

Two novel supramolecular metallogels constructed by platinum(II) coordination and pillar[5]arene-based host–guest interactions

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1. Materials and methods

All reagents were commercially available and used as supplied without further purification. Compounds **2**,^{S1} **3**,^{S2} **4**,^{S3} and **6**^{S4} were synthesized by published literature procedures. NMR spectra were recorded with a Bruker Avance DMX 500 spectrophotometer with use of the deuterated solvent as the lock and the residual solvent or TMS as the internal reference. Low-resolution electrospray ionization (LRESI) mass spectra were obtained on a Bruker Esquire 3000 plus mass spectrometer (Bruker-Franzen Analytik GmbH Bremen, Germany) equipped with an ESI interface and an ion trap analyzer. High-resolution electrospray ionization mass spectra (HRESI-MS) were obtained on a Bruker 7-Tesla FT-ICR mass spectrometer equipped with an electrospray source (Billerica, MA, USA). MALDI-TOF MS was performed with a Bruker UltrafleXtreme instrument. Dynamic light scattering (DLS) was carried out on a Malvern Nanosizer S instrument at room temperature. Scanning electron microscopy (SEM) investigations were carried out on a JEOL 6390LV instrument. Rheological behaviors of gels were analyzed by using an ARG-2 rheometer (TA Instruments, USA). The disc-shaped gels with thickness of 1 mm and diameter of 20 mm were adhere to the plates and surrounded by silicone oil. Frequency sweeps were performed to the samples with strain amplitude of 0.05%.

2. Synthetic route to compound **1**

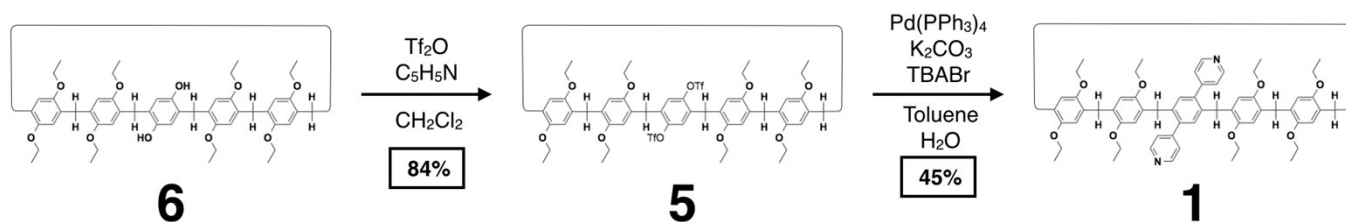


Figure S1. Synthetic route to compound **1**.

Trifluoromethanesulfonic anhydride (2 mL) was added dropwise to a mixture of **6** (500 mg, 0.69 mmol) and C₅H₅N (dry, 1 mL) in CH₂Cl₂ (dry, 20 mL), cooled to 0 °C and then allowed to stir at room temperature for 12 h. The reaction mixture was concentrated under vacuum and subjected to silica gel chromatography (1:1 hexane/CH₂Cl₂) to give **5** as a white powder (520 mg, 84%). The ¹H NMR spectrum of **5** is shown in Figure S2. ¹H NMR (500 MHz, CDCl₃, 298 K) δ = 7.38 (1H, s), 6.79 (1H, s), 6.76 (1H, s), 6.75 (1H, s), 6.67 (1H, s), 3.95–3.79 (14H, m), 1.42–1.22 (12 H, m). The ¹³C NMR spectrum of **5** is shown in Figure S3. ¹³C NMR (125 MHz, CDCl₃, 298 K) δ = 148.88, 148.77, 148.75, 148.54, 145.05, 133.11, 128.96, 127.77, 127.02, 123.77, 123.19, 119.13, 115.95, 113.96, 113.66, 113.16, 62.81, 62.77, 62.36, 52.22, 29.64, 28.59, 28.12, 14.12, 14.04, 13.53. LRESIMS is shown in Figure S4: *m/z* 1116.4 [M + NH₄⁺]⁺. HRESIMS: *m/z* calcd for [M + H]⁺ C₅₃H₆₁O₁₄F₆S₂⁺, 1099.34069; found 1099.34181; error 1 ppm.

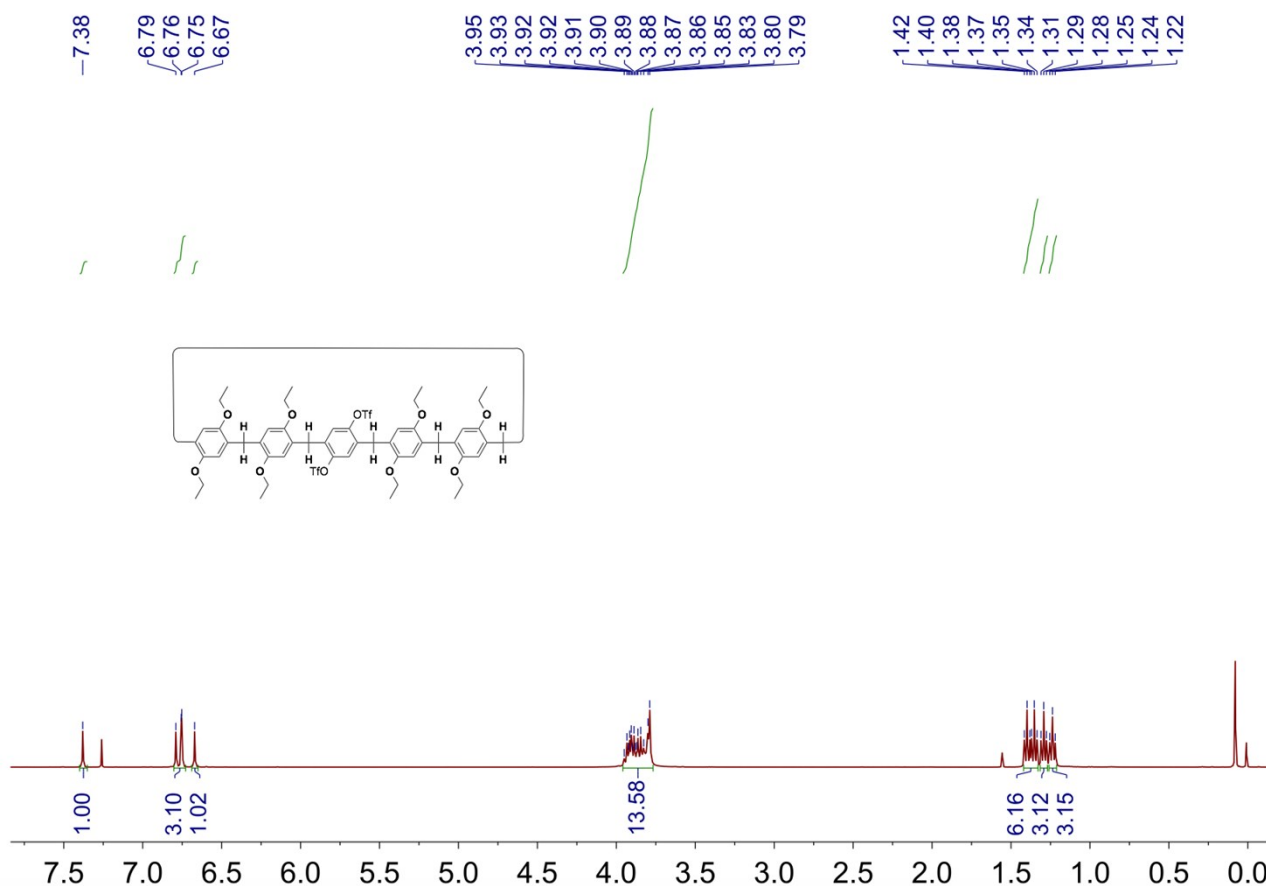


Figure S2. ¹H NMR spectrum (500 MHz, CDCl₃, 298 K) of **5**.

