[Supporting Information]

"Bottom-Up" Construction of Chiral Porous Organic Polymers for Heterogeneous Asymmetric Organocatalysis: MacMillan Catalyst Built-in Nanoporous Organic Frameworks

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A. General information

Chemicals and solvents were purchased from commercial suppliers and, if necessary, purified before use by standard techniques. All equipment was thoroughly oven-dried. Chemicals and solvents were purchased from commercial suppliers and, if necessary, purified before use by standard techniques. Thin-layer chromatography (TLC) plates were visualized by exposure to ultraviolet light and/or immersion in a solution of phosphomolybdic acid in methanol followed by heating on a hot plate. Flash column chromatography (FCC) was carried out with silica gel (200–300 mesh). ¹H and ¹³C liquid NMR spectra were recorded on a Bruker Avance III 400 MHz spectrometer. The chemical shifts δ are given in ppm (parts per million) relative to tetramethylsilane (TMS) and the coupling constants J are given in Hz. All the spectra were recorded in CDCl₃ as solvent at room temperature. TMS served as the internal standard ($\delta = 0.00$ ppm) for ¹H NMR while CDCl₃ as the internal standard ($\delta = 77.0$ ppm) for ¹³C NMR. HRMS data were obtained ESI ionization sources. Enantiomeric excess (ee) values were determined by HPLC on a Agilent Technologies 1200 Series and Waters 1525 Delta system with Daicel chiral OJ-H and AD-H columns and with *i*-PrOH/*n*-hexane as the eluent. Elemental analysis was performed on an Elementar Analysensysteme GmbH VarioEL V3.00 elemental analyzer. FT-IR spectra were recorded on a Nicolet NEXUS 670 instrument. The nitrogen adsorption and desorption isotherms were measured at 77 K using a Micromeritics ASAP 2020M system. Solid-state NMR experiments were performed on a Bruker Avance II WB 400 MHz spectrometer. The ¹³C CP/MAS NMR spectra were recorded with the contact time of 2 ms (ramp 100) and the pulse delay of 3 s. Surface morphologies and microstructures of the synthesized materials were examined with a Hitachi S-4800 scanning electron microscope (SEM) and with a JEOL JEM-2010 transmission electron microscope (TEM) operated at 200 KV. Thermal properties of the synthesized materials were evaluated on a STA PT1600 Linseis thermogravimetric analysis (TGA) instrument in

the temperature range of 25 to 800 °C under nitrogen atmosphere with a heating rate of 10 °C/min. Powder X-ray diffraction (PXRD) data were collected with a Rigaku D/MAX-2400 X-ray diffractometer operated at 40 kV and 100 mA with Cu K α radiation at a scan rate of 15°/min.

B. Materials and synthesis

Materials. All reagents were purchased from commercial sources and used as received without further purification. THF and toluene were dried over Nabenzophenone prior to distillation. DMF was dried by calcium hydride and used after distillation. [Pd(PPh₃)₂Cl₂] and 1,3,5-triethynylbenzene was prepared and purified according to the literature procedures. All anhydrous reactions were carried out under dry nitrogen by using Schlenk tube techniques. All catalytic reactions were performed in a 5 mL centrifuge tube.

(I) Synthesis of the chiral functional building block (FBB-Mac)^[1]



