

Copper-Catalysed Enantioselective Michael Addition of Malonic Esters to β -Trifluoromethyl- α,β -Unsaturated Imines

Miguel Espinosa,^a Jorge Herrera, Gonzalo Blay,*^a Luz Cardona, M.Carmen Muñoz,^b and José R. Pedro*^a

*a Departament de Química Orgànica, Facultat de Química, Universitat de València,
C/Dr. Moliner, 50, E-46100 Burjassot (València), Spain*

*b Departament de Física Aplicada, Universitat Politècnica de València, Camí de Vera
s/n, E-46022-València, Spain*

SUPPLEMENTARY INFORMATION

Table of Contents:

General Experimental Methods	S2
General procedure for the enantioselective conjugate addition and characterisation data for compounds 3	S2
Synthetic transformations of compound 3a	S21
References	S22
NMR spectra and chiral analysis chromatograms for compounds 3	S23
Ortep plot for the X-ray structure of compounds 2a and 3a	S52
Table S-1. Enantioselective conjugate addition of dimethyl malonate to imine 2a catalysed by trivalent metal-pyBOX complexes.	S53

General Experimental Methods

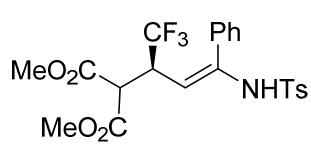
Reactions were carried out under nitrogen in round bottom flasks oven-dried overnight at 120 °C. Commercial reagents were used as purchased. Dichloromethane was distilled from CaH₂. 4 Å molecular sieves (8-12 mesh, beads Aldrich 208604) were dried at the flame under vacuum (oil pump) and stored in a closed flask and used before a week. Reactions were monitored by TLC analysis using Merck Silica Gel 60 F-254 thin layer plates. Flash column chromatography was performed on Merck silica gel 60, 0.040-0.063 mm. Melting points were determined in capillary tubes. NMR spectra were run at 300 MHz for ¹H and at 75 MHz for ¹³C NMR using residual nondeuterated solvent (CHCl₃) as internal standard (δ 7.26 and 77.0 ppm, respectively), and at 282 MHz for ¹⁹F NMR using CFCl₃ as internal standard. Chemical shifts are given in ppm. The carbon type was determined by DEPT experiments. High resolution mass spectra (ESI) were recorded on a Q-TOF spectrometer equipped with an electrospray source with a capillary voltage of 3.3 kV (ESI). Specific optical rotations were measured using sodium light (D line 589 nm). Chiral HPLC analyses were performed in a chromatograph equipped with a UV diode-array detector using chiral stationary phase columns from Daicel or from Phenomenex. *N*-Tosyl unsaturated imines **2** were prepared according to the procedure described by A. D. Smith.¹

General procedure for the enantioselective conjugate addition of methyl malonate to β -trifluoromethyl α,β -unsaturated *N*-sulfonylimines **2**

Cu(OTf)₂ (4.5 mg, 0.0125 mmol) was dried in a Schlenk tube under vacuum. **BOX7** (4.4 mg, 0.0125 mmol) was added and the tube was filled with nitrogen. CH₂Cl₂ (0.55 mL) was added via syringe and the mixture was stirred for 30 min. A solution of imine **2** (0.125 mmol) dissolved in dry CH₂Cl₂ (0.5 mL), was added via syringe, followed by 4 Å MS (110 mg) and dimethyl malonate (34 μ L, 0.3 mmol). The mixture was stirred at room temperature for the indicated time and chromatographed on silica gel eluting with hexane/EtOAc mixtures to give compounds **3**.

Racemic compounds for comparative purpose were prepared by following the same procedure, using La(OTf)₃-pyBOX (rac) at 40 °C.

Dimethyl (*S,E*)-2-(1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-phenylbut-3-en-2-yl)malonate (**3a**)



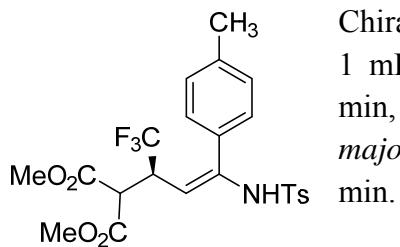
Chiral HPLC analysis: Chiralpak AD-H, hexane-*i*PrOH 80:20, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 8.4 min, *minor enantiomer (R)* tr = 14.0 min; *Z*-diastereomer: *major enantiomer* tr = 12.4 min, *minor enantiomer* tr = 9.4 min.

Major *E*-diastereomer: White solid, m.p. 159-161 °C (hexane-EtOAc); $[\alpha]_D^{20}$ -54.0 (c 1.0, CHCl₃) for the mixture of diastereomers; white solid, M.p. 153.4-160.2 °C (hexane-

EtOAc); ^1H NMR (300 MHz, CDCl_3) δ 7.76 (2H, d, $J = 8.4$ Hz, Ar), 7.40-7.27 (5H, m, Ar), 7.10 (2H, m, Ar), 6.21 (1H, s, NH), 5.57 (1H, d, $J = 10.8$ Hz, =CH), 3.73 (3H, s, MeO), 3.68 (1H, d, $J = 8.4$ Hz, CHCO_2Me), 3.64 (1H, m, CHCF_3), 3.63 (3H, s, MeO), 2.45 (3H, s, Me-Ar); ^{13}C NMR (75 MHz, CDCl_3) δ 166.7 (C), 166.4 (C), 144.2 (C), 141.3 (C), 135.9 (C), 134.0 (C), 129.6 (CH), 129.5 (CH), 128.7 (CH), 128.6 (CH), 127.7 (CH), 125.4 (C, q, $J_{\text{C}-\text{F}} = 264.8$ Hz), 102.9 (CH, q, $J_{\text{C}-\text{F}} = 2.4$ Hz), 52.93 (CH₃), 52.90 (CH₃), 51.0 (CH), 42.7 (CH, q, $J_{\text{C}-\text{F}} = 27.9$ Hz), 21.5 (CH₃); ^{19}F NMR (282 MHz, CDCl_3) $\delta = -70.1$ (s, CF₃) ppm; HRMS (ESI) m/z 486.1197, $\text{C}_{22}\text{H}_{23}\text{F}_3\text{NO}_6\text{S}$ requires 486.1193.

Minor Z-diastereomer: ^1H NMR (300 MHz, CDCl_3) δ 7.95 (s, 1H), 7.59 (2H, d, $J = 8.4$ Hz, Ar), 7.41 (2H, dd, $J = 8.1, 1.5$ Hz, Ar), 7.36-7.26 (3H, m, Ar), 7.22 (2H, d, $J = 8.4$ Hz, Ar), 5.22 (1H, d, $J = 11.1$ Hz, =CH), 3.81 (3H, s, MeO), 3.76-3.48 (2H, m, CH-CF₃, CHCO_2Me), 3.68 (3H, s, MeO), 2.39 (s, 3H); ^{19}F NMR (282 MHz, CDCl_3) $\delta = -69.8$ (s, CF₃) ppm.

Dimethyl (*S,E*)-2-(1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-(*p*-tolyl)but-3-en-2-yl)malonate (3b)

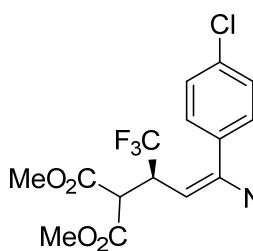


Chiral HPLC analysis: Lux Amylose-1, hexane-*iPrOH* 85:15, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 13.4 min, *minor enantiomer (R)* tr = 16.1 min; *Z*-diastereomer: *major enantiomer* tr = 14.5 min, *minor enantiomer* tr = 11.9 min.

Major E-diastereomer: White solid, m.p. 138-146 °C (hexane-EtOAc); $[\alpha]_D^{20} -38.6$ (*c* 0.95, CHCl_3) for the mixture of diastereomers; ^1H NMR (300 MHz, CDCl_3) δ 7.75 (2H, d, $J = 8.4$ Hz, Ar), 7.32 (2H, d, $J = 8.4$ Hz, Ar), 7.13 (2H, d, $J = 8.1$ Hz, Ar), 6.97 (2H, d, $J = 8.1$ Hz, Ar), 6.18 (1H, s, NH), 5.52 (1H, d, $J = 10.8$ Hz, =CH), 3.72 (3H, s, MeO), 3.67 (1H, d, $J = 8.1$ Hz, CHCO_2Me), 3.64 (1H, m, CHCF_3), 3.63 (3H, s, MeO), 2.45 (3H, s, Me-Ar), 2.33 (3H, s, Me-Ar); ^{13}C NMR (75 MHz, CDCl_3) δ 166.8 (C), 166.5 (C), 144.2 (C), 141.3 (C), 139.5 (C), 135.9 (C), 131.2 (C), 129.6 (CH), 129.5 (CH), 128.4 (CH), 127.7 (CH), 125.4 (C, q, $J_{\text{C}-\text{F}} = 249.7$ Hz), 102.6 (CH, q, $J_{\text{C}-\text{F}} = 2.0$ Hz), 52.94 (CH₃), 52.91 (CH₃), 51.0 (CH), 42.7 (CH, q, $J_{\text{C}-\text{F}} = 27.9$ Hz), 21.5 (CH₃), 21.3 (CH₃); ^{19}F NMR (282 MHz, CDCl_3) $\delta = -70.2$ (s, CF₃) ppm; HRMS (ESI) m/z 500.1356 (M+H)⁺ $\text{C}_{23}\text{H}_{25}\text{F}_3\text{NO}_6\text{S}$ requires 500.1349.

Minor Z-diastereomer: ^1H NMR (300 MHz, CDCl_3), representative signals taken from the ^1H NMR of the diastereomer mixtures, δ 7.91 (1H, s, NH), 7.86 (2H, d, $J = 8.1$ Hz, Ar), 7.60 (2H, d, $J = 8.1$ Hz, Ar), 7.25 (2H, d, $J = 8.1$ Hz, Ar), 7.22 (2H, d, $J = 8.1$ Hz, Ar), 5.16 (1H, d, $J = 10.8$ Hz, =CH), 3.82-3.60 (2H, m, CH-CF₃, CHCO_2Me), 3.76 (3H, s, MeO), 3.67 (3H, s, MeO), 2.43 (3H, s, Me-Ar), 2.39 (3H, s, Me-Ar); ^{19}F NMR (282 MHz, CDCl_3) $\delta = -69.9$ (s, CF₃) ppm.

Dimethyl (S,E)-2-(4-(4-chlorophenyl)-1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)but-3-en-2-yl)malonate (3c)

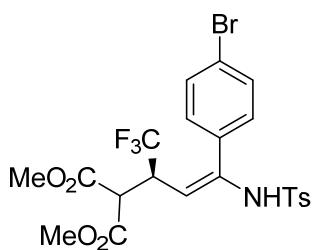


Chiral HPLC analysis: Lux Amylose-1, hexane-*i*PrOH 95:05, 1 mL/min, *E*-diastereomer: *major enantiomer* (*S*) tr = 38.4 min, *minor enantiomer* (*R*) tr = 47.9 min; *Z*-diastereomer: *major enantiomer* tr = 48.4 min, *minor enantiomer* tr = 33.9 min.

Major *E*-diastereomer: White solid, m.p. 142-150 °C (hexane-EtOAc); $[\alpha]_D^{20}$ -21.3 (c 0.95, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 7.70 (2H, d, *J* = 8.4 Hz, Ar), 7.34-7.24 (4H, m, Ar), 7.04 (2H, d, *J* = 8.7 Hz, Ar), 6.42 (1H, s, NH), 5.52 (1H, d, *J* = 10.8 Hz, =CH), 3.72 (3H, s, MeO), 3.67 (1H, d, *J* = 8.4 Hz, CHCO₂Me), 3.64 (3H, s, MeO), 3.55 (1H, m, CHCF₃), 2.44 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.6 (C), 166.5 (C), 144.3 (C), 140.4 (C), 135.9 (C), 135.6 (C), 132.3 (C), 130.2 (CH), 129.7 (CH), 129.0 (CH), 127.6 (CH), 125.4 (C, q, *J*_{C-F} = 278 Hz), 104.23 (CH, q, *J*_{C-F} = 2.5 Hz), 53.04 (CH₃), 52.99 (CH₃), 50.9 (CH), 42.6 (CH, q, *J*_{C-F} = 28.0 Hz), 21.6 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.1 (s, CF₃) ppm; HRMS (ESI) *m/z* 520.0795 (M+H)⁺, C₂₂H₂₂ClF₃NO₆S requires 520.0803.

Minor *Z*-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixtures, δ 8.01 (1H, s, NH), 7.57 (2H, d, *J* = 8.4 Hz, Ar), 7.40-7.19 (6H, m, Ar), 5.19 (1H, d, *J* = 11.4 Hz, =CH), 3.80 (3H, s, MeO), 3.76 (1H, d, *J* = 7.2 Hz, CHCO₂Me), 3.68 (3H, s, MeO), 3.51 (1H, m, CHCF₃), 2.39 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.8 (s, CF₃) ppm.

Dimethyl (S,E)-2-(4-(4-bromophenyl)-1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)but-3-en-2-yl)malonate (3d)



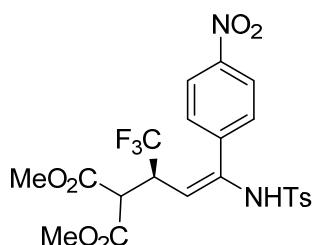
Chiral HPLC analysis: Chiralpak IC, hexane-*i*PrOH 95:05, 1 mL/min, *E*-diastereomer: *major enantiomer* (*S*) tr = 47.9 min, *minor enantiomer* (*R*) tr = 57.1 min; *Z*-diastereomer: *major enantiomer* tr = 31.9 min, *minor enantiomer* tr = 40.2 min.

Major *E*-diastereomer: Yellow solid, m.p. 130-133 °C (hexane-EtOAc); $[\alpha]_D^{20}$ -12.8 (c 1.02, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 7.71 (2H, d, *J* = 8.4 Hz, Ar), 7.45 (2H, d, *J* = 8.4 Hz, Ar), 7.31 (2H, d, *J* = 8.4 Hz, Ar), 7.04 (2H, d, *J* = 8.4 Hz, Ar), 6.29 (1H, s, NH), 5.53 (1H, d, *J* = 10.8 Hz, =CH), 3.73 (3H, s, MeO), 3.68 (1H, d, *J* = 8.4 Hz, CHCO₂Me), 3.65 (3H, s, MeO), 3.55 (1H, m, CHCF₃), 2.45 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.6 (C), 166.5 (C), 144.4 (C), 140.4 (C), 135.9 (C), 132.8 (C), 132.0 (CH), 130.4 (CH), 129.7 (CH), 127.6 (CH), 125.4 (C, q, *J*_{C-F} = 278 Hz), 124.0 (C), 104.3 (CH, q, *J*_{C-F} = 2.3 Hz), 53.07 (CH₃), 53.01

(CH₃), 50.9 (CH), 42.6 (CH, q, *J*_{C-F} = 28.0 Hz), 21.6 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.0 (s, CF₃) ppm; HRMS (ESI) *m/z* 564.0295 (M+H)⁺, C₂₂H₂₂BrF₃NO₆S requires 564.0298.

Minor Z-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixtures, δ 8.01 (1H, s, NH), 7.58 (2H, d, *J* = 8.4 Hz, Ar), 7.41 (2H, d, *J* = 8.4 Hz, Ar), 7.31 (2H, d, *J* = 8.4 Hz, Ar), 7.25 (2H, d, *J* = 8.4 Hz, Ar), 5.21 (1H, d, *J* = 11.4 Hz, =CH), 3.80 (3H, s, MeO), 3.77 (1H, d, *J* = 7.2 Hz, CHCO₂Me), 3.68 (3H, s, MeO), 3.52 (1H, m, CHCF₃), 2.40 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.8 (s, CF₃) ppm.

Dimethyl (*S,E*)-2-(1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-(4-nitrophenyl)but-3-en-2-yl)malonate (3e)

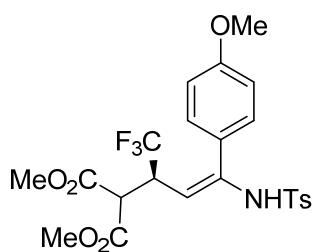


Chiral HPLC analysis: Chiraldak IC, hexane-*i*PrOH 90:10, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 60.4 min, *minor enantiomer (R)* tr = 69.2 min; *Z*-diastereomer: *major enantiomer* tr = 50.2 min, *minor enantiomer* tr = 94.8 min.

Major *E*-diastereomer: Orange oil; [α]_D²⁰ 1.1 (*c* 1.0, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 8.14 (2H, d, *J* = 9.0 Hz, Ar), 7.68 (2H, d, *J* = 8.1 Hz, Ar), 7.37-7.28 (4H, m, Ar), 6.80 (1H, s, NH), 5.58 (1H, d, *J* = 11.1 Hz, =CH), 3.73 (3H, s, MeO), 3.68 (1H, d, *J* = 8.4 Hz, CHCO₂Me), 3.65 (3H, s, MeO), 3.49 (1H, m, CHCF₃), 2.45 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.5 (C), 166.4 (C), 148.2 (C), 144.6 (C), 140.1 (C), 139.6 (C), 135.7 (C), 130.2 (CH), 129.7 (CH), 127.6 (CH), 125.2 (C, q, *J*_{C-F} = 279 Hz), 123.8 (C), 106.5 (CH, q, *J*_{C-F} = 2.1 Hz), 53.2 (CH₃), 53.1 (CH₃), 50.7 (CH), 42.6 (CH, q, *J*_{C-F} = 28.2 Hz), 21.6 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.9 (s, CF₃) ppm; HRMS (ESI) *m/z* 531.1034 (M+H)⁺, C₂₂H₂₂ClF₃N₂O₈S requires 531.1043.

Minor *Z*-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 8.19 (1H, s, NH), 8.14 (2H, d, *J* = 9.0 Hz, Ar), 7.60 (4H, m, Ar), 7.25 (2H, d, *J* = 8.0 Hz, Ar), 5.40 (1H, d, *J* = 10.8 Hz, =CH), 3.83 (3H, s, MeO), 3.80 (1H, d, *J* = 5.7 Hz, CHCO₂Me), 3.69 (3H, s, MeO), 3.49 (1H, m, CHCF₃), 2.40 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 168.2 (C), 166.9 (C), 148.2 (C), 144.4 (C), 143.2 (C), 140.0 (C), 136.5 (C), 129.7 (CH), 128.7 (CH), 126.9 (CH), 125.2 (C, q, *J*_{C-F} = 279 Hz), 123.3 (C), 114.8 (CH, q, *J*_{C-F} = 2.1 Hz), 54.0 (CH₃), 53.4 (CH₃), 50.7 (CH), 41.8 (CH, q, *J*_{C-F} = 29.0 Hz), 21.4 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.9 (s, CF₃) ppm; ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.6 (s, CF₃) ppm.

Dimethyl (*S,E*)-2-(1,1,1-trifluoro-4-(4-methoxyphenyl)-4-((4-methylphenyl)sulfonamido)but-3-en-2-yl)malonate (3f)

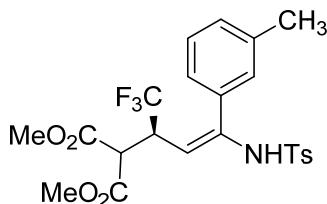


Chiral HPLC analysis: Chiralpak IC, hexane-*i*PrOH 90:10, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 44.2 min, *minor enantiomer (R)* tr = 63.8 min; *Z*-diastereomer: *major enantiomer* tr = 38.0 min, *minor enantiomer* tr = 50.9 min.

Major *E*-diastereomer: Yellow oil; [α]_D²⁰ -16.3 (c 1.0, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 7.75 (2H, d, *J* = 8.1 Hz, Ar), 7.32 (2H, d, *J* = 8.1 Hz, Ar), 7.03 (2H, d, *J* = 8.7 Hz, Ar), 6.84 (2H, d, *J* = 8.7 Hz, Ar), 6.16 (1H, s, NH), 5.48 (1H, d, *J* = 10.8 Hz, =CH), 3.79-3.66 (2H, m, CH-CF₃, CHCO₂Me), 3.80 (3H, s, MeO), 3.73 (3H, s, MeO), 3.64 (3H, s, MeO), 2.45 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.8 (C), 166.5 (C), 160.3 (C), 144.1 (C), 141.2 (C), 136.0 (C), 130.0 (CH), 129.6 (CH), 127.7 (CH), 125.6 (C, q, *J*_{C-F} = 279 Hz, CF₃), 114.1 (CH), 102.7 (C, q, *J*_{C-F} = 2.0 Hz, CF₃), 55.2 (CH₃), 52.96 (CH₃), 52.91 (CH₃), 51.1 (CH), 42.6 (CH, q, *J*_{C-F} = 27.8 Hz, CF₃), 21.6 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.2 (s, CF₃) ppm; HRMS (ESI) *m/z* 516.1294 (M+H)⁺, C₂₃H₂₅F₃NO₇S requires 516.1298.

