

Supporting Information

Nickel-Catalyzed Selective C-5 Fluorination of 8-Aminoquinolines with NFSI

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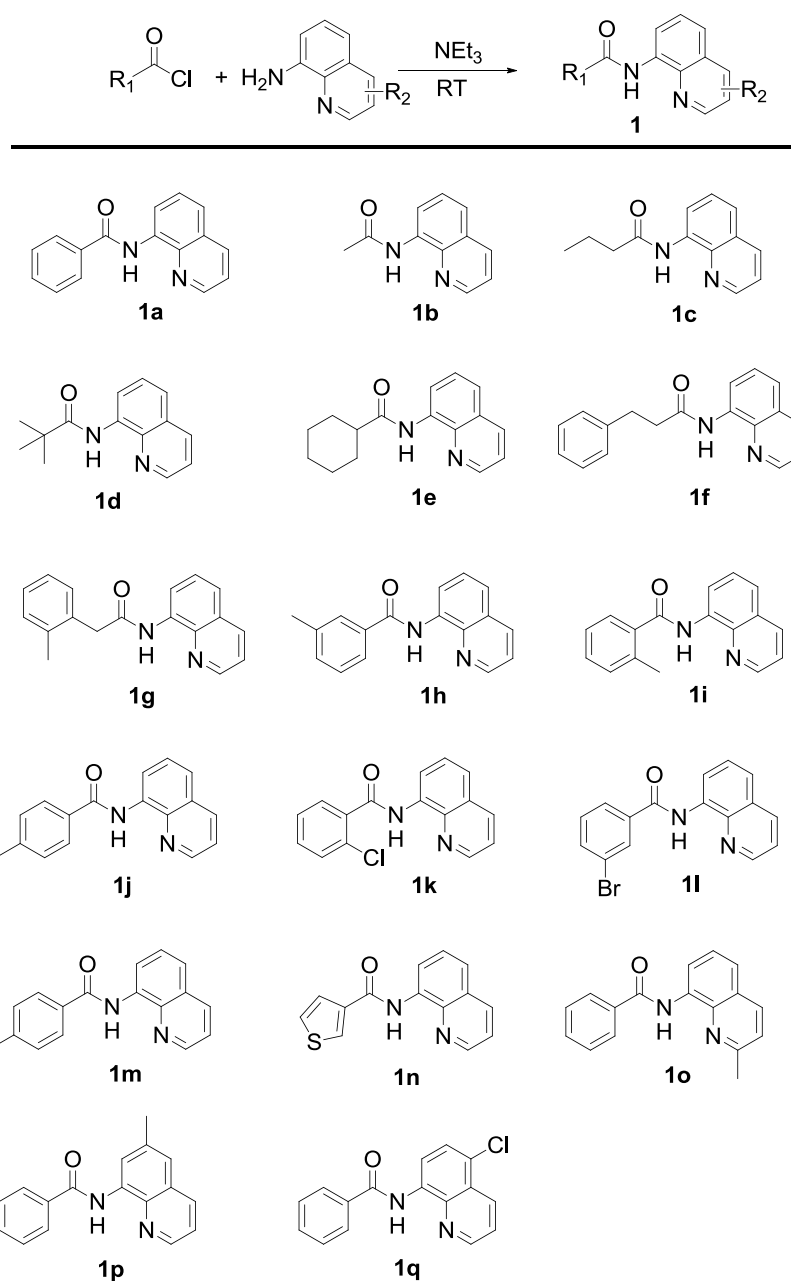
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1. General information

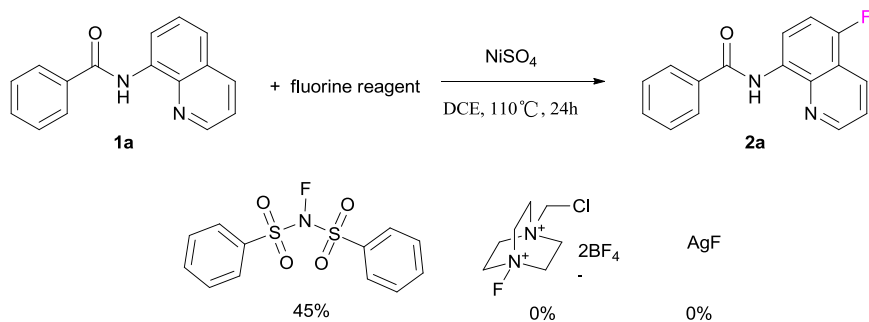
All reagents were purchased from commercial sources and used without treatment, unless otherwise indicated. All the solvents were dried and newly distilled. NMR spectra were obtained on a Bruker AMX 400 system using chloroform-d as deuterated solvents. The ^1H -NMR spectra were recorded at 400 MHz in CDCl_3 , the ^{13}C -NMR spectra were recorded at 100 MHz in CDCl_3 , and the ^{19}F -NMR spectra were recorded at 376 MHz in CDCl_3 with TMS as internal standard. All shifts were given in ppm. All coupling constants (J values) were reported in Hertz (Hz). Single crystal X-ray diffraction data were collected using a Bruker-AXS SMART APEX2 CCD diffractometer (Mo $K\alpha$, $\lambda = 0.71073 \text{ \AA}$). Column chromatography was performed on silica gel 100-200 mesh or 200-300 mesh. Ethyl acetate and petroleum ether were used for column chromatography.

2. Preparation of substrated amides 1a-1q.¹

Preparation of amides **1a-1q**: A solution of 8-aminoquinoline (1.44 g, 10.0 mmol) and NEt₃ (1.01g, 11.0 mmol) in dichloromethane (10 mL) was added dropwise to a stirring solution of an acid chloride (11.0 mmol) in dichloromethane (40 mL). The resulting mixture was stirred at 25°C for 3 hours. Then, the mixture was quenched with saturated aq. NaHCO₃ (50mL), and was extracted with dichloromethane for three times (3 x 50 mL). The organic layer was dried over Na₂SO₄. After filtration and evaporation, the amides were purified by column chromatography on silica gel.

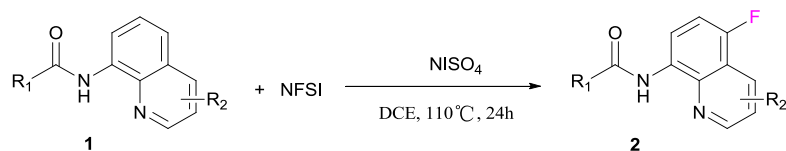


3. Screening of fluorine reagents



N-(8-quinoliny) benzamide **1a** (0.2 mmol, 1.0 equiv), fluorine reagent (0.4 mmol, 2.0 equiv), NiSO_4 (0.04 mmol, 0.2 equiv) were mixed in DCE (1.0 mL) and stirred for 24 h in a sealed tube at 110°C . The resulting organic solution was concentrated under reduced pressure and further purified by flash chromatography (SiO_2 , petroleum ether/ethyl acetate gradient), yielding the target product **2a**.

4. General procedure for preparation of regioselective C-5 fluorinated N-(8-quinolinyl) amides



N-(8-quinolinyl) amide **1** (0.2 mmol, 1.0 equiv), NFSI (0.4 mmol, 2.0 equiv), NiSO₄ (0.04 mmol, 0.2 equiv) were mixed in DCE (1.0 mL) and stirred for 24 h in a sealed tube at 110 °C. The resulting organic solution was concentrated under reduced pressure and further purified by flash chromatography (SiO₂, petroleum ether/ethyl acetate gradient), yielding the target product **2** and yields in parentheses were determined by ¹⁹F NMR spectroscopy with fluorobenzene as an internal standard.

5. The single crystal X-ray diffraction study of N-(5-fluoro-quinolin-8-yl) benzamide **2a**.

Single-crystal X-ray structure of **2a**

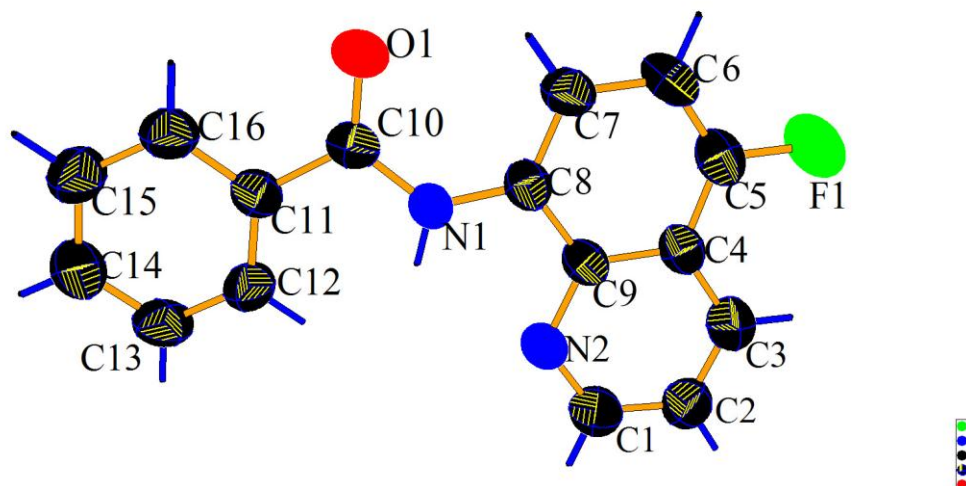


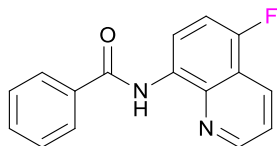
Table 1. Crystal data and structure refinement for **2a**.