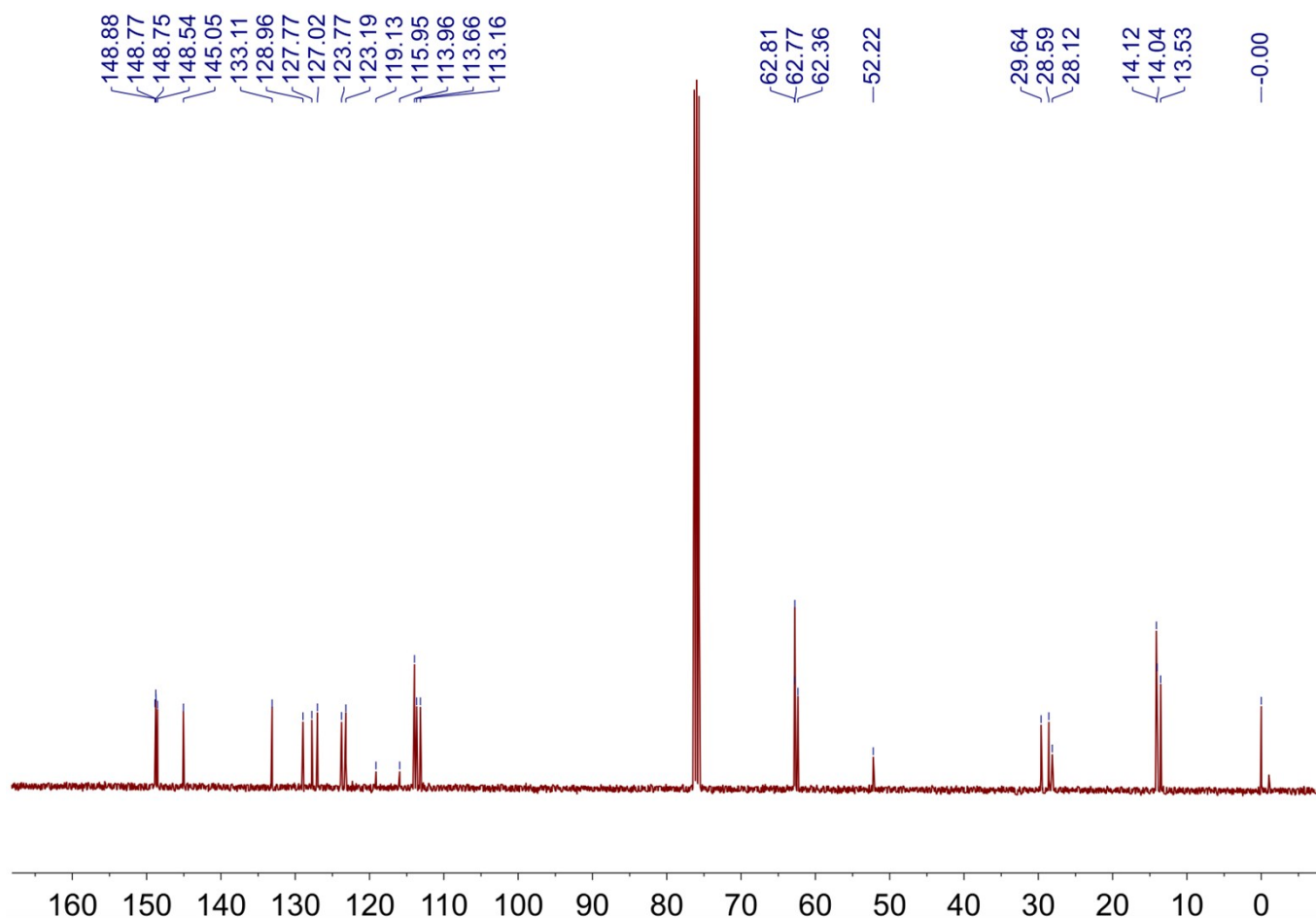


Figure S3. ^{13}C NMR spectrum (125 MHz, CDCl_3 , 298 K) of **5**.

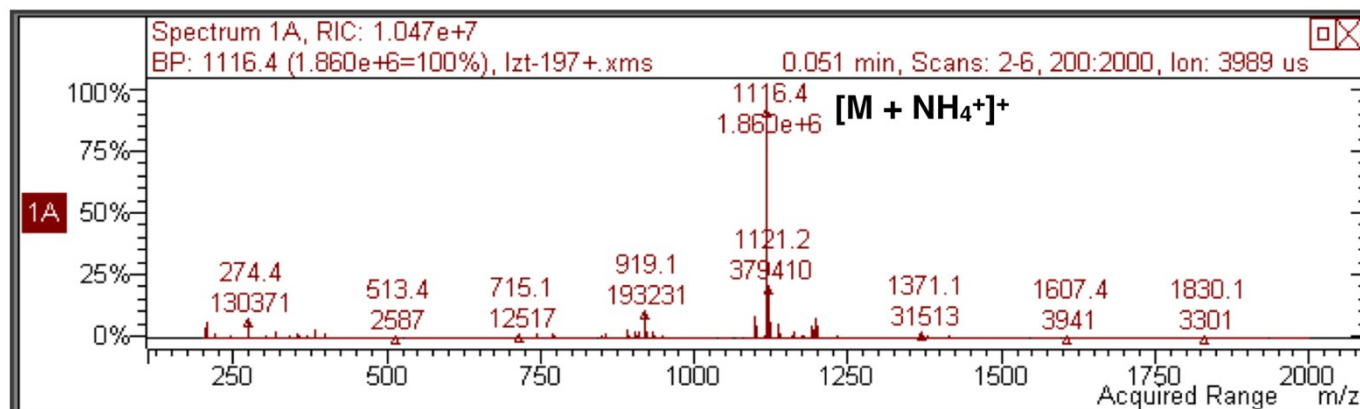


Figure S4. LRESIMS spectrum of **5**.

$\text{Pd}(\text{PPh}_3)_4$ (266 mg, 0.230 mmol) was added to a mixture of **5** (1.04 g, 1.01 mmol), 4-pyridinyl boronic acid (738 mg, 6.00 mmol), and K_2CO_3 (2.49 g, 1.80 mmol) in 80.0 mL of a mix solvent (Toluene/ H_2O , 3:1 v/v). The mixture was stirred at 100 °C for 24 h. After cooling to room temperature, the excess solvent was removed on a rotary evaporator at reduced pressure. After adding 200 mL of CH_2Cl_2 , the solution was washed with H_2O (2 \times 100 mL), brine (100 mL), and dried (Na_2SO_4) then concentrated under vacuum, and finally subjected to silica gel chromatography ($\text{CH}_2\text{Cl}_2/\text{MeOH}$, 100:1 v/v) to give **1** (0.37 g,

45%). The ^1H NMR spectrum of **5** is shown in Figure S5. ^1H NMR (500 MHz, CDCl_3 , 298 K) δ = 8.45 (4H, s), 6.89 (6H, s), 6.86 (2H, s), 6.74 (2H, s), 6.53 (2H, s), 5.95 (2H, s), 3.95–3.56 (26H, m), 1.46–1.43 (6H, t, J = 7.5 Hz), 1.26–1.24 (6H, t, J = 5.0 Hz), 1.17–1.15 (6H, t, J = 5.0 Hz), 0.97–0.94 (6H, t, J = 7.5 Hz). The ^{13}C NMR spectrum of **1** is shown in Figure S6. ^{13}C NMR (125 MHz, CDCl_3 , 298 K) δ = 150.02, 149.99, 149.69, 149.52, 149.28, 138.59, 136.91, 131.45, 129.83, 128.98, 128.64, 126.81, 124.24, 115.22, 115.16, 114.97, 114.58, 64.12, 64.05, 63.23, 33.10, 30.96, 29.95, 29.61, 15.34, 15.19, 14.86, 14.56. MOLDI-TOF is shown in Figure S7: m/z 957.120 $[\text{M} + \text{H}]^+$.