To a suspension of 3,5-diiodo-(L)-tyrosine dehydrate 1 (4.69 g, 10 mmol) in icecooled dry methanol (30 mL) was added dropwise thionyl chloride (2.38 g, 20 mmol). After the solution was stirred at room temperature overnight, the solvent was removed under reduced pressure to give quantitatively 3,5-diiodo-(L)-tyrosine methyl ester hydrochloride 2 as colorless crystalline solid, which was directly used in the next step without further purification. To a solution of 33% ethanolic MeNH₂ (1.53 g, 15 mmol) was added 2 (2.42 g, 5 mmol) and the resulting solution was stirred at room temperature until the amino ester was consumed as determined by TLC. After removal of the organic solvents under vacuum, the residue was then suspended in Et₂O and concentrated. The Et₂O addition–removal cycle was repeated several times to remove excess MeNH₂. The obtained amide hydrochloride **3** was treated with saturated aqueous NaHCO₃ (20 mL) and stirred for 30 min. The resulting mixture was filtered and the residue was desiccated under vacuum. To this residue was added MeOH (25 mL), acetone (40 mL) and *p*TSA (50 mg). The resulting solution was heated to reflux for 24 h and concentrated under vacuum to afford the crude product, which was recrystallized further from MeOH to provide **FBB-Mac** as white solid in 70% overall yield. ¹H NMR (400 MHz, d₆-DMSO): δ (ppm) 9.31 (s, 1H), 7.64 (s, 2H), 3.57 (d, J = 7.3 Hz, 2H), 3.34 (s, 1H), 2.87 (d, J = 13.7 Hz, 2H), 2.64 (s, 3H), 1.20 (s, 3H), 1.18 (s, 3H). 13C NMR (100 MHz, d6-DMSO): δ (ppm) 172.5, 153.5, 139.7, 135.2, 86.8, 75.1, 58.9, 35.7, 27.1, 24.9, 24.7. RMS m/z calcd for C₁₃H₁₆I₂N₂O₂ (M+H): 486.9374, found: 486.9360.





To a solution of 1,4-diiodobenzene **1** (2.00 g, 6.06 mmol) in THF (10 mL) and NEt₃ (6 mL) was added $PdCl_2(PPh_3)_2$ (255 mg, 0.36 mmol) and CuI (72 mg, 0.36 mmol). The solution was degassed with argon for 30 min. 2-Methyl-3-butyn-2-ol (510 mg, 60.06 mmol) was then added dropwise and the solution was stirred overnight at room temperature. The solvent was removed by rotary evaporation and the residue was treated with water and extracted with dichloromethane. The organic extracts were washed with water, brine, and dried over magnesium sulfate. The solvent was then

removed by rotary evaporation and the residue was purified by column chromatography on silica gel, eluting with ethyl acetate-petroleum ether (v/v 10/60) to give 2 (820 mg, 47 % yield). PdCl₂(PPh₃)₂ (22 mg, 0.0315 mmol) and CuI (30 mg, 0.16 mmol) were added to a solution of 1,3,5-triethynylbenzene (95 mg, 0.63 mmol) and 2 (540 mg, 1.89 mmol) in NEt₃ (35 mL). The mixture was stirred for 2 h and then filtered through a glass sinter. The filtrate was reduced to dryness in vacuo. The residue was purified by column chromatography on silica gel, eluting with ethyl acetate-petroleum ether (v/v 10/10) Removal of the solvent afforded 3 as brown solid (318 mg, 81 % yield). To a solution of 3 (318 mg, 0.51 mmol) in anhydrous toluene (10 mL) was added sodium hydride (48 mg, 2.04 mmol). The solution was refluxed for 3 h at 130 °C. The solvent was then removed by rotary evaporation and the residue was treated with NH₄Cl and extracted with dichloromethane. The organic extracts were washed with water, and then brine, and dried over magnesium sulfate. The solvent was removed by rotary evaporation And the residue was purified by column chromatography on silica gel, eluting with ethyl acetate-petroleum ether (v/v 10/60) to give 1,3,5-tris((4-ethynylphenyl)ethynyl)benzene 4 (172 mg, 75 % yield) as light yellow solid. ¹H NMR (400 MHz, CDCl₃): δ (ppm) 7.65 (s, 3H); 7.48 (s, 12H); 3.19 (s, 3H). ¹³CNMR (100 MHz, CDCl₃): δ (ppm) 134.2, 132.1, 131.6, 123.9, 123.2, 122.4, 90.1, 89.6, 83.2, 79.2.

(III) Synthesis of MacMillan catalyst-based porous organic polymers (Mac-CPOP-1 and Mac-CPOP-2)^[3]

The Mac-CPOPs frameworks were synthesized via palladium catalyzed Sonogashira-Hagihara coupling reaction of functional building block (FBB-Mac) with different structural building blocks: (1,3,5-triethynylbenzene (SBB-I) or 1,3,5-tris((4ethynylphenyl)ethynyl)benzene (SBB-II).