Identification code	2a
Empirical formula	C ₁₆ H ₁₁ F N ₂ O
Formula weight	266.27
Temperature	273(2) K
Wavelength	0.71073 Å
Crystal system, space group	Monoclinic, P 2 ₁ /c
Unit cell dimensions	a = 21.229(10) Å alpha = 90 deg. b = 3.9190(19) Å beta = 105.861(11) deg. c = 15.628(8) Å gamma = 90 deg.
Volume	1250.7(10) Å ³
Z, Calculated density	4, 1.414 Mg/m ³
Absorption coefficient	0.100 mm ⁻¹

F(000)	552
Crystal size	0.220 x 0.200 x 0.180 mm
Theta range for data collection	0.997 to 25.654 deg.
Limiting indices	-25<=h<=25, -4<=k<=4, -19<=l<=14
Reflections collected / unique	7250 / 2342 [R(int) = 0.0644]
Completeness to theta = 25.242	99.6 %
Absorption correction	Semi-empirical from equivalents
Refinement method	Full-matrix least-squares on F ²
Data / restraints / parameters	2342 / 0 / 181
Goodness-of-fit on F ²	0.994
Final R indices [I>2sigma(I)]	R1 = 0.0737, wR2 = 0.2007
R indices (all data)	R1 = 0.1172, wR2 = 0.2307
Extinction coefficient	n/a
Largest diff. peak and hole	0.212 and -0.184 e.A ⁻³

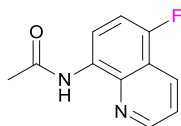
6. Characterization data of the products

2a:



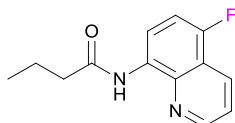
White solid, isolated yield: 45%; ^{19}F NMR yield: 60%. ^1H NMR (400 MHz, CDCl_3) δ : 10.57 (s, 1H), 8.96-8.90 (m, 2H), 8.50 (dd, $J = 8.6$ Hz, 5.5 Hz, 1H), 8.10 (d, $J = 7.0$ Hz, 2H), 7.67-7.55 (m, 4H), 7.32 (t, $J = 9.2$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ : 165.34 (s), 153.01 (d, $J = 251.2$ Hz), 149.14 (s), 138.97 (d, $J = 3.0$ Hz), 134.99 (s), 131.91 (s), 131.12 (d, $J = 4.0$ Hz), 129.86 (d, $J = 3.6$ Hz), 128.83 (s), 127.24 (s), 121.77 (d, $J = 2.6$ Hz), 118.82 (d, $J = 18.3$ Hz), 116.04 (d, $J = 7.7$ Hz), 110.52 (d, $J = 19.7$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.00 (s).

2b:



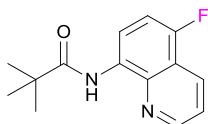
White solid, isolated yield: 40%; ^{19}F NMR yield: 52%. ^1H NMR (400 MHz, CDCl_3) δ : 9.62 (s, 1H), 8.88 (d, $J = 4.1$ Hz, 1H), 8.74 (dd, $J = 8.6$, 5.5 Hz, 1H), 8.46 (d, $J = 8.4$ Hz, 1H), 7.55 (dd, $J = 8.4$, 4.2 Hz, 1H), 7.25 (t, $J = 9.2$ Hz, 1H), 2.37 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ : 168.61 (s), 152.80 (d, $J = 250.8$ Hz), 148.94 (s), 138.44 (d, $J = 2.1$ Hz), 131.04 (d, $J = 4.1$ Hz), 129.82 (d, $J = 3.7$ Hz), 121.64 (d, $J = 2.5$ Hz), 118.71 (d, $J = 18.1$ Hz), 115.94 (d, $J = 7.6$ Hz), 110.42 (d, $J = 19.6$ Hz), 25.03 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.40 (s).

2c:



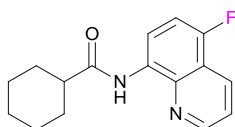
White solid, isolated yield: 44%; ^{19}F NMR yield: 58%. ^1H NMR (400 MHz, CDCl_3) δ : 9.65 (s, 1H), 8.88 (d, $J = 4.2$ Hz, 1H), 8.77 (dd, $J = 8.4$, 5.6 Hz, 1H), 8.46 (d, $J = 8.4$ Hz, 1H), 7.55 (dd, $J = 8.4$, 4.2 Hz, 1H), 7.25 (t, $J = 9.2$ Hz, 1H), 2.56 (t, $J = 7.5$ Hz, 2H), 1.94-1.82 (m, 2H), 1.08 (t, $J = 7.3$ Hz, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ : 171.58 (s), 152.74 (d, $J = 250.6$ Hz), 148.93 (s), 138.54 (d, $J = 2.9$ Hz), 131.10 (d, $J = 3.9$ Hz), 129.78 (d, $J = 3.6$ Hz), 121.60 (d, $J = 2.6$ Hz), 118.72 (d, $J = 18.0$ Hz), 115.86 (d, $J = 7.6$ Hz), 110.43 (d, $J = 19.6$ Hz), 40.09 (s), 19.13 (s), 13.82 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.55 (s).

2d:



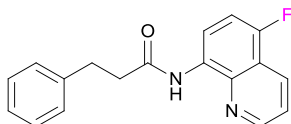
White solid, isolated yield: 46%; ^{19}F NMR yield: 60%. ^1H NMR (400 MHz, CDCl_3) δ : 10.09 (s, 1H), 8.89 (dd, $J = 4.2, 1.5$ Hz, 1H), 8.77 (dd, $J = 8.6, 5.5$ Hz, 1H), 8.46 (dd, $J = 8.4, 1.5$ Hz, 1H), 7.55 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.23 (t, $J = 9.2$ Hz, 1H), 1.45 (s, 9H). ^{13}C NMR (100 MHz, CDCl_3) δ : 177.10 (s), 152.71 (d, $J = 250.5$ Hz), 149.04 (s), 138.94 (d, $J = 3.0$ Hz), 131.22 (d, $J = 4.1$ Hz), 129.76 (d, $J = 3.6$ Hz), 121.59 (d, $J = 2.5$ Hz), 118.72 (d, $J = 18.1$ Hz), 115.65 (d, $J = 7.6$ Hz), 110.42 (d, $J = 19.4$ Hz), 40.27 (s), 27.73 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.81 (s).

2e:



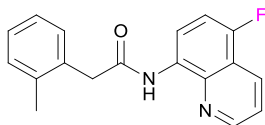
White solid, isolated yield: 46%; ^{19}F NMR yield: 62%. ^1H NMR (400 MHz, CDCl_3) δ : 9.78 (s, 1H), 8.90 (dd, $J = 4.3, 1.6$ Hz, 1H), 8.79 (dd, $J = 8.7, 5.5$ Hz, 1H), 8.50 (dd, $J = 8.4, 1.5$ Hz, 1H), 7.58 (dd, $J = 8.4, 4.3$ Hz, 1H), 7.25 (t, $J = 9.2$ Hz, 1H), 2.55-2.46 (m, 1H), 2.21-2.03 (m, 2H), 1.99-1.83 (m, 2H), 1.81-1.59 (m, 3H), 1.49-1.26 (m, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ : 174.71 (s), 152.68 (d, $J = 250.7$ Hz), 148.89 (s), 138.61 (d, $J = 2.6$ Hz), 131.16 (d, $J = 4.0$ Hz), 129.81 (d, $J = 3.7$ Hz), 121.58 (d, $J = 2.5$ Hz), 118.71 (d, $J = 18.0$ Hz), 115.90 (d, $J = 7.6$ Hz), 110.45 (d, $J = 19.5$ Hz), 46.82 (s), 29.75 (s), 25.77 (s), 25.75 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.73 (s).

2f:



White solid, isolated yield: 41%; ^{19}F NMR yield: 54%. ^1H NMR (400 MHz, CDCl_3) δ : 9.61 (s, 1H), 8.85 (dd, $J = 4.2, 1.6$ Hz, 1H), 8.76 (dd, $J = 8.6, 5.4$ Hz, 1H), 8.45 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.54 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.36-7.30 (m, 4H), 7.27-7.19 (m, 2H), 3.22-3.12 (t, $J = 7.8$ Hz, 2H), 2.95-2.85 (t, $J = 7.8$ Hz, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ : 170.57 (s), 152.82 (d, $J = 250.9$ Hz), 148.92 (s), 140.71 (s), 138.51 (d, $J = 3.0$ Hz), 130.96 (d, $J = 4.1$ Hz), 129.78 (d, $J = 3.7$ Hz), 128.57 (s), 128.39 (s), 126.27 (s), 121.63 (d, $J = 2.5$ Hz), 118.71 (d, $J = 18.1$ Hz), 115.96 (d, $J = 7.7$ Hz), 110.43 (d, $J = 19.6$ Hz), 39.66 (s), 31.48 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.34 (s).

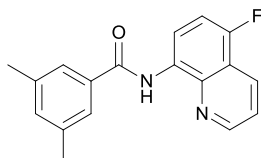
2g:



White solid, isolated yield: 39%; ^{19}F NMR yield: 45%. ^1H NMR (400 MHz, CDCl_3) δ : 9.67 (s, 1H), 8.76-8.68 (m, 2H), 8.40 (dd, $J = 8.4, 1.6$ Hz, 1H), 7.48 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.41-.29 (m, 5H), 7.20

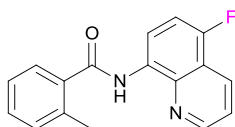
(t, $J = 9.2$ Hz, 1H), 3.92 (s, 2H), 2.42 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ : 169.21 (s), 152.90 (d, $J = 250.9$ Hz), 149.00 (s), 138.71 (d, $J = 2.7$ Hz), 137.25 (s), 133.09 (s), 130.91 (d, $J = 4.0$ Hz), 130.81 (s), 130.59 (s), 129.59 (d, $J = 3.6$ Hz), 127.83 (s), 126.64 (s), 121.54 (d, $J = 2.6$ Hz), 118.62 (d, $J = 18.2$ Hz), 115.75 (d, $J = 7.8$ Hz), 110.32 (d, $J = 19.6$ Hz), 43.21 (s), 19.71 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.20 (s).

