# Supporting information

# Efficient Chiral <sup>1</sup>H NMR Analysis of Indoloquinazoline Alkaloids

# Phaitanthrins A, Cephalanthrin-A and Their Analogues with A

# **Chiral Phosphoric Acid**

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#### 1. General information.

All commercial reagents were used as received without further purification unless otherwise stated. <sup>1</sup>H NMR (500 MHz) spectra were recorded in acetone-D<sub>6</sub> or CDCl<sub>3</sub> solutions using a 500 MHz spectrometer at 25 °C. Chemical shifts were reported in parts per million (ppm,  $\delta$ ) relative to residual TMS. **2q**, **2r**, **2s** were purchased from J&K Chemical Ltd. (Shanghai); **2a-2m** were synthesized according the procedures we developed previously (reference 7 in the main text); the chiral phosphoric acids were synthesized by known procedures (reference 11 in the main text).

# 2. Evaluating the chiral recognition abilities of chiral sensors 1a-1f (0.01 mmol) with 2a (0.01 mmol) in acetone-D<sub>6</sub> at 25 °C.



# 3. Copies of <sup>1</sup>H NMR spectra of chiral sensors 1a-1f (0.01 mmol, 1.0 equiv.) and racemic Cephalanthrin-A (0.01 mmol, 1.0 equiv.).

# 3.1. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of racemic 2a.



# 3.2. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1a and guest 2a.



3.3. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1b and guest 2a.



# 3.4. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1c and guest 2a.



# 3.6. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of (R)-BINOL 1e and guest 2a.



3.7. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of quinidine 1f and guest 2a.



# 3.8. <sup>1</sup>H NMR (500 MHz, DMSO) of CPA 1d and guest 2a.



3.9. <sup>1</sup>H NMR (500 MHz, CD<sub>3</sub>OD) of CPA 1d and guest 2a.



4. Copies of <sup>1</sup>H NMR spectra of 1d (0.01 mmol, 1.0 equiv.) and racemic 2 (0.01 mmol, 1.0 equiv.).

4.1. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and racemic guest 2b.





4.3. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and racemic guest 2d.



4.5. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and racemic guest 2f.



#### 4.7. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and racemic guest 2h.



4.9. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and racemic guest 2j.



4.11. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and racemic guest 2l.

4.12. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d (0.015 mmol, 1.5 equiv.) and racemic guest 2m (0.01 mmol, 1.0 equiv.).



# 4.13. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of racemic 2n.



4.14. <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) of racemic 2n.







# 4.17. <sup>13</sup>C NMR (126 MHz, CDCl<sub>3</sub>) of racemic 20.



4.18. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of CPA 1d (0.01 mmol, 1.0 equiv.) and racemic guest 20 (0.01 mmol, 1.0 equiv.).



# 4.19. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of racemic 2p.





4.21. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of CPA 1d (0.01 mmol, 1.0 equiv.) and racemic guest 2p(0.01 mmol, 1.0 equiv.).





4.23. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of CPA 1d (0.01 mmol, 1.0 equiv.) and racemic guest 2q (0.02 mmol, 2.0 equiv.).





4.25. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of CPA 1d (0.01 mmol, 1.0 equiv.) and racemic guest 2r (0.015 mmol, 1.5 equiv.).







4.27. <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>) of CPA 1d (0.01 mmol, 1.0 equiv.) and racemic guest 2s (0.02 mmol, 2.0 equiv.).



# 5. Copies of <sup>1</sup>H NMR spectra/HPLC data of 1d (0.01 mmol, 1.0 equiv.) and 2 (0.01 mmol, 1.0 equiv.) with different optical purities.

For chiral HPLC analysis, a wide range of nonracemic **2a** were converted into corresponding esters. The procedure is as follows:



SOCl<sub>2</sub> (1.2 mL) was added slowly to a solution of **2a** in anhydrous CH<sub>3</sub>OH (4.0 mL) at 0 °C, the resulting mixture was stirred at the same temperature for 4 hours followed by the addition of saturated aqueous NaHCO<sub>3</sub> solution (60 mL), and the reaction mixture was extracted with CH<sub>2</sub>Cl<sub>2</sub> (4×10 mL). The combined organic phase was washed with brine (80 mL), and then dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, filtered and the solvent was removed in vacuo. The residue was purified through flash column chromatography on a silica gel (eluent: dichloromethane : ethyl acetate = 15/1-10/1) to yield **2a'**.

5.1. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2a (Sample 1, ee 79.2%).



# 5.2. HPLC of guest 2a' (Sample 1, ee 81.2%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.592	3137113	70627	10.891	14.304	9.4065
2	19.414	30213247	247556	18.539	27.008	90.5935

5.3. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2a (Sample 2, ee 65.1%).



5.4. HPLC of guest 2a' (Sample 2, ee 66.1%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.549	4761352	107576	10.880	14.581	16.9586
2	19.612	23314923	195381	18.688	27.509	83.0414

5.5. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2a (Sample 3, ee 51.1%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.528	7030258	157159	10.848	15.040	23.6499

192142

17.5

20.0

22.5

18.784

25.0

27.5

27.211

30.0

32.5

76.3501

15.0

0.0

2

2.5

5.0

19.728

7.5

10.0

12.5

22696076

5.7. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2a (Sample 4, ee 38.6%).



5.8. HPLC of guest 2a' (Sample 4, ee 38.9%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.430	8015810	181334	10.763	15.125	30.5327
2	19.728	18237362	159155	18.741	26.517	69.4673

5.9. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2a (Sample 5, ee 26.1%).



