Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2017

SUPPORTING INFORMATION

Volatiles from the fungal microbiome of the marine sponge *Callyspongia* cf. *flammea*

Lena Barra,^a Paul Barac,^b Gabriele M. König,^b Max Crüsemann^b and Jeroen S. Dickschat^{*a}

a. Kekulé-Institute for Organic Chemistry and Biochemistry, University of Bonn, Gerhard-Domagk-Straße 1, D-53121 Bonn, Germany

E-mail: dickschat@uni-bonn.de

b. Institute of Pharmaceutical Biology, University of Bonn, Nußallee 6, D-53115 Bonn, Germany

TABLE OF CONTENTS

1. LIST OF IDENTIFIED VOLATILES	3 - 5
2. HEADSPACE EXTRACT OF Sporormiella sp. 293 K05	6
3. HEADSPACE EXTRACT OF <i>Botrytis</i> sp. 293 K02	7
4. MASS SPECTRA OF ISOTORQUATONE (15) AND TORQUATONE	8
5. IDENTIFICATION OF ISOTORQUATONE BY GC-MS	9
6. DETERMINATION OF THE ABSOLUTE CONFIGURATION OF 15	10
7. MASS SPECTRA OF DESMETHYL ANALOGS 16, 17 and 19	11
8. RESULTS OF FEEDING EXPERIMENTS	12
9. BIOACTIVITY TESTS AGAINST BACTERIA	13
10. NMR SPECTRA OF SYNTHETIC COMPOUNDS	14-43

1. LIST OF IDENTIFIED VOLATILES

Compound ^a	Þ	/(Lit.)℃	ldent. ^d	occurrence (strain) ^e
furan-3-carbaldehyde (34)	816		ms, std	St
butyl acetate (35)	819	814 ¹	ms, ri, std	St
furan-2-carbaldehyde	835	835 ²	ms, ri, std	St
2-furanmethanol	854	850 ³	ms, ri, std	D, St
ethyl 2-methylbutyrate (45)	856	861 ⁴	ms, ri	E
ethylbenzene	860	858 ⁵	ms, ri, std	Е, В
m-xylene	868	8665	ms, ri, std	E, B
protoanemonin (6)	881	880 ⁶	ms, ri	D
cyclohexanol (46)	886	886 ⁷	ms, ri, std	E
2-acetylfuran	909	909 ⁸	ms, ri, std	D, Sp
2,5-hexanedione	927	931 ⁹	ms, ri, std	St,
2-hydroxy-5-methylhexan-3-one (36)	942	944 ¹⁰	ms, ri	St
4-methyl-γ-butyrolactone (37)	954	958 ¹¹	ms, ri	St
benzaldehyde	959	952 ⁸	ms, ri, std	B, Sp
1-octen-3-ol (38)	978	975 ¹²	ms, ri, std	St, E
1-ethyl-4-methylbenzene	968	965 ¹³	ms, ri, std	В
3-octanon (39)	986	983 ¹⁴	ms, ri, std	St, E
6-methylhept-5-en-2-one (51)	987	981 ⁸	ms, ri, std	В
1,3,4-trimethylbenzene	993	995 ⁸	ms, ri, std	Е, В
2-acetylpyrrole	1058	1054 ⁸	ms, ri, std	D, St, Sp
3,4-dimethylpentan-4-olid (47)	1064	1063 ¹⁵	ms, ri, std	E
acetophenone	1066	1059 ⁸	ms, ri, std	St, Sp
linalool (40)	1100	1095 ⁸	ms, ri, std	St, B
nonanal (7)	1104	1101 ¹⁶	ms, ri, std	D, St, E, B, Sp
2-phenylethanol	1114	1107 ⁸	ms, ri, std	St, Sp
phenylacetonitrile (10)	1138	1134 ⁸	ms, ri, std	D
1,2-dimethoxybenzene (41)	1146	1146 ¹⁷	ms, ri, std	St
2-methylisoborneol (42)	1184	1178 ⁸	ms, ri, std	St, B
decanal (8)	1202	1201 ⁸	ms, ri, std	D, St, E, B, Sp

Table S1. Identified volatile compounds and their occurrence in the investigated strains.

2-phenyloxyethanol	1219	1221 ⁸	ms, ri	St, B, Sp
benzothiazole (43)	1224	1223 ¹⁸	ms, ri	St, E, Sp
undecanal (9)	1307	1305 ¹⁴	ms, ri, std	D, E
3,4-dimethoxystyrene (11)	1364	1368 ¹⁹	ms, ri, std	D
1,3,4-trimethoxybenzene (44)	1368	1373	ms, std	St
dodecanal (48)	1418	1411 ²⁰	ms, ri, std	Е, В
geranylactone (12)	1454	1455 ²¹	ms, ri, std	D, St, B
dauca-4(11),8-diene (13)	1539	1537 ²²	ms, ri	D
hexadecane (49)	1600	1600 ⁸	ms, ri, std	E
(1(10) <i>E</i> ,5 <i>E</i>)-germacradien-11-ol (14)	1649	1638 ²³	ms, ri, std	D
tetradecanol (50)	1676	1676 ⁸	ms, ri, std	E
isotorquatone (15)	1808		ms, std	D
chartabomone (16)	1853		ms, std	D
dichotomone (17)	1884		ms, std	D