SBB-I (100)0.67 mmol), FBB-Mac (486 mg, mg, 1.00 mmol), bis-(triphenylphosphine)palladium(II) dichloride (75 mg), and copper iodide (45 mg) were dissolved in a mixture of dried dimethylformamide (5 mL) and Et₃N (5 mL). The reaction mixture was heated to 80 °C and stirred for 72 h under nitrogen atmosphere. The mixture was then cooled to room temperature, and the precipitated Mac-CPOP-1 polymer was filtered and washed four times (once each) with chloroform, water, methanol, and acetone to remove any unreacted substrates or catalyst residues. Further purification of the Mac-CPOP-1 polymer was carried out by Soxhlet extraction with methanol for 48 h. The product was dried in vacuum for 24 h at 70 °C (308 mg, yield: 93%). Elemental analysis (%) Found: C 68.59, H 4.72, N 6.25. Solid-state ¹³C CP/MAS NMR: δ 173, 155, 130, 123, 107, 90, 75, 59, 35, 25 ppm. IR (KBr): 3440, 3065, 2205, 1610, 1538, 1500, 1409, 826 cm⁻¹.



SBB-II (63 mg, 0.14 mmol), **FBB-Mac** (102 mg, 0.21 mmol), bis-(triphenylphosphine)palladium(II) dichloride (30 mg), and copper iodide (15 mg) were dissolved in a mixture of dried dimethylformamide (2 mL) and Et₃N (2 mL). The reaction mixture was heated to 80 °C and stirred for 72 h under nitrogen atmosphere. The mixture was then cooled to room temperature, and the precipitated **Mac-CPOP-2** polymer was filtered and washed four times (once each) with chloroform, water, methanol, and acetone to remove any unreacted substrates or catalyst residues. Further purification of the **Mac-CPOP-2** polymer was dried in vacuum for 24 h at 70 °C (105 mg, yield: 95%). Elemental analysis (%) Found: C 71.11, H 4.27, N 3.39. Solid-state ¹³C CP/MAS NMR: δ 173, 155, 139, 131, 123, 90, 75, 59, 35, 31, 24 ppm. IR (KBr): 3434, 3035, 2922, 2858, 2205, 1676, 1575, 1390, 833 cm⁻¹.



C. Physical properties for Mac-CPOP-1 and Mac-CPOP-2

Table S1. Physical properties for the synthesized CPOPs polymers

Polymer	Buildin	g Blocks	SA _{BET}		V _{total}	Catalyst Loading
	Functional	Structural		$(m^2 g^{-1})^a$	$(cm^3 g^{-1})^b$	(mmol g ⁻¹) ^c
Mac-CPOP-1	FBB-Mac	SBB-I	3:2	5.9	0.19	2.23
Mac-CPOP-2	FBB-Mac	SBB-II	3:2	468	0.62	1.21

^a Surface area (SA) calculated from the N₂ adsorption isotherm using the BET method. ^b Total pore volume at $P/P_{o} = 0.99$. ^c Catalyst loading calculated from elemental analysis.





Figure S1. N₂ adsorption–desorption isotherms for Mac-CPOP-1 measured at 77 K.

E. The geometries of the reactants and products

The geometries of the reactants and products of the the Diels-Alder reaction were optimized with the Gaussian 09 package^[4], using a B3LYP density function and the 6-31G(d, p) method. The three dimensional sizes of the reactants and products were approximately measured based the optimized geometries, such as $7.1 \times 9.1 \times 1.0$ Å for (*E*)-cinnamaldehyde, $5.4 \times 4.8 \times 2.8$ Å for cyclopentadiene, $10.2 \times 6.4 \times 5.2$ Å for *endo* cycloaddition product, and $9.3 \times 6.6 \times 5.2$ Å for *exo* cycloaddition product.



Figure S2. The geometries of the reactants and products

F. ¹³C CP/MAS NMR spectrum for Mac-CPOP-1



Figure S3. ¹³C CP/MAS NMR spectra of **Mac-CPOP-1**. The spectrum was recorded at an MAS rate of 10 kHz. Asterisks denote spinning sidebands. the peak at ca. 102 ppm and 107 ppm corresponds to the carbon atoms of phenylbenzo[b]furan structure, which was formed under Pd/Cu catalytic conditions.



G. FT-IR spectra for fresh and recycled catalysts

Figure S4. FT-IR spectra of **Mac-CPOP-2** as the fresh catalyst (in black) and as the recycled catalyst after six-time catalytic reactions (in red).