Minor *Z*-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 7.60 (2H, d, *J* = 8.1 Hz, Ar), 7.23 (2H, d, *J* = 8.1 Hz, Ar), 6.93 (2H, d, *J* = 9.0 Hz, Ar), 6.79 (2H, d, *J* = 9.0 Hz, Ar), 5.09 (1H, d, *J* = 11.4 Hz, =CH), 2.43 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.9 (s, CF₃) ppm.

Dimethyl (*S,E*)-2-(1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-(*m*-tolyl)but-3-en-2-yl)malonate (3g)

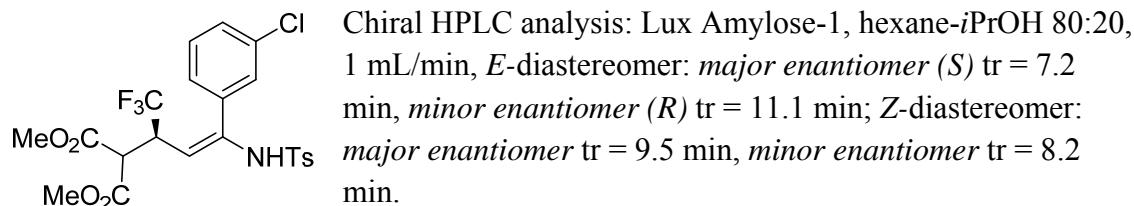


Chiral HPLC analysis: Lux Amylose-1, hexane-*i*PrOH 80:20, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 7.0 min, *minor enantiomer (R)* tr = 10.7 min; *Z*-diastereomer: *major enantiomer* tr = 9.5 min, *minor enantiomer* tr = 7.8 min.

Major *E*-diastereomer: White solid, m.p. 117-120 °C (hexane-EtOAc); [α]_D²⁰ -40.7 (c 1.0, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 7.75 (2H, d, *J* = 8.1 Hz, Ar), 7.32 (2H, d, *J* = 8.1 Hz, Ar), 7.21-7.13 (2H, m, Ar), 6.89 (1H, d, *J* = 7.5 Hz, Ar), 6.81 (1H, s, Ar), 6.18 (1H, s, NH), 5.55 (1H, d, *J* = 10.8 Hz, =CH), 3.76-3.67 (2H, m, CH-CF₃, CHCO₂Me), 3.73 (3H, s, MeO), 3.63 (3H, s, MeO), 2.45 (3H, s, Me-Ar), 2.28 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.8 (C), 166.5 (C), 144.2 (C), 141.4 (C), 138.5 (C), 136.0 (C), 134.0 (C), 130.2 (CH), 129.6 (CH), 129.1 (CH), 128.7 (CH), 127.7 (CH), 125.6 (CH), 125.4 (C, q, *J*_{C-F} = 255 Hz), 102.9 (CH, q, *J*_{C-F} = 2.0 Hz), 52.93 (CH₃), 52.91 (CH₃), 51.1 (CH), 42.6 (CH, q, *J*_{C-F} = 27.9 Hz), 21.5 (CH₃), 21.3 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.2 (s, CF₃) ppm; HRMS (ESI) *m/z* 500.1354 (M+H)⁺, C₂₃H₂₅F₃NO₆S requires 500.1349.

Minor Z-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 7.90 (1H, s, NH), 7.58 (2H, d, J = 8.4 Hz, Ar), 7.35-6.75 (6H, m, Ar), 5.22 (1H, d, J = 11.4 Hz, =CH), 3.82-3.60 (2H, m, CH-CF₃, CHCO₂Me), 3.80 (3H, s, MeO), 3.68 (3H, s, MeO), 2.43 (3H, s, Me-Ar), 2.26 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.8 (s, CF₃) ppm.

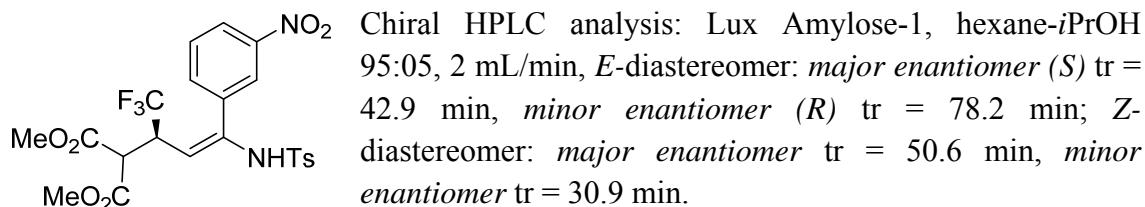
Dimethyl (S,E)-2-(4-(3-chlorophenyl)-1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)but-3-en-2-yl)malonate (3h)



Major E-diastereomer: yellow solid, m.p. 100-107 °C (hexane-EtOAc); [α]_D²⁰ -20.8 (c 0.96, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 7.71 (2H, d, J = 8.4 Hz, Ar), 7.32 (2H, d, J = 8.4 Hz, Ar), 7.31-7.17 (2H, m, Ar), 7.05 (1H, dt, J = 7.2, 1.5 Hz, Ar), 6.93 (1H, t, J = 1.5 Hz, Ar), 6.34 (1H, s, NH), 5.58 (1H, d, J = 10.8 Hz, =CH), 3.74 (3H, s, MeO), 3.68 (1H, d, J = 8.1 Hz, CHCO₂Me), 3.64 (3H, s, MeO), 3.56 (1H, m, CHCF₃), 2.45 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.6 (C), 166.4 (C), 144.4 (C), 140.1 (C), 135.8 (C), 135.5 (C), 134.5 (C), 130.0 (CH), 129.7 (CH), 129.6 (CH), 128.8 (CH), 127.6 (CH), 127.0 (CH), 125.4 (C, q, J_{C-F} = 280 Hz), 104.9 (CH, q, J_{C-F} = 2.0 Hz), 53.03 (CH₃), 52.98 (CH₃), 50.9 (CH), 42.6 (CH, q, J_{C-F} = 28.0 Hz), 21.5 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.0 (s, CF₃) ppm; HRMS (ESI) *m/z* 520.0801 (M+H)⁺, C₂₂H₂₂ClF₃NO₆S requires 520.0803.

Minor Z-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 8.00 (1H, s, NH), 7.57 (2H, d, J = 8.4 Hz, Ar), 7.33-7.20 (6H, m, Ar), 5.26 (1H, dd, J = 10.8, 0.6 Hz, =CH), 3.81 (3H, s, MeO), 3.78 (1H, d, J = 6.3 Hz, CHCO₂Me), 3.69 (3H, s, MeO), 3.56 (1H, m, CH-CF₃), 2.39 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.7 (s, CF₃) ppm.

Dimethyl (S,E)-2-(1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-(3-nitrophenyl)but-3-en-2-yl)malonate (3i)

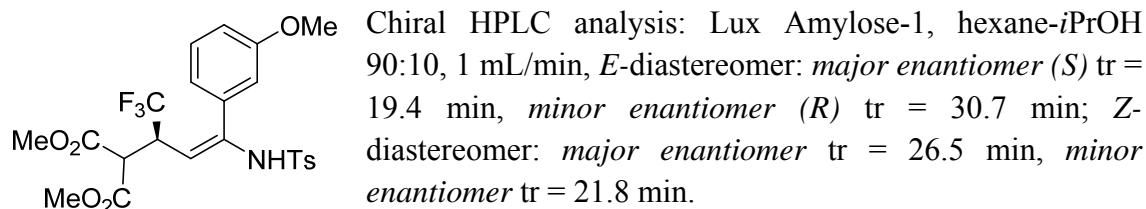


Major E-diastereomer: Yellow oil; [α]_D²⁰ -9.5 (c 0.97, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 8.18 (1H, m, Ar), 7.82 (1H, ddd, J = 7.8, 1.8, 1.2 Hz, Ar), 7.76 (1H, t, J = 1.8 Hz, Ar), 7.65 (2H, d, J = 8.0 Hz, Ar), 7.54 (1H, t, J = 8.0 Hz, Ar), 7.30 (2H, d, J = 8.0 Hz, Ar), 6.62 (1H, s, NH), 5.63 (1H, d, J = 11.1 Hz, =CH), 3.76 (3H, s, MeO), 3.67 (1H, d, J = 7.4 Hz, CHCO₂Me), 3.65 (3H, s, MeO), 3.48 (1H, m, CHCF₃), 2.44 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.5 (C), 166.4

(C), 148.1 (C), 144.7 (C), 139.5 (C), 135.7 (C), 135.2 (CH), 135.1 (C), 129.8 (CH, overlaped signals), 127.5 (CH), 125.3 (C, q, $J_{C-F} = 279$ Hz), 124.2 (CH), 124.1 (CH), 106.9 (CH, q, $J_{C-F} = 2.0$ Hz), 53.17 (CH₃), 53.12 (CH₃), 50.7 (CH), 42.6 (CH, q, $J_{C-F} = 28.2$ Hz), 21.5 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) $\delta = -69.9$ (s, CF₃) ppm; HRMS (ESI) *m/z* 531.1036 (M+H)⁺, C₂₂H₂₂ClF₃N₂O₈S requires 531.1043.

Minor Z-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 8.19 (1H, s, NH), 8.16 (1H, m, Ar), 8.11 (1H, t, $J = 1.9$ Hz, Ar), 7.63-7.53 (3H, m, Ar), 7.50 (1H, t, $J = 8.1$ Hz, Ar), 7.23 (2H, d, $J = 8.0$ Hz, Ar), 5.38 (1H, dd, $J = 10.8, 0.6$ Hz, =CH), 3.83 (3H, s, MeO), 3.81 (1H, d, $J = 6.3$ Hz, CHCO₂Me), 3.71 (3H, s, MeO), 3.58 (1H, m, CH-CF₃), 2.39 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 168.2 (C), 167.0 (C), 148.0 (C), 144.4 (C), 139.8 (C), 138.5 (C), 136.6 (C), 134.1 (CH), 129.7 (CH), 129.2 (CH), 126.9 (CH), 125.3 (C, q, $J_{C-F} = 279$ Hz), 123.9 (CH), 122.7 (CH), 113.8 (CH, q, $J_{C-F} = 2.4$ Hz), 54.0 (CH₃), 53.4 (CH₃), 50.7 (CH), 41.9 (CH, q, $J_{C-F} = 28.29$ Hz), 21.4 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) $\delta = -69.6$ (s, CF₃) ppm.

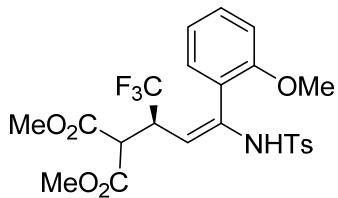
Dimethyl (S,E)-2-(1,1,1-trifluoro-4-(3-methoxyphenyl)-4-((4-methylphenyl)sulfonamido)but-3-en-2-yl)malonate (3j)



Major E-diastereomer: Yellow solid, m.p. 102-105 °C (hexane-EtOAc); $[\alpha]_D^{20} -40.3$ (*c* 0.95, CHCl₃) for the mixture of diastereomers; ¹H NMR (300 MHz, CDCl₃) δ 7.82 (2H, d, $J = 8.4$ Hz, Ar), 7.38 (2H, d, $J = 8.4$ Hz, Ar), 7.28-7.25 (1H, m, Ar), 6.93 (1H, ddd, $J = 8.4, 2.4, 1.2$ Hz, Ar), 6.71-6.69 (2H, m, Ar), 6.31 (1H, s, NH), 5.65 (1H, d, $J = 10.8$ Hz, =CH), 3.88 (1H, d, $J = 8.7$ Hz, CHCO₂Me), 3.80 (3H, s, MeO), 3.79 (3H, s, MeO), 3.76-3.75 (1H, m, CH-CF₃), 3.70 (3H, s, MeO), 2.50 (3H, s, Me-Ar); ¹³C NMR (75 MHz, CDCl₃) δ 166.8 (C), 166.6 (C), 159.6 (C), 144.2 (C), 141.1 (C), 135.9 (C), 135.3 (C), 129.9 (CH), 129.6 (CH), 127.7 (CH), 125.4 (C, q, $J_{C-F} = 257.3$ Hz), 120.6 (CH), 115.7 (CH), 113.6 (CH), 102.9 (CH, q, $J_{C-F} = 2.0$ Hz), 55.2 (CH₃), 52.95 (CH₃), 52.94 (CH₃), 51.0 (CH), 42.6 (CH, q, $J_{C-F} = 27.9$ Hz), 21.5 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) $\delta = -70.2$ (s, CF₃) ppm; HRMS (ESI) *m/z* 516.1294 (M+H)⁺, C₂₃H₂₅F₃NO₇S requires 516.1298.

Minor Z-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 7.99 (1H, s, NH), 7.65 (2H, d, $J = 8.4$ Hz, Ar), 7.37-7.32 (1H, m, Ar), 7.06 (dt, $J = 7.8, 1.2$ Hz, Ar), 6.75-6.65 (2H, m, Ar), 5.31 (1H, d, $J = 11.4$ Hz, =CH), 3.89-3.67 (2H, m, CH-CF₃, CHCO₂Me), 3.86 (3H, s, MeO), 3.80 (3H, s, MeO), 3.75 (3H, s, MeO), 2.45 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) $\delta = -69.8$ (s, CF₃) ppm.

Dimethyl (S,E)-2-(1,1,1-trifluoro-4-(2-methoxyphenyl)-4-((4-methylphenyl)sulfonamido)but-3-en-2-yl)malonate (3k)

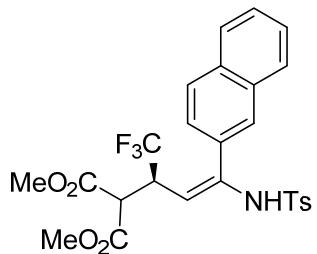


Chiral HPLC analysis: Chiralpak AD-H, hexane-*iPrOH* 90:10, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 21.6 min, *minor enantiomer (R)* tr = 47.2 min; *Z*-diastereomer: *major enantiomer* tr = 38.3 min, *minor enantiomer* tr = 32.7 min.

Major *E*-diastereomer: Yellow solid, m.p. 129-133 °C (hexane-EtOAc); $[\alpha]_D^{20} -32.2$ (*c* 0.92, CHCl_3) for the mixture of diastereomers; ^1H NMR (300 MHz, CDCl_3) δ 7.72 (2H, d, *J* = 8.4 Hz, Ar), 7.28-7.25 (2H, m, Ar), 7.03-7.00 (2H, m, Ar), 6.89 (1H, dt, *J* = 7.5, 1.2 Hz, Ar), 6.82 (1H, dd, *J* = 8.4, 1.2 Hz, Ar), 6.18 (1H, s, NH), 5.69 (1H, d, *J* = 10.8 Hz, =CH), 3.75 (3H, s, MeO), 3.69-3.55 (2H, m, CH-CF₃, CHCO_2Me), 3.65 (3H, s, MeO), 3.60 (3H, s, MeO), 2.42 (3H, s, Me-Ar); ^{13}C NMR (75 MHz, CDCl_3) δ 166.8 (C), 166.6 (C), 156.7 (C), 143.8 (C), 138.8 (C), 136.1 (C), 131.2 (C), 131.0 (C), 129.3 (CH), 128.9 (CH), 127.9 (CH), 127.1 (CH), 125.0 (C, q, $J_{\text{C-F}} = 258.8$ Hz), 120.6 (CH), 111.0 (CH), 113.6 (CH), 105.6 (CH, q, $J_{\text{C-F}} = 2.0$ Hz), 55.1 (CH₃), 52.9 (CH₃), 51.1 (CH), 42.8 (CH, q, $J_{\text{C-F}} = 27.8$ Hz), 21.5 (CH₃); ^{19}F NMR (282 MHz, CDCl_3) δ = -70.2 (s, CF₃) ppm; HRMS (ESI) *m/z* 516.1302 (M+H)⁺, $\text{C}_{23}\text{H}_{25}\text{F}_3\text{NO}_7\text{S}$ requires 516.1298.

Minor *Z*-diastereomer: ^1H NMR (300 MHz, CDCl_3), representative signals taken from the ^1H NMR of the diastereomer mixture, δ 7.81 (2H, d, *J* = 8.1 Hz, Ar), 7.38-6.49 (7H, m, Ar, NH), 5.44 (1H, d, *J* = 10.8 Hz, =CH), 3.82-3.60 (2H, m, CH-CF₃, CHCO_2Me), 3.83 (3H, s, MeO), 3.73 (3H, s, MeO), 3.55 (3H, s, MeO), 2.31 (3H, s, Me-Ar); ^{19}F NMR (282 MHz, CDCl_3) δ = -69.5 (s, CF₃) ppm.

Dimethyl (S,E)-2-(1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-(naphthalen-2-yl)but-3-en-2-yl)malonate (3l)



Chiral HPLC analysis: Lux Amylose-1, hexane-*iPrOH* 80:20, 1 mL/min, *E*-diastereomer: *major enantiomer (S)* tr = 11.3 min, *minor enantiomer (R)* tr = 13.8 min; *Z*-diastereomer: *major enantiomer* tr = 12.3 min, *minor enantiomer* tr = 9.4 min

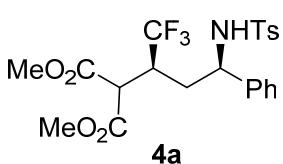
Major *E*-diastereomer: Yellow solid, m.p. 98-103 °C (hexane-EtOAc); $[\alpha]_D^{20} 1.0$ (*c* 0.96, CHCl_3) for the mixture of diastereomers; ^1H NMR (300 MHz, CDCl_3) δ 7.80-7.74 (5H, m, Ar), 7.59-7.58 (1H, m, Ar), 7.53-7.49 (2H, m, Ar), 7.30 (2H, d, *J* = 8.1 Hz Ar), 7.14 (1H, dd, *J* = 8.1, 1.8 Hz, Ar), 6.38 (1H, s, NH), 5.65 (1H, d, *J* = 10.8 Hz, =CH), 3.82-3.69 (2H, m, CH-CF₃, CHCO_2Me), 3.76 (3H, s, MeO), 3.60 (3H, s, MeO), 2.44 (3H, s, Me-Ar); ^{13}C NMR (75 MHz, CDCl_3) δ 166.7 (C), 166.5 (C), 144.2 (C), 141.4 (C), 136.0 (C), 133.3 (C), 132.8 (C), 131.2 (C), 129.6 (CH), 128.64 (CH), 128.57 (CH), 128.3 (CH), 127.7 (CH), 127.1 (CH), 126.6 (CH), 125.5 (CH), 125.4 (C, q, $J_{\text{C-F}} = 264.8$ Hz), 123.7 (CH), 104.0 (CH, q, $J_{\text{C-F}} = 2.0$ Hz), 53.0 (CH₃), 52.9 (CH₃), 51.1 (CH), 42.7

(CH, q, $J_{C-F} = 28.5$ Hz), 21.5 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.0 (s, CF₃) ppm; HRMS (ESI) *m/z* 536.1346 (M+H)⁺, C₂₆H₂₅F₃NO₆S requires 536.1349.

Minor Z-diastereomer: ¹H NMR (300 MHz, CDCl₃), representative signals taken from the ¹H NMR of the diastereomer mixture, δ 8.05 (1H, s, NH), 7.87-6.94 (11H, m, Ar), 5.37 (1H, d, $J = 11.1$ Hz, =CH), 3.82 (3H, s, MeO), 3.80-3.60 (2H, m, CH-CF₃, CHCO₂Me), 3.68 (3H, s, MeO), 2.34 (3H, s, Me-Ar); ¹⁹F NMR (282 MHz, CDCl₃) δ = -69.6 (s, CF₃) ppm.

