Electronic Supplementary Material (ESI) for Organic & Biomolecular Chemistry. This journal is © The Royal Society of Chemistry 2017

Bronsted-base-catalyzed remote cascade reactivity of 2,4-dienones – asymmetric synthesis of tetrahydrothiophenes

Artur Przydacz,¹ Rafał Kowalczyk,² and Łukasz Albrecht^{1,*}

¹Institute of Organic Chemistry

Lodz University of Technology

90-924 Łódź, Poland

E-mail: lukasz.albrecht@p.lodz.pl

www.a-teamlab.p.lodz.pl

² Department of Organic Chemistry

Wroclaw University of Science and Technology

50-370 Wrocław, Poland

Contents

1. General methods	S2
2. Synthetic procedures	S3
2.1 Procedure for synthesis of S-Acetyl-2-mercapto-1-phenylethanone	S3
2.2 Procedure for synthesis of 2-mercapto-1-phenylethanone 3	S5
2.3 Brønsted-base-catalyzed remote cascade – general procedure	S7
3. Crystal and X-ray data for (2 <i>S</i> ,3 <i>S</i> ,4 <i>S</i>)-cyclohexyl 4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)-4-	
phenyltetrahydrothiophene-2-carboxylate	S12
4. NMR data	S13
5. HPLC traces	S34

1. General methods

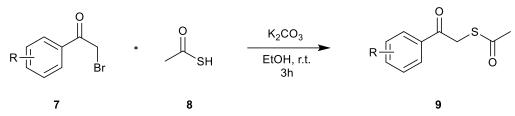
NMR spectra were acquired on a Bruker Ultra Shield 700 instrument, running at 700 MHz for 1H and 176 MHz for 13C, respectively. Chemical shifts (δ) are reported in ppm relative to residual solvent signals (CDCl3: 7.26 ppm for 1H NMR, 77.16 ppm for 13C NMR). Mass spectra were recorded on a Bruker Maxis Impact spectrometer using electrospray (ES+) ionization (referenced to the mass of the charged species). Optical rotations were measured on a Perkin-Elmer 241 polarimeter and [α]D values are given in deg·cm·g⁻¹·dm⁻¹; concentration c is listed in g·(100 mL)⁻¹. Analytical thin layer chromatography (TLC) was performed using pre-coated aluminum-backed plates (Merck Kieselgel 60 F254) and visualized by ultraviolet irradiation. The enantiomeric ratio (er) of the products was determined by chiral stationary phase HPLC (Daicel Chiralpak IA and ID column). Unless otherwise noted, analytical grade solvents and commercially available reagents were used without further purification. For flash chromatography (FC) silica gel (Silica gel 60, 230-400 mesh, Fluka). 2,4-Dienones **2a-d**, **2h**,¹ **2e**, **2f** and **2i**² were prepared applying Heck reaction of the suitable acrylate and vinyl bromide, according to literature precedents. Dienone **2g** was prepared following procedure described by Alexakis.³

¹ R. Kowalczyk, P. J. Boratyński, A. J. Wierzba and J. Bąkowicz, RSC Adv., 2015, 5, 66681.

² D. Duvvuru, J.-F. Betzer, P. Retailleau, G. Frison and A. Marinetti, *Adv. Synth. Catal.*, 2011, **353**, 483.
3 M. Tissot, D. Poggiali, H. Hénon, D. Müller, L. Guénée, M. Mauduit and A. Alexakis, *Chem. Eur. J.*, 2012, **18**, 8731.

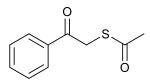
2. Synthetic procedures





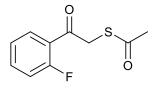
S-Acetyl-2-mercaptoacetophenones **9** were prepared according to the literature data.⁴ NMR spectra of compounds **9a** and **9e** were in accordance with the literature data.⁴

To the solution of thioacetic acid **8** (764 μ L, 10 mmol, 1 equiv.) in ethanol (20 mL, 0.5 M) anhydrous potassium carbonate (1.4 g, 10 mmol, 1 equiv.) was added and the resulting mixture was stirred for 20 min. at room temperature. Subsequently, a solution of corresponding 2-bromoacetophenone **7** (10 mmol 1equiv.) in ethanol (2M) was added dropwise at room temperature and the reaction mixture was stirred at the same temperature until full substrate consumption indicated by TLC analysis (ca. 3h). Formed precipitate was filtered off, and washed with ethanol. Combined washings were concentrated *in vacuo*, dissolved in CHCl₃ (20 mL), washed with distilled water (2×10 mL) and brine (10 mL), dried over Na₂SO₄ and concentrated *in vacuo* to afford the crude product. Pure compound **9** was isolated by the flash chromatography (eluent: Hex:AcOEt 10:1).



9a S-Acetyl-2-mercaptoacetophenone^[4]

Following the general procedure **9a** was isolated as light yellow oil; yield: 90%; ¹H NMR (700 MHz, CDCl₃): δ 7.99 (m, 2H), 7.60 (m, 1H), 7.48 (m, 2H), 4.40 (s, 2H), 2.41 (s, 3H).

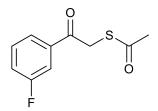


9b S-Acetyl-2-mercapto-2'-fluoroacetophenone

Following the general procedure **9b** was isolated as light yellow oil; yield: 85%; ¹H NMR (700 MHz, CDCl₃): δ 7.87 (td, *J* = 7.6, 1.8 Hz, 1H), 7.55 (dddd, *J* = 8.3, 7.1, 5.1, 1.9 Hz, 1H), 7.24 (ddd, *J* = 7.8, 7.3, 1.1 Hz, 1H), 7.16 (ddd, *J* = 11.1, 8.3, 1.1 Hz, 1H), 4.35 (d, *J* = 2.9 Hz, 2H), 2.38

(s, 3H); ¹³C NMR (176 MHz, CDCl₃) δ 194.3, 191.3 (d, *J* = 4.1 Hz), 162.0 (d, *J* = 254.6 Hz), 135.3 (d, *J* = 9.0 Hz), 131.2 (d, *J* = 2.5 Hz), 124.8 (d, *J* = 3.2 Hz), 124.6 (d, *J* = 13.0 Hz), 116.8 (d, *J* = 23.6 Hz), 40.9 (d, *J* = 9.0 Hz), 30.3.

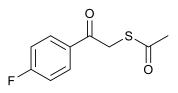
⁴ Y. Z. Adamczewska, J. M. Barker, P. R. Huddlestone and M. L. Wood, Synth. Commun. 1996, 26, 1083.



9c S-Acetyl-2-mercapto-3'-fluoroacetophenone

Following the general procedure **9c** was isolated as light yellow oil; yield: 97% ¹H NMR (700 MHz, CDCl₃) δ 7.77 (ddd, *J* = 7.8, 1.6, 1.0 Hz, 1H), 7.66 (ddd, *J* = 9.2, 2.6, 1.6 Hz, 1H), 7.46 (ddd, *J* = 8.4, 7.8, 5.5 Hz, 1H), 7.29 (tdd, *J* = 8.2, 2.6, 1.0 Hz, 1H), 4.35 (s, 2H), 2.40 (s, 3H); ¹³C NMR (176 MHz, CDCl₃) δ 194.0, 192.1 (d, *J* = 2.0 Hz), 162.9 (d, *J* = 248.6

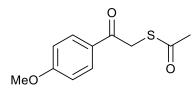
Hz), 137.6 (d, *J* = 6.3 Hz), 130.5 (d, *J* = 7.7 Hz), 124.3 (d, *J* = 2.9 Hz), 120.8 (d, *J* = 21.3 Hz), 115.3 (d, *J* = 22.7 Hz), 36.6, 30.2



9d S-Acetyl-2-mercapto-4'-fluoroacetophenone

Following the general procedure **9d** was isolated as light yellow oil; yield: 96%; ¹H NMR (700 MHz, CDCl₃) δ 8.02 (dd, *J* = 8.9, 5.3 Hz, 2H), 7.14 (dd, *J* = 8.9, 8.3 Hz, 2H), 4.35 (s, 2H), 2.40 (s, 3H); ¹³C NMR (176 MHz, CDCl₃) δ 193.0 (d, *J* = 410.3 Hz), 166.8, 165.4,

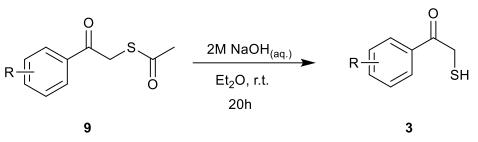
132.0 (d, J = 3.1 Hz), 131.3 (d, J = 9.5 Hz), 116.0 (d, J = 22.2 Hz), 36.4, 30.2.



9e S-Acetyl-2-mercapto-4'-methoxyacetophenone^[4]

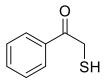
Following the general procedure **9e** was isolated as light yellow oil; yield: 51%.

2.2 Procedure for synthesis of 2-mercapto-1-phenylethanone 3

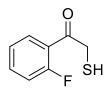


2-Mercaptoacetophenones **3** were prepared according to the literature procedure.⁵ NMR spectra of **3a** were in accordance with literature data.⁵

To the solution of *S*-acetyl-2-mercapto-1-phenylethanone **9** (1.75 g, 9 mmol, 1 equiv.) in Et₂O (9 mL, 1M) 2M aqueous solution of sodium hydroxide (9 mL, 2 equiv.) were added and the resulting biphasic mixture was stirred vigorously at room temperature for 20 h. Next, the aqueous phase was separated, acidified with 1M aqueous solution of HCl and extracted with CHCl₃ (2×15 mL). Organic phase was washed with distilled water (2×10 mL), dried over Na₂SO₄ and concentrated *in vacuo* to afford the crude product. Pure compound was isolated by the flash chromatography (eluent: Hex:AcOEt 10:1).



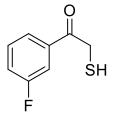
3a 2-Mercapto-1-phenylethanone⁵ Colorless oil; yield: 55%.



3b 2-Mercapto-1-(2'-fluorophenyl)ethanone

Following the general procedure **3b** was isolated as light yellow oil; yield: 49%; ¹H NMR (700 MHz, CDCl₃) δ 7.97 – 7.88 (m, 1H), 7.55 (dddd, *J* = 8.2, 7.1, 5.1, 1.9 Hz, 1H), 7.29 – 7.20 (m, 1H), 7.15 (ddd, *J* = 11.3, 8.3, 1.0 Hz, 1H), 3.93 (ddd, *J* = 7.7, 2.4, 0.6 Hz, 2H), 2.04 (td, *J* = 7.7, 1.0 Hz, 1H); ¹³C NMR (176 MHz,

CDCl₃) δ 193.0 (d, J = 4.3 Hz), 161.8 (d, J = 254.3 Hz), 135.2 (d, J = 9.0 Hz), 131.4 (d, J = 2.5 Hz), 124.8 (d, J = 3.5 Hz), 123.7 (d, J = 13.0 Hz), 116.7 (d, J = 23.8 Hz), 35.4 (d, J = 8.9 Hz).

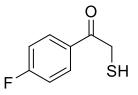


3c 2-Mercapto-1-(3'-fluorophenyl)ethanone

Following the general procedure **3c** was isolated as light yellow solid; yield: 58%; ¹H NMR (700 MHz, CDCl₃) δ 7.72 (ddd, *J* = 7.8, 1.6, 1.0 Hz, 1H), 7.64 (dddd, *J* = 9.3, 2.7, 1.6, 0.4 Hz, 1H), 7.46 (dddd, *J* = 8.2, 7.8, 5.5, 0.4 Hz, 1H), 7.29 (tdd, *J* = 8.2, 2.6, 1.0 Hz, 1H), 3.92 (d, *J* = 7.5 Hz, 2H), 2.11 (t, *J* = 7.4 Hz, 1H); ¹³C NMR (176 MHz, CDCl₃) δ 193.6, 162.9 (d, *J* = 248.6 Hz), 137.1 (d, *J* = 6.4 Hz), 130.6 (d, *J* = 7.6 Hz), 124.3 (d, *J* = 3.1 Hz), 120.7 (d, *J* = 21.2 Hz), 115.3

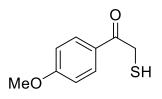
(d, J = 22.4 Hz), 31.2.

⁵ C.-C. Han, R. Balakumar, Tetrahedron Lett. 2006, 47, 8255.



