Donor-Acceptor Conjugated Ladder Polymer via Aromatization-

Driven Thermodynamic Annulation

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1. General information

Starting materials and reagents were purchased from Aldrich, Alfa Aesar, TCI, and Acros, and were used as received without further purification unless specified. Toluene and DMF were dried using Inert Technology pure solvent system (PureSolv-MD-5a) and used without further treatment. Anhydrous THF was distilled with Na/benzophenone before uses. Anhydrous 1,2dichloroethane and 1,4-Dioxane were purchased from Acros. 4,7-dibromo-5,6bis(bromomethyl)-2,1,3-benzothiadiazole¹ 3,9-dibromo-5,11-dihydro-5,11-di(1and octylnonyl)indolo[3,2-b] carbazole (S1)² were synthesized according to reported procedures in literatures. Analytical thin-layer chromatography (TLC) tests were performed on glass that was precoated with silica gel 60-F₂₅₄ (Sorbtech). Flash column chromatography was carried out using a Biotage[®] Isolera[™] Prime with various sizes of SiO₂ Biotage ZIP[®] cartridges. UV-visible absorption spectra and fluorescence emission spectra were recorded using a Shimadzu UV-2600 and a Horiba Fluoromax-4 spectrometer, respectively. UV-visible-Near IR spectra were measured on a Hitachi U-4100 spectrometer. ¹H and ¹³C NMR spectra were obtained on a 500 MHz Varian Inova at room temperature unless specified. The NMR data were processed in MestReNova 6.1.0. Chemical shifts were reported in ppm relative to the signals corresponding to the residual non-deuterated solvents (CDCl₃: δ 7.26 for ¹H and 77.16 for ¹³C at room temperature. Size exclusion chromatography (SEC) was performed on Tosoh EcoSEC (HLC-8320GPC) in THF solution at 40 °C with a flow rate of 0.4 mL/min through TSKgel SuperHM-M and TSKgel SuperH-RC columns. The molecular weights were calculated based on the retention time, using a calibration curve constructed from polystyrene standards. Preparative SEC was performed in chloroform solution at room temperature using a JAI recycling preparative HPLC (LC-92XXII NEXT SERIES). High-resolution Thermal gravimetric analysis (TGA) was recorded under nitrogen atmosphere with heating rate of 10 °C min⁻¹ from 25 to 900 °C using TA Q500. High-resolution Matrix-assisted laser desorption/ionization (HR-MALDI) mass spectra and low-resolution atmospheric pressure chemical ionization (LR-APCI) mass spectra were measured on Applied Biosystems 4800 MALDI-TOF and a Thermo Scientific LCQ-DECA, respectively. Grazing-incidence wide-angle X-ray scattering (GIWAXS) measurements were carried out in Sector 8-ID-E at the Advanced Photon Source, Argonne National Laboratory.³ Beamline 8-ID-E operates at an energy of 10.91 keV and the images were collected from a Pilatus 1MF camera (Dectris), with two exposures for different vertical positions of the detector. Using the GIXSGUI package⁴ for MATLAB (Mathworks), data are corrected for X-ray polarization, detector sensitivity and geometrical solid-angle. The beam size is 0.8 mm × 0.5 mm and the resolution ($\Delta E/E$) is 1 × 10⁻⁴. Sample detector distance is 278 mm. Solid-state samples for GIWAXS were deposited on silicon wafers by spin-casting with the solutions (5 mg/mL in CHCl₃) at a spin rate of 1500 rpm and annealed at 250 °C or 500 °C for 30 min.

