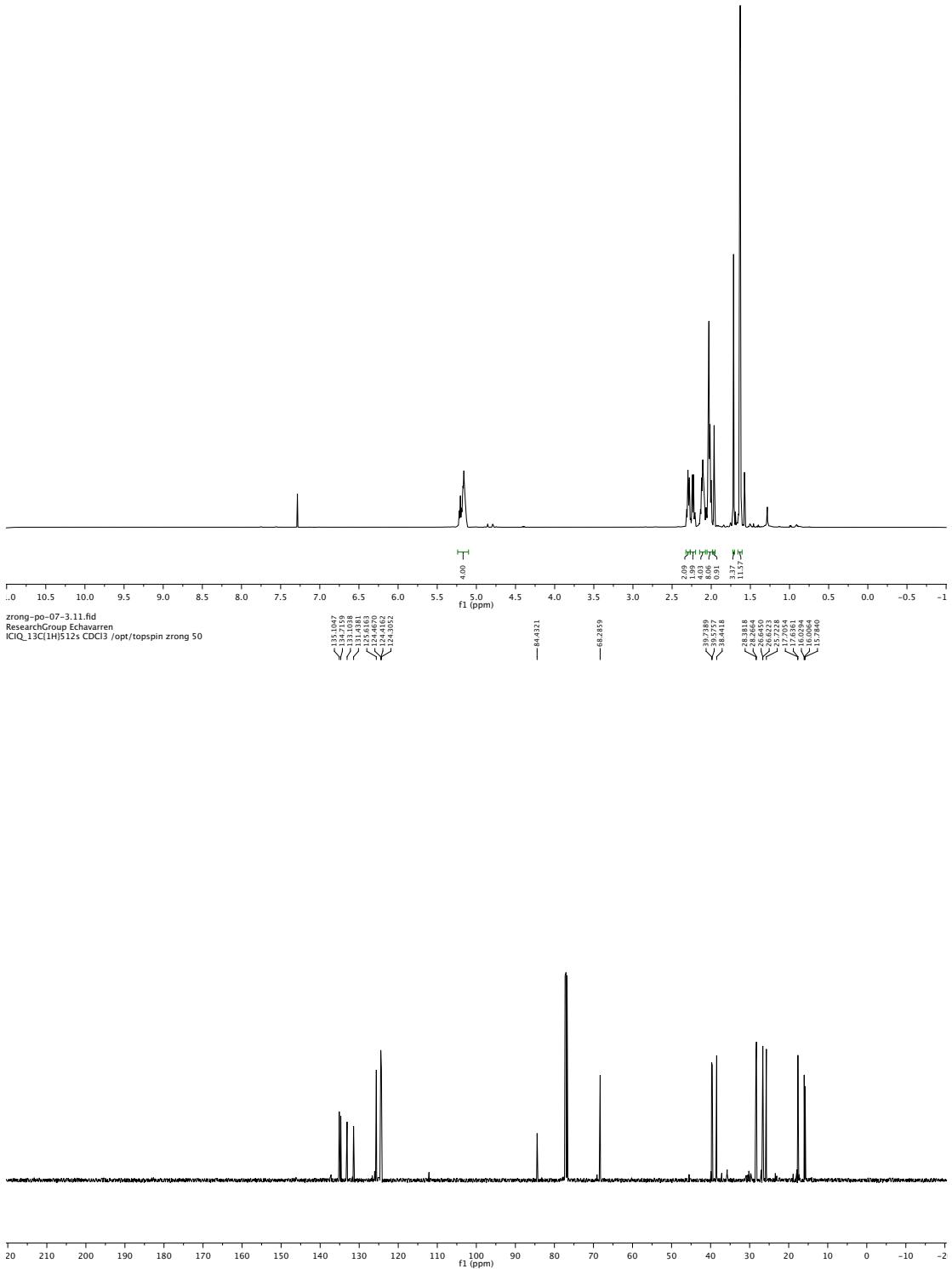
zrpo-07-2.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrpng 97

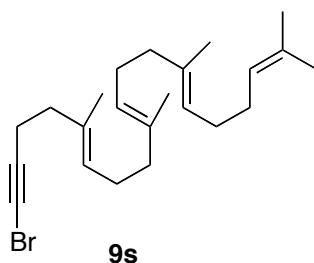




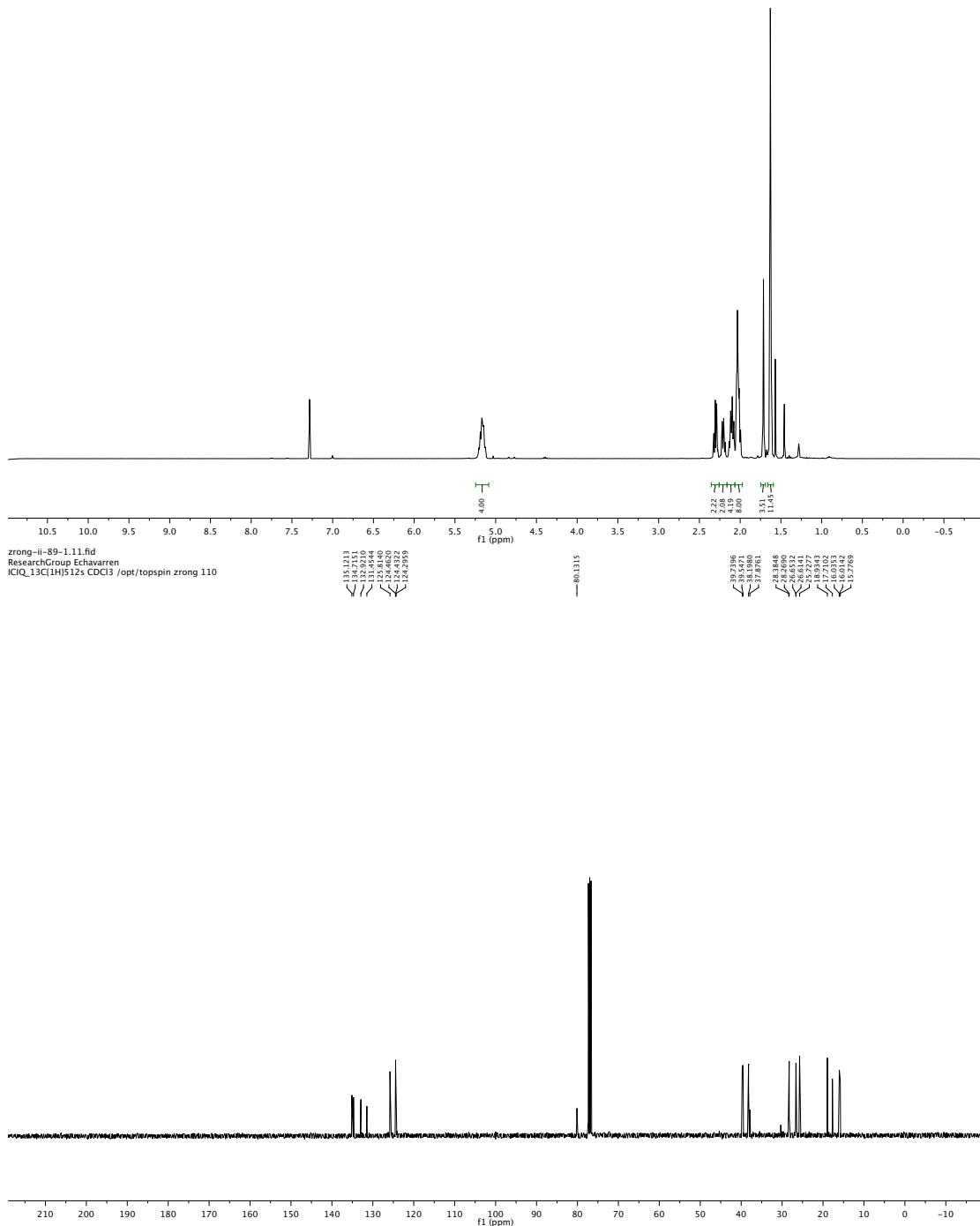
S26

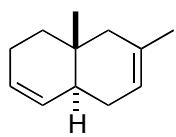
zrong-po-07-3.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 50





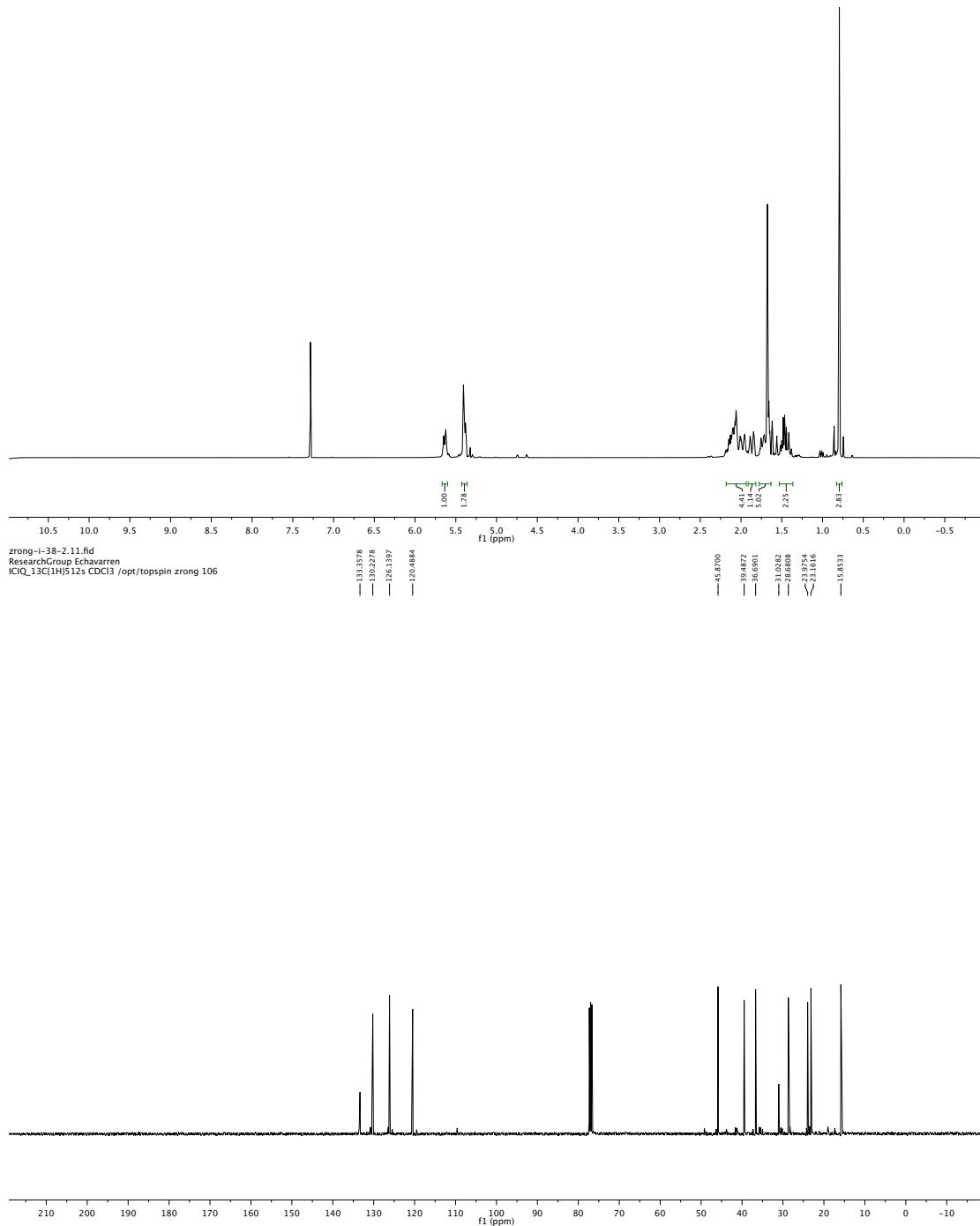
zrong-ii-89-1.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 110