2h:



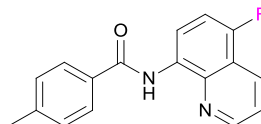
White solid, isolated yield: 43%; ^{19}F NMR yield: 53%. ^1H NMR (400 MHz, CDCl_3) δ : 10.49 (s, 1H), 8.98 – 8.89 (m, 2H), 8.49 (d, $J = 8.4$ Hz, 1H), 7.68 (s, 2H), 7.58 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.30 (t, $J = 9.2$ Hz, 1H), 7.24 (s, 1H), 2.47 (s, 6H). ^{13}C NMR (100 MHz, CDCl_3) δ : 165.80 (s), 152.94 (d, $J = 251.0$ Hz), 149.11 (s), 138.99 (d, $J = 3.1$ Hz), 138.51 (s), 135.05 (s), 133.53 (s), 131.25 (d, $J = 4.2$ Hz), 129.82 (d, $J = 3.7$ Hz), 124.98 (s), 121.70 (d, $J = 2.6$ Hz), 118.81 (d, $J = 18.2$ Hz), 116.04 (d, $J = 7.7$ Hz), 110.52 (d, $J = 19.6$ Hz), 21.39 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.26 (s).

2i:



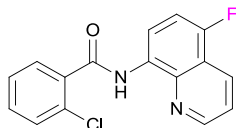
White solid, isolated yield: 44%; ^{19}F NMR yield: 57%. ^1H NMR (400 MHz, CDCl_3) δ : 10.06 (s, 1H), 8.93 (dd, $J = 8.5, 5.5$ Hz, 1H), 8.86 (d, $J = 4.1$ Hz, 1H), 8.50 (dd, $J = 8.4, 1.4$ Hz, 1H), 7.70 (d, $J = 7.6$ Hz, 1H), 7.57 (dd, $J = 8.4, 4.3$ Hz, 1H), 7.48-7.29 (m, 4H), 2.62 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ : 168.06 (s), 153.05 (d, $J = 251.3$ Hz), 149.12 (s), 138.82 (d, $J = 3.1$ Hz), 136.71 (s), 136.46 (s), 131.42 (s), 131.26 (d, $J = 4.0$ Hz), 130.41 (s), 129.81 (d, $J = 3.6$ Hz), 127.23 (s), 126.05 (s), 121.75 (d, $J = 2.5$ Hz), 118.80 (d, $J = 18.1$ Hz), 116.01 (d, $J = 7.7$ Hz), 110.46 (d, $J = 19.6$ Hz), 20.24 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -128.93 (s).

2j:



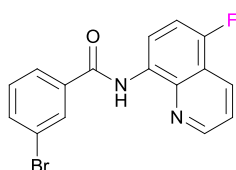
White solid, isolated yield: 45%; ^{19}F NMR yield: 58%. ^1H NMR (400 MHz, CDCl_3) δ : 10.54 (s, 1H), 8.99-8.88 (m, 2H), 8.49 (d, $J = 8.4$ Hz, 1H), 7.99 (d, $J = 7.7$ Hz, 2H), 7.58 (dd, $J = 8.3, 4.2$ Hz, 1H), 7.38 (d, $J = 7.7$ Hz, 2H), 7.30 (t, $J = 9.2$ Hz, 1H), 2.48 (s, 3H). ^{13}C NMR (101 MHz, CDCl_3) δ : 165.32 (s), 152.92 (d, $J = 251.1$ Hz), 149.08 (s), 142.40 (s), 138.97 (d, $J = 2.8$ Hz), 132.19 (s), 131.24 (d, $J = 4.0$ Hz), 129.83 (d, $J = 3.8$ Hz), 129.48 (s), 127.25 (s), 121.71 (d, $J = 2.4$ Hz), 118.82 (d, $J = 18.1$ Hz), 115.94 (d, $J = 7.6$ Hz), 110.53 (d, $J = 19.6$ Hz), 21.56 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.27 (s).

2k:



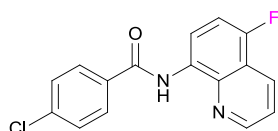
White solid, isolated yield: 43%; ^{19}F NMR yield: 54%. ^1H NMR (400 MHz, CDCl_3) δ : 10.35 (s, 1H), 8.94 (dd, $J = 8.6, 5.4$ Hz, 1H), 8.88 (dd, $J = 4.2, 1.5$ Hz, 1H), 8.49 (dd, $J = 8.4, 1.5$ Hz, 1H), 7.85 (dd, $J = 7.1, 2.1$ Hz, 1H), 7.60-7.51 (m, 2H), 7.51-7.41 (m, 2H), 7.30 (d, $J = 9.2$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ : 164.72 (s), 153.26 (d, $J = 251.7$ Hz), 149.23 (s), 138.92 (d, $J = 3.2$ Hz), 135.63 (s), 131.60 (s), 131.15 (s), 130.97 (d, $J = 4.2$ Hz), 130.55 (s), 130.14 (s), 129.79 (d, $J = 3.7$ Hz), 127.19 (s), 121.79 (d, $J = 2.7$ Hz), 118.81 (d, $J = 18.1$ Hz), 116.47 (d, $J = 7.8$ Hz), 110.46 (d, $J = 19.7$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ : -128.40 (s)

2l:



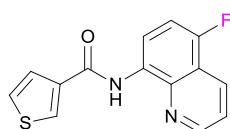
White solid, isolated yield: 41%; ^{19}F NMR yield: 50%. ^1H NMR (400 MHz, CDCl_3) δ : 10.51 (s, 1H), 8.95 (d, $J = 4.2$ Hz, 1H), 8.89 (dd, $J = 8.6, 5.4$ Hz, 1H), 8.51 (d, $J = 8.5$ Hz, 1H), 8.23 (s, 1H), 8.01 (d, $J = 7.5$ Hz, 1H), 7.75 (d, $J = 7.9$ Hz, 1H), 7.60 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.49-7.30 (m, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ : 163.76 (s), 153.21 (d, $J = 251.8$ Hz), 149.26 (s), 138.95 (s), 136.99 (s), 134.85 (s), 130.78 (d, $J = 4.0$ Hz), 130.57 (s), 130.34 (s), 129.92 (d, $J = 3.6$ Hz), 125.66 (s), 123.08 (s), 121.85 (d, $J = 2.5$ Hz), 118.84 (d, $J = 18.3$ Hz), 116.27 (d, $J = 7.8$ Hz), 110.50 (d, $J = 19.8$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ : -128.39 (s).

2m:



White solid, isolated yield: 33%; ^{19}F NMR yield: 42%. ^1H NMR (400 MHz, CDCl_3) δ : 10.53 (s, 1H), 8.94 (d, $J = 4.1$ Hz, 1H), 8.89 (dd, $J = 8.6, 5.5$ Hz, 1H), 8.50 (d, $J = 8.4$ Hz, 1H), 8.04 (d, $J = 8.4$ Hz, 2H), 7.64-7.52 (m, 3H), 7.30 (d, $J = 9.1$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ : 164.20 (s), 153.14 (d, $J = 251.6$ Hz), 149.20 (s), 138.93 (d, $J = 3.1$ Hz), 138.19 (s), 133.38 (s), 130.88 (d, $J = 4.0$ Hz), 129.94 (d, $J = 3.6$ Hz), 129.10 (s), 128.67 (s), 121.83 (d, $J = 2.5$ Hz), 118.85 (d, $J = 18.3$ Hz), 116.15 (d, $J = 7.7$ Hz), 110.53 (d, $J = 19.7$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ : -128.57 (s).

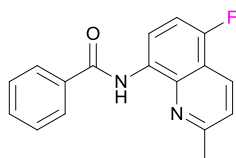
2n:



White solid, isolated yield: 32%; ^{19}F NMR yield: 40%. ^1H NMR (400 MHz, CDCl_3) δ : 10.36 (s, 1H),

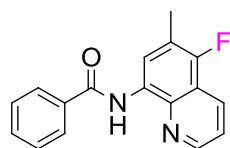
8.92 (d, $J = 4.2$ Hz, 1H), 8.86 (dd, $J = 8.6, 5.4$ Hz, 1H), 8.49 (d, $J = 8.4$ Hz, 1H), 8.22-8.15 (m, 1H), 7.70 (d, $J = 5.0$ Hz, 1H), 7.58 (dd, $J = 8.4, 4.2$ Hz, 1H), 7.47 (dd, $J = 5.0, 3.0$ Hz, 1H), 7.30 (t, $J = 9.2$ Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ : 160.90 (s), 152.96 (d, $J = 251.2$ Hz), 149.11 (s), 138.79 (d, $J = 3.0$ Hz), 138.19 (s), 131.02 (d, $J = 4.0$ Hz), 129.85 (d, $J = 3.6$ Hz), 128.93 (s), 126.78 (s), 126.27 (s), 121.74 (d, $J = 2.5$ Hz), 118.82 (d, $J = 18.2$ Hz), 115.99 (d, $J = 7.6$ Hz), 110.53 (d, $J = 19.7$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.10 (s).

2o:



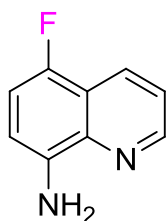
White solid, isolated yield: 42%; ^{19}F NMR yield: 55%. ^1H NMR (400 MHz, CDCl_3) δ : 10.62 (s, 1H), 8.88 (dd, $J = 8.6, 5.5$ Hz, 1H), 8.36 (d, $J = 8.5$ Hz, 1H), 8.09 (d, $J = 6.9$ Hz, 2H), 7.65-7.56 (m, 3H), 7.44 (d, $J = 8.5$ Hz, 1H), 7.21 (t, $J = 9.2$ Hz, 1H), 2.82 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ : 165.17 (s), 158.32 (s), 153.28 (d, $J = 250.7$ Hz), 138.52 (s), 135.19 (s), 131.79 (s), 130.46 (d, $J = 4.2$ Hz), 129.88 (d, $J = 3.4$ Hz), 128.83 (s), 127.19 (s), 122.55 (d, $J = 2.4$ Hz), 116.87 (d, $J = 18.2$ Hz), 116.03 (d, $J = 7.8$ Hz), 109.52 (d, $J = 19.7$ Hz), 25.50 (s). ^{19}F NMR (376 MHz, CDCl_3) δ : -129.19 (s).