2. Synthesis



S2 (3,9-dibromo-5,11-dihydro-5,11-di(1-octylnonyl)indolo[3,2-b]carbazole-2,8-dicarb aldehyde): In a 150 mL sealed tube, DMF (8.7 mL, 112.5 mmol) was added in anhydrous 1,2-DCE (30 mL) and cooled down to 0 °C under N2. To the mixture was added phosphorus oxychloride (10.5 mL, 112.5 mmol) dropwise at 0 °C. After the reaction mixture was warmed to room temperature, compound S1 (4.0 g, 4.5 mmol) was added into the flask, and the mixture was stirred at 95 °C for 48 h. The reaction was cooled down to room temperature and quenched with water. The aqueous mixture was basified to pH = 7 by 2M KOH solution. The mixture was extracted with CH_2Cl_2 (3 × 50 mL), and the combined organic layer was dried over MgSO₄, filtered through Celite, and concentrated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, hexane:CH₂Cl₂ = 100:0 to 50:50) to give the product S2 (2.7 g, 62%) as a yellow solid. ¹H NMR (500MHz, CDCl₃): δ 10.46 (s, 2H), 8.83, 8.81 (two s[†], 2H), 8.27, 8.24, 8.13, 8.10 (four s[†], 2H), 7.86, 7.77 (two s[†], 2H), 4.68, 4.55 (two quint[†], 2H), 2.40, 2.26 (two m[†], 4H), 2.05 (m, 4H), 1.13 (m, 46H), 0.78 (t, J = 7.0 Hz, 12H). [†]Peak separations were observed due to the atropisomers by hindered rotation of α -branched alkyl chain. ¹³C NMR (125 MHz, CDCl₃): δ 191.98, 147.06, 143.82, 139.06, 138.93, 135.15, 125.15, 124.68, 124.36, 124.32, 123.72, 123.30, 123.11, 122.97, 122.76, 122.41, 122.04, 121.69, 104.08, 13.68, 103.39, 103.00, 100.96, 100.59, 57.94, 57.59, 33.77, 33.44, 31.84, 29.44, 29.23, 27.36, 26.99, 26.66, 22.70, 14.48, 14.19, 13.84. ¹³C NMR showed multiple separated peaks due to the atropisomers. HR-MALDI: calcd for $C_{54}H_{78}Br_2N_2O_2$ [M+H]⁺ m/z = 947.4488; found m/z = 947.4431.



S3 (3,9-dibromo-2,8-diethenyl-5,11-dihydro-5,11-di(1-octylnonyl)indolo[3,2-b]carbazole):

To the mixture of methyltriphenylphosphonium bromide (4.0 g, 11.2 mmol) in THF (80 mL) was added *t*-BuOK (1.18 g, 10.5 mmol) in small portions over 10 min at room temperature. Compound **S2** (2.5 g, 2.63 mmol) was added into the flask (covered with aluminum foil) with several crystals of 2,6-di-t-butyl-4-methylphenol (BHT). The mixture was stirred at reflux for 6 h. The reaction was quenched with water and diluted with CH₂Cl₂. The mixture was extracted with CH₂Cl₂ (3 × 30 mL) and the combined organic layer was dried over MgSO₄, filtered through Celite, and concentrated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, hexane) to give the product **S3** (2.5 g, 99%) as a yellow solid. ¹H NMR (500MHz, CDCl₃): δ 8.38, 8.35 (two s[†], 2H), 8.14, 8.10, 8.00, 7.97 (four s[†], 2H), 7.75, 7.59 (two s[†], 1H), 7.26 (dd, $J_1 = 17.5$ Hz, $J_2 = 11.0$ Hz, 2H), 5.82 (dd, $J_1 = 17.5$ Hz, $J_2 = 11.0$ Hz, 2H), 5.34 (d, J = 11.0 Hz, 2H), 4.67, 4.48 (two br[†], 2H), 2.42, 2.26 (two m[†], 4H), 2.00 (m, 4H), 1.13 (m, 46H), 0.79 (t, J = 7.0 Hz, 12H). [†] Peak separations were observed due to the atropisomers by hindered rotation of α -branched alkyl chain. ¹³C NMR (125 MHz, CDCl₃): δ

143.69, 142.79, 140.14, 138.25, 137.34, 137.12, 136.60, 136.36, 134.40, 127.47, 123.83, 123.55, 122.41, 122.17, 121.51, 120.92, 117.96, 117.78, 117.60, 114.77, 114.55, 113.85, 113.22, 112.49, 112.31, 105.12, 102.25, 101.95, 99.47, 99.26, 57.38, 57.02, 56.43, 33.88, 33.64, 31.88, 29.58, 29.46, 29.31, 27.04, 22.72, 14.21, 14.16. ¹³C NMR showed multiple separated peaks due to the atropisomers. HR-MALDI: calcd for $C_{56}H_{82}Br_2N_2$ [M+H]⁺ m/z = 944.4923; found m/z = 944.4905.