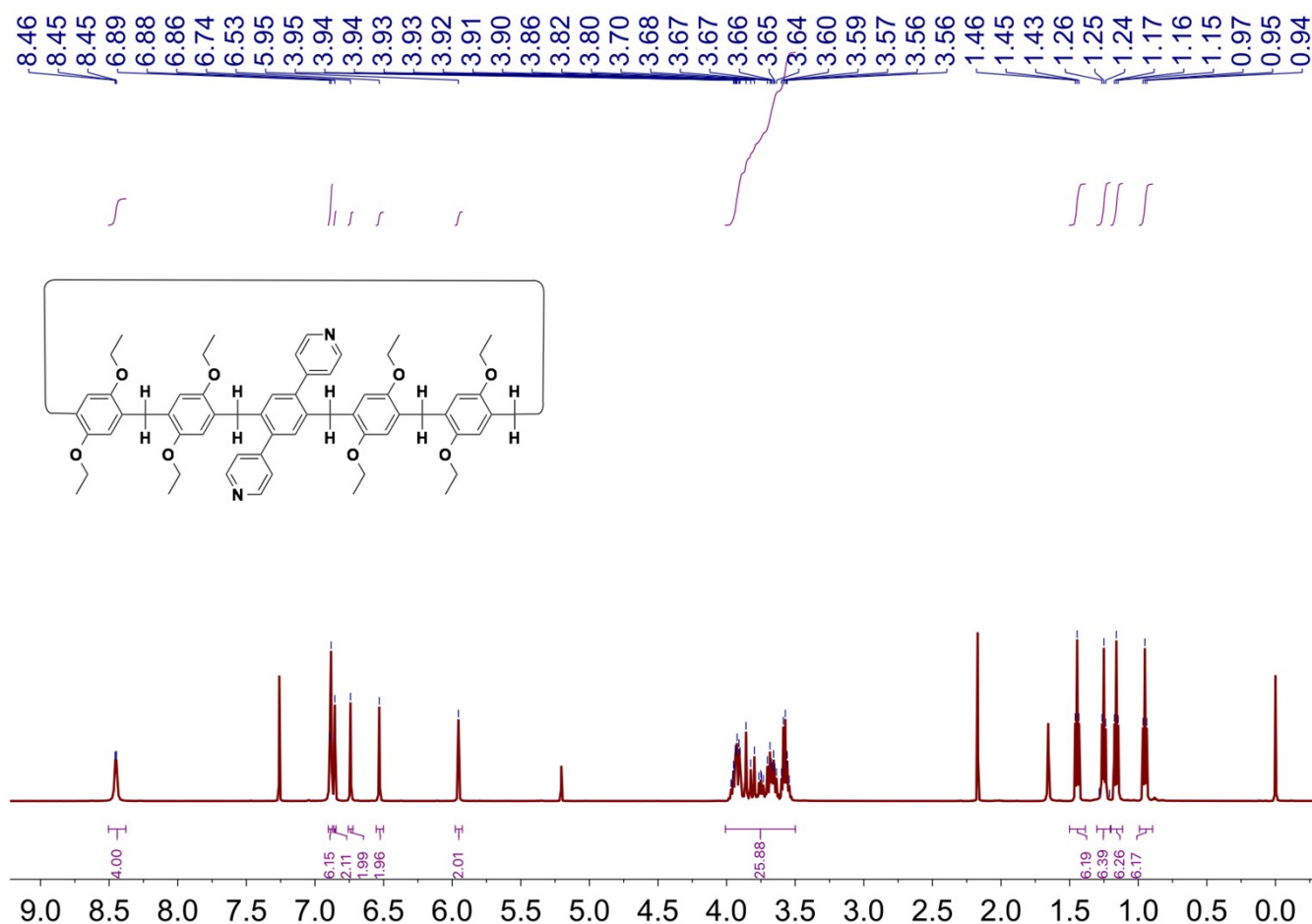


Figure S5. ^1H NMR spectrum (500 MHz, CDCl_3 , 298 K) of **1**.

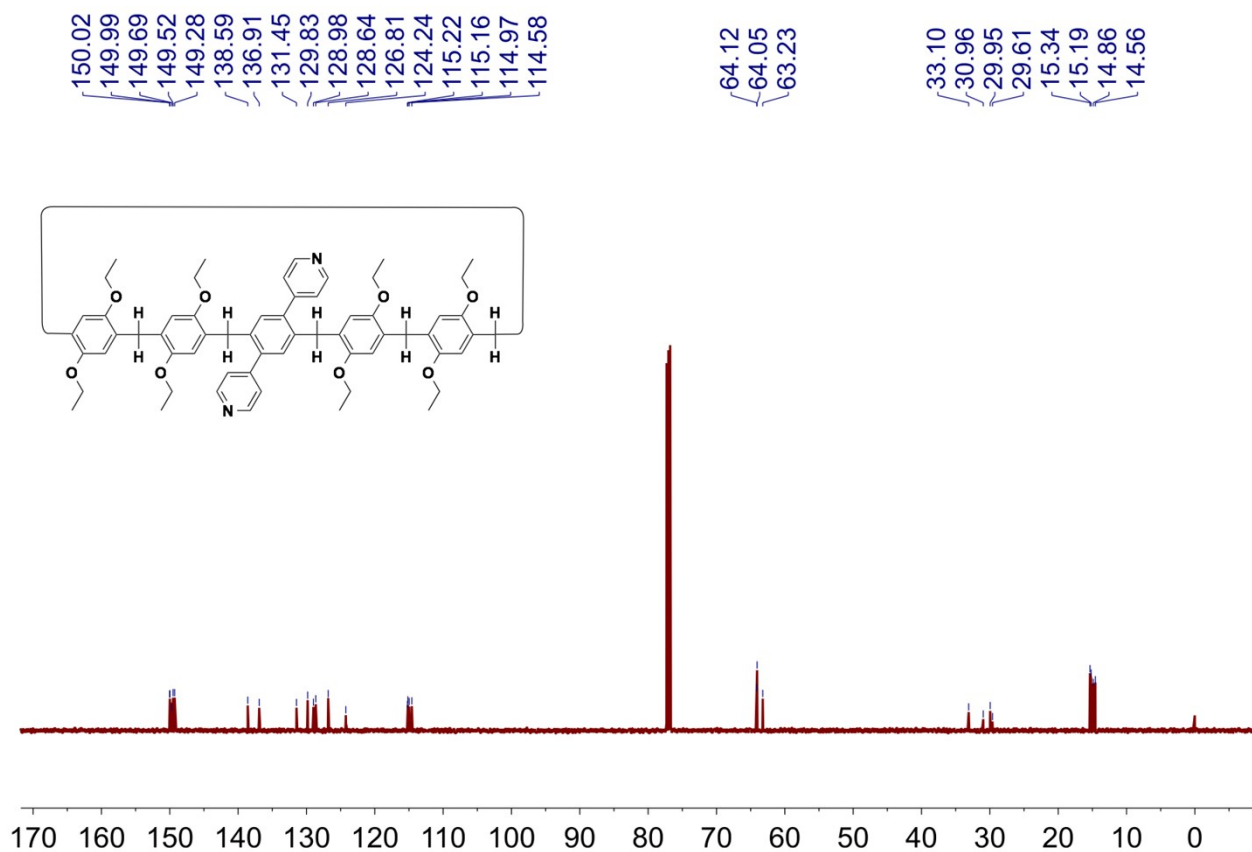


Figure S6. ^{13}C NMR spectrum (125 MHz, CDCl_3 , 298 K) of **1**.