2p:



White solid, isolated yield: 45%; ^{19}F NMR yield: 59%. ^1H NMR (400 MHz, CDCl_3) δ 10.54 (s, 1H), 8.85 (m, 2H), 8.44 (dd, $J = 8.4, 1.4$ Hz, 1H), 8.10 (dd, $J = 7.8, 1.4$ Hz, 2H), 7.76-7.40 (m, 4H), 2.53 (d, $J = 2.4$ Hz, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 165.30 (s), 150.44 (d, $J = 248.1$ Hz), 148.09 (s), 137.79 (d, $J = 2.8$ Hz), 135.05 (s), 131.86 (s), 130.46 (d, $J = 4.1$ Hz), 129.31 (d, $J = 4.2$ Hz), 128.83 (s), 127.23 (s), 121.72 (d, $J = 2.6$ Hz), 118.82 (d, $J = 18.2$ Hz), 115.99 (d, $J = 7.6$ Hz), 110.53 (d, $J = 19.7$ Hz), 14.82 (d, $J = 3.6$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ -133.73 (s).

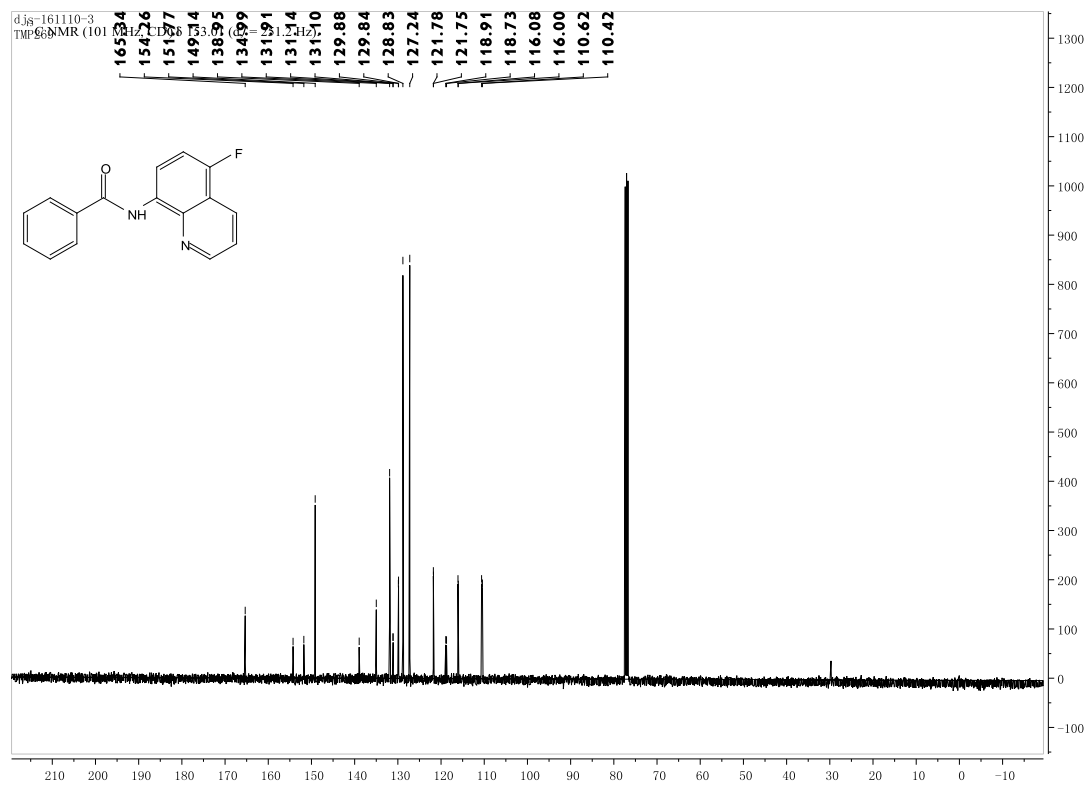
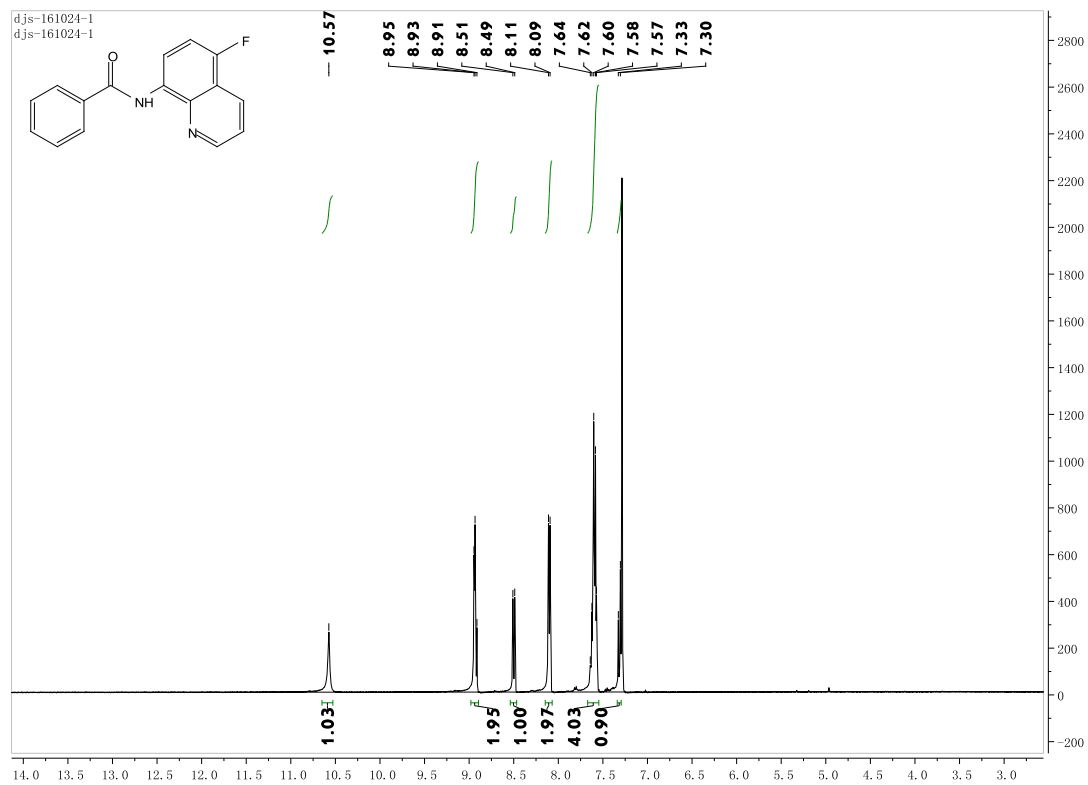
9:

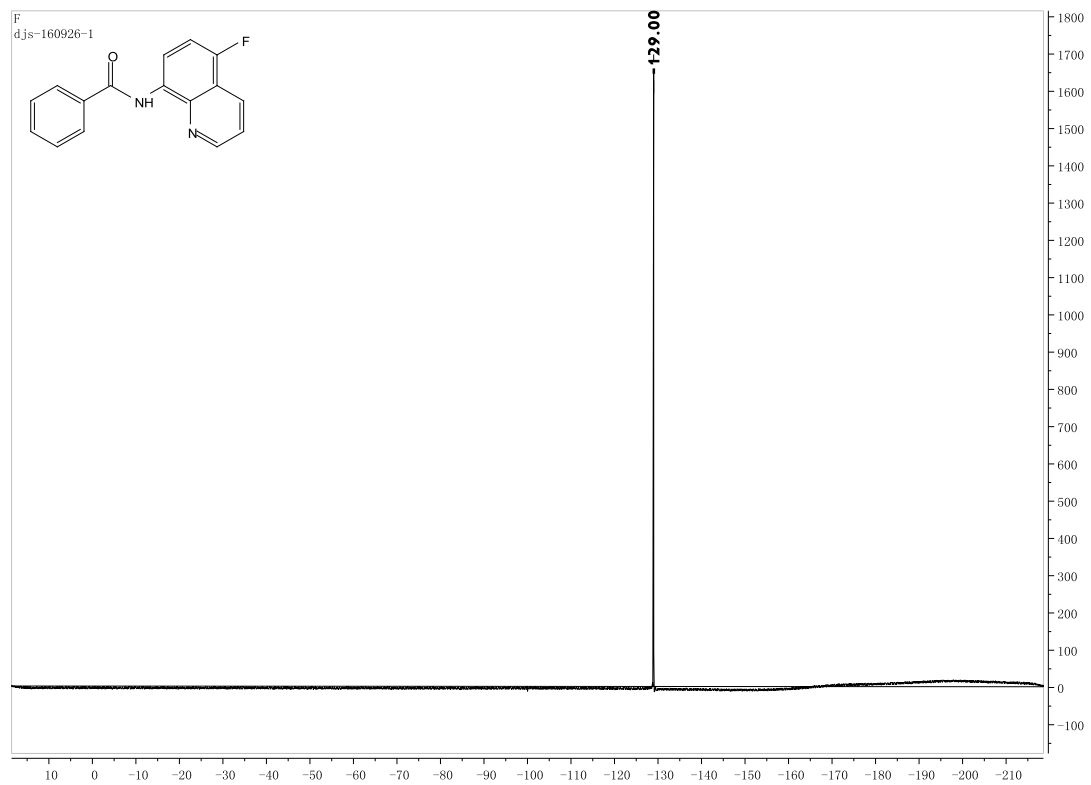


Yellow solid. ^1H NMR (400 MHz, CDCl_3) δ : 8.84 (dd, $J = 4.2, 1.7$ Hz, 1H), 8.37 (dd, $J = 8.5, 1.7$ Hz, 1H), 7.47 (dd, $J = 8.5, 4.2$ Hz, 1H), 7.06 (dd, $J = 9.9, 8.3$ Hz, 1H), 6.84 (dd, $J = 8.3, 4.9$ Hz, 1H), 4.85 (s, 2H). ^{13}C NMR (100 MHz, CDCl_3) δ : 149.97 (d, $J = 243.4$ Hz), 148.17 (s), 140.25 (d, $J = 3.2$ Hz), 137.99 (d, $J = 2.6$ Hz), 129.38 (d, $J = 3.7$ Hz), 121.31 (d, $J = 2.7$ Hz), 119.27 (d, $J = 17.7$ Hz), 110.57 (d, $J = 20.1$ Hz), 108.50 (d, $J = 7.5$ Hz). ^{19}F NMR (376 MHz, CDCl_3) δ : -137.37 (s).

