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Supporting Information

Synthesis and Evaluation of Selenium-containing Indole Chalcone and Diarylketone Derivatives as Tubulin Polymerization Inhibition Agents

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1. Chemistry

All chemicals were analytically pure without further purification as commercially available unless otherwise noted. Nuclear magnetic resonance spectra for proton (¹H) and carbon (¹³C) were recorded in CDCl₃ on a Bruker AvanceIII spectrometer, and chemical shifts are expressed in parts per million (ppm) with TMS as the internal standard. Melting points (MP) were determined on SRS-OptiMelt automated melting point instrument. The purity of each compound (>95%)was determined on high-performance liquid chromatography (HPLC), which were recorded using an Agilent LC–MS 6120 instrument and were run on Eclipse Plus C8 column (4.6 × 150 mm, 5 μ m) with two different solvent gradients (acetonitrile/water=90:10), and a flow rate of 0.20mL/min.

1.1. Synthesis procedure of 1-(4-hydroxy-3-methoxy-5-nitrophenyl)ethan-1-one (2)

To a solution of 1-(4-hydroxy-3-methoxyphenyl)ethan-1-one 1 (8.31g, 50.0mmol) in 100 mL of acetic acid at 0 °C, HNO₃ (4ml, 60%) were added in dropwise and then the reaction was stirred at room temperature for 3h. The resulting mixture was poured into ice water and the precipitate was collected by filtration, dried in vacuum oven to yield 2 as a yellow solid used in the following step without further purification. Yield: 92%.

1.2. Synthesis procedure of 1-(3,4-dimethoxy-5-nitrophenyl)ethan-1-one (3)

Compound **2** (6.33g, 30.0mmol) was dissolved in 100ml of acetone, followed by addition of K₂CO₃ (10.37g, 75.0mmol), dimethylsulfate (4.27ml, 45.0mmol), and then the reaction mixture was stirred at 80°C for two days. After the mixture was cooled to room temperature, ammonium hydroxide was added to remove the excess dimethylsulfate. The mixture was extracted with ethyl acetate, washed with water (twice) and brine (once), dried over anhydrous Na₂SO₄, concentrated in vacuo to produce a dark oil, which was purified by column chromatography (petroleum ether: ethyl acetate= 10: 1) on silica gel to afford the intermediate compound **3** as white solid, Yield: 90%. ¹H NMR (400 MHz, CDCl₃) δ 7.88 (s, 1H), 7.72 (s, 1H), 4.04 (s, 3H), 4.01 (s, 3H), 2.63 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 195.01 (s), 154.01 (s), 146.47 (s), 144.14 (s), 132.11 (s), 116.80 (s), 114.32 (s), 61.99 (s), 56.51 (s), 26.18 (s).

1.3. Synthesis procedure of 1-(3-amino-4,5-dimethoxyphenyl)ethan-1-one (4)

To a stirred solution of **3** (2.25g, 10.0mmol) in acetic acid (25ml), iron power (2.80g, 50.0mmol) and 1ml HCl were added. After stirred for 8h at room temperature, the precipitates were filtered, vacuumed and purified by column chromatographic (petroleum ether: ethyl acetate, 8:1) to provide compound **4** as white solid, yield: 72%. ¹H NMR (400 MHz, CDCl₃) δ 7.00 (d, *J* = 8.3 Hz, 1H), 6.95 (d, *J* = 9.8 Hz, 1H), 4.08 (s, 2H), 3.86 (d, *J* = 6.9 Hz, 6H), 2.49 (d, *J* = 10.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 197.52 (s), 152.64 (s), 140.50 (s), 139.68 (s), 133.03 (s), 109.69 (s), 101.92 (s), 59.86 (s), 55.77 (s), 26.36 (s).

1.4. Synthesis procedure of 1-(3,4-dimethoxy-5-selenocyanatophenyl)ethan-1-one (5)

To a suspension of 4 (195mg, 1.0 mmol) in water, 10% HCl (2.5ml) was added in -5 °C. NaNO₂ (100mg, 1.2 mmol) was added and the reaction mixture was stirred for 30 min until the suspension completely dissolved and the solution was adjusted to the pH 6 by the addition of aqueous solution of NaOAc. After half one hour later, KSeCN (217mg, 1.5mmol) was added to the solution and the reaction mixture was stirred for another 3 hours. The mixture was filtered to get a brown yellow solid **5**, which was used in next step without further purification. Yield: 88%.

1.5. Synthesis procedure of 1-(3,4-dimethoxy-5-(methylselanyl)phenyl)ethan-1-one (6)

To a solution of **5** (285mg, 1.0mmol) in 10ml of MeOH, NaBH₄ (38mg, 1.0 mmol) and CH₃I (92µL, 1.5 mmol) were added at room temperature in sequence. After stirred 2 minutes, the reaction mixture was diluted with saturated NaHSO₄ solution and then extracted with ethyl acetate, the combined extracts were dried, evaporated the solvent and purified by column chromatography (petroleum ether: ethyl acetate/5: 1) to yield the key intermediate **6** as white solid. Yield: 81%. ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 1.9 Hz, 1H), 7.40 (d, *J* = 1.9 Hz, 1H), 3.95 (s, 3H), 3.93 (s, 3H), 2.60 (s, 3H), 2.34 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 196.70 (s), 152.07 (s), 150.67 (s), 133.77 (s), 127.80 (s), 121.24 (s), 109.95 (s), 60.17 (s), 56.03 (s), 26.42 (s), 5.14 (s).

1.6. General procedure for the preparation of 12, 13 and 14.

To a stirred solution of ketone **6** (137 mg, 0.5 mmol) and aldehyde **7** or **8**, **10** and **11** (0.5 mmol), which were synthesized according to a literature method,¹⁻² in methanol (8 mL), piperidine (151 μ L, 1.5mmol) and HOAc (2ml) was added and the resulting mixture was stirred at 95°C for 36-48h. On completion, the slurry was partitioned between water and ethyl acetate, and the mixture was extracted with ethyl acetate (twice). The organic phase were dried over anhydrous Na₂SO₄ and concentrated in vacuum. The crude product was purified by column chromatography (petroleum ether: ethyl acetate=8:1) on silica gel, and then crystallized from ethyl acetate to give target compounds **12**, **13** and **14**).

1.6.1. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(1H-indol-3-yl)prop-2-en-1-one (12a)

Yellow solid, yield: 52%. mp:191.1°C -192.0°C. ¹H NMR (400 MHz, CDCl₃) δ 8.85 (s, 1H), 8.10 (d, *J* = 15.5 Hz, 1H), 7.99 (s, 1H), 7.63 (s, 1H), 7.53 (t, *J* = 7.5 Hz, 2H), 7.47 (s, 2H), 7.32 (s, 2H), 3.95 (d, *J* = 3.6 Hz, 6H), 2.35 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.81 (s, 2H), 152.19 (s, 2H), 149.96 (s, 1H), 139.00 (s, 3H), 137.29 (s, 2H), 135.71 (s, 1H), 130.33 (s, 4H), 127.70 (s, 2H), 125.39 (s, 2H), 123.59 (s, 4H), 121.85 (s, 3H), 120.78 (s, 3H), 120.56 (s, 4H), 117.81 (s, 4H), 114.49 (s, 2H), 112.07 (s, 4H), 110.31 (s, 3H), 60.24 (s, 2H), 56.11 (s, 3H), 5.25 (s, 2H). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₂₀ H₁₉ NO₃ Se, 402.0604; found, 402.0612. Purity: 98.5% (by HPLC).

1.6.2. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(1-methyl-1H-indol-3-yl)prop-2-en-1-one (12b)

Yellow solid, yield: 44%. mp:153.9°C -154.6°C. ¹H NMR (400 MHz, CDCl₃) δ 8.09 (d, *J* = 15.5 Hz, 1H), 8.00 (d, *J* = 7.0 Hz, 1H), 7.56 (d, *J* = 1.7 Hz, 1H), 7.50 (t, *J* = 7.7 Hz, 3H), 7.39 (d, *J* = 3.6 Hz, 1H), 7.36 (dd, *J* = 7.4, 1.6 Hz, 1H), 7.33 (dd, *J* = 6.7, 1.5 Hz, 1H), 3.98 (s, 6H), 3.86 (s, 3H), 2.38 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.51 (s), 152.17 (s), 149.97 (s), 138.65 (s), 138.32 (s), 135.85 (s), 134.47 (s), 127.57 (s), 126.20 (s), 123.22 (s), 121.64 (s), 120.83 (s), 120.65 (s), 116.99 (s), 113.03 (s), 110.41 (s), 110.19 (s), 60.20 (s), 56.12 (s), 33.29 (s), 5.24 (s). HRMS (ESI) (*m/z*) [M+H] ⁺ calcd for C₂₁ H₂₁ NO₃ Se, 416.0761; found, 416.0770. Purity: 98.5% (by HPLC). Purity: 99.7% (by HPLC).