H. SEM images for Mac-CPOP-1



Figure S5. SEM images for Mac-CPOP-1

I. Powder X-Ray diffraction for Mac-CPOP-2



Figure S6. Powder X-ray diffraction pattern of **Mac-CPOP-2**. No intensive diffraction peaks were observable.

J. TGA for Mac-CPOP-1 and Mac-CPOP-2



Figure S7. TGA analysis of Mac-CPOP-2 and Mac-CPOP-2 under nitrogen atmosphere.

K. ¹³C CP/MAS NMR spectra for fresh and recycled catalysts



Figure S8. Solid-state ¹³C CP/MAS NMR spectra of **Mac-CPOP-2** as fresh catalyst (in black), and as recycled catalysts after six-time catalytic reactions (in red). Each ¹³C CP/MAS spectra was recorded at an MAS rate of 10.0 kHz and reported relative to Me₄Si. Asterisks denote spinning sidebands. It was evident that the structure of **Mac-CPOP-2** catalyst was not destroyed after catalytic reactions. However, little change in the high-field region (0 to 50 ppm) of ¹³C solid-state NMR spectra should not be overlooked.



L. N_2 adsorption-desorption isotherms of the recycled Mac-CPOP-2

Figure S9. N_2 adsorption-desorption isotherms of **Mac-CPOP-2** as recycled catalyst after three-time catalytic reactions and six-time catalytic reactions. We found that the BET surface of **Mac-CPOP-2** had decreased 180 m²g⁻¹ after the sixth recycled used, which could be due to the partial blocking of the polymeric nanopores.

M. General procedure for the asymmetric Diels-Alder reaction^[1]

To a stirred solution of **Mac-CPOP-2** (0.04 mmol, 33 mg, 20 mol%) in a 3/1 H₂O/MeOH mixture (1.0 mL), trifluoroacetic acid (0.05 mmol) was added and the mixture was stirred for 5 min at room temperature. α , β -Unsaturated aldehyde (0.2 mmol, 1.0 eq) used soon after purification and cyclopentadiene (1.0 mmol, 5.0 eq) were added in this order. The mixture was then stirred at ambient temperature in open air. After the reaction was completed (monitored by TLC), the mixture was centrifugated and the solid catalyst was washed with MeOH (2 x 5.0 mL) and EtOAc (3 x 5.0 mL). After the evaporation of the combined organic phase under vacuum, the residue was purified by flash column chromatography with petroleum ether/ethyl acetate mixture as eluent to give the product as oil. Ratio of *endo* and *exo* was determined by 400 MHz ¹H liquid NMR. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H or AD-H column.

(1S, 2S, 3S, 4R)-3-Phenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-Phenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 1).



Prepared according to the general procedure with (*E*)-cinnamaldehyde and cyclopentadiene. The products were collected as a colorless oil in 95% yield after silica gel chromatography (hexane:ethyl acetate = 10:1); *endo* 81% ee, *exo* 75% ee. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (70/30 hexane/i-PrOH; flow rate 0.75 ml/min) *endo* isomer [T_R1 = 14.69 min (major), T_R2 = 9.27 min (minor)], *exo* isomer [T_R1 = 23.18 min (major), T_R2 = 18.03 min (minor)]. Spectroscopic data are in agreement with the published data.^[1]

(1S, 2S, 3S, 4R)-3-(2-Naphtyl)-bicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-(2-Naphtyl)-bicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 2).^[5]



