# Wholly biomass derivable sustainable polymers by ring-

# opening metathesis polymerisation of monomers obtained

# from furfuryl alcohol and itaconic anhydride

## **Supplementary Information**

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The original data associated with this research is available at: DOI: 10.15124/f4cf9761-ba74-4de4-9903-16261b5baa7c



Not all <sup>13</sup>C peaks within the cetyl chain are resolved or have been assigned.

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.54 (1H, dd, *J* 5.8, 1.7 Hz, H1), 6.48 (1H, d, *J* 5.8 Hz, H2), 5.03 (1H, dd, *J* 4.7, 1.6 Hz, H6), 4.78 (1 H, d, *J* 10.7 Hz, H7), 4.59 (1H, d, *J* 10.7 Hz, H7), 4.05 (2H, td, *J* 6.8, 1.7 Hz, H11), 2.6–2.4 (2H, m, H5+H9), 2.28 (1H, d, *J* 14.6 Hz, H9), 1.7–1.5 (2H, m, H12), 1.47 (1H, d, *J* 12.2 Hz, H5), 1.4–1.1 (26H, m, H13-25), 0.87 (3H, t, *J* 6.8 Hz, H26).

 $^{13}$ C NMR (CDCl<sub>3</sub>, 101 MHz)  $\delta_{\rm C}$  177.2 (C8), 169.7 (C10), 138.1 (C1), 130.8 (C2), 94.2 (C3), 78.8 (C6), 68.9 (C7), 65.7 (C11), 53.6 (C12), 52.2 (C4), 40.3 (C9), 36.8 (C5), 32.1, 29.8, 29.7, 29.6, 29.5, 29.3, 28.5, 26.0, 22.8, 14.3 (C26).

FTIR (neat, ATR) v<sub>max</sub> 3013, 2952, 2913, 2850, 1768 and 1739 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>26</sub>H<sub>42</sub>NaO<sub>5</sub> 457.2924, Found 457.2929







<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.6–6.5 (1H, m, H1), 6.49 (1H, d, *J* 5.8 Hz, H2), 5.04 (1H, dd, *J* 4.7, 1.3 Hz, H6), 4.79 (1H, d, *J* 10.7 Hz, H7), 4.60 (1H, d, *J* 10.7 Hz, H7), 4.06 (2H, td, *J* 6.9, 1.6 Hz, H11), 2.6–2.4 (2H, m, H5+H9), 2.28 (1H, d, *J* 14.6 Hz, H9), 1.7–1.5 (2H, m, H12), 1.48 (1H, d, *J* 12.2 Hz, H5), 1.4-1.2 (10H, m, H13-H17), 0.88 (3H, t, *J* 6.8 Hz, H18).

 $^{13}$ C NMR (CDCl<sub>3</sub> 101 MHz)  $\delta_{C}$  177.1 (C8), 169.6 (C10), 138.0 (C1), 130.7 (C2), 94.1 (C3), 78.7 (C6), 68.7 (C7), 65.5 (C11), 52.0 (C4), 40.1 (C9), 36.7 (C5), 31.8 (C12), 29.2 (C13+C14), 28.4 (C15), 25.8 (C16), 22.6 (C17), 14.1 (C18).

FTIR (neat, ATR) v<sub>max</sub> 2955, 2926, 2856, 1774 and 1730 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>18</sub>H<sub>26</sub>NaO<sub>5</sub> 345.1672, Found 345.1663







<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.55 (1H, dd, *J* 5.8, 1.6 Hz, H1), 6.49 (1H, d, *J* 5.9 Hz, H2), 5.04 (1H, dd, *J* 4.7, 1.5 Hz, H6), 4.79 (1H, d, *J* 10.7 Hz, H7), 4.60 (1H, d, *J* 10.7 Hz, H7), 4.10 (2H, td, *J* 7.0, 1.4 Hz, H11), 2.6–2.4 (2H, m, H5+H9), 2.28 (1H, d, *J* 14.6 Hz, H9), 1.7-1.6 (1H, m, H13), 1.6–1.4 (3H, m, H5+H12), 0.91 (6H, d, *J* 6.7, H14).