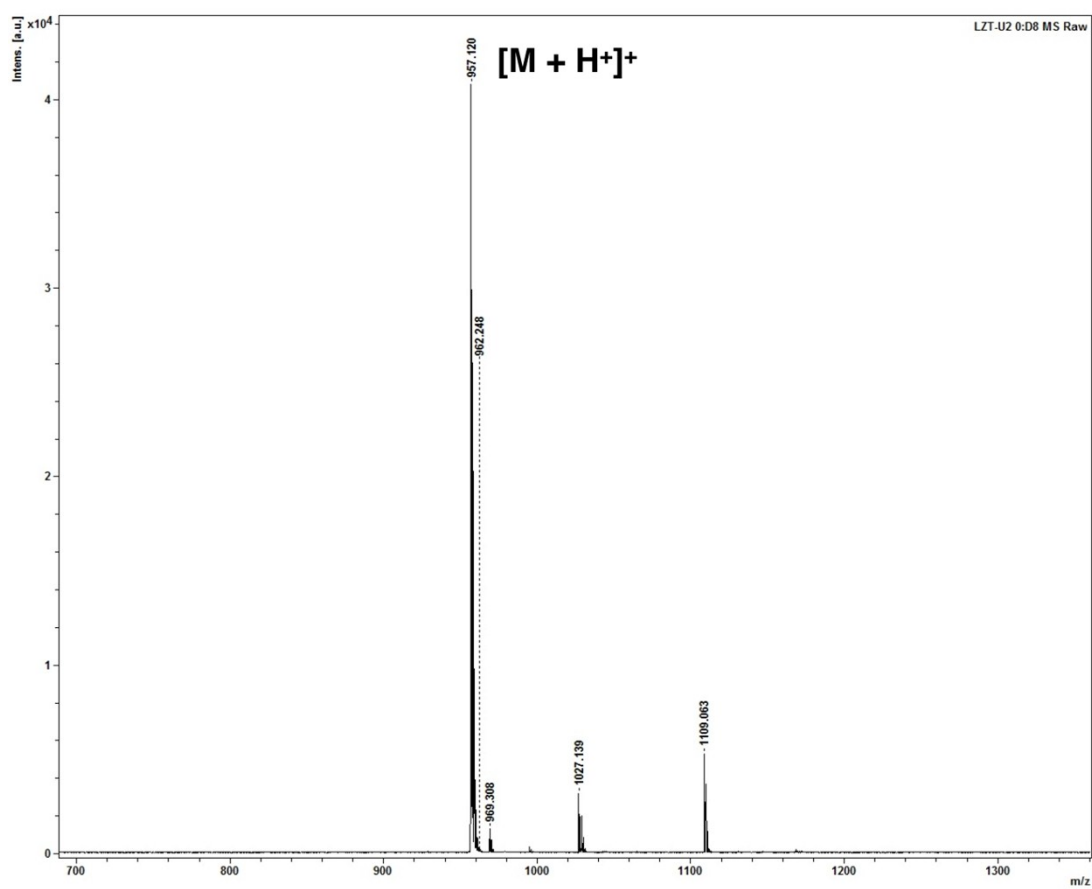


Figure S7. MOLDI-TOF spectrum of **1**.

3. $^{31}\text{P}\{^1\text{H}\}$ NMR spectra of **3** and assemblies

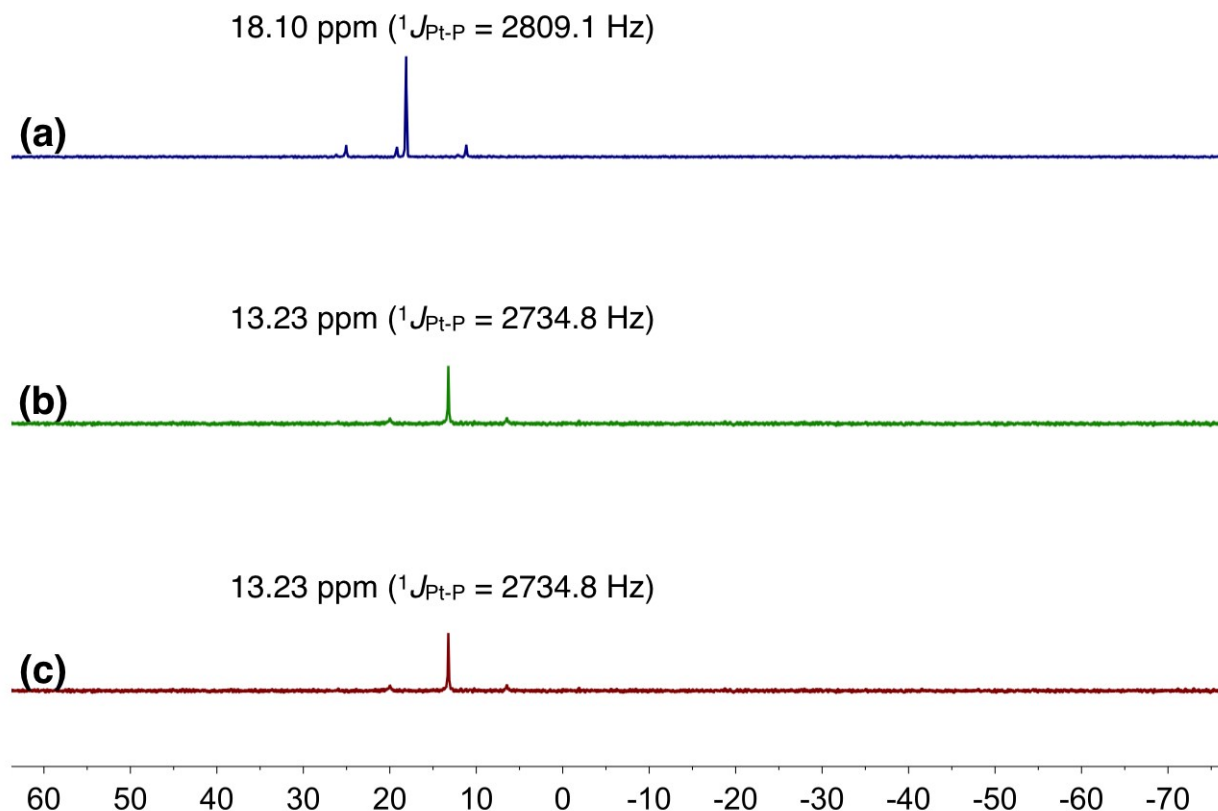


Figure S8. Partial $^{31}\text{P}\{^1\text{H}\}$ NMR spectra (202.3 MHz, acetone- d_6 , 298 K): (a) **3**; (b) **1** + **3**; (c) **1** + **3** + **2**. $c = 5.00$ mM.

4. ^1H NMR spectra of **1**, **2**, and **1** + **2**

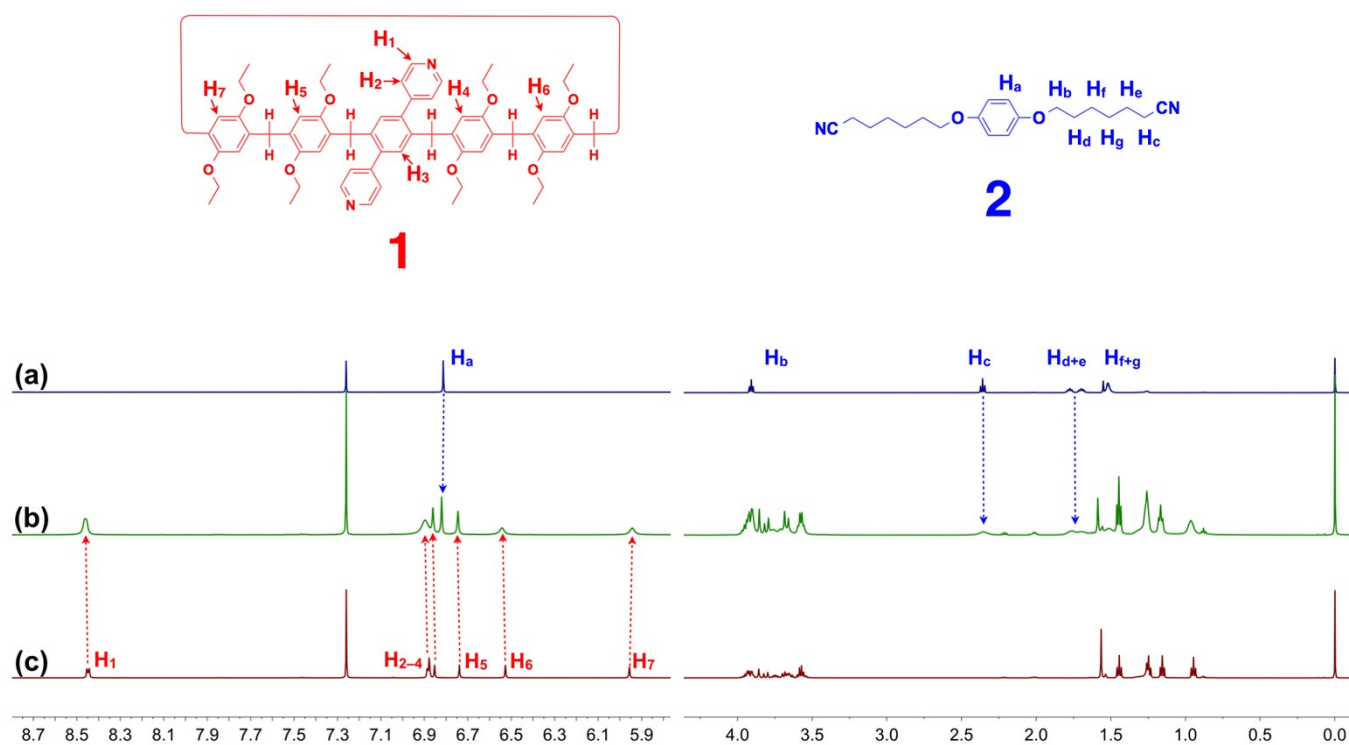


Figure S9. Partial ^1H NMR spectra (500 MHz, CDCl_3 , 298 K): (a) **2**; (b) **1** + **2**; (c) **1**. $c = 5.00$ mM.