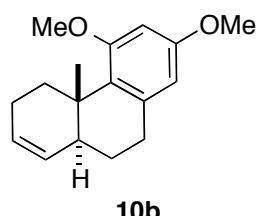




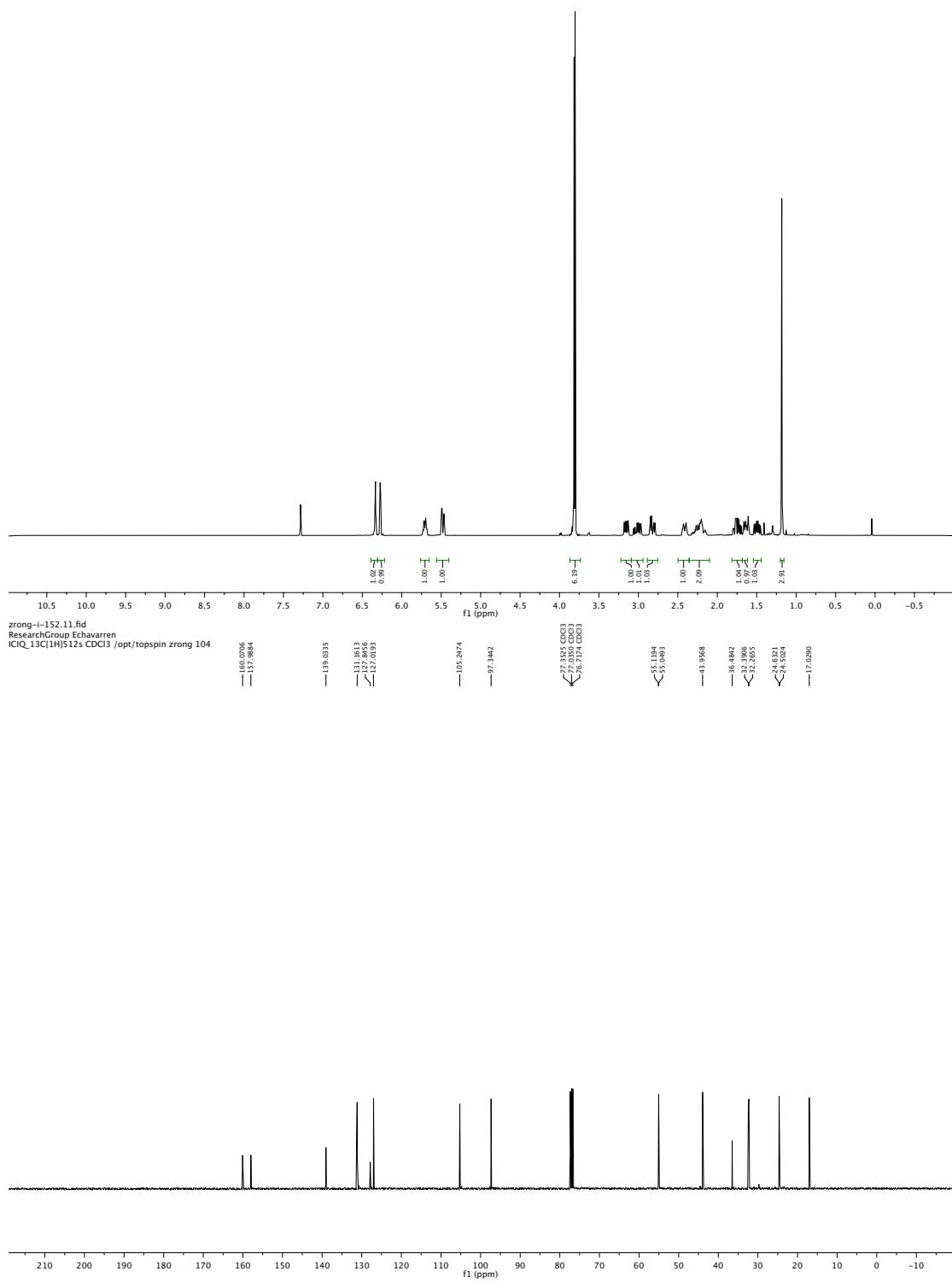
10a

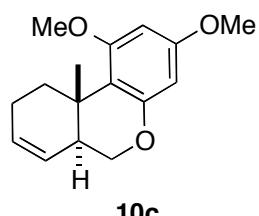
zrong-i-38-2.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 106



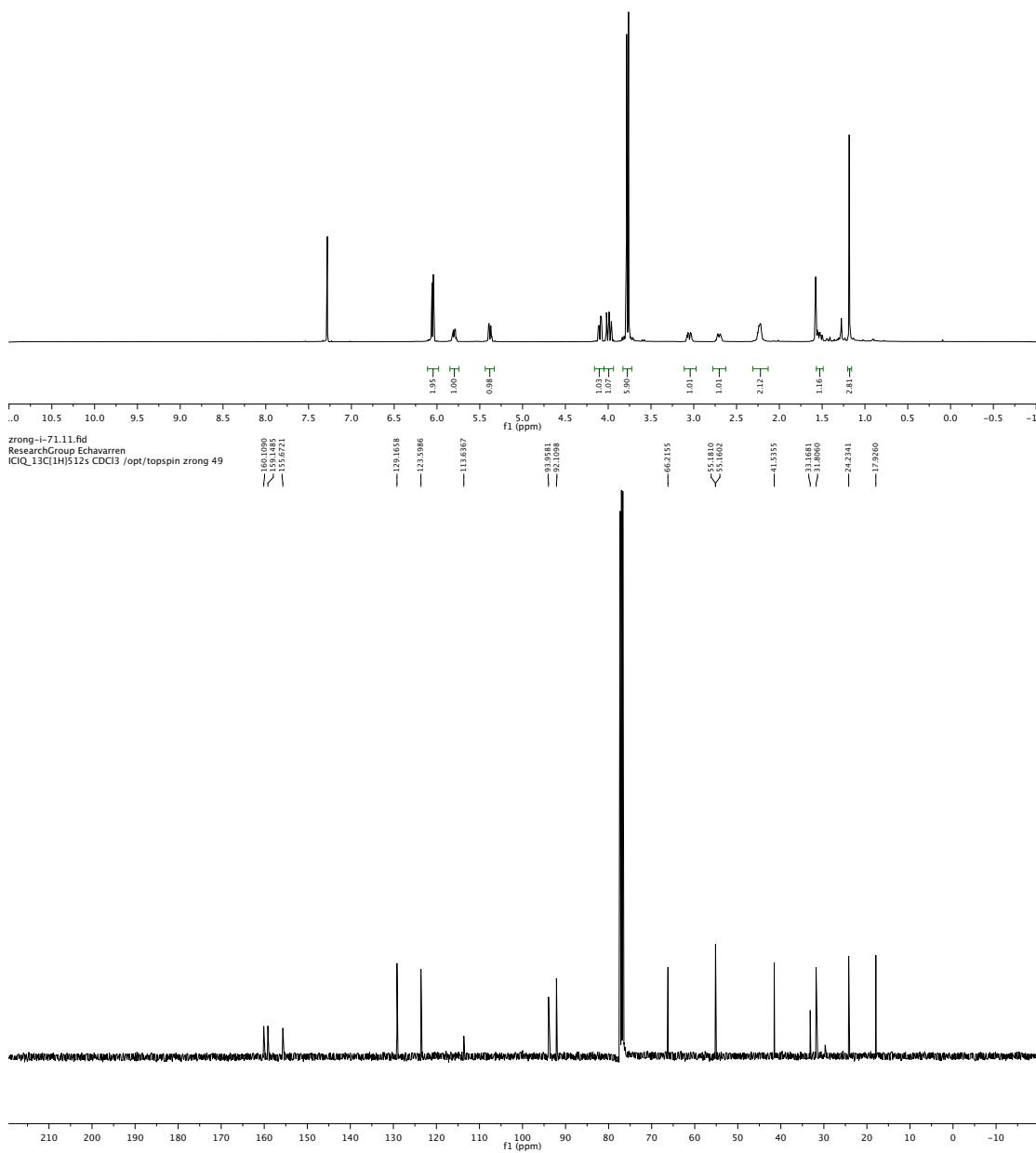


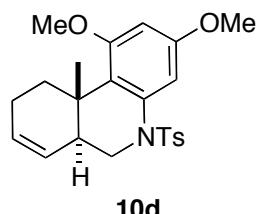
zrong-i-152.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 104



