SUPPLEMENTARY INFORMATION

Structural elucidation of major selective androgen receptor modulator (SARM) metabolites for doping control

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Supporting Figures



Figure S1: ¹⁹F NMR spectrum of purified sulphated metabolite 5.



Figure S2: Retention time comparison of the urinary natural and the synthesized purified cyano SARMsderived metabolite **5**. Extracted ion chromatograms on a C18 column for the urinary, synthesized and mixed samples (top right corner of each chromatogram: signal intensities).



Figure S3: Synthesis of purified sulphated metabolite **7**. (i) NIS, MeOH/THF, 91%; (ii) Cul, Cs₂CO₃, 1,10-phenanthrolin-mono hydrate, MeOH, 29%; (iii) BBr₃, DCM, 40%; (iv) SO₃-NMe₃, NaHCO₃, NaOH, H₂O, 45%.



Figure S4: ¹⁹F NMR spectrum of purified sulphated metabolite 7.



Figure S5: Comparison of the urinary natural and the synthesized purified nitro-SARMs-derived metabolite **7** (SARMs **S1**): UHPLC-MS co-injection using a C18 column for the urinary, synthesized and mixed samples (top right corner of each chromatogram: signal intensities).

General

All non-aqueous reactions were performed using flame- or oven dried glassware under an atmosphere of dry nitrogen. All reagents and solvents were purchased from Sigma-Aldrich or Fischer Scientific and were used without further purification. HPLC grade solvents were used for HPLC purification and mass spectrometry grade for UHPLC-ESI-MS analysis. Solutions were concentrated in vacuo on a Heidolph or a IKA rotary evaporator. Thin Layer Chromatography (TLC) was performed on silica gel 60 F-254 plates. Visualization of the developed chromatogram was performed using fluorescence quenching or staining with CAM (Cerium Ammonium Molybdate), Ninhydrin, Ehrlich reagent (4-(Dimethylamino)benzaldehyde) or Vanillin. Chromatographic purification of products was accomplished using flash column chromatography on Merck silica gel 60 (40-63 µm) or preparative reverse phase HPLC on an Agilent HPLC-1100 series system equipped with a Waters Atlantis T3 preparative column (10x100 mm, 5 µm) at a 2,5 mL/min flow rate. All synthesized compounds were ≥95% pure as determined by NMR. NMR spectra were recorded on an Agilent 400 MHz spectrometer (¹H NMR: 399.97 MHz, ¹³C NMR: 100.58 MHz) or Varian 300 MHz spectrometer (¹³C NMR: 75.43 MHz). Chemical shifts are reported in parts per million (ppm) on the δ scale from an internal standard. Multiplicities are abbreviated as follows: s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet. High-resolution mass spectra were acquired on a SYNAPT G2-S High Definition Mass Spectrometry (HDMS) using an electrospray ionization (ESI) source with a AQCUITY UPLC I-class system and equipped with a Waters ACQUITY UPLC BEH C18 column (2.1 × 75 mm, 1.7 µm particle size) or Waters ACQUITY UPLC HSS T3 column (1.8 × 100 mm, 2.1 µm particle size).

UHPLC-MS/MS analysis

The analysis was performed on an Acquity UPLC system hyphenated with a Synapt G2 Q-TOF, both from Waters Corporation (Milford, MA, USA). The system was controlled using the MassLynx software v 4.1, also from Waters. The separation was performed on a Kinetex biphenyl column (1.7 μ m, 100 x 2.1 mm) from Phenomenex (Torrance, CA, USA) and an Acquity BEH C18 column (1.7 μ m, 100 x 2.1 mm) from

Waters Corporation. The mobile phase consisted of A) 0.1 % formic acid in MilliQ water and B) methanol and the flow rate was 0.5 mL/min. For the biphenyl column the column temperature was 40 °C and the gradient was 0-1 min, 5% B; 1-11 min, 5-95% B; 11-12 min, 95% B; 12 min, 95-5% B, 12-14 min, 5% B. For the C18 column, the temperature was set to 65 °C and the gradient was 0-1 min, 5% B; 1-6 min, 5-95% B; 6-8 min, 95% B; 8 min, 95-5% B, 8-10 min, 5% B.