1.6.3. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(4-methoxy-1Hindol-3-yl)prop-2-en-1-one (12c)

Yellow solid, yield: 56%. mp:162.5°C -163.4°C. ¹H NMR (400 MHz, CDCl₃) δ 8.86 (s, 1H), 8.45 (d, *J* = 15.7 Hz, 1H), 7.66 (d, *J* = 2.1 Hz, 1H), 7.55 (d, *J* = 1.6 Hz, 1H), 7.53 (d, *J* = 15.8 Hz, 1H), 7.48 (d, *J* = 1.6 Hz, 1H), 7.20 (t, *J* = 8.0 Hz, 1H), 7.05 (d, *J* = 8.1 Hz, 1H), 6.67 (d, *J* = 7.8 Hz, 1H), 4.01 (s, 3H), 3.98 (s, 3H), 3.96 (s, 3H), 2.36 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.45 (s), 154.80 (s), 152.13 (s), 149.87 (s), 140.65 (s), 138.45 (s), 135.80 (s), 127.43 (s), 125.73 (s), 124.27 (s), 121.12 (s), 119.01 (s), 116.08 (s), 115.01 (s), 110.53 (s), 105.02 (s), 101.89 (s), 60.21 (s), 56.05 (s), 55.35 (s), 5.22 (s). HRMS (ESI) (*m/z*) [M+H] ⁺ calcd for C₂₁ H₂₁ NO₄ Se, 432.0710; found, 432.0708. Purity: 97.2% (by HPLC).

1.6.4. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(4-methoxy-1methyl-1H-indol-3-yl)prop-2-en-1-one (12d)

Yellow solid, yield: 60%. mp:138.8°C -140.1°C. ¹H NMR (400 MHz, CDCl₃) δ 8.44 (d, *J* = 15.7 Hz, 1H), 7.54 (d, *J* = 1.7 Hz, 1H), 7.52 (s, 1H), 7.47 (d, *J* = 1.5 Hz, 1H), 7.45 (d, *J* = 12.3 Hz, 1H), 7.23 (t, *J* = 8.0 Hz, 1H), 6.97 (d, *J* = 8.2 Hz, 1H), 6.68 (d, *J* = 7.9 Hz, 1H), 4.01 (s, 3H), 3.97 (s, 3H), 3.96 (s, 3H), 3.83 (s, 3H), 2.36 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 190.15 (s), 154.84 (s), 152.09 (s), 149.74 (s), 140.49 (s), 139.43 (s), 135.92 (s), 130.05 (s), 127.32 (s), 123.87 (s), 121.03 (s), 118.10 (s), 116.64 (s), 113.49 (s), 110.50 (s), 103.19 (s), 101.81 (s), 60.18 (s), 56.04 (s), 55.37 (s), 33.56 (s), 5.22 (s). HRMS (ESI) (*m/z*) [M+H] ⁺ calcd for C₂₂ H₂₃ NO₄ Se, 446.0866; found, 446.0869. Purity: 98.2% (by HPLC).

1.6.5. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(5-methoxy-1Hindol-3-yl)prop-2-en-1-one (12e)

Yellow solid, yield: 41%. mp:201.7°C -202.5°C. ¹H NMR (400 MHz, CDCl₃) δ 8.60 (s, 1H), 8.11 (d, *J* = 15.5 Hz, 1H), 7.63 (s, 1H), 7.55 (d, *J* = 1.8 Hz, 1H), 7.50 (d, *J* = 1.9 Hz, 1H), 7.48 (d, *J* = 15.7 Hz, 1H), 7.44 (d, *J* = 2.3 Hz, 1H), 7.37 (d, *J* = 8.8 Hz, 1H), 6.99 (dd, *J* = 8.8, 2.4 Hz, 1H), 3.98 (s, 3H), 3.97 (s, 3H), 3.94 (s, 3H), 2.37 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.74 (s), 155.68 (s), 152.18 (s), 149.86 (s), 139.12 (s), 135.73 (s), 132.14 (s), 130.46 (s), 127.82 - 127.25 (m), 126.15 (s), 120.59 (s), 117.12 (s), 114.43 - 113.93 (m), 113.40 (s), 112.74 (s), 110.22 (s), 102.69 (s), 60.24 (s), 56.05 (s), 55.85 (s), 5.11 (s). HRMS

(ESI) (m/z) [M+H] ⁺ calcd for C₂₁ H₂₁ NO₄ Se, 432.0710; found, 432.0711. Purity: 99.0% (by HPLC).

1.6.6. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(5-methoxy-1methyl-1H-indol-3-yl)prop-2-en-1-one (12f)

Yellow solid, yield: 55%. mp:168.5°C -169.9°C. ¹H NMR (400 MHz, CDCl₃) δ 8.06 (d, *J* = 15.5 Hz, 1H), 7.51 (d, *J* = 1.1 Hz, 1H), 7.47 (d, *J* = 1.4 Hz, 1H), 7.46 (s, 1H), 7.40 (d, *J* = 3.0 Hz, 1H), 7.38 (d, *J* = 10.3 Hz, 1H), 7.26 (d, *J* = 2.9 Hz, 1H), 6.98 (dd, *J* = 8.9, 2.1 Hz, 1H), 3.95 (s, 6H), 3.92 (s, 3H), 3.82 (s, 3H), 2.35 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.49 (s), 155.70 (s), 152.16 (s), 149.75 (s), 138.84 (s), 135.89 (s), 134.52 (s), 133.36 (s), 127.59 (s), 126.90 (s), 120.51 (s), 116.17 (s), 112.99 (s), 112.55 (s), 110.98 (s), 110.19 (s), 102.82 (s), 60.21 (s), 56.05 (s), 55.87 (s), 33.54 (s), 5.11 (s). HRMS (ESI) (*m/z*) [M+H] ⁺ calcd for C₂₂ H₂₃ NO₄ Se, 446.0866; found, 446.0865. Purity: 95.8% (by HPLC).

1.6.7. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(6-methoxy-1Hindol-3-yl)prop-2-en-1-one (12g)

Yellow solid, yield: 39%. mp:179.5°C -180.5°C. ¹H NMR (400 MHz, CDCl₃) δ 8.90 (s, 1H), 8.04 (d, *J* = 15.5 Hz, 1H), 7.84 (d, *J* = 8.7 Hz, 1H), 7.52 (s, 1H), 7.50 (s, 1H), 7.48 (d, *J* = 13.8 Hz, 2H), 6.95 (d, *J* = 8.7 Hz, 1H), 6.92 (s, 1H), 3.95 (s, 3H), 3.93 (s, 3H), 3.84 (s, 3H), 2.34 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.95 (s), 157.27 (s), 152.17 (s), 149.91 (s), 139.28 (s), 138.37 (s), 135.76 (s), 129.95 (s), 127.73 (s), 121.18 (s), 120.73 (s), 119.43 (s), 117.40 (s), 114.51 (s), 111.41 (s), 110.28 (s), 95.60 (s), 60.25 (s), 56.10 (s), 55.67 (s), 5.25 (s). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₂₁ H₂₁ NO₄ Se, 432.0710; found, 432.0716. Purity: 97.8% (by HPLC).

1.6.8. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(6-methoxy-1methyl-1H-indol-3-yl)prop-2-en-1-one (12h)

Yellow solid, yield: 48%. mp:166.0°C -167.0°C. ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, J = 15.5 Hz, 1H), 7.84 (d, J = 8.7 Hz, 1H), 7.51 (d, J = 1.6 Hz, 1H), 7.46 (d, J = 1.7 Hz, 1H), 7.42 (d, J = 15.5 Hz, 1H), 7.38 (s, 1H), 6.96 (dd, J = 8.7, 2.2 Hz, 1H), 6.82 (d, J = 2.1 Hz, 1H), 3.95 (s, 6H), 3.91 (s, 3H), 3.78 (s, 3H), 2.36 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.58 (s), 157.19 (s), 152.15 (s), 149.83 (s), 139.30 (s), 138.82 (s), 135.90 (s), 134.08 (s),

127.57 (s), 121.39 (s), 120.69 (s), 120.18 (s), 116.70 (s), 113.13 (s), 110.97 (s), 110.28 (s), 93.97 (s), 60.22 (s), 56.11 (s), 55.76 (s), 33.33 (s), 5.26 (s). HRMS (ESI) (*m/z*) [M+H] ⁺ calcd for C₂₂ H₂₃ NO₄ Se, 446.0866; found, 446.0853. Purity: 98.6% (by HPLC).