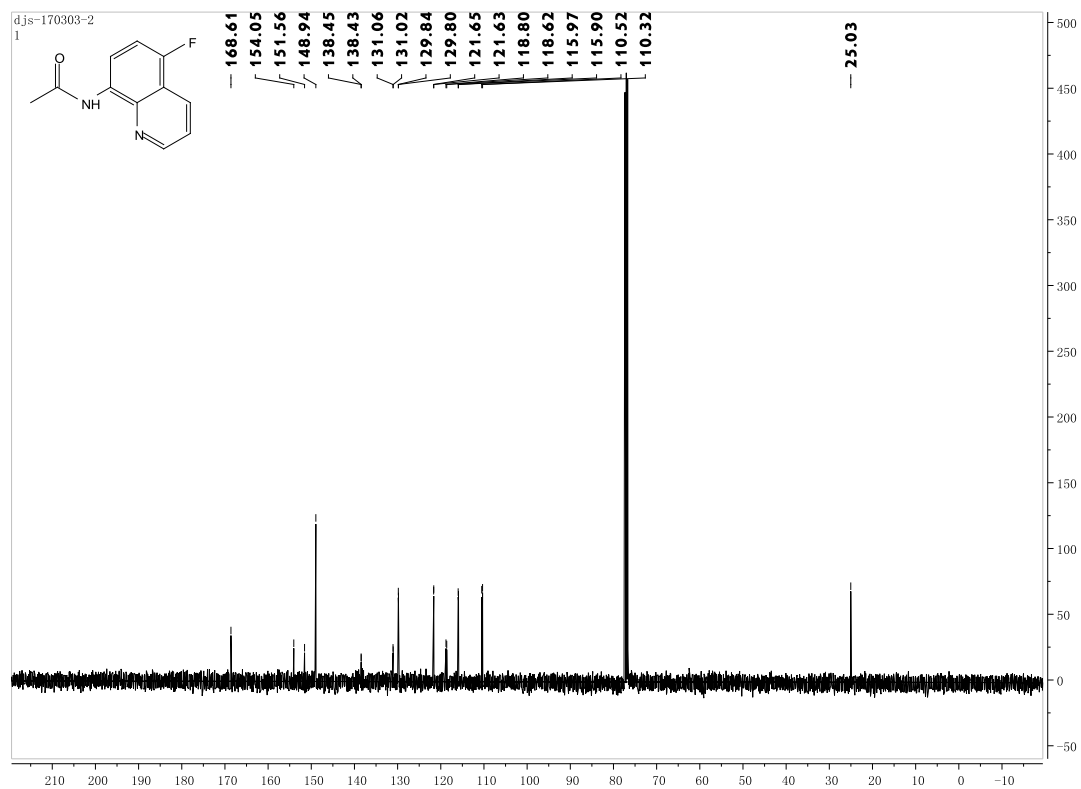
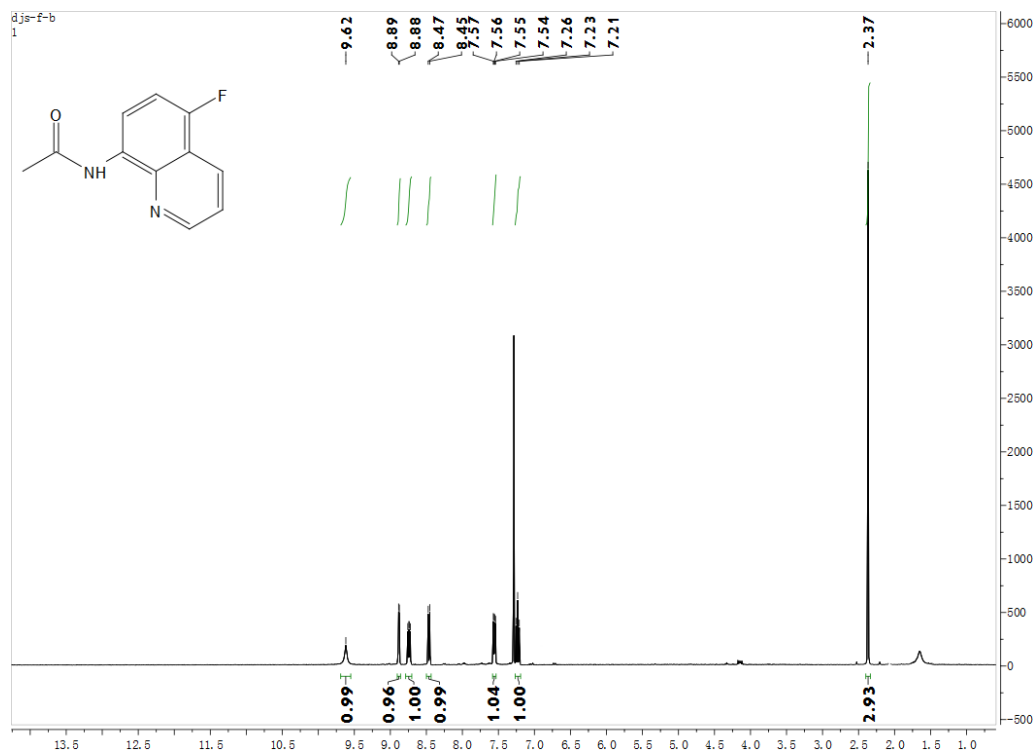
7. Spectra for the Products

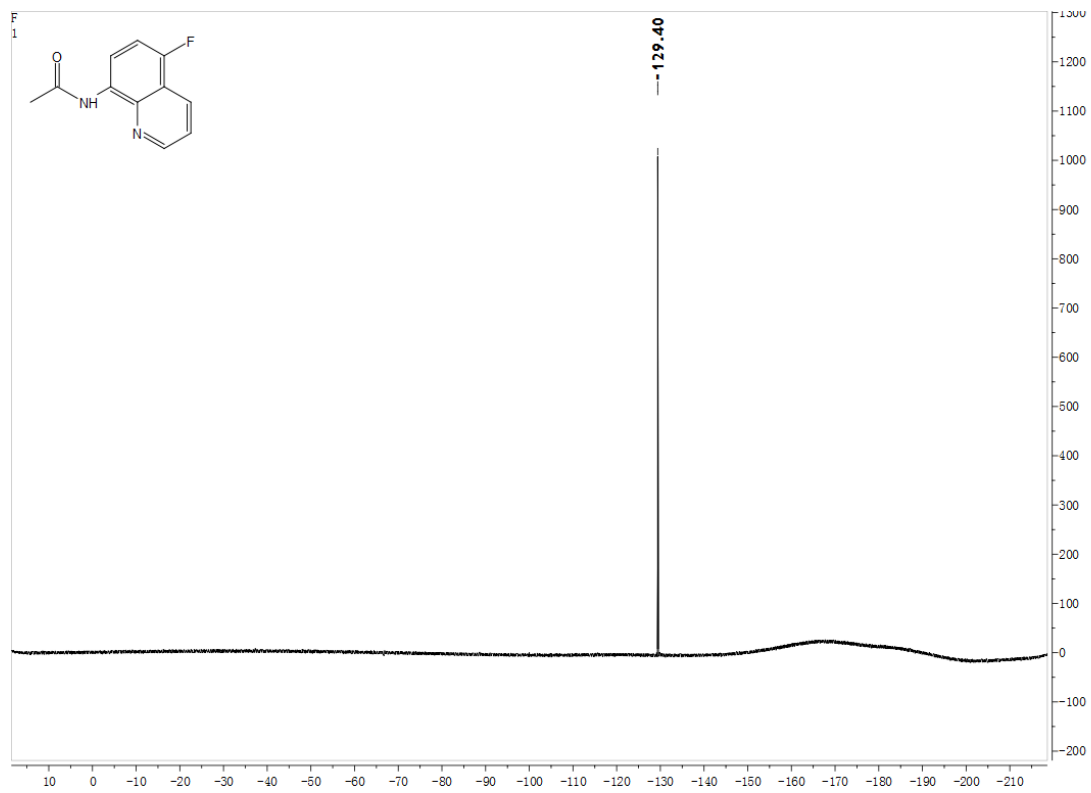
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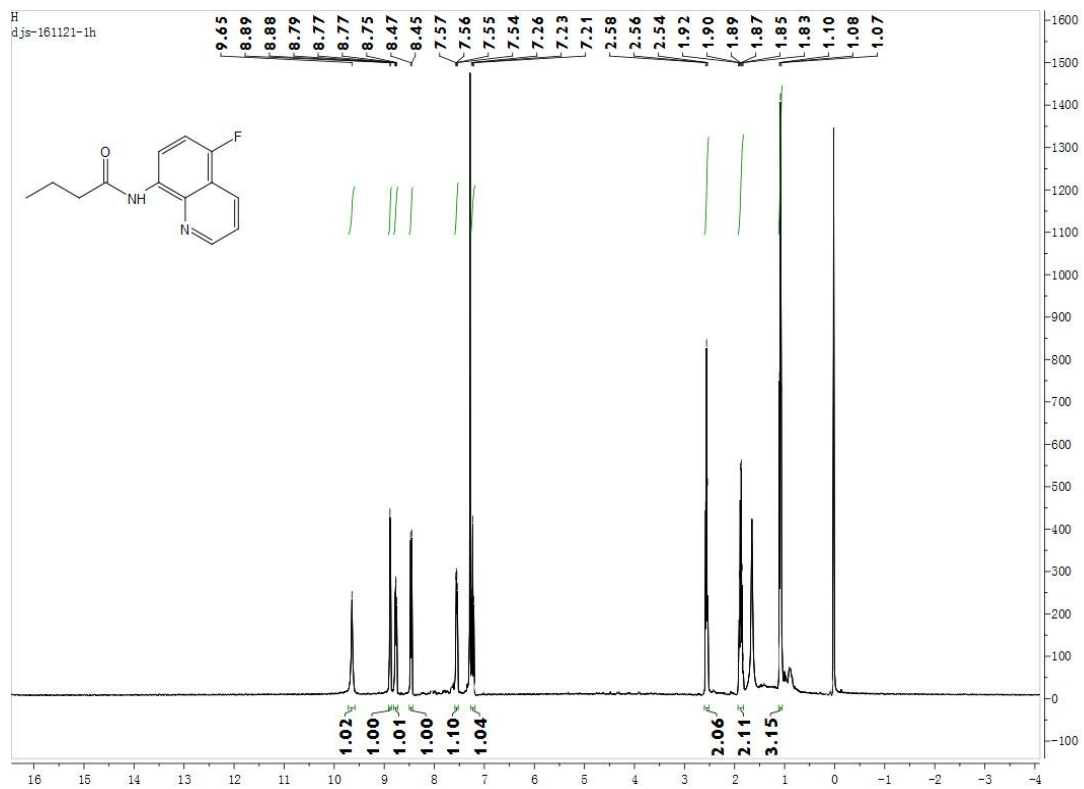