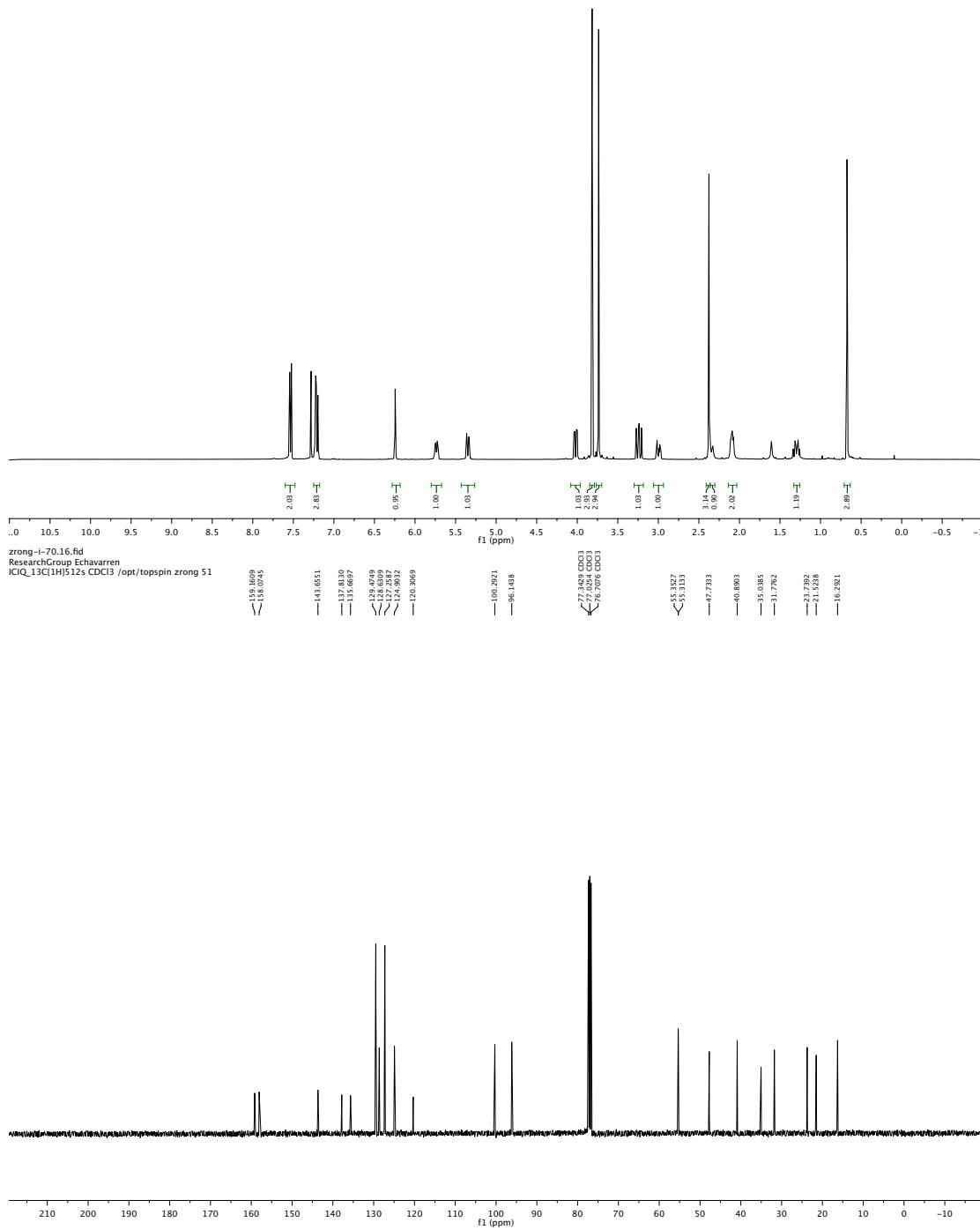


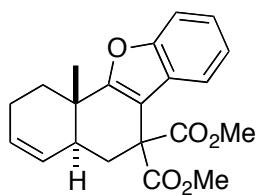
zrong-i-71.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p32s CDCl₃ /opt/topspin zrong 49





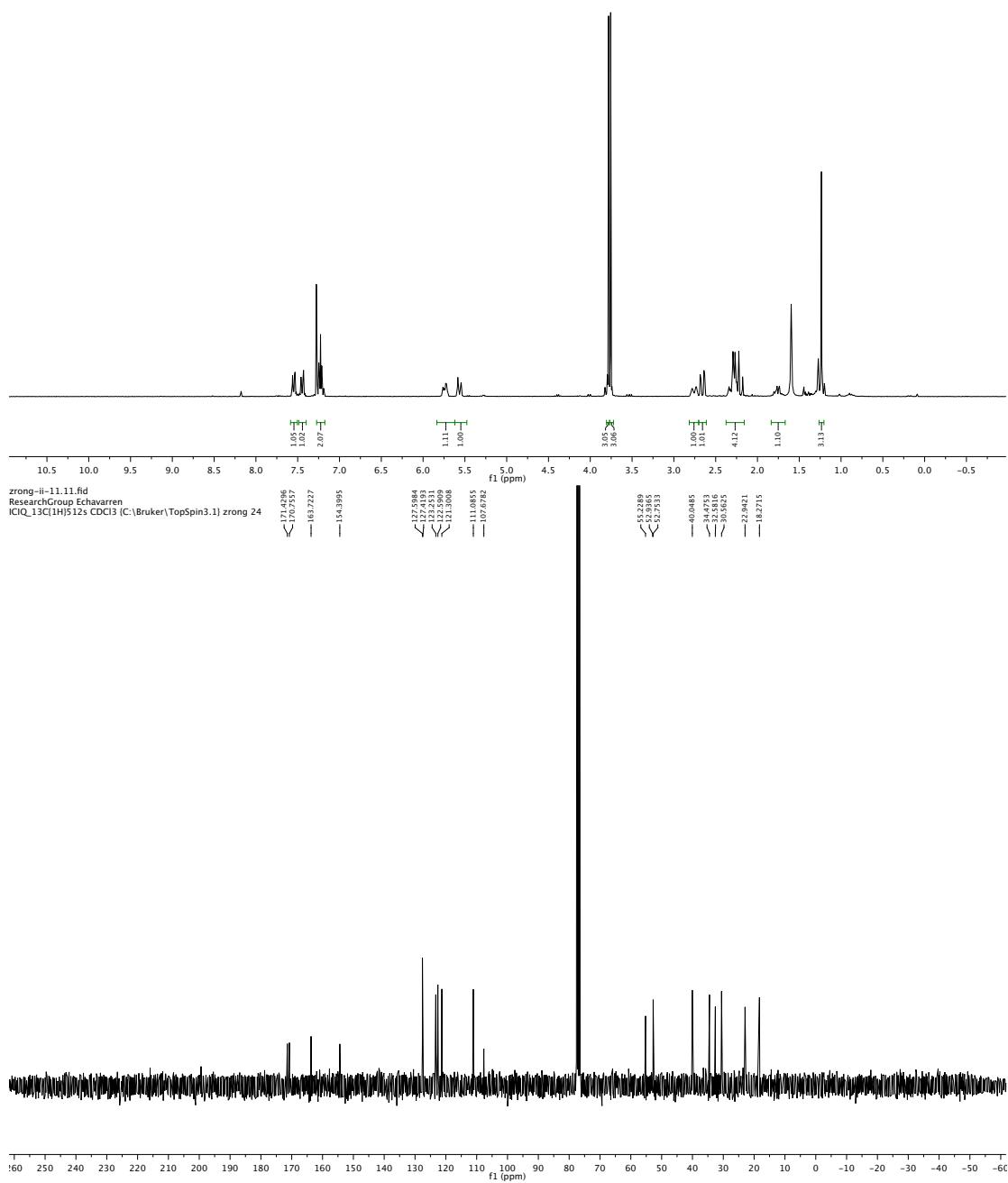
zrong-i-70.10.fid
 ResearchGroup Echavarren
 ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 51

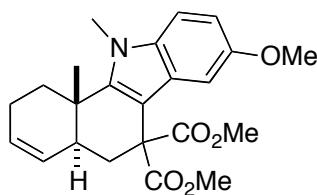




10e

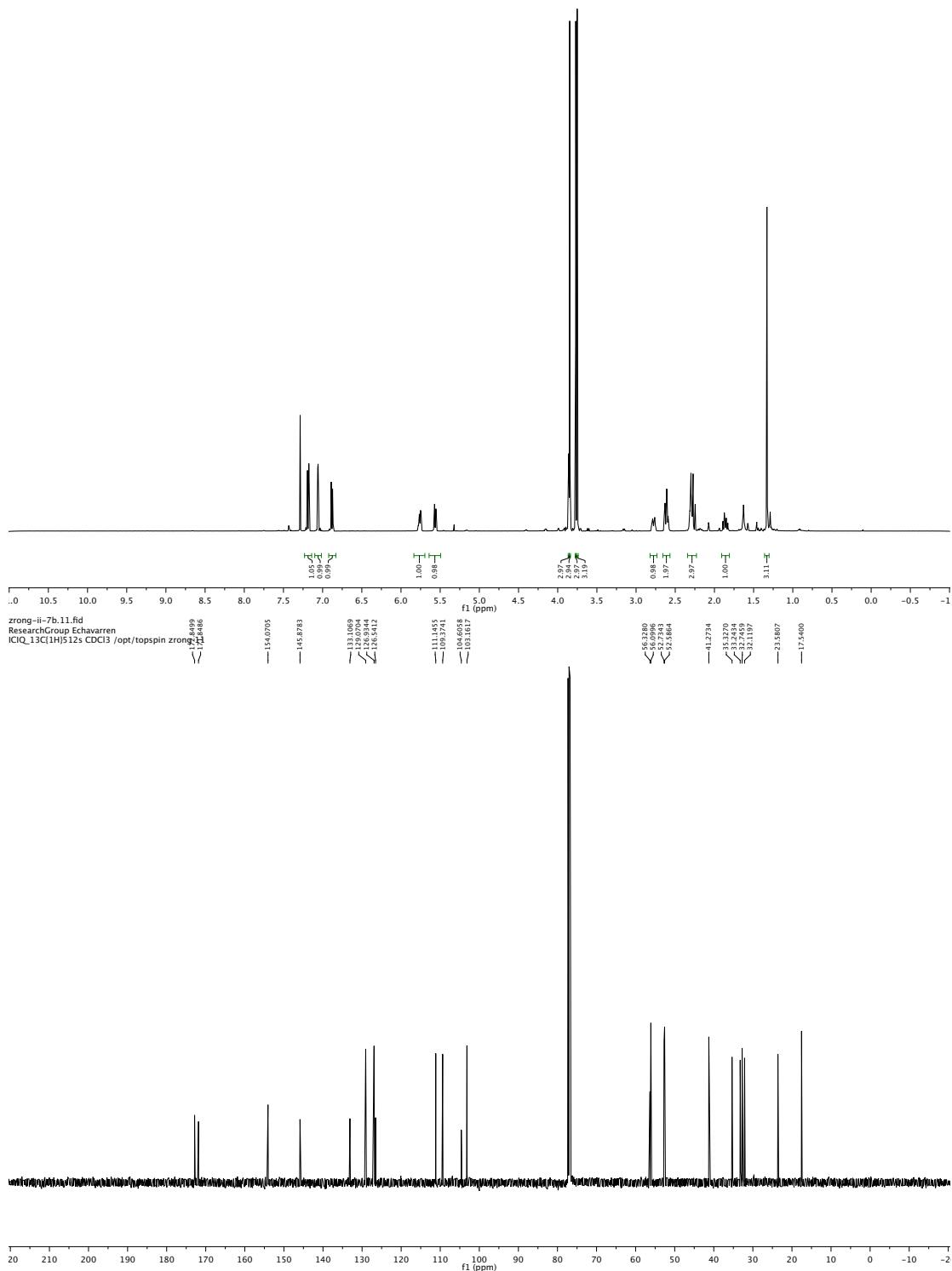
zrong-ii-11.10.fid
ResearchGroup Echavarren
ICIQ_1H20p8s CDCl3 (C:\Bruker\TopSpin3.1) zrong 24