1.6.9. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(1H-indol-4-yl)prop-2-en-1-one (13a)

Yellow solid, yield: 67%. mp:131.7°C -132.6°C. ¹H NMR (400 MHz, CDCl₃) δ 8.58 (s, 1H), 8.24 (d, *J* = 15.7 Hz, 1H), 7.66 (d, *J* = 15.6 Hz, 1H), 7.55 (s, 1H), 7.49 (t, *J* = 7.2 Hz, 3H), 7.35 (s, 1H), 7.25 (d, *J* = 2.7 Hz, 1H), 6.88 (s, 1H), 3.96 (s, 3H), 3.94 (s, 3H), 2.34 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.65 (s), 152.22 (s), 150.32 (s), 143.97 (s), 136.41 (s), 135.23 (s), 127.87 (s), 127.39 (s), 127.12 (s), 125.84 (s), 122.14 (s), 122.07 (s), 121.11 (s), 120.89 (s), 113.69 (s), 110.49 (s), 101.40 (s), 60.25 (s), 56.11 (s), 5.25 (s). HRMS (ESI) (*m/z*) [M+H] ⁺ calcd for C₂₀ H₁₉ NO₃ Se, 402.0604; found, 402.0617. Purity: 98.0% (by HPLC).

1.6.10. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(1-methyl-1Hindol-4-yl)prop-2-en-1-one (13b)

Yellow solid, yield: 59%. mp:130.3°C -131.0°C. ¹H NMR (400 MHz, CDCl₃) δ 8.19 (d, *J* = 15.7 Hz, 1H), 7.62 (d, *J* = 15.7 Hz, 1H), 7.50 (s, 1H), 7.47 (d, *J* = 7.4 Hz, 1H), 7.44 (s, 1H), 7.38 (d, *J* = 8.1 Hz, 1H), 7.23 (d, *J* = 8.4 Hz, 1H), 7.16 (d, *J* = 2.8 Hz, 1H), 6.78 (d, *J* = 2.7 Hz, 1H), 3.92 (s, 3H), 3.91 (s, 3H), 3.81 (s, 3H), 2.31 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.57 (s), 152.20 (s), 150.21 (s), 143.89 (s), 137.27 (s), 135.25 (s), 130.42 (s), 127.94 (s), 127.83 (s), 127.16 (s), 122.10 (s), 121.60 (s), 120.98 (s), 120.51 (s), 111.81 (s), 110.38 (s), 99.77 (s), 60.25 (s), 56.11 (s), 33.12 (s), 5.25 (s). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₂₁ H₂₁ NO₃ Se, 416.0761; found, 416.0766. Purity: 99.4% (by HPLC).

1.6.11. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(1H-indol-5yl)prop-2-en-1-one (14a)

Yellow solid, yield: 50%. mp:150.3°C -151.8°C. ¹H NMR (400 MHz, CDCl₃) δ 8.56 (s, 1H), 7.99 (d, J = 15.5 Hz, 1H), 7.93 (s, 1H), 7.59 – 7.52 (m, 2H), 7.48 (d, J = 4.8 Hz, 1H), 7.46 (d, J = 9.2 Hz, 1H), 7.43 (d, J = 8.5 Hz, 1H), 7.27 (s, 1H), 6.63 (d, J = 2.4 Hz, 1H), 3.98 (d, J = 12.0 Hz, 3H), 3.96 – 3.90 (m, 3H), 2.36 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.56 (s), 152.19 (s), 150.26 (s), 147.17 (s), 137.37 (s), 135.40 (s), 128.34 (s), 127.73 (s), 126.98 (s),

125.52 (s), 123.04 (s), 122.03 (s), 121.14 (s), 119.13 (s), 111.73 (s), 110.51 (s), 103.60 (s), 60.23 (s), 56.12 (s), 5.33 (s). HRMS (ESI) (m/z) [M+H] ⁺ calcd for C₂₀ H₁₉ NO₃ Se, 402.0604; found, 402.0612. Purity: 99.4% (by HPLC).

1.6.12. (E)-1-(3,4-dimethoxy-5-(methylselanyl)phenyl)-3-(1-methyl-1Hindol-5-yl)prop-2-en-1-one (14b)

Yellow solid, yield: 57%. mp:142.7°C -143.6°C. ¹H NMR (400 MHz, CDCl₃) δ 7.98 (d, *J* = 15.5 Hz, 1H), 7.90 (s, 1H), 7.57 (d, *J* = 8.4 Hz, 1H), 7.53 (s, 1H), 7.46 (t, *J* = 7.6 Hz, 2H), 7.34 (d, *J* = 8.5 Hz, 1H), 7.08 (d, *J* = 2.4 Hz, 1H), 6.55 (d, *J* = 2.1 Hz, 1H), 3.95 (s, 6H), 3.81 (s, 3H), 2.35 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 189.42 (s), 152.17 (s), 150.13 (s), 147.19 (s), 138.13 (s), 135.45 (s), 130.21 (s), 128.83 (s), 127.71 (s), 126.43 (s), 123.25 (s), 121.62 (s), 120.99 (s), 118.86 (s), 110.39 (s), 109.86 (s), 102.21 (s), 60.24 (s), 56.12 (s), 33.05 (s), 5.34 (s). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₂₁ H₂₁ NO₃ Se, 416.0759; found, 416.0761. Purity: 99.2% (by HPLC).

1.7. Synthesis procedure of 4-bromo-2-methoxyphenol (16)

To a solution of 2-methoxyphenol (**15**, 1.24g, 10.0mmol) and NBS (2.13g, 12.0mmol) in CH3CN at 0 °C for 4h. After the reaction, the solvent diluted with Na₂SO₃ aqueous solution and then extracted with ethyl acetate, the combined extracts were dried. The solvent evaporated and the residue purified by column chromatography (petroleum ether: ethyl acetate=10: 1) to yield 16 as colorless oil, yield: 89%. ¹H NMR (400 MHz, CDCl₃) δ 6.95 (dt, J = 5.1, 2.1 Hz, 2H), 6.77 (d, J = 8.3 Hz, 1H), 5.84 (s, 1H), 3.79 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 147.34 (s), 144.86 (s), 124.16 (s), 115.90 (s), 114.26 (s), 111.65 (s), 56.17 (s).

1.8. Synthesis procedure of 4-bromo-2-methoxy-6-nitrophenol (17)

Intermediate 17 were obtained by the same procedure as compound 2. Yellow solid, yield: 88%. ¹H NMR (400 MHz, CDCl₃) δ 7.76 (s, 1H), 7.12 (s, 1H), 3.87 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 150.76 (s), 145.74 (s), 134.08 (s), 120.79 (s), 118.15 (s), 110.89 (s), 57.01 (s).

1.9. Synthesis procedure of 5-bromo-1,2-dimethoxy-3-nitrobenzene (18)

Intermediate **18** were obtained by the same procedure as compound **3**. Colorless oil, yield: 92%. ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 2.2 Hz, 1H), 7.21 (d, *J* = 2.2 Hz, 1H), 3.96 (s, 3H), 3.93 (s, 3H). ¹H NMR (400 MHz, CDCl₃) δ 7.46 (d, *J* = 2.2 Hz, 1H), 7.21 (d, *J* = 2.2 Hz, 1H), 7.21 (d, *J* = 2.2 Hz, 1H), 3.96 (s, 3H), 3.93 (s, 3H).

1.10. Synthesis procedure of 5-bromo-2,3-dimethoxyaniline (19)

Intermediate **19** were obtained by the same procedure as compound**4**. White solid, yield: 75%. ¹H NMR (400 MHz, CDCl₃) δ 6.53 (s, 1H), 6.45 (s, 1H), 3.88 (s, 2H), 3.82 (s, 3H), 3.79 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 152.38 (s), 140.67 (s), 133.77 (s), 115.68 (s), 110.50 (s), 104.73 (s), 58.84 (s), 54.90 (s).

1.11. Synthesis procedure of 5-bromo-1,2-dimethoxy-3-selenocyanatobenzene (20)

Intermediate **20** were obtained by the same procedure as compound **5**. Brown yellow solid, yield: 92%. ¹H NMR (400 MHz, CDCl₃) δ 7.33 (d, *J* = 2.0 Hz, 1H), 7.02 (d, *J* = 2.0 Hz, 1H), 3.90 (s, 3H), 3.87 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 152.56 (s), 144.02 (s), 122.79 (s), 120.33 (s), 118.09 (s), 116.27 (s), 100.99 (s), 60.80 (s), 56.28 (s).