Prepared according to the general procedure

with 3-naphthalen-2-yl-propenal, and cyclopentadiene. The products were collected as a colorless oil in 70% yield after silica gel chromatography (hexane:ethyl acetate=10:1); *endo* 73% ee, *exo* 75% ee. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (70/30 hexane/i-PrOH; flow rate 0.75 ml/min); endo isomer [T_R1= 12.24 min (major), T_R2 = 9.03 min (minor)], exo isomer [T_R1 = 16.97 min (major), T_R2 = 10.96 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.99 (d, J = 2.0 Hz, 1H), 7.83–7.32 (m, 7H), 6.39 (dd, J = 5.6, 3.2 Hz, 1H), 6.11 (dd, J = 5.6, 2.9 Hz, 1H), 3.93–3.90 (m, 1H), 3.33 (s, 1H), 3.28-3.27 (m, 1H), 2.75–2.74(m, 1H), 1.92-1.61 (m, 2H). *endo* isomer: δ 9.67 (d, J = 2.1 Hz, 1H), 7.83–7.32 (m, 7H), 6.49 (dd, J = 5.6, 3.3 Hz, 1H), 6.23 (dd, J = 5.6, 2.8 Hz, 1H), 3.40 (s, 1H), 3.28-3.27 (m, 2H), 3.10–3.09 (m, 1H), 1.92-1.61 (m, 2H) ¹³C NMR (100 MHz, CDCl₃) δ 203.3, 202.7, 141.1, 140.0, 139.2, 136.6, 136.3, 133.8, 133.5, 133.2, 132.2, 132.1, 128.2, 127.7, 127.7, 127.6, 127.5, 127.5, 126.7, 126.6, 126.1, 126.0, 125.9, 125.5, 125.4, 124.8, 60.8, 59.2, 48.5, 48.3, 47.6, 47.2, 45.9, 45.6, 45.5, 45.2.

(1S, 2S, 3S, 4R)-3-o-Methoxyphenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-o-Methoxyphenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 3).^[6]



Prepared according to the general procedure

with 3-(4-methoxyphenyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 64% yield after silica gel chromatography (hexane:ethyl acetate=10:1); *endo* 91% ee, *exo* 88% ee. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (90/10 hexane/i-PrOH; flow rate 0.75 ml/min); endo isomer [T_R1 = 22.78 min (major), T_R2 = 18.97 min (minor)], exo isomer [T_R1 = 30.59 min (major), T_R2 = 20.12 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.91 (d, J = 2.1 Hz, 1H), 7.07 (d, J = 8.4 Hz, 2H), 6.79 (d, J = 8.8 Hz, 2H), 6.34 (dd, J = 5.6, 3.2 Hz, 1H), 6.08 (dd, J = 5.6, 2.8 Hz, 1H), 3.78 (s, 3H), 3.67–3.65 (m, 1H), 3.21–3.18 (m, 1H), 3.02 (t, J = 6.1 Hz, 1H), 2.54-2.53 (m, 1H), 1.63-1.54 (m, 2H). *endo* isomer: δ 9.58 (d, J = 2.3 Hz, 1H), 7.19 (d, J = 8.8 Hz, 2H), 6.85 (d, J = 8.8 Hz, 2H), 6.42 (dd, J = 5.6, 3.2 Hz, 1H), 6.17 (dd, J = 5.6, 2.7 Hz, 1H), 3.80 (s, 3H), 3.32 (s, 1H), 3.21–3.18 (m, 1H), 3.07 (s, 1H), 2.95–2.93 (m, 1H), 1.79 (d, J = 8.7 Hz, 1H), 1.63-1.54 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 203.7, 202.9, 158.1, 158.0, 139.2, 136.5, 136.3, 135.6, 134.6, 133.7, 128.8, 128.3, 114.0, 113.5, 60.9, 59.7, 55.3, 55.2, 48.7, 48.6, 47.6, 47.1, 45.5, 45.1, 45.1, 44.8.

(1S, 2S, 3S, 4R)-3-p-Chlorophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-p-Chlorophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 4).^[6]



Cl Prepared according to the general procedure with 3-(4-chlorophenyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 76% yield after silica gel chromatography (hexane:ethyl acetate=6:1); *endo* 95% ee, *exo* 93% ee. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (90/10 hexane/i-PrOH; flow rate 0.75 ml/min); endo isomer [T_R1= 21.18 min (major), T_R2 = 26.19 min (minor)], exo isomer [T_R1 = 30.12 min (major), T_R2 = 47.91 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.91 (d, J = 2.0 Hz, 1H), 7.30-7.06 (m, 4 H), 6.36 (dd, J = 5.6, 3.2 Hz, 1H), 6.05 (dd, J = 5.6, 2.8 Hz, 1H), 3.71–3.68 (m, 1H), 3.24–3.10 (m, 1H), 3.05 (d, J = 4.8 Hz, 1H), 2.54 (d, J = 5.2 Hz, 1H), 1.65–1.55 (m, 2H). *endo* isomer: δ 9.60 (d, J = 2.0 Hz, 1H), 7.30-7.06 (m, 4 H), 6.42 (dd, J = 5.6, 2.8 Hz, 2.0 Hz, 2.