The samples were introduced to the q-TOF using negative electrospray ionization. The capillary voltage was set to -2.50 kV and the cone voltage was 30 V. The source temperature was 120 °C, the cone gas flow 50 L/min and the desolvation gas flow 800 L/min. The instrument was operated in MSE mode, the scan range was m/z 50-1200, and the scan time was 0.2 s. In low energy mode, the collision energy was 4 V and in high energy mode the collision energy was ramped between 20-40 V. A solution of sodium formate (0.5 mM in 2-propanol:water, 90:10, v/v) was used to calibrate the instrument and a solution of leucine-encephalin (2 ng/µl in acetonitrile:0.1% formic acid in water, 50:50, v/v) was used for the lock mass correction.

To control the mass accuracy, a known standard mix consisting of leucine-enkephalin and meloxicam was analyzed before and after every sample set.

Chemical Synthesis

Synthesis of 4-amino-5-hydroxy-2-(trifluoromethyl)benzonitrile (4)



4-Amino-5-methoxy-2-(trifluoromethyl)benzonitrile **13** (35 mg, 160 μ mol) was dissolved in DCM (4 mL) and BBr₃ (243 μ L, 1M solution in heptane, 6 eq) was added dropwise to the reaction mixture at -78 °C. The mixture was stirred at -78 °C and slowly warmed to 25 °C within 4 h. The reaction mixture was then stirred for another 16 h at 25 °C. Upon full consumption of all starting material the reaction mixture was quenched by slow addition of MeOH (2 mL). Silica gel was added to the reaction mixture, the solvent evaporated *in vacuo* and purified by silica gel chromatography using a gradient of MeOH in DCM (1-5%) to afford 4-amino-5-hydroxy-2-(trifluoromethyl)benzonitrile **4** (11 mg, 54 μ mol, 34%).

¹H NMR (400 MHz, CD₃OD) δ (ppm) = 6.99 (s, 2H); ¹⁹F NMR (376 MHz, CD₃OD) δ (ppm) = 62.29 (s, CF₃); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) = 145.7, 142.0, 125.2 (q, ${}^{2}J_{CF}$ = 31.9 Hz), 120.6 (q, ${}^{1}J_{CF}$ = 272 Hz, CF₃), 117.7, 116.9, 110.6 (q, ${}^{3}J_{CF}$ = 4.8 Hz), 94.1 (q, ${}^{4}J_{CF}$ = 2.4 Hz); HRMS (ESI -) calculated for C₈H₄F₃N₂O⁻ (M-H)⁻: 201.0281, found: 201.0278.

Synthesis of 2-amino-5-cyano-4-(trifluoromethyl)phenyl hydrogen sulfate (5)



NaHCO₃ (6.7 mg, 79 µmol, 4 eq) and a sulfur trioxide-trimethyl complex (6.9 mg, 50 µmol, 2.5 eq) were added to a solution of 4-amino-5-hydroxy-2-(trifluoromethyl)benzonitrile **4** (4.0 mg, 20 µmol) and NaOH (2.4 mg, 59 µmol, 3 eq) in water (500 µL). The mixture was heated at 40 °C and stirred for 3 days. Upon full consumption of all starting material the solvent was removed and co-evaporated with MeOH. The crude reaction mixture was subjected to HPLC purification [r.t. = 15 min, 0-5 min (0% B), 5-20 min (0-100% B) at a flow of 2.5 mL/min; buffer A = ammonium acetate 5 mM (water) and buffer B = ammonium acetate 5 mM (MeOH)] to afford 2-amino-5-cyano-4-(trifluoromethyl)phenyl hydrogen sulfate **5** (4.0 mg, 14 µmol, 72%).