1.12. Synthesis procedure of (5-bromo-2,3-dimethoxyphenyl)(methyl)selane (21)

Intermediate **21** were obtained by the same procedure as compound **6**. White solid, yield: 80%. ¹H NMR (400 MHz, CDCl₃) δ 6.87 (s, 2H), 3.83 (s, 3H), 3.83 (s, 3H), 2.25 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 152.75 (s), 145.34 (s), 129.63 (s), 121.87 (s), 117.29 (s), 113.31 (s), 60.00 (s), 56.09 (s), 5.03 (s).

1.13. General procedure for the preparation of 23.

To a solution of (5-bromo-2,3-dimethoxyphenyl)(methyl)selane **21** (310mg, 1.0 mmol) in 20mL of anhydrous THF in nitrogen atmosphere at -78°C, n-butyllithium (0.8mL, 2.5M in hexane) was added in dropwise. After the solution was stirred for 0.5h, **22a-22c**, which were synthesized according to literature methods, ³ was added, and then the reaction mixture was stirred for overnight in darkness. A saturated NH₄Cl solution was added and the mixture was extracted by ethyl acetate. The solvent was evaporated and the crude product was purified by flash column chromatography to get colorless oil **23**.

1.13.1. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-(phenylsulfonyl)-1Hindol-3-yl)methanol (23a)

Colorless oil, yield: 50%. ¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, *J* = 8.4 Hz, 1H), 7.88 (d, *J* = 7.7 Hz, 2H), 7.54 (t, *J* = 7.3 Hz, 1H), 7.46 (d, *J* = 4.0 Hz, 2H), 7.43 (d, *J* = 6.9 Hz, 2H), 7.31 (t, *J* = 7.8 Hz, 1H), 7.18 (t, *J* = 7.6 Hz, 1H), 6.81 (d, *J* = 17.8 Hz, 2H), 5.96 (s, 1H), 3.86 (s, 3H), 3.77 (s, 3H), 2.16 (s, 3H).

1.13.2. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-(phenylsulfonyl)-1Hindol-4-yl)methanol (23b)

Colorless oil, yield: 70%. ¹H NMR (400 MHz, CDCl₃) δ 7.93 (d, J = 8.2 Hz, 1H), 7.88 – 7.83 (m, 2H), 7.53 (d, J = 3.8 Hz, 1H), 7.50 (d, J = 7.4 Hz, 1H), 7.40 (t, J = 7.7 Hz, 2H), 7.30 – 7.25 (m, 1H), 7.21 (d, J = 7.3 Hz, 1H), 6.78 – 6.71 (m, 3H), 6.02 (s, 1H), 3.80 (s, 3H), 3.70 (s, 3H), 2.08 (s, 3H).

1.13.3. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-(phenylsulfonyl)-1Hinden-5-yl)methanol (23c)

Colorless oil, yield: 70%. ¹H NMR (400 MHz, CDCl₃) δ 7.93 (d, J = 8.6 Hz, 1H), 7.85 (d, J = 7.7 Hz, 2H), 7.54 (d, J = 3.7 Hz, 1H), 7.53 – 7.48 (m, 2H), 7.41 (t, J = 7.8 Hz, 2H), 7.31 (dd, J = 8.6, 1.4 Hz, 1H), 6.76 (d, J = 10.2 Hz, 2H), 6.62 (d, J = 3.6 Hz, 1H), 5.81 (d, J = 2.5 Hz, 1H), 3.82 (s, 3H), 3.76 (s, 3H), 2.17 (s, 3H).

1.14. General procedure for the preparation of 24.

The colorless oil **23** was dissolved in THF, and IBX (1.2eq) was added. After stirred for 2 days, the mixture was diluted with ethyl acetate and water, the organic phase was separated, washed with saturated NaCl solution (twice) and dried with anhydrous sodium sulfate. The solvent was removed in vacuum, and the residue was purified by flash column chromatography (petroleum ether: ethyl acetate=8:1) to afford compounds **24**.

1.14.1. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-(phenylsulfonyl)-1Hindol-3-yl)methanone (24a)

White oil, yield: 82%. ¹H NMR (400 MHz, CDCl₃) δ 8.18 (d, *J* = 7.5 Hz, 1H), 8.01 – 7.91 (m, 2H), 7.86 (d, *J* = 7.7 Hz, 2H), 7.51 (t, *J* = 7.1 Hz, 1H), 7.40 (t, *J* = 7.5 Hz, 2H), 7.37 – 7.28 (m, 2H), 7.24 (d, *J* = 7.7 Hz, 2H), 3.90 (s, 3H), 3.84 (s, 3H), 2.19 (s, 3H). ¹³C NMR (101

MHz, CDCl₃) δ 188.32 (s), 151.14 (s), 148.90 (s), 136.59 (s), 134.42 (s), 134.04 (s), 133.59 (s), 131.94 (s), 128.63 (s), 127.49 (s), 127.24 (s), 126.01 (s), 125.09 (s), 123.90 (s), 121.89 (s), 120.19 (s), 119.50 (s), 112.18 (s), 109.76 (s), 59.20 (s), 55.06 (s), 3.99 (s).

1.14.2. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-(phenylsulfonyl)-1Hindol-4-yl)methanone (24b)

White oil, yield: 83%. ¹H NMR (400 MHz, CDCl₃) δ 8.23 (d, J = 8.3 Hz, 1H), 7.91 (d, J = 8.0 Hz, 2H), 7.70 (d, J = 3.7 Hz, 1H), 7.57 – 7.52 (m, 2H), 7.46 (t, J = 7.5 Hz, 2H), 7.39 (t, J = 7.9 Hz, 1H), 7.31 (d, J = 2.5 Hz, 1H), 7.21 (d, J = 5.4 Hz, 1H), 7.04 (d, J = 3.6 Hz, 1H), 3.95 (s, 3H), 3.85 (s, 3H), 2.14 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 194.95 (s), 151.93 (s), 150.12 (s), 138.09 (s), 135.47 (s), 134.14 (s), 130.48 (s), 129.45 (s), 128.09 (s), 126.84 (s), 126.39 (s), 123.72 (s), 122.75 (s), 120.31 (s), 117.23 (s), 113.57 (s), 111.69 (s), 109.28 (s), 102.01 (s), 60.22 (d, J = 4.0 Hz), 56.06 (s), 4.98 (s).

1.14.3. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-(phenylsulfonyl)-1Hindol-5-yl)methanone (24c)

White oil, yield: 88%. ¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, *J* = 8.6 Hz, 1H), 7.93 (s, 1H), 7.84 (d, *J* = 8.2 Hz, 2H), 7.73 (d, *J* = 8.7 Hz, 1H), 7.60 (d, *J* = 3.5 Hz, 1H), 7.51 (t, *J* = 7.2 Hz, 1H), 7.40 (t, *J* = 7.7 Hz, 2H), 7.14 (d, *J* = 10.7 Hz, 2H), 6.68 (d, *J* = 3.6 Hz, 1H), 3.88 (s, 3H), 3.80 (s, 3H), 2.12 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 195.30 (s), 151.91 (s), 149.85 (s), 138.00 (s), 136.97 (s), 134.30 (s), 134.23 (s), 133.21 (s), 130.35 (s), 129.47 (s), 127.76 (s), 127.72 (s), 126.87 (s), 126.48 (s), 124.26 (s), 122.55 (s), 113.21 (s), 111.77 (s), 109.61 (s), 60.21 (s), 56.11

1.15. General procedure for the preparation of 25.

To a solution of 24a-24c (51.5mg, 0.1mmol) in 10mL of methanol, NaOH (12mg, 0.3mmol) was added and the reaction was stirred for 3h. The mixture was extracted with ethyl acetate (twice) and the organic phase was washed with saturated brine (twice) and dried over Na₂SO₄. The solvent was evaporated in vacuum and the crude product was purified by column chromatography (petroleum ether: ethyl acetate/5:1) over silica gel to give the target compounds 25.

1.15.1. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1H-indol-3-yl)methanone (25a)

White soild, yield: 77%. mp:217.6°C -218.9°C. ¹H NMR (400 MHz, DMSO) δ 8.25 (dd, *J* = 6.5, 1.9 Hz, 1H), 8.09 (s, 1H), 7.53 (dd, *J* = 6.6, 1.7 Hz, 1H), 7.28 (d, *J* = 1.4 Hz, 1H), 7.28 – 7.22 (m, 2H), 7.20 (d, *J* = 1.4 Hz, 1H), 3.89 (s, 3H), 3.83 (s, 3H), 2.28 (s, 3H). ¹³C NMR (101 MHz, DMSO) δ 189.14 (s), 151.97 (s), 147.94 (s), 137.69 (s), 137.02 (s), 136.04 (s), 127.78 (s), 126.78 (s), 123.62 (s), 122.36 (s), 121.90 (s), 119.71 (s), 115.32 (s), 112.66 (s), 110.97 (s), 59.95 (s), 56.36 (s), 4.74 (s). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₁₈ H₁₇NO₃ Se, 376.0447; found, 376.0441. Purity: 98.9% (by HPLC).