5. ^1H and $^{31}\text{P}\{^1\text{H}\}$ NMR spectra of **4** and assemblies

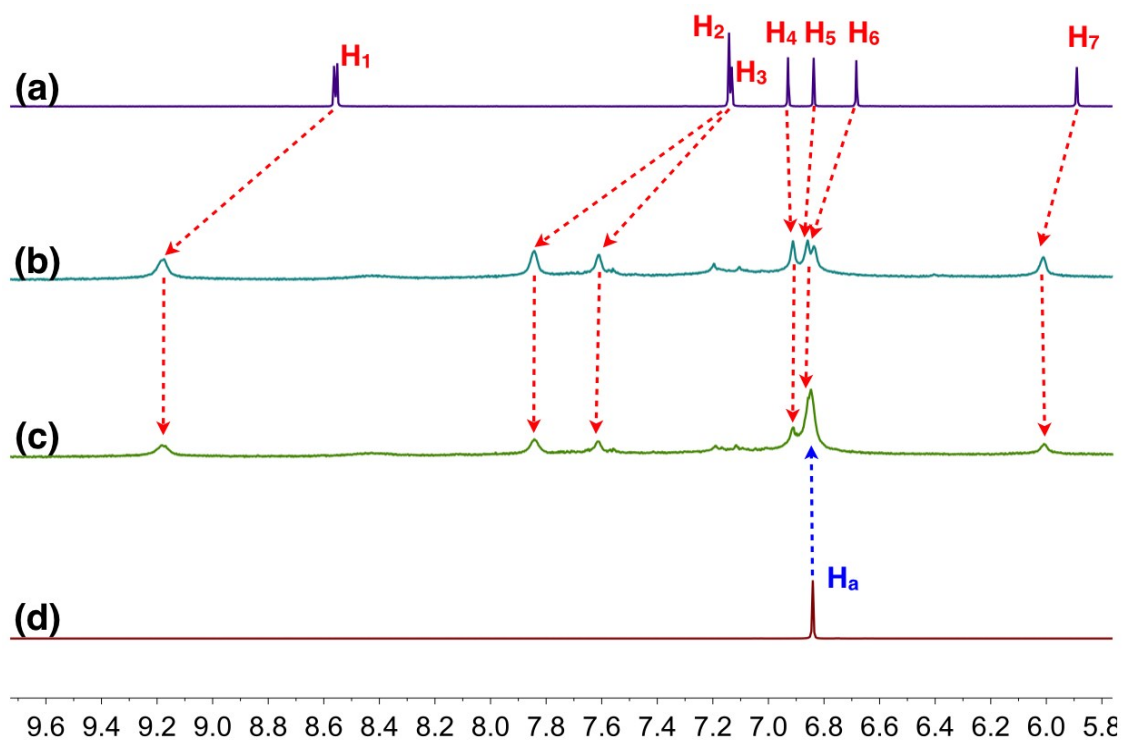


Figure S10. Partial ^1H NMR spectra (500 MHz, acetone- d_6 , 298 K): (a) **1**; (b) **1 + 4**; (c) **1 + 4 + 2**; (d) **2**. $c = 5.00$ mM.

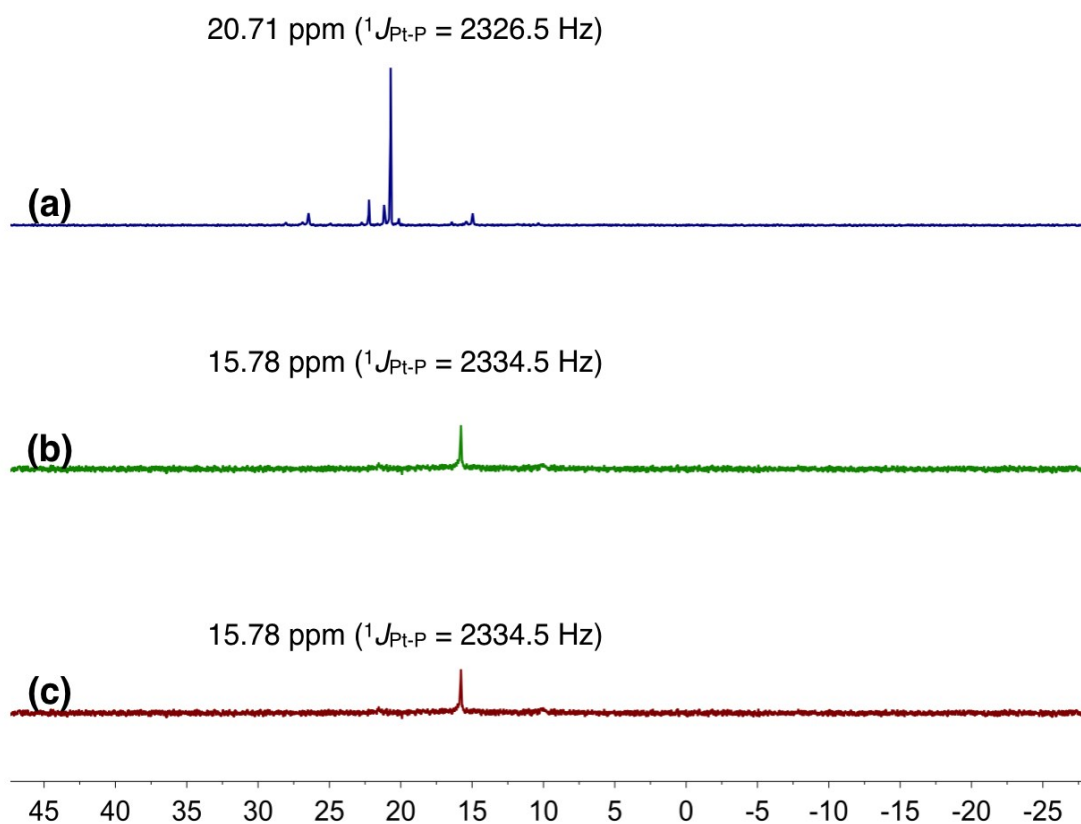


Figure S11. Partial $^{31}\text{P}\{^1\text{H}\}$ NMR spectra (202.3 MHz, acetone- d_6 , 298 K): (a) **4**; (b) **1 + 4**; (c) **1 + 4 + 2**. $c = 5.00$ mM.