3d Mercapto-1-(4'-fluorophenyl)ethanone

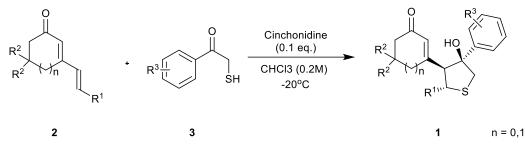
Following the general procedure 3d was isolated as light yellow oil; yield: 52%; ¹H NMR (700 MHz, CDCl₃) δ 7.98 (dd, J = 8.9, 5.3 Hz, 2H), 7.15 (dd, J = 8.9, 8.3 Hz, 2H), 3.92 (d, J = 7.4 Hz, 2H), 2.11 (t, J = 7.4 Hz, 1H); 13 C NMR (176 MHz, CDCl₃) δ 193.29, 166.02 (d, *J* = 256.0 Hz), 131.46 (d, J = 3.0 Hz), 116.02 (d, J = 21.9 Hz), 31.02.



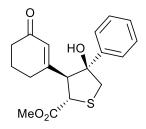
3e Mercapto-1-(4'-methoxyphenyl)ethanone

Following the general procedure **3e** was isolated as yellow solid; yield: 50%; ¹H NMR (700 MHz, CDCl₃) δ 7.97 – 7.88 (m, 2H), 6.94 (d, J = 8.9 Hz, 2H), 3.90 (d, J = 7.3 Hz, 2H), 3.87 (s, 3H); ¹³C NMR (176 MHz, CDCl₃) δ 193.4, 163.9, 130.9, 128.0, 114.0, 55.6, 30.8.

2.3 Brønsted-base-catalyzed remote cascade – general procedure

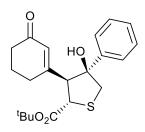


To the solution of corresponding 2-mercapto-1-phenylethanone **3** (0.15 mmol, 1.5 equiv.) in CHCl₃ (0.5 mL, 0.3 M) placed in a standard glass vial cinchonidine **4c** (0.01 mmol, 0.1 equiv.) was added in one portion at room temperature and the resulting mixture was stirred for 10 minutes at the same temperature. Subsequently, the reaction mixture was cooled down to -20 °C, the corresponding 2,4-dienone **2** (0.1 mmol, 1 equiv.) was added in one portion and the reaction mixture was stirred at -20 °C. Upon reaction completion (as indicated by ¹H NMR) the reaction mixture was directly subjected to the flash chromatography on silica (eluent: Hex:AcOEt 4:1) to afford pure **1**.



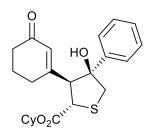
1a (2*S*,3*S*,4*S*)-Methyl **4**-hydroxy-**3**-(**3**-oxocyclohex-**1**-en-**1**-yl)-**4**phenyltetrahydrothiophene-**2**-carboxylate. Following the general procedure **1a** (> 20:1 dr) was obtained in 90% yield as white foam. ¹H NMR (700 MHz, CDCl₃) δ 7.51 – 7.47 (m, 2H), 7.36 (td, *J* = 8.1, 7.7, 1.6 Hz, 2H), 7.31 – 7.27 (m, 1H), 5.94 (dt, *J* = 3.6, 1.5 Hz, 1H), 4.34 (dd, *J* = 10.6, 1.7 Hz, 1H), 3.76 (d, *J* = 12.0 Hz, 1H), 3.74 (d, *J* = 1.6 Hz, 3H), 3.47 (dd, *J* = 10.5, 2.6 Hz, 1H), 3.14 (dt, *J* = 4.3, 1.4 Hz, 1H), 3.06 (d, *J* = 11.9

Hz, 1H), 2.26 – 2.12 (m, 3H), 1.76 – 1.68 (m, 1H), 1.67 – 1.57 (m, 2H); ¹³C NMR (176 MHz, CDCl₃) δ 199.5, 172.2, 160.3, 140.5, 129.0, 128.6, 128.0, 125.0, 85.3, 62.6, 52.9, 48.7, 47.1, 37.3, 30.6, 22.5 HRMS calculated for [C₁₈H₂₀O₄S+H]⁺: 333.1156; found: 333.1148. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} = 16.0 min, τ_{minor} = 36.2 min (97:3 er). [α_D^{20}] = - 31.3 (c 1.0, CHCl₃).



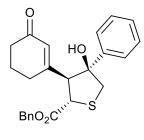
1b (2*S*,3*S*,4*S*)-*tert*-Butyl **4**-hydroxy-**3**-(**3**-oxocyclohex-**1**-en-**1**-yl)-**4**phenyltetrahydrothiophene-**2**-carboxylate. Following the general procedure **1b** (> 20:1 dr) was obtained in 98% yield as a white solid. ¹H NMR (700 MHz, CDCl₃) δ 7.51 – 7.47 (m, 2H), 7.35 (t, *J* = 7.8 Hz, 2H), 7.30 – 7.26 (m, 1H), 5.93 (d, *J* = 1.9 Hz, 1H), 4.22 (d, *J* = 10.3 Hz, 1H), 3.74 (d, *J* = 11.8 Hz, 1H), 3.42 (d, *J* = 10.4 Hz, 1H), 3.03 (d, *J* = 11.9 Hz, 1H), 2.94 (s, 1H), 2.21 (dddd, *J* = 26.1, 13.0, 8.1, 5.2 Hz, 3H), 1.74 (td, *J*

= 7.9, 4.9 Hz, 1H), 1.71 – 1.60 (m, 2H), 1.45 (s, 9H); ¹³C NMR (176 MHz, CDCl₃) δ 199.3, 170.7, 160.5, 140.5, 128.9, 128.6, 128.0, 125.0, 85.6, 82.5, 62.6, 50.0, 47.1, 37.3, 30.5, 28.0, 22.5. HRMS calculated for [C₂₁H₂₆O₄S+H]⁺: 375.1625; found: 375.1636. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} = 8.5 min, τ_{minor} = 20.2 min (96:4 er). [α_D^{20}] = - 11.5 (c 0.5, CHCl₃).



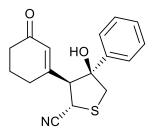
1c (2*S*,3*S*,4*S*)-Cyclohexyl 4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)-4phenyltetrahydrothiophene-2-carboxylate. Following the general procedure 1c (> 20:1 dr) was obtained in 96% yield as a white solid. ¹H NMR (700 MHz, CDCl₃) δ 7.52 – 7.47 (m, 2H), 7.36 (dd, *J* = 8.5, 7.1 Hz, 2H), 7.31 – 7.27 (m, 1H), 5.93 (d, *J* = 1.6 Hz, 1H), 4.79 (ddt, *J* = 12.9, 8.7, 3.9 Hz, 1H), 4.30 (d, *J* = 10.4 Hz, 1H), 3.76 (d, *J* = 11.8 Hz, 1H), 3.46 (d, *J* = 10.4 Hz, 1H), 3.05 (d, *J* = 11.9 Hz, 1H), 2.96 (s, 1H), 2.28 – 2.12 (m,

3H), 1.88 – 1.77 (m, 2H), 1.76 – 1.67 (m, 3H), 1.67 – 1.59 (m, 1H), 1.52 (dt, J = 9.8, 3.6 Hz, 1H), 1.48 – 1.41 (m, 2H), 1.39 – 1.32 (m, 2H), 1.27 (dd, J = 9.9, 3.4 Hz, 1H); ¹³C NMR (176 MHz, CDCl₃) δ 199.3, 171.1, 160.3, 140.5, 129.0, 128.6, 128.0, 125.0, 85.5, 74.4, 62.6, 49.3, 47.1, 37.3, 31.4, 31.3, 30.6, 25.3, 23.6, 23.5, 22.5. HRMS calculated for [C₂₃H₂₈O₄S+H]⁺: 401.1782; found: 401.1789. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} = 11.8 min, τ_{minor} = 29.8 min (95:5 er). [α_D^{20}] = - 24.4 (c 1.0, CHCl₃).



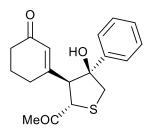
1d (2*S*,3*S*,4*S*)-Benzyl 4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)-4phenyltetrahydrothiophene-2-carboxylate. Following the general procedure 1d (> 20:1 dr) was obtained in 33% yield as a beige solid. ¹H NMR (700 MHz, CDCl₃) δ 7.50 – 7.47 (m, 2H), 7.38 – 7.32 (m, 7H), 7.32 – 7.28 (m, 1H), 5.91 (d, *J* = 1.7 Hz, 1H), 5.26 – 5.11 (m, 2H), 4.37 (d, *J* = 10.3 Hz, 1H), 3.77 (d, *J* = 11.9 Hz, 1H), 3.48 (d, *J* = 10.3 Hz, 1H), 3.05 (d, *J* = 11.9 Hz, 1H), 2.75 (s, 1H), 2.22 – 2.10 (m, 3H), 1.71 – 1.53 (m, 4H);

¹³C NMR (176 MHz, CDCl₃) δ 199.1, 171.5, 159.8, 140.2, 135.2, 129.1, 128.7, 128.6, 128.5, 128.2, 128.1, 125.0, 85.5, 67.6, 62.7, 48.9, 47.2, 37.3, 30.4, 22.4. HRMS calculated for [C₂₄H₂₄O₄S+H]⁺: 409.1469; found: 409.1470. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹τ_{major} = 20.4 min, t_{minor} = 40.0 min (95:5 er). [α_D^{20}] = - 19.9 (c 1.0, CHCl₃).



1e (2*S*,3*S*,4*S*)-4-Hydroxy-3-(3-oxocyclohex-1-en-1-yl)-4phenyltetrahydrothiophene-2-carbonitrile. Following the general procedure 1e (> 20:1 dr) was obtained in 87% yield as a yellow oil. ¹H NMR (700 MHz, Acetonitrile-d₃) δ 7.53 – 7.48 (m, 2H), 7.40 – 7.34 (m, 2H), 7.32 – 7.28 (m, 1H), 5.73 (d, *J* = 1.7 Hz, 1H), 4.60 (d, *J* = 11.4 Hz, 1H), 3.98 (s, 1H), 3.86 (s, 1H), 3.42 (d, *J* = 11.4 Hz, 1H), 3.10 (d, *J* = 11.7 Hz, 1H), 2.27 (dddd, *J* = 18.2, 7.3, 4.5, 1.7 Hz, 1H), 2.13 (dd, *J* = 7.4, 6.0

Hz, 2H), 1.82 – 1.76 (m, 1H), 1.72 (ddd, J = 13.6, 6.2, 4.4 Hz, 1H), 1.69 – 1.63 (m, 1H); ¹³C NMR (176 MHz, Acetonitrile₃₃) δ 199.3, 158.6, 141.7, 130.8, 129.4, 128.8, 126.4, 120.1, 118.3, 85.6, 65.2, 47.2, 37.9, 34.7, 30.0, 23.3. HRMS calculated for $[C_{17}H_{17}NO_2S+H]^+$: 300.1053; found: 300.1048. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} =12.3 min, τ_{minor} = 21.5 min (79:21 er). $[\alpha_D^{20}]$ = 4.3 (c 0.5, CHCl₃).



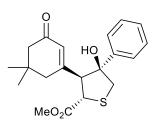
If 3-((2*S*,3*S*,4*S*)-2-Acetyl-4-hydroxy-4-phenyltetrahydrothiophen-3yl)cyclohex-2-enone. Following the general procedure If (5:1 dr) was obtained in 84% yield as a yellow oil. ¹H NMR (700 MHz, Chloroformd) δ 7.55 – 7.52 (m, 2H), 7.40 (t, *J* = 7.8 Hz, 2H), 7.33 – 7.30 (m, 1H), 6.09 (t, *J* = 1.1 Hz, 1H), 4.68 (d, *J* = 10.3 Hz, 1H), 4.17 (d, *J* = 1.6 Hz, 1H), 3.67 (d, *J* = 10.4 Hz, 1H), 3.50 (dd, *J* = 11.8, 1.2 Hz, 1H), 3.11 (d, *J* = 11.7 Hz, 1H), 2.67 (dt, *J* = 17.8, 6.0 Hz, 1H), 2.42 (ddd, *J* = 13.4, 7.4, 5.7 Hz,

2H), 2.07 (tt, J = 6.0, 1.8 Hz, 2H), 1.79 (s, 3H), 1.75 – 1.65 (m, 1H); ¹³C NMR (176 MHz, Chloroform-*d*) δ 208.3, 199.1, 161.4, 141.0, 128.9, 128.2, 127.2, 124.8, 86.4, 66.5, 54.2, 47.5, 37.6, 32.7, 26.9, 23.0. HRMS calculated for [C₁₈H₂₀O₃S+H]⁺: 317.1206; found: 317.1197. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{minor} = 32.2 min, τ_{major} = 55.4 min (29:71 er). [α_D^{20}] = 3.3 (c 0.5, CHCl₃).