1.15.2. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1H-indol-4-yl)methanone (25b)

White soild, yield: 71%. mp:139.5°C -140.4°C. ¹H NMR (400 MHz, CDCl₃) δ 8.64 (s, 1H), 7.60 (d, J = 8.1 Hz, 1H), 7.47 (d, J = 7.4 Hz, 1H), 7.35 – 7.30 (m, 3H), 7.23 (t, J = 7.8 Hz, 1H), 6.96 (s, 1H), 3.96 (s, 3H), 3.86 (s, 3H), 2.18 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 196.23 (s), 151.84 (s), 149.65 (s), 136.65 (s), 135.39 (s), 129.33 (s), 127.53 (s), 127.35 (s), 126.43 (s), 124.32 (s), 122.69 (s), 120.77 (s), 115.55 (s), 111.88 (s), 103.55 (s), 60.21 (s), 56.05 (s), 4.97 (s). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₁₈ H₁₇ NO₃ Se, 376.0447; found, 376.0442. Purity: 98.7% (by HPLC).

1.15.3. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1H-indol-5-yl)methanone (25c)

White soild, yield: 69%. mp:160.0°C -161.2°C. ¹H NMR (400 MHz, CDCl₃) δ 8.68 (s, 1H), 8.16 (s, 1H), 7.76 (dd, J = 8.5, 1.5 Hz, 1H), 7.45 (d, J = 8.6 Hz, 1H), 7.31 – 7.28 (m, 1H), 7.27 (d, J = 0.7 Hz, 2H), 6.66 (s, 1H), 3.96 (s, 3H), 3.87 (s, 3H), 2.21 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 196.20 (s), 151.83 (s), 149.34 (s), 138.31 (s), 135.43 (s), 129.80 (s), 127.50 (s), 127.26 (s), 125.81 (s), 124.99 (s), 124.23 (s), 122.35 (s), 111.87 (s), 110.97 (s), 104.25 (s), 60.18 (s), 56.08 (s), 5.03 (s). HRMS (ESI) (m/z) [M+H] ⁺ calcd for C₁₈ H₁₇NO₃ Se, 376.0447; found, 376.0438. Purity: 98.7% (by HPLC).

1.16. Synthesis procedure of (3,4-dimethoxy-5-(methylselanyl)phenyl) -(1methyl-1H-indol-4-yl)methanol (26) Intermediate **26** was obtained by the same procedure as compound **23**.Colorless oil, yield: 72%. ¹H NMR (400 MHz, CDCl₃) δ 7.26 (d, *J* = 8.1 Hz, 1H), 7.21 (t, *J* = 7.6 Hz, 1H), 7.14 (d, *J* = 7.0 Hz, 1H), 7.03 (d, *J* = 3.0 Hz, 1H), 6.90 (d, *J* = 16.6 Hz, 2H), 6.53 (d, *J* = 2.9 Hz, 1H), 6.15 (s, 1H), 3.81 (s, 3H), 3.77 (s, 6H), 2.18 (s, 3H).

1.17. (3,4-dimethoxy-5-(methylselanyl)phenyl)(1-methyl-1H-indol-4-yl)methanone (27)

Compound 27 was obtained by the same procedure as compound 24. Yellow oil, yield: 83%. 1H NMR (400 MHz, CDCl3) δ 7.56 (d, J = 8.2 Hz, 1H), 7.48 (d, J = 7.3 Hz, 1H), 7.33 (s, 2H), 7.31 – 7.27 (m, 1H), 7.20 (d, J = 3.1 Hz, 1H), 6.89 (d, J = 3.0 Hz, 1H), 3.97 (s, 3H), 3.88 (s, 3H), 3.87 (s, 3H), 2.20 (s, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 196.03 (s), 151.83 (s), 149.63 (s), 137.50 (s), 135.47 (s), 131.01 (s), 129.43 (s), 127.90 (s), 127.46 (s), 123.86 (s), 122.66 (s), 120.31 (s), 113.52 (s), 111.89 (s), 102.04 (s), 60.19 (s), 56.05 (s), 33.03 (s), 4.98 (s). HRMS (ESI) (*m*/*z*) [M+H] ⁺ calcd for C₁₉ H₁₉ NO₃ Se, 390.0604; found, 390.0595. Purity: 98.3% (by HPLC).

2. Biological Assays⁴

2.1. Cell Lines and Culture.

The human cancer cell lines (A549, HELA, HEPG2, RKO, HCT116, MGC803, MDAMB231) used in this study were purchased from the Laboratory Animal Service Center at Sun Yat-sen University (Guangzhou, China). Cell lines A549, HELA, RKO, and MDAMB231 were cultivated in DMEM containing 10% (v/v) heat-inactivated fetal bovine serum (FBS), 100 units/mL penicillin, and 100 μ g/mL streptomycin. Cell lines HCT116 and HEPG2 were cultivated in RPMI 1640 medium containing 10% (v/v) heat-inactivated FBS, 100 units/mL penicillin, and 100 mg/mL streptomycin. The cells were incubated at 37 °C under a 5% CO₂ and 90% relative humidity (RH) atmosphere. *2.2.* **MTT Assay.**

Cells grownin the logarithmic phase were seeded into 96-well plates (5×10^3 cells/well) for 24 h, and then exposed to different concentrations of the test compounds for 48 h. After attached cells were incubated with 5 mg/mL MTT (Sigma, USA) for another 4 h, the suspension was discarded, and subsequently the dark blue crystals (formazan) were solubilized in dimethyl

sulfoxide (DMSO). The absorbance of the solution at 570 nm was measured using a multifunction microplate reader (Molecular Devices, Flex Station 3), and each experiment was performed at least in triplicate. IC_{50} values, which represent the drug concentrations required to cause 50% cancer cell growth inhibition, were used to express the cytotoxic effects of each compound and were calculated with GraphPad Prism Software version 5.02 (GraphPad Inc., La Jolla, CA, USA).

2.3. In vitro Tubulin Polymerization

Assay. A tubulin polymerization assay was performed by measuring the increase in fluorescence intensity, which can be easily recorded due to the incorporation of a fluorescent reporter, DAPI (4',6-diamidino-2-phenylindole), a fluorophore that is known to be a DNA intercalator. A commercial kit (cytoskeleton, cat. #BK011P) was used for the tubulin polymerization. The final buffer used for tubulin polymerization contained 80.0 mM piperazine-N, N'-bis(2-ethanesulfonic acid) sequisodium salt (pH 6.9), 2.0 mM MgCl2, 0.5 mM EGTA, 1 mM GTP, and 10.2% glycerol. First, 5 µL of the tested compounds at the indicated concentrations was added, and the mixture was warmed to 37 °C for 1 min; then, the reaction was initiated by the addition of 55 µL of the tubulin solution. The fluorescence intensity enhancement was recorded every 60 sec for 90 min in a multifunction microplate reader (Molecular Devices, Flex Station 3) (emission wavelength at 410 nm, excitation wavelength at 340 nm). The area under the curve was used to determine the concentration that inhibited tubulin polymerization by 50% (IC₅₀) and was calculated using GraphPad Prism Software version 5.02 (GraphPad Inc., La Jolla, CA, USA).

2.4. Immunofluorescence Microscopy.

In a 10 mm confocal culture dish, 3×10^4 cells were grown for 24 h and then incubated in the presence/absence of compound **25b** at the indicated concentrations for another 12 h. After being washed with phosphate-buffered solution (PBS) and fixed in 4% pre-warmed (37 °C) paraformaldehyde for 15 min, the cells were permeabilized with 0.5% Triton X-100 for 15 min and blocked for 30 min in 10% goat serum. Then, the cells were incubated with mouse anti-tubulin antibody (CST, USA) at 4 °C overnight, washed with PBS three times, and incubated with goat anti-mouse IgG/Alexa-Fluor 488 antibody (Invitrogen, USA) for 1 h. The samples were immediately visualized on a Zeiss LSM 570 laser scanning confocal

microscope (Carl Zeiss, Germany) after the nuclei were stained with Hoechst 33342 (Sigma,

USA) in the dark at room temperature for 30 min.