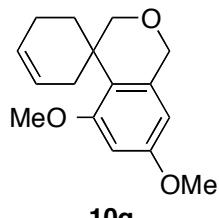




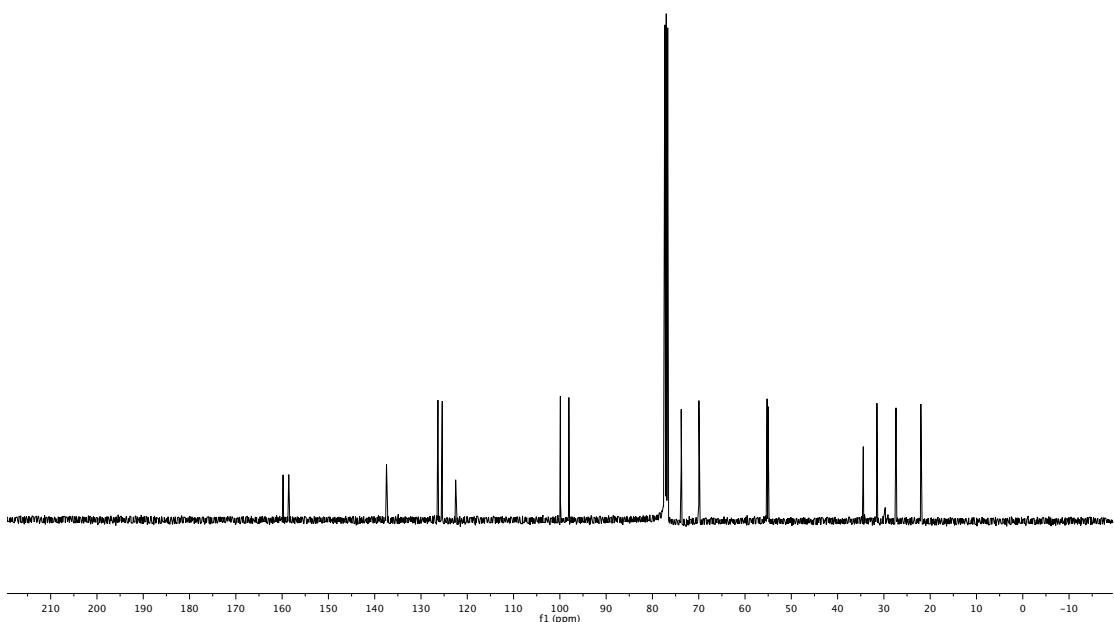
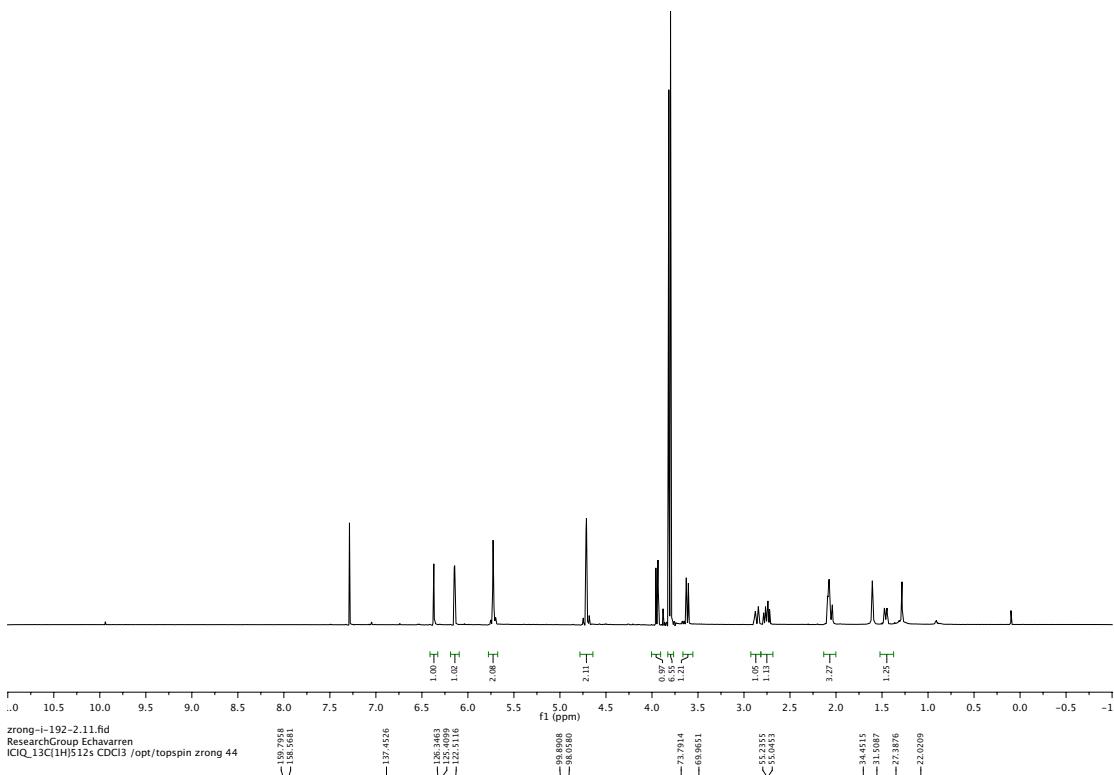
10f

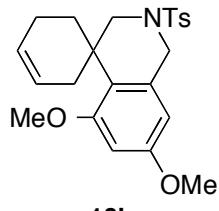
zrong-ii-7b.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 11



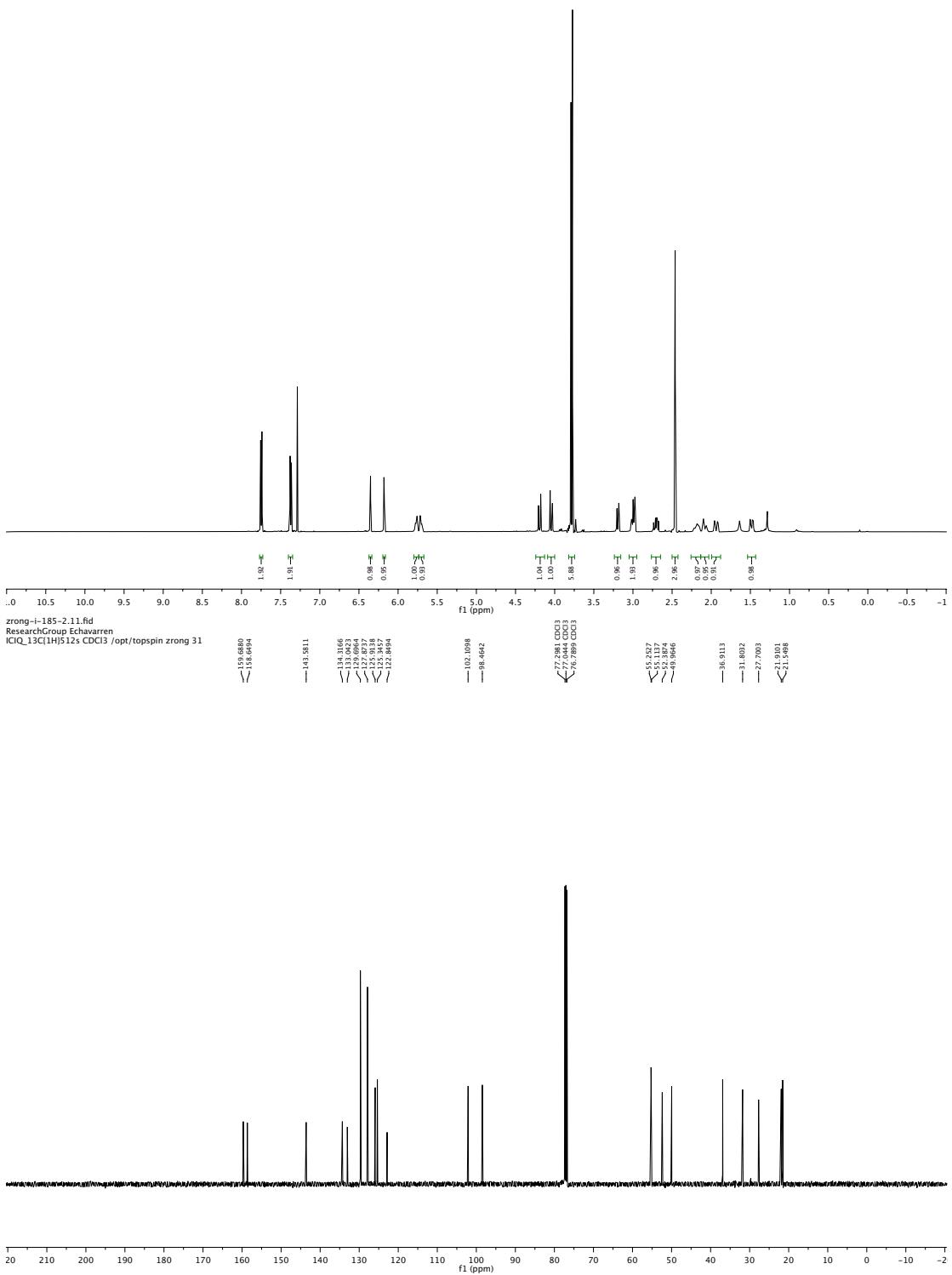


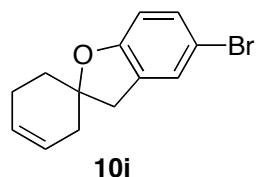
zrong-i-192-2.10.fid
ResearchGroup_Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 44