^aCompound numbers refer to compound numbers in main text. Unidentified compounds and artifacts are not listed. Compounds which have also been identified from the medium are marked in italics. ^bRetention index on a HP5-MS fused silica capillary column. ^cRetention index on the same or a similar column from tabulated data in the literature. ^dIdentification based on ms: mass spectrum (mass spectral match factor >850), ri: retention index on same or similar column (maximum deviation of 10 points), std: comparison to a synthetic or commercially available standard. ^eLetters refer to fungal strains: *D. cejpii* (D), *Stachylidium* sp. (St), *Emericella* sp. (E), *Sporormiella* sp. (Sp), *Botrytis* sp. (B).

References

- 1 C. A. Citron, L. Barra, J. Wink and J. S. Dickschat, Org. Biomol. Chem., 2015, 13, 2673.
- 2 N. S. Radulovic, N. D. Dordevic and R. M. Palic, J. Serbian Chem. Soc., 2010, 75, 1653.
- 3 D. Ansorena, I. Astiasaran and J. Bello, J. Agric. Food Chem., 2000, 48, 2395.
- 4 J. C. Leffingwell and E. D. Alford, *Electron. J. Environ. Agric. Food Chem.*, 2005, **4**, 899.
- 5 E. Engel, C. Baty, D. L. Corre, I. Souchon and N. Martin, *J. Agric. Food Chem.*, 2002, **50**, 6459.
- 6 N. Radulovic, N. Dordevic, M. Markovic and R. Palic, *Bull. Chem. Soc. Ethio.*, 2010, **24**, 67.
- 7 J. A. Pino, J. Mesa, Y. Munoz, M. P. Marti and R. Marbot, *J. Agric. Food Chem.*, 2005, **53**, 2213.
- 8 R. P. Adams, *Identification of Essential Oil Components by Gas Chromatography/ Mass Spectrometry*, Allured, Carol Stream, **2009**.
- 9 I. Jerkovic and Z. Marijanovic, *Molecules*, 2010, **15**, 3744.
- 10 C. A. Citron, P. Rabe and J. S. Dickschat, J. Nat. Prod., 2012, 75, 1765.
- 11 M. Garcia-Estaban, D. Ansorena, I. Astiasaran, D. Martin and J. Ruiz, *J. Sci. Food Agric.*, 2004, **84**, 1364.
- 12 S. Cavar, M. Maksimovic and M. E. Solic, *Biologica Nyssana*, 2010, 1, 99.
- 13 Z. Wang, M. Fingas, J. Chromatogr. A, 1995, 712, 321.

- 14 M. Miyazawa, S. Marumoto, T. Kobayashi, S. Yoshida and Y. Utsumi, *Rec. Nat. Prod.*, 2011, **5**, 221.
- 15 C. A. Citron, C. Junker, B. Schulz, J. S. Dickschat, Angew. Chem. Int. Ed., 2014, 53, 4346.
- 16 A. Bertoli, M. Lepnardi, J. Krzyzanowska, W. Oleszek, L. Pistelli, *Acta Biochem. Polonica*, 2011, **58**, 581.
- 17 W. N. Setzer, J. A. Noletto, R. O. Lawton, *Flavour Fragr. J.*, 2006, 21, 244.
- 18 T. Nawrath, G. F. Mgode, B. Weetjens, S. H. E. Kaufmann and S. Schulz, *Beilstein J. Org. Chem.*, 2012, **8**, 290.
- 19 C. X. Zhao, X. N. Li, Y. Z. Liang, H. Z. Fang, L. F. Huang and F. Q. Guo, *Chemom. Intell. Lab. Syst.*, 2006, **82**, 218.
- 20 N. Radulovic, P. Blagojevic and R. Palic, *Molecules*, 2010, **15**, 6168.
- 21 N. E. Sandoval-Montemayor, A. Garcia, E. Elizondo-Trevino, E. Garza-Gonzales, L. Alvarez, and M. del Rayo Camacho-Corona, *Molecules*, 2012, **17**, 11173.
- 22 G. M. Petrovic, J. G. Stamenkovic, I. R. Kostevski, G. S. Stojanovic, V. D. Mitic and B. K. Zlatkovic, *Chem. Biodivers.*, 2017, DOI: 10.1002/cbdv.201600367.
- 23 P. Rabe, C. A. Citron and J. S. Dickschat, ChemBioChem, 2013, 14, 2345.

2. HEADSPACE EXTRACT OF Sporormiella sp. 293 K05



Figure S1. Volatiles produced by *Sporormiella*. A) Gas chromatogram of the headspace extract, B) structures of the detected volatiles. Asterisks indicate compounds originating from the medium.