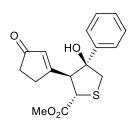
1g 3-((2*S***,3***S***,4***S***)-4-Hydroxy-2,4-diphenyltetrahydrothiophen-3yl)cyclohex-2-enone. Following the general procedure 1g** (> 20:1 dr) was obtained in 35% yield as a colorless oli.¹H NMR (700 MHz, CDCl₃) δ 7.55 – 7.51 (m, 2H), 7.44 (dd, *J* = 8.1, 1.3 Hz, 2H), 7.38 (dd, *J* = 8.4, 7.1 Hz, 2H), 7.34 – 7.29 (m, 2H), 7.29 – 7.24 (m, 2H), 5.89 – 5.87 (m, 1H), 4.91 (d, *J* = 11.3 Hz, 1H), 3.92 (d, *J* = 12.0 Hz, 1H), 3.20 (d, *J* = 11.3 Hz, 1H), 3.16 (d, *J* = 12.0 Hz, 1H), 3.03 (d, *J* = 1.8 Hz, 1H), 2.10 (dd, *J* =

7.4, 6.0 Hz, 2H), 1.98 (dddd, J = 18.3, 7.7, 4.6, 1.6 Hz, 1H), 1.74 – 1.68 (m, 1H), 1.64 – 1.58 (m, 1H), 1.58 – 1.51 (m, 1H); ¹³C NMR (176 MHz, Chloroform-*d*) δ 199.3, 160.5, 141.4, 139.2, 129.7, 128.7, 128.6, 128.0, 127.9, 127.9, 124.9, 85.7, 69.4, 53.3, 47.3, 37.2, 30.6, 22.4. HRMS calculated for [C₂₂H₂₂O₂S+H]⁺: 351.1414; found: 351.1410. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} =10.7 min, τ_{minor} = 14.8 min (84:16 er). [α_D^{20}] = 24.0 (c 1.0, CHCl₃).



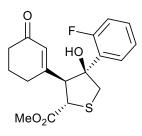
1h (2*S*,3*S*,4*S*)-Methyl 3-(5,5-dimethyl-3-oxocyclohex-1-en-1-yl)-4hydroxy-4-phenyltetrahydrothiophene-2-carboxylate. Following the general procedure **1h** (> 20:1 dr) was obtained in 98% yield as a yellow oil. ¹H NMR (700 MHz, CDCl₃) δ 7.52 – 7.47 (m, 2H), 7.35 (t, *J* = 7.4 Hz, 2H), 7.30 – 7.25 (m, 1H), 5.98 (s, 1H), 4.34 (d, *J* = 10.5 Hz, 1H), 3.74 (d, *J* = 1.0 Hz, 3H), 3.72 (d, *J* = 11.8 Hz, 1H), 3.47 (d, *J* = 10.6 Hz, 1H), 3.09 (s, 1H), 3.06 (d, *J* = 11.8 Hz, 1H), 2.21 (d, *J* = 18.4 Hz, 1H),

2.10 – 1.98 (m, 2H), 1.64 (d, *J* = 18.3 Hz, 1H), 0.82 (s, 3H), 0.61 (s, 3H); ¹³C NMR (176 MHz, CDCl₃) δ 199.6, 172.2, 157.9, 140.3, 128.6, 128.3, 128.0, 125.2, 85.4, 62.1, 52.9, 50.8, 49.0, 47.6, 44.6, 33.3, 28.4, 27.5. HRMS calculated for [C₂₀H₂₄O₄S+H]⁺: 361.1469; found: 361.1473. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹τ_{major} = 15.7 min, τ_{minor} = 20.1 min (96:4 er). [α_D^{20}] = -18.4 (c 1.0, CHCl₃).



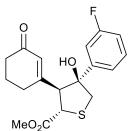
1i (2*S*,3*S*,4*S*)-Methyl **4**-hydroxy-3-(3-oxocyclopent-1-en-1-yl)-4phenyltetrahydrothiophene-2-carboxylate. Following the general procedure **1i** (> 20:1 dr) was obtained in 90% yield as a yellow oil. ¹H NMR (700 MHz, CDCl₃) δ 7.52 (dd, *J* = 7.4, 1.3 Hz, 2H), 7.39 – 7.34 (m, 2H), 7.32 – 7.28 (m, 1H), 6.00 – 5.96 (m, 1H), 4.35 (d, *J* = 10.1 Hz, 1H), 3.85 (d, *J* = 10.2 Hz, 1H), 3.77 (d, *J* = 1.2 Hz, 3H), 3.72 (d, *J* = 11.9 Hz, 1H), 3.19 (s, 1H), 3.07 (d, *J* = 11.9 Hz, 1H), 2.40 (dd, *J* = 18.9, 7.1 Hz, 1H), 2.23 – 2.07 (m,

2H), 2.06 – 1.99 (m, 1H); ¹³C NMR (176 MHz, CDCl₃) δ 209.3, 176.1, 172.2, 140.3, 133.0, 128.7, 128.1, 125.0, 85.5, 58.7, 53.1, 48.9, 47.6, 35.0, 31.9. HRMS calculated for [C₁₇H₁₈O₄S+H]⁺: 319.0999; found: 319.0998. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹τ_{major} = 13.8 min, τ_{minor} = 26.9 min (92:8 er). [α_D^{20}] = - 14.8 (c 1.0, CHCl₃).



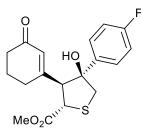
1j (25,35,45)-Methyl 4-(2-fluorophenyl)-4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)tetrahydrothiophene-2-carboxylate. Following the general procedure **1j** (> 20:1 dr) was obtained in 95% yield as a colorless solid. ¹H NMR (700 MHz, CDCl₃) δ 7.63 (td, *J* = 8.0, 1.8 Hz, 1H), 7.33 – 7.28 (m, 1H), 7.15 (td, *J* = 7.6, 1.2 Hz, 1H), 7.06 (ddd, *J* = 12.4, 8.1, 1.2 Hz, 1H), 5.96 (d, *J* = 1.4 Hz, 1H), 4.38 (d, *J* = 10.7 Hz, 1H), 3.99 (d, *J* = 11.6 Hz, 1H), 3.85 (d, *J* = 10.7 Hz, 1H), 3.75 (s, 3H), 3.23 (s, 1H), 2.96 (dd, *J* = 11.6, 1.3

Hz, 1H), 2.35 – 2.28 (m, 1H), 2.25 – 2.11 (m, 2H), 1.75 (tddd, J = 9.3, 7.3, 4.4, 2.3 Hz, 2H), 1.66 – 1.58 (m, 1H); ¹³C NMR (176 MHz, CDCl₃) δ 199.5, 171.8, 160.4, 158.5, 130.2 (d, J = 8.8 Hz), 129.0, 128.4 (d, J = 3.2 Hz), 127.3 (d, J = 10.9 Hz), 124.6 (d, J = 3.3 Hz), 116.2 (d, J = 23.5 Hz), 83.5 (d, J = 5.1 Hz), 59.3 (d, J = 5.5 Hz), 52.9, 48.3, 44.4 (d, J = 5.6 Hz), 37.3, 30.2, 22.5. HRMS calculated for [C₁₈H₁₉FO₄S+H]⁺: 351.1061; found: 351.1055. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ $\tau_{major} = 11.3$ min, $t_{minor} = 14.9$ min (92:8 er). [α_D^{20}] = - 10.5 (c 0.5, CHCl₃).



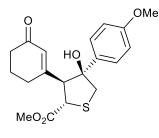
1k (2*S*,3*S*,4*S*)-Methyl 4-(3-fluorophenyl)-4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)tetrahydrothiophene-2-carboxylate. Following the general procedure 1k (> 20:1 dr) was obtained in 98% yield as a light yellow oil. ¹H NMR (700 MHz, CDCl₃) δ 7.34 (td, *J* = 8.2, 5.9 Hz, 1H), 7.26 (ddt, *J* = 5.4, 4.6, 1.9 Hz, 2H), 7.00 (tdd, *J* = 8.3, 2.5, 1.1 Hz, 1H), 5.94 (d, *J* = 1.7 Hz, 1H), 4.31 (d, *J* = 10.2 Hz, 1H), 3.76 (s, 3H), 3.72 (d, *J* = 11.9 Hz, 1H), 3.47 (d, *J* = 10.2 Hz, 1H), 3.12 (s, 1H), 3.06 (d, *J* = 11.9 Hz, 1H), 2.30 – 2.18 (m,

3H), 1.83 – 1.62 (m, 3H); ¹³C NMR (176 MHz, CDCl₃) δ 199.3, 172.0, 162.3, 159.6, 143.3 (d, J = 7.0 Hz), 130.2 (d, J = 8.0 Hz), 129.2, 120.6 (d, J = 3.0 Hz), 115.0 (d, J = 21.0 Hz), 112.7 (d, J = 23.3 Hz), 85.0, 62.5, 53.0, 48.6, 47.1, 37.3, 30.4, 22.5. HRMS calculated for [C₁₈H₁₉FO₄S+H]⁺: 351.1061; found: 351.1051. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ $\tau_{major} = 11.9 \text{ min}$, $t_{minor} = 21.4 \text{ min}$ (89:11 er). [α_D^{20}] = -12.7 (c 0.5, CHCl₃).



1I (2*S*,3*S*,4*S*)-Methyl 4-(4-fluorophenyl)-4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)tetrahydrothiophene-2-carboxylate. Following the general procedure 1I (> 20:1 dr) was obtained in 87% yield as a light yellow oil. ¹H NMR (700 MHz, CDCl₃) δ 7.52 – 7.45 (m, 2H), 7.09 – 7.03 (m, 2H), 5.93 (q, *J* = 2.8, 2.2 Hz, 1H), 4.29 (d, *J* = 10.2 Hz, 1H), 3.76 (s, 3H), 3.73 (d, *J* = 11.9 Hz, 1H), 3.46 (d, *J* = 10.2 Hz, 1H), 3.04 (d, *J* = 11.9 Hz, 1H), 3.00 – 2.96 (m, 1H), 2.27 – 2.18 (m, 3H), 1.82 – 1.62 (m, 3H); ¹³C NMR

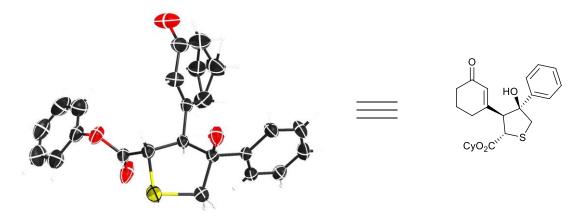
(176 MHz, CDCl₃) δ 199.20, 172.08, 163.04, 161.63, 159.70, 136.11 (d, *J* = 3.2 Hz), 129.19, 126.93 (d, *J* = 8.1 Hz), 115.52 (d, *J* = 21.3 Hz), 85.10, 62.40, 53.45, 52.99, 48.48, 47.22, 37.26, 30.55, 22.49. HRMS calculated for [C₁₈H₁₉FO₄S+H]⁺: 351.1061; found: 351.1060. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} = 12.5 min, τ_{minor} = 28.1 min (90:10 er). [α_D^{20}] = - 8.6 (c 0.5, CHCl₃).



1m (2*S*,3*S*,4*S*)-Methyl 4-hydroxy-4-(4-methoxyphenyl)-3-(3-oxocyclohex-1-en-1-yl)tetrahydrothiophene-2-carboxylate. Following the general procedure 1m (> 20:1 dr) was obtained in 85% yield as a colorless solid. ¹H NMR (700 MHz, CDCl₃) δ 7.42 – 7.38 (m, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 5.94 (d, *J* = 1.6 Hz, 1H), 4.31 (d, *J* = 10.4 Hz, 1H), 3.80 (s, 3H), 3.75 (d, *J* = 0.6 Hz, 3H), 3.73 (d, *J* = 11.8 Hz, 1H), 3.45 (d, *J* = 10.4 Hz, 1H), 3.02 (d, *J* = 11.9 Hz, 1H), 2.82 (d, *J* = 5.2 Hz,

1H), 2.26 – 2.18 (m, 3H), 1.80 – 1.73 (m, 1H), 1.72 – 1.63 (m, 2H); ¹³C NMR (176 MHz, CDCl₃) δ 199.3, 172.2, 160.2, 159.2, 132.3, 129.0, 126.2, 113.9, 85.2, 62.3, 55.3, 52.9, 48.5, 47.1, 37.3, 30.6, 22.5. HRMS calculated for [C₁₉H₂₂O₅S+H]⁺: 363.1261; found: 363.1259. The er was determined by HPLC using a Chiralpak ID column [hexane/*i*-PrOH (80:20)]; flow rate 1mL×min⁻¹ τ_{major} = 23.6 min, τ_{minor} = 37.1 min (72:28 er). [α_D^{20}] = -4.9 (c 1.0, CHCl₃).