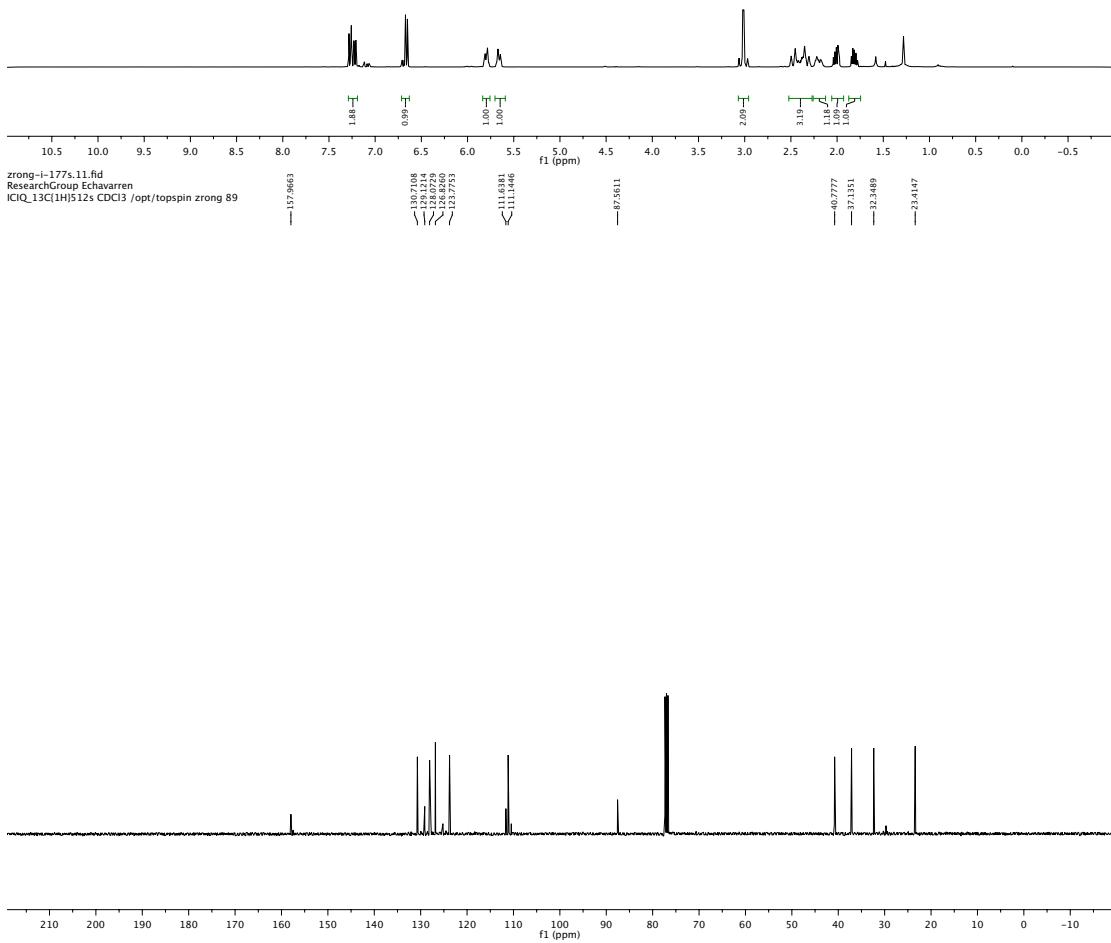


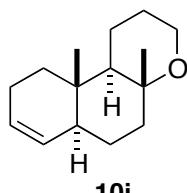
zrong-i-185-2.10.fid
 ResearchGroup_Echavarren
 ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 31



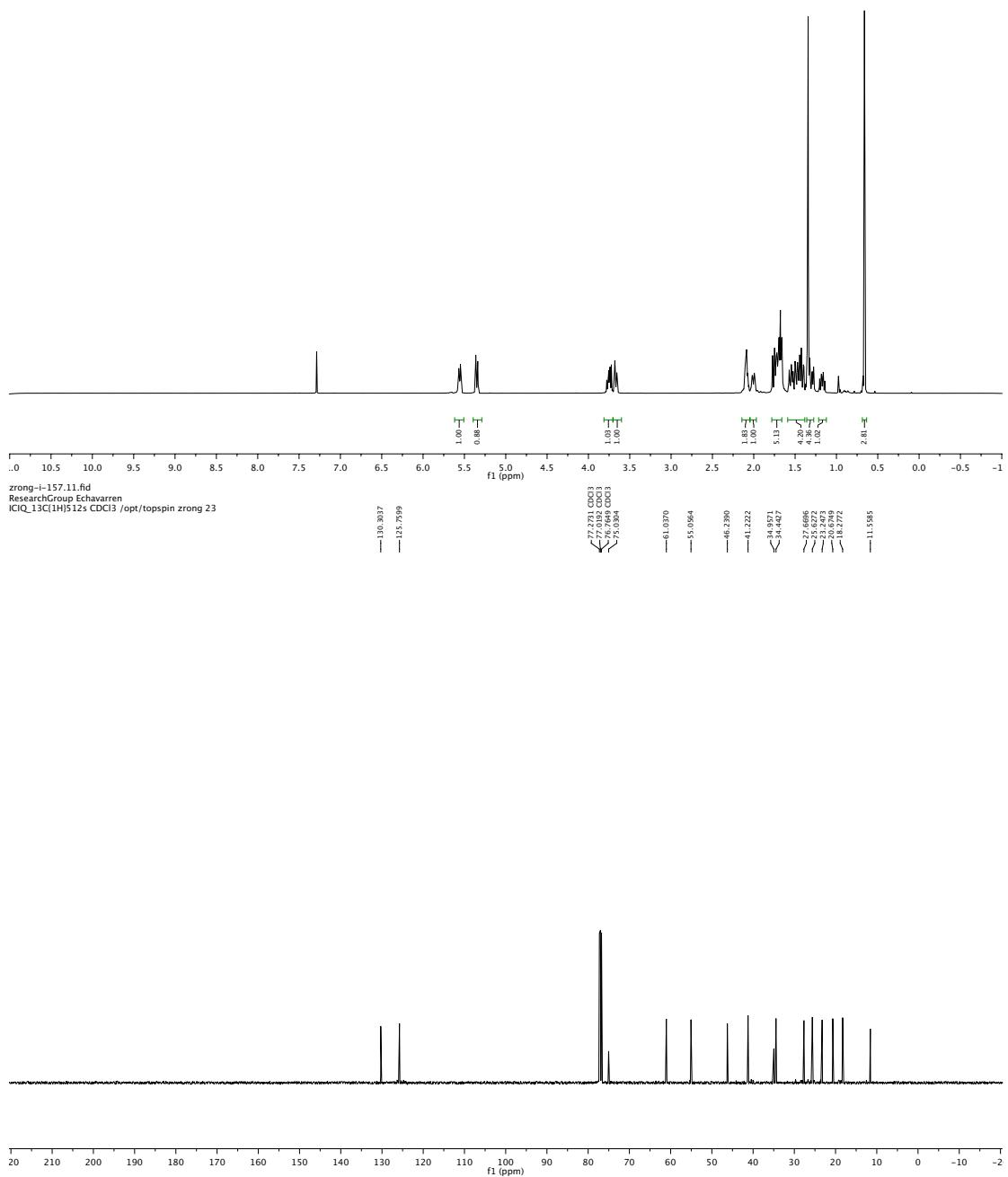


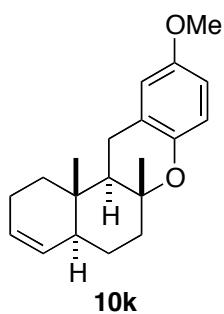
zrong-i-177s.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 89



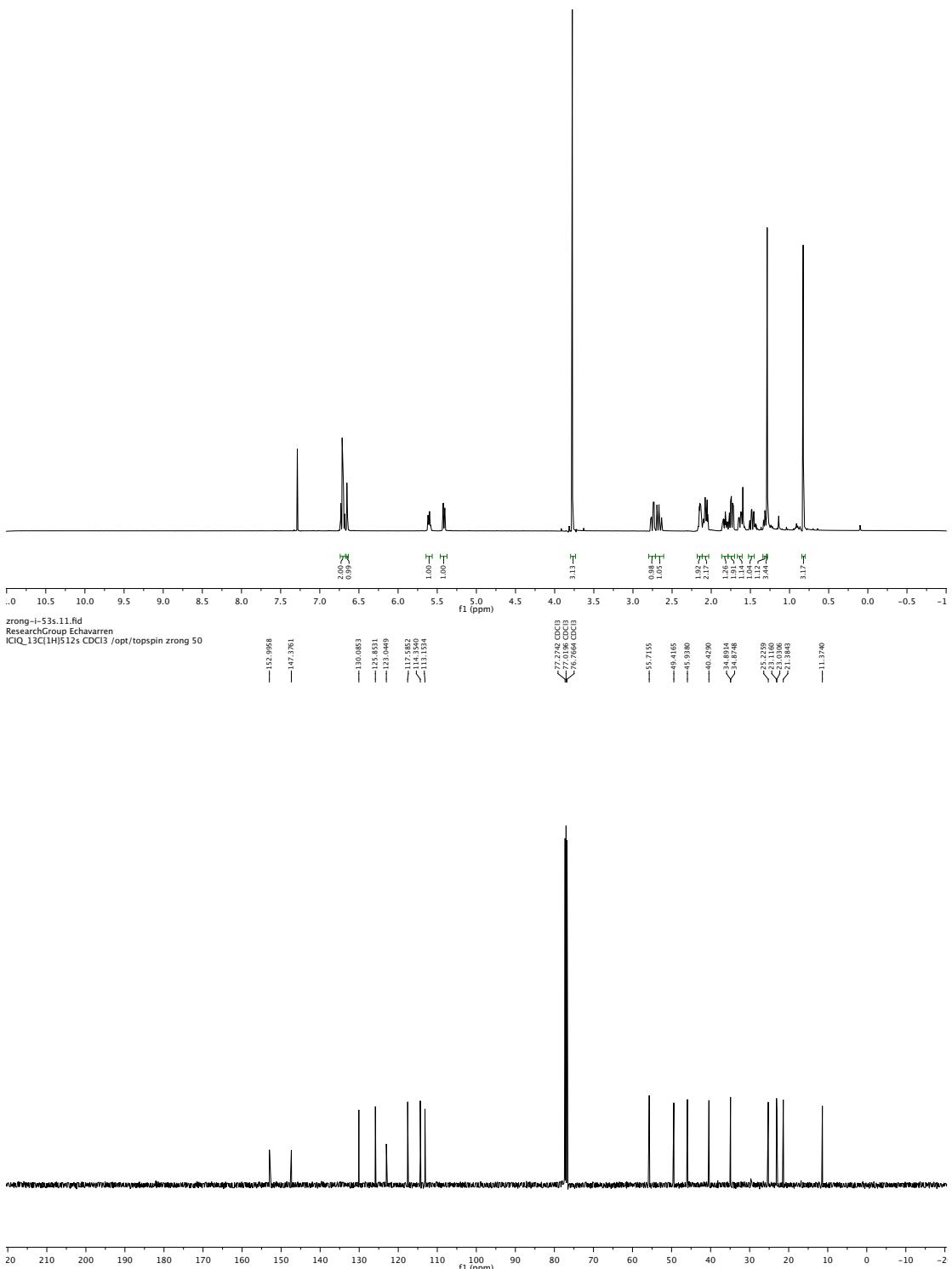


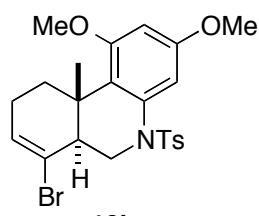
zrong-i-157.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 23