6. Partial DOSY NMR spectra of supramolecular assemblies

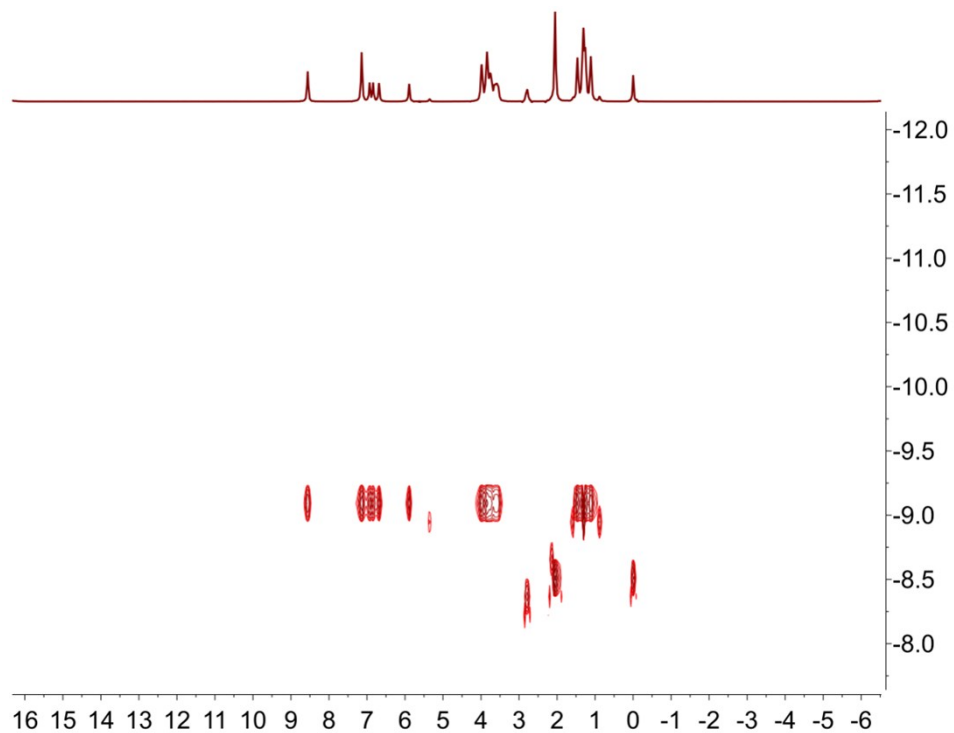


Figure S12. Partial DOSY NMR spectrum (500 MHz, acetone-*d*₆, 298 K) of **1** at 5.00 mM.

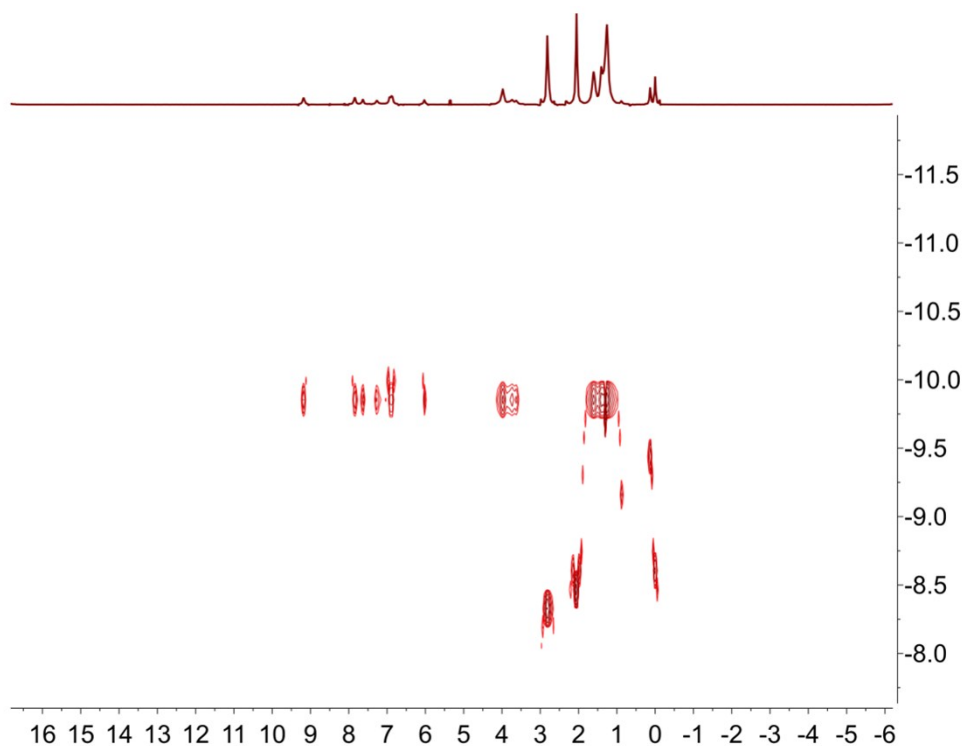


Figure S13. Partial DOSY NMR spectrum (500 MHz, acetone-*d*₆, 298 K) of **1** + **3** at 5.00 mM.

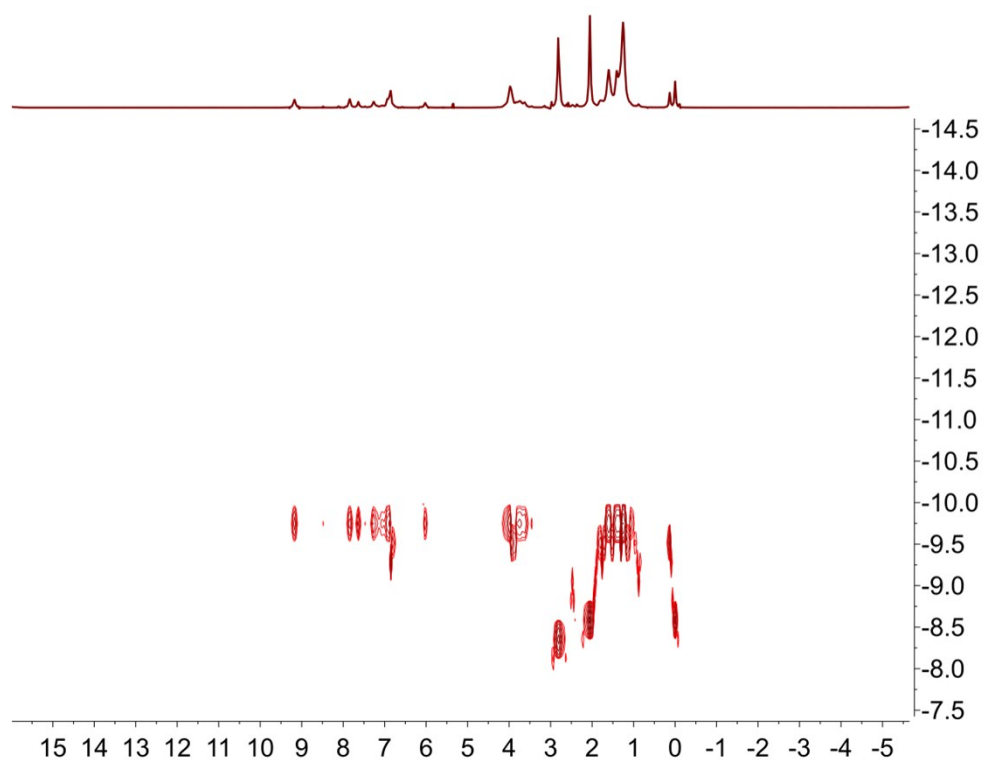


Figure S14. Partial DOSY NMR spectrum (500 MHz, acetone- d_6 , 298 K) of **1** + **3** + **2** at 5.00 mM.

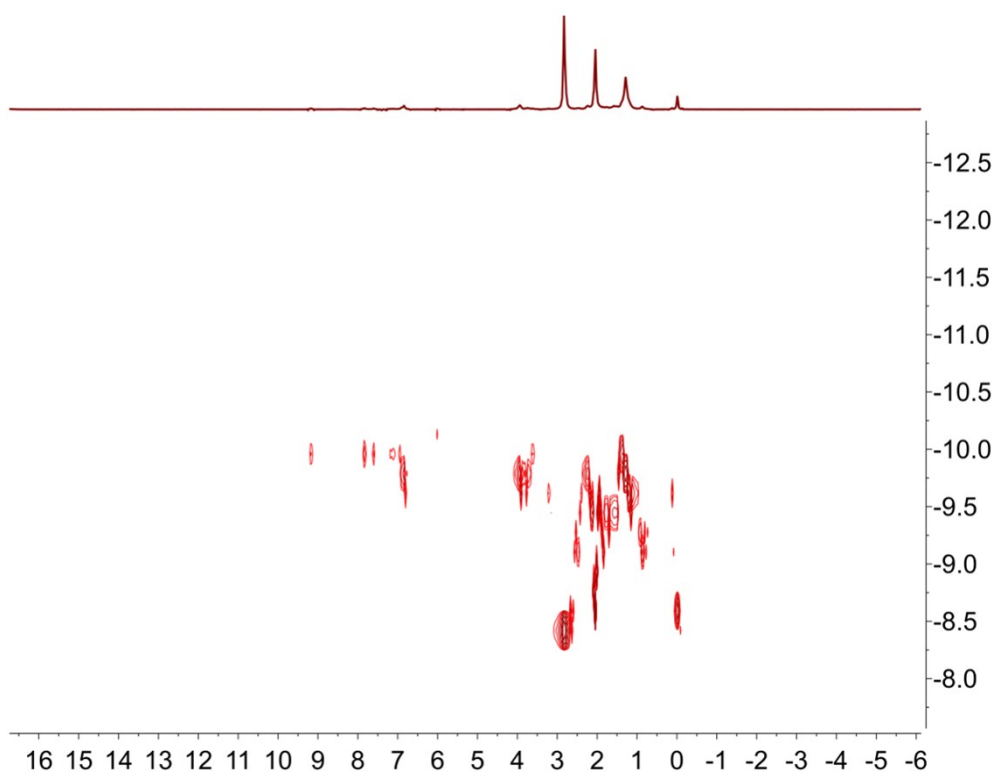


Figure S15. Partial DOSY NMR spectrum (500 MHz, acetone- d_6 , 298 K) of **1** + **4** at 5.00 mM.