¹H NMR (400 MHz, CD₃OD) δ (ppm) = 7.69 (s, 1H), 7.12 (s, 1H); ¹⁹F NMR (376 MHz, CD₃OD) δ (ppm) = 63.29 (s, CF₃); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) = 145.8, 139.8, 129.7 (q, ²*J*_{CF} = 31.8 Hz), 127.2, 122.8 (q, ¹*J*_{CF} = 272 Hz, CF₃), 116.3, 112.3 (q, ³*J*_{CF} = 4.8 Hz), 93.4 (q, ⁴*J*_{CF} = 2.1 Hz); HRMS (ESI-) calculated for C₈H₄F₃N₂O₄S⁻ (M-H)⁻: 280.9849, found: 280.9844.

Synthesis of 2-amino-5-nitro-4-(trifluoromethyl)phenol (6)



2-Methoxy-4-nitro-5-(trifluoromethyl)aniline **16** (40 mg, 169 µmol) was dissolved in DCM (3 mL) and BBr₃ (254 µL, 1M solution in heptane, 6eq) was added dropwise into the reaction mixture at -78 °C. The mixture was stirred at -78 °C and slowly warmed to 25 °C within 4 h. The reaction mixture was then stirred for another 16 h at 25 °C. Upon full consumption of all starting material the reaction mixture was quenched by slow addition of MeOH (2 mL). Silica gel was added in the reaction mixture. The reaction mixture was evaporated. The crude material was purified by silica gel column by using gradient mixture of 1-5% of MeOH in DCM to afford 2-amino-5-nitro-4-(trifluoromethyl)phenol **6** (15 mg, 68 µmol, 40%).

¹H NMR (400 MHz, CD₃OD) δ (ppm) = 7.44 (s, 1H), 7.02 (s, 1H); ¹⁹F NMR (376 MHz, CD₃OD) δ (ppm) = 60.55 (s, CF₃); ¹³C NMR (75 MHz, CD₃OD) δ (ppm) = 144.9, 142.4, 136.0, 123.0 (q, ¹*J*_{CF} = 271 Hz, CF₃), 117.1 (q, ²*J*_{CF} = 33.3 Hz), 111.1, 110.6 (q, ³*J*_{CF} = 6.4 Hz); HRMS (ESI+) calculated for C₇H₆F₃N₂O₃⁺ (M+H)⁺: 223.0325, found: 223.0327.

Synthesis of 2-amino-5-nitro-4-(trifluoromethyl)phenyl hydrogen sulfate (7)



Sodium bicarbonate (9.8 mg, 120 µmol, 4 eq) and a sulfur trioxide-trimethyl complex (10 mg, 73 µmol, 2.5 eq) were added to a solution of 2-amino-5-nitro-4-(trifluoromethyl)phenol **6** (6.5 mg, 29 µmol) and NaOH (3.5 mg, 88 µmol, 3 eq) in water (500 µL). Upon full consumption of all starting material the solvent was removed and co-evaporated with MeOH. The crude reaction mixture was subjected to HPLC purification [r.t.= 16 min, 0-5 min (0% buffer B), 5-20 min (0-100% buffer B); buffer A = ammonium acetate 5 mM (water) and buffer B = ammonium acetate 5 mM (MeOH)] to afford 2-amino-5-nitro-4-(trifluoromethyl)phenyl hydrogen sulfate **7** (4 mg, 10 µmol, 45%).

¹H NMR (400 MHz, CD₃OD) δ (ppm) = 8.14 (s, 1H), 7.15 (s, 1H); ¹⁹F NMR (376 MHz, CD₃OD) δ (ppm) = 61.19 (s, CF₃); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) = 146.2, 138.8, 135.1, 122.5 (q, ¹*J*_{CF} = 272 Hz, CF₃), 121.9 (q, ²*J*_{CF} = 33.2 Hz), 120.1, 112.5 (q, ³*J*_{CF} = 6.5 Hz); HRMS (ESI-) calculated for C₇H₄F₃N₂O₆S⁻ (M-H)⁻: 300.9748, found: 300.9754.