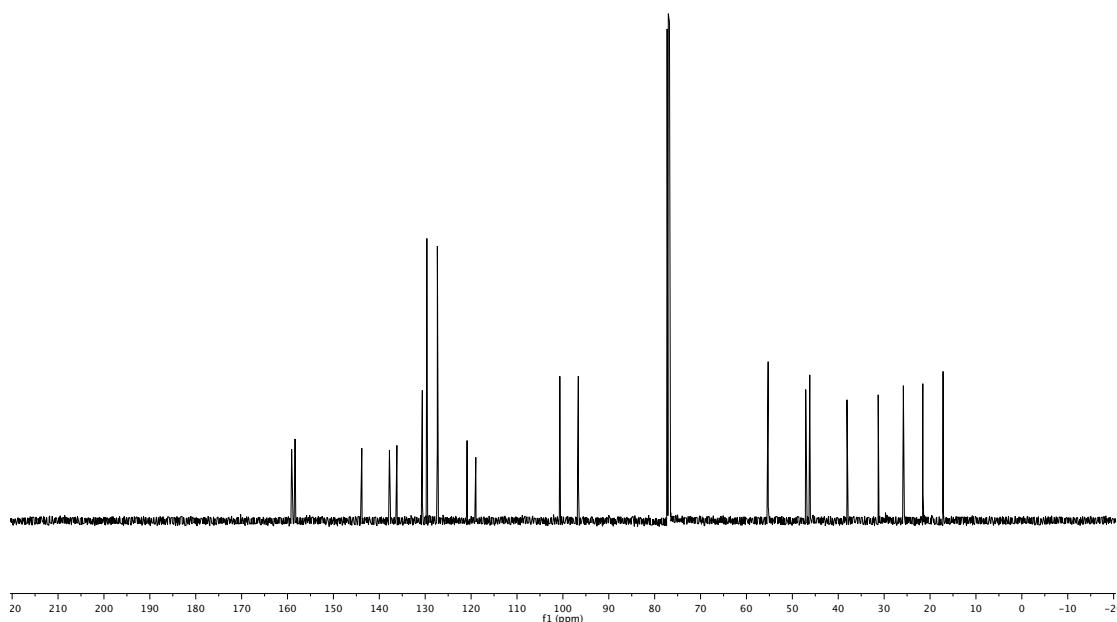
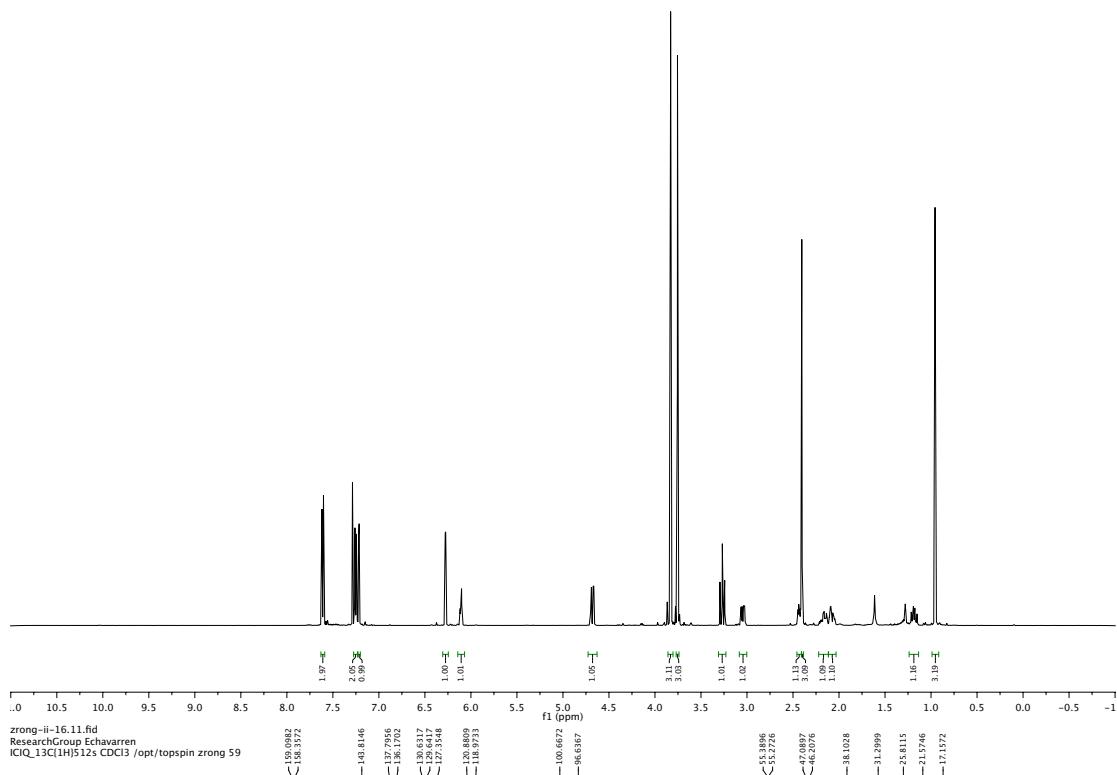


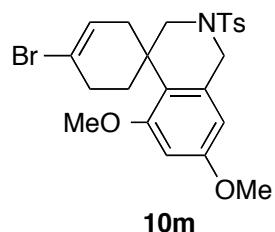
zrong-i-53s.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 50



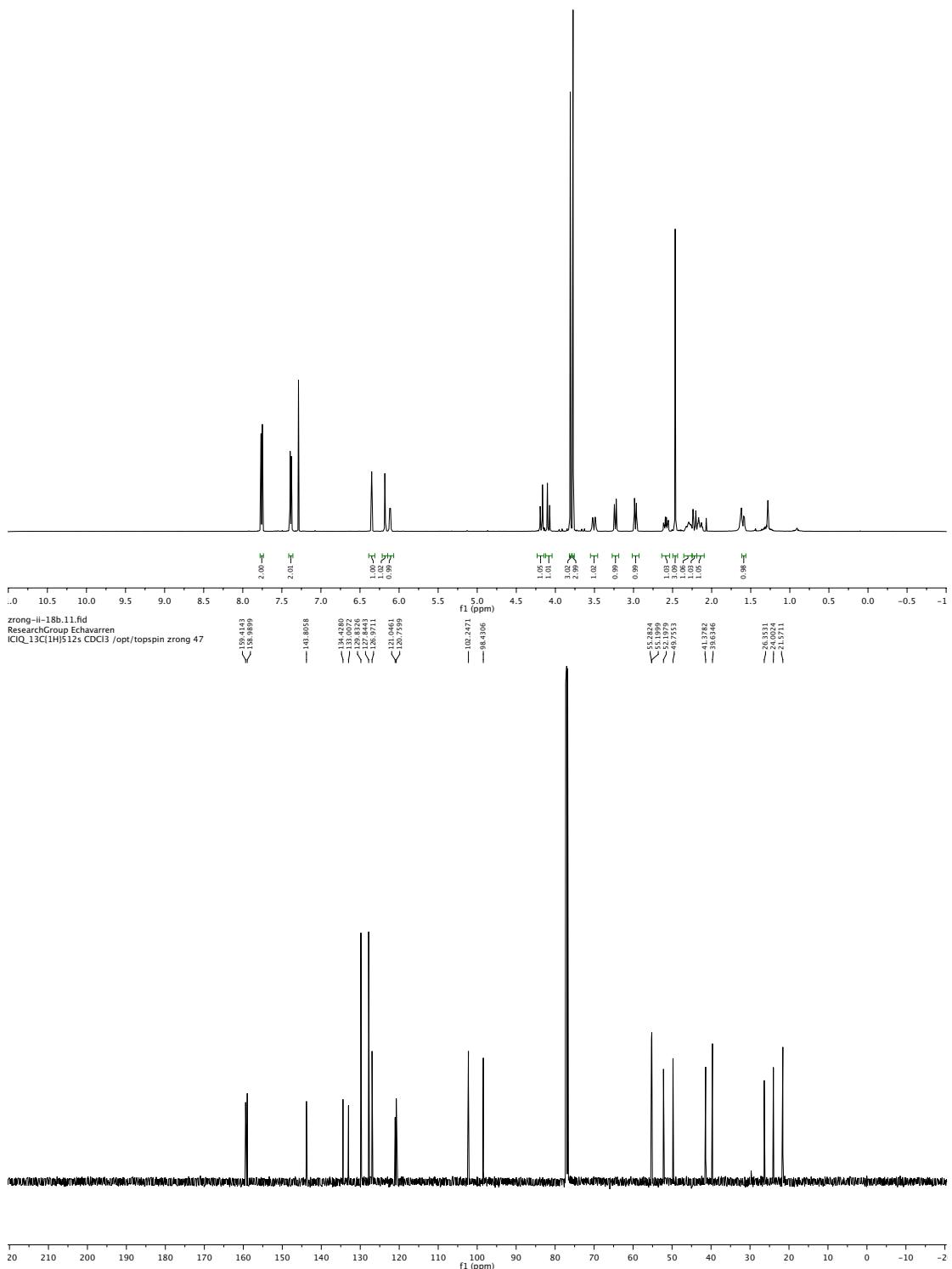


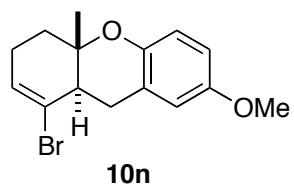
zrong-II-16.10.Rd
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 59