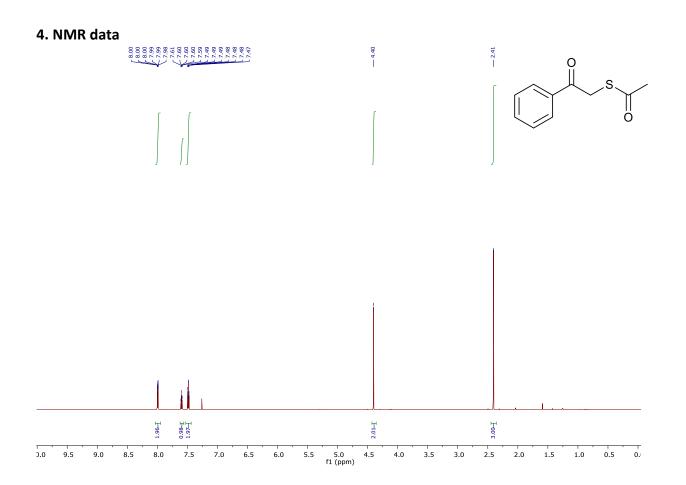
3. Crystal and X-ray data for (2*S*,3*S*,4*S*)-cyclohexyl 4-hydroxy-3-(3-oxocyclohex-1-en-1-yl)-4-phenyltetrahydrothiophene-2-carboxylate

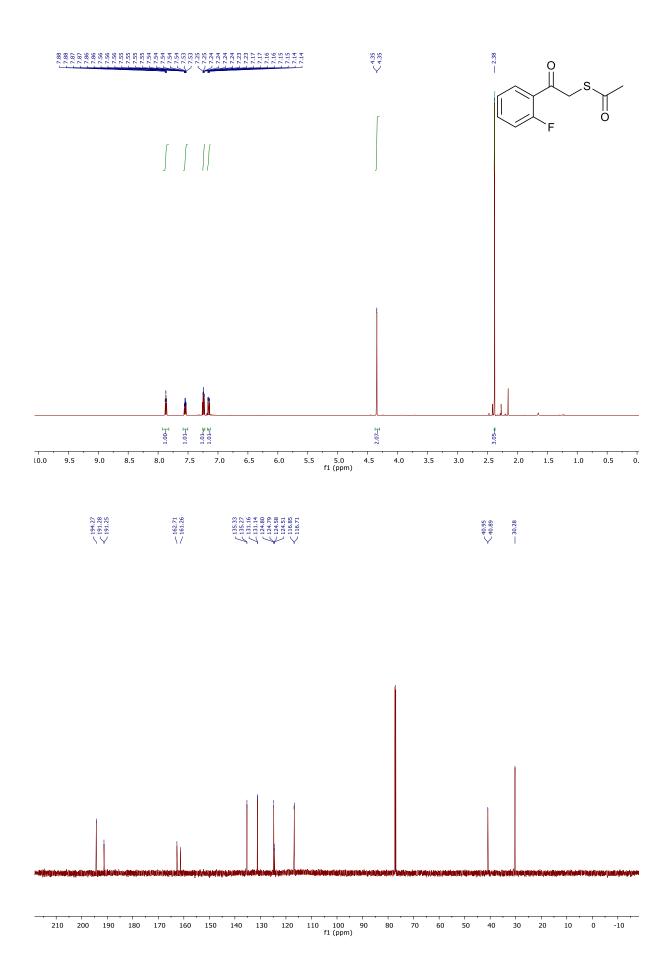


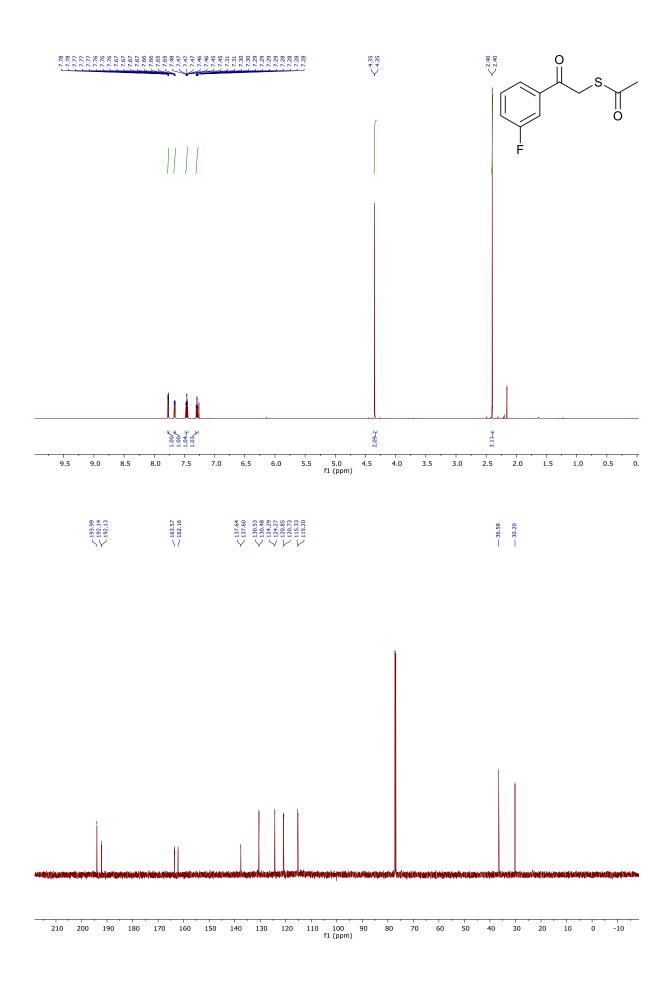
Formula $C_{23}H_{22}O_4S_1$, triclimic, space group P1, Z = 1, cell constants a = 5.6715(2) Å, b = 8.7531(2) Å, c = 10.1807(2) Å, α = 84.0710(10)°, β = 82.0360(10)°, γ = 83.6930(10)°, V = 495.503(17) Å³. The data was collected on a Bruker Smart Apex2 diffractometer at 100 K using Incoatec IµS Cu-K α (λ =1.54178 Å) as a source of radiation. The integration of the data yielded a total of 5975 reflections to a θ angle of 84.6°, of which 3276 were independent (Rint =1. 5%,) and 3241 were greater than 2σ (F²). The final anisotropic full-matrix least-squares refinement on F² with 265 variables converged at R₁ = 3.64%, for the observed data and wR₂ = 9.17% for all data. The hydrogen atoms were placed in calculated positions and refined isotropically by using a riding model. The goodness-of-fit was 1.119.

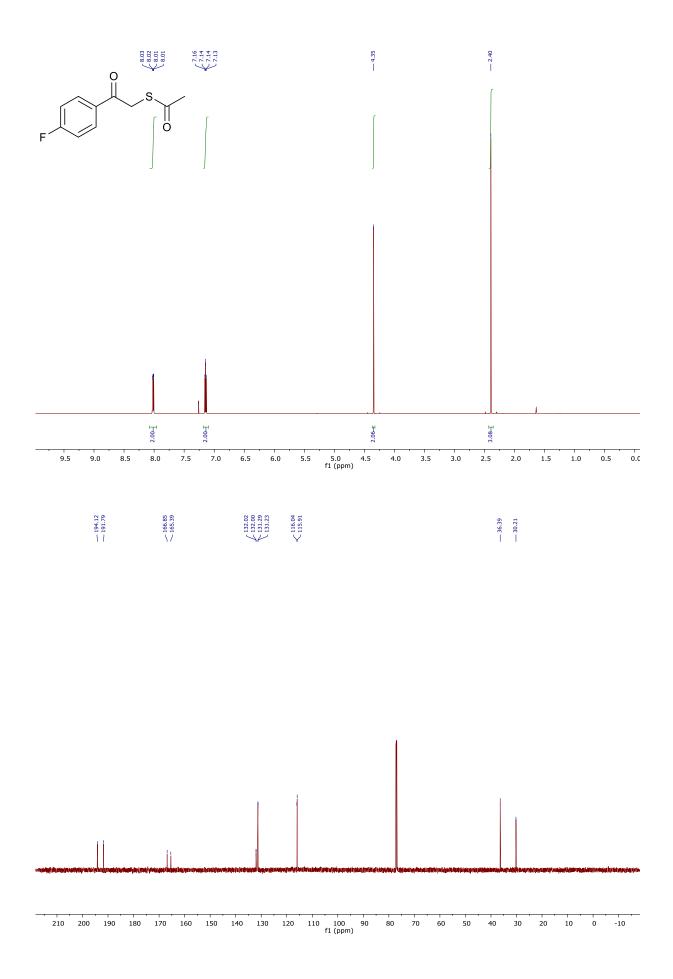
The absolute configuration of X was determined from anomalous scattering, by calculating the Flack parameter: -0.004(9) from 1299 selected quotients (Parsons' method).

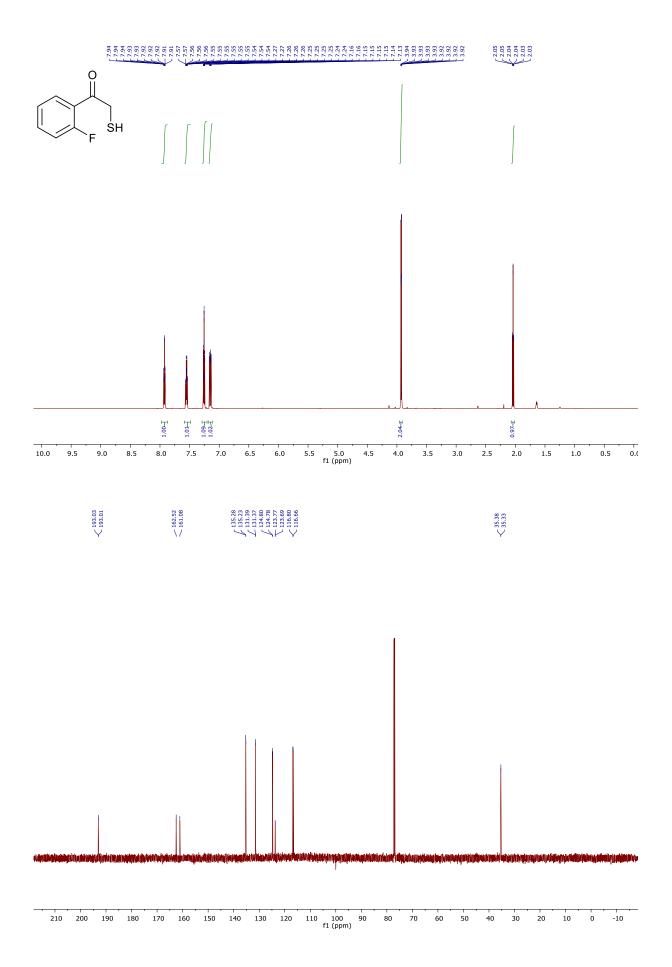
CCDC 1548930 contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from The Cambridge Crystallographic Data Centre via www.ccdc.cam.ac.uk/data request/cif.