Synthesis of 4-amino-5-iodo-2-(trifluoromethyl)benzonitrile (12)



4-Amino-2-(trifluoromethyl)benzonitrile **11** (400 mg, 2.15 mmol) was dissolved in a 1:1 mixture of THF (10 mL) and methanol (10 mL). *N*-lodosuccinimide (531 mg, 2.36 mmol, 1.1 eq) was then added, followed by addition of about 5 mol % of p-toluenesulfonic acid monohydrate (18.5 mg, 108 μmol). The solution was stirred for 16 h, then concentrated. The crude was dissolved in ethyl acetate (30 mL) and the solution was washed with water twice, then washed with brine and evaporated to afford solid. The crude product was purified by silica gel column chromatography by using gradient mixture of 10-30% EtOAc in hexane to afford 4-amino-5-iodo-2-(trifluoromethyl)benzonitrile **12** (625 mg, 2.00 mmol, 93%).

¹H NMR (400 MHz, (CD₃OD) δ (ppm) = 8.11 (s, 1H), 7.08 (s, 1H); ¹⁹F NMR (376 MHz, ((CD₃)₂CO) = δ (ppm) = 63.38 (s, CF₃); ¹³C NMR (100 MHz, ((CD₃)₂CO) δ (ppm) = 152.7, 145.3, 132.9 (q, ²*J*_{CF} = 33 Hz), 122.8 (q, ¹*J*_{CF} = 272 Hz, CF₃), 115.1, 110.5 (q, ³*J*_{CF} = 5.0 Hz), 95.9, 83.1 (q, ⁴*J*_{CF} = 1.3 Hz); HRMS (ESI+) calculated for C₈H₅F₃IN₂⁺ (M+H)⁺: 312.9444, found: 312.9443.

Synthesis of 4-amino-5-methoxy-2-(trifluoromethyl)benzonitrile (13)¹



4-amino-5-iodo-2-(trifluoromethyl)benzonitrile **12** (150 mg, 481 µmol), Cul (9.2 mg, 48 µmol, 0.1 eq), Cs_2CO_3 (313 mg, 961 µmol, 2 eq) and 1 ,10-phenanthrolin-mono hydrate (17 mg, 96 µmol, 0.2 eq) were mixed in 3 mL of methanol and flushed with nitrogen for 5 minutes. The reaction mixture was heated at 90 °C for 20 h. The solvent was removed and the crude product was purified by silica gel column chromatography by using gradient mixture of 10-30% EtOAc in hexane to afford 4-amino-5-methoxy-2-(trifluoromethyl)benzonitrile **13** (52 mg, 240 µmol, 50%). The analytical data of **13** perfectly agree with reference data.

Synthesis of 2-iodo-4-nitro-5-(trifluoromethyl)aniline (15)



5-Amino-2-nitrobenzotrifluoride **14** (1.00 g, 4.85 mmol) was dissolved in a 1:1 mixture of THF (10 mL) and methanol (10 mL). N-iodosuccinimide (1.20 g, 5.34 mmol, 1.1 eq) were then added, followed by addition of about 5 mol % of p-toluenesulfonic acid monohydrate (41.8 mg, 243 μ mol). The solution was stirred for 16 h, then concentrated. The crude was dissolved in ethyl acetate (30 mL) and the solution was washed with water twice, then washed with brine and evaporated to afford solid. The crude product was purified by silica gel column chromatography by using gradient mixture of 10-30% EtOAc in hexane to afford 2-lodo-4-nitro-5-(trifluoromethyl)aniline **15** (1.47 g, 4.41 mmol, 91%).