1 (2,8-diethenyl-3,9-bis(4,4,5,5-tetramethyl-1,3,2-dioxaborolan-2-yl)-5,11-dihydro-5,11-di(1-octylnonyl)indolo[3,2-b]carbazole): To the mixture of compound S3 (1.07 g, 1.1 mmol) in anhydrous THF (20 mL) at -78 °C was added *n*-BuLi (1.4 mL, 2.31 mmol, 1.6 M in hexane) dropwise over 1 h and stirred for another 1 h at -78 °C. 2-isopropoxy-4,4,5,5-tetramethyl-1,3,2-dioxaborolane (0.53 mL, 2.64 mmol) was added into the mixture, and the mixture was allowed to room temperature and stirred for 24 h. The reaction was quenched with water and diluted with CH₂Cl₂. The mixture was extracted with CH₂Cl₂ (3 × 30 mL) and the combined organic layer was dried over MgSO₄, filtered through Celite, and concentrated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, hexane:CH₂Cl₂ = 100:0 to 50:50) to give the product **1** (0.81g, 71%) as a yellow solid. ¹H NMR (500MHz, CDCl₃): δ 8.46 (m, 2H), 8.20 (m, 2H), 8.17, 8.05 (two s[†], 1H), 8.01 (s, 1H), 7.85 (s, 1H), 7.76 (dd, *J*₁ = 17.0 Hz, *J*₂ = 11.0 Hz, 2H), 5.85 (d, *J* = 17.0 Hz, 2H), 5.25 (d, *J* = 11.0 Hz, 2H), 4.71 (bs, 2H), 2.44, 2.37 (two m[†],

4H), 2.02 (m, 4H), 1.43 (s, 24H), 1.14 (m, 46H), 0.80 (t, J = 7.0 Hz, 12H). [†]Peak separations were observed due to the atropisomers by hindered rotation of α-branched alkyl chain. ¹³C NMR (125 MHz, CDCl₃): 151.67, 142.52, 138.76, 134.70, 133.83, 126.38, 125.67, 125.02, 124.16, 118.46, 116.21, 116.06, 115.88, 111.45, 102.36, 101.98, 99.55, 99.19, 83.73, 56.88, 56.39, 34.09, 33.77, 31.91, 29.69, 29.51, 29.38, 27.12, 26.98, 25.10, 22.73, 14.18. ¹³C NMR showed multiple separated peaks due to the atropisomers. HR-MALDI: calcd for C₆₈H₁₀₆B₂N₂O₄ [M+H]⁺ *m/z* = 1037.8417; found *m/z* = 1037.8499.



2 (4,7-dibromo-5,6-diethenyl-2,1,3-benzothiadiazole): The mixture of compound 4,7-dibromo-5,6-bis(bromomethyl)-2,1,3-benzothiadiazole (3.0 g, 6.3 mmol) and PPh₃ (4.1 g, 15.8 mmol) in DMF (40 mL) was stirred at 80 °C for 18 h. After being cooled to room temperature, the reaction mixture was concentrated under reduced pressure. The residue and paraformaldehyde (4.3 g) were suspended in THF (70 mL) under N₂. While stirring, *t*-BuOK (2.1 g, 18.9 mmol) was added in small portions over 10 min. After 30 min, the reaction was quenched with water and extracted with CH₂Cl₂ (3 × 30 mL). The combined organic layer was dried over MgSO₄, filtered through Celite, and concentrated under reduced pressure. The residue was purified by flash column chromatography (SiO₂, hexane) to give the product **S1** (1.7 g, 80%) as a white solid. ¹H NMR (500 MHz, CDCl₃): δ 6.70 (dd, $J_1 = 17.5$ Hz, $J_2 = 12.0$ Hz, 2H), 5.76 (d, $J_1 = 12.0$ Hz, J2 = 1.5 Hz, 2H), 5.61 (dd, $J_1 = 17.5$ Hz, $J_2 = 1.5$ Hz, 2H). ¹³C NMR (125 MHz, CDCl₃): δ 152.31, 140.56, 134.42, 124.49, 113.72. LR-APCI: calcd for C₁₀H₆Br₂N₂S [M+H]⁺ m/z = 346.87; found m/z = 346.93.

3. Size Exclusion Chromatography (SEC)



Fig. S1. Size exclusion chromatogram of **DACP** after purification by preparative recycling SEC. $M_n^{\text{SEC}} = 43 \text{ kg/mol}$, PDI = 4.34. It is worthy to note that aggregation feature was observed as a shoulder peak in the high molecular weight region.



Fig. S2. Size exclusion chromatogram of DALP after Soxhlet extraction from CHCl₃. $M_n^{\text{SEC}} = 21 \text{ kg/mol}$, PDI = 2.95.