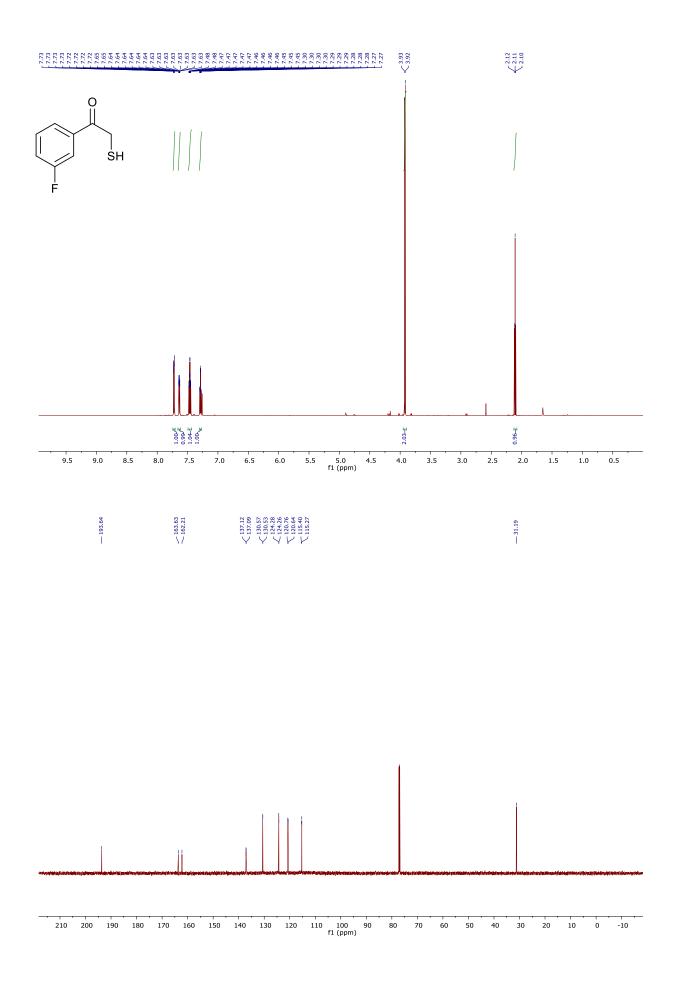


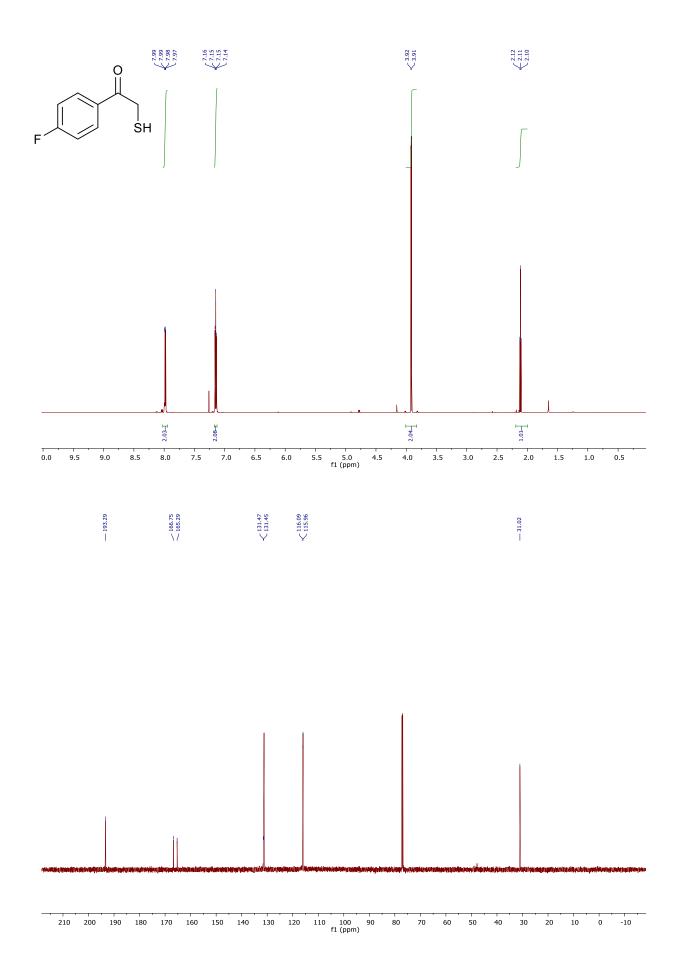


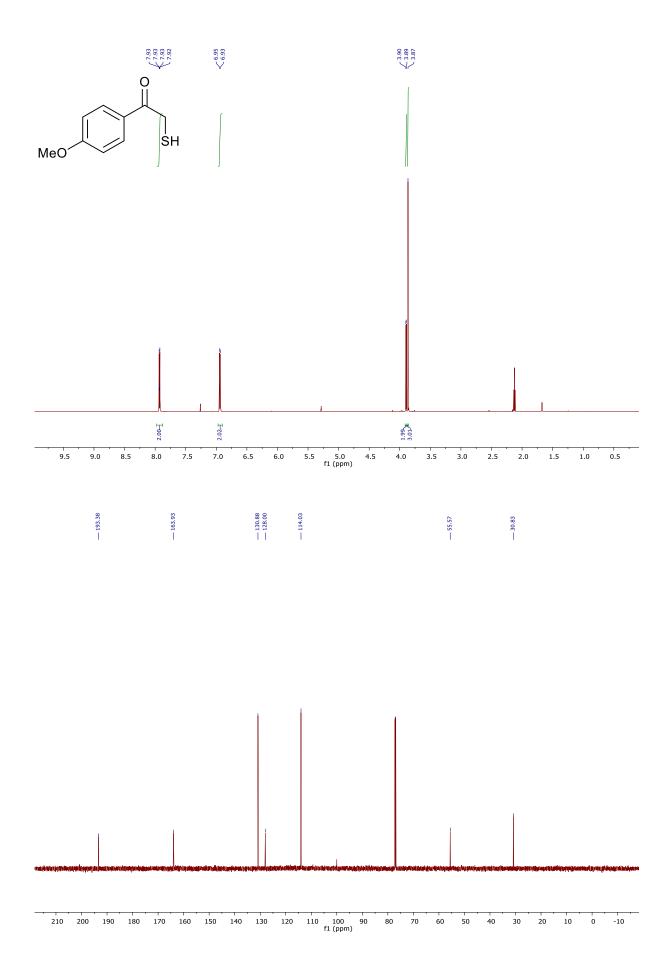


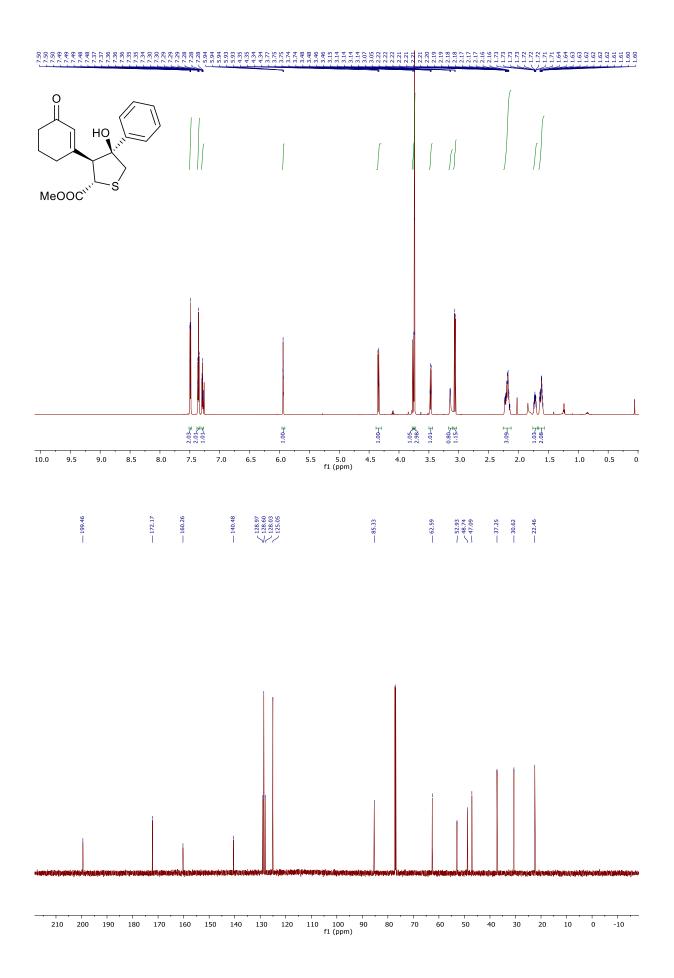




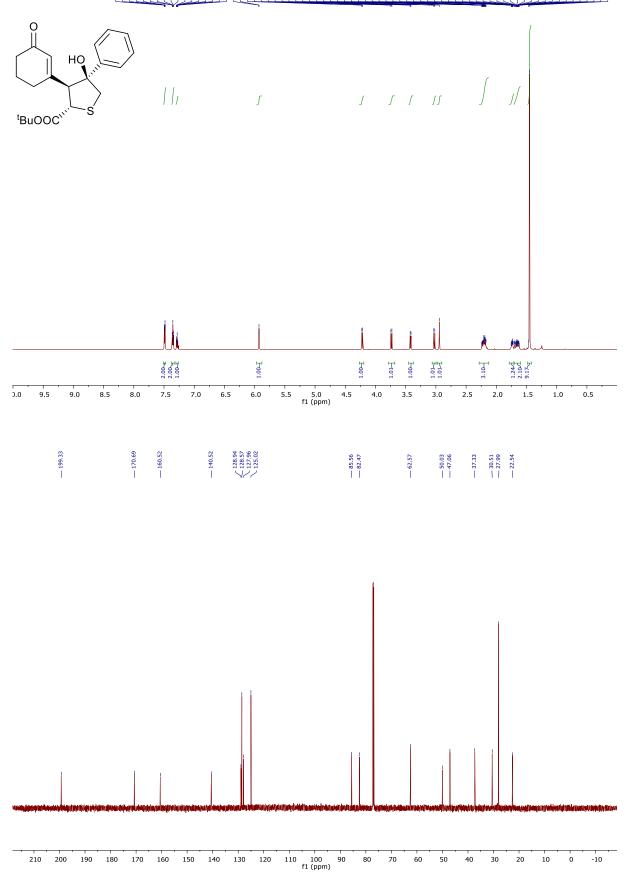


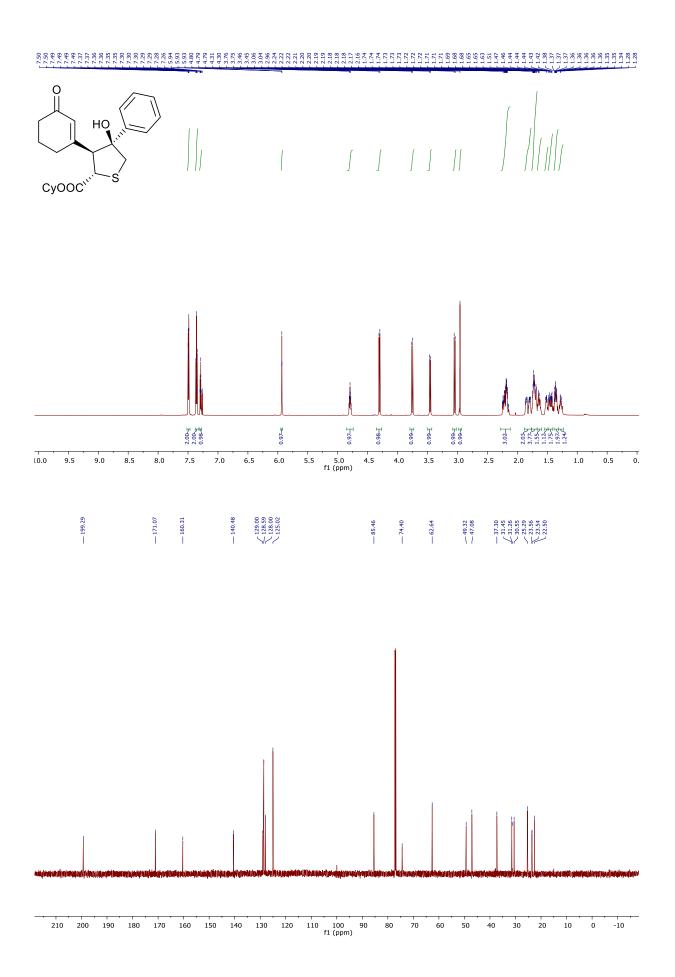


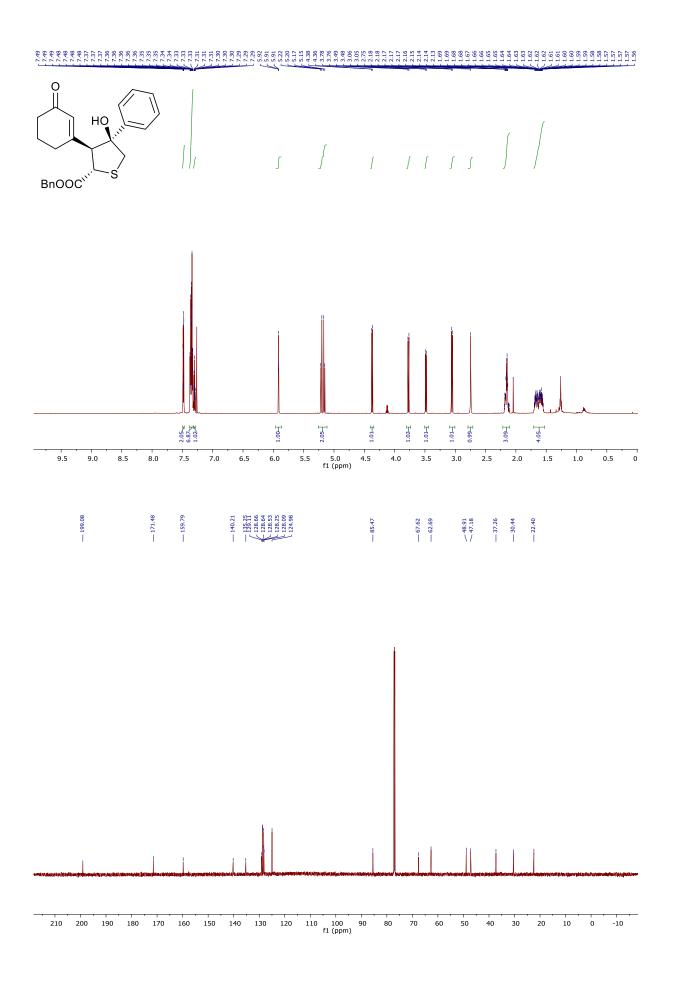


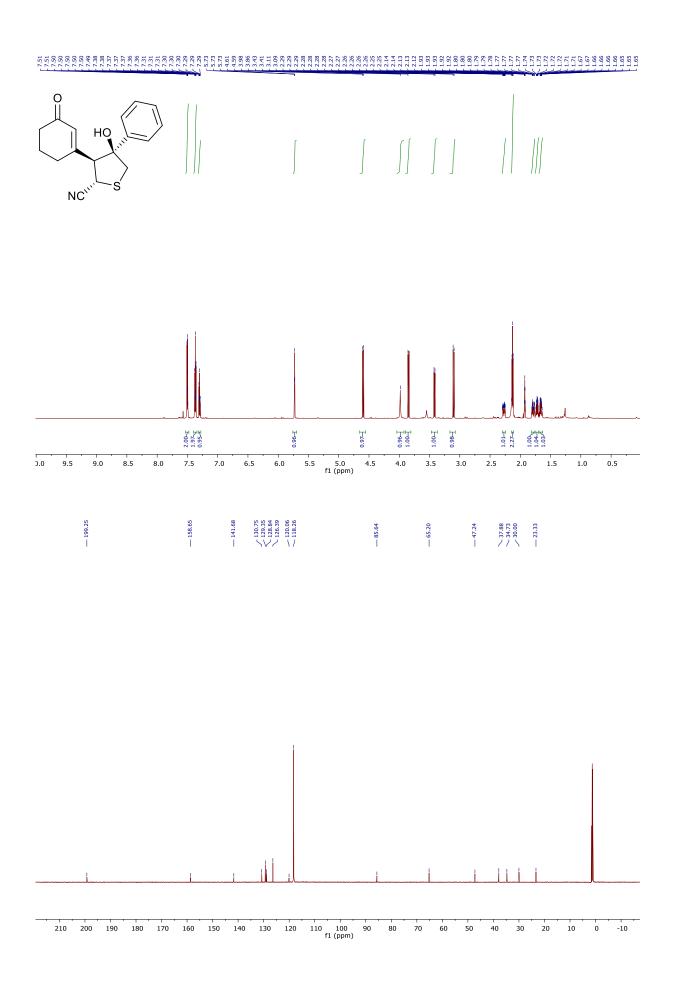


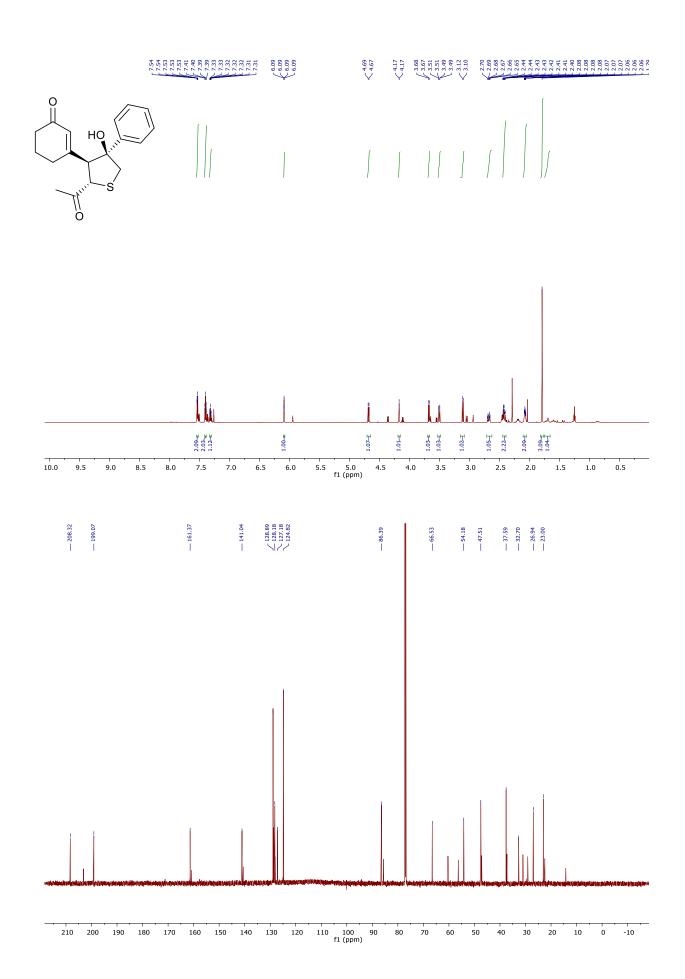
77.50 77.449 77.748 77.748 77.728 77.728 77.728 77.728 77.728 77.728 77.728 77.729 77.728 77.729 77.