¹H NMR (400 MHz, (CD₃OD) δ (ppm) = 8.43 (s, 1H), 7.11 (s, 1H); ¹⁹F NMR (376 MHz, CD₃OD) δ (ppm) = 61.95 (s, CF₃); ¹³C NMR (100 MHz, CD₃OD) δ (ppm) = 153.7, 137.8, 135.5, 125.8 (q, ²*J*_{CF} = 33 Hz), 122.3 (q, ¹*J*_{CF} = 273 Hz, CF₃), 110.7 (q, ³*J*_{CF} = 7.0 Hz), 80.9; HR-MS (ESI+) calculated for C₇H₃F₃IN₂O₂⁻ (M-H)⁻: 330.9197, found: 330.9193.

Synthesis of 2-methoxy-4-nitro-5-(trifluoromethyl)aniline (16)



2-lodo-4-nitro-5-(trifluoromethyl)aniline **15** (60 mg, 180 μ mol), Cul (3.4 mg, 18 μ mol, 0.1 eq), Cs₂CO₃ (118 mg, 361 μ mol, 2 eq) and 1 ,10-phenanthrolin-mono hydrate (6.5 mg, 36 μ mol, 0.2 eq) were mixed in 3 mL of methanol and flushed with nitrogen for 5 minutes. The reaction mixture was heated at 90 °C for 20 h. The solvent was removed and the crude product was purified by silica gel column chromatography by using gradient mixture of 10-30% EtOAc in hexane to afford 2-methoxy-4-nitro-5-(trifluoromethyl)aniline **16** (12.3 mg, 52.1 μ mol, 29%) and 4-nitro-3-(trifluoromethyl)aniline (16.3 mg, 78 μ mol, 43%).

¹H NMR (400 MHz, CD₃OD) δ (ppm) = 7.61 (s, 1H), 7.05 (s, 1H), 3.97 (s, 3H, OMe); ¹⁹F NMR (376 MHz, CDCl₃) δ (ppm) = 59.29 (s, CF₃); ¹³C NMR (100 MHz, CDCl₃) δ (ppm) = 146.9, 140.9, 122.4 (q, ¹ J_{CF} = 273 Hz, CF₃), 119.0 (q, ² J_{CF} = 33.8 Hz), 111.4 (q, ³ J_{CF} = 6.3 Hz), 108.4, 107.6, 56.3; HRMS (ESI-) calculated for C₈H₆F₃N₂O₃⁻ (M-H)⁻: 235.0336, found: 235.0329.

References

1 X. Zhang, X. Li, G. F. Allan, T. Sbriscia, O. Linton, S. G. Lundeen and Z. Sui, *J. Med. Chem.*, 2007, **50**, 3857-3869.

NMR Spectra

4-Amino-5-hydroxy-2-(trifluoromethyl)benzonitrile (4) $^{1}\mathrm{H}$ NMR, $^{13}\mathrm{C}$ NMR, $^{19}\mathrm{F}$ NMR

2-Amino-5-cyano-4-(trifluoromethyl)phenyl hydrogen sulfate (5) $^{1}\mathrm{H}$ NMR, $^{13}\mathrm{C}$ NMR, $^{19}\mathrm{F}$ NMR

2-Amino-5-nitro-4-(trifluoromethyl)phenol (6) ¹H NMR, ¹³C NMR, ¹⁹F NMR

2-Amino-5-nitro-4-(trifluoromethyl)phenyl hydrogen sulfate (7) $^{1}\mathrm{H}$ NMR, $^{13}\mathrm{C}$ NMR, $^{19}\mathrm{F}$ NMR

4-Amino-5-iodo-2-(trifluoromethyl)benzonitrile (**12**) ¹H NMR, ¹³C NMR, ¹⁹F NMR

2-Iodo-4-nitro-5-(trifluoromethyl)aniline (**15**) ¹H NMR, ¹³C NMR, ¹⁹F NMR

2-Methoxy-4-nitro-5-(trifluoromethyl)aniline (**16**) ¹H NMR, ¹³C NMR, ¹⁹F NMR









