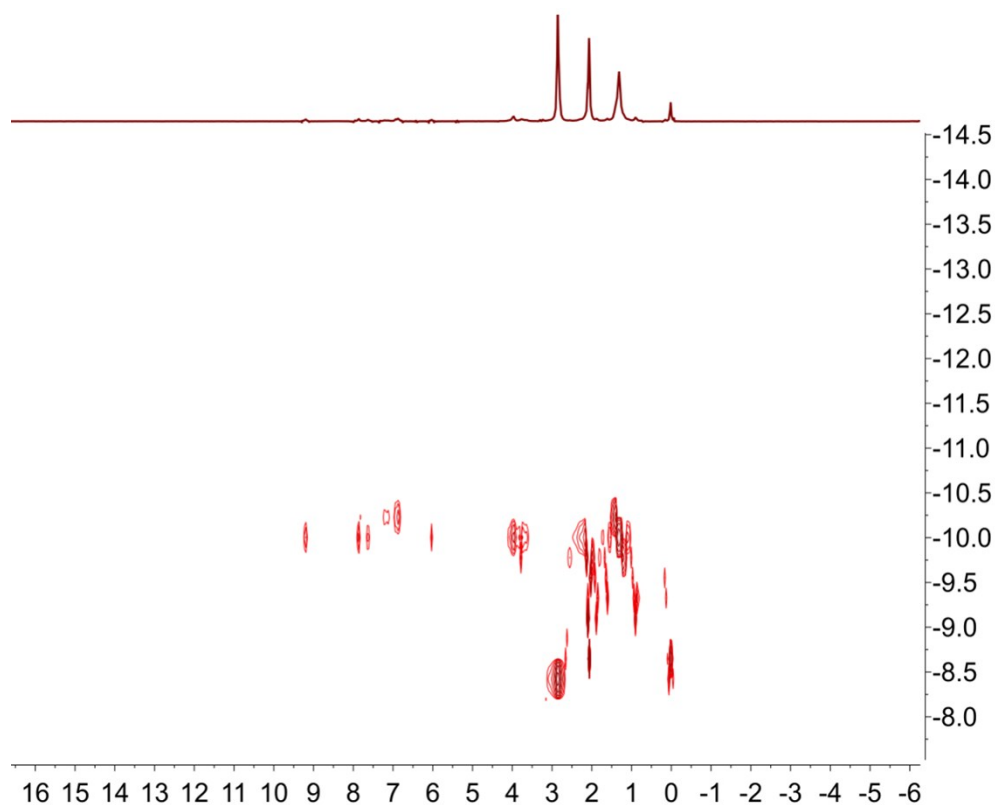


Figure S16. Partial DOSY NMR spectrum (500 MHz, acetone- d_6 , 298 K) of **1** + **4** + **2** at 5.00 mM.

7. DOSY and DLS results of **1**, **1** + **4**, and **1** + **4** + **2**

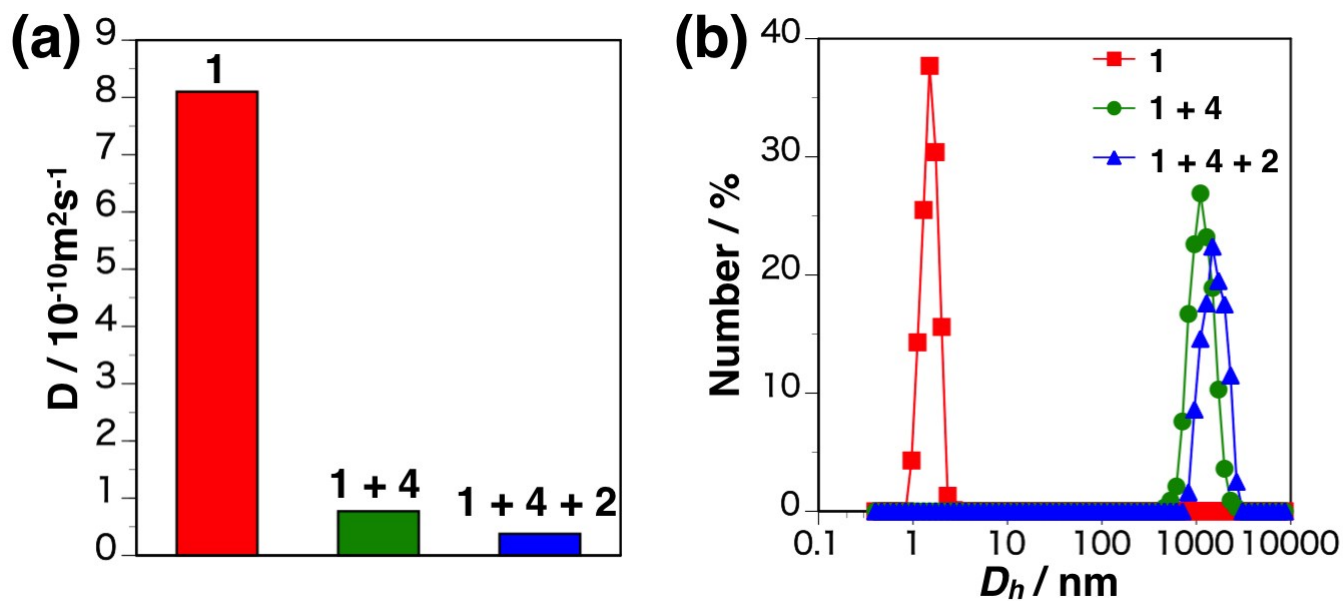


Figure S17. (a) Diffusion coefficient D values (500 MHz, acetone- d_6 , 298 K) of **1**, **1** + **4**, and **1** + **4** + **2**. (b) Size distributions of **1**, **1** + **4**, and **1** + **4** + **2**. $c = 5.00$ mM.

8. SEM images of **1** + **4** and **1** + **4** + **2**

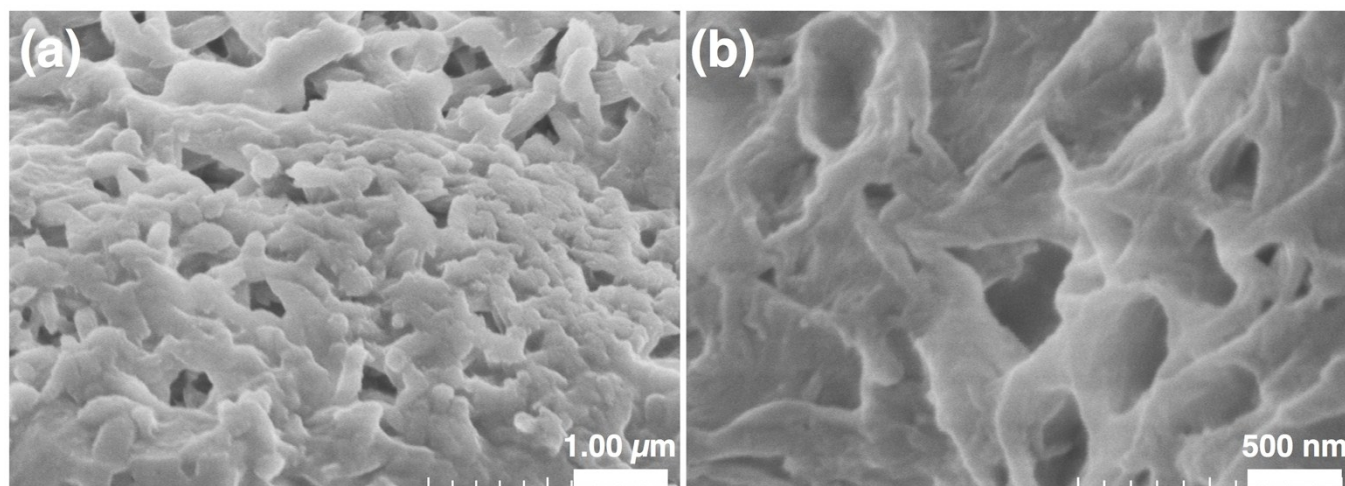


Figure S18. SEM image of (a) **1** + **4** and (b) **1** + **4** + **2** in acetone at 100 mM.