5.10. HPLC of guest 2a' (Sample 5, ee 26.0%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.356	12345298	279349	10.709	14.997	36.9798
2	19.736	21038580	177489	18.688	27.051	63.0202

5.11. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2a (Sample 6, ee 11.0%).



5.12. HPLC of guest 2a' (Sample 6, ee 11.4%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.378	10945741	251330	10.773	15.168	44.3138
2	19.986	13754757	123222	18.955	26.112	55.6862

5.13. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 7, ee 80.2%).



5.14. HPLC of guest 2f (Sample7, ee 82.5%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	11.297	35709491	1591188	10.624	12.800	91.2619
2	15.330	3419074	132513	14.656	16.469	8.7381

5.15. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 8, ee 60.0%).



5.16. HPLC of guest 2f (Sample 8, ee 59.2%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.622	35890342	1614320	10.048	12.267	79.6094
2	14.435	9192717	350725	13.739	16.395	20.3906

5.17. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 9, ee 52.7%).



5.18. HPLC of guest 2f (Sample 9, ee 52.8%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.581	17042169	860334	10.037	12.171	76.4180
2	14.415	5259086	202128	13.760	16.267	23.5820

5.19. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 10, ee 40.8%).



5.20. HPLC of guest 2f (Sample 10, ee 41.7%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.757	18745435	953030	10.208	12.203	70.8530
2	14.663	7711347	301972	14.037	16.213	29.1470

5.21. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 11, ee 32.5%).



5.22. HPLC of guest 2f (Sample 11, ee 32.5%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.457	10705423	566954	9.899	11.509	66.2670
2	14.218	5449570	220488	13.547	15.413	33.7330

5.23. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 12, ee 21.2%).



5.24. HPLC of guest 2f (Sample 12, ee 20.6%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.768	11302331	582636	10.219	12.149	60.2997
2	14.686	7441270	290554	14.037	16.075	39.7003

5.25. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 13, ee 10.5%).



5.26. HPLC of guest 2f (Sample 13, ee 9.6%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.790	16737062	856071	10.240	12.149	54.7864
2	14.672	13812622	541397	13.984	16.448	45.2136

5.27. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 14, ee -42.2%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.354	8648303	472606	9.835	11.488	28.0206
2	14.080	22215786	891115	13.515	16.096	71.9794

5.29. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d and guest 2f (Sample 15, ee -55.0%).



5.30. HPLC of guest 2f (Sample 15, ee -55.7%).



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.327	7208227	400148	9.845	11.232	22.1710
2	14.028	25303762	1016915	13.419	16.032	77.8290

# 6. Copies of <sup>1</sup>H NMR spectra/HPLC data for the fast evaluation of reaction conditions in catalytic asymmetric aldol reaction between acetone and tryptanthrin.

General procedure: tryptanthrin (0.1 mmol) and catalyst (20 mmol %) were added to 0.8 mL of acetone and stirred under the specific temperature until the reaction was completed; then acetone was added to the reaction mixture until the volume of resulting solution reached 10 mL; 1.0 mL of the reaction mixture solution was subsequently removed to a flask and dried for <sup>1</sup>H NMR analysis in the presence of 0.01 mmol **1d**.

(1). amino acid salt (20 mol%) neat acetone, conditions (2). 1 equiv. of 1d directly chiral <sup>1</sup> H NMR analysis <i>without purification</i>							
entry	cat.	temp.	S:R (by NMR)	S:R (by HPLC)	absolute error		
1	$H_2N \leftarrow CO_2K$	0°C	80.7 : 19.3	81.7 : 18.3	2.0		
2		0°C	51.8 : 48.2	50.6 : 49.4	2.4		
3	CO <sub>2</sub> K	0°C	73.0 : 27.0	72.6 : 27.4	0.8		
4	O <sup>t</sup> -Bu CO <sub>2</sub> K	0°C	75.8 : 24.2	76.3 : 23.7	1.0		
5	H <sub>2</sub> N CO <sub>2</sub> K	-10 °C	87.7 : 12.3	88.6 : 11.4	1.8		



6.1. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of reaction mixture without purification.

6.2. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of entry 1, *S*:*R*= 80.7 : 19.3.



6.3. HPLC of entry 1, *S*:*R*= 81.7 : 18.3.





6.4. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of entry 2, *S*:*R* = 51.8 : 48.2.

#### 6.5. HPLC of entry 2, *S*:*R* = 50.6 : 49.4.



6.6. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of entry 3, *S*:*R* =73.0 : 27.0.



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.413	31340187	1535056	9.643	12.085	72.5607
2	14.112	11851517	483419	13.461	15.808	27.4393

6.8. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of entry 4, *S*:*R* =75.8 : 24.2.



#### 6.9. HPLC of entry 4, *S*:*R* =76.3 : 23.7.



Peak	Ret. Time	Area	Height	Peak Start	Peak End	Area%
1	10.726	28020828	1415441	10.123	12.181	76.2566
2	14.587	8724599	342495	14.016	16.149	23.7434



6.10. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of entry 5, *S*:*R* =87.7 : 12.3.

#### 6.11. HPLC of entry 5, *S*:*R* = 88.6 : 11.4.



# 7. Copies of <sup>1</sup>H NMR spectra of control experiments.

7.1. <sup>1</sup>H NMR (500 MHz, ACETONE-D<sub>6</sub>) of CPA 1d (0.01 mmol, 1.0 equiv.) and guest 4 (0.01 mmol, 1.0 equiv.).



7.1. <sup>1</sup>H NMR (500 MHz, ACETONE-D6) of CPA 1d (0.01 mmol, 1.0 equiv.) and guest 5 (0.01 mmol, 1.0 equiv.).