Synthetic transformations of compound 3a

Dimethyl 2-((2*S*,4*R*)-1,1,1-trifluoro-4-((4-methylphenyl)sulfonamido)-4-phenylbutan-2-yl)malonate (4a)

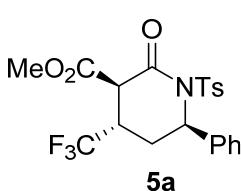


To a sample of compound (*S,E*)-**3a** (52.0 mg, 0.11 mmol, *E/Z* 96:4, ee = 89%/69%), dissolved in dry CH₂Cl₂ (3.3 mL) under nitrogen atmosphere was added triethylsilane (50 µL, 0.428 mmol) followed by BF₃·Et₂O (67 µL, 0.471 mmol). After stirring for 48 h at room temperature, the mixture was chromatographed on silica gel eluting with hexane:EtOAc (80:20) to give 48.1 mg (92%) of compound **4a**, as a c.a. 88:12 of two diastereomers. Chiral HPLC analysis: Lux Amylose-1, hexane-*i*PrOH 90:10, 1 mL/min, (**2*S,4R***) **major diastereomer** (ee = 87%), **major enantiomer** tr = 16.4 min, **minor enantiomer** tr = 15.0 min; (**2*S,4S***) **minor diastereomer** unresolved tr = 8.3 min. Chiralpak IC, hexane-*i*PrOH 95:05, 2 mL/min, (**2*S,4R***) **major diastereomer**, tr > 120 min; (**2*S,4S***) **minor diastereomer** (ee = 89%) **major enantiomer** tr = 37.6 min, **minor enantiomer** tr = 35.8 min;

(2*S,4R*)-4a (major): colorless oil; [α]_D²⁰ 7.8 (c 0.97, CHCl₃) for the diastereomer mixture; ¹H NMR (300 MHz, CDCl₃) δ 7.52 (2H, d, *J* = 8.1 Hz, Ar), 7.20-7.13 (3H, m, Ar), 7.10 (2H, d, *J* = 8.1 Hz, Ar), 7.02-7.6.90 (2H, m, Ar), 5.13 (1H, d, *J* = 8.1 Hz, NH), 4.47 (1H, q, *J* = 7.8 Hz, CHPh), 3.73 (3H, s, MeO), 3.69 (1H, d, *J* = 5.4 Hz, CHCO₂Me), 2.83 (1H, m, CHCF₃), 2.34 (3H, s, Me-Ar), 2.32-2.10 (2H, m, CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 167.1 (C), 167.0 (C), 143.1 (C), 138.8 (C), 137.2 (C), 129.3 (CH), 128.6 (CH), 128.5 (C), 127.9 (CH), 127.0 (CH), 126.8 (CH), 126.6 (C, q, *J*_{C-F} = 278 Hz), 56.4 (CH), 53.1 (CH₃), 52.8 (CH₃), 49.9 (CH), 40.0 (CH, q, *J*_{C-F} = 26.8 Hz), 33.4 (CH₂), 21.4 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -68.5 (s, CF₃) ppm; HRMS (ESI) *m/z* 488.1357 (M+H)⁺, C₂₂H₂₅F₃NO₆S requires 488.1349.

(2*S,4S*)-4a (minor): ¹H NMR (300 MHz, CDCl₃), representative signals taken from the diastereomer mixture δ 7.58 (2H, d, *J* = 8.4 Hz, Ar), 7.40-6.90 (7H, m, Ar), 5.95 (1H, d, *J* = 6.9 Hz, NH), 4.45 (1H, m, CHPh), 3.81 (3H, s, MeO), 3.72 (3H, s, MeO), 2.83 (1H, m, CHCF₃), 2.34 (3H, s, Me-Ar), 2.32-2.10 (2H, m, CH₂); ¹⁹F NMR (282 MHz, CDCl₃) δ = -70.1 (s, CF₃) ppm.

Methyl (**3*R,4S,6R***)-2-oxo-6-phenyl-1-tosyl-4-(trifluoromethyl)piperidine-3-carboxylate.

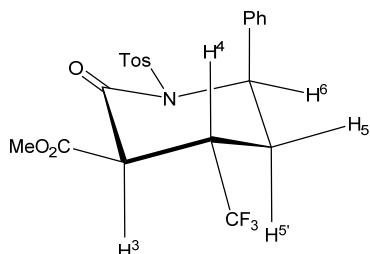


A 25% solution of tetraethylammonium hydroxyde in MeOH (24 µL, 0.14 mmol) was added to a solution of compound **4a** (28.0 mg, 0.037 mmol, ee = 87%) in dimethylsulfoxide (1.6 mL) under nitrogen, and the reaction flask was introduced in a bath at 80 °C. After 14 h, the reaction mixture was diluted with EtOAc (75 mL), washed with water (5 × 5 mL), brine (5 mL), and dried over MgSO₄. Purification by column chromatography eluting with hexane:EtOAc (80:20) gave 13.2 mg (78%) of compound **5a**. Chiral HPLC analysis: Lux Amylose-1, hexane-*i*PrOH 90:10, 1 mL/min,

major enantiomer tr = 24.0 min, *minor enantiomer* tr = 22.1 min. White solid, m.p. 177-179 °C (hexane-EtOAc); $[\alpha]_D^{20}$ -4.5 (*c* 1.0, CHCl₃, ee = 87%); ¹H NMR (300 MHz, CDCl₃) δ 7.56 (2H, d, *J* = 8.5 Hz, Ar), 7.40-7.32 (3H, m, Ar), 7.20-7.14 (4H, m, Ar), 5.93 (1H, t, *J* = 3.8 Hz, CH-Ph), 3.78 (3H, s, OMe), 3.65 (1H, d, *J* = 11.4 Hz, CHCO₂Me), 3.11 (1H, m, CHCF₃), 2.40 (3H, s, Me-Ar), 2.35-2.28 (2H, m, CH₂); ¹³C NMR (75 MHz, CDCl₃) δ 168.3 (C), 164.3 (C), 145.4 (C), 138.1 (C), 134.5 (C), 129.7 (CH), 129.0 (CH), 128.5 (C), 126.5 (CH), 125.6 (C, q, *J*_{C-F} = 278 Hz), 58.3 (CH), 53.4 (CH₃), 52.8 (CH₃), 50.2 (CH), 37.1 (CH, q, *J*_{C-F} = 28.5 Hz), 29.9 (CH₂), 21.7 (CH₃); ¹⁹F NMR (282 MHz, CDCl₃) δ = -73.1 (s, CF₃) ppm; HRMS (ESI) *m/z* 456.1087 (M+H)⁺, C₂₁H₂₁F₃NO₅S requires 456.1077.

Determination of the relative stereochemistry of compounds **4a** and **5a**

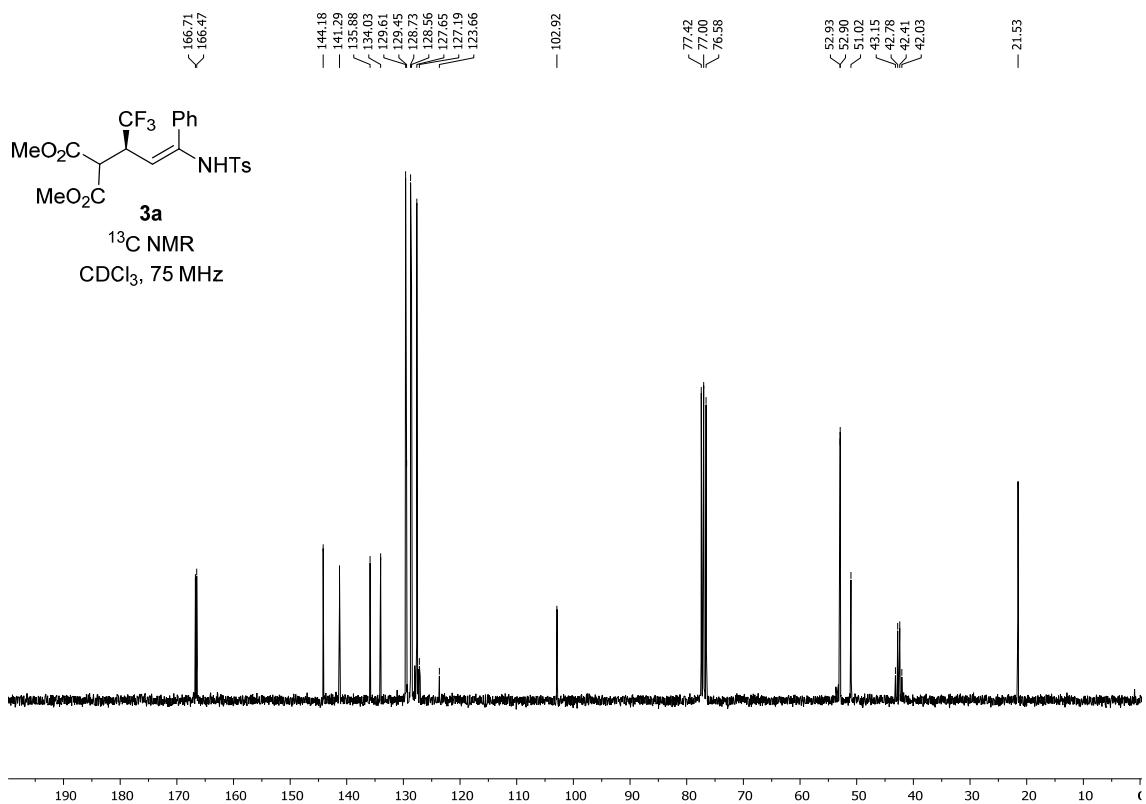
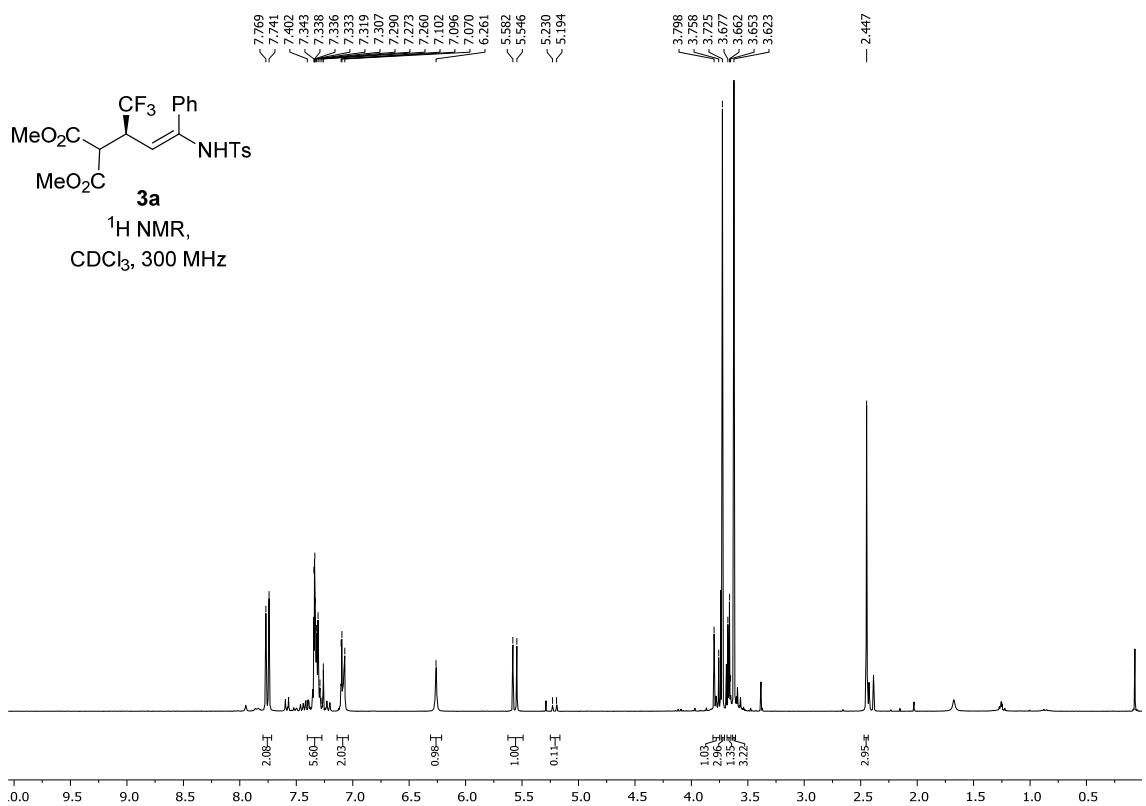
The relative stereochemistry of compound **5a**, and hence, of its precursor, the major diastereomer of compound **4a**, was established considering the coupling constants of the ring-attached protons (see figure):

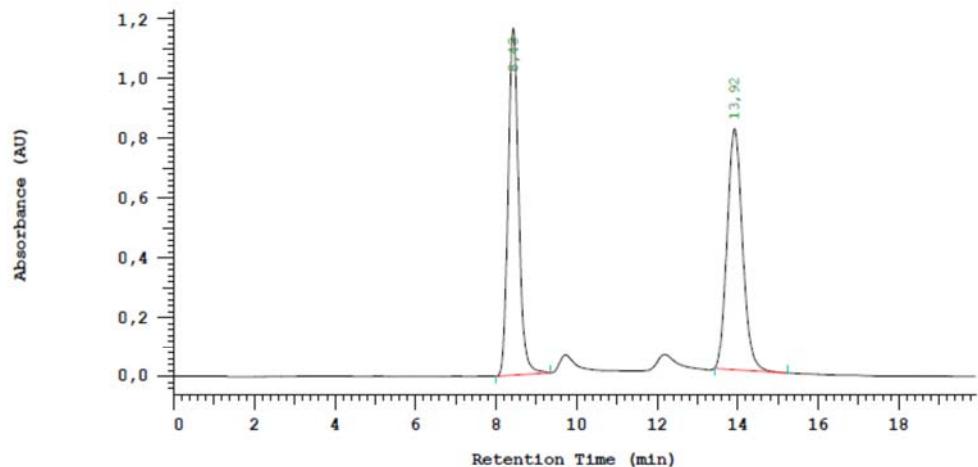
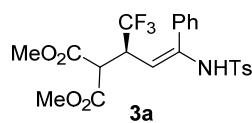


H6 5.93 ppm (t) *J*_{6,5} = 3.8 Hz (eq-eq), *J*_{6,5'} = 3.8 Hz (eq-ax)
 H3 3.65 ppm (d) *J*_{3,4} = 11.4 Hz (ax-ax),

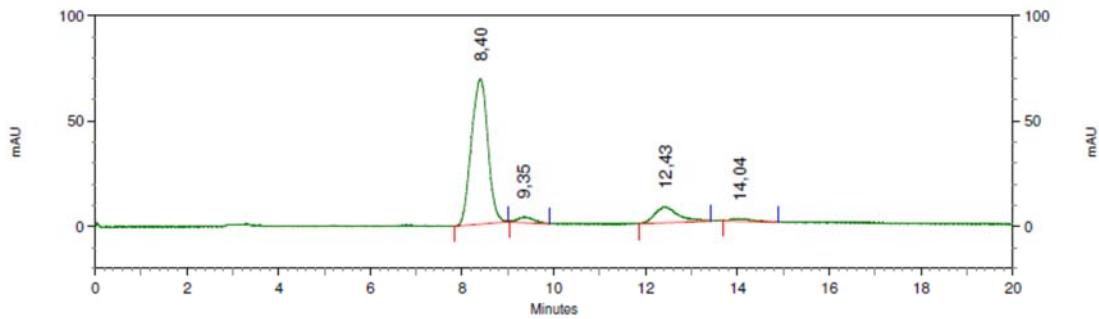
Figure S1. Coupling constants in compound **5a**

1 D. G. Stark, L. C. Morrill, P.-P. Yeh, A. M.Z. Slawin, T. J. C. O'Riordan, A. D. Smith, *Angew. Chem. Int. Ed.* **2013**, 52, 11642.

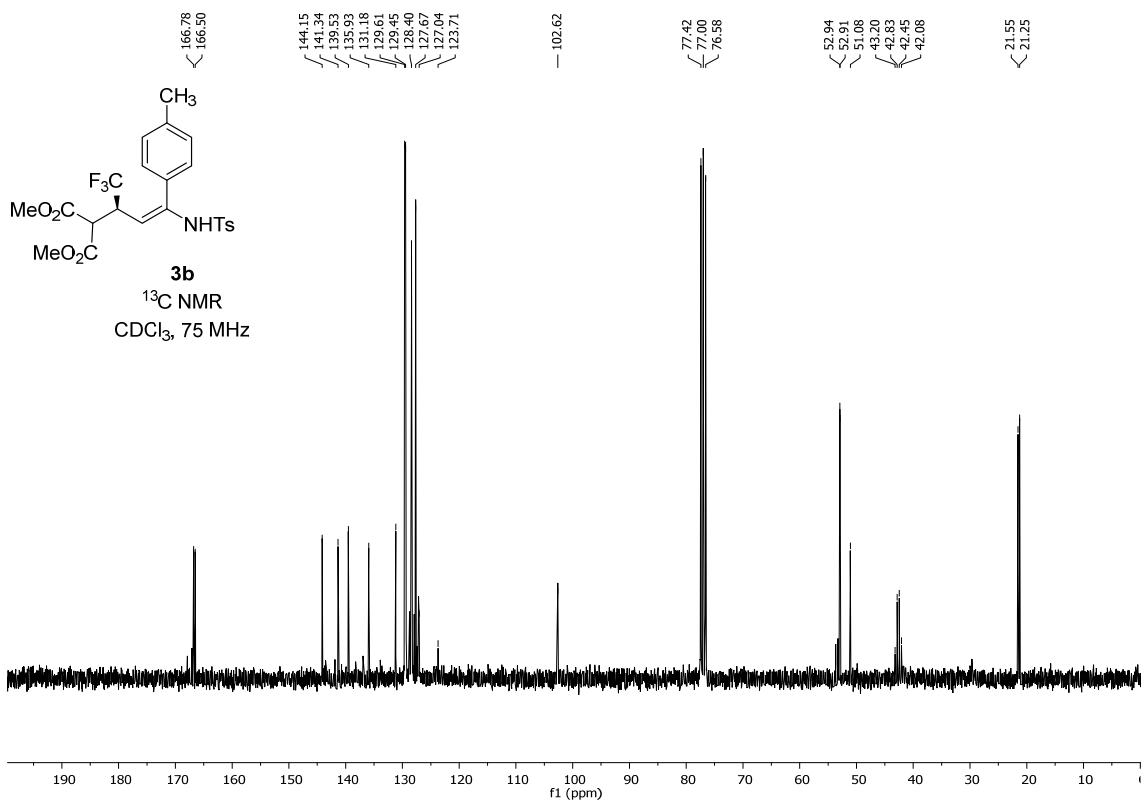
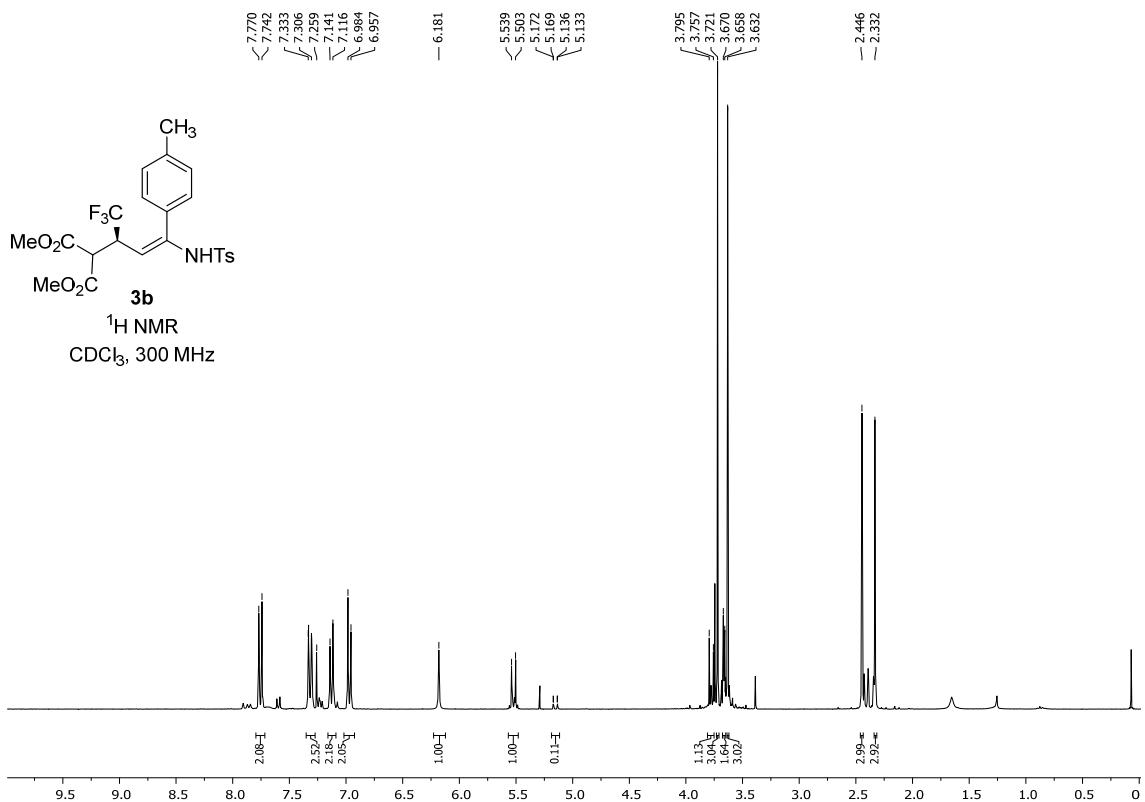