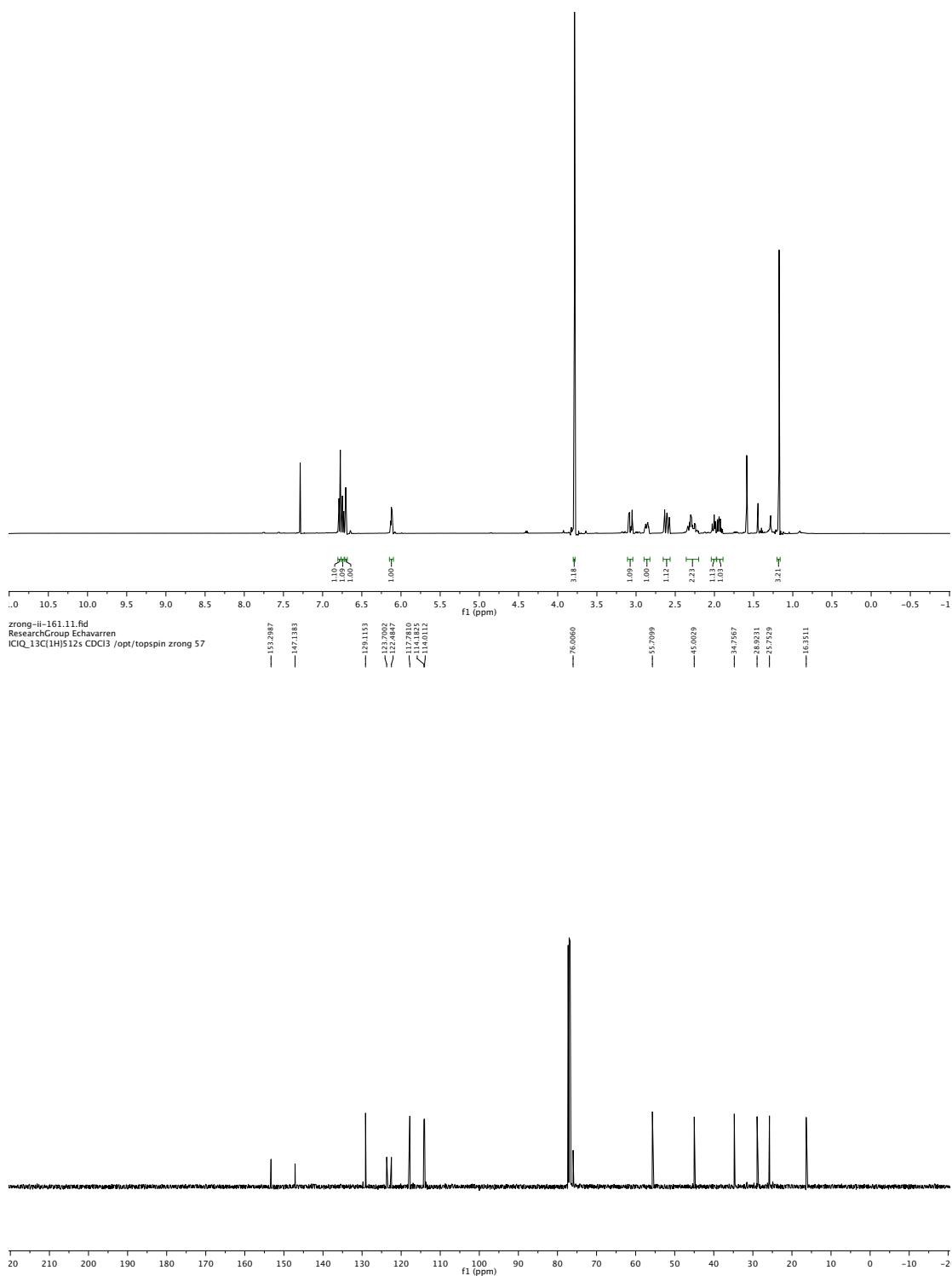


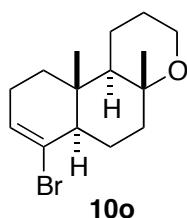
zrong-ii-18b.10.fid
 ResearchGroup_Echavarren
 ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 47



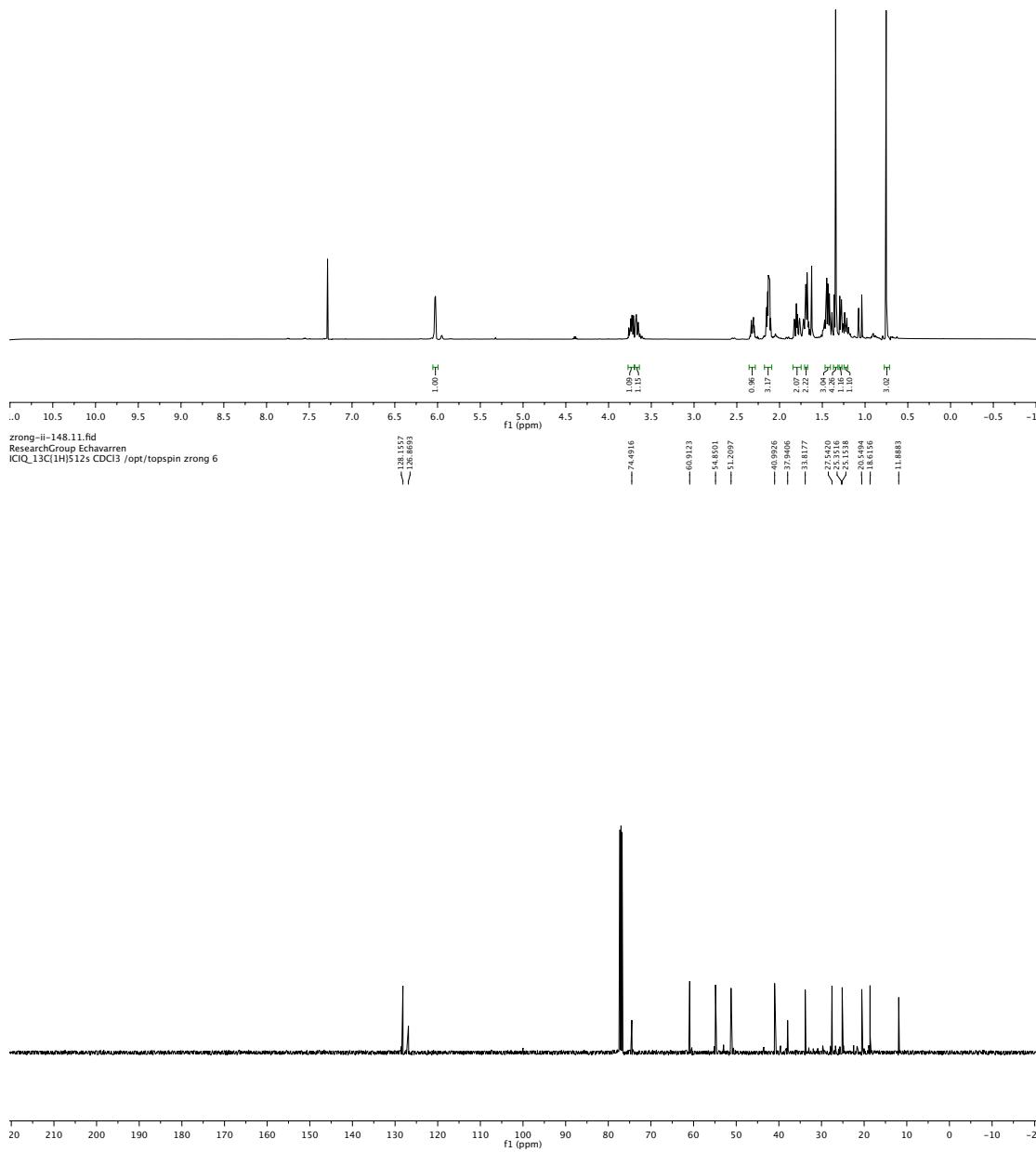


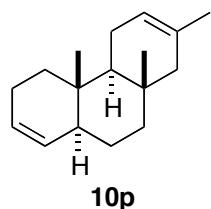
zrong-ii-161.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl₃ /opt/topspin zrong 57



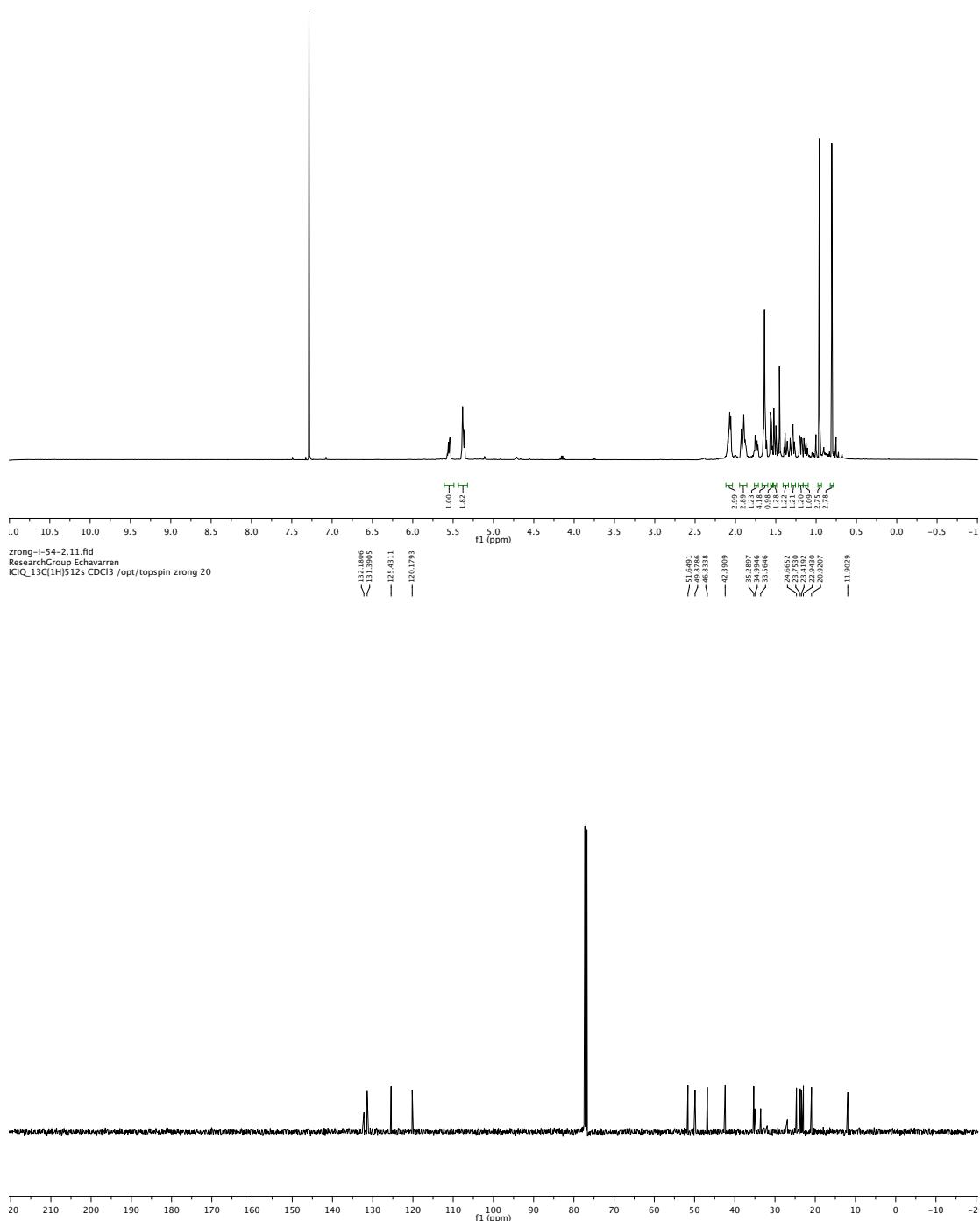


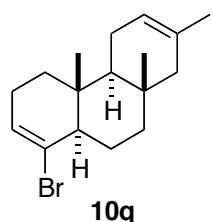
zrong-ii-148b.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 6