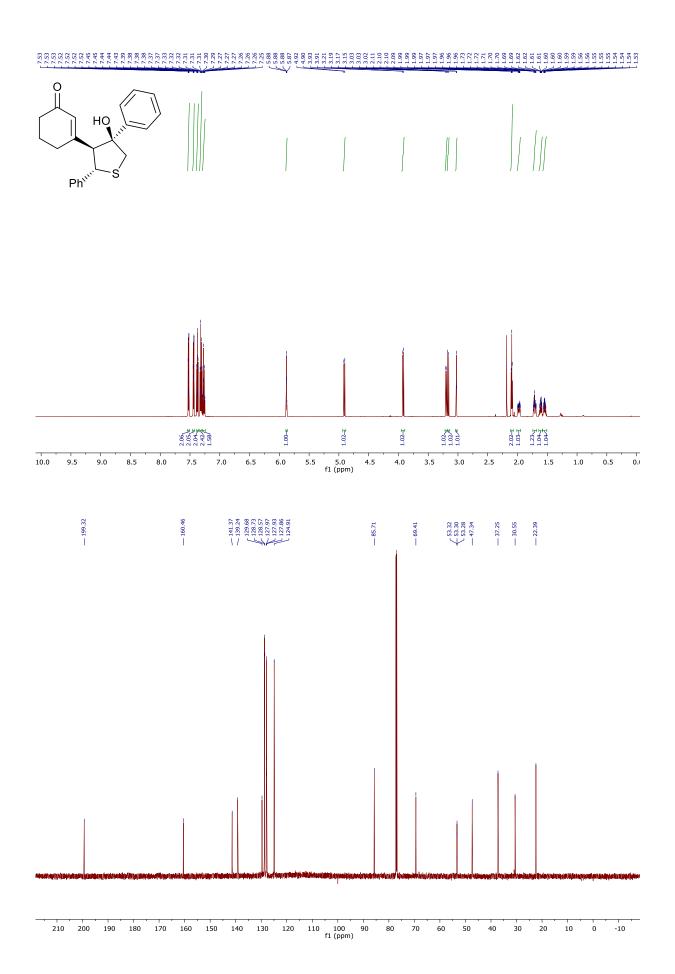


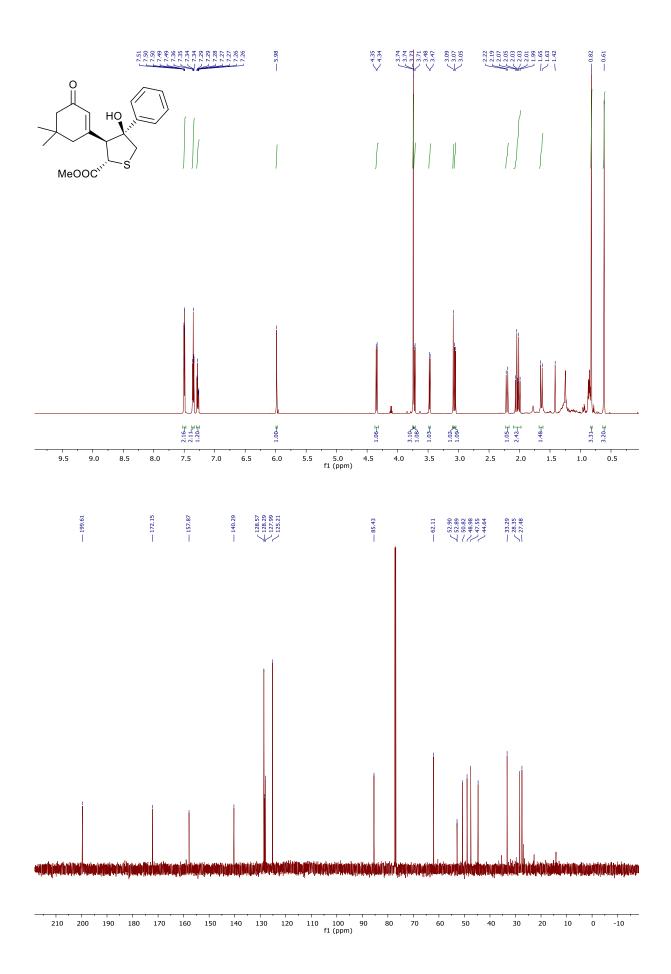


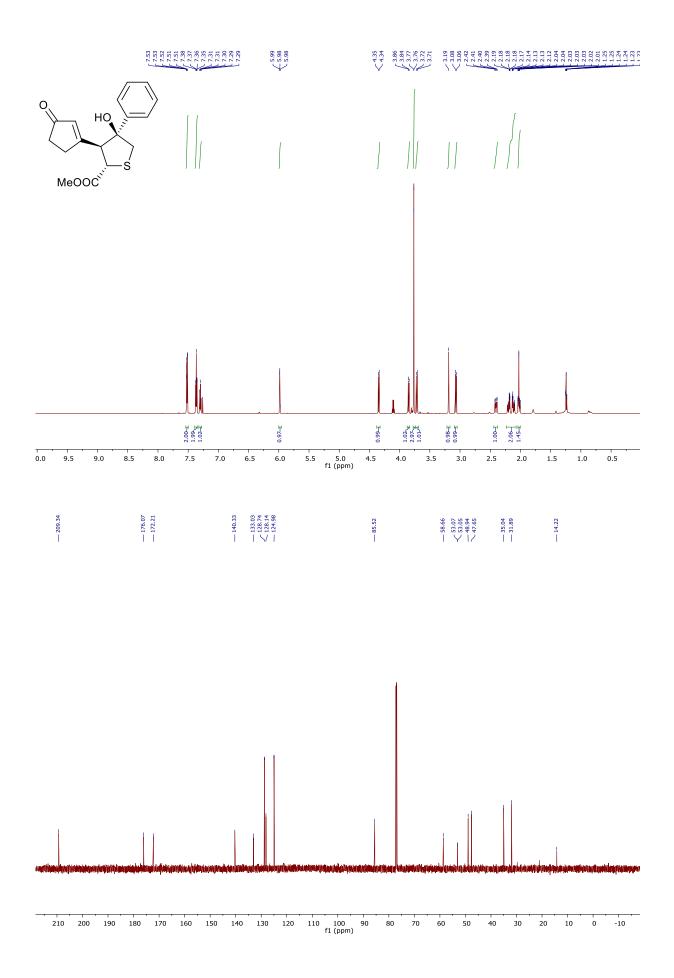


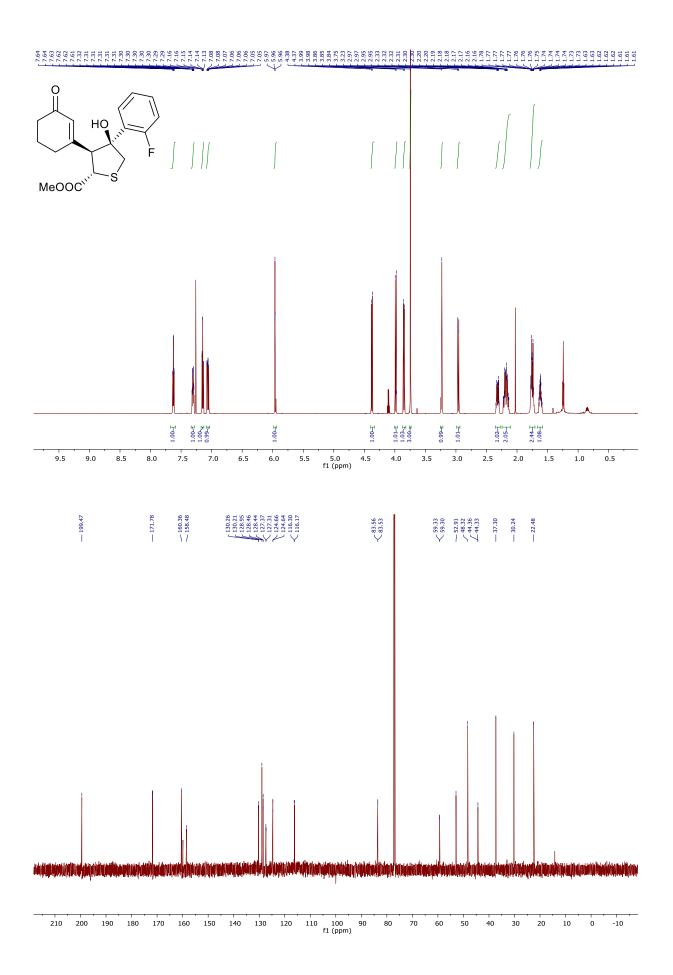


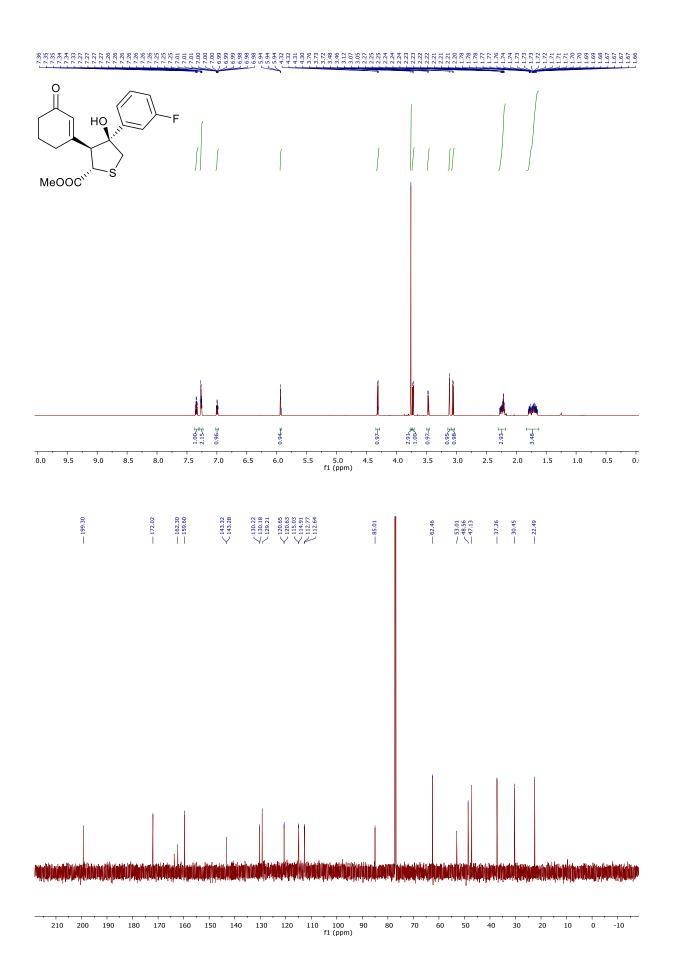


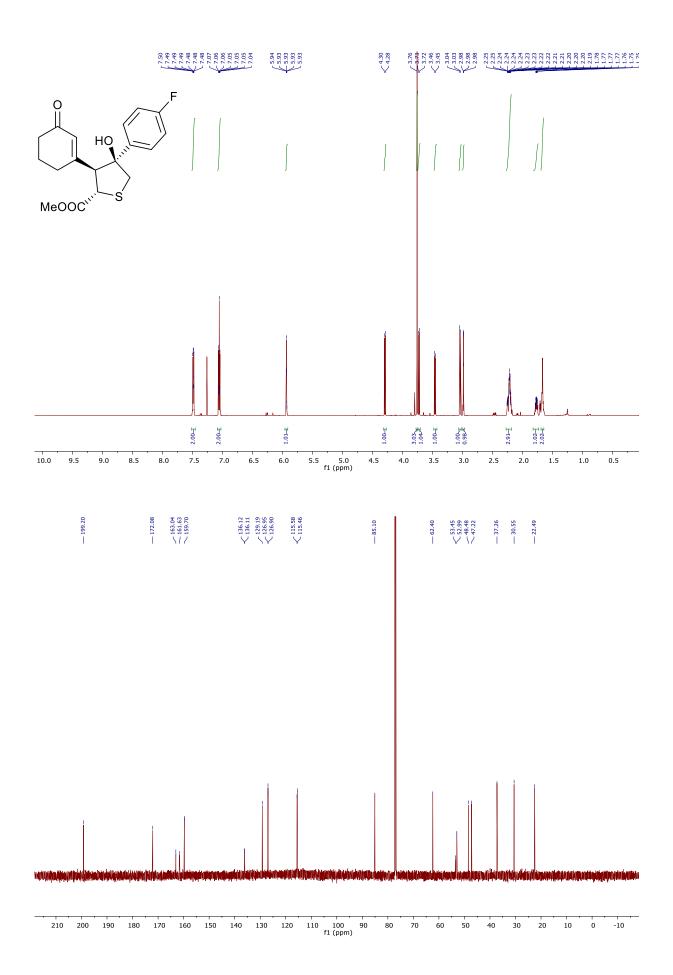


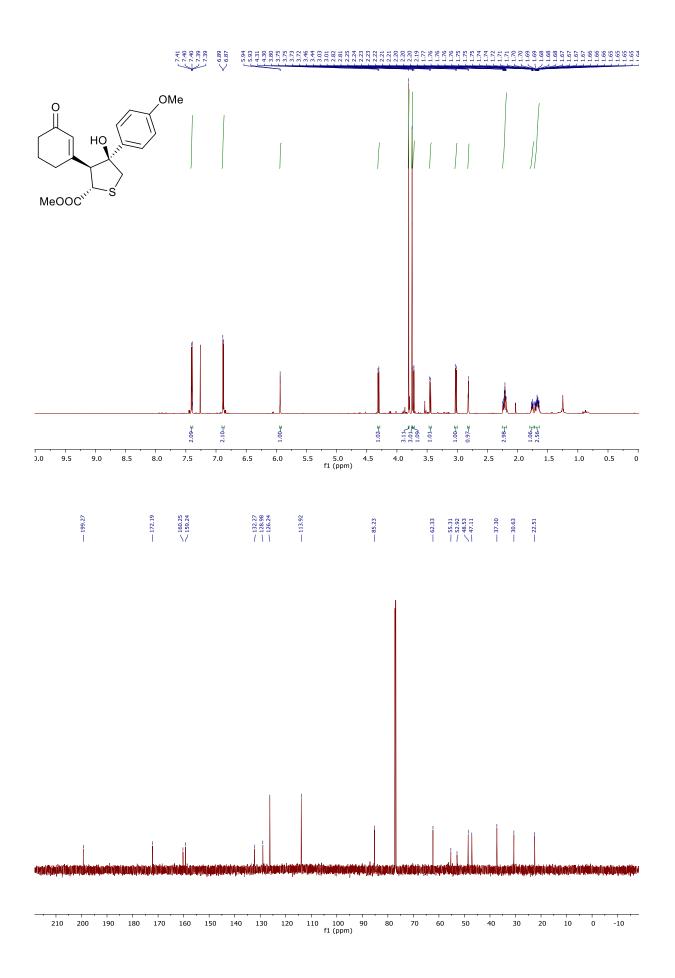






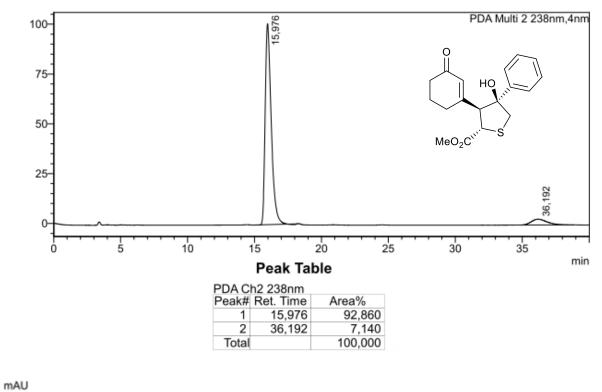


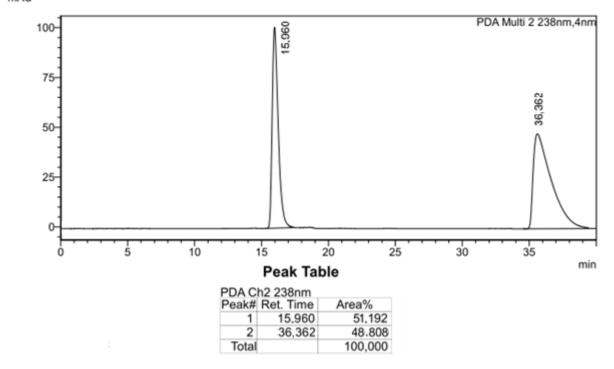