2.5. Cell Cycle Analysis.

A549 cells were seeded in 6-well plates $(3 \times 10^5 \text{ cells/well})$, incubated in the presence/absence of compound **25b** at the indicated concentrations for 24 or 49 h, harvested by centrifugation, and then fixed in ice-cold 70% ethanol overnight. After the ethanol was removed the next day, the cells were re-suspended in ice-cold PBS, treated with RNAse A (Keygen Biotech, China) at 37 °C for 30 min, and then incubated with the DNA staining solution propidium iodide (PI, Keygen Biotech, China) at 4 °C for 30 min. Approximately 10,000 events were detected by flow cytometry (Beckman Coulter, Epics XL) at 488 nm. The data regarding the number of cells in different phases of the cell cycle were analysed using EXPO32 ADC analysis software.

2.6. Effect of 24b on cell apoptosis progression and decreased the mitochondrial membrane potential of A549 cells

The A549 cell sample was prepared following the same protocols used for the cell cycle analysis. After incubation, cells were harvested and incubated with 5 μ L of Annexin-V/FITC (Keygen Biotech, China) in binding buffer (10 mM HEPES, 140 mM NaCl, and 2.5 mM CaCl₂ at pH 7.4) at room temperature for 15 min. PI solution was then added to the medium for another 10 min-incubation. Almost 10,000 events were collected for each sample and analysed by flow cytometry (Beckman Coulter, Epics XL). The percentage of apoptotic cells was calculated using EXPO32 ADC Analysis software.

A lipophilic cationic dye, 5,5',6,6'-tetrachloro-1,1',3,3'-tetraethyl-benzimidazolcarbocyanine (JC-1, Beyotime, China) was used to monitor the level of MMP in the cells. At normal state, the MMP is high and JC-1 appears as aggregates, which indicated by red fluorescence. When apoptosis occurs, the MMP reduced and JC-1 displayed as monomers, which indicated by green fluorescence. For the fluorescence microscopy detection, A549 cells were plated in 6-well plates (3×10^5 cells/well) and incubated for 24 h, and treated with 25b at the indicated concentrations for another 48 h. Then, the cells were stained with 2 μ M JC-1 at 37 °C for 30 min, washed with PBS, and then the cell nuclei were stained with Hoechst 33342 (Sigma, USA) for 10 min in the dark. The cell images were immediate detected by a fluorescence microscopy (EVOS FL Auto).

SI1. NMR spectrums of target compounds

12a



12b





12c

12d 20161121-zs-k18-h ^{8.46}
^{8.42} -2.36 -13000 CH₃ 12000 0 11000 H₃C H₃C 10000 0 CH3 S 9000 H₃C H (d) 7.45 8000 s) 96 (s) 83 7000 A (d) E (d) B (d) 8.44 7.54 6.68 H D (t) 7.23 M (s) 2 36 I 4 s) 1 6000 s) 97 5000 G (d) 7.47 4000 3000 -2000 1000 W 00110 9 8 7 fl (ppn) 0 3.04 2.82 2.86 83 -1000 Ņ 6 fl (ppm) 16 15 14 13 12 11 10 5 ó -1 -2 -3 -4 3 2 20161121-zs-k18-C og V. 158.92 135.92 15.92 15.92 15.92 15.92 15.92 15.92 15.92 15.92 15.92 15.95 60.18 56.04 55.37 -33.56 -190.1 -5.22 -800 750 o o -700 H₃C 650 H₃C C 600 CH3 Se H₂C 550 $\begin{array}{l} {}^{13} C \ NMR \left({101 \ MHz, \ CDC4} \right) \delta \ 190.15 \ (s), \ 154.84 \ (s), \ 152.09 \ (s), \ 149.74 \ (s), \ 140.49 \ (s), \ 139.43 \ (s), \ 135.92 \ (s), \ 130.05 \ (s), \ 127.32 \ (s), \ 123.87 \ (s), \ 121.03 \ (s), \ 118.10 \ (s), \ 118.40 \ (s), \ 113.49 \ (s), \ 110.50 \ (s), \ 103.19 \ (s), \ 101.81 \ (s), \ 60.18 \ (s), \ 55.37 \ (s), \ 33.56 \ (s), \ 43.42 \ (s), \ 118.10 \ (s)$ 500 450 R (s) M (s) J (s) 130.05 116.64 103.19 V (s) 149.74 D (s) 56.04 400 W (s) 152.09
 T
 (s)
 Q
 (s)
 L
 (s)
 I
 (s)

 139.43
 127.32
 113.49
 101.81
 C (s) 55.37 B (s) 33.56 Y (s) 190.15 A (s) 5. 22 350 E (s) 60.18 X (s) 154.84 S (s) P (s) K (s) 135.92 123.87 110.50 300 0 (s) 121.03 250 -200 150 -100 -50 -0 -50 120 110 100 fl (ppm) 80 70 50 40 20 60 30 210 200 190 170 160 150 140 130 90 10 ò -10 180

12e



12f 20161120-zs-k19-h 3.95 3.92 3.82 -2.35 -19000 18000 Ö 17000 H₃C 16000 H₃C -15000 CH2 14000 H₃C -13000 $\begin{array}{c} ^{1}\mathrm{H}\ \mathrm{NMR}\ (400\ \mathrm{MHz},\ \mathrm{CDCls}) \in 8.06\ (\mathrm{d}, J=15.5\ \mathrm{Hz},\ \mathrm{Hh}),\ 7.51\ (\mathrm{d}, J=1.1\ \mathrm{Hz},\ \mathrm{HH}),\ 7\frac{4}{7} \left(\frac{\mathrm{d}}{\mathrm{d}} J=1.4\ \mathrm{Hz},\ \mathrm{Hz},\ \mathrm{Hz}),\ 7.40\ (\mathrm{d}, J=3.0\ \mathrm{Hz},\ \mathrm{HH}),\ 7.38\ (\mathrm{d}, J=10.3\ \mathrm{Hz},\ \mathrm{HH}),\ 7.26\ (\mathrm{d}, J=2.9\ \mathrm{Hz},\ \mathrm{Hz}),\ 6.98\ (\mathrm{d}, J=8.9,\ 2.1\ \mathrm{Hz},\ \mathrm{HH}),\ 3.95\ (\mathrm{s},\ \mathrm{SH}),\ 3.92\ (\mathrm{s},\ \mathrm{SH}),\ 2.35\ (\mathrm{s},\ \mathrm{SH}). \end{array}$ 12000 E (s) 7.46 -11000 C (d) 7.51] (s) 92 10000 A (d) B (dd) 8.06 6.98 L (s) 2.35 I 3 (s) 95 9000 D (d) 7.47 8000 (s) 82 7000 F (d) 7.40 6000 H (d) 7.26 5000 4000 3000 2000 1000 -0 1.00 0.96 0.98 0.98 0.98 85 95 07 -1000 in mini ŝ 16 15 14 13 12 11 10 9 8 7 6 fl (ppm) 5 -1 -2 -3 3 2 Ó 20161120-zs-k19-C 4 -1600 102.89 102.89 112.55 11 000 60.21 56.05 55.87 -33.54 -155.7 -152.1 -149.7 -5.11 -189. 1500 1400 0 H₃C 1300 H₃C 0 1200 CH3 H₃C -1100 $\begin{array}{c} {}^{11}\text{C}\ \text{NMR}\ (101\ \text{MHz},\ \text{CDCh})\ 5\ 189.49\ (s),\ 155.70\ (s),\ 152.16\ (s),\ 149.75\ (s),\ 13884\ (\phi),\ 135.89\ (s),\ 13$ 1000 900 V (s) 155.70 T (s) 0 (s) 138.84 126.90 I (s) 110.19 C (s) 56.05 800 W (s) 189.49 M (s) H (s) 116.17 102.82 B (s) 33.54 U (s) 152.16 P (s) 127.59 D (s) 60.2 A (s) 5.11 700 F (s) 149.75 Q (s) 133.36 (s) 1.98 (s) . 87 E 600 (s) 89 500 99 400 300 200 100 and to be delay Alinability Want -0 -100 110 100 fl (ppm) 70 40 20 180 130 120 90 80 60 50 30 10 0 210 200 190 170 160 150 140 -10



12h







13b







25a

25c

SI3. HPLC chromatograms of target compounds.