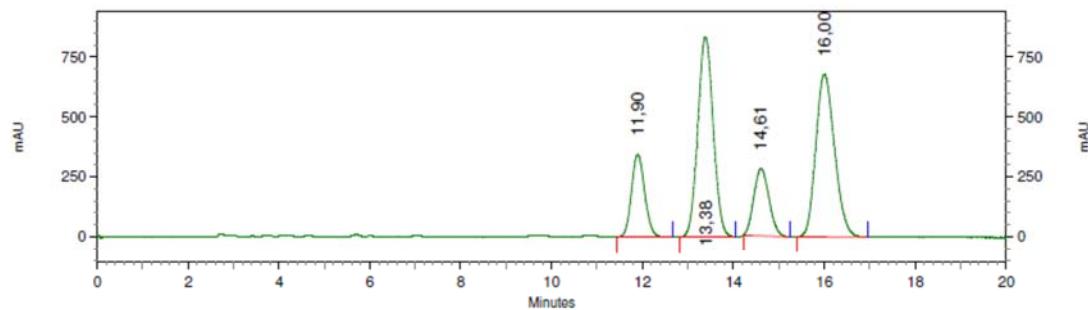
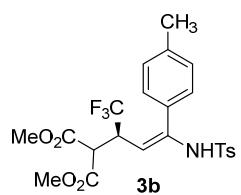


No.	RT	Area	Area %	Name
1	8,43	10584244	49,555	
2	13,92	10774280	50,445	
21358524			100,000	



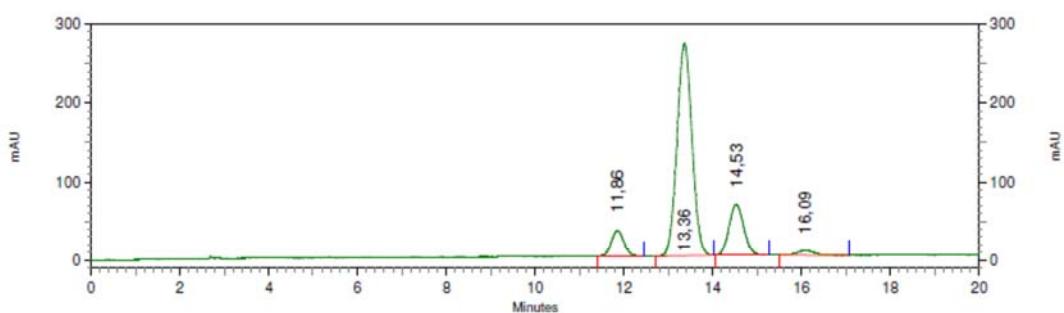
6: 280 nm, 4 nm Results		
Retention Time	Area	Area Percent
8,40	6869947	81,639
9,35	281947	3,351
12,43	1096977	13,036
14,04	166126	1,974





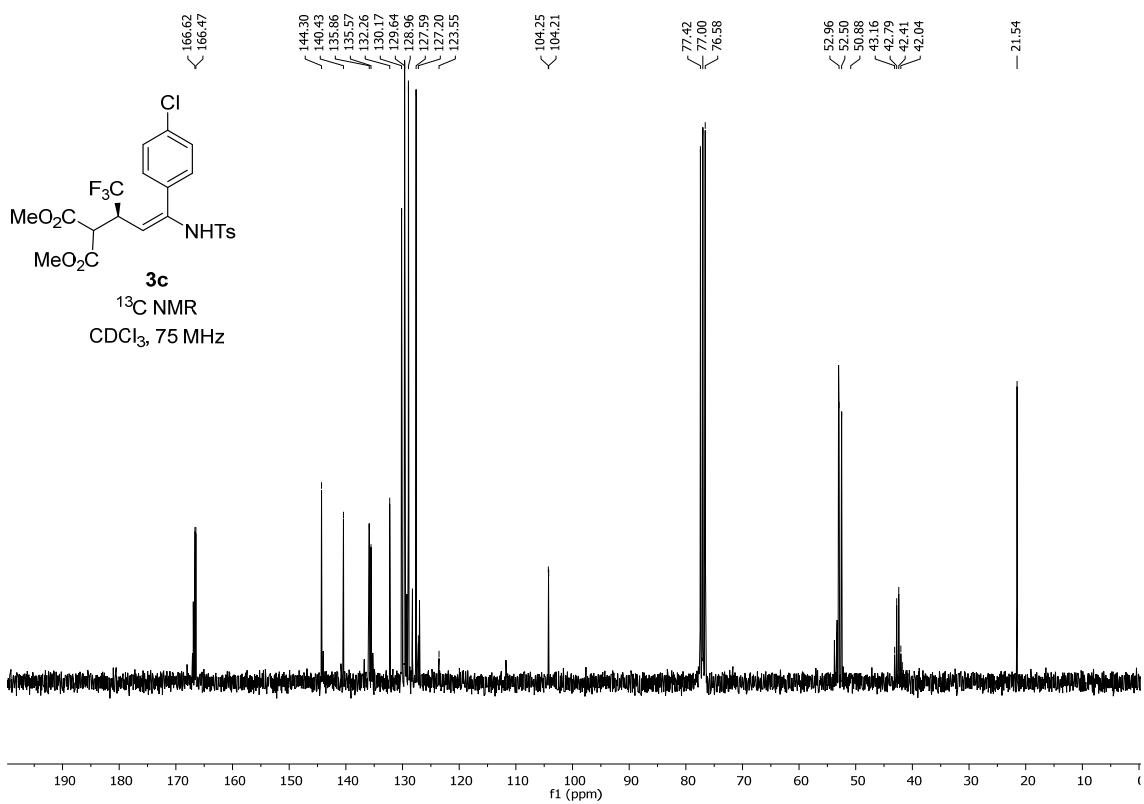
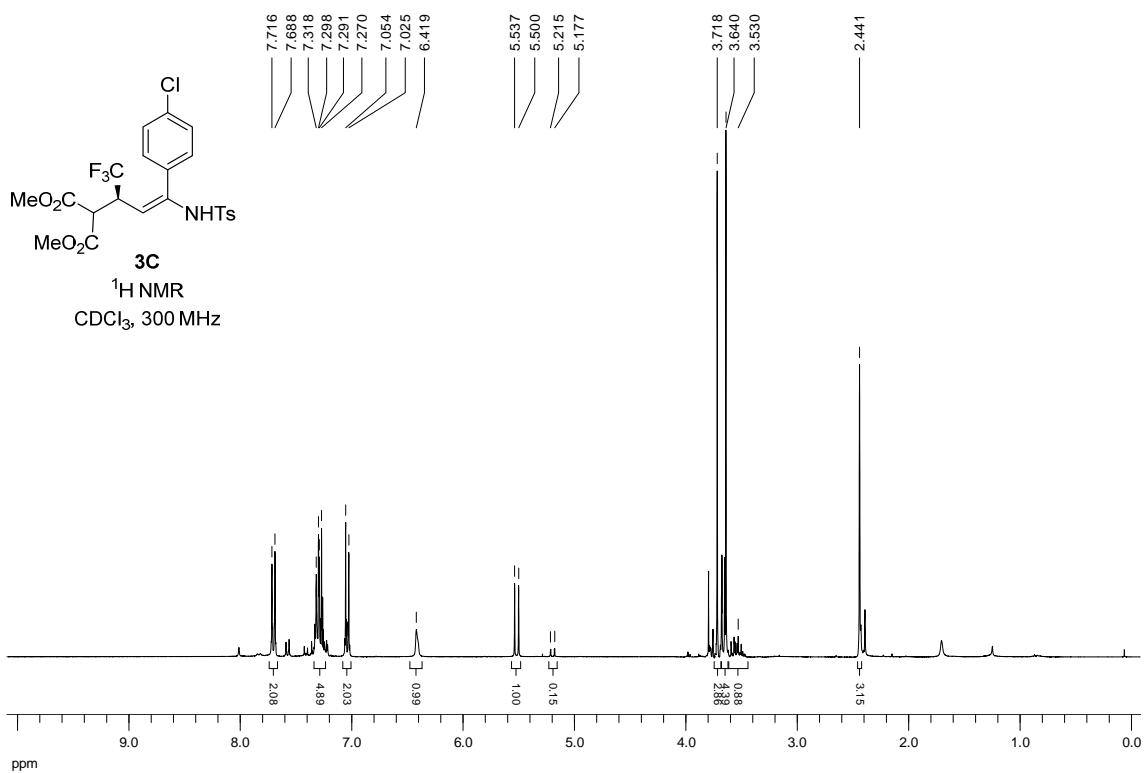
3: 240 nm, 4 nm Results

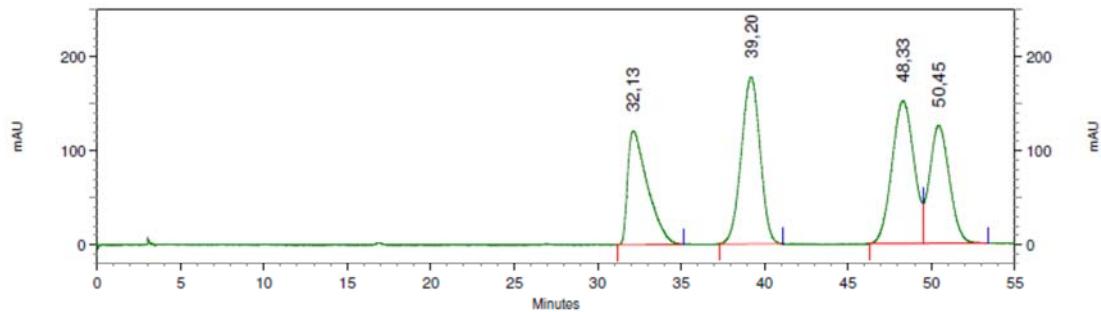
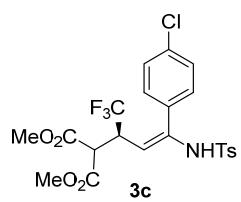
Retention Time	Area	Area Percent
11, 90	28064804	13,267
13, 38	79174978	37,428
14, 61	26352385	12,458
16, 00	77944696	36,847



7: 270 nm, 4 nm Results

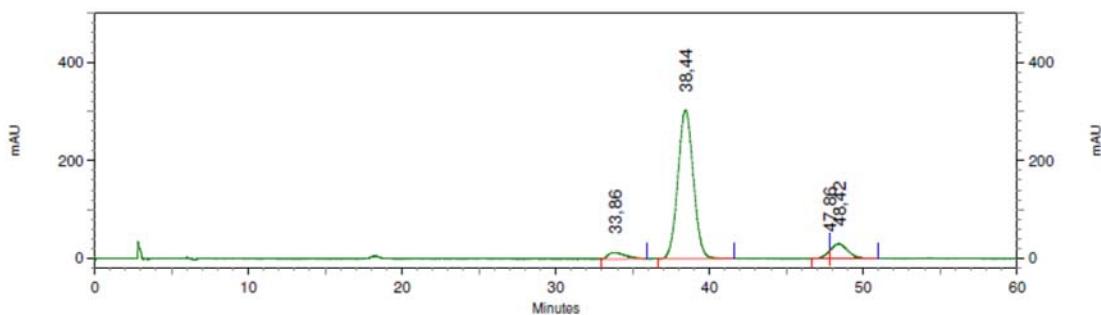
Retention Time	Area	Area Percent
11, 86	2621585	7,556
13, 36	25321055	72,982
14, 53	6091668	17,558
16, 09	660655	1,904





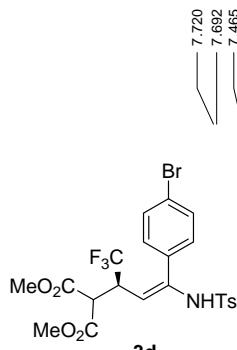
13: 250 nm, 4 nm
Results

Retention Time	Area	Area Percent
32, 13	40066660	20, 800
39, 20	55590059	28, 859
48, 33	55632362	28, 881
50, 45	41336988	21, 460

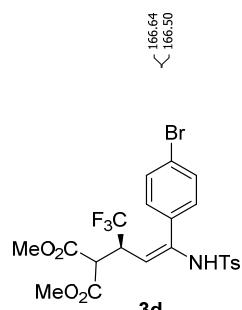
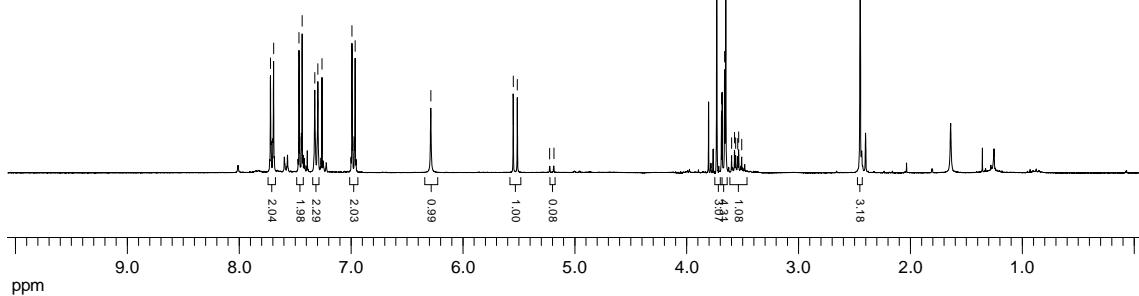


12: 220 nm, 4 nm
Results

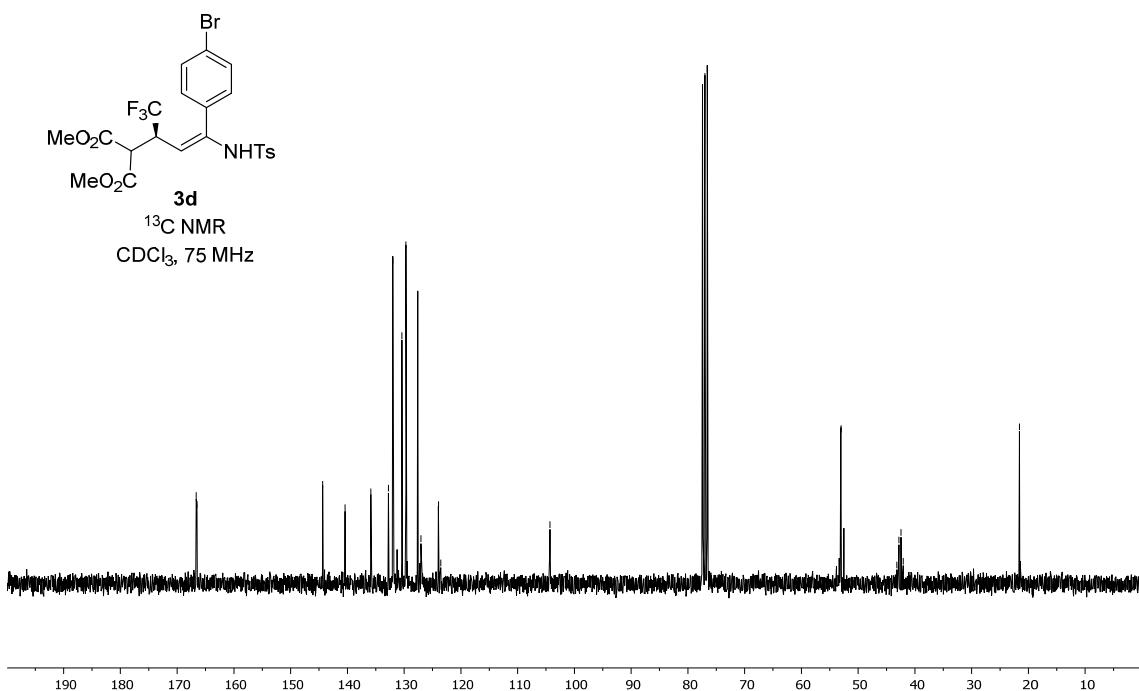
Retention Time	Area	Area Percent
33, 86	4277468	4, 322
38, 44	84913788	85, 800
47, 86	1448248	1, 463
48, 42	8327505	8, 414

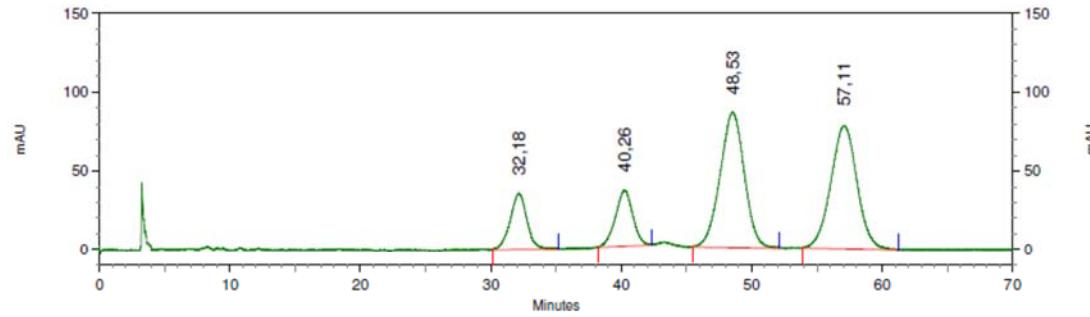
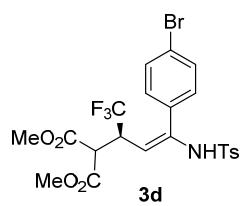