3.3 Hz, 1H), 6.18 (dd, J = 5.6, 2.8 Hz, 1H), 3.36 (s, 1H), 3.24-3.10 (m, 2H), 2.93–2.91 (m, 1H), 1.76 (d, J = 8.8 Hz, 1H), 1.65–1.55 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 203.1, 202.4, 142.1, 141.2, 139.1, 136.5, 136.3, 133.8, 132.1, 131.9, 129.2, 129.1, 128.6, 128.2, 61.0, 59.6, 48.4, 48.2, 47.6, 47.1, 45.5, 45.1, 45.0, 44.7.

(1S, 2S, 3S, 4R)-3-p-Bromophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-p-Bromophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 5).^[5]



Prepared according to the general procedure

with 3-(4-bromophenyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 80% yield after silica gel chromatography (hexane:ethyl acetate=6:1); *endo* 80% ee, *exo* 74% ee. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (90/10 hexane/i-PrOH; flow rate 0.75 ml/min); endo isomer [T_R1= 20.42 min (major), T_R2 = 9.94 min (minor)], exo isomer [T_R1 = 27.55 min (major), T_R2 = 11.81 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.91 (d, J = 2.0 Hz, 1H), 7.37 (d, J = 8.4 Hz, 3H), 7.02 (d, J = 8.4 Hz, 2H), 6.36 (dd, J = 5.6, 3.2 Hz, 1H), 6.05 (dd, J = 5.6, 2.8 Hz, 1H), 3.71–3.68 (m, 1H), 3.24–3.10 (m, 1H), 3.05 (d, J = 4.8 Hz, 1H), 2.54 (d, J = 5.2 Hz, 1H), 1.65–1.55 (m, 2H). *endo* isomer: δ 9.60 (d, J = 2.0 Hz, 1H), 7.43 (d, J = 8.4 Hz, 2H), 7.15 (d, J = 8.4 Hz, 2H), 6.42 (dd, J = 5.6, 3.3 Hz, 1H), 6.18 (dd, J = 5.6, 2.8 Hz, 1H), 3.36 (s, 1H), 3.24–3.10 (m, 2H), 2.93–2.91 (m, 1H), 1.76 (d, J = 8.8 Hz, 1H), 1.65–1.55 (m, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 202.9, 202.2, 142.6, 141.6, 139.1, 136.5, 136.2, 133.8, 131.6, 131.2, 129.6, 129.1, 120.1, 120.0, 60.9, 59.5, 48.3, 48.2, 47.6, 47.0, 45.4, 45.2, 45.0, 44.8.

(1S, 2S, 3S, 4R)-3-o-Chlorophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-o-Chlorophenylbicyclo [2.2.1]hex-5-ene-2-carboxaldehyde (Table 4, entry 6).



Prepared according to the general procedure

with 3-(*o*-Chlorophenyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 88% yield after silica gel chromatography (hexane:ethyl acetate=6:1); *endo* 93% ee, *exo* 55% ee. The Products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral AD-H column (90/10 hexane/i-PrOH; flow rate 0.5 mL/min; endo isomer [T_R1= 14.72 min (major), T_R2 = 11.98 min (minor)], exo isomer (T_R1 = 13.72 min (major), T_R2 = 12.64 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.88 (d, J = 1.6 Hz, 1H), 7.38–6.97 (m, 5H), 6.40 (dd, J = 5.3, 3.2 Hz, 1H), 5.93 (dd, J = 5.3, 2.7 Hz, 1H), 4.18–4.16 (m, 1H), 3.30–3.26 (m, 2H), 2.63 (d, J = 5.2 Hz, 1H), 1.69 (d, J = 8.8 Hz, 1H), 1.56 (d, J = 8.8 Hz, 1H). *endo* isomer: δ 9.55 (d, J = 3.4 Hz, 1H), 7.38–6.97 (m, 5H), 6.47 (dd, J = 5.2, 3.4 Hz, 2H), 6.19 (dd, J = 5.5, 2.5 Hz, 1H), 3.37 (d, J = 4.4 Hz, 1H), 3.30–3.26 (m, 1H), 3.12 (s, 1H), 2.86 (dt, J = 10.0, 4.9 Hz, 1H), 1.77 (d, J = 8.8 Hz, 1H), 1.62 (d, J = 8.8 Hz, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 204.5, 202.3, 140.7, 139.3, 138.8, 136.4, 136.4, 135.0, 134.9, 134.2, 129.8, 129.4, 128.0, 127.5, 127.5, 127.1, 126.9, 126.2, 58.5, 58.1, 48.2, 47.8, 47.4, 47.3, 46.7, 46.1, 43.2, 42.0.