4. Lippert-Mataga Solvatochromism Plot

The positive solvatochromism of **DALP** was demonstrated by a Lippert-Mataga plot using the Lippert-Mataga equation (1).⁵

$$\bar{\nu}_A - \bar{\nu}_E = \frac{2(\mu_G - \mu_E)^2}{hca^3}\Delta f + c \tag{1}$$

Herein, v_A and v_E are the absorption maximum and emission maximum in wavenumbers, μ_g and μ_E are, respectively, the ground and excited state dipole moments, *h* is Planck's constant, *c* is the speed of light in vacuum, *a* is the Onsager cavity radius, and Δf is the orientation polarizability. Stokes shift is generally described by the difference of absorption maximum and emission maximum. However, since **DALP** exhibits optically weak HOMO-LUMO transitions in the absorption spectrum, the Stokes shift of **DALP** represents the difference of the weak HOMO-LUMO absorption and the emission maximum. Unfortunately, it was difficult to obtain an accurate wavelength of the weak HOMO-LUMO absorptions in various solvents because the weak absorption shoulder (468 nm) observed in toluene became less characteristic in more polar solvents. Although the difference between absorption maxima and emission maxima does not depicts Stokes shift of **DALP**, the difference, which is more apparent, was plotted as a function of Δf , which is defined as the equation (2).⁵

$$\Delta f = \left(\frac{\varepsilon - 1}{2\varepsilon + 1}\right) - \left(\frac{n^2 - 1}{2n^2 + 1}\right) \tag{2}$$

where, ε is dielectric constant, and *n* is refractive index of a solvent.

Table S1. Summary of dielectric constants (ε) ,⁶ refractive indices (n),⁶ orientation polarizabilities (Δf) , absorptions, emissions, and the difference between the absorptions and emissions in various solvents.

	3	п	Δf	$\lambda_{abs}(nm)$	$\lambda_{em}(nm)$	$\Delta v (\text{cm}^{-1})$
Toluene	2.38	1.4969	0.013	468	569	3792
Chlorobenzene	5.62	1.5248	0.143	469	591	4401
Chloroform	4.81	1.4459	0.148	467	590	4464
1,2-Dichlorobenzene	9.93	1.5514	0.186	470	608	4829
Tetrahydrofuran	7.58	1.4072	0.210	468	604	4811
Dichloromethane	8.93	1.4242	0.217	466	616	5225
1,2-Dichloroethane	10.37	1.4448	0.221	466	618	5278







Fig. S4. Lippert-Mataga plot of DALP. The red line represents the linear fit ($R^2 = 0.8821$).

5. Charge Transfer Complex







Fig. S6. UV-vis-NIR spectra of DALP doped with various amounts of F_4TCNQ (from 0 wt% to 23.2 wt%).



Fig. S7. Structural formula of CP, which is the polymeric precursor of LP.

6. Grazing Incidence Wide-Angle X-ray Scattering (GIWAXS)



Fig. S8. GIWAXS image of as-cast film of DALP.

7. Cyclic Voltammetry (CV)



Fig. S9. Cyclic Voltammograms of DALP in DCM (0.1M [*n*-Bu₄N]PF₆) at scan rate of 20 mV/s.

8. ¹H and ¹³C NMR Spectra



Fig. S10. ¹H NMR of S2 (500 MHz, CDCl₃, RT).



Fig. S11. ¹³C NMR of S2 (125 MHz, CDCl₃, RT).



Fig. S12. ¹H NMR of S3 (500 MHz, CDCl₃, RT).



Fig. S13. ¹³C NMR of S3 (125 MHz, CDCl₃, RT).



Fig. S14. ¹H NMR of 1 (500 MHz, CDCl₃, RT).



Fig. S15. ¹³C NMR of **1** (125 MHz, CDCl₃, RT).



Fig. S16. ¹H NMR of **2** (500 MHz, CDCl₃, RT).





Fig. S17. ¹³C NMR of 2 (125 MHz, CDCl₃, RT).



Fig. S18. ¹H NMR of DACP (500 MHz, CDCl₃, RT).



12.0 11.5 11.0 10.5 10.0 6.0 5.5 5.0 (ppm) 1.5 1.0 9.5 9.0 8.5 8.0 7.5 7.0 6.5 4.5 4.0 3.5 3.0 2.5 2.0 0.5 0.0 -0.!

Fig S19. ¹H NMR of DALP (500 MHz, CDCl₃, RT).

9. Reference

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