3d
 ^1H NMR
 CDCl_3 , 300 MHz



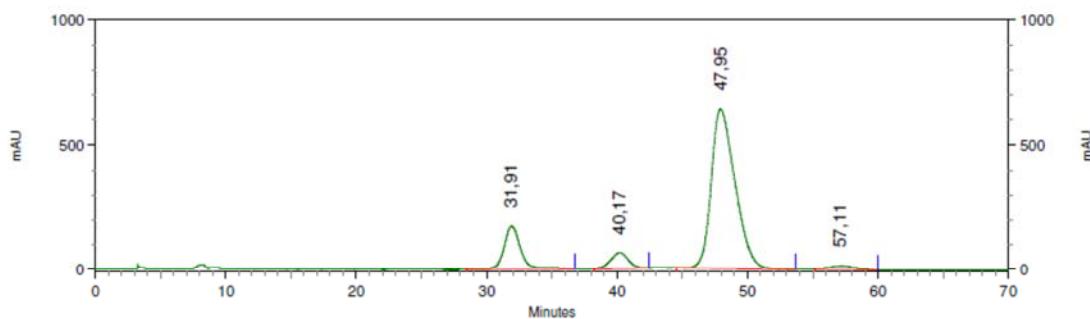
3d
 ^{13}C NMR
CDCl₃, 75 MHz





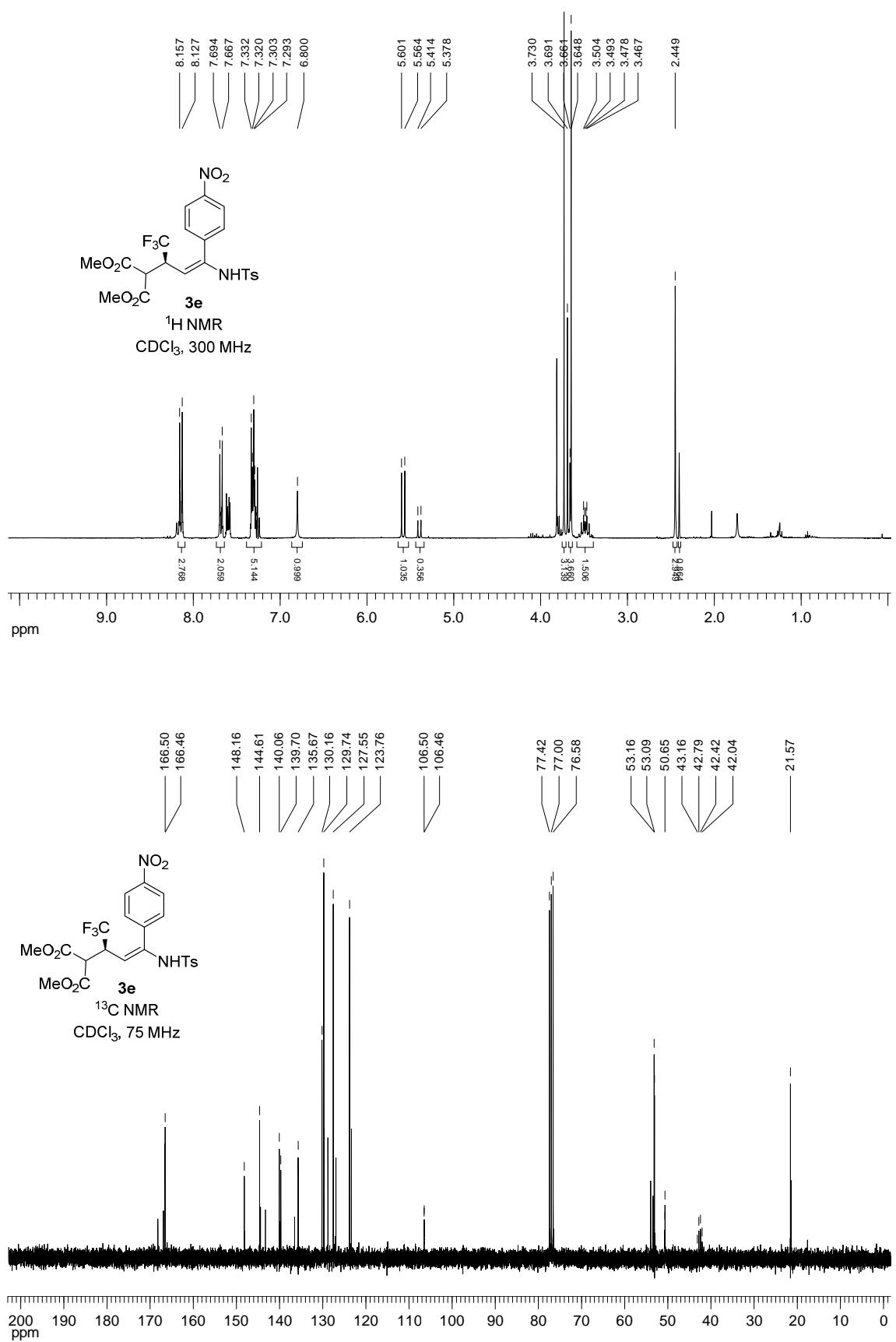
3: 220 nm, 4 nm Results

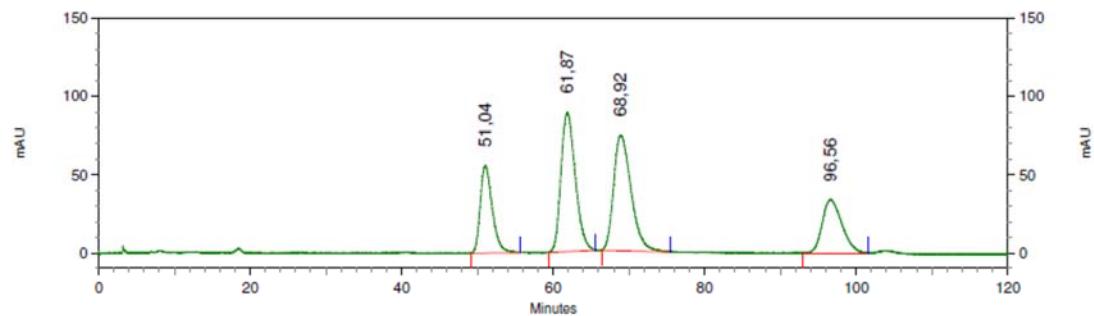
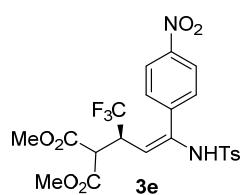
Retention Time	Area	Area Percent
32,18	12831961	11,192
40,26	13210649	11,523
48,53	44536148	38,845
57,11	44070703	38,440



2: 240 nm, 4 nm Results

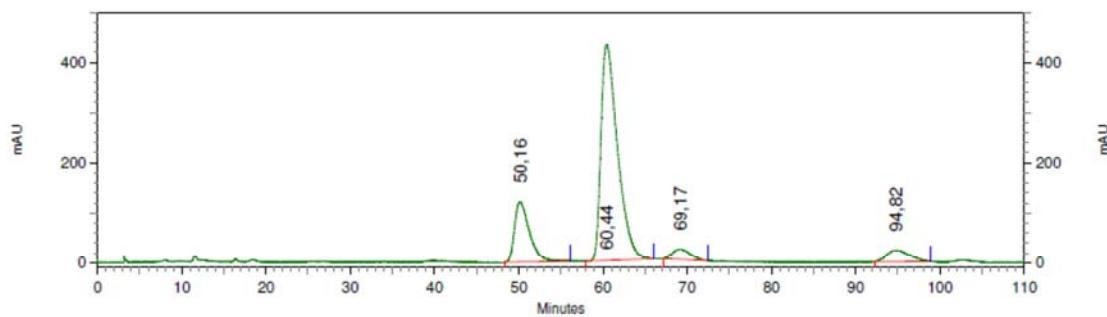
Retention Time	Area	Area Percent
31,91	61799367	14,738
40,17	24333799	5,803
47,95	325343204	77,586
57,11	7855823	1,873





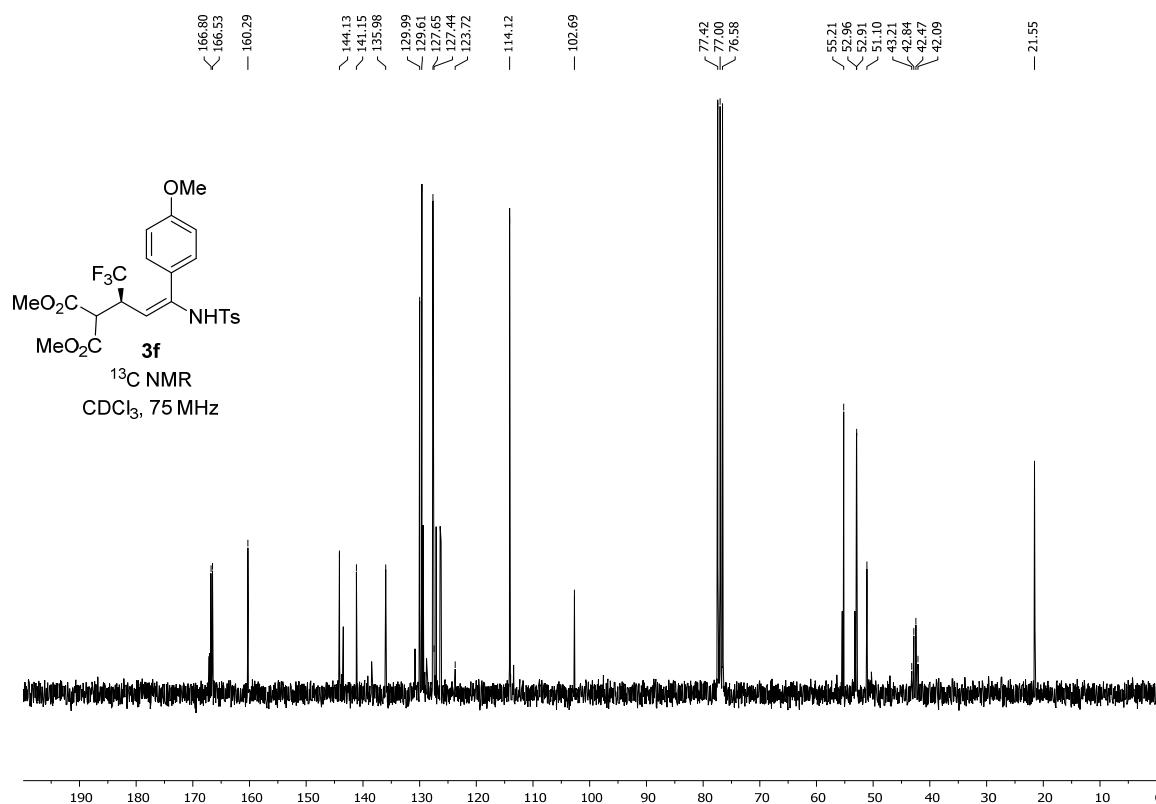
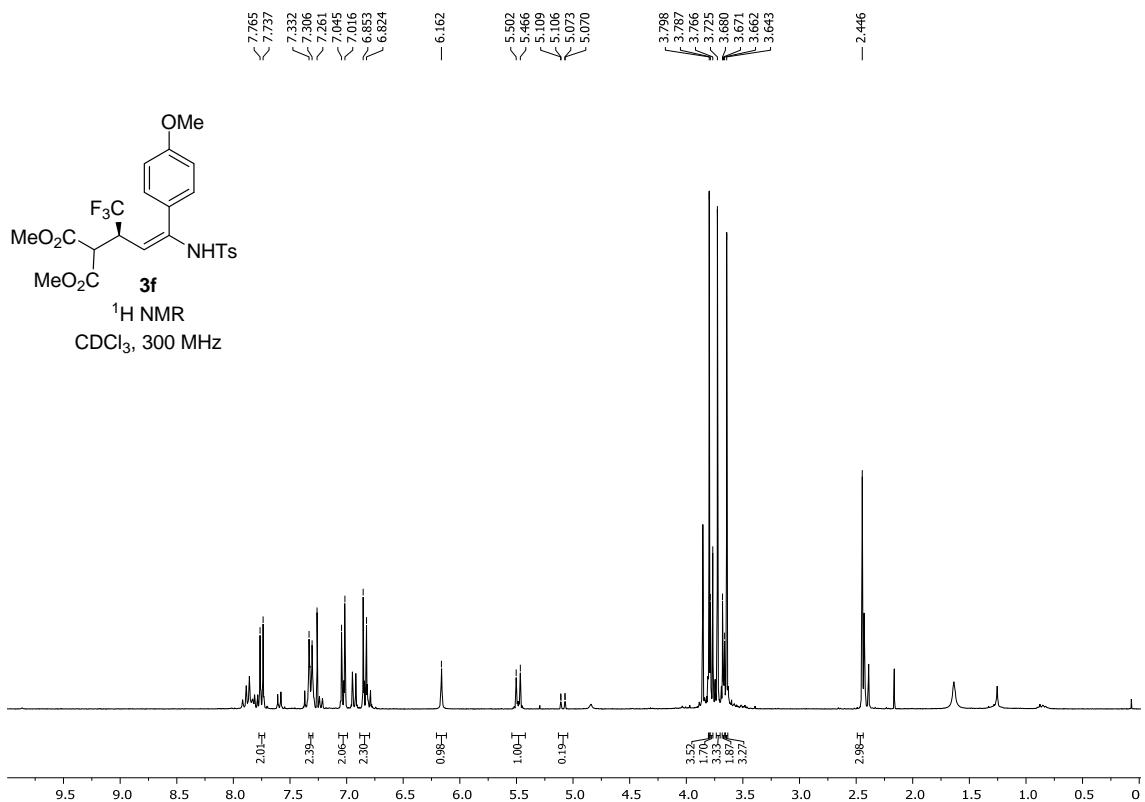
2: 250 nm, 4 nm Results
Retention Time

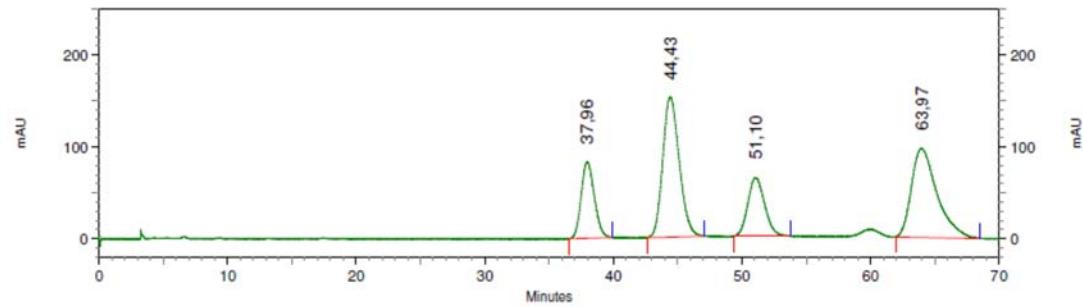
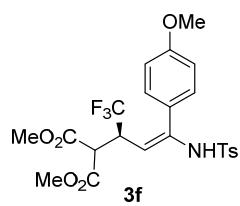
Retention Time	Area	Area Percent
51, 04	24653841	17, 171
61, 87	46674020	32, 507
68, 92	47423624	33, 029
96, 56	24830845	17, 294



2: 250 nm, 4 nm Results
Retention Time

Retention Time	Area	Area Percent
50, 16	60239223	18, 377
60, 44	237092235	72, 329
69, 17	12432840	3, 793
94, 82	18034406	5, 502

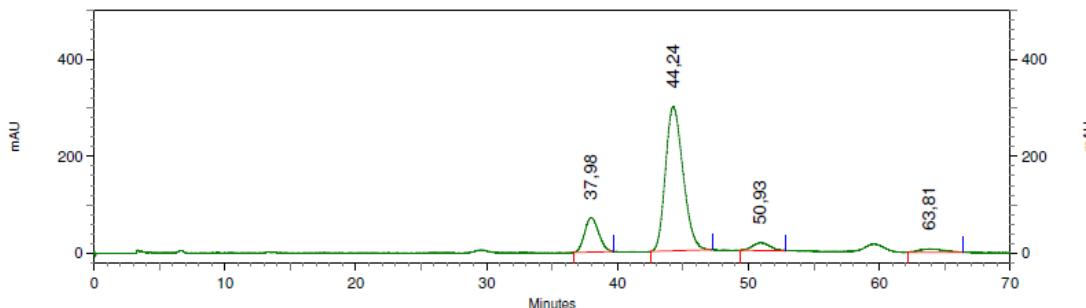




13: 250 nm, 4 nm

Results

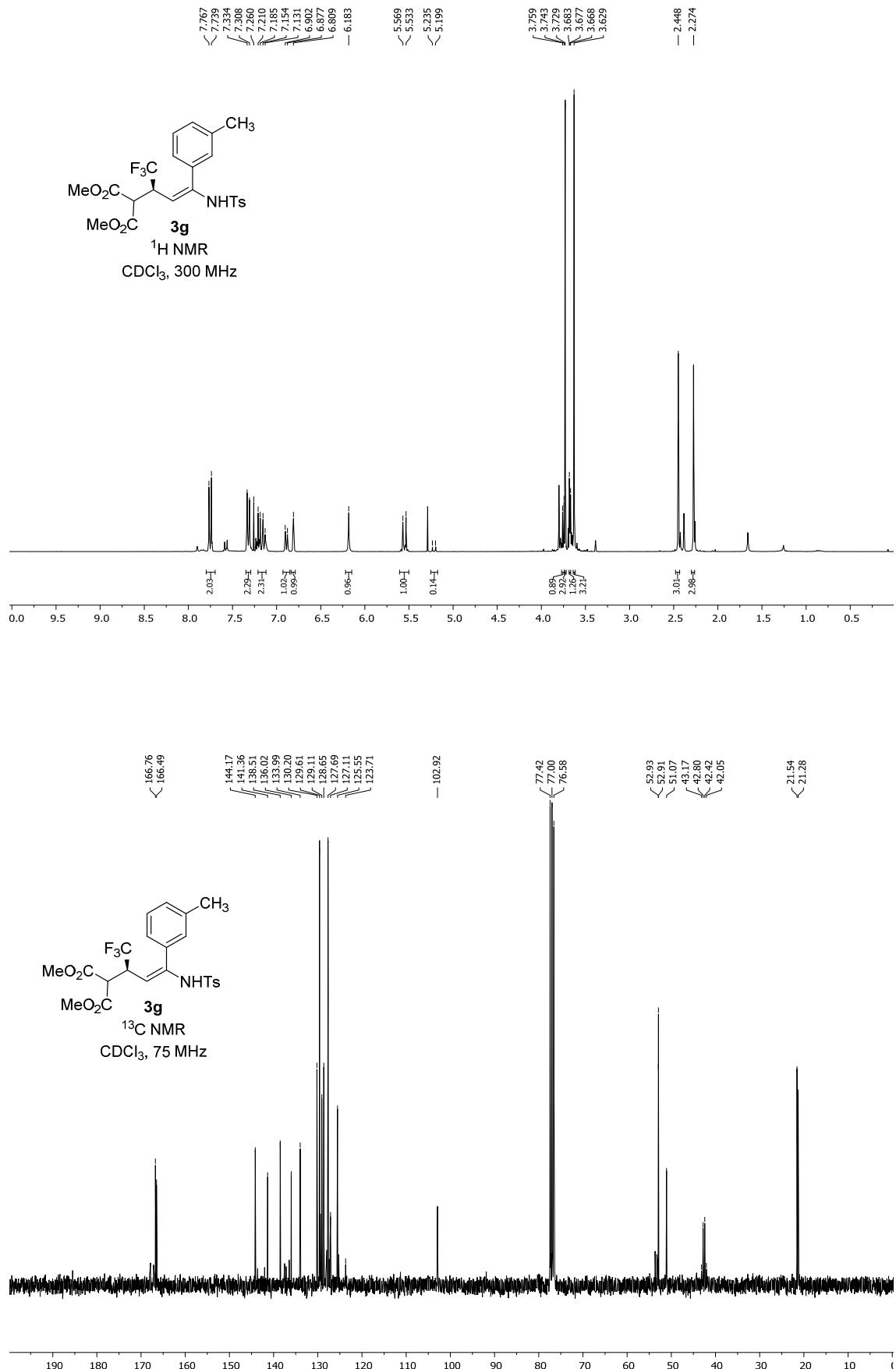
Retention Time	Area	Area Percent
37, 96	23906621	15, 390
44, 43	54493135	35, 081
51, 10	23247941	14, 966
63, 97	53687149	34, 562

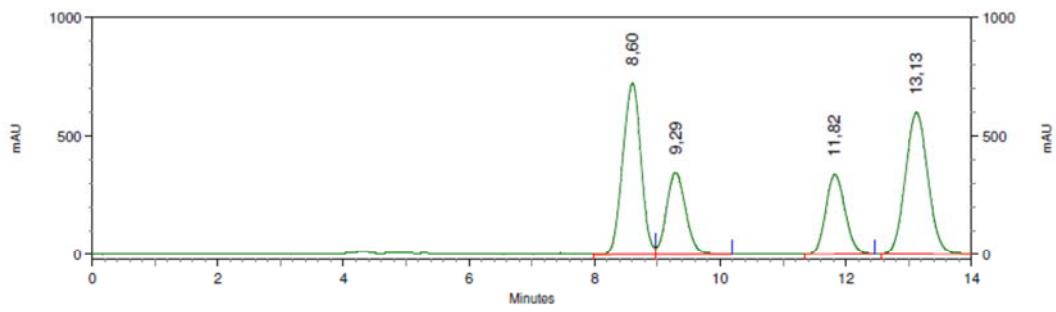
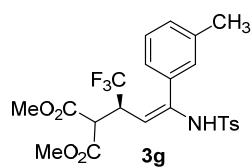


13: 250 nm, 4 nm

Results

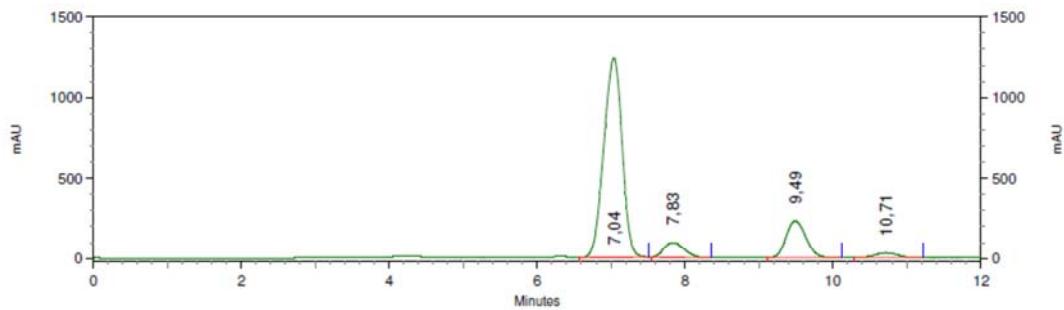
Retention Time	Area	Area Percent
37, 98	21218148	14, 931
44, 24	111411062	78, 400
50, 93	5991092	4, 216
63, 81	3486368	2, 453