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	80
1	8.165	BV	0.1970	67.00445	5.17786	1.4714
2	9.340	VB	0.2856	4486.91357	234.13512	98.5286
总量	: :			4553.91802	239.31297	

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰 #	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 %
1	8.179	BV	0.2088	73.55861	5.40434	0.3007
2	10.057	BB	0.3095	2.43891e4	1187.92786	99.6993

总量:

2.44626e4 1193.33220

峰	保留时间	类型	峰宽 [min]	峰面积	峰高	峰面积
#	[min]	-	[min]	[mAU^S]		
1	8.138	BV	0.1933	67.26139	5.32593	0.3697
2	9.151	VV	0.2798	1.76861e4	930.24316	97.2094
3	10.125	VB	0.4454	440.45602	13.12862	2.4209
总量	:			1.81938e4	948.69771	

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰 (R留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积
1	8.179	BV	0.2005	70.70094	5.41041	0.7568
2	9.957	BB	0.3013	9181.02930	451.31256	98.2700
3	10.914	BB	0.3214	90.92791	3.96668	0.9733

总量:

9342.65815 460.68965

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰(呆留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	용
1	8.145	BV	0.2003	76.09689	5.90598	0.9855
2	9.090	VB	0.2803	7645.70947	404.81860	99.0145
总量	:			7721.80637	410.72459	

12f

峰(#	保留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积 8
1	8.361	MM	0.2887	85.30326	4.92386	4.1880
2	9.709	BB	0.3106	1951.53650	97.06363	95.8120

总量:

2036.83976 101.98749

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积	
#	# [min]		[min] [mAU*s]		[mAU]	8	
1	8.138	BV	0.1829	55.53136	4.80186	0.6977	
2	9.106	VB	0.2789	7784.39307	411.21252	97.8075	
3	3 10.026	BB	0.3509	118.96532	4.83712	1.4947	

总量:

7958.88974 420.85151

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	00
1	8.152	BV	0.1965	68.20457	5.28724	0.5732
2	9.053	VV	0.4694	88.49655	2.54707	0.7437
3	9.751	VB	0.2976	1.17428e4	581.52692	98.6831
总量	:			1.18995e4	589.36122	

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	8
1	8.166	BV	0.2678	106.48360	5.65262	2.0353
2	9.289	VB	0.2933	5125.34082	263.03516	97.9647
总量				5231.82442	268.68777	

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	90
1	8.135	BV	0.1986	67.54684	5.23152	0.2208
2	10.175	BB	0.3088	3.04227e4	1450.09656	99.4347
3	13.656	BB	0.3643	105.40075	4.29526	0.3445
总量	:			3.05957e4	1459.62333	

14a					
DAD1 D, Sig=254,4 Ref=600,	100 (20140301-17_LC 2016-11	I-18 20-36-02\20161118-ZS	-K14.D)		
mAU 1		69			
1		5			
E 003					
000					
500 -					
400					
300 -					
200					
100					
		133			
		,			
1		7.5	40.5	 	 00.6

峰	保留	时间	类型	峰宽	峰面积	峰高	峰面积
#	[1	min]		[min]	[mAU*s]	[mAU]	8
			-				
1	. 1	8.133	BV	0.2138	80.13026	5.63687	0.5709
2	2	9.189	VB	0.2903	1.39568e4	713.20044	99.4291
总量	:				1.40370e4	718.83731	

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰 (4) (#)	R留时间 [min]	类型	峰宽 [min]	峰面积 [mAU*s]	峰高 [mAU]	峰面积
						1
1	8.138	BV	0.1960	63.72637	4.95745	0.7339
2	9.977	BB	0.3129	8619.78613	414.03085	99.2661
总量	:			8683.51250	418.98830	

Intermediate

2

SI3. HPLC chromatograms of target compounds 12a

12d

12c

12f

12g

13a

14b

峰保	留时间	类型 峰	宽 屿	峰面积	峰高	峰面积
#	[min]	[m	in] [m	AU*s]	[mAU]	oto
-	-					<mark> </mark>
1	7.347 E	BB 0.	2049 2	2.68031	1.64424	0.3287
2	7.894 E	BB 0.	1914 5	4.67888	4.32884	0.7925
3	8.837 E	BB 0.	2953 682	2.19580	359.69595	98.8788
总量:			689	9.55499	365.66903	

25b

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰 保留时间 类型	峰宽	峰面积	峰高	峰面积
# [min]	[min]	[mAU*s]	[mAU]	90
1 7.851 BV	0.1720	52.59585	4.71885	0.4622
2 8.443 VV	0.2959	97.33435	4.81411	0.8553
3 9.098 VB	0.2817	1.12302e4	591.02191	98.6825
总量:		1.13801e4	600.55487	

-
2
3
5

27

信号 1: DAD1 D, Sig=254,4 Ref=600,100

峰	保留时间	类型	峰宽	峰面积	峰高	峰面积
#	[min]		[min]	[mAU*s]	[mAU]	olo
		-				
1	8.713	BB	0.4158	54.50431	1.74787	1.6700
2	10.058	BB	0.2803	3209.15015	171.52037	98.3300
总量	:			3263.65446	173.26824	

SI4. HR-MS of target compounds

12a

	402	0612							7
2.400e6-									
2.200e6-									
2.000e6-									
1.800e6-									
1.600e6-		400.0626							
1.400e6-		100.0020							
1.200e6-									
1.000e6-									
8.000e5-	404.0610	403.0646							
6.000e5-	398.0618	399.0625							
4.000e5-		440.0186							
2.000e5-	401.0647								
01,,	200 200 0 10		700.0 800.0	000 0 100	0.0 1100.0	1200.0	1200 0 14	000	2
100.0 20	0.0 300.0 40	0.0 500.0 600.0	700.0 000.0	900.0 100	00.0 1100.0	1200.0	1300.0 14	00.0	
Rank Score Formu	ula (M)	lon	1	leas. m/z	Pred. m/z D	f. (mDa)	Df. (ppm)	Iso	DBE
1 97.53 C20 H	19 N O3 Se	[M+H]+		402.0612	402.0604	0.8	1.99	100.00	12.0

Event#: 1 MS(E+) Ret. Time : 1.147 -> 1.147 Scan# : 173 -> 173

12b

vent#: 1 MS(E+) R	et. Time : 1.200 -> 1.20	0 Scan#: 181 -> 181	
	416.0	1770	
3.000e6			
2.500e6-			
2.000e6		414.0778	
1.500e6	Į.		
1.000e6	418.0781	417.0788	
5.000e5	412.0788	-413.0792 -415.0799	
0-1	200.0 300.0 400.0	0 500.0 600.0 700.0 800.0 900.0 1000.0 1100.0 1200.0 1300.0 1400.	0

Rank	Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1	97.10 C21 H21 N O3 Se	[M+H]+	416.0770	416.0761	0.9	2.16	100.00	12.0

12c

Event#: 1 MS(E+) Ret. Time : 1.040 -> 1.040	Scan# : 157 -> 157
432.0	0708
2.000e6	
1.800e6	
1.600e6	
1.400e6	430 0742
1.200e6	
1.000e6	
8.000e5 434.0722	433.0753
6.000e5-	428 0742
4.000e5	4200742
2.000e5	-431.0738
100.0 200.0 300.0 400.0	500.0 600.0 700.0 800.0 900.0 1000.0 1100.0 1200.0 1300.0 1400.0

Rank	Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1	100.00 C21 H21 N O4 Se	[M+H]+	432.0708	432.0710	-0.2	-0.46	100.00	12.0

Event#: 1100 / 1100 - 1100	Scall# . 171-2 171
446.0	0869
2.500e6	
2.000e6-	444.0884
1.500e6-	
1.000e6 448.0876	447.0905
5.000e5- 258.9871	
0 ¹	500.0 600.0 700.0 800.0 900.0 1000.0 1100.0 1200.0 1300.0 1400.0

Event#: 1 MS(E+) Ret. Time : 1.133 -> 1.133 Scan# : 171 -> 171

Rank Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1 100.00 C22 H23 N O4 Se	[M+H]+	446.0869	446.0866	0.3	0.67	100.00	12.0

12e

12d

Event#: 1 MS(E+)	Ret. Time : 1.067 -> 1.067	Scan#: 161 -> 161
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	432.0)71 <mark>1</mark>									
2.000e6											
1.800e6											
1.600e6											
1.400e6											
1.200e6		430.0729									
1.000e6											
8.000e5	404.0715	433 0750									
6.000e5	434.0715	400.0700									
4.000e5	428.0750	429.0748									
2.000e5	Ì	-431.0749									
0 ¹											
100.0	200.0 300.0 400.0	500.0 60	0.0 700.0	800.0	900.0	1000.0	1100.0	1200.0	1300.0	1400.0	
						_					_

Rank	Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1	99.48 C21 H21 N O4 Se	[M+H]+	432.0711	432.0710	0.1	0.23	99.48	12.0

12f

Event#: 1 MS(E+)	Ret. Time : 1.147	-> 1.147	Scan#:1	73 -> 17	3								
		446.	0865										
2.400e6- 2.200e6- 2.000e6- 1.800e6-													
1.600e6- 1.400e6- 1.200e6- 1.000e6- 8.000e5- 6.000e5- 2.000e5- 0	44 44 214.0880	48.0888 13.0901	444.088 447.087 442.089 445.087	4 4 3 8									
100.0	200.0 300.0	400.0	500.0	600.0	700.0	800.0	900.0	1000.0	1100.0	1200.0	1300.0	1400.0	
Rank Score F	ormula (M)		lon			Mea	s. m/z	Pred. m/	z Df. (r	nDa) D	of. (ppm)	Iso	DBE
1 100.00 C	22 H23 N O4 Se		[M+	H]+		446	6.0865	446.086	6	-0.1	-0.22	100.00	12.0

