

Supporting Information

Fingerprinting antibiotics with PAE-based fluorescent sensor arrays

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1. General Information.

Chemicals were purchased from commercial laboratory suppliers. Reagents were used without further purification unless otherwise noted.

Solvents were purchased from commercial laboratory suppliers and if necessary distilled prior use. Absolute solvents were dried by a MB SPS-800 using drying columns.

Analytical thin layer chromatography (TLC) was performed on Macherey & Nagel Polygram® SIL G/UV254 precoated plastic sheets. Components were visualized by observation under UV light (254 nm or 365 nm) or in the case of UV-inactive substances by using the suitably colouring solutions. The following colouring solutions were used for the visualization of UV-inactive substances:

KMnO₄ solution: 2.0 g KMnO₄, 10.0 g K₂CO₃, 0.3 g NaOH, 200 mL distilled water.

Cer solution: 10.0 g Ce₂(SO)₃, 25 g phosphomolybdic acid hydrate, 1 L distilled water, 50 mL conc. H₂SO₄.

Flash column chromatography was carried out using silica gel S (0.032 mm-0.062 mm), purchased from Sigma Aldrich, according to G. Nill, unless otherwise stated.¹

¹H NMR spectra were recorded at room temperature on the following spectrometers: Bruker Avance III 300 (300 MHz), Bruker Avance III 400 (400 MHz), Bruker Avence III 500 (500 MHz) and Bruker Avance III 600 (600 MHz). The data were interpreted in first order spectra. The spectra were recorded in CDCl₃, D₂O or DMSO-d₆ as indicated in each case. Chemical shifts are reported in δ units relative to the solvent residual peak or TMS.² The following abbreviations are used to indicate the signal multiplicity: s (singlet), d (doublet), t (triplet), q (quartet), quin (quintet), sext (sextet), dd (doublet of doublet), dt (doublet of triplet), ddd (doublet of doublet of doublet), etc., bs (broad signal), m (multiplet).

¹³C NMR spectra were recorded at room temperature on the following spectrometers: Bruker Avance III 300 (75 MHz), Bruker Avance III 400 (100 MHz), Bruker Avence III 500 (125

MHz) and Bruker Avance III 600 (150 MHz). The spectra were recorded in CDCl_3 , D_2O or DMSO-d_6 as indicated in each case. Chemical shifts are reported in δ units relative to the solvent signal or TMS.²

High resolution mass spectra (HR-MS) were either recorded on a Bruker ApexQehybrid 9.4 T FT-ICR-MS (ESI^+ , DART^+), a Finnigan LCQ (ESI^+) or a JEOL JMS-700 (EI^+) mass spectrometer at the Organisch-Chemisches Institut der Universität Heidelberg.

Absorption and emission spectra were recorded using a Jasco V660 and Jasco FP6500 spectrometer. Emission data for sensing were recorded on a CLARIOstar (firmware version 1.13) Platereader from BMG Labtech using the corresponding software (software version 5.20 R5). Data were analysed with CLARIOstar MARS Data Analysis Software (software version 3.10 R5) from BMG Labtech.

IR spectra were recorded on a JASCO FT/IR-4100. Substances were applied as a film, solid or in solution. The obtained data was processed with the software JASCO Spectra Manager™ II.

Fluorescence lifetimes τ were acquired by an exponential fit according to the least mean square with commercially available software HORIBA Scientific Decay Data Analyses 6 (DAS6) version 6.4.4. The luminescence decays were recorded with a HORIBA Scientific Fluorocube single photon counting system operated with HORIBA Scientific DataStation version 2.2.

Quantum yields Φ were measured by using the comparative method with quinine sulfate in 0.1 N sulfuric acid as a reference ($\Phi = 0.54$) according to the literature, the average values of three measurements were calculated for each sample.³

Dialysis was realized with regenerated cellulose tubular membranes (ZelluTrans, Carl Roth[®]) with a molecular weight cut-off of 3500 Da against deionized (DI) water.

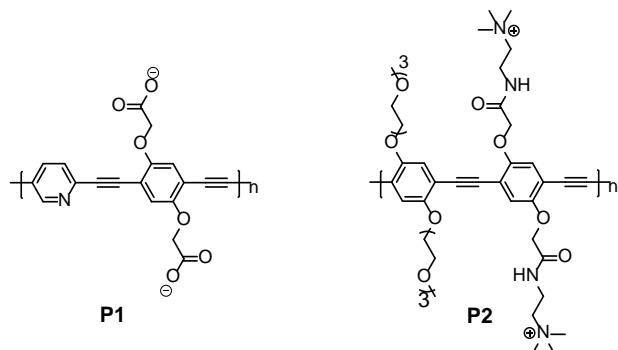
Gel Permeation Chromatography (GPC): Number- (M_n) and weight-average (M_w) molecular weights and polydispersities (PDI, M_w/M_n) were determined by GPC versus polystyrene standards. Measurements were carried out at room temperature in chloroform or THF with PSS-

SDV columns (8.0 mm x 30.0 mm, 5 μ m particles, 10^2 -, 10^3 - and 10^5 - \AA pore size) on a Jasco PU-2050 GPC unit equipped with a Jasco UV-2075 UV- and a Jasco RI-2031 RI-detector.

Linear discriminant analysis (LDA) and principal component analysis (PCA). Both methods were carried out in this study by using SYSTAT (version 13.0). In LDA, all variables were used in the model (complete mode) and the tolerance was set as 0.001. The fluorescence response patterns were transformed to canonical patterns. The Mahalanobis distances of each individual pattern to the centroid of each group in a multidimensional space were calculated and the assignment of the case was based on the shortest Mahalanobis distance. PCA is a mathematical transformation used to extract variance between entries in a data matrix by reducing the redundancy in the dimensionality of the data. It takes the data points for all analytes and generates a set of orthogonal eigenvectors (principal components, PCs) for maximum variance.⁴

2. Synthetic Details and Analytical Data.

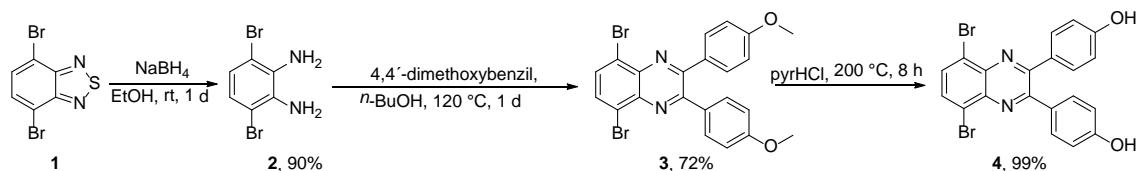
2.1 Synthesis of P1/P2



Compound **P1** was synthesized according to the literature.⁵

Compound **P2** was synthesized according to the literature.⁶

2.2 Synthesis of P3/P4



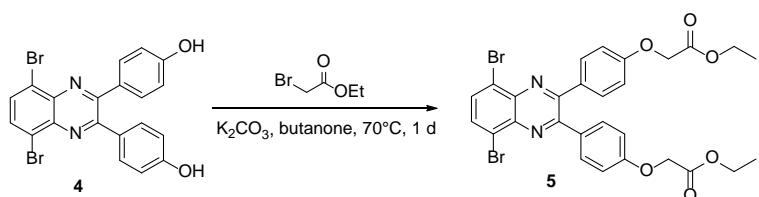
Compound **1** was synthesized according to the literature.⁷

Synthesis of 2. Compound **1** (1.50 g, 5.10 mmol) was dissolved in EtOH (50 ml) and cooled with an ice/water bath. NaBH₄ (10.2 mmol, 3.86 g) was added in small portions and the reaction mixture was stirred over night. The solidified reaction mixture was evaporated to dryness and cautiously diluted with water (100 mL). Water and Et₂O were added, the aqueous layer was separated and extracted with CH₂Cl₂. The combined organic layers were washed with brine, dried over MgSO₄ and concentrated in vacuo to afford compound **2** (1.23 g, 90%) as yellow solid. The product was used without further purification in the next step. ¹H NMR (300 MHz, DMSO-D₆): δ = 6.63 (s, 2 H), 5.00 (s, 4 H) ppm.

Synthesis of 3. Compound **2** (1.23 g, 4.63 mmol) and 4,4'-dimethoxybenzil (2.75 g, 10.2 mmol) were dissolved in *n*-BuOH (50 ml). A few drops of acetic acid were added and the reaction mixture was stirred at 120°C for 1 d. The solidified reaction mixture was suspended in

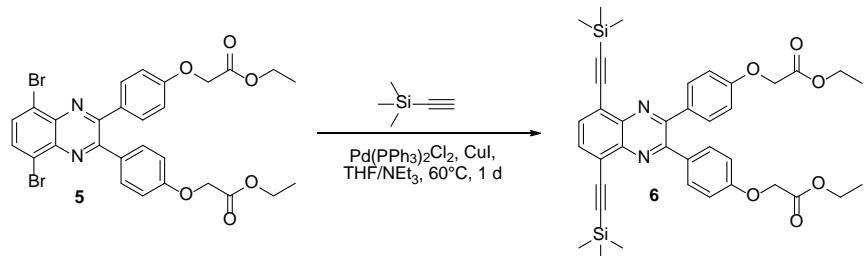
EtOH and filtrated. The residue was washed with copious amounts of hot ethanol and dried in vacuo to afford compound **3** (1.67 g, 72%) as ocre solid (m. p. 188-190 °C). ¹H NMR (400 MHz, CDCl₃): δ = 7.86 (s, 2H), 7.65-7.70 (m, 4H), 6.89-6.93 (m, 4H), 3.86 (s, 6H) ppm. ¹³C NMR (100 MHz, CDCl₃): δ = 160.91, 153.53, 139.09, 132.57, 131.71, 130.62, 123.49, 113.90, 55.35 ppm. IR (cm⁻¹): ν 3079, 3001, 2956, 2929, 2836, 1900, 1603, 1577, 1576, 1560, 1558, 1537, 1526, 1525, 1522, 1512, 1458, 1451, 1443, 1436, 1415, 1382, 1331, 1305, 1292, 1248, 1236, 1218, 1179, 1170, 1111, 1064, 1025, 1015, 1008, 987, 962, 948, 899, 844, 837, 829, 813, 801, 792, 758, 737, 718, 673, 670, 660, 652, 635, 627, 604, 592, 568, 555, 552, 533, 521, 500, 492, 470, 463. HR-MS (EI⁺): *m/z* calcd. for C₂₂H₁₆Br₂N₂O₂⁺ 497.95730 [M·]⁺; found 497.95808. C₂₂H₁₆Br₂N₂O₂ (499.18): calcd. C 52.83, H 3.22, N 5.60; found C 52.46, H 3.38, N 5.68.

Synthesis of 4. Compound **3** (1.50 g, 3.00 mmol) and pyridine hydrochloride (1.04 g, 90.0 mmol) was stirred at 200°C for 8 h. The cooled reaction mixture was dissolved in Et₂O and quenched with H₂O. The aqueous phase was extracted with Et₂O and the combined organic layers were washed with water and brine. Evaporation in vacuo afforded compound **4** (1.41 g, 99%) as light yellow solid (m. p. 302-304 °C). The product was used without further purification in the next step. ¹H NMR (400 MHz, DMSO-D₆): δ = 9.89 (br. s, 2H), 8.05 (s, 2H), 7.42-7.48 (m, 4H), 6.75-6.83 (m, 4H) ppm.



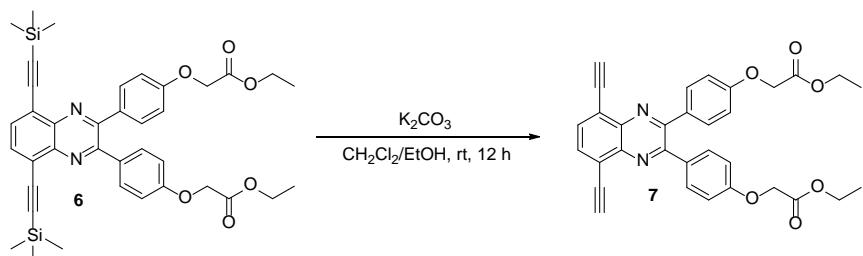
Synthesis of 5. Compound **4** (500 mg, 1.06 mmol) and ethylbromo acetate (0.26 mL, 389 mg, 2.33 mmol) were dissolved in degassed butanone. K₂CO₃ (1.46 g. 10.6 mmol) was added and the mixture was stirred at 70°C for 1 d until TLC showed a complete conversion. The reaction mixture was quenched with H₂O and the aqueous phase was extracted with ethyl acetate. The combined organic layers were dried over MgSO₄, filtered and concentrated in vacuo. The crude

product was dissolved in ethyl acetate, filtered over a silica gel plug and evaporated at 80°C and 0.09 mbar for 2 h to afford compound 5 (605 mg, 89%) as yellow slowly solidifying oil (m. p. 131-133 °C). ¹H NMR (600 MHz, CDCl₃): δ = 7.88 (s, 2H), 7.63-7.69 (m, 4H), 6.88-6.93 (m, 4H), 4.67 (s, 4H), 4.30 (q, *J* = 7.2 Hz, 4H), 1.32 (t, *J* = 7.2 Hz, 6H) ppm. ¹³C NMR (150 MHz, CDCl₃): δ = 168.85, 159.37, 153.55, 139.41, 133.09, 132.09, 131.75, 123.80, 114.89, 65.66, 61.83, 14.49 ppm. IR (cm⁻¹): ν 3080, 2999, 2979, 2938, 2908, 1749, 1725, 1606, 1578, 1540, 1511, 1473, 1440, 1415, 1386, 1332, 1307, 1292, 1262, 1235, 1204, 1173, 1114, 1078, 1069, 1055, 1026, 1014, 986, 947, 932, 899, 875, 850, 834, 824, 814, 801, 758, 735, 720, 698, 653, 640, 607, 591, 572, 551, 530, 520, 492, 467. HR-MS (DART+): m/z calcd. for C₂₈H₂₄Br₂N₂O₆⁺ 641.99956 [M·]⁺; found 642.00065. C₂₈H₂₄Br₂N₂O₆ (644.32): calcd. C 52.20, H 3.75, Br 24.80, N 4.35, O 14.90; found C 52.68, H 4.01, N 4.40.

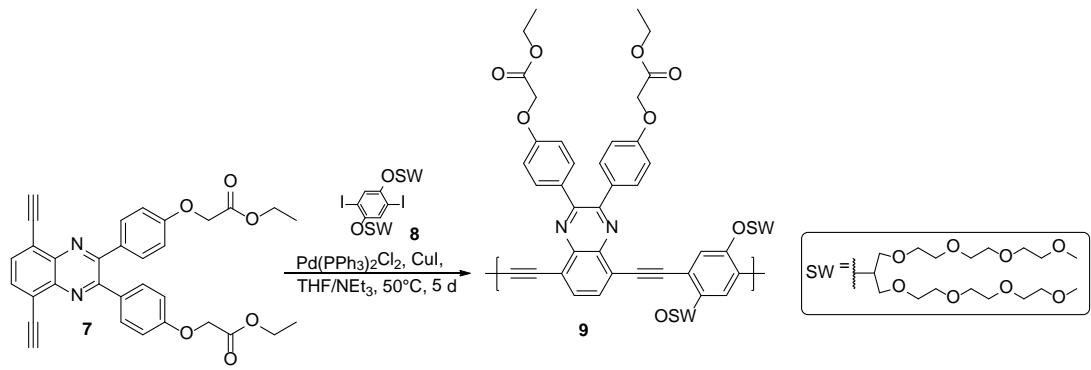


Synthesis of 6. Compound 5 (350 mg, 543 μmol) was dissolved in a degassed mixture of THF/NEt₃ (1:1, 3 mL/3 mL). PdCl₂(PPh₃)₂ (19 mg, 27 μmol) and CuI (5.2 mg, 27 μmol) were added, then TMS-acetylene (168 μL, 117 mg, 1.20 mmol) was added dropwise and the resulting mixture was stirred for 1 d at 60°C until TLC showed a complete conversion. The reaction mixture was filtrated over Celite® and evaporated in vacuo. The crude product was purified by flash chromatography on silica gel [petroleum ether/ethyl acetate (5/1)] to give compound 6 (312 mg, 85%) as yellow solid (m. p. 163 – 165 °C). ¹H NMR (600 MHz, CDCl₃): δ = 7.79 (s, 1H), 7.70 (d, *J* = 8.8 Hz, 2H), 6.88 (d, *J* = 8.8 Hz, 2H), 4.66 (s, 2H), 4.29 (q, *J* = 7.2 Hz, 2H), 1.32 (t, *J* = 7.2 Hz, 3H), 0.35 (s, 9H) ppm. ¹³C NMR (150 MHz, CDCl₃): δ = 168.59, 158.85, 152.04, 140.78, 132.83, 132.23, 131.71, 123.16, 114.31, 103.46, 101.28, 65.36, 61.49, 14.17, -

0.04 ppm. IR (cm^{-1}): ν 2957, 2933, 2898, 2154, 1755, 1604, 1581, 1537, 1511, 1478, 1463, 1437, 1411, 1377, 1341, 1301, 1283, 1271, 1247, 1217, 1194, 1176, 1115, 1081, 1062, 1026, 966, 952, 929, 839, 757, 733, 722, 698, 673, 661, 649, 634, 618, 575, 542, 526, 496, 480, 456. HR-MS (ESI $^+$): m/z calcd. for $\text{C}_{38}\text{H}_{43}\text{N}_2\text{O}_6\text{Si}_2^+$ 679.26542 [M+H] $^+$; found 679.26551. $\text{C}_{38}\text{H}_{42}\text{N}_2\text{O}_6\text{Si}_2$ (678.93): calcd. C 67.23, H 6.24, N 4.13; found C 66.94, H 6.15, N 3.67.

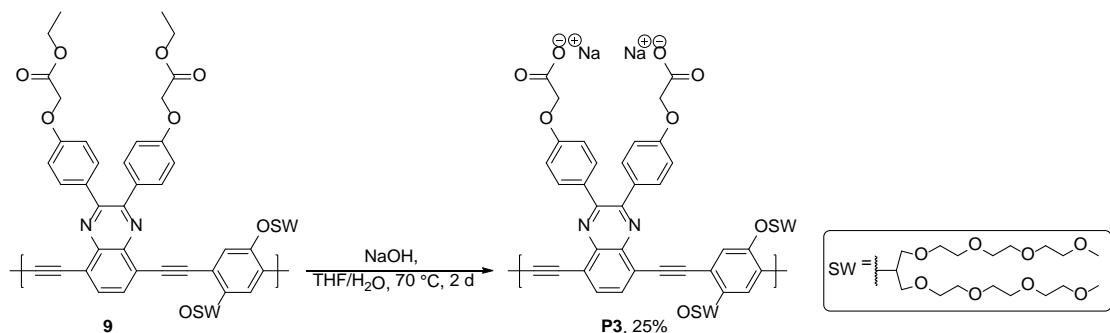


Synthesis of 7. Compound **6** (300 mg, 442 μmol) was dissolved in a degassed mixture of EtOH/ CH_2Cl_2 (1:1, 10 mL/5 mL). K_2CO_3 (610 mg, 4.42 mmol) was added and the resulting mixture was stirred for 12 h at ambient temperature until TLC showed a complete conversion. H_2O and CH_2Cl_2 were added, the aqueous layer was separated and extracted with CH_2Cl_2 . The combined organic layers were washed with H_2O and brine, dried over MgSO_4 and concentrated in vacuo. The crude product was dissolved in CH_2Cl_2 , filtered over a silica gel plug and concentrated in vacuo to give compound **7** (181 mg, 76%) as yellowish solid (m. p. 162 $^\circ\text{C}$ decomposition). ^1H NMR (300 MHz, CDCl_3): $\delta = \delta = 7.87$ (s, 1H), 7.58-7.68 (m, 2H), 6.86-6.92 (m, 2H), 4.65 (s, 2H), 4.30 (q, $J = 7.1$ Hz, 2H), 3.63 (s, 1H), 1.32 (t, $J = 7.1$ Hz, 3H) ppm. ^{13}C NMR (75 MHz, CDCl_3): $\delta = 168.74, 159.06, 153.14, 141.20, 133.67, 132.29, 131.90, 123.00, 114.68, 85.34, 80.21, 77.16, 65.57, 61.65, 14.34$ ppm. IR (cm^{-1}): ν 3304, 3243, 2982, 2938, 1752, 1716, 1606, 1581, 1567, 1540, 1511, 1476, 1446, 1415, 1382, 1338, 1308, 1295, 1261, 1211, 1179, 1142, 1114, 1092, 1077, 1064, 1051, 1029, 1013, 964, 938, 925, 855, 838, 826, 804, 770, 741, 702, 661, 632, 615, 570, 534, 518, 470. HR-MS (ESI $^+$): m/z calcd. for $\text{C}_{32}\text{H}_{27}\text{N}_2\text{O}_6^+$ 535.18636 [M+H] $^+$; found 535.18687. $\text{C}_{32}\text{H}_{26}\text{N}_2\text{O}_6$ (499.18): calcd. C 71.90, H 4.90, N 5.24; found C 71.86, H 5.49, N 4.63.

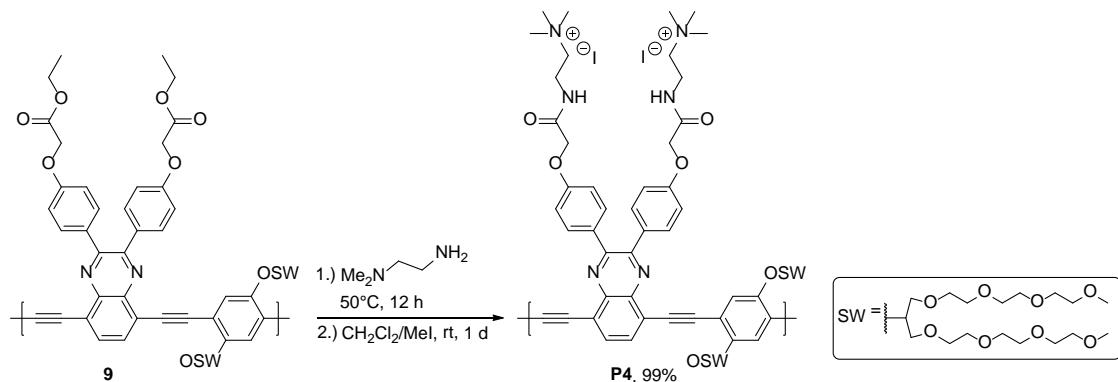


Compound 8 was synthesized according to the literature.⁸

Synthesis of 9. Monomer **7** (200 mg, 374 µmol) and monomer **8** (410 mg, 374 µmol) were dissolved in a mixture of degassed THF/CHCl₃/NEt₃ (3:1:4, 1.5 mL/0.5 mL/2 mL). Pd(PPh₃)₄ (5.3 mg, 7.5 µmol) and CuI (2.9 mg, 15 µmol) were added and the mixture was stirred at 50 °C for 5 d. Saturated aqueous NH₄Cl and CH₂Cl₂ were added, the aqueous layer was separated and extracted with CH₂Cl₂. The combined organic layers were dried over MgSO₄, filtered and concentrated in vacuo. Two times, the crude product was dissolved in a small amount of CHCl₃ and slowly added to an excess of pentane to give **9** as dark red solid (328 mg, 63%). The *M_n* was estimated to be 2.1 × 10⁴ with a PDI of 1.5. ¹H NMR (300 MHz, CDCl₃): δ = 7.88-8.05 (m, 2 H), 7.63-7.76 (m, 4 H), 7.34-7.50 (m, 2 H), 6.89-7.01 (m, 4 H), 4.60-4.77 (m, 6 H), 4.24-4.36 (m, 4 H), 3.44-3.88 (m, 56 H), 3.27-3.39 (m, 12 H), 1.28-1.36 (m, 12 H) ppm. IR (cm⁻¹): ν 2917, 2872, 2364, 2340, 2328, 1758, 1738, 1605, 1513, 1497, 1468, 1447, 1407, 1380, 1337, 1303, 1274, 1248, 1200, 1180, 1108, 1032, 958, 843, 620, 577. Due to low solubility, ¹³C NMR spectrum could not be obtained.

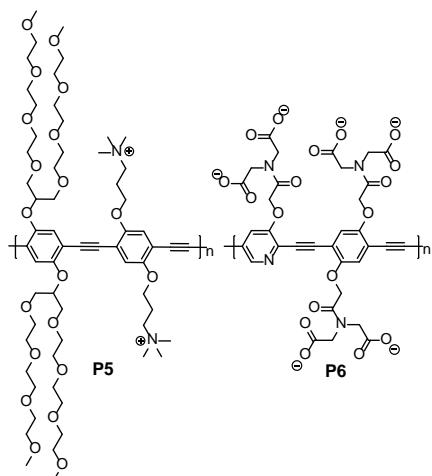


Synthesis of P3. Compound **9** (114 mg, 83 μmol) was dissolved in H_2O (20 mL), NaOH (65 mg, 1.66 mmol) was added and the resulting mixture was stirred at 70 °C for 2 d. After adjusting a pH of 7 (HCl) the aqueous mixture was dialyzed against DI H_2O for 3 d. Freeze-drying gave **P3** as spongy red solid (28 mg, 25%). The M_n and PDI result from **9**. ^1H NMR (500 MHz, D_2O): δ = 7.72-8.15 (m, 4 H), 7.41-7.68 (m, 2 H), 6.36-7.09 (m, 4 H), 3.05-4.03 (m, 74 H) ppm. IR (cm^{-1}): ν 3429, 3263, 3038, 2871, 1668, 1604, 1532, 1510, 1474, 1445, 1411, 1380, 1348, 1296, 1246, 1179, 954, 924, 841, 661, 577, 528. Due to low solubility, ^{13}C NMR spectrum could not be obtained.



Synthesis of P4. Compound **9** (113 mg, 81.9 μmol) was dissolved in $\text{N,N}'\text{-dimethyl-ethylene diamine}$ (15 mL) and stirred at 70 °C for 12 h. The reaction mixture was evaporated to dryness and washed with copious amounts of *n*-pentane. The crude product was dissolved in CH_2Cl_2 (10 mL). After addition of MeI (5 mL) the reaction mixture was stirred for 1 d at ambient temperature. All volatiles were removed under reduced pressure. The residue was dissolved in H_2O and dialyzed against DI H_2O for 3 d. Freeze-drying afforded **P4** as spongy red solid (141 mg, 99%). The M_n and PDI result from **9**. ^1H NMR (500 MHz, D_2O): δ = 6.91-7.77 (m, 10 H), 7.63-7.75 (m, 2 H), 4.41-4.74 (m, 6 H), 3.10-3.84 (m, 4 H) ppm. IR (cm^{-1}): ν 3392, 2921, 2874, 1720, 1646, 1603, 1511, 1444, 1348, 1297, 1200, 1175, 1073, 944, 878, 838, 775, 675, 647, 601, 576, 529. Due to low solubility, ^{13}C NMR spectrum could not be obtained.

2.3 Synthesis of P5/P6



Compound **P5**, **P6** was synthesized according to the literature.⁵

2.4 GPC and Optical Data of P1-P6

Table S1. Additional analytical data of P1-P6.

No.	M _n ^a [g/mol]	M _w ^a [g/mol]	PDI ^a	P _n	λ _{max,abs.} ^b [nm]	λ _{max,em.} ^b [nm]	Φ ^b [%]
P1	6.9 × 10 ³	1.3 × 10 ⁴	1.9	17	415	536	2
P2	1.1 × 10 ⁴	1.7 × 10 ⁴	1.5	15	404	460	4
P3	2.1 × 10 ⁴	3.2 × 10 ⁴	1.5	13	477	546	2
P4	2.1 × 10 ⁴	3.2 × 10 ⁴	1.5	13	403	550	4
P5	1.4 × 10 ⁴	5.5 × 10 ⁴	3.9	11	410	459	37
P6	1.1 × 10 ⁴	1.8 × 10 ⁴	1.5	12	390	443	8

^aDetermined by gel permeation chromatography of the corresponding organosoluble precursors.

^bMeasured in KH₂PO₄/Na₂HPO₄ buffer solution.

3. Fluorescence Titration of Polymers with Surfactant

Fluorescence titration of weakly fluorescent PAE (**P1-P4**) with increasing concentration of surfactants (SDBS and CTMA) at different pH condition (pH3, pH7, and pH13 buffer) were carried out in a Jasco V660 and V670 UV/VIS spectrometer. The quenching titration and determination of K_{sv} constants of **P5/P6** have been reported previously.⁵

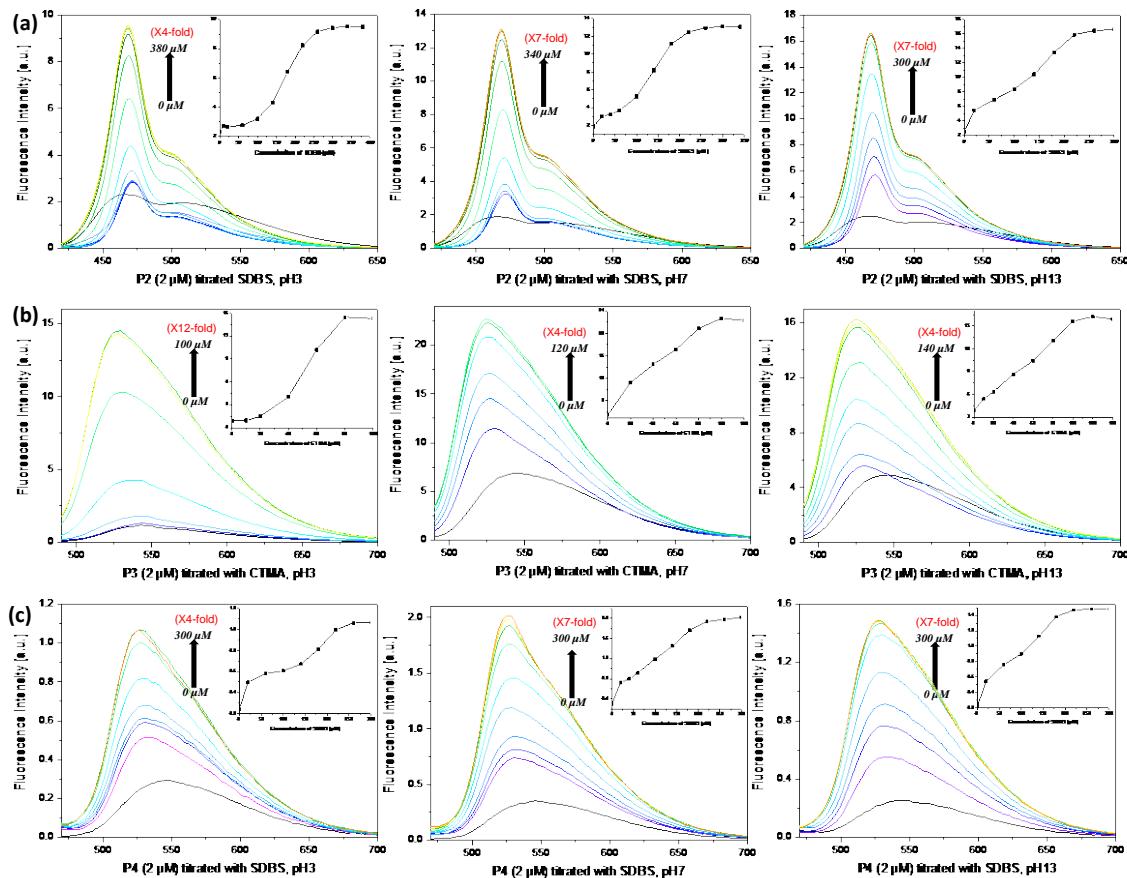


Figure S1. (a) **P2** (2 μ M, black line) with **SDBS**, (b) **P3** (2 μ M, black line) with **CTMA**, (c) **P4** (2 μ M, black line) with **SDBS** at pH 3, pH 7, and pH 13. The inset graph shows the change of I_{Fl} with increasing surfactant concentration.

4. Fluorescence Response Patterns

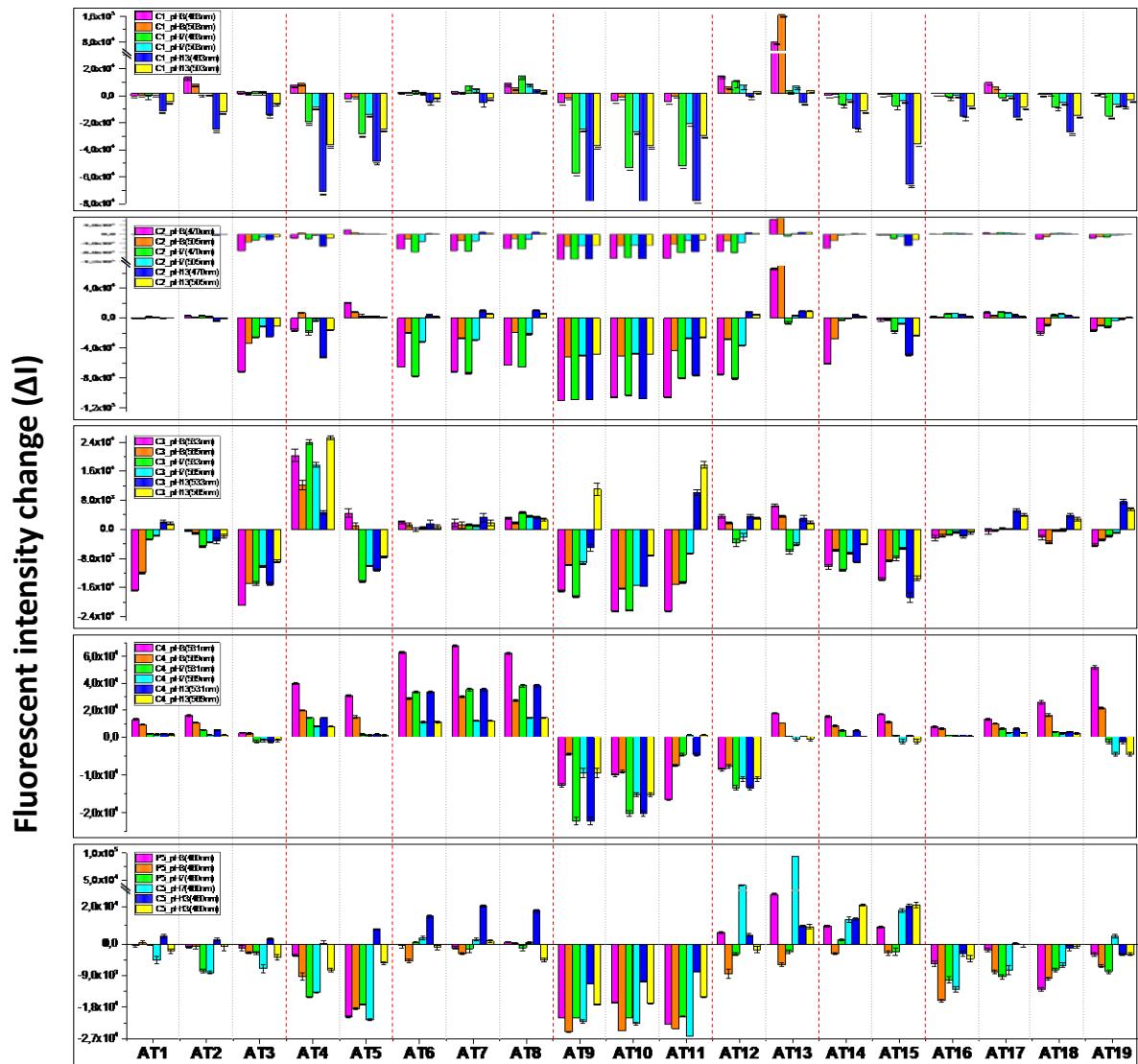


Figure S2. Fluorescence response pattern ΔI obtained by **C1-C4** (2 μM , at pH 3, pH 7 and pH 13, buffered) and **P5, C5** (0.5 μM , at pH 3, pH 7 and pH 13, buffered) treated with antibiotics **AT1-AT19** ($c = 5 \text{ mM}$). Each value is the average of six independent measurements; each error bar shows the standard error of these measurements. The black dotted line shows the type of each antibiotic, red dotted line shows the seven families of antibiotics.

AT19	109	-669	-	16612	-7441	-9020	-4239	-17071	-9301	-11435	-3075	-1100	999	-3948	-3344	-1630	-1103	7663	5358	50916	2032	-826	-4710	-24	-29	60	72	-7	11
AT19	41	-762	16623	-7762	-8622	-5054	-16212	-9324	-12997	-3751	-1441	490	-4703	-2732	-1910	-1038	8423	6079	52965	2163	-898	-3983	-813	206	60	71	-9	11	
Control	-9	4	175	477	905	853	-684	-691	-2154	-194	871	196	-21	28	357	-326	723	-168	350	-295	-241	-77	-138	-309	64	68	41	20	
Control	107	154	-1250	-390	1143	-17	542	161	1223	-143	363	126	-70	118	-209	169	108	340	-652	251	333	134	68	302	64	67	43	20	
Control	-36	1	402	306	969	553	1427	667	-2340	-395	525	-41	203	136	-359	-245	-213	567	581	387	-180	144	-7	6	64	68	42	20	
Control	148	-157	298	157	518	86	-230	-22	-1431	225	35	186	67	136	-338	225	-472	-493	702	-310	-242	-29	-181	-214	66	67	42	20	
Control	-99	10	-676	155	-536	-640	-578	256	-785	-484	-114	-422	202	-148	676	292	449	-360	161	110	51	-23	-339	-103	64	65	42	20	
Control	-94	-2	-77	-279	1363	-1241	-498	-781	382	161	-48	-263	-45	73	429	188	122	-161	-47	-825	-238	137	199	217	64	67	43	20	

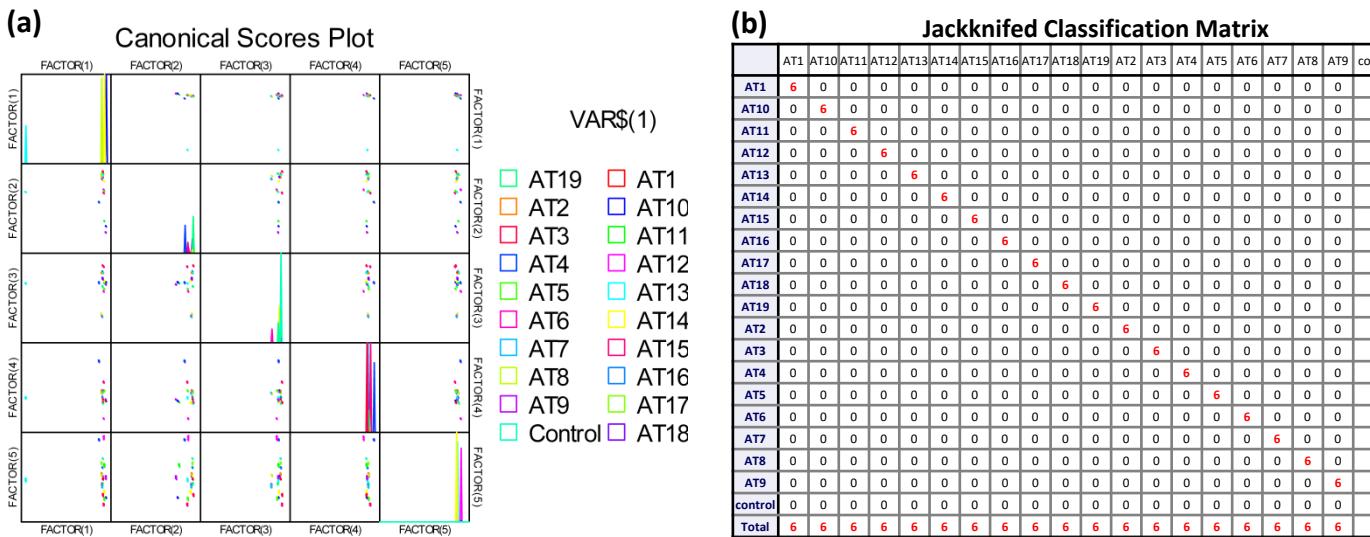


Figure S3. (a) Correlations of canonical fluorescence response patterns from an array of sensor element S1-S24 against 19 antibiotics. The 95% confidence ellipses for the individual acids are also shown. (b) Jackknifed classification matrix showed the 100% correct classification.

Table S3. Training matrix of fluorescence response pattern from an array of sensor element **S25-S30** against 19 antibiotics. LDA was carried out and resulting in 6 factors of the canonical scores and group generation. Jackknifed classification matrix showed the 100% correct classification.

AT6	48.25	40.80	-123.53	-2.91	6.00	-5.68	-21.37	-0.57	16
AT6	49.12	40.29	-122.82	-3.16	5.17	-6.30	-21.33	0.69	16
AT6	48.56	39.63	-123.32	-2.39	5.98	-6.13	-23.87	0.27	16
AT6	48.19	42.53	-121.63	-2.60	4.72	-7.01	-21.33	0.36	16
AT6	48.67	40.34	-124.90	-2.71	8.10	-6.75	-22.13	0.12	16
AT7	50.50	41.62	-126.67	1.16	14.31	-13.39	-2.56	-6.54	17
AT7	49.45	41.84	-128.43	-0.22	11.37	-10.32	-3.58	-4.42	17
AT7	51.38	39.64	-126.31	2.25	11.90	-11.02	-3.51	-6.29	17
AT7	50.81	41.45	-126.55	2.06	11.61	-10.84	-2.60	-5.65	17
AT7	49.91	40.54	-128.98	1.10	12.70	-12.62	-2.74	-6.18	17
AT7	49.73	41.68	-127.75	-1.27	11.60	-11.15	-4.91	-6.12	17
AT8	40.81	56.71	-113.77	2.22	9.09	7.33	-4.06	4.05	18
AT8	42.70	56.16	-114.99	2.15	9.66	8.46	-4.30	4.60	18
AT8	42.75	58.41	-114.40	1.73	7.64	7.64	-3.48	4.32	18
AT8	41.89	57.93	-114.36	1.21	8.53	7.38	-4.69	0.35	18
AT8	43.81	56.83	-114.67	0.82	10.27	7.14	-5.02	2.68	18
AT8	41.40	57.26	-115.71	0.07	9.17	7.16	-4.79	0.37	18
AT9	111.43	-230.15	3.91	17.73	-6.57	5.20	-13.86	-11.41	19
AT9	112.10	-228.42	5.60	19.05	-6.32	3.29	-15.96	-9.92	19
AT9	111.32	-227.81	5.53	18.15	-5.30	1.87	-15.35	-8.04	19
AT9	112.27	-228.41	4.40	17.32	-8.15	3.24	-15.31	-9.40	19
AT9	112.60	-229.61	4.74	18.34	-6.45	3.87	-14.48	-10.78	19
AT9	111.66	-229.66	5.06	18.12	-7.22	4.75	-15.62	-11.42	19
AT10	117.88	-195.94	12.75	24.03	4.84	-23.42	-23.94	12.05	2
AT10	118.35	-196.92	12.53	24.15	4.27	-22.95	-24.28	12.47	2
AT10	118.12	-196.55	12.72	24.45	3.27	-22.86	-24.75	12.02	2
AT10	118.26	-196.72	14.04	24.73	3.99	-22.87	-24.54	12.30	2
AT10	116.98	-196.79	13.72	24.66	4.49	-23.05	-24.82	12.43	2
AT10	117.53	-195.85	11.95	23.91	3.35	-22.74	-24.99	12.49	2
AT11	83.26	-181.26	5.41	-15.53	-47.73	-5.50	13.83	-10.51	3
AT11	82.61	-181.88	4.57	-15.63	-48.04	-4.58	14.37	-10.88	3
AT11	82.58	-181.82	4.76	-16.33	-47.69	-5.57	15.12	-10.41	3
AT11	82.94	-181.37	4.11	-14.57	-48.10	-5.73	12.55	-9.98	3
AT11	82.69	-181.74	5.33	-15.10	-49.22	-3.32	14.75	-11.73	3
AT11	82.08	-181.70	4.25	-15.13	-48.41	-4.63	14.08	-12.21	3
AT12	89.17	13.94	-17.52	106.37	-25.11	62.73	20.43	20.37	4
AT12	88.10	13.98	-19.32	108.35	-25.81	62.59	16.91	19.50	4
AT12	87.82	14.36	-19.44	107.67	-25.25	60.45	20.32	20.19	4
AT12	88.92	12.48	-19.06	107.67	-25.55	61.51	19.25	19.46	4
AT12	88.39	13.28	-17.40	105.26	-27.81	60.56	18.51	19.65	4
AT12	89.68	11.22	-17.13	106.95	-25.79	58.66	16.44	20.41	4
AT13	-1319.48	-14.05	5.63	7.78	-3.28	-3.72	-1.86	0.59	5
AT13	-1319.91	-14.83	5.60	8.96	-4.19	-6.78	1.01	0.73	5
AT13	-1314.87	-14.36	6.18	7.04	-2.06	-4.50	-0.11	-1.83	5
AT13	-1320.85	-14.37	5.49	7.97	-3.02	-3.50	-0.03	0.00	5
AT13	-1321.95	-14.49	6.54	9.48	-4.21	-6.75	-0.81	-0.49	5
AT13	-1318.72	-14.97	4.79	8.16	-3.92	-6.72	0.54	-1.04	5
AT14	93.85	45.56	12.37	42.21	11.36	-38.56	51.78	-21.35	6
AT14	93.34	47.20	9.28	41.76	9.25	-37.18	53.34	-22.05	6
AT14	93.58	48.42	12.37	45.06	7.92	-36.45	53.13	-23.26	6
AT14	95.33	45.62	11.51	41.82	8.55	-36.79	52.55	-19.63	6
AT14	94.84	47.39	9.28	42.78	9.14	-37.26	53.75	-20.99	6
AT14	95.25	47.27	10.21	41.85	8.05	-37.12	52.41	-20.94	6
AT15	82.37	-7.79	61.20	38.74	98.84	-24.21	1.97	-2.86	7
AT15	80.07	-5.31	61.51	38.31	100.51	-23.82	-0.75	-1.91	7
AT15	80.70	-6.99	59.68	39.96	100.48	-26.88	1.50	-3.43	7
AT15	80.58	-7.64	60.66	37.60	98.55	-24.18	-0.51	-1.43	7
AT15	81.14	-6.03	61.57	37.82	102.39	-25.90	2.43	-3.31	7
AT15	81.66	-9.53	60.13	38.87	100.40	-25.42	0.81	-2.36	7
AT16	70.44	65.59	50.62	-23.04	-32.60	8.74	-5.10	0.84	8
AT16	68.03	64.47	51.63	-25.08	-33.61	7.82	-3.73	-1.01	8
AT16	68.24	64.99	50.19	-26.38	-32.37	7.71	-3.55	1.47	8

6. Principal Component Analysis (PCA)

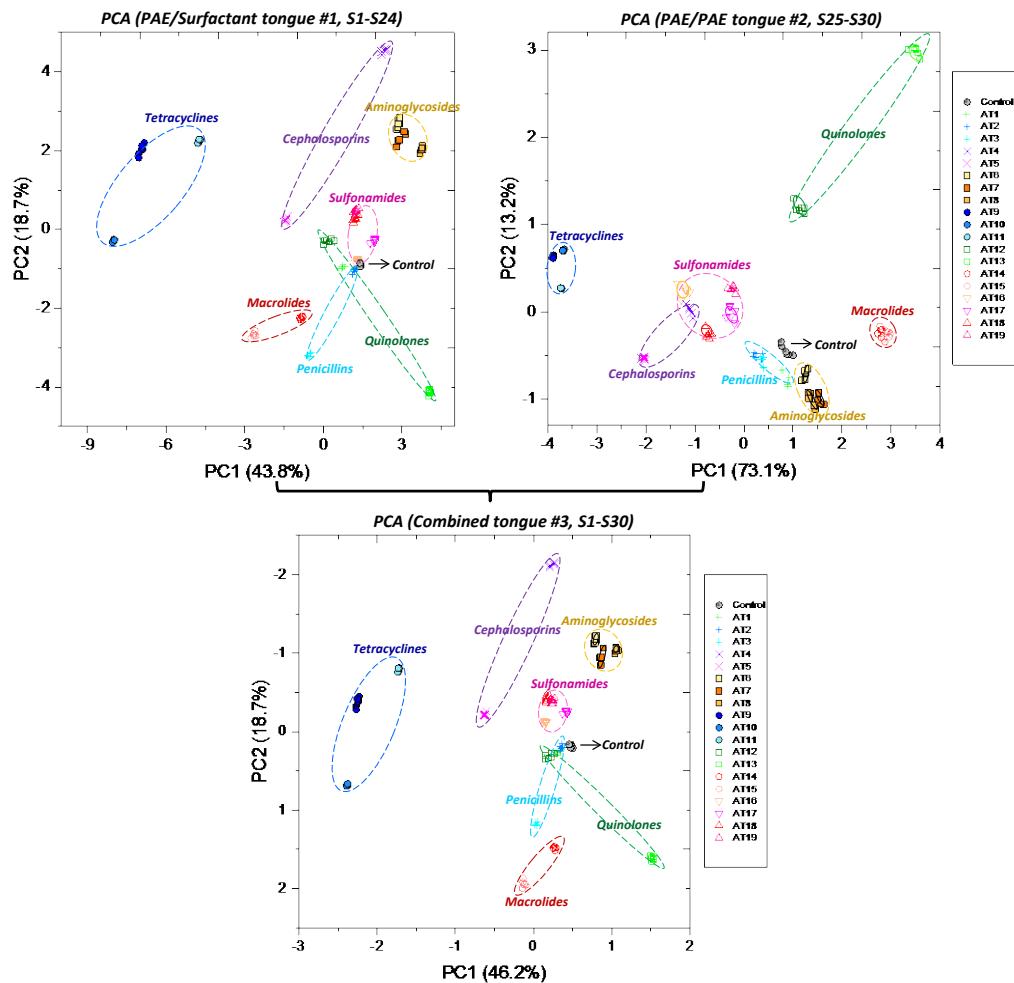


Figure S7. 2D PCA plot for the first two principal component obtained with an array of **S1-S24** (left, PAE/surfactant tongue #1), **S25-S30** (right, PAE/PAE tongue #2) and combined tongue of **S1-S30** (bottom, tongue #3) treated with antibiotics **AT1-AT19** ($c = 5 \text{ mM}$) with 95% confidence ellipses. Each point represents the response pattern for a single antibiotic to the array. Each antibiotic was shown with their individual shape (triangle, square, circle etc.) and similar color.

7. Optimization Process

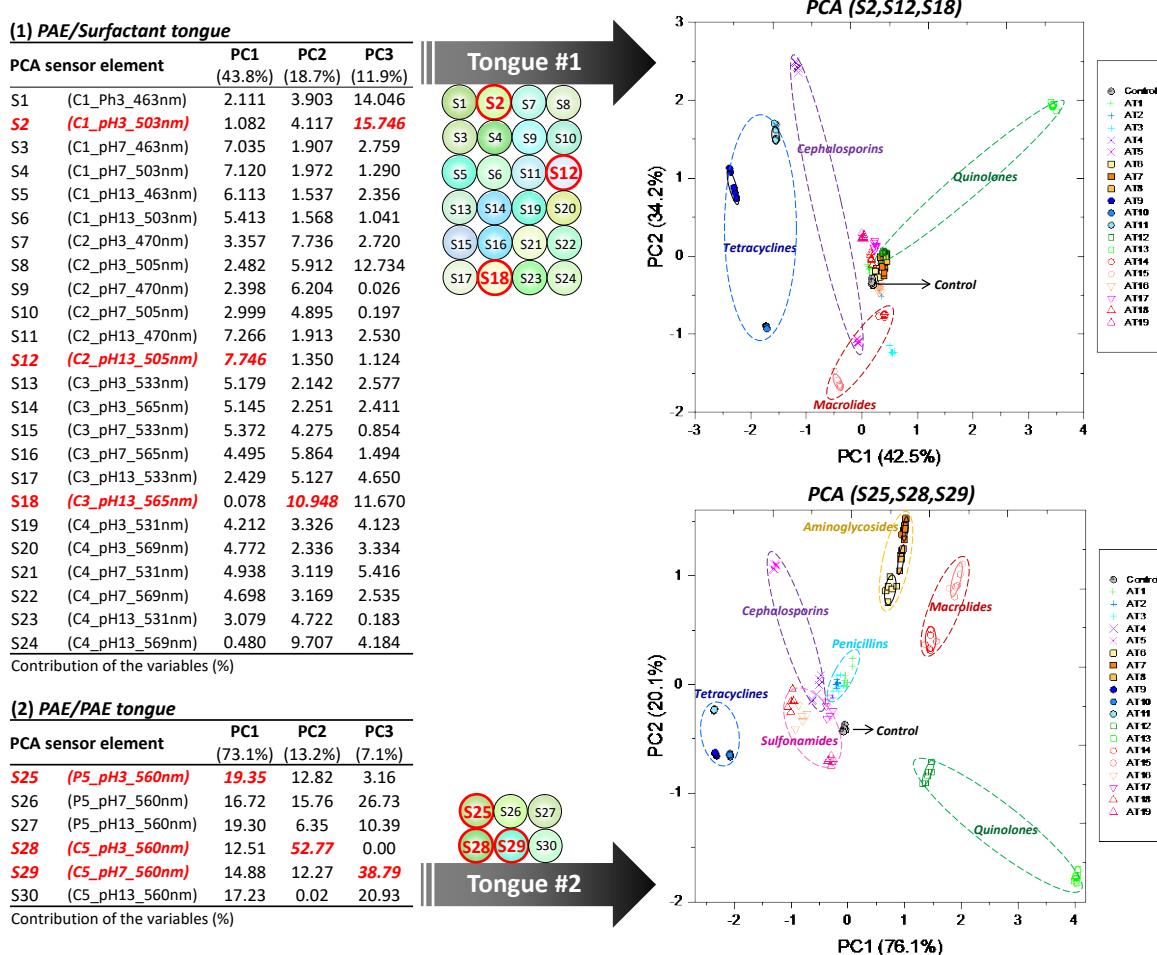


Figure S8. Optimization and selection of the best three sensing elements from the PAE/surfactant tongue (**S1-S24**) and PAE/PAE tongue (**S25-S30**) based on contribution of the variables of PCA (left). The resulting PCA plots are shown (right).

8. Others

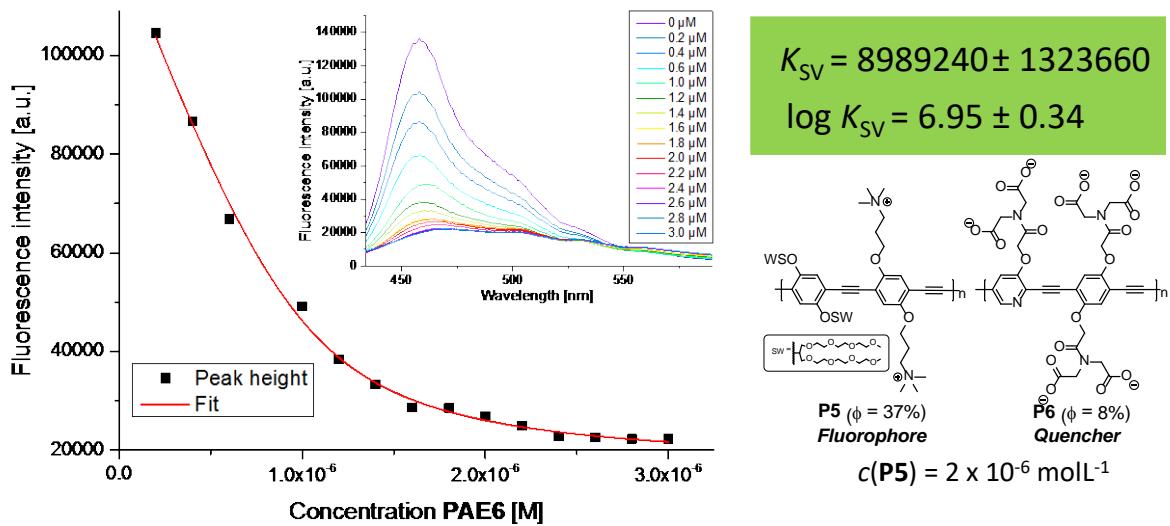


Figure S9. Complex titrations and determination of K_{SV} constants with modified Ster-Volmer equation. The titrations were performed in $\text{KH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$ buffered solution ($\text{pH} = 7$).

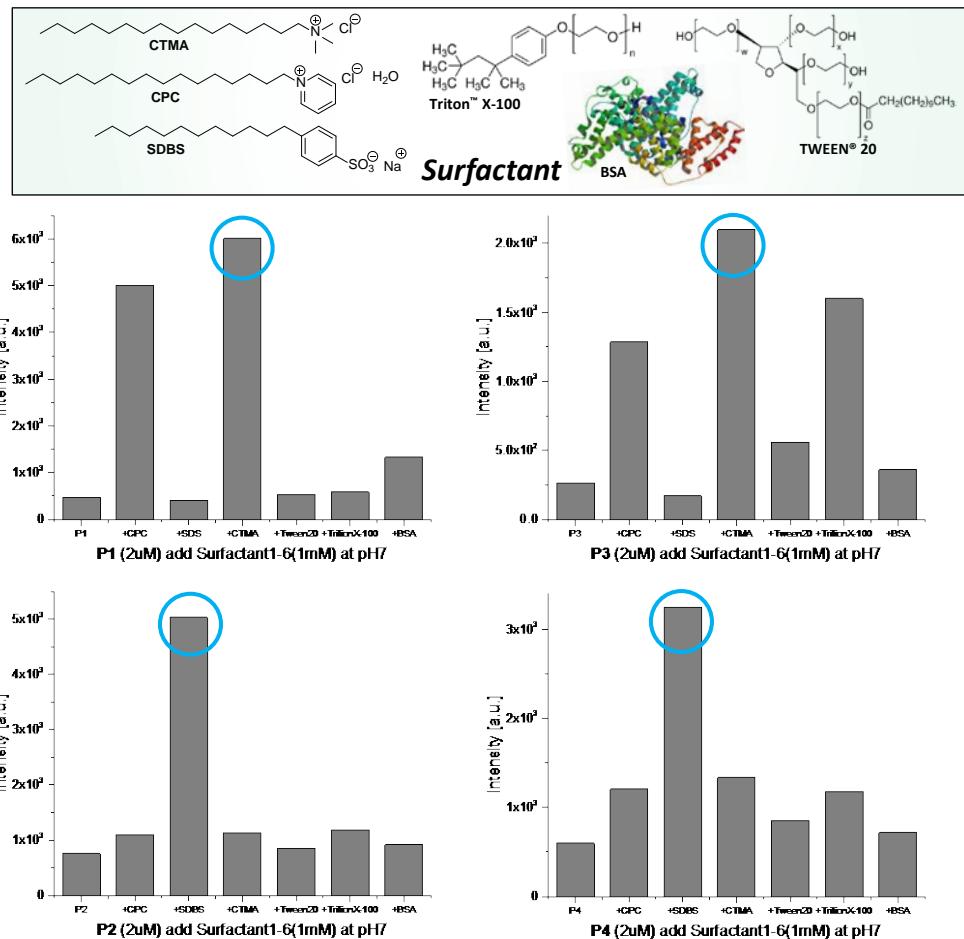


Figure S10. Screening process of various surfactants.

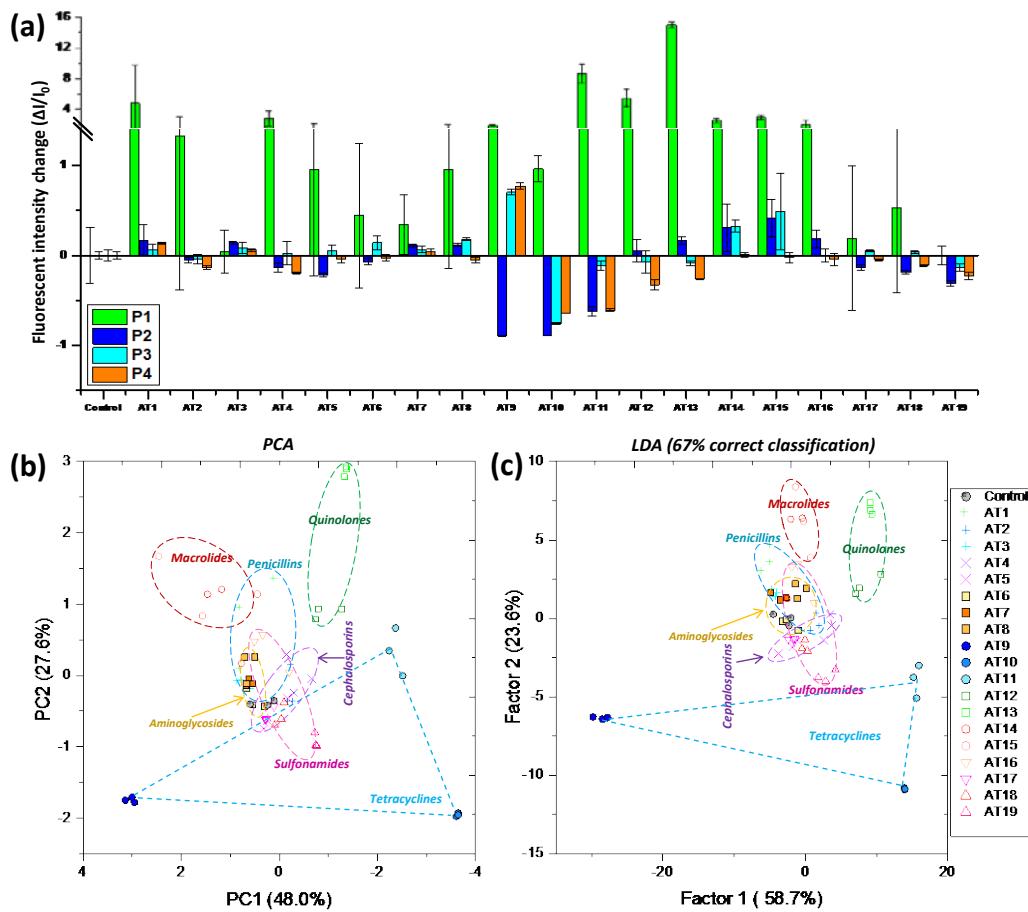
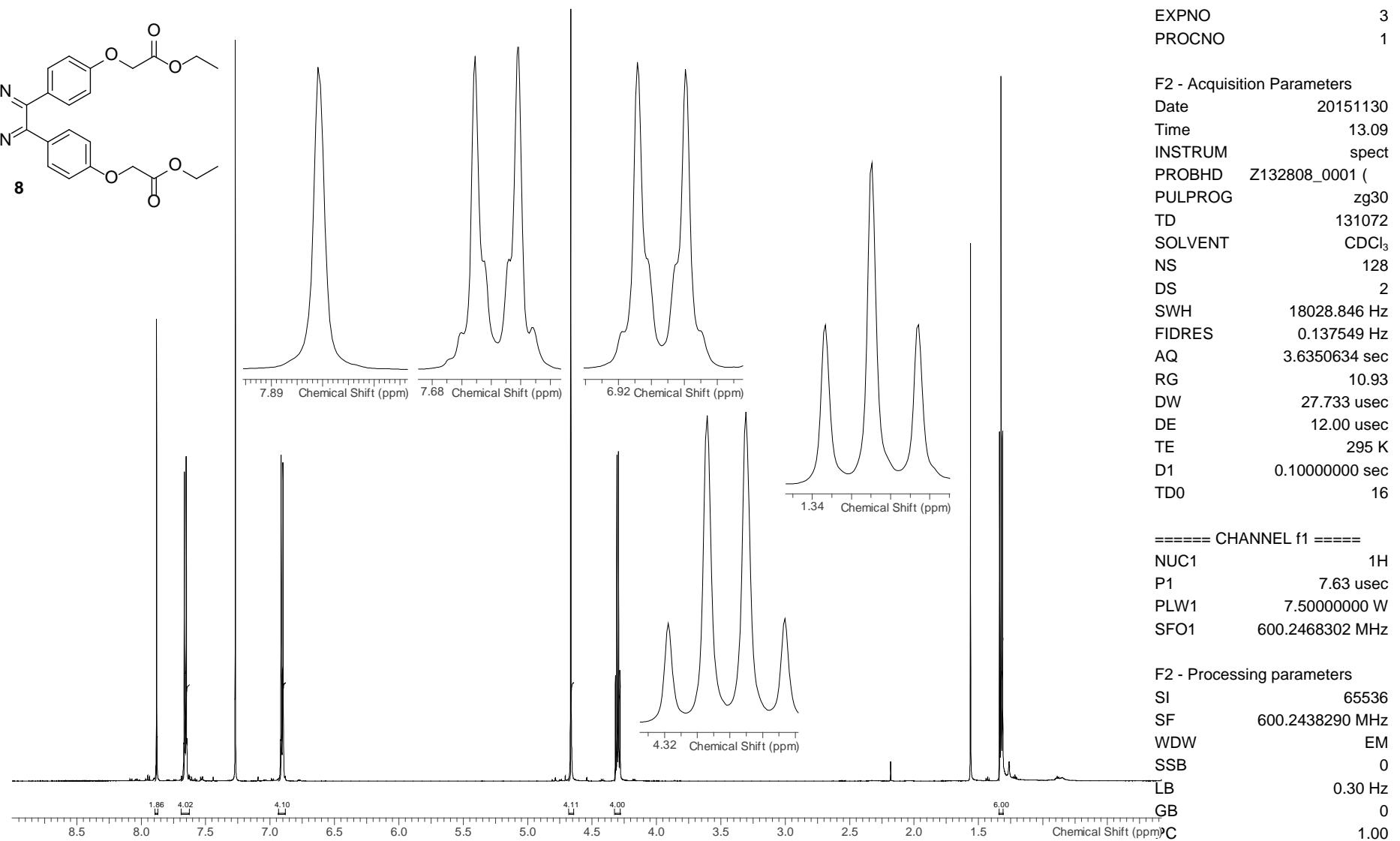
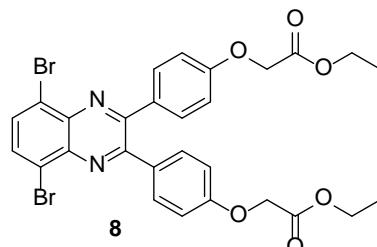
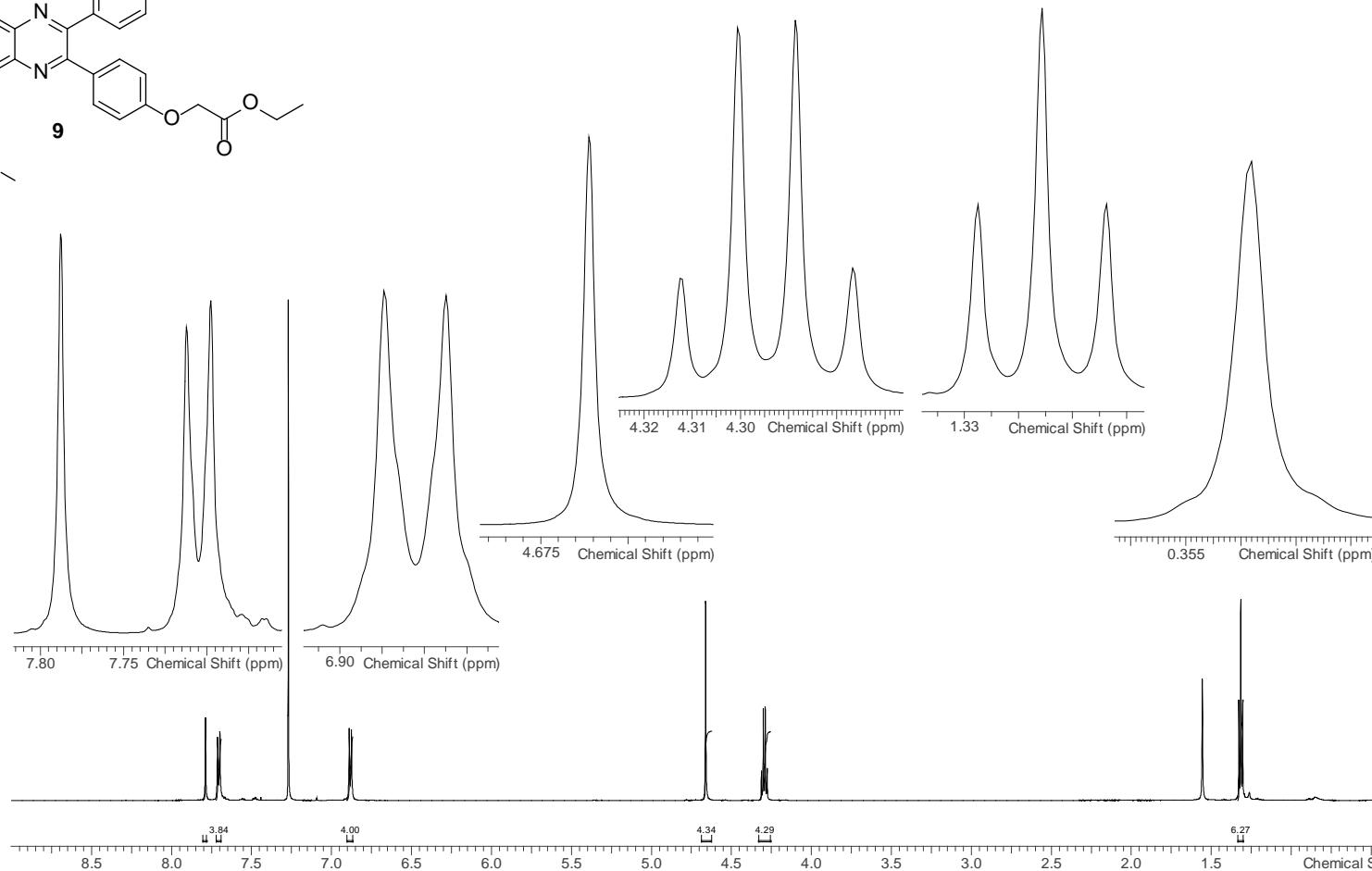
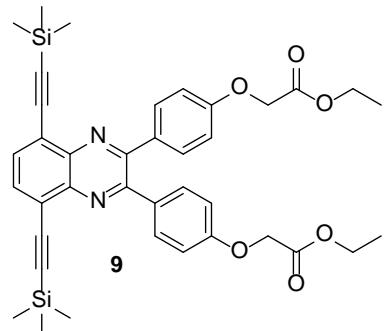


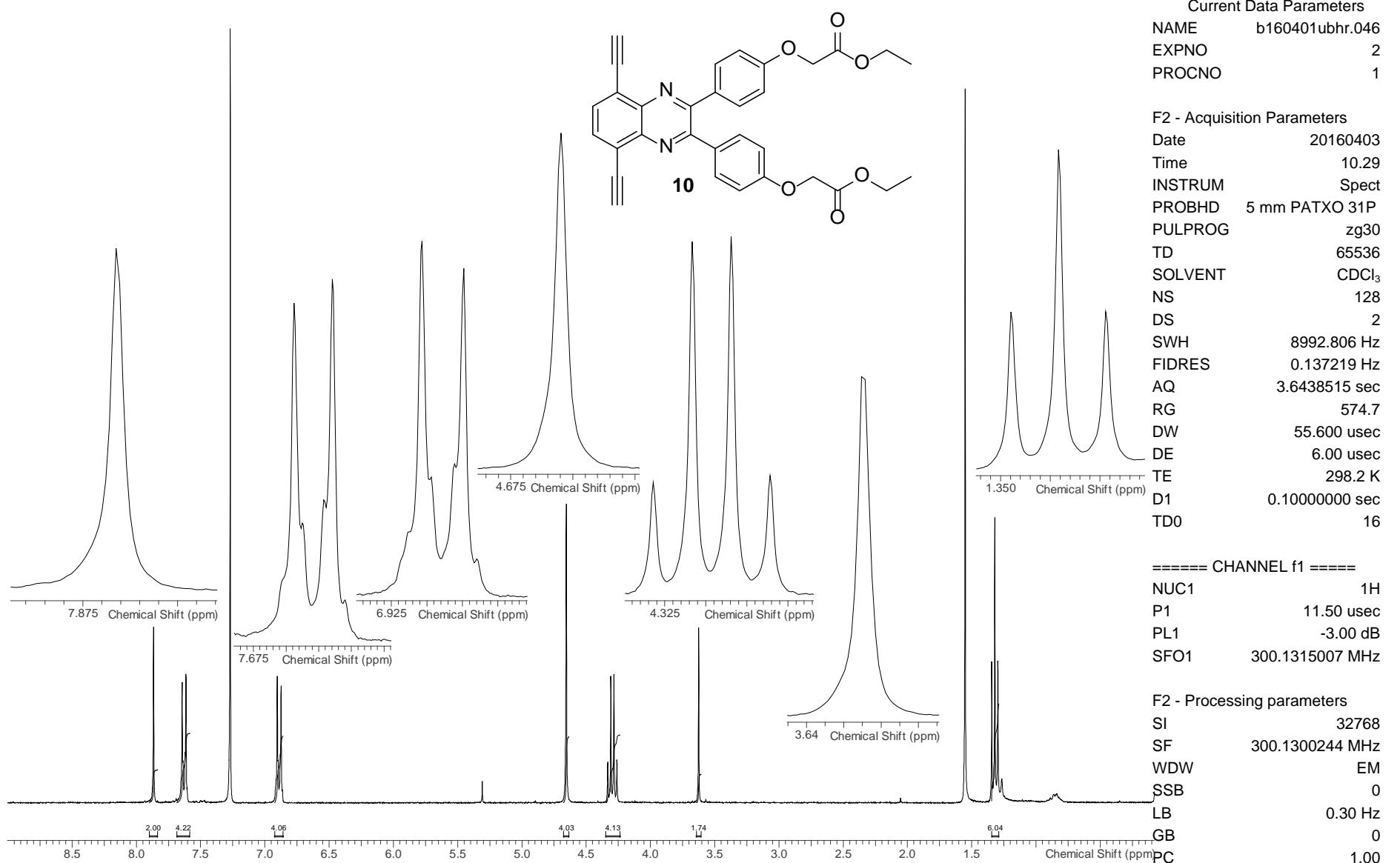
Figure S11. (a) Fluorescence intensity change $\Delta I/I_0$ obtained by weakly fluorescent **P1-P4** (2 μM , at pH 7, buffered) treated with antibiotics **AT1-AT19** ($c = 5 \text{ mM}$). Each value is the average of three independent measurements; each error bar shows the standard error (SD) of these measurements. (b) PCA plot and (c) LDA plot from first the first two factors obtained with **P1-P4** (2 μM , at pH7, buffered) treated with antibiotics **AT1-AT19** ($c = 5 \text{ mM}$). Cross-validated LDA showed 67% correct accuracy for all antibiotics.

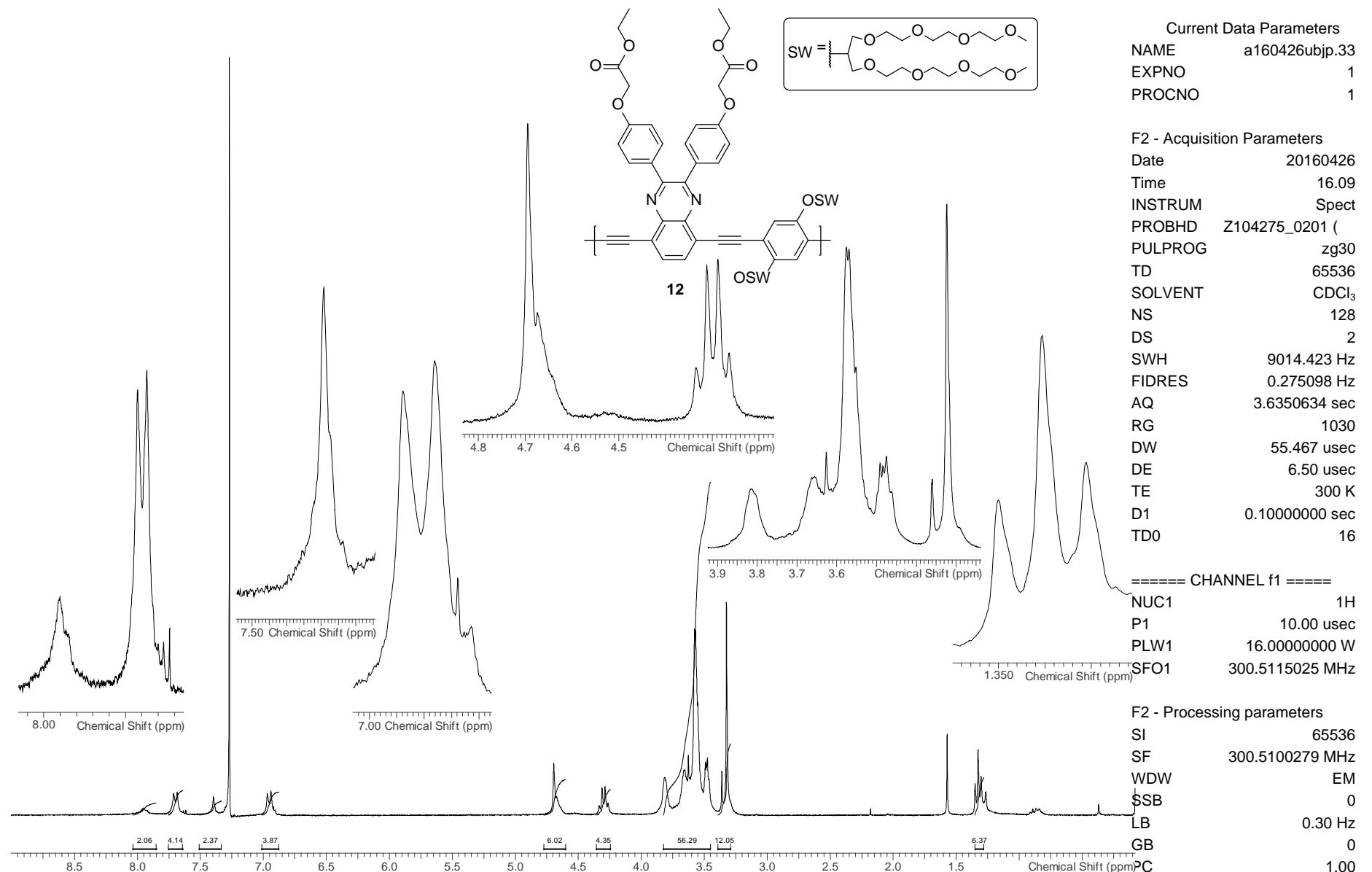
9. $^1\text{H-NMR}$ Spectra

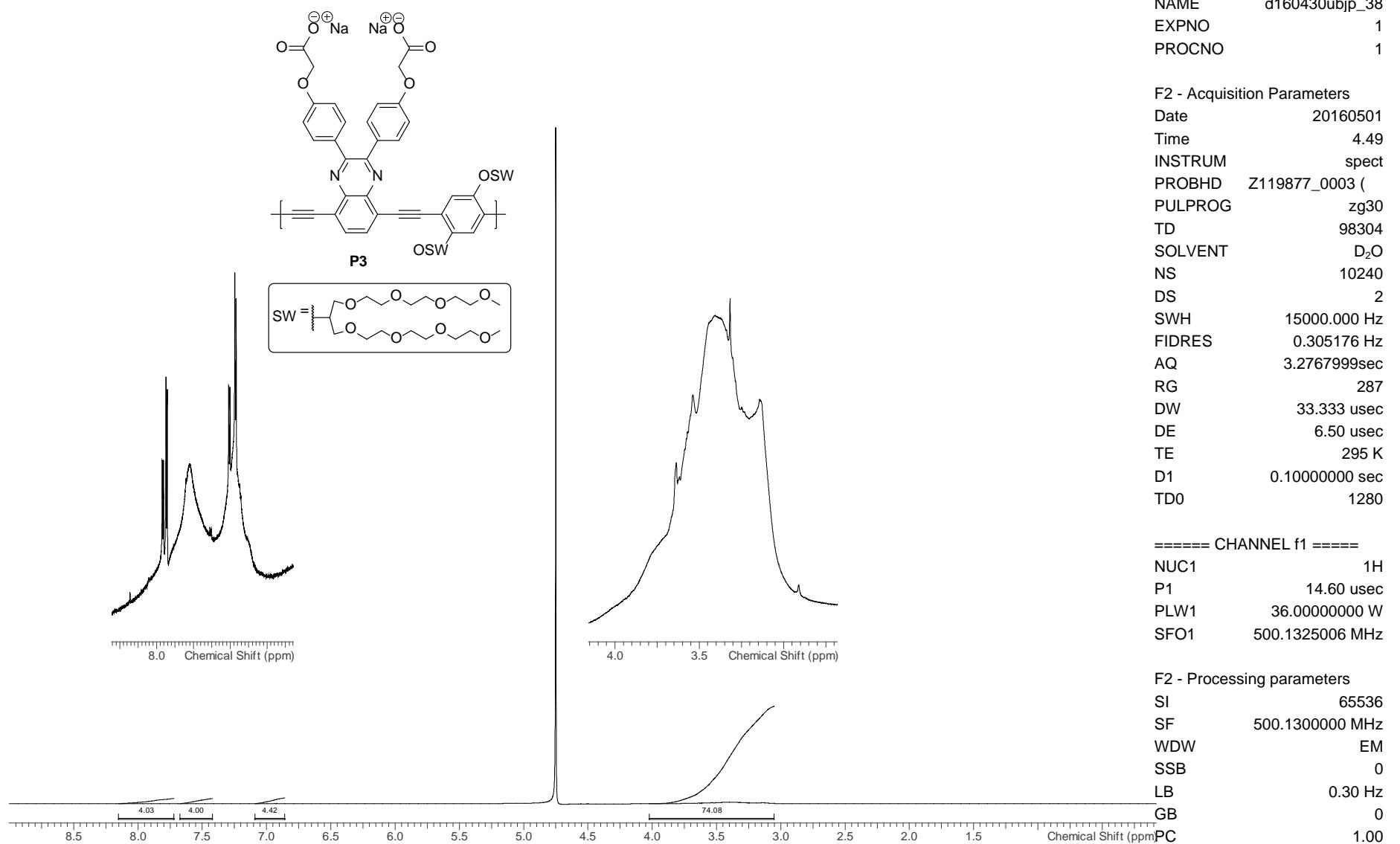


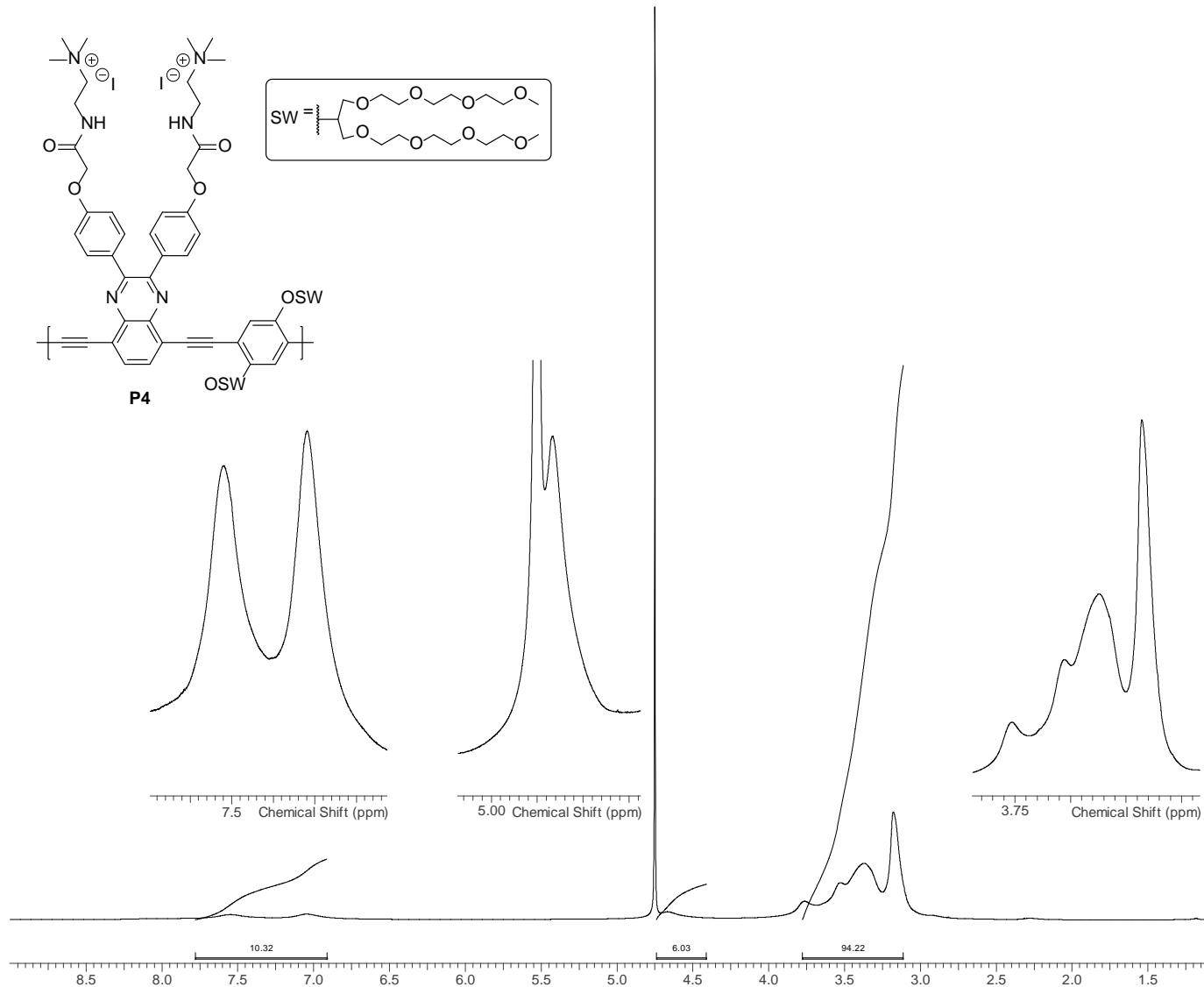
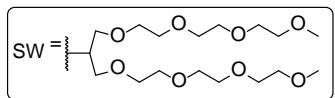
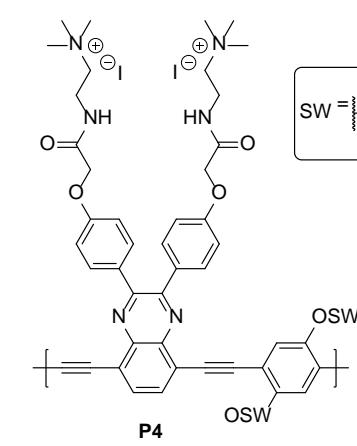


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10. Supplemental References

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