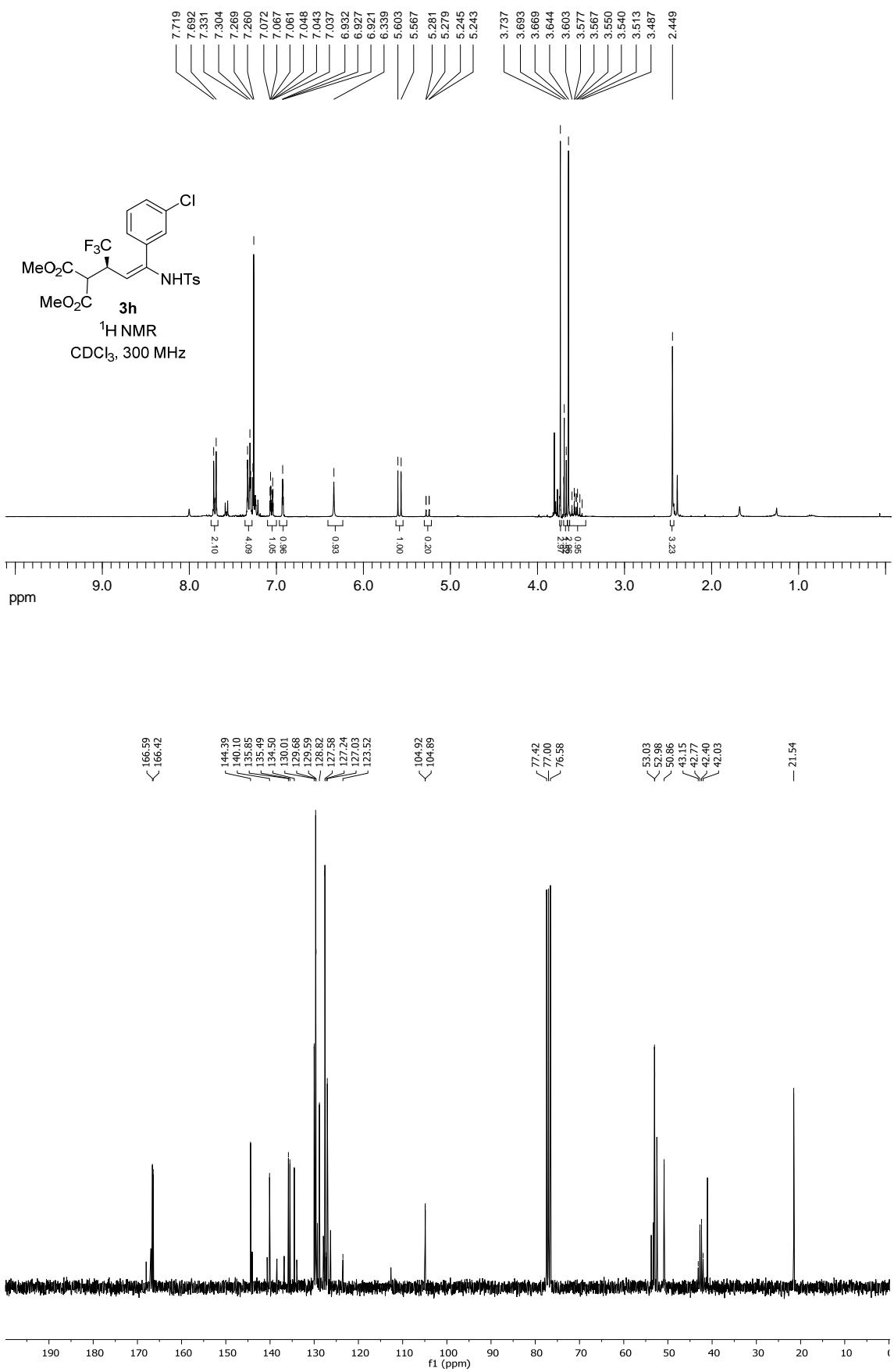
5: 250 nm, 4 nm Results

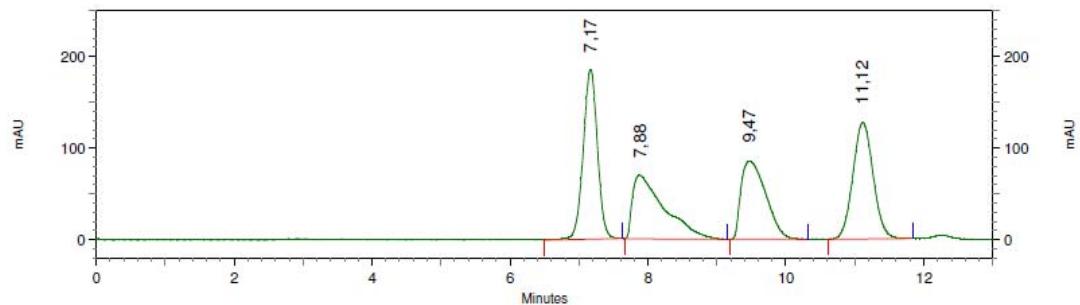
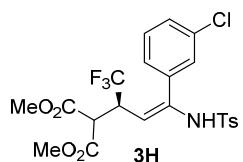
Retention Time	Area	Area Percent
8, 60	58401268	32, 954
9, 29	29577670	16, 690
11, 82	29173969	16, 462
13, 13	60066740	33, 894



7: 260 nm, 4 nm Results

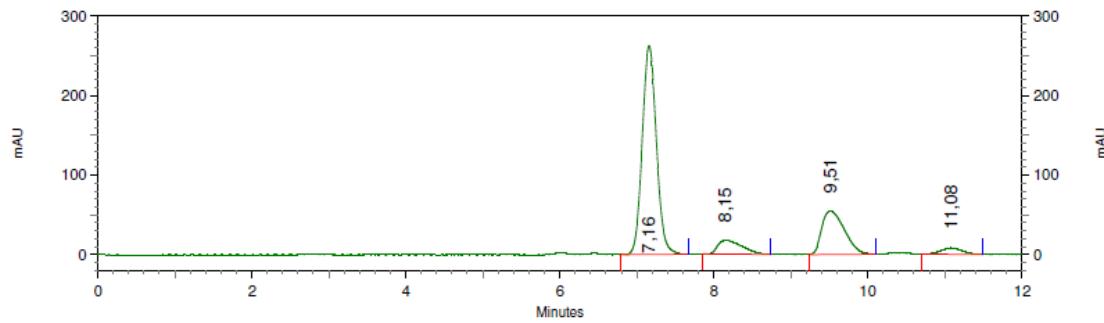
Retention Time	Area	Area Percent
7, 04	85744726	75, 785
7, 83	7274374	6, 429
9, 49	17477257	15, 447
10, 71	2646307	2, 339





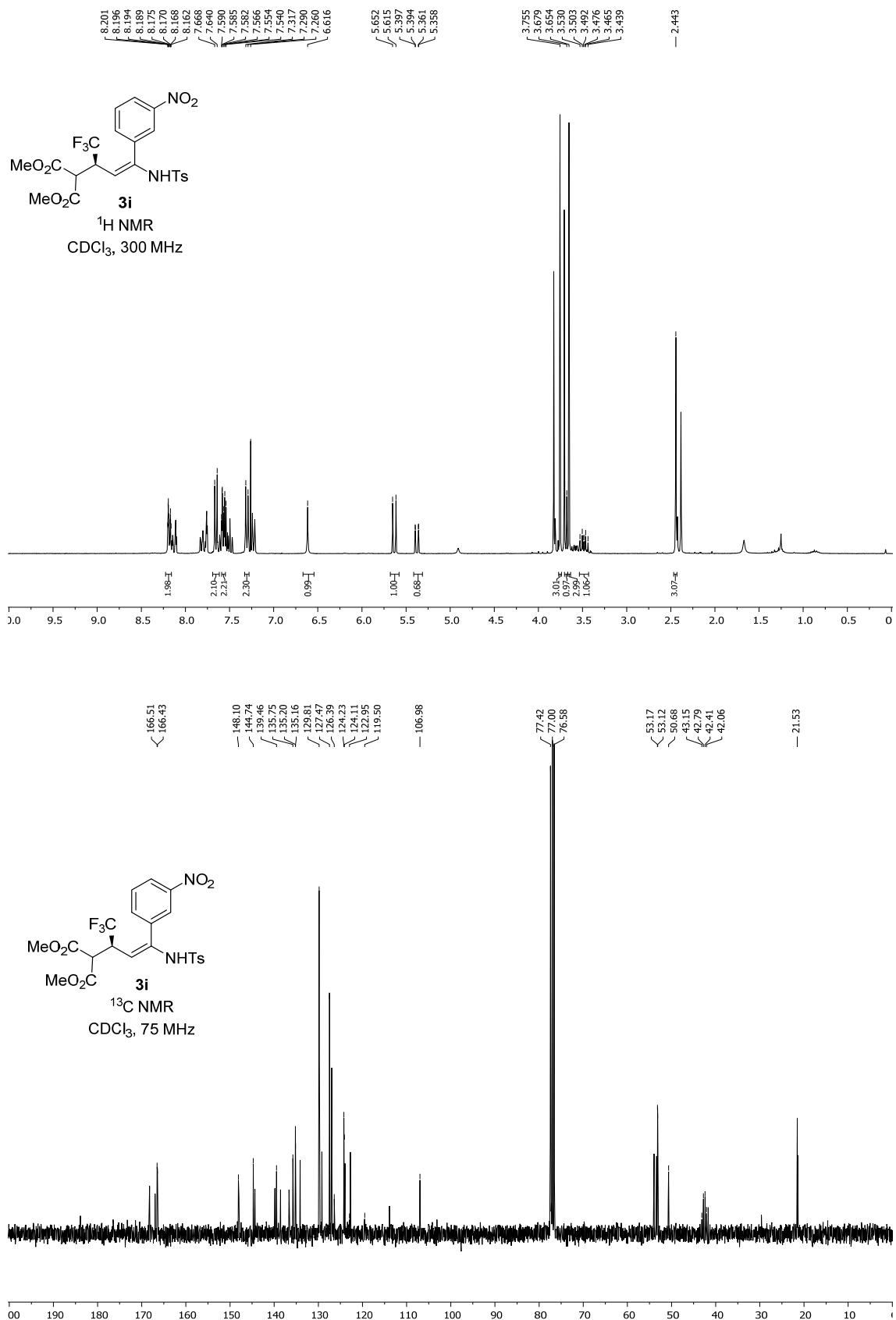
8: 270 nm, 4 nm Results
Retention Time

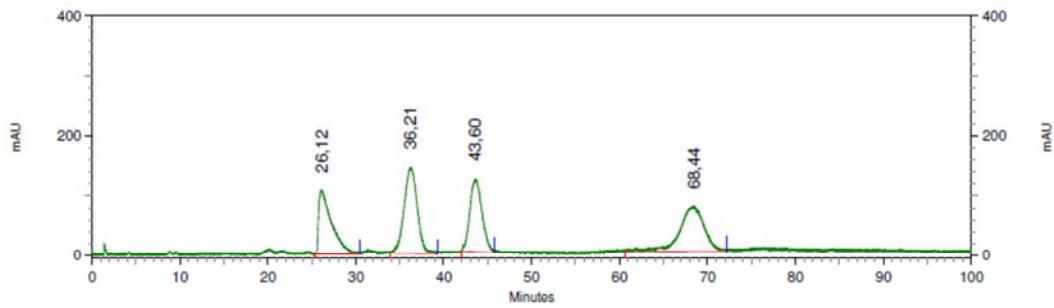
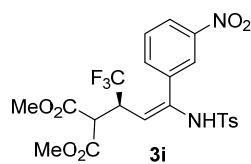
	Area	Area Percent
7, 17	10579681	27,533
7, 88	8899930	23,162
9, 47	8513973	22,157
11, 12	10431744	27,148



8: 270 nm, 4 nm Results
Retention Time

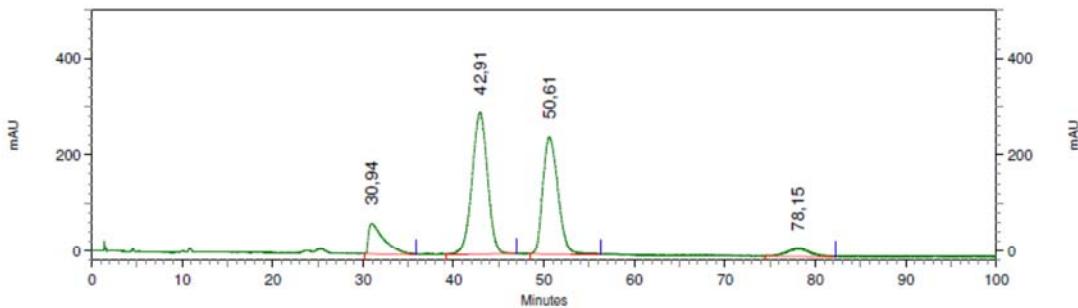
	Area	Area Percent
7, 16	13408638	66,894
8, 15	1539248	7,679
9, 51	4467929	22,290
11, 08	628647	3,136





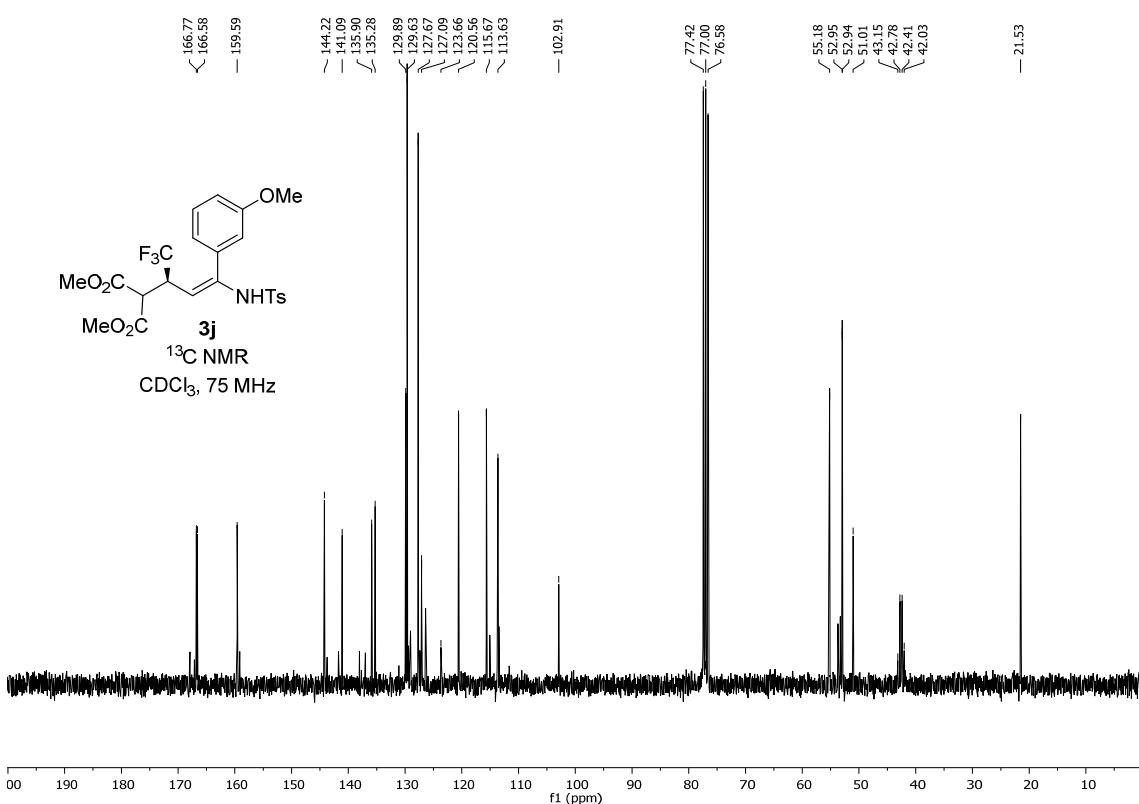
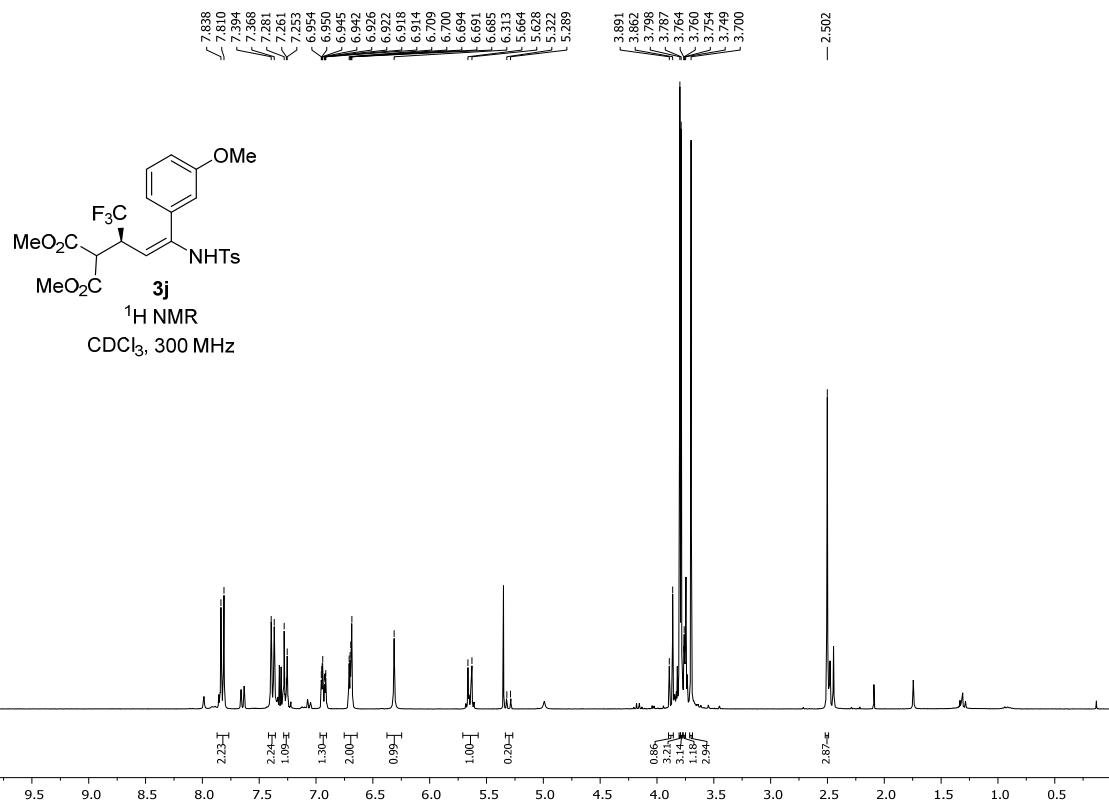
2: 240 nm, 4 nm Results
Retention Time

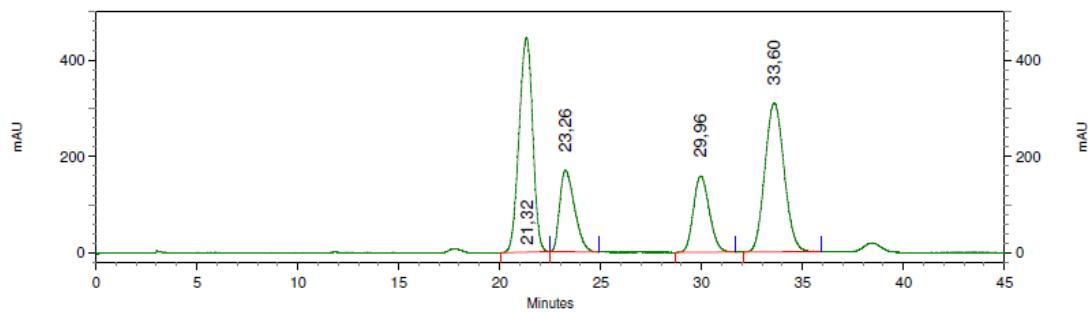
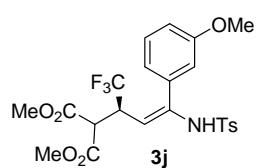
Retention Time	Area	Area Percent
26, 12	44278354	20, 864
36, 21	60686427	28, 596
43, 60	47134471	22, 210
68, 44	60124210	28, 331



2: 240 nm, 4 nm Results
Retention Time

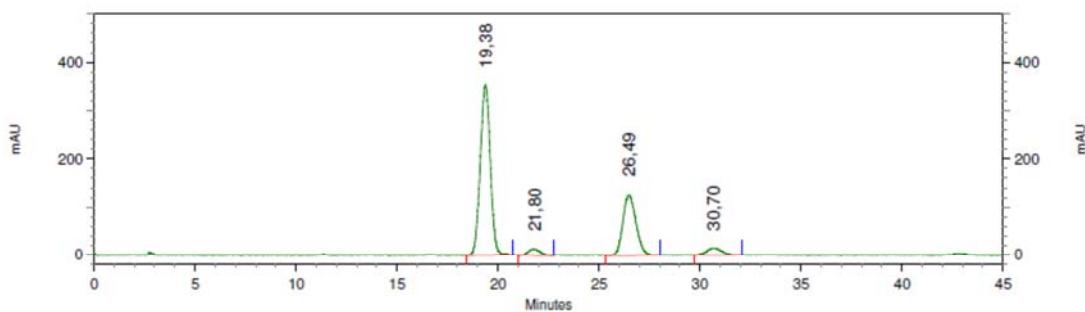
Retention Time	Area	Area Percent
30, 94	30715284	10, 243
42, 91	141508952	47, 190
50, 61	114674553	38, 242
78, 15	12969103	4, 325





