Electronic Supplementary Information

Modulation in the Acidity Constant of Acridine Dye with Cucurbiturils: Stimuli-Responsive pK_a Tuning and Dye Relocation into Live Cells

Raman Khurana,^{a,b} Nilotpal Barooah,^a Achikanath C. Bhasikuttan,^{a,b} Jyotirmayee Mohanty^{a,b*} ^aRadiation & Photochemistry Division, Bhabha Atomic Research Centre, Mumbai 400 085, India; ^bHomi Bhabha National Institute, Anushaktinagar, Mumbai, 400 094, India



Figure S1. Absorption spectra of acridine $(2.45 \times 10^{-6} \text{ M})$ in water at different *pH*s. (1) 2.6, (2) 4.0, (3) 4.5, (4) 5.0, (5) 5.6, (6) 6.1, (7) 7.0, (8) 8.1 and (9) 9.1. Inset: variation in absorbance with pH at 354 nm.



Figure S2. (A) Absorption spectra of Ac $(4.5 \times 10^{-6} \text{ M})$ in aqueous solution at different CB7 concentrations at pH 11. [CB7]/ μ M: (1) 0, (2) 5, (3) 20, (4) 77, (5) 113, (6) 157 and (7) 206. (B) Fluorescence spectra of Ac $(4.5 \times 10^{-6} \text{ M})$ in aqueous solution at different CB7 concentrations at pH 11. [CB7]/mM: (1) 0.0, (2) 0.02, (3) 0.04, (4) 0.11, (5) 0.16, (6) 0.21, (7) 0.46, (8) 0.90 and (9) 1.35.



Figure S3. CB7 Concentration dependent normalized absorption spectra of acridine dye at pH ~3.5.

Method M1: 1:1 host: guest binding model

In the present systems, the binding constants (*K*) for the different forms of the dye with the CB7 host were estimated at suitable pH conditions by the fluorescence titration method assuming 1:1 complexation stoichiometry according to eq. S1, which afforded satisfactory fitting results.¹⁻³

$$I_{f} = I_{Dye}^{0} \frac{[Dye]_{eq}}{[Dye]_{0}} + I_{CB7:Dye}^{\infty} \frac{[CB7:Dye]_{eq}}{[Dye]_{0}}$$
(S1)

Where, I_{Dye}^{0} and $I_{CB7:Dye}^{\infty}$ are the extrapolated fluorescence intensities of the uncomplexed and complexed form of the dye, respectively, [Dye]₀ and [CB7]₀ are the respective total concentrations of dye (AcH⁺, Ac forms, as applicable) and CB7

host, and [Dye]_{eq} is the concentration of uncomplexed dye in the solution. Exchange of the dye during its excited-state lifetime (<31 ns), i.e., the conversion of the uncomplexed dye to the complexed one or vice versa, can be excluded since the corresponding rate constants are very small for macrocyclic host molecules.^{2,3} For fitting, the change in the fluorescence intensity (ΔI_f^{λ}) was plotted against the total host concentration and the obtained titration curve was fitted according to the rearranged eq. S2:^{1, 3}

$$\Delta I_f^{\lambda} = \left(1 - \frac{[Dye]_{eq}}{[Dye]_0}\right) \left(I_{CB7:Dye}^{\infty} - I_{Dye}^0\right)$$
(S2)

where, [Dye]_{eq} is expressed as

$$[Dye]_{eq} = \{K_{eq}[Dye]_0 - K_{eq}[SCX6]_0 - 1 + \sqrt{(K_{eq}[Dye]_0 + K_{eq}[CB7]_0 + 1)^2 - 4K_{eq}^2[Dye]_0[CB7]_0\}} / 2K_{eq}$$
(S3)

Method M2: 1:2 host: guest binding model⁴

$$CB + Dye \xrightarrow{\kappa_1} CB : Dye$$
(S4a)

CB : Dye + Dye
$$\leftarrow$$
 CB : (Dye)₂ (S4b)

$$K_{(\text{ternary complex})} = K_1 \times K_2$$

The fluorescence intensity of the system, I, is a function of the intensities of the free guest (I_g) and the 1:1 and 1:2 host-guest complexes (I_{gh} and I_{ggh} , respectively):

$$I = x_g I_g + x_{gh} I_{gh} + 2 x_{ggh} I_{ggh}$$
(S5)

 x_{g} , x_{gh} and x_{ggh} are the mole fractions of the free guest and the 1:1 and 1:2 host-guest complexes.