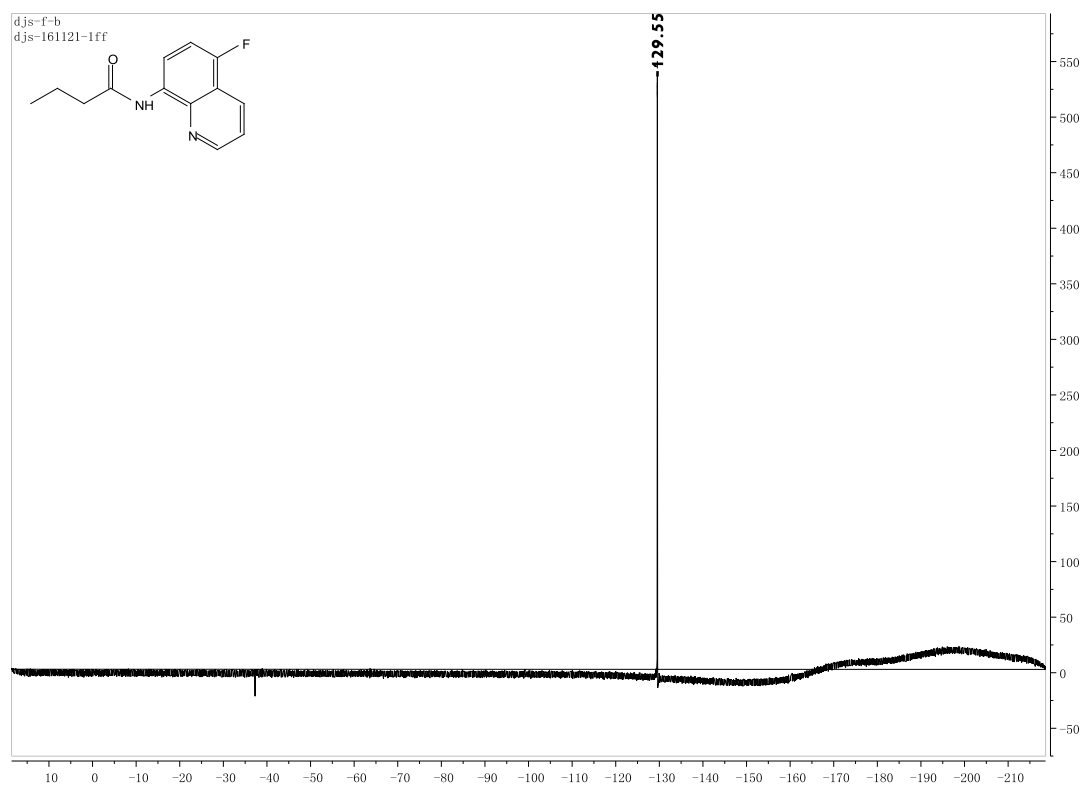
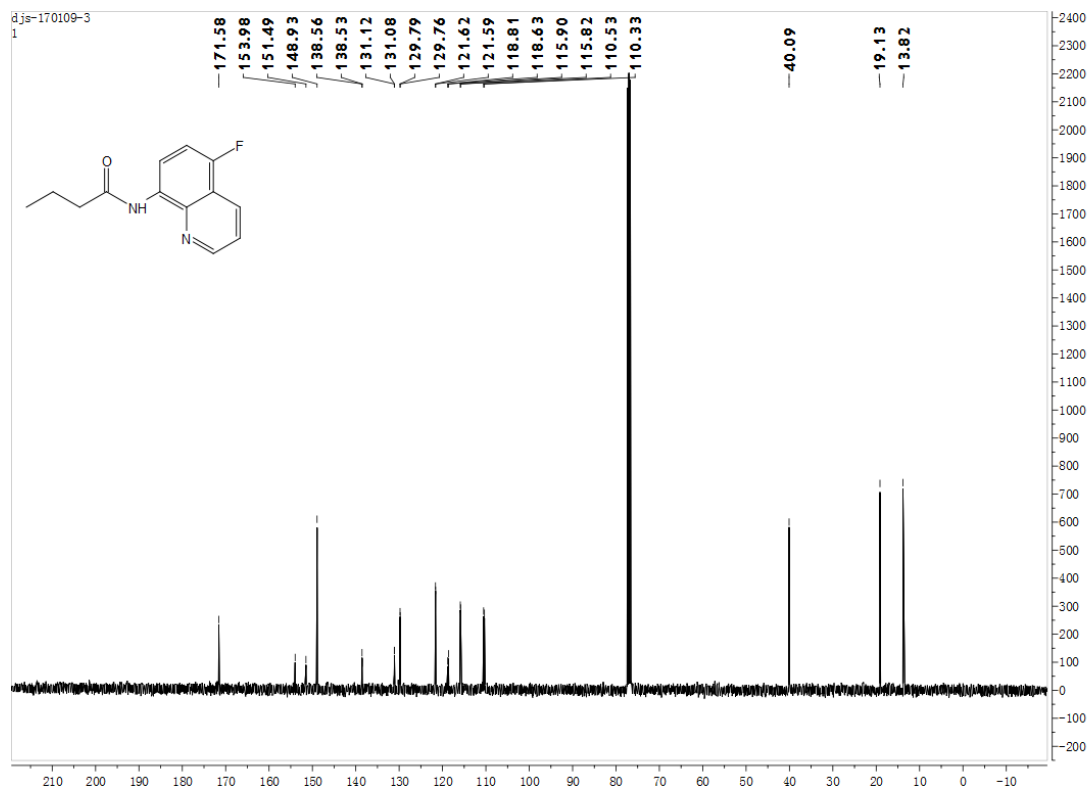
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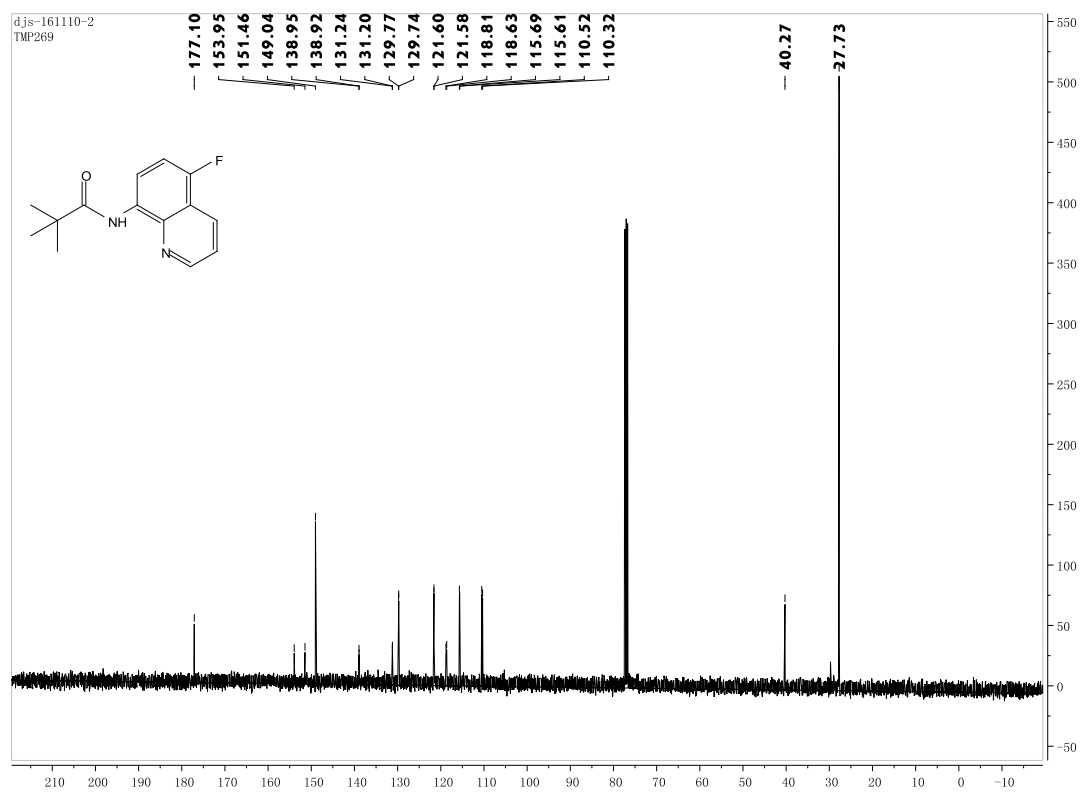
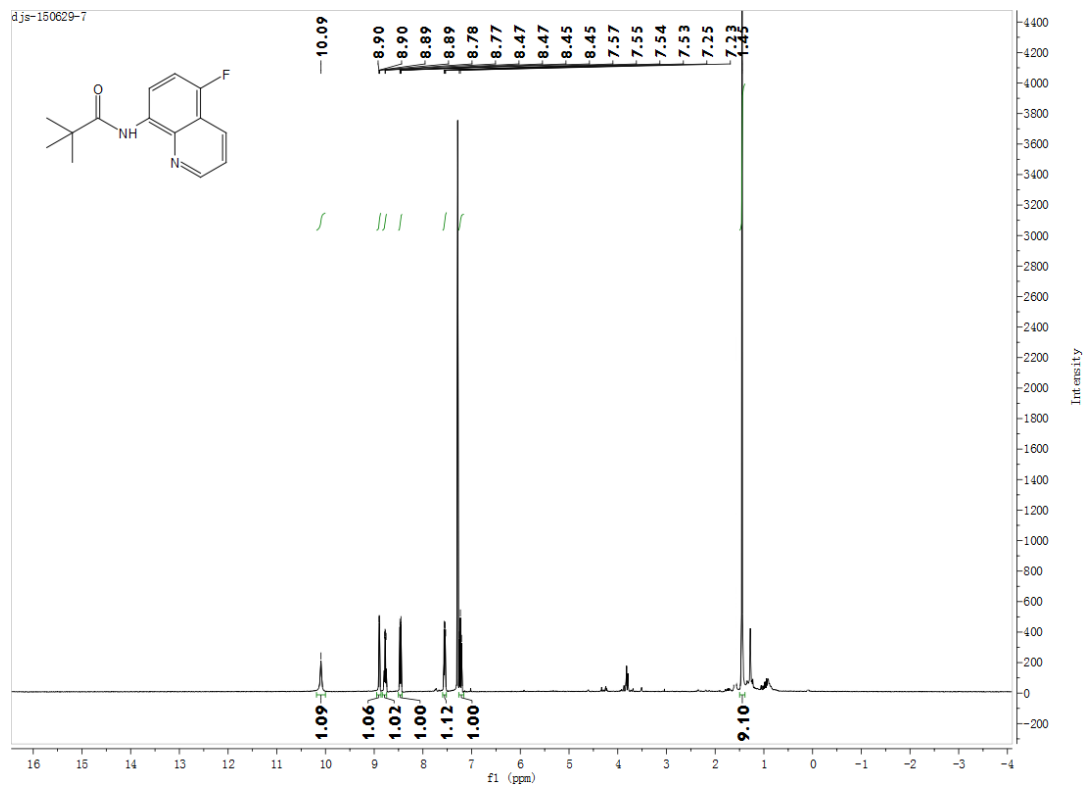