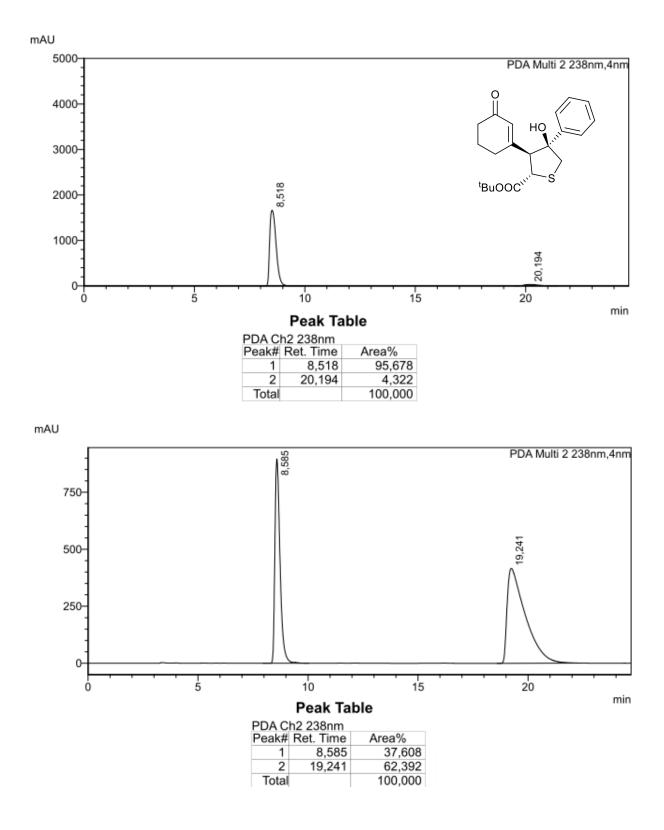


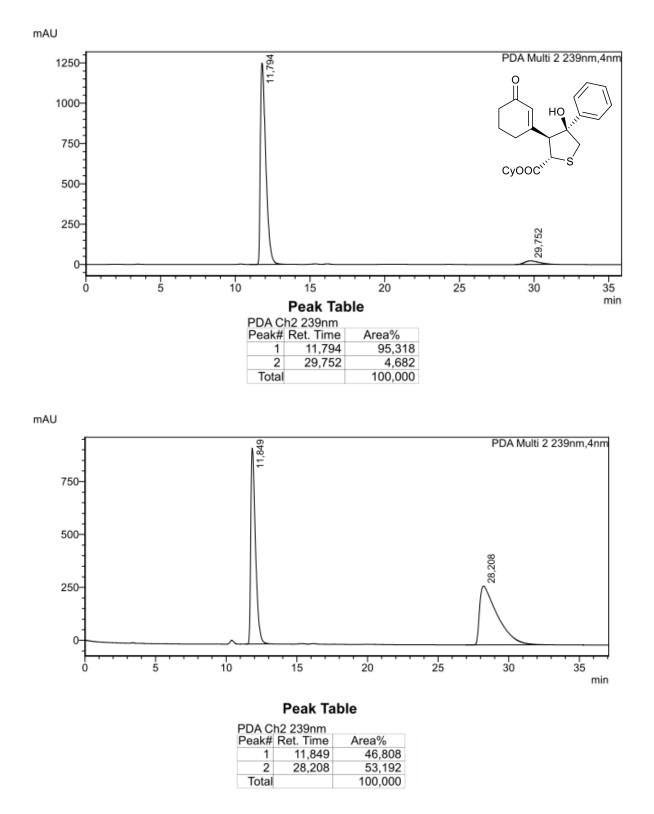
5. HPLC traces



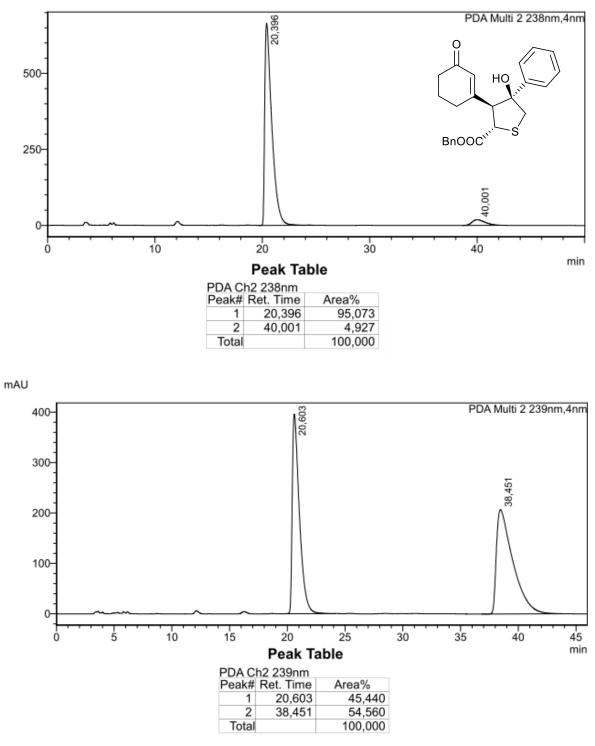




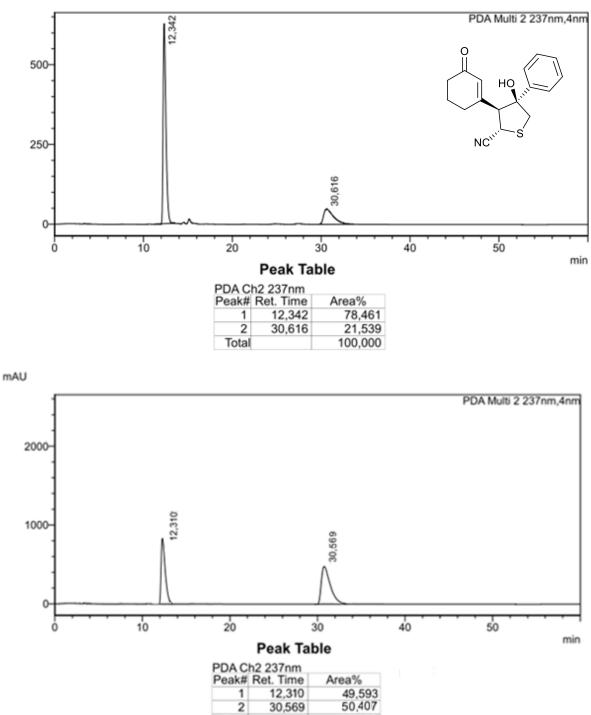








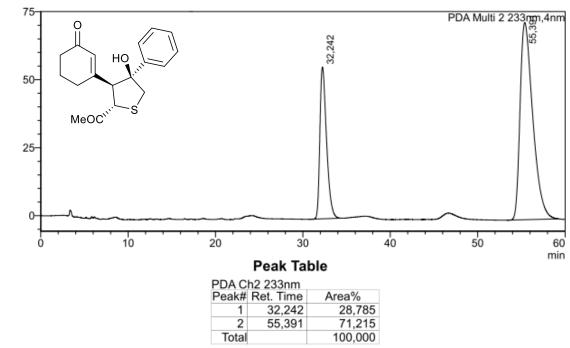


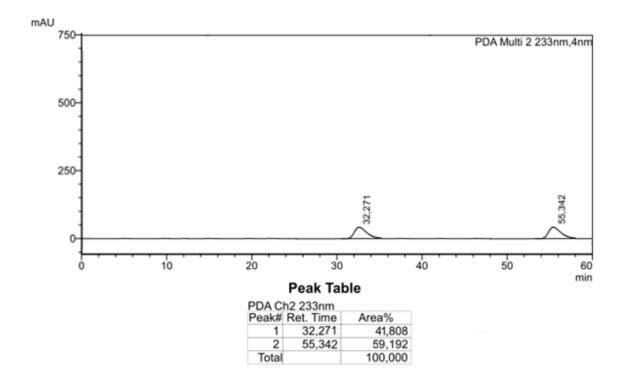


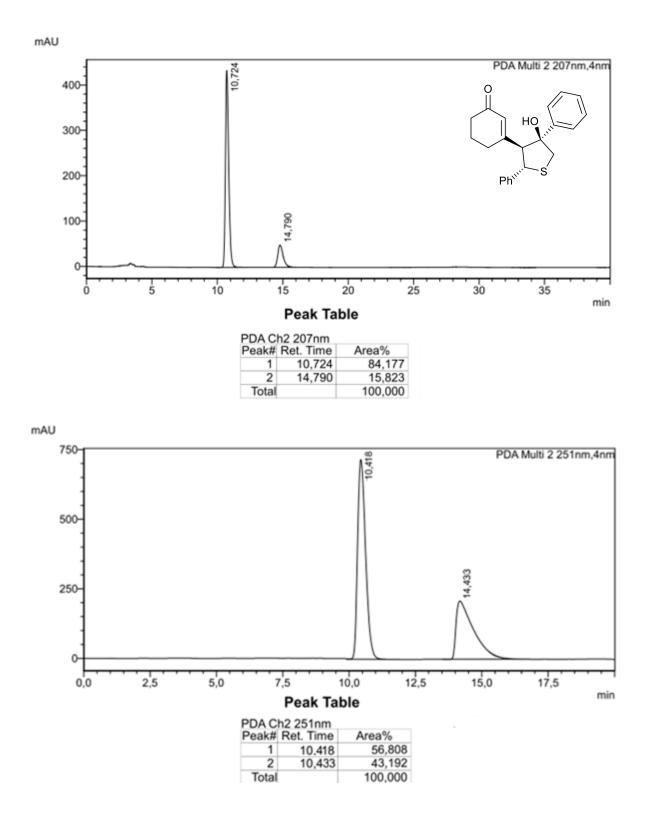
Total

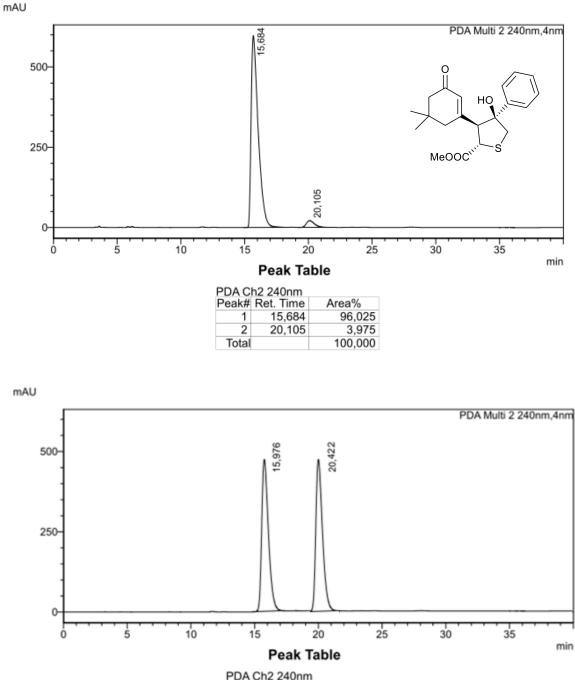
100,000