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 101 MHz)  $\delta_{C}$  177.1 (C8), 169.5 (C10), 138.0 (C1), 130.6 (C2), 94.0 (C3), 78.7 (C6), 68.6 (C7), 63.9 (C11), 52.0 (C4), 40.0 (C9), 37.0 (C5), 36.6 (C13), 24.9 (C12), 22.4 (C14), 22.3 (C14).

FTIR (neat, ATR) v<sub>max</sub> 2958, 2872, 1772 and 1729 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>15</sub>H<sub>20</sub>NaO<sub>5</sub> 303.1203, Found 303.1197







<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.55 (1H, dd, *J* 5.8, 1.7 Hz, H1), 6.49 (1 H, d, *J* 5.8 Hz, H2), 5.04 (1H, dd, *J* 4.7, 1.7 Hz, H6), 4.79 (1H, d, *J* 10.8 Hz, H7), 4.60 (1H, d, *J* 10.7 Hz, H7), 4.07 (2H, td, *J* 6.7, 1.9 Hz, H11), 2.6–2.4 (2H, m, H5+H9), 2.28 (1H, d, *J* 14.5 Hz, H9), 1.7–1.5 (2H, m, H12), 1.48 (1H, d, *J* 12.2 Hz, H5), 1.4–1.3 (2 H, m, H13), 0.92 (3H, t, *J* 7.4 Hz, H14).

 $^{13}\text{C}$  NMR (CDCl<sub>3</sub> 101 MHz)  $\delta_{\text{C}}$  177.1 (C8), 169.6 (C10), 138.0 (C1), 130.6 (C2), 94.0 (C3), 78.6 (C6), 68.6 (C7), 65.1 (C11), 52.0 (C4), 40.0 (C9), 36.6 (C5), 30.4 (C12), 19.0 (C13), 13.6 (C14).

FTIR (neat, ATR) v<sub>max</sub> 2960, 2785, 1772 and 1728 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>14</sub>H<sub>18</sub>NaO<sub>5</sub> 289.1046, Found 289.1048







<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.54 (1H, dd, *J* 5.8, 1.7 Hz, H1), 6.49 (1H, d, *J* 5.8 Hz, H2), 5.03 (1H, dd, *J* 4.7, 1.6 Hz, H6), 4.79 (1H, d, *J* 10.7 Hz, H7), 4.59 (1H, d, *J* 10.7 Hz, H7), 2.53 (1H, dd, *J* 12.2, 4.8 Hz, H5), 2.45 (1H, d, *J* 14.2 Hz, H9), 2.16 (1H, d, *J* 14.2 Hz, H9), 1.57 (2H, d, *J* 2.5 Hz, H5), 1.45 (9H, s, H12).

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 101 MHz) δ<sub>C</sub> 177.2 (C8), 168.9 (C10), 138.0 (C1), 130.9 (C2), 94.2 (C3), 82.2 (C11), 78.8 (C6), 68.7 (C7), 52.2 (C4), 41.5 (C9), 36.7 (C5), 28.0 (C12).

FTIR (neat, ATR) v<sub>max</sub> 3070, 2980, 2938, 1774 and 1716 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>14</sub>H<sub>18</sub>NaO<sub>5</sub> 289.1046, Found 289.1043





#### Isopropyl ester 8g



<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.55 (1H, dd, *J* 5.8, 1.6 Hz, H1), 6.49 (1H, d, *J* 5.8 Hz, H2), 5.1-4.8 (2H, m, H11+H6), 4.79 (1H, d, *J* 10.8 Hz, H7), 4.60 (1H, d, *J* 10.7 Hz, H7), 2.6-2.5 (1H, m, H5), 2.49 (1H, d, *J* 14.3 Hz, H9), 2.24 (1H, d, *J* 14.3 Hz, H9), 1.48 (1H, d, *J* 12.1 Hz, H5), 1.24 (3H, d, *J* 6.3 Hz, H12), 1.23 (3H, d, *J* 6.3 Hz, H12).

<sup>13</sup>C NMR (CDCl<sub>3</sub>, 101 MHz)  $\delta_{C}$  177.1 (C8), 169.0 (C10), 138.0 (C1), 130.6 (C2), 94.0 (C3), 78.7 (C6), 69.0 (C7), 68.6 (C11), 52.0 (C4), 40.2 (C9), 36.6 (C5), 21.7 (C12), 21.6 (C12).