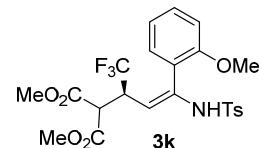
5: 250 nm, 4 nm Results
Retention Time

Retention Time	Area	Area Percent
21, 32	83433064	35, 597
23, 26	34464298	14, 704
29, 96	34604433	14, 764
33, 60	81878917	34, 934

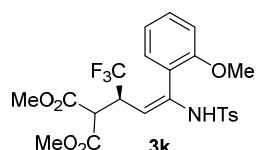
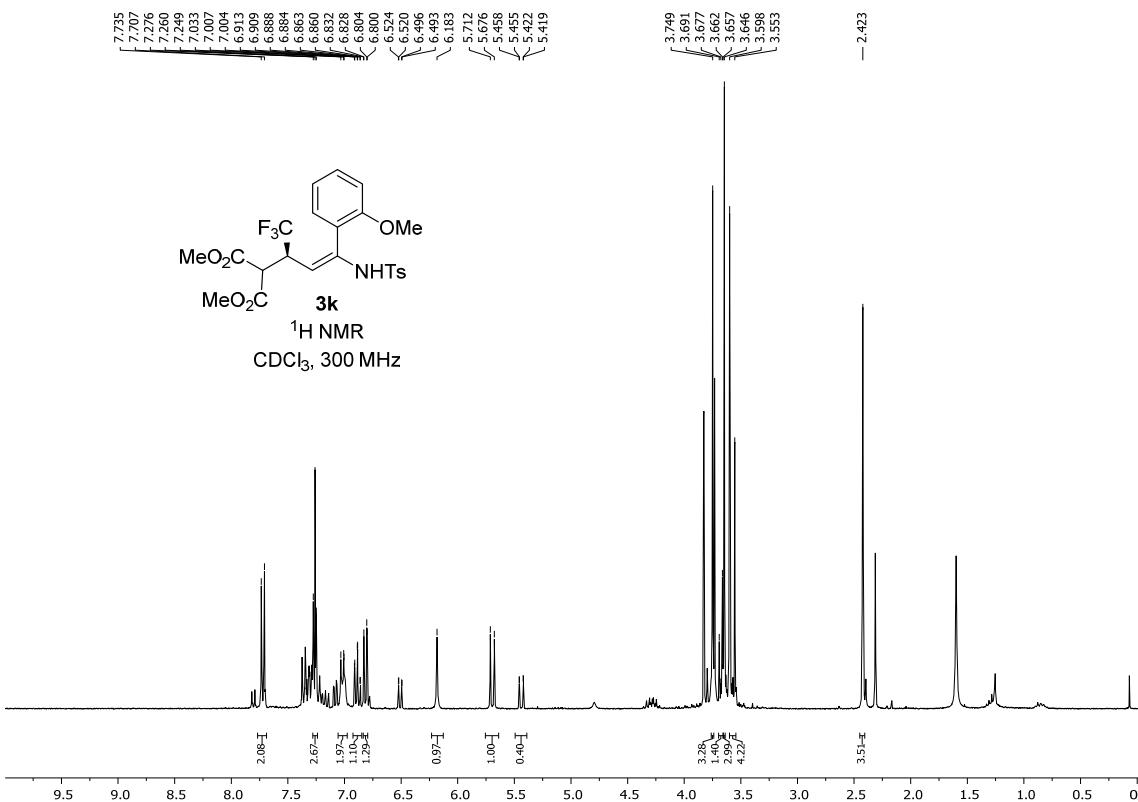


5: 250 nm, 4 nm Results
Retention Time

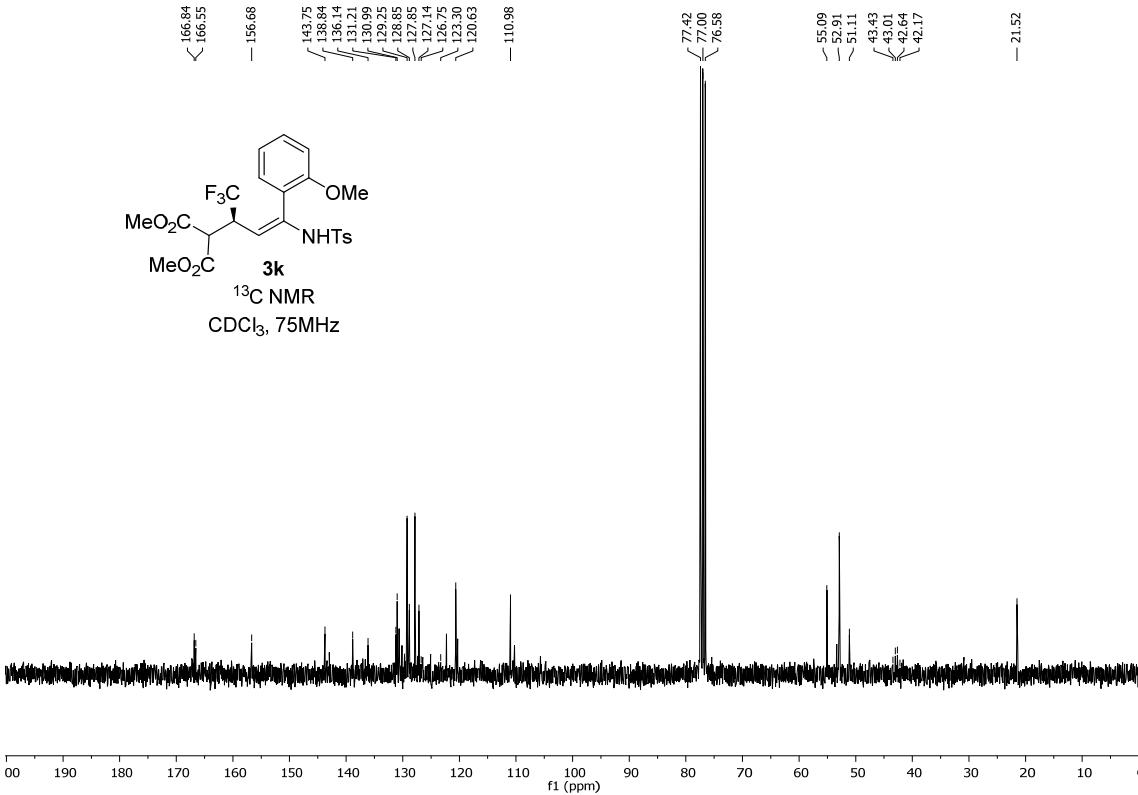
Retention Time	Area	Area Percent
19, 38	49004176	63, 381
21, 80	2251250	2, 912
26, 49	22751614	29, 426
30, 70	3310239	4, 281

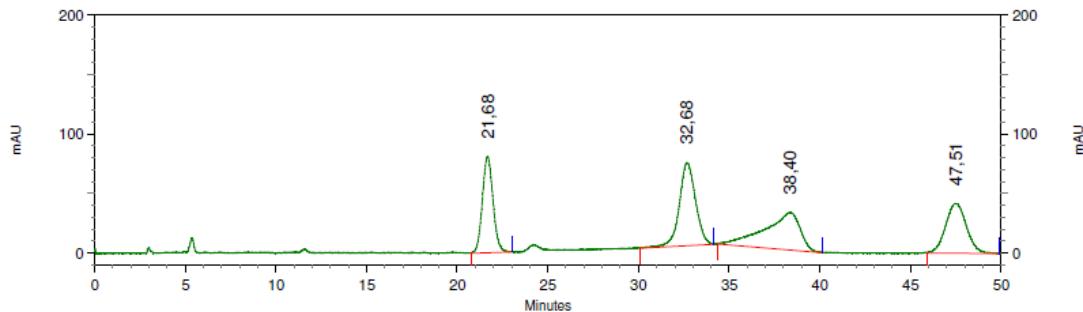
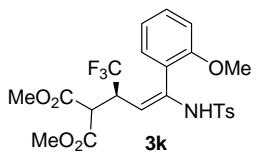


¹H NMR
CDCl₃, 300 MHz



¹³C NMR
CDCl₃, 75MHz



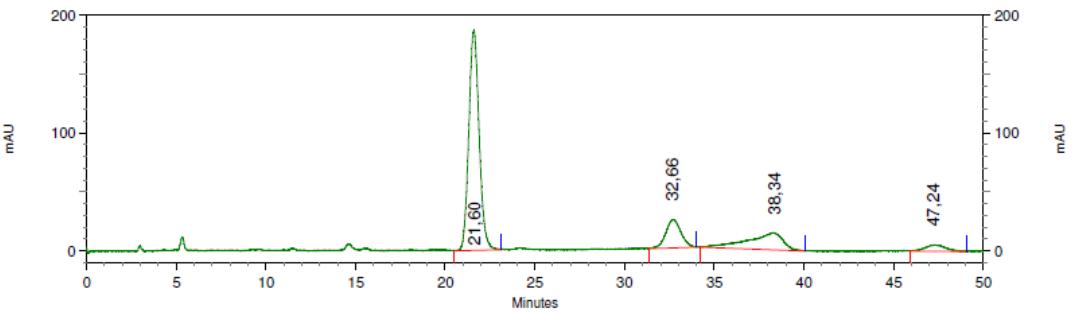


5: 250 nm, 4 nm Results
Retention Time

Area

Area Percent

21, 68	13274995	21, 691
32, 68	17429237	28, 479
38, 40	17140324	28, 007
47, 51	13356633	21, 824

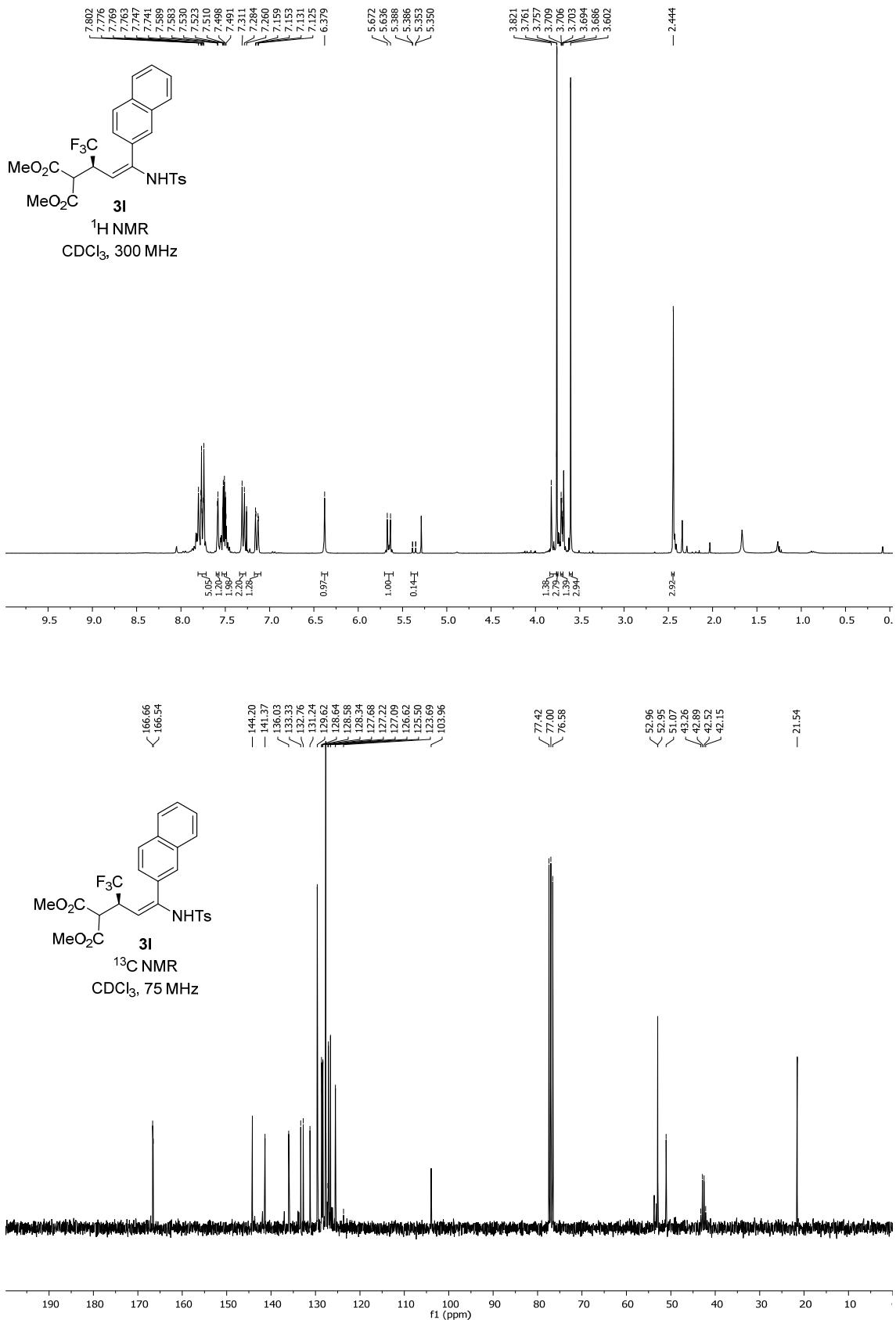


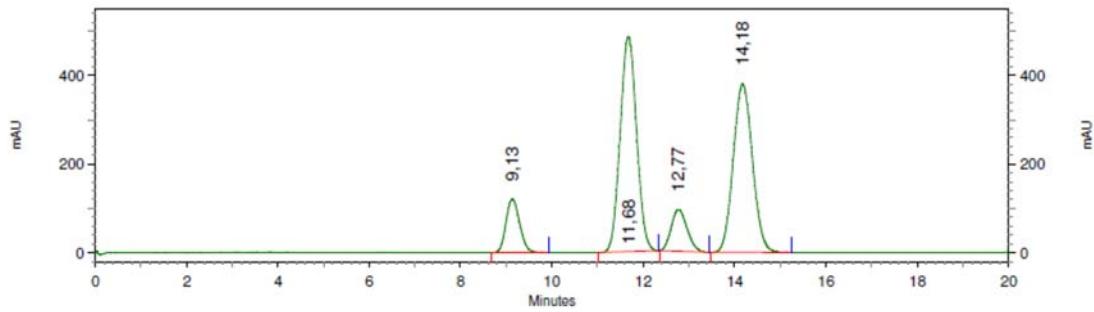
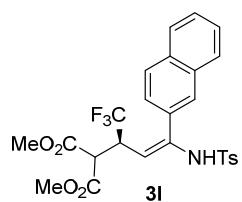
5: 250 nm, 4 nm Results
Retention Time

Area

Area Percent

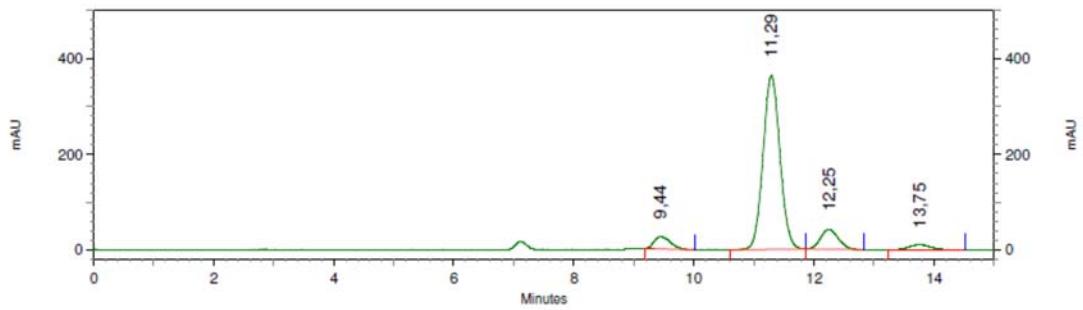
21, 60	29945092	65, 926
32, 66	5642658	12, 423
38, 34	8015853	17, 647
47, 24	1818970	4, 005





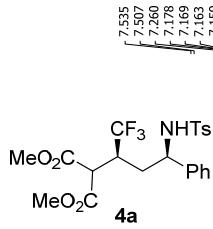
1: 280 nm, 4 nm Results

Retention Time	Area	Area Percent
9,13	10055071	8,990
11,68	48362788	43,240
12,77	9063996	8,104
14,18	44365277	39,666



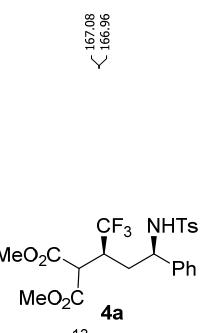
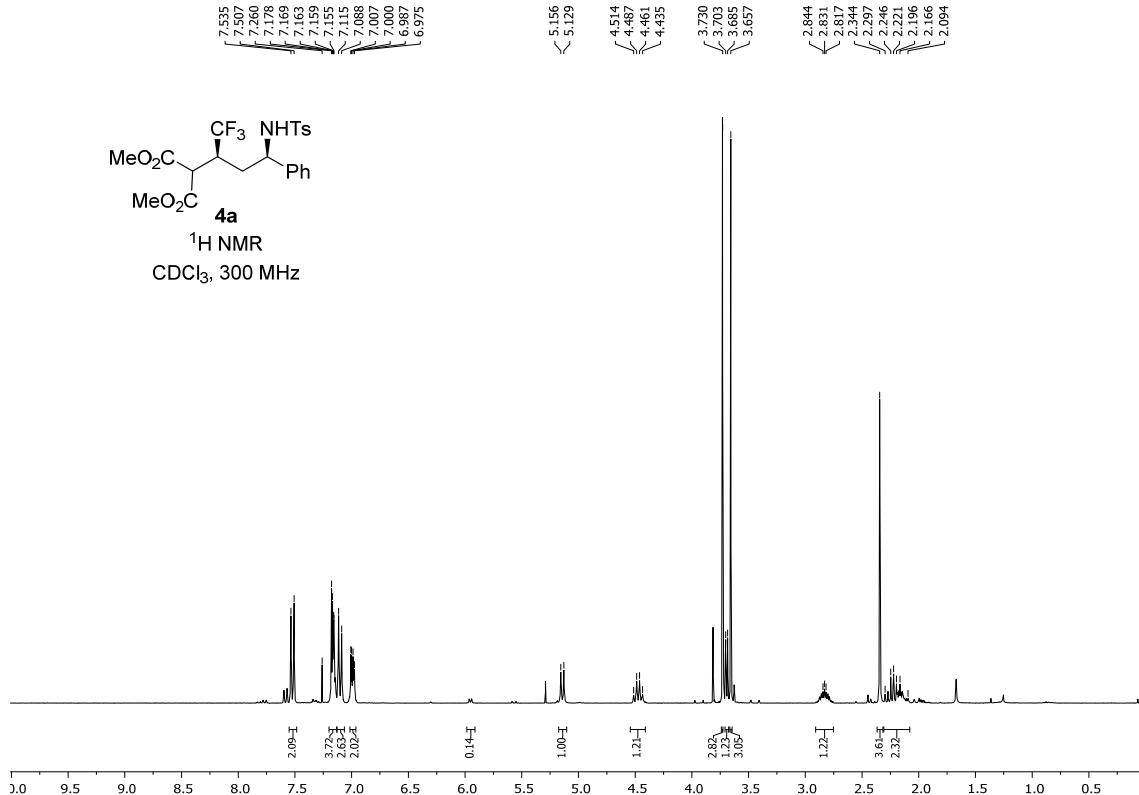
6: 280 nm, 4 nm Results

Retention Time	Area	Area Percent
9,44	1956733	5,608
11,29	28423861	81,464
12,25	3431501	9,835
13,75	1079387	3,094