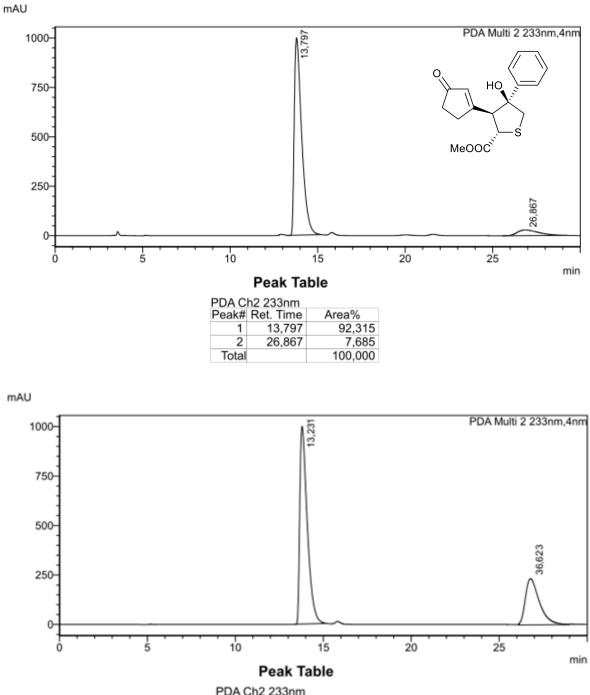




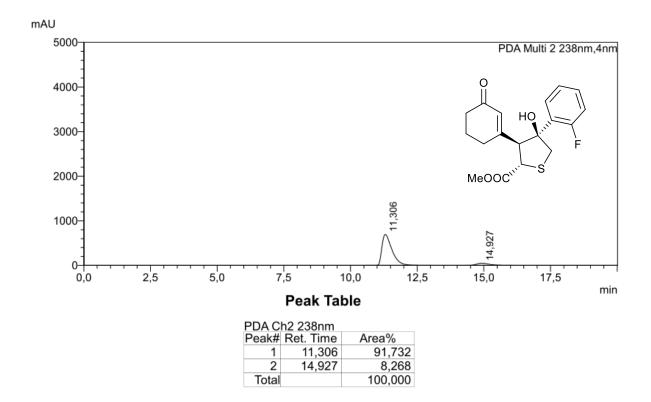


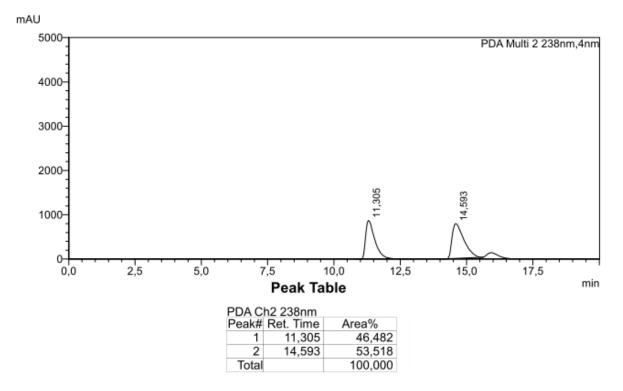


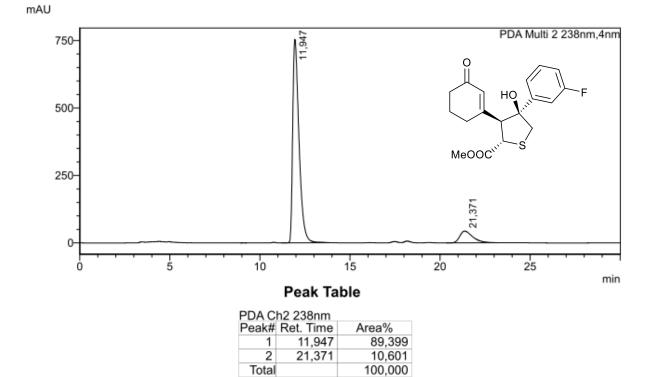
Peak# Ret. Time Area	a%
1 15,976 50	0,401
2 20.422 49	9,599
Total 100	0,000

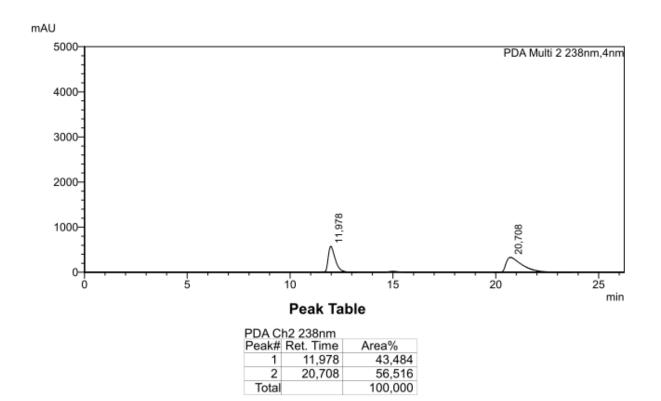


PDA C	h2 233nm	
Peak#	Ret. Time	Area%
1	13,231	48.928
2	36,623	51,072
Total		100,000









S44