(1S, 2S, 3S, 4R)-3-p-Nitrophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-p-Nitrophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 7).^[5]



Prepared according to the general procedure

with 3-(4-nitrophenyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 90% yield after silica gel chromatography (hexane:ethyl acetate=6:1); *endo* 80% ee, *exo* 68% ee. The products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral AD-H column (90/10 hexane/i-PrOH; flow rate 0.5 mL/min; endo isomer [T_R1= 35.25 min (major), T_R2 = 33.81 min (minor)], exo isomer (T_R1 = 38.20 min (major), T_R2 = 47.06 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.92 (d, J = 1.5 Hz, 1H), 8.09 (d, J = 8.8 Hz, 2H), 7.29 (d, J = 8.8 Hz, 2H), 6.40 (dd, J = 5.6, 3.2 Hz, 1H), 6.05 (dd, J = 5.6, 2.8 Hz, 1H), 3.89–3.87 (m, 1H), 3.30 (s, 1H), 3.25 (s, 1H), 2.62 (d, J = 4.8 Hz, 1H), 1.62 (d, J = 5.2 Hz, 2H). *endo* isomer: δ 9.64 (d, J = 1.5 Hz, 1H), 8.15 (d, J = 8.8 Hz, 2H), 7.43 (d, J = 8.4 Hz, 2H), 6.44 (dd, J = 5.6, 3.3 Hz, 1H), 6.20 (dd, J = 5.6, 2.8 Hz, 1H), 3.43 (s, 1H), 3.21–3.18 (m, 2H), 2.98–2.94 (m, 1H), 1.77 (d, J = 8.8 Hz, 1H), 1.70 (dd, J = 8.9, 1.6 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 202.0, 201.5, 151.6, 150.5, 146.5, 146.4, 139.0, 136.9, 135.9, 133.9, 128.6, 128.2, 123.8, 123.3, 61.1, 59.5, 48.4, 47.9, 47.6, 47.1, 45.6, 45.5, 45.2, 45.0.

(1S, 2S, 3S, 4R)-3-p-Cyanophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)- 3-p-Cyanophenylbicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 8).



CN Prepared according to the general procedure with 3-(4-cyanophenyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 80% yield after silica gel chromatography (hexane:ethyl acetate=6:1); *endo* 86% ee, *exo* 63% ee. The Products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (90/10 hexane/i-PrOH; flow rate 0.75 ml/min); endo isomer [T_R1= 23.76 min (major), T_R2 = 16.74 min (minor)], exo isomer [T_R1 = 33.28 min (major), T_R2 = 19.67 min (minor)].

¹H NMR (400 MHz, CDCl₃) *exo* isomer: δ 9.91 (d, J = 1.6 Hz, 1H), 7.54 (d, J = 8.4 Hz, 2H), 7.24 (d, J = 8.0 Hz, 2H), 6.39 (dd, J = 5.6, 3.2 Hz, 1H), 6.04 (dd, J = 5.6, 2.8 Hz, 1H), 3.83–3.81 (m, 1H), 3.28 (s, 1H), 3.23 (s, 1H), 2.59 (d, J = 5.2 Hz, 1H), 1.62 (d, J = 14.0 Hz, 2H). *endo* isomer: δ 9.62 (d, J = 1.6 Hz, 1H), 7.60 (d, J = 8.0 Hz, 2H), 7.37 (d, J = 8.4 Hz, 2H), 6.43 (dd, J = 5.5, 3.3 Hz, 1H), 6.19 (dd, J = 5.6, 2.8 Hz, 1H), 3.41 (s, 1H), 3.16 (d, J = 3.2 Hz, 2H), 2.96–2.88 (m, 1H), 1.75 (d, J = 8.8 Hz, 1H), 1.68 (dd, J = 8.8, 1.5 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 202.2, 201.6, 149.4, 148.3, 139.0, 136.8, 136.0, 133.9, 132.4, 131.9, 128.6, 128.1, 118.8, 110.2, 110.1, 61.0, 59.4, 48.3, 47.8, 47.6, 47.1, 45.7, 45.4, 45.3, 44.9.