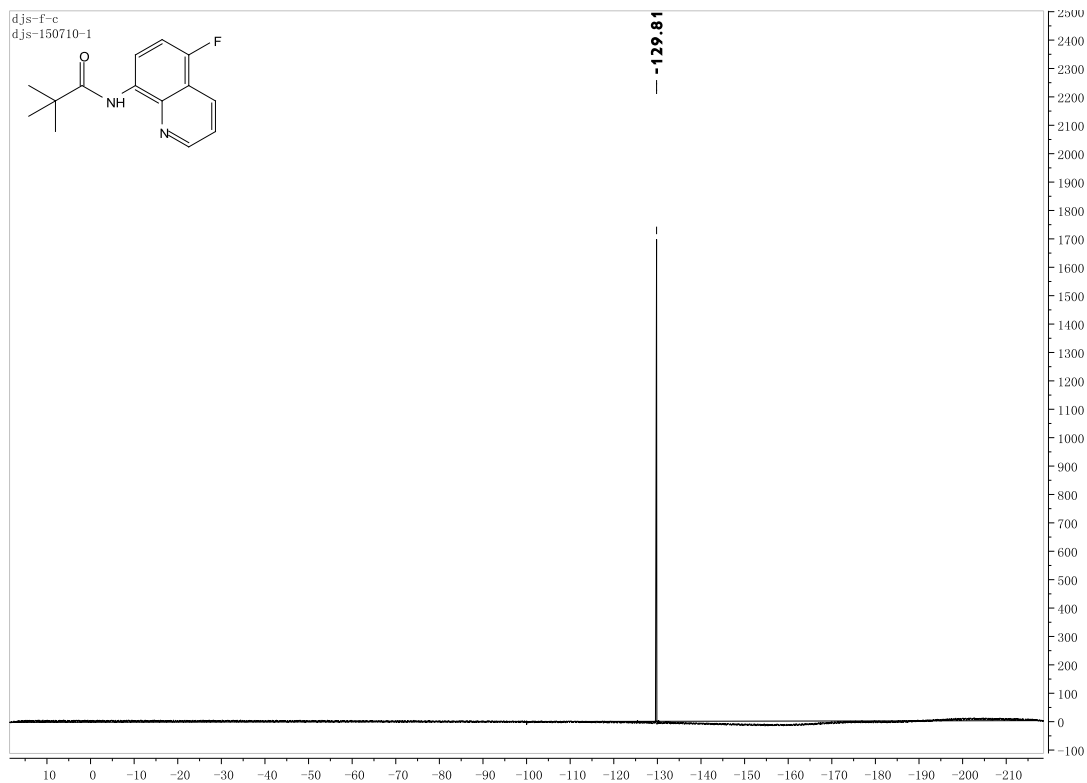
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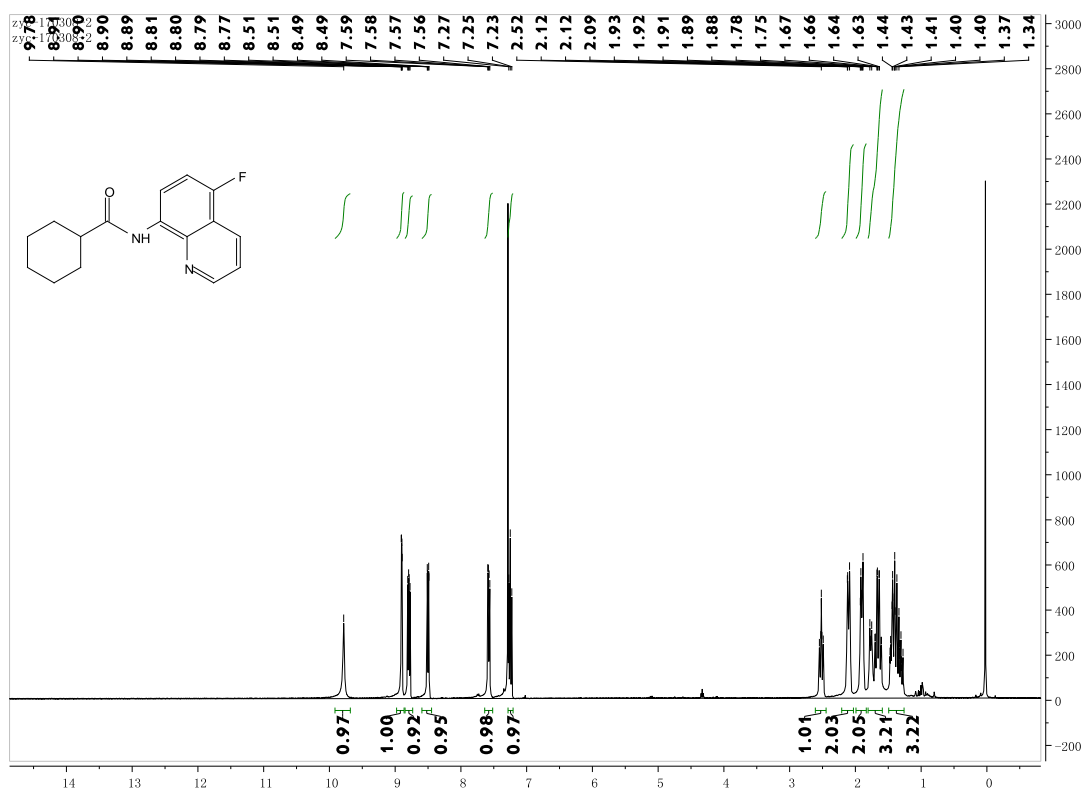