LVent#. I WO(L+) Re	t. Time . 1.000 -> 1.000	Scall# . 105 -> 10	5						
	432.0	0716							
3.000e6-									
2.500e6									
2.000e6-		430.0723							
1.500e6-									
1.000e6	434.0728	433.0710							
	428.0738	429.0742							
5.000e5 200	0.0708 432.4274	431.0770							
0 ¹ 100.0 20	00.0 300.0 400.0	500.0 600.0	700.0 800.0	900.0	1000.0	100.0 1200	.0 1300.0	1400.0	
Rank Score Form	ula (M)	lon	Me	as. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1 99.03 C21 H	H21 N O4 Se	[M+H]+	43	2.0716	432.0710	0.6	1.39	100.00	12.0

Event#: 1 MS(E+) Ret. Time : 1.080 -> 1.080 Scan# : 163 -> 163

12h

EVent#: MS(E+) Ret. Ime: 1,133 -> 1,133 Scan#: 1/1	-> 171
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446.	0853			
3.000e6				
2.500e6				
2.000e6	444.0862			
1.500e6 448.0874 1.000e6 442.0879 5.000e5 214.0860 446.4479	447.0897 443.0901 445.0883			
0 ¹ 100.0 200.0 300.0 400.0	500.0 600.0 700.0	800.0 900.0	1000.0 1100.0 120	0.0 1300.0 1400.0
Rank Score Formula (M) 2 95.22 C22 H23 N O4 Se	lon [M+H]+	Meas. m/z 446.0853	Pred. m/z Df. (mDa) 446.0866 -1.3	Df. (ppm) Iso DBE

13a

Event#: 1 MS(E+) Ret. Time : 1.0	053 -> 1.053 Scan# : 159 -> 159						
1.400e6-	402.0617						
1.200e6-							
1.000e6-							
8.000e5	400.0629						
6.000e5							
4.000e5 40	403.0665						
2.000e5-	438.0132	825.1028	0954				
0 ¹ , 100.0 200.0 300	0.0 400.0 500.0 600.0 700	0.0 800.0 900.0	1000.0 11	00.0 1200.0	0 1300.0	1400.0	
Rank Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1 94.42 C20 H19 N O3 S	6e [M+H]+	402.0617	402.0604	1.3	3.23	100.00	12.0

12g

	416.0	0766						
2.500e6-								
2.000e6								
1.500e6-		414.0777						
1.000e6-	418.0780	417.0812						
5.000e5-	415.0813 285.0018	413.0811 -412.0815						
01, 11	00.0 200.0 300.0 400.	0 500.0 600.0	700.0 800.0 900	0.0 1000.0 1	100.0 1200.	0 1300.0	1400.0	
Rank Sco 1 99.	ore Formula (M) .50 C21 H21 N O3 Se	lon [M+H]+	Meas. m/ 416.076	z Pred. m/z 6 416.0761	Df. (mDa) 0.5	Df. (ppm) 1.20	lso 100.00	

Event#: 1 MS(E+) Ret. Time : 1.240 -> 1.240 Scan# : 187 -> 187

14a

Event# 1 MS(E+)	Ret Time : 1 040 -> 1 040	Scan# : 157 -> 157
EVenter. TIVIO(E.)	Het. Hille . 1.040 - 1.040	Scarim . 107 - 107

	402.0	612						
1.600e6								
1.400e6								
1.200e6								
1.000e6		400.0630						
8.000e5-	Í							
6.000e5-	404.0604	403.0618						
4.000e5-	398.0641	399.0675						
2.000e5-	401.0630-	424.0433		823.100	825 0958			
0 ¹	200.0 300.0 400	.0 500.0	600.0 700	.0 800.0	900.0 100	0.0 1100.0	1200.0 1300.0	1400.0

Rank	Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1	97.53 C20 H19 N O3 Se	[M+H]+	402.0612	402.0604	0.8	1.99	100.00	12.0

14b

Event#: 1 MS(E+) F	Ret. Time : 1.147 -> 1.14	7 Scan#: 173 -> 17	3					
	416.0	759						
2.500e6-								
2.000e6		414.0779						
1.500e6-	/							
1.000e6-	418.0756 412.0787	417.0768						
5.000e5-	415.0808-	-413.0809						
0	200.0 300.0 400.0	500.0 600.0	700.0 800.0	900.0 1000.	.0 1100.0 120	0.0 1300.0	1400.0	
Rank Score For 1 100.00 C21	mula (M) IH21 N O3 Se	lon [M+H]+	Me 41	as. m/z Pred. 6.0759 416.0	m/z Df. (mDa)	Df. (ppm) -0.48	lso 100.00	

13b

25a

	376.	0441						
1.600e6								
1.400e6								
1.200e6-								
1.000e6		374.0454						
8.000e5-								
6.000e5	373 0449	378.0440						
4.000e5-	364.1019	377.0440						
2.000e5	345,1547	372.0473 375.0468						
0 ¹ , 100	0.0 200.0 300.0	400.0 500.0 600.0	700.0 800.0 900	.0 1000.0 1	100.0 1200.0	0 1300.0	1400.0	
Rank Score	E_Formula (M)	lon	Meas. m/	z Pred. m/z	Df. (mDa)	Df. (ppm)	lso	DBE
1 98.5	C18 H17 N O3 Se	[M+H]+	376.044	1 376.0447	-0.6	-1.60	100.00	11.0

Event#: 1 MS(E+) Ret. Time : 1.037 -> 1.037 Scan# : 208 -> 208

25b

Event#: 1 MS(E+) Ret. Time : 0.962 -> 0.962 Scan# : 193 -> 193

37	76.0442
1.800e6	
1.600e6	
1.400e6	
1.200e6	
1.000e6	374.0433
8.000e5	
6.000e5	377.0460
4.000e5 283.1128	378.0440
2.000e5 165.0541	398.0273
100.0 200.0 300.0	400.0 500.0 600.0 700.0 600.0 900.0 1000.0 1100.0 1200.0 1300.0 1400.0

Rank	Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1	99.17 C18 H17 N O3 Se	[M+H]+	376.0442	376.0447	-0.5	-1.33	100.00	11.0

25c

	376.	438	
1	070.	100	
1.100e6-			
1.000e6-			
9.000e5-			
8.000e5-		374.0438	
7.000e5-			
6.000e5-			
5.000e5-			
4.000e5-	377.0445	378.0426	
3.000e5-	372 0449		
2.000e5-	214 0901	-373.0480	
1.000e5- 375	5.0472	773.0611	
01		771.0542	
100.0	200.0 300.0 4	00.0 500.0 600.0 700.0 800.0 900.0	1000.0 1100.0 1200.0 1300.0 1400.0

Rank	Score Formula (M)	lon	Meas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
1	96.53 C18 H17 N O3 Se	[M+H]+	376.0438	376.0447	-0.9	-2.39	100.00	11.0

	390.059	95	-						
2.500e6-									
2.000e6									
1.500e6-	3	88.0598							
1.000e6	392.0599	391.0617							
5.000e5-	386.0610 3 390.3875	389.0652							
0 100.0 200.0	274 10 300.0 400.0	005.0002 0 500.0 600.0	700.0 800.0	900.0	1000.0 11	100.0 1200.	0 1300.0	1400.0	
L									
Rank Score Formula	(M)	lon	M	eas. m/z	Pred. m/z	Df. (mDa)	Df. (ppm)	Iso	DBE
96.72 C19 H19	N 03 Se	[M+H]+		90.0595	390.0604	-0.9	-2.31	100.00	11.0

Event#: 1 MS(E+) Ret. Time : 1.057 -> 1.057 Scan# : 212 -> 212

Reference;

- C. J. Trabbic, J. H. Overmeyer, E. M. Alexander, E. J. Crissman, H. M. Kvale, M. A. Smith, P. W. Erhardt, W. A. Maltese, *J. Med. Chem.*, 2015, 58, 2489-512.
- 2 M. S. Pedras, A. Abdoli, *Bioorg. Med. Chem.*, 2013, 21, 4541-4549.
- 3 M. Z. Zhan, Q. Chen, N. Mulholland, *Eur. J. Med. Chem.*, 2012, **53**, 283-291.
- 4 J. Yan, J. Chen, S. Zhang, J. Hu, L. Huang, X. Li, J. Med. Chem., 2016, 59, 5264–5283.

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