Figure S2. Volatiles produced by *Botrytis*. A) Gas chromatogram of the headspace extract, B) structures of the detected volatiles. Asterisks indicate compounds originating from the medium.



5. IDENTIFICATION OF ISOTORQUATONE BY GC-MS



Figure S4. Total ion chromatograms of A) headspace extract of *D. cejpii*, B) synthetic **15**, C) synthetic **18**.

6. DETERMINATION OF THE ABSOLUTE CONFIGURATION OF 15



Figure S5. Analysis of the absolute configuration of **15** by HPLC on a homochiral stationary phase. A) mixture of synthetic (*R*)-**15** and (*S*)-**15**, B) synthetic (*S*)-**15**, C) natural product from *D. cejpii*.



Figure S6. Mass spectra of A) natural desmethyl analogue **16**, B) natural desmethyl analogue **17**, C) synthetic **16**, D) synthetic **17**, E) synthetic **19**.

8. RESULTS OF FEEDING EXPERIMENTS



Figure S7. Results of feeding experiments in *D. cejpii*. A) Total ion chromatogram of the headspace-extract of *D. cejpii* after feeding of (*methyl*-²H₃)-L-methionine, B) mass spectrum of (${}^{2}H_{18}$)-**15** after feeding of (*methyl*-²H₃)-L-methionine, C) mass spectrum of **15** after feeding of (${}^{2-13}C$)acetate.

9. BIOACTIVITY TESTS AGAINST BACTERIA

Compound	B. megaterium DSM 32 ^a	<i>E. coli</i> DSM 498 ^a
15	0	0
(S)- 15	0	0
16	3	0
17	3	2 ^b
18	0	0
19	2 ^b	1 ^b
25	2 ^b	0
pos. control (ertapenem)	10	8

 Table S2.
 Bioactivity tests against bacteria.

^a Radii of inhibition zones in mm, ^b partial inhibition.



10. NMR SPECTRA OF SYNTHETIC COMPOUNDS

Figure S8. ¹H NMR spectrum of **15** (500 MHz, CDCl₃).



Figure S9. ¹³C NMR spectrum of **15** (125 MHz, CDCl₃).



Figure S10. 13 C DEPT 135 spectrum of 15 (125 MHz, CDCl₃).



Figure S11. ¹H NMR spectrum of 16 (500 MHz, CDCl₃).

0=

НО



Figure S12. ¹³C NMR spectrum of 16 (125 MHz, CDCl₃).



Figure S13. ¹³C DEPT 135 spectrum of **16** (125 MHz, CDCl₃).



Figure S14. ¹H NMR spectrum of 17 (500 MHz, CDCl₃).



Figure S15. ¹³C NMR spectrum of **17** (125 MHz, CDCl₃).



Figure S16. ¹³C DEPT 135 spectrum of **17** (125 MHz, CDCl₃).





Figure S17. ¹H NMR spectrum of **18** (500 MHz, C₆D₆).



Figure S18. ¹³C NMR spectrum of **18** (125 MHz, C₆D₆).



Figure S19. ¹³C DEPT 135 spectrum of 18 (125 MHz, C₆D₆).



Figure S20. ¹H NMR spectrum of **19** (500 MHz, CDCl₃).

0=

0



Figure S21. ¹³C NMR spectrum of **19** (125 MHz, CDCl₃).



Figure S22. 13 C DEPT 135 spectrum of 19 (125 MHz, CDCl₃).



Figure S23. ¹H NMR spectrum of 21 (500 MHz, CDCl₃).



Figure S24. ¹³C NMR spectrum of 21 (125 MHz, CDCl₃).



Figure S25. ¹³C DEPT 135 spectrum of **21** (125 MHz, CDCl₃).



Figure S26. ¹H NMR spectrum of 22 (500 MHz, CDCl₃).



Figure S27. ¹³C NMR spectrum of 22 (125 MHz, CDCl₃).



Figure S28. ¹³C DEPT 135 spectrum of 22 (125 MHz, CDCl₃).





Figure S29. ¹H NMR spectrum of 23 (400 MHz, CDCl₃).



Figure S30. ¹³C NMR spectrum of 23 (100 MHz, CDCl₃).



Figure S31. 13 C DEPT 135 spectrum of 23 (100 MHz, CDCl₃).



Figure S32. ¹H NMR spectrum of 24 (500 MHz, CDCl₃).



Figure S33. ¹³C NMR spectrum of 24 (125 MHz, CDCl₃).



Figure S34. ¹³C DEPT 135 spectrum of 24 (125 MHz, CDCl₃).



Figure S35. ¹H NMR spectrum of 25 (500 MHz, CDCl₃).



Figure S36. ¹³C NMR spectrum of 25 (125 MHz, CDCl₃).



Figure S37. ¹³C DEPT 135 spectrum of 25 (125 MHz, CDCl₃).