FTIR (neat, ATR) v<sub>max</sub> 2982, 1772 and 1723 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>13</sub>H<sub>16</sub>NaO<sub>5</sub> 275.0890, Found 275.0888







This compound was formed as a 1:1 mixture of two diastereomers.

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 400 MHz)  $\delta_{H}$  6.53 (1H, dd, *J* 5.8, 1.6 Hz, H1), 6.49 (1H, dd, *J* 5.8, 1.9 Hz, H2), 5.02 (1H, dd, *J* 4.7, 1.6 Hz, H6), 4.82 (1H, d, *J* 10.6 Hz, H7), 4.59 (1H, d, *J* 10.6 Hz, H7), 2.6–2.4 (2H, m, H5+H9), 2.17 (1H, dd, *J* 14.6, 2.2 Hz, H9), 1.9–1.6 (4H, m, H12+H18), 1.6–1.4 (1H, m, H5), 1.36 (3H, s, H17), 1.3–1.0 (5H, m, H13+H14+H15), 0.9–0.8 (9H, m, H16+H19).

 $^{13}$ C NMR (CDCl<sub>3</sub>, 101 MHz)  $\delta_{\rm C}$  (101 MHz) 177.2 (C8), 168.86 and 168.85 (C10), 137.86 and 137.83 (C1), 130.99 and 130.97 (C2), 94.3 (C3), 87.4 (C11), 78.69 and 78.68 (C6), 68.85 and 68.83 (C7), 52.09 and 52.08 (C4), 41.60 and 41.57 (C9), 39.33 and 39.31 (C14), 37.90 and 37.88 (C12), 36.88 and 36.86 (C5), 30.84 and 30.81 (C18), 27.93 and 27.92 (C15), 23.20 and 23.16 (C17), 22.73, 22.71, 22.69 and 22.68 (C16), 21.45 and 21.43 (C13), 8.15 and 8.12 (C19).

FTIR (neat, ATR) v<sub>max</sub> 2953, 2870, 1775 and 1721 cm<sup>-1</sup>

HRMS (ESI) m/z: [M+Na]<sup>+</sup> Calculated for C<sub>20</sub>H<sub>30</sub>NaO<sub>5</sub> 373.1985, Found 373.1985.





## Polymerised cetyl ester 9b (100:1 homopolymer)







## IR Spectrum



## <sup>1</sup>H NMR Spectrum



<u>SEC</u>



## IR Spectrum



## Polymerised isoamyl ester 9d (100:1 homopolymer)

## <sup>1</sup>H NMR Spectrum



<u>SEC</u>



## IR Spectrum



## Polymerised linalooyl ester 9h (100:1 homopolymer)

<sup>1</sup>H NMR Spectrum



<u>SEC</u>



## IR Spectrum







#### NMR monitoring of the block copolymerisation of monomers 8a and 8b

Additional stacked <sup>1</sup>H NMR plot showing more detail of the alkylidene region during the initial stages of polymerisation of **8a**. The change in alkylidene signal reflecting monomer, dimer, and then oligomer bound to ruthenium, is apparent.



Graph showing total monomer conversion throughout the experiment as a function of time.

Plot of carbene integral versus time



## Individual spectra from Figure 7



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20.0	19.5	19.0	18.5	18.0	17.5 f1 (ppm)	17.0	16.5	16.0	15.5	15.
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t = 534 minutes



t = 1488 minutes

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20.0	19.5	19.0	18.5	18.0	17.5 f1 (ppm)	17.0	16.5	16.0	15.5	15.





t = 1691 minutes





15.445	15.430	15.415	15.400	15.385 fi	15.370 L (ppm)	15.355	15.340	15.325	15.310



## <u>DSC</u>



## Random copolymer of monomers 8a and 8b





Sample: YB124B random cetyl methyl copol Size: 6.7000 mg Method: H-C-H



File: ...\YB124B cetyl-methyl random copol.001 Operator: jwc Run Date: 16-Nov-2016 23:12 Instrument: DSC Q2000 V24.10 Build 122