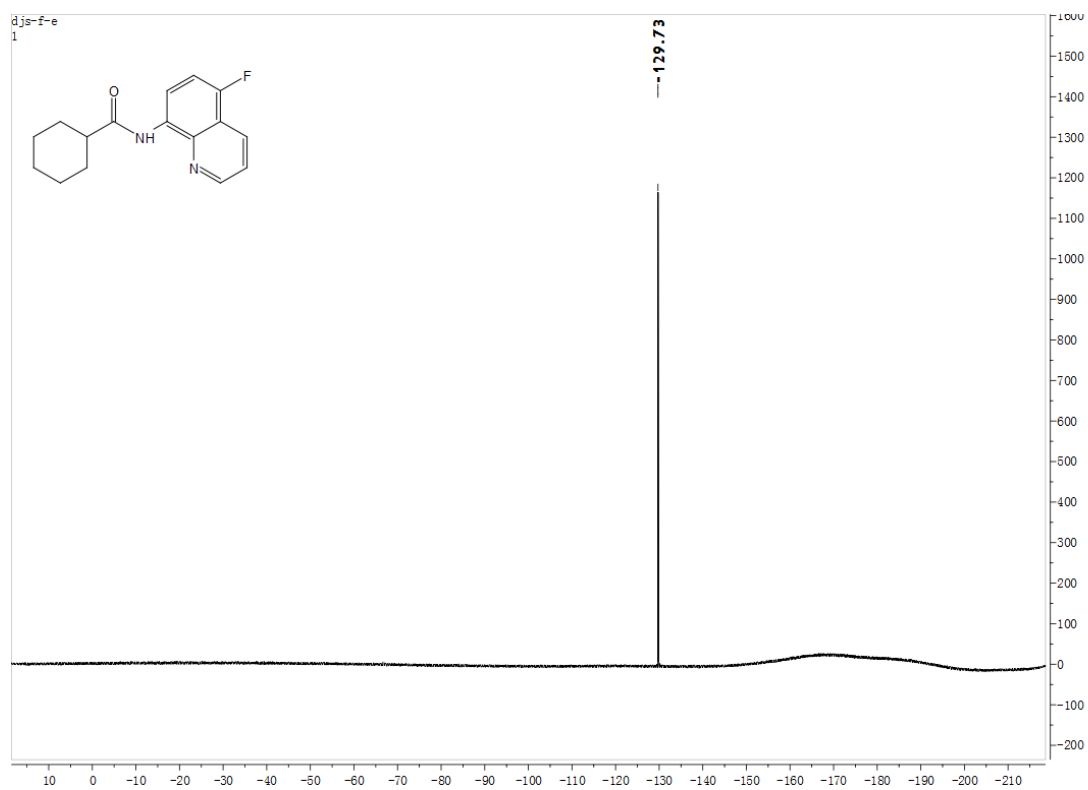
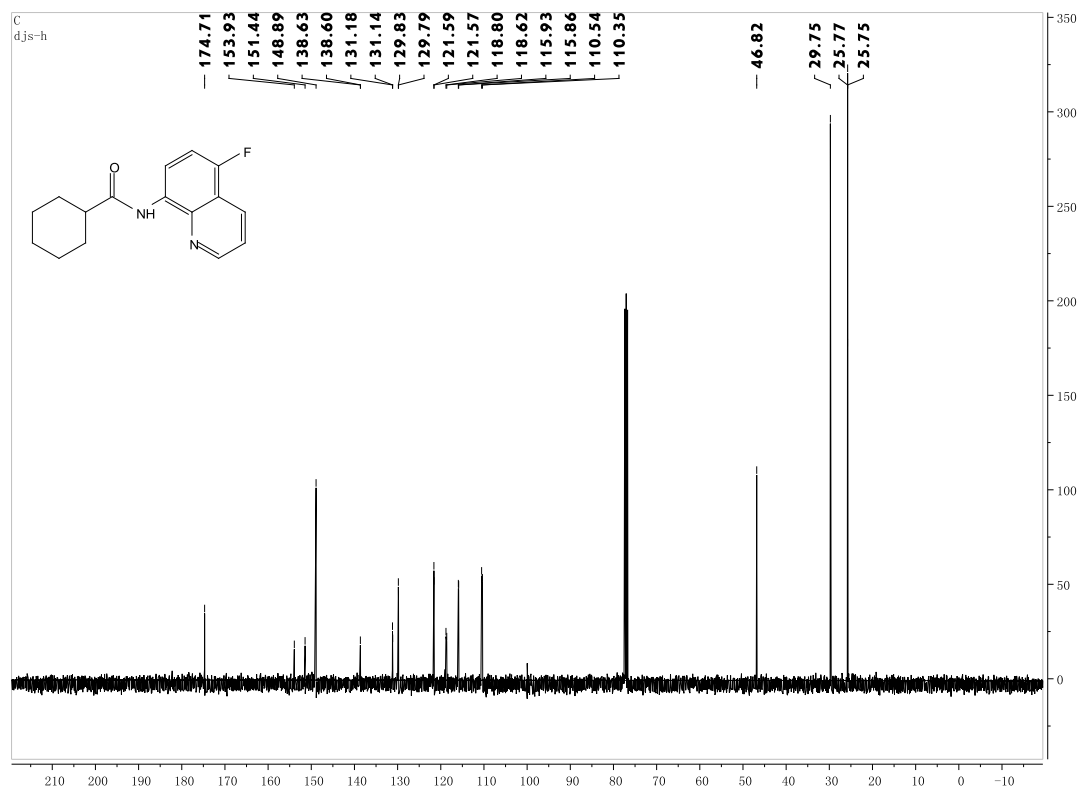
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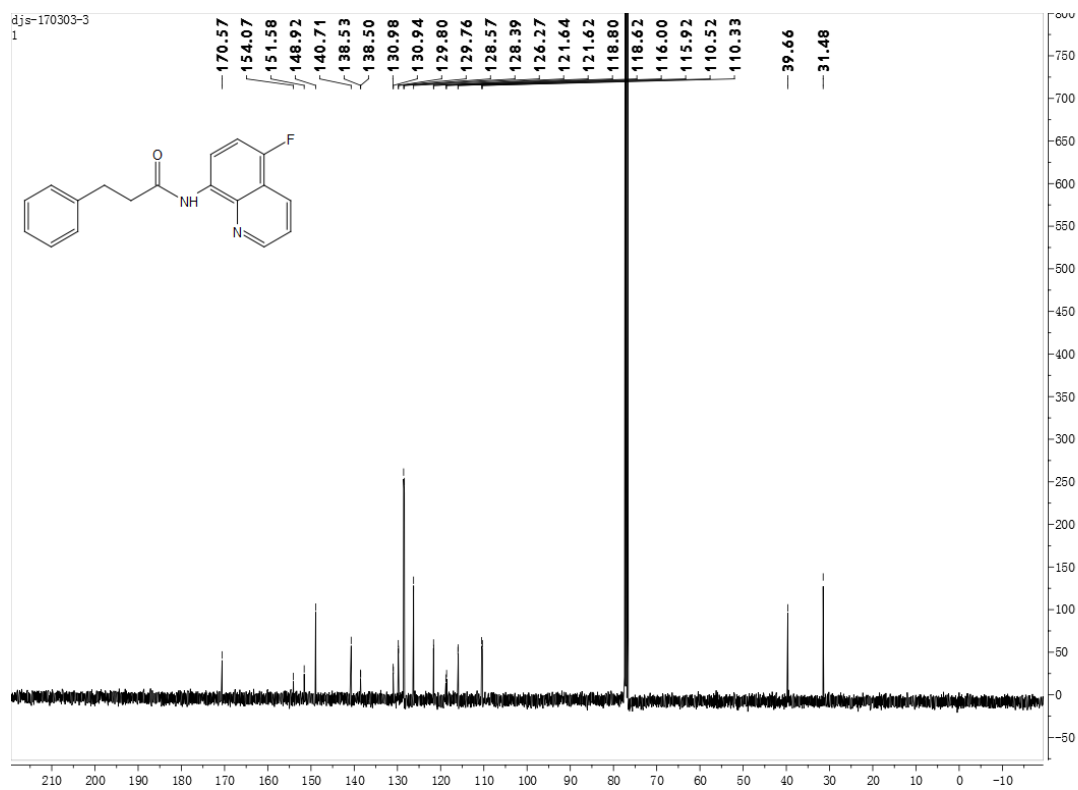
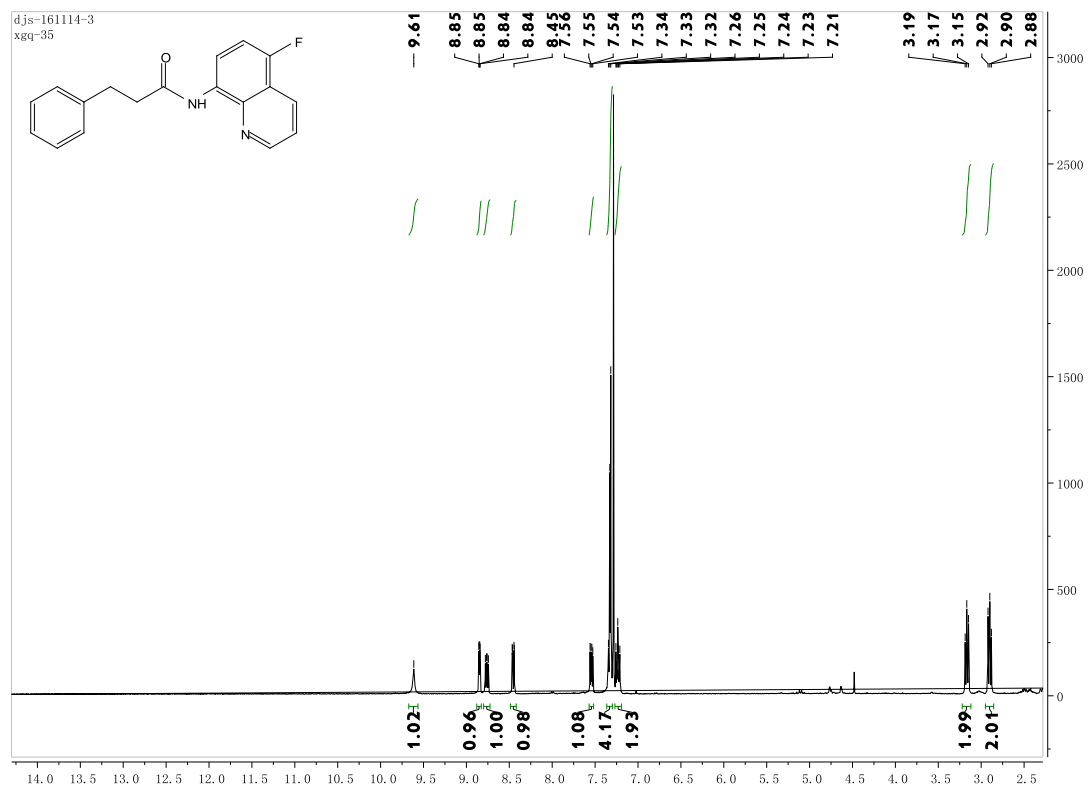