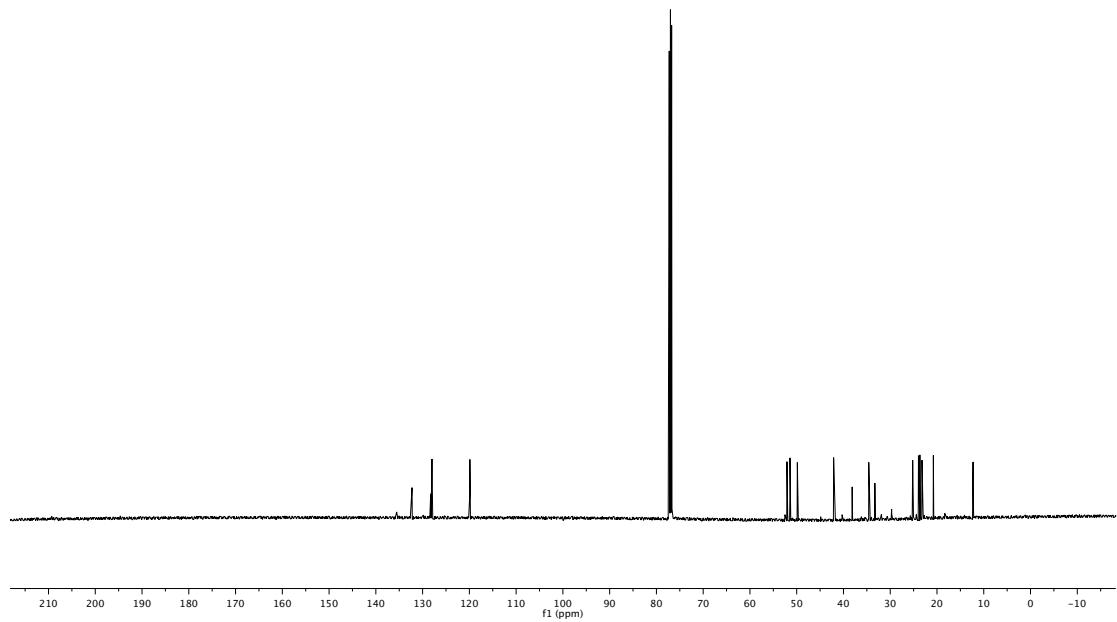
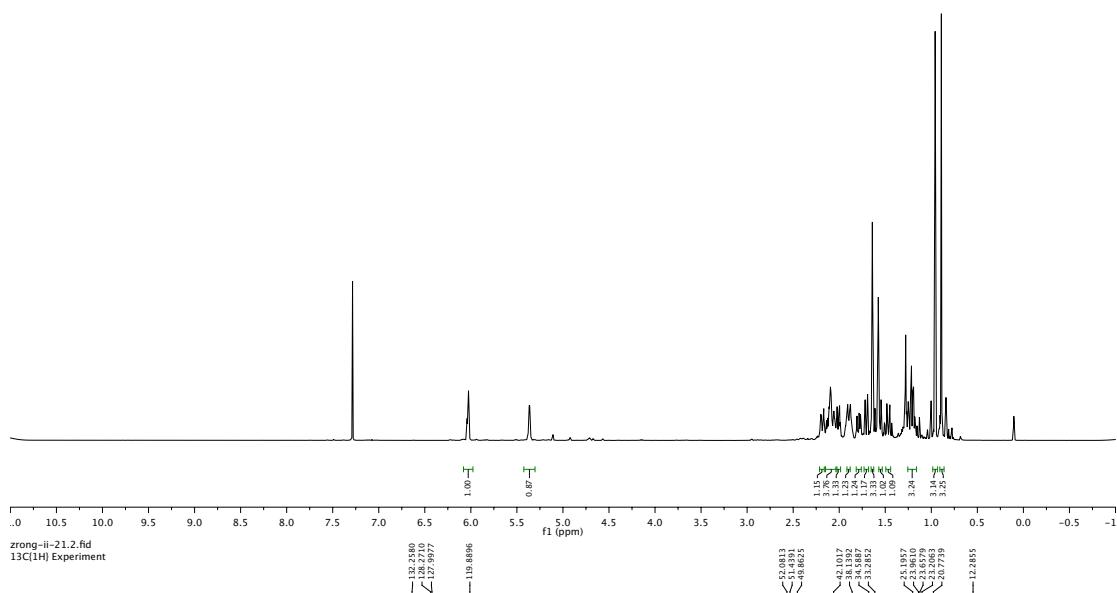


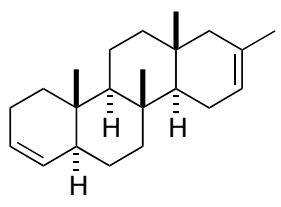
zrong-i-54-2.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 20





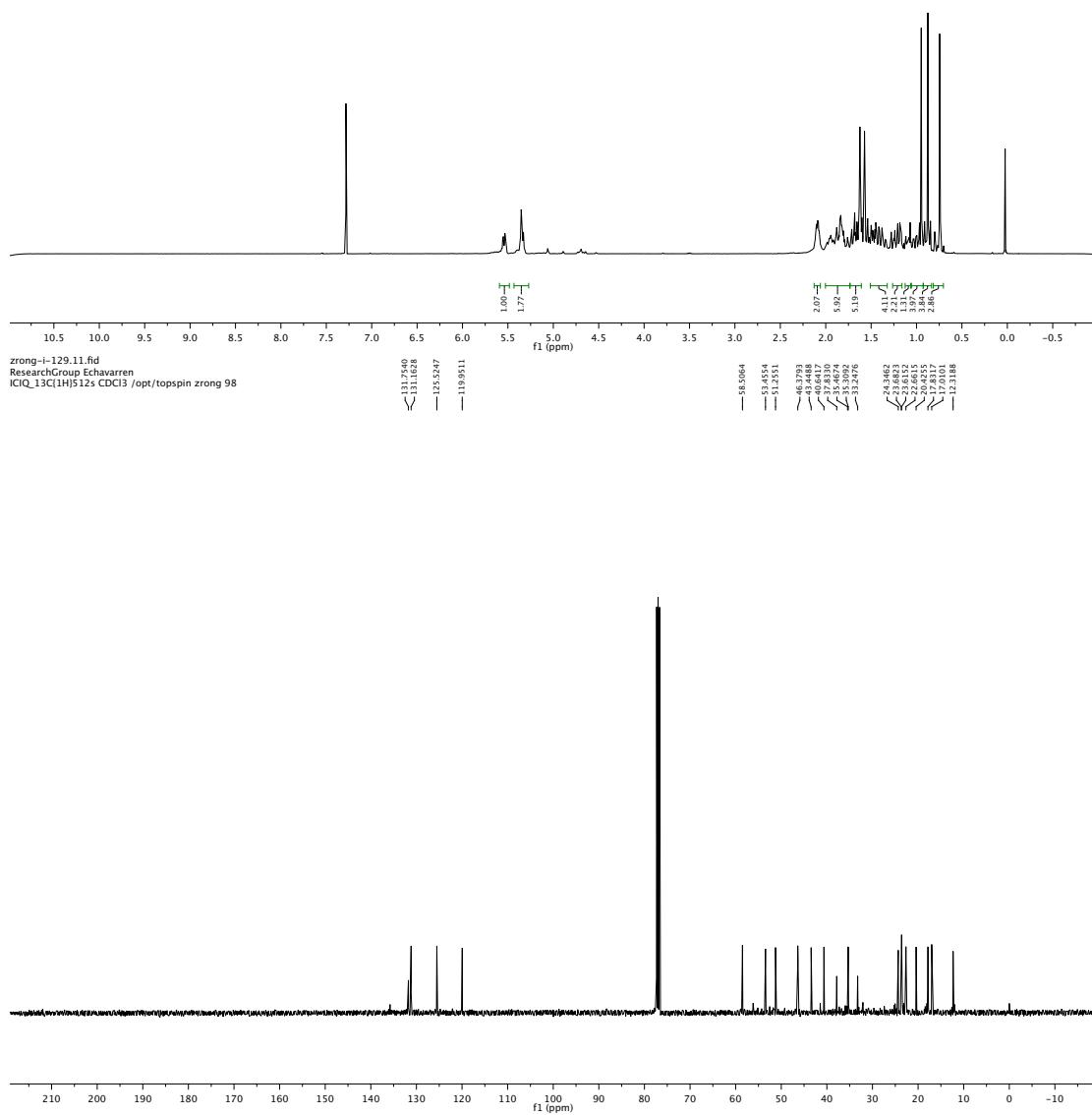
zrong-ii-21.1.fid
1H Experiment

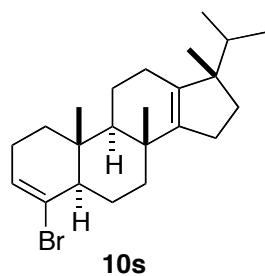




10r

zrong-i-129.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCl3 /opt/topspin zrong 98





zrong-ii-91.10.fid
ResearchGroup Echavarren
ICIQ_1H12p8s CDCI3 /opt/topspin zrong 1