(1S, 2S, 3S, 4R)-3-(2-Furyl)bicyclo[2.2.1]hex-5-ene-2-carboxaldehyde and (1R, 2S, 3S, 4S)-3-(2-Furyl)bicyclo[2.2.1]hex-5-ene-2-carboxaldehyde (Table 2, entry 9).^[5]



with 3-(2-furyl)propenal, and cyclopentadiene. The products were collected as a pale yellow oil in 66% yield after silica gel chromatography (hexane:ethyl acetate=6:1); *endo* 73% ee, *exo* 73% ee. The Products were converted to the corresponding alcohols with NaBH₄ and the enantiomers were separated by HPLC using a Daicel chiral OJ-H column at 230 nm (90/10 hexane/i-PrOH; flow rate 0.75 ml/min); endo isomer [T_R1= 18.67 min (major), T_R2 = 10.58 min (minor)], exo isomer [T_R1 = 21.61 min (major), T_R2 = 17.59 min (minor)]. Spectroscopic data are in agreement with the published data.^[1]

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N. NMR and HPLC spectra for products







wangcha-(30-1(1-3) 10 2010413 5 mr FABEO BBC 5 mr FABEO BC 4223,465 Hz 0.11440 Hz 4223,465 Hz 0.14440 Hz 400 Lusz 400 Lusz 400 Lusz 1.0000000 sec 4 CRAINEL f1

1 HANNEL f1 14,70 usec -1.00 dB 13.75590801 W 400.1334710 MHz 32768 400.130118 MHz 0 0.30 Hz 0 1.00











wangcha-cmp-1-Br 1 2010056 6.01 5 am FAIBO B8-253 6 CT13 1 8 221.06 B8-15 8 221.08 B8-0.115483 H8-60.000 Usec 6.00 Usec 1.0000000 sec UNNEL f1

1 ANNEL f1 12.00 usec -3.00 dB 22.99425682 W 400.1320710 MHz 32769 400.1300010 MHz EM 0.30 Hz 0.100

































90 80 70 60 50 40 30 20

ppm

180 170 160 150 140 130 120 110 100

200 190



HPLC spectra



	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	9.274	926	64.06	4.16
2	14.692	8812	260.79	39.61
3	18.025	1599	50.40	7.19
4	23.182	10913	187.73	49.04



	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	9.032	8010	538.01	5.19
2	10.960	11725	696.67	7.60
3	12.241	52397	1779.38	33.96
4	16.967	82165	1672.96	53.25









	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	21.184	51342	1058.76	47.18
2	26.192	1202	29.82	1.10
3	30.121	54197	756.89	49.80
4	47.911	2090	26.48	1.92



	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	9.941	4140	154.29	5.16
2	11.812	5252	204.81	6.54
3	20.416	36160	907.93	45.05
4	27.549	34712	739.83	43.25



	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	11.975	526636	33827	2.09
2	12.637	2078794	141081	8.25
3	13.715	7247752	315944	28.77
4	14.715	15337777	611528	60.89



	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.)	(microvolt)	
1	33.814	3451714	83878	4.82
2	35.246	30960900	729467	43.20
3	38.199	31351674	635806	43.75
4	47.063	5902902	92527	8.24





	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	16.741	952	21.88	2.88
2	19.674	3511	72.65	10.62
3	23.764	13033	234.69	39.44
4	33.282	15551	224.10	47.06

min



	Retention Time	Area	Height	% Area
	(minute)	(microvolt*sec.	(microvolt)	
)		
1	10.575	2236	72.09	7.11
2	17.590	1985	67.54	6.31
3	18.670	114320	482.39	45.53
4	21.613	12912	376.53	41.05

The recyclability of Mac-CPOP-2:







mir













Run 5



Run 6