The mole fractions are further defined as follows:

$$x_{g} = \frac{\left[G\right]}{\left[G\right]_{0}} \tag{S6}$$

$$x_{\rm gh} = \frac{[{\rm HG}]}{[{\rm G}]_0} \qquad \Rightarrow x_{\rm gh} = \frac{K_1 [{\rm H}]_0}{1 + K_1 [{\rm G}] + K_1 K_2 [{\rm G}]^2} \cdot \frac{[{\rm G}]}{[{\rm G}]_0} \tag{S7}$$

$$x_{\rm gh} = \frac{[{\rm HG}_2]}{[{\rm G}]_0} \implies x_{\rm ggh} = \frac{K_1 K_2 [{\rm H}]_0}{1 + K_1 [{\rm G}] + K_1 K_2 [{\rm G}]^2} \cdot \frac{[{\rm G}]^2}{[{\rm G}]_0}$$
(S8)

Thus, the fluorescence intensity of the system is obtained through substitution of eq. (S6-S8) into eq. (S5).

$$I = \frac{[G]}{[G]_{0}} \cdot \frac{\left(1 + K_{1}[G] + K_{1}K_{2}[G]^{2}\right)I_{g} + K_{1}[H]_{0}I_{gh} + 2K_{1}K_{2}[H]_{0}[G]I_{ggh}}{1 + K_{1}[G] + K_{1}K_{2}[G]^{2}}$$
(S9)

The *K* value of CB8:(AcH⁺)₂ has been estimated from the binding isotherm (inset of Fig. 3B) by using eq. S9.



Figure S4. (A) ITC isotherm for titration of AcH⁺ with CB7 in water at 25 °C. (B) ITC isotherm for titration of CB8 with AcH⁺ in water at 25 °C.



Figure S5. Decay traces of AcH⁺ in solutions at pH ~3.5 in the absence (1) and presence (2) of 250 μ M CB7 (A) and 70 μ M CB8 (B). 'L' represents excitation lamp profile. λ_{ex} = 374 nm.



Figure S6. ¹H NMR spectra (500 MHz) of ~100 μ M acridine dye in the absence (a) and in the presence (b) of 1mM CB7 and (c) 80 μ M CB8 in D₂O at pD 4.5. Inset: Pictorial representation of ACH⁺.

Method M3:

The pH-dependent absorbance changes at 355 nm (inset of Fig. S1) were fitted according to the following relation (eq. S10), 3

$$A_{\rm obs} = \frac{A_{\rm AcH^+}^{\infty}}{\{1+10^{\rm pH-pK_a}\}} + \frac{A_{\rm Ac}^{\infty}}{\{1+10^{\rm pK_a-pH}\}}$$
(S10)

where A_{obs} is the observed absorbance at any pH, and $A_{AcH^+}^{\infty}$ and A_{Ac}^{∞} are the extrapolated absorbances of the AcH⁺ and Ac forms, respectively. From this analysis, the p K_a value of the dye in its ground state was found to be 5.4 ± 0.1, which matches well with the reported value.^{3, 5}



Figure S7. Fluorescence spectra of CB7:AcH⁺ complex (A) and CB8:AcH⁺ complex (B) with increasing temperature. T/ $^{\circ}$ C: (1) 20, (2) 30, (3) 40, (4) 50, (5) 60, (6) 70 and (8) 80.



Ac (Ac form)

Ac-CB7 (AcH⁺ form)

Ac-CB7-AD (Ac form)

Figure S8. Fluorescence microscopic images recorded from CHO cell lines at pH 7.4 after treating them with uncomplexed acridine dye (10 μ M) (**a**); acridine dye (10 μ M)-CB7 (1mM) (**b**); and acridine dye-CB7 (1mM)-AD (100 μ M) (c).

References

- 1. J. Mohanty, A. C. Bhasikuttan, W. M. Nau and H. Pal, *J. Phys. Chem. B*, 2006, **110**, 5132-5138.
- 2. M. Shaikh, J. Mohanty, P. K. Singh, W. M. Nau and H. Pal, *Photochem. Photobiol. Sci.*, 2008, **7**, 408–414.
- 3. A. Jadhav, V. S. Kalyani, N. Barooah, D. D. Malkhede and J. Mohanty, *ChemPhysChem*, 2015, **16**, 420-427.
- 4. M. Sayed, F. Biedermann, V. D. Uzunova, K. I. Assaf, A. C. Bhasikuttan, H. Pal, W. M. Nau and J. Mohanty, *Chem. Eur. J.*, 2015, **21**, 691 696.
- 5. M.-J. Ji, J.-G. Kim and U. S. Shin, Bull. Korean Chem. Soc., 2012, 33, 2489.