## SUPPLEMENTARY INFORMATION

## Modulation of i-Motif Thermal Stability by Insertion of Anthraquinone Monomers

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Circular Dichroism (CD) spectral analysis of human telomeric DNA (ON1) and anthraquinone-modified i-motif variants (ON2-ON33, Fig. S1-S3):



Figure S1. Circular dichroism (CD) spectra of human telomeric i-motif DNA (ON1) and anthraquinone-modified i-motif variants (ON2-ON13) measured at 20 °C. For sample composition see table 1.



Wavelength / nm

Figure S2. CD spectra of human telomeric i-motif DNA (ON1) and anthraquinone-modified i-motif variants (ON14-ON29) measured at 20 °C. For sample composition see table 1.



Wavelength / nm

Figure S3. CD spectra of human telomeric i-motif DNA (ON1) and anthraquinone-modified i-motif variants (ON30-ON33) measured at 20 °C. For sample composition see table 1.

Normalized UV absorption melting curves and first derivatives of human telomeric i-motif DNA (ON1) and anthraquinone-modified i-motif variants (ON2-ON33, Fig S4-S6):



**Figure S4.** Normalized UV absorption melting curves (Up) and first derivatives (Down) versus temperature difference (0.5 °C/min) from 25-65 °C for i–motifs (**ON1-ON13**) at 295 nm (4  $\mu$ M) in potassium buffer (100 mM KCl + 20 mM K<sub>2</sub>HPO<sub>4</sub> (pH= 5.5) and 1 mM K<sub>2</sub>EDTA). Melting temperatures are within the uncertainty  $\pm$  0.5 °C as determined by repetitive experiments and *T<sub>m</sub>* values were calculated taking an average of the two melting curves.



**Figure S5.** Normalized UV absorption melting curves (Up) and first derivatives (Down) versus temperature difference (0.5 °C/min) from 25-65 °C for i–motifs (**ON1** and **ON14-ON29**) at 295 nm (4  $\mu$ M) in potassium buffer (100 mM KCl + 20 mM K<sub>2</sub>HPO<sub>4</sub> (pH= 5.5) and 1 mM K<sub>2</sub>EDTA). Melting temperatures are within the uncertainty ± 0.5 °C as determined by repetitive experiments and  $T_m$  values were calculated taking an average of the two melting curves.





**Figure S6.** Normalized UV absorption melting curves (Up) and first derivatives (Down) versus temperature difference (0.5 °C/min) from 25-65 °C for i–motifs (**ON1** and **ON30-ON33**) at 295 nm (4  $\mu$ M) in potassium buffer (100 mM KCl + 20 mM K<sub>2</sub>HPO<sub>4</sub> (pH= 5.5) and 1 mM K<sub>2</sub>EDTA). Melting temperatures are within the uncertainty ± 0.5 °C as determined by repetitive experiments and  $T_m$  values were calculated taking an average of the two melting curves.

Table S1. MALDI-TOF mass spectrometry data (calculated and found mass) of i-Motifs

Oligo	bases replaced	Sequence	M <sub>calcd</sub>	M <sub>found</sub>
ON1	wild type	5'-CCC TAA CCC TAA CCC TAA CCC T-3'	6504.44	6504.13
ON2	T <sub>10</sub>	5'-CCC TAA CCC H <sub>18</sub> AA CCC TAA CCC T-3'	6617.43	6618.68
ON3	$A_{11}$	5'-CCC TAA CCC TH <sub>18</sub> A CCC TAA CCC T-3'	6608.43	6608.07
ON4	A <sub>12</sub>	5'-CCC TAA CCC TAH <sub>18</sub> CCC TAA CCC T-3'	6608.43	6609.25
ON5	T <sub>10</sub>	5'-CCC TAA CCC H <sub>14</sub> AA CCC TAA CCC T-3'	6617.43	6617.38
ON6	$A_{11}$	5'-CCC TAA CCC TH <sub>14</sub> A CCC TAA CCC T-3'	6608.43	6608.94
ON7	A <sub>12</sub>	5'-CCC TAA CCC TAH <sub>14</sub> CCC TAA CCC-T-3'	6608.43	6608.07
ON8	$T_{10}$	5'-CCC TAA CCC H <sub>15</sub> AA CCC TAA CCC T-3'	6617.43	6618.29
ON9	$A_{11}$	5'-CCC TAA CCC TH <sub>15</sub> A CCC TAA CCC T-3'	6608.43	6609.22
<b>ON10</b>	A <sub>12</sub>	5'-CCC TAA CCC TAH <sub>15</sub> CCC TAA CCC T-3'	6608.43	6609.30
<b>ON11</b>	$T_{10}$	5'-CCC TAA CCC H <sub>26</sub> AA CCC TAA CCC T-3'	6617.43	6617.49
<b>ON12</b>	$A_{11}$	5'-CCC TAA CCC TH <sub>26</sub> A CCC TAA CCC T-3'	6608.43	6608.65
<b>ON13</b>	$A_{12}$	5'-CCC TAA CCC TAH <sub>26</sub> CCC TAA CCC-T-3'	6608.43	6608.36
<b>ON14</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>18</sub> CCC TAA CCC H <sub>18</sub> AA CCC T-3'	6721.56	6722.74
<b>ON15</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>14</sub> CCC TAA CCC H <sub>14</sub> AA CCC T-3'	6721.56	6722.85
<b>ON16</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>15</sub> CCC TAA CCC H <sub>15</sub> AA CCC T-3'	6721.56	6728.08
<b>ON17</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>26</sub> CCC TAA CCC H <sub>26</sub> AA CCC T-3'	6721.56	6721.66
<b>ON18</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>18</sub> CCC TAA CCC H <sub>14</sub> AA CCC T-3'	6721.56	6723.44
ON19	$A_{6}T_{16}$	5'-CCC TAH <sub>18</sub> CCC TAA CCC H <sub>15</sub> AA CCC T-3'	6721.56	6725.84
<b>ON20</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>18</sub> CCC TAA CCC H <sub>26</sub> AA CCC T-3'	6721.56	6721.66
<b>ON21</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>14</sub> CCC TAA CCC H <sub>18</sub> AA CCC T-3'	6721.56	6722.85
<b>ON22</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>14</sub> CCC TAA CCC H <sub>15</sub> AA CCC T-3'	6721.56	6724.04
<b>ON23</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>14</sub> CCC TAA CCC H <sub>26</sub> AA CCC T-3'	6721.56	6722.76
<b>ON24</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>15</sub> CCC TAA CCC H <sub>18</sub> AA CCC T-3'	6721.56	6723.74
<b>ON25</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>15</sub> CCC TAA CCC H <sub>14</sub> AA CCC-T-3'	6721.56	6722.26
<b>ON26</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>15</sub> CCC TAA CCC H <sub>26</sub> AA CCC T-3'	6721.56	6722.18
<b>ON27</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>26</sub> CCC TAA CCC H <sub>18</sub> AA CCC T-3'	6721.56	6719.88
<b>ON28</b>	$A_{6}T_{16}$	5'-CCC TAH <sub>26</sub> CCC TAA CCC H <sub>14</sub> AA CCC T-3'	6721.56	6722.26
ON29	$A_{6}T_{16}$	5'-CCC TAH <sub>26</sub> CCC TAA CCC H <sub>15</sub> AA CCC T-3'	6721.56	6718.12
<b>ON30</b>	$A_{18}$	5'-CCC TAA CCC TAA CCC TAH <sub>18</sub> CCC T-3'	6608.43	6608.28
ON31	$A_{18}$	5'-CCC TAA CCC TAA CCC TAH <sub>14</sub> CCC-T-3'	6608.43	6612.98
ON32	$A_{18}$	5'-CCC TAA CCC TAA CCC TAH <sub>15</sub> CCC T-3'	6608.43	6608.36
<b>ON33</b>	A <sub>18</sub>	5'-CCC TAA CCC TAA CCC TAH <sub>26</sub> CCC T-3'	6608.43	6609.83

## Maldi-TOF Mass Spectrometry Data of Oligonucleotides (ON1-ON33)















































































**ON29** 