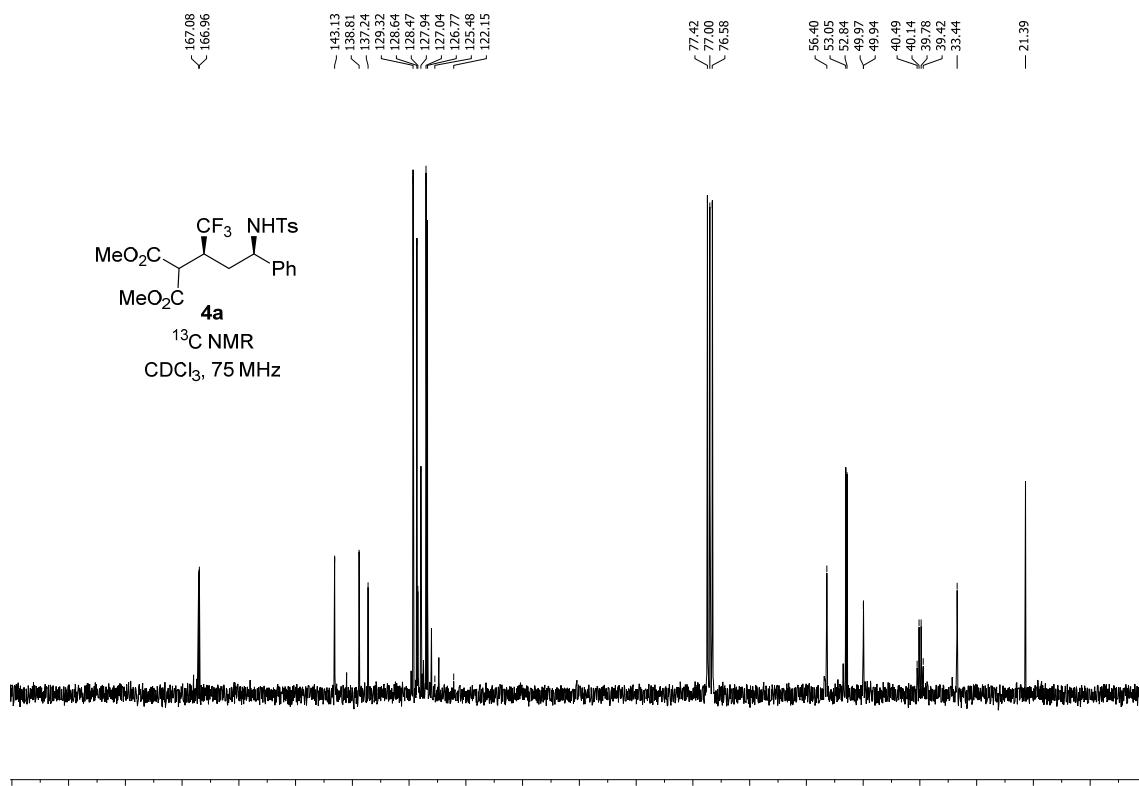
¹H NMR

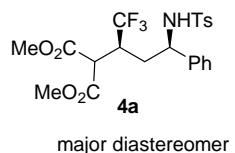
CDCl₃, 300 MHz



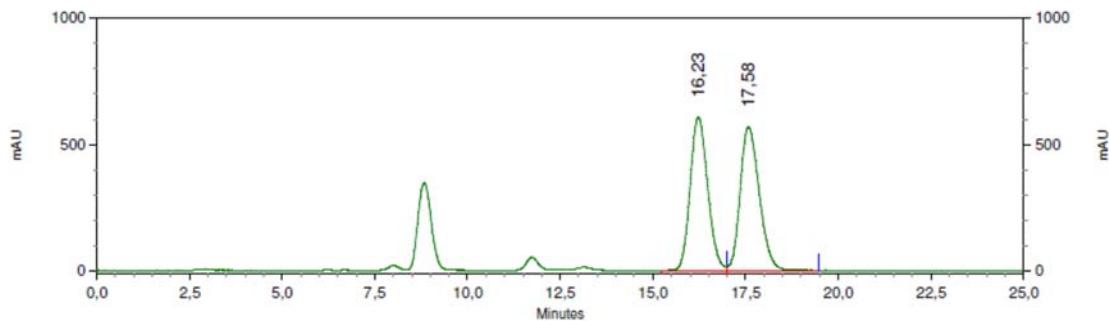
4a
¹³C NMR

¹³C NMR
CDCl₃, 75 MHz





Lux-Amylose 1

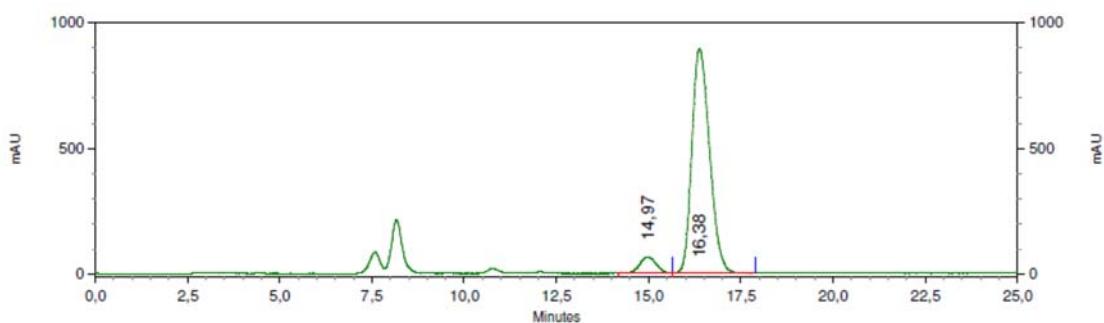


3: 240 nm, 4 nm Results
Retention Time

16, 23
17, 58

Area
80079138
81640326

Area Percent
49, 517
50, 483

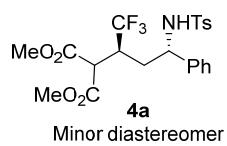


3: 240 nm, 4 nm Results
Retention Time

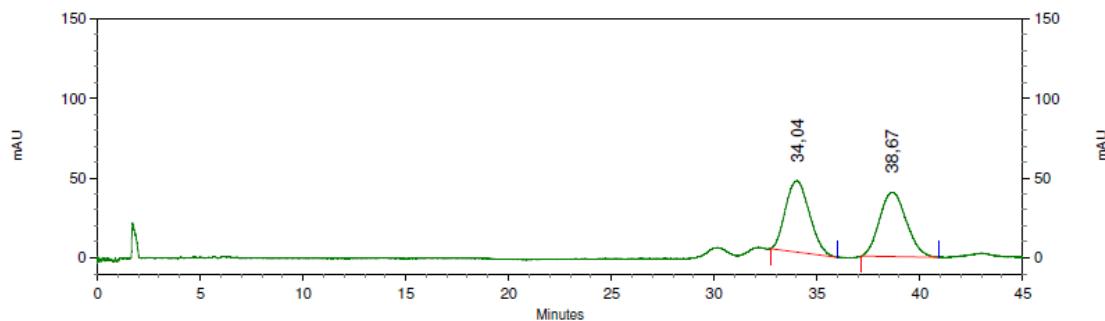
14, 97
16, 38

Area
8051805
119921166

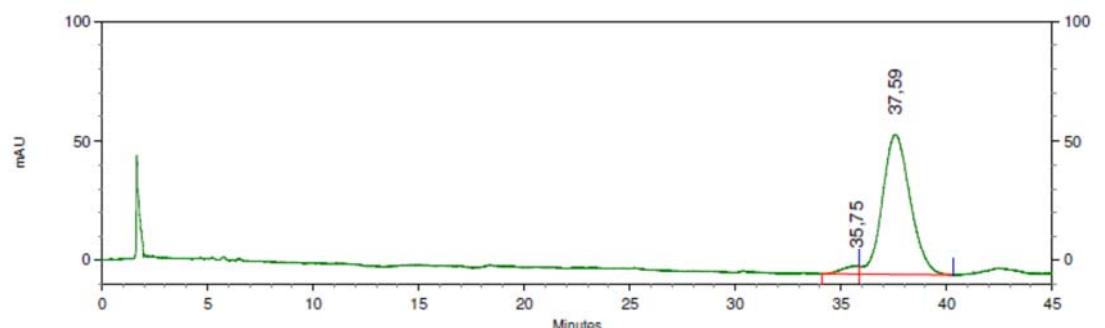
Area Percent
6, 292
93, 708



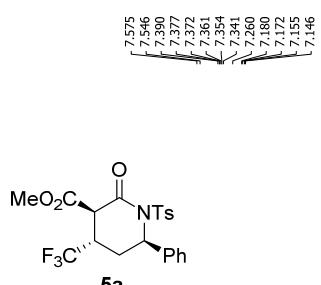
Chiraldpak IC



6: 220 nm, 4 nm Results		Area	Area Percent
Retention Time			
34, 04		14547165	49, 809
38, 67		14658732	50, 191



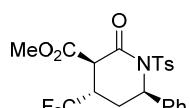
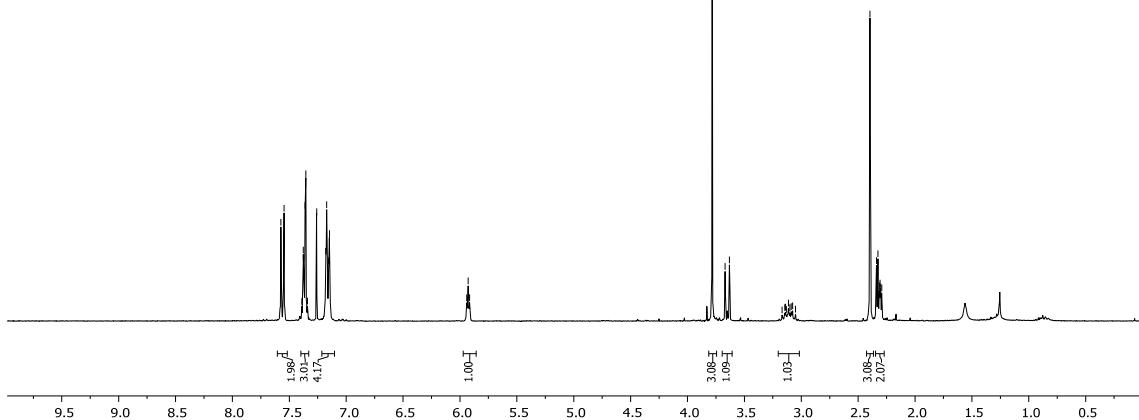
6: 220 nm, 4 nm Results		Area	Area Percent
Retention Time			
35, 75		696319	3, 104
37, 59		21738335	96, 896



5a
 ^1H NMR
 CDCl_3 , 300 MHz

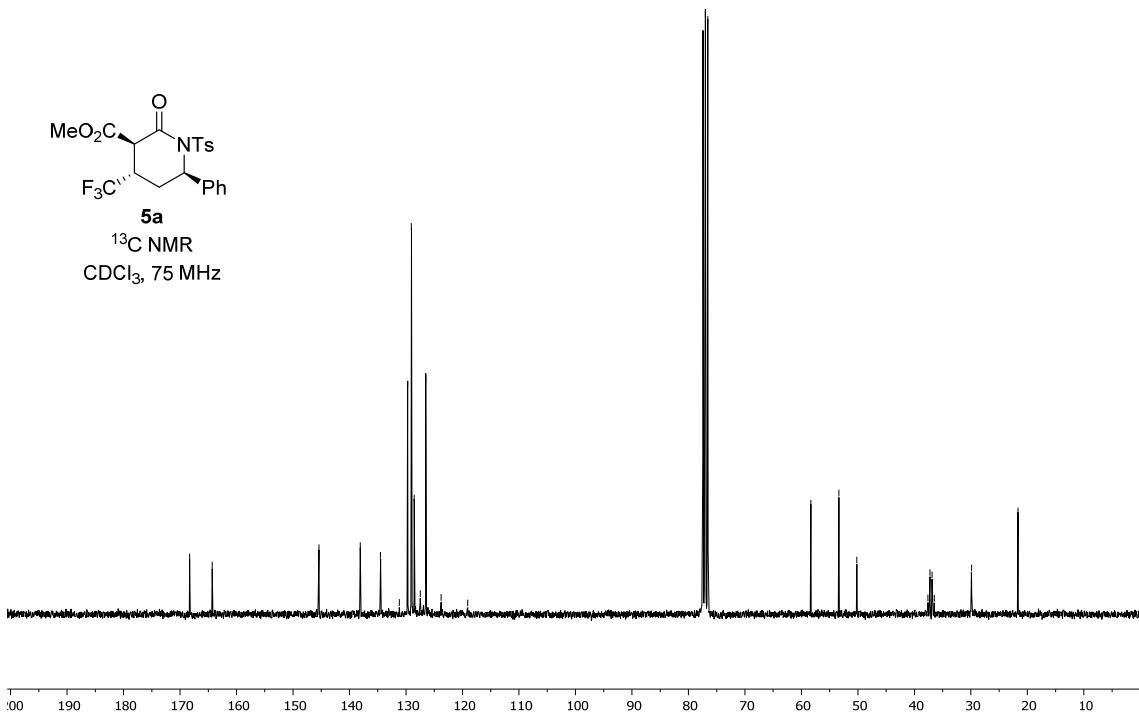
¹H NMR

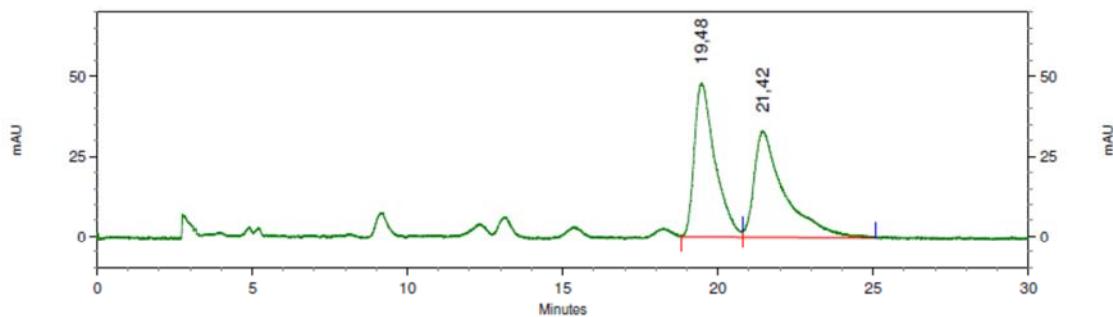
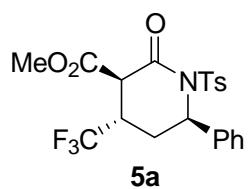
CDCl_3 , 300 MHz



5a

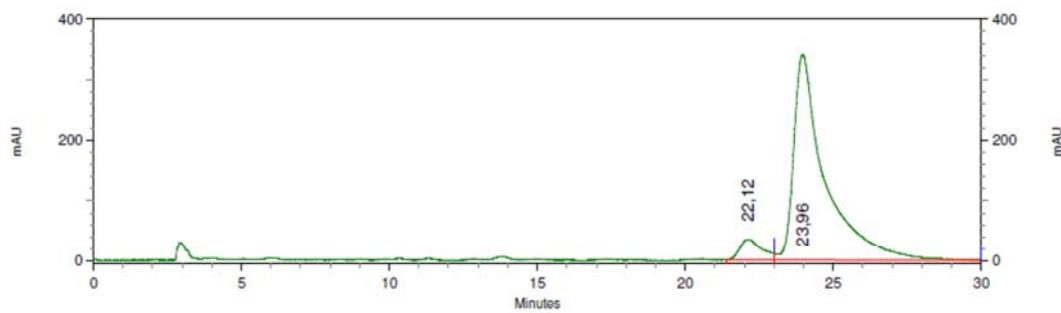
¹³C NMR
CDCl₃ 75 MHz





13: 250 nm, 4 nm
Results

Retention Time	Area	Area Percent
19, 48	8876376	49, 765
21, 42	8960049	50, 235



12: 220 nm, 4 nm
Results

Retention Time	Area	Area Percent
22,12	7086461	6,584
23,96	100551429	93,416

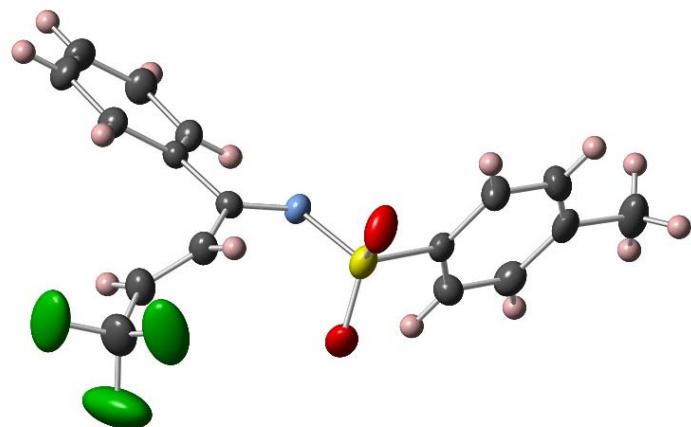


Figure 2S. Ortep plot for the X-ray structure of compound **2a**. The thermal ellipsoids are drawn at the 50% probability level.

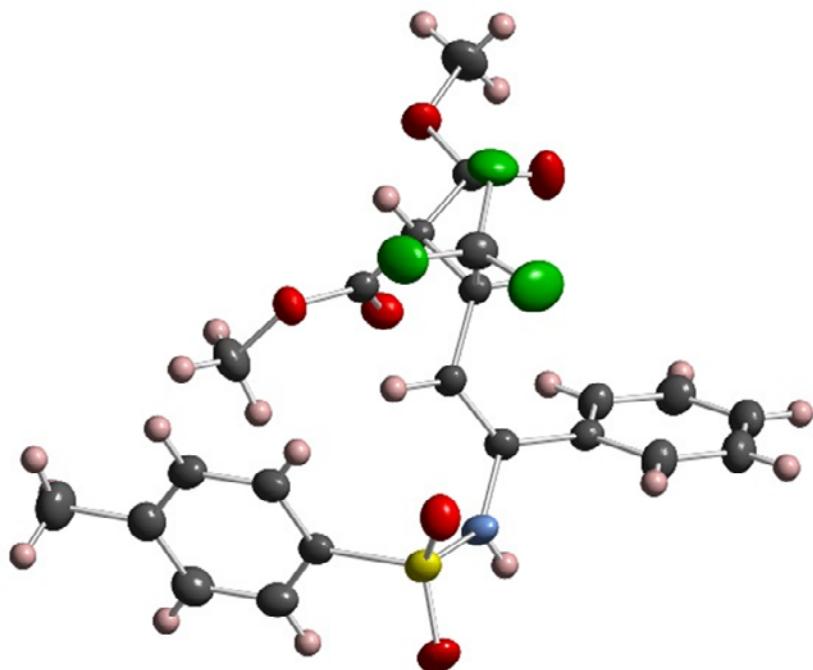
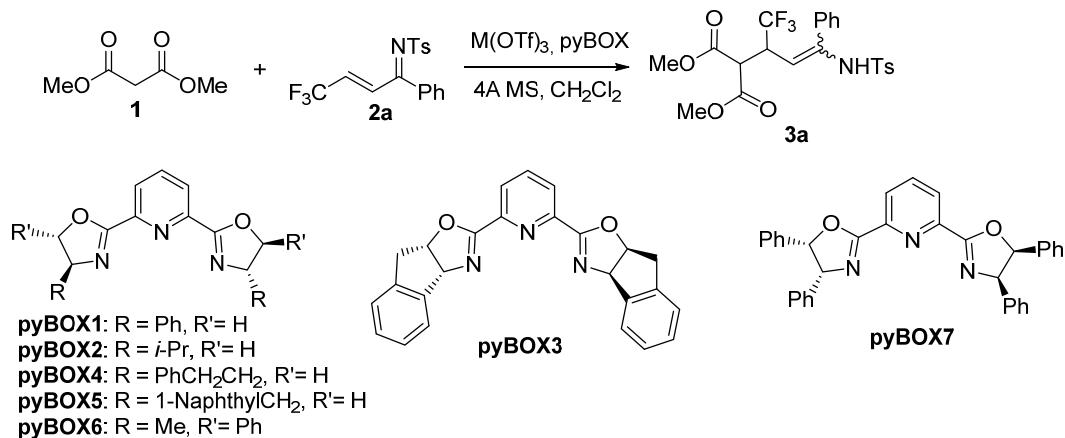


Figure 3S. Ortep plot for the X-ray structure of compound **3a**. The thermal ellipsoids are drawn at the 50% probability level.

Table S-1. Enantioselective conjugate addition of dimethyl malonate **2** to imine **2a** catalyzed by trivalent metal complexes.^a



Entry	La(OTf) ₃	pyBOX	t (h)	Yield (%) ^[b]	dr (E:Z)	ee (%) (E/Z) ^c
1	La(OTf) ₃	pyBOX1	16h	>99	72:28	75/34
2	La(OTf) ₃	pyBOX2	43h	>99	78:23	-15/-2
3	La(OTf) ₃	pyBOX3	48	>99	77:23	45/-3
4	La(OTf) ₃	pyBOX4	40h	>99	82:18	-18/-9
5	La(OTf) ₃	pyBOX5	40h	>99	82:18	-28/-11
6	La(OTf) ₃	pyBOX6	37h	79	89:11	-16/5
7	La(OTf) ₃	pyBOX7	44h	86	73:27	-76/-39
8	Sc(OTf) ₃	pyBOX1	96	-- ^d	--	--
9	Yb(OTf) ₃	pyBOX1	96	19	43:57	69/42
10	In(OTf) ₃	pyBOX1	96	-- ^d	--	--

^a Reaction conditions: **2** (0.3 mmol), **11** (0.12 mmol), ligand (0.012 mmol, M(OTf)₃ (0.012 mmol), 4Å MS (110 mg), CH₂Cl₂ (1.1 mL). ^b Yield of isolated product. ^c Determined by HPLC with chiral stationary phases. ^d Little advance of the reaction was observed after the indicated time.