9. Multi-responsiveness of gels

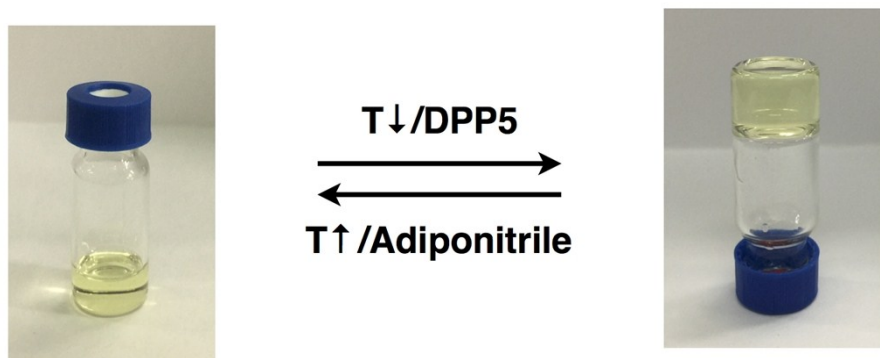


Figure S19. The gel-sol transitions of the supramolecular gel triggered by different stimuli.

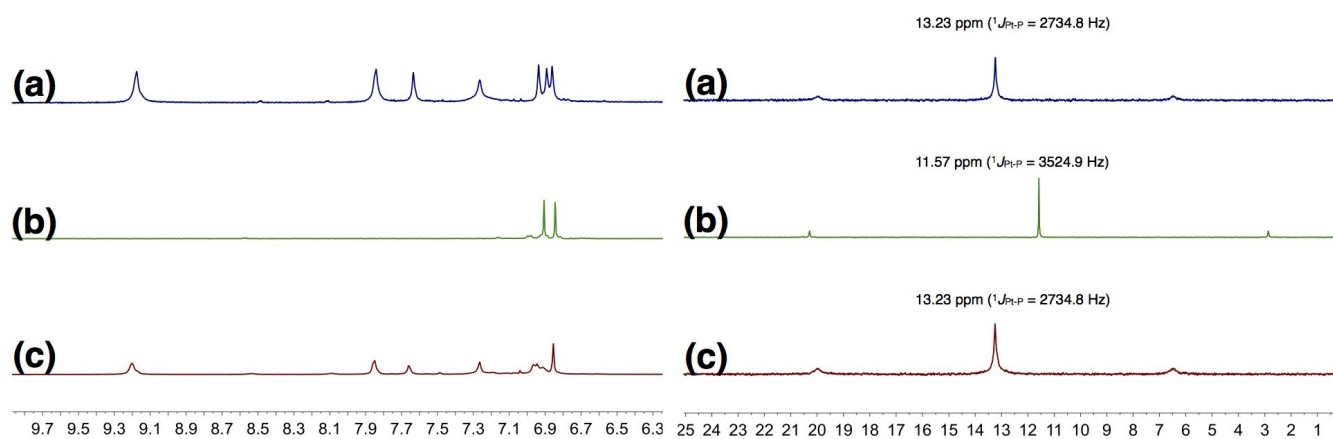


Figure S20. Partial ^1H NMR (500 MHz, acetone- d_6 , 298 K) (left) and $^{31}\text{P}\{^1\text{H}\}$ NMR (202.3 MHz, acetone- d_6 , 298 K) (right) spectra of (a) **1** + **3** + **2**; (b) after the addition of 2 equiv of TBABr to a; (c) after subsequent addition of 4 equiv of AgOTf to b. $c = 5.00$ mM.

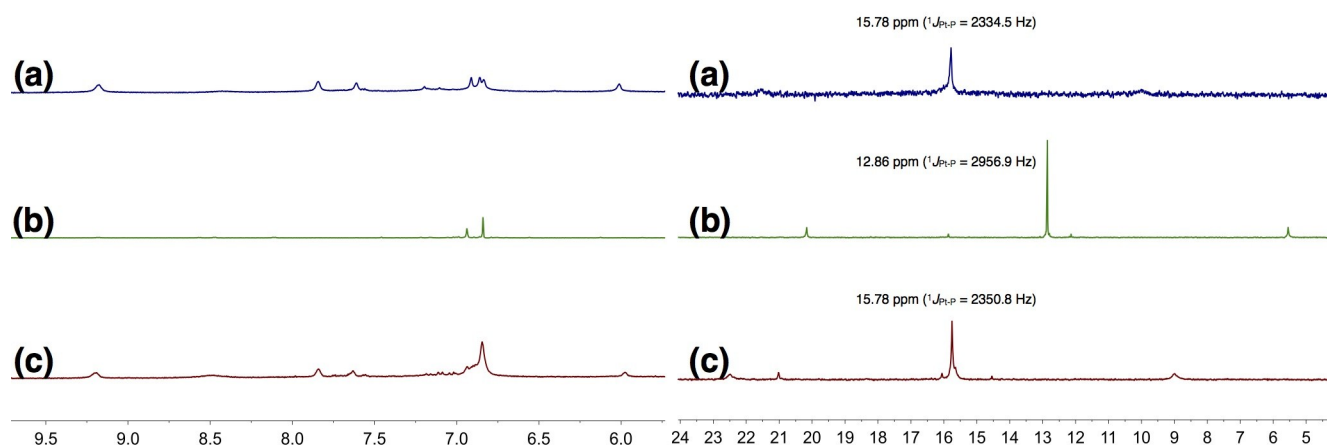


Figure S21. Partial ^1H NMR (500 MHz, acetone- d_6 , 298 K) (left) and $^{31}\text{P}\{^1\text{H}\}$ NMR (202.3 MHz, acetone- d_6 , 298 K) (right) spectra of (a) **1** + **4** + **2**; (b) after the addition of 2 equiv of TBABr to a; (c) after subsequent addition of 4 equiv of AgOTf to b. $c = 5.00$ mM.

10. X-ray analysis data for **1**

Crystallographic data: prism, colourless, $0.448 \times 0.350 \times 0.252$ mm³, $\text{C}_{61}\text{H}_{68}\text{N}_2\text{O}_8 \cdot 2\text{C}_3\text{H}_6\text{O}$, FW 1073.33, orthorhombic, space group $P-1$, $a = 25.3293(7)$, $b = 20.1695(5)$, $c = 23.6895(6)$ Å, $\alpha = 90^\circ$, $\beta = 90^\circ$, $\gamma = 90^\circ$, $V = 12102.5(5)$ Å³, $Z = 8$, $D_c = 1.178$ g cm⁻³, $T = 150(1)$ K, $\mu = 0.078$ mm⁻¹, 28854 measured reflections, 9828 independent reflections, 1359 parameters, 59 restraints, $F(000) = 4608$, $R_1 = 0.1727$, $wR_1 = 0.3001$ (all data), $R_2 = 0.0889$, $wR_2 = 0.2365$ [$I > 2\sigma(I)$], max. residual density 0.792 e \cdot Å⁻³, and goodness-of-fit (F^2) = 1.015. CCDC 1522035.

11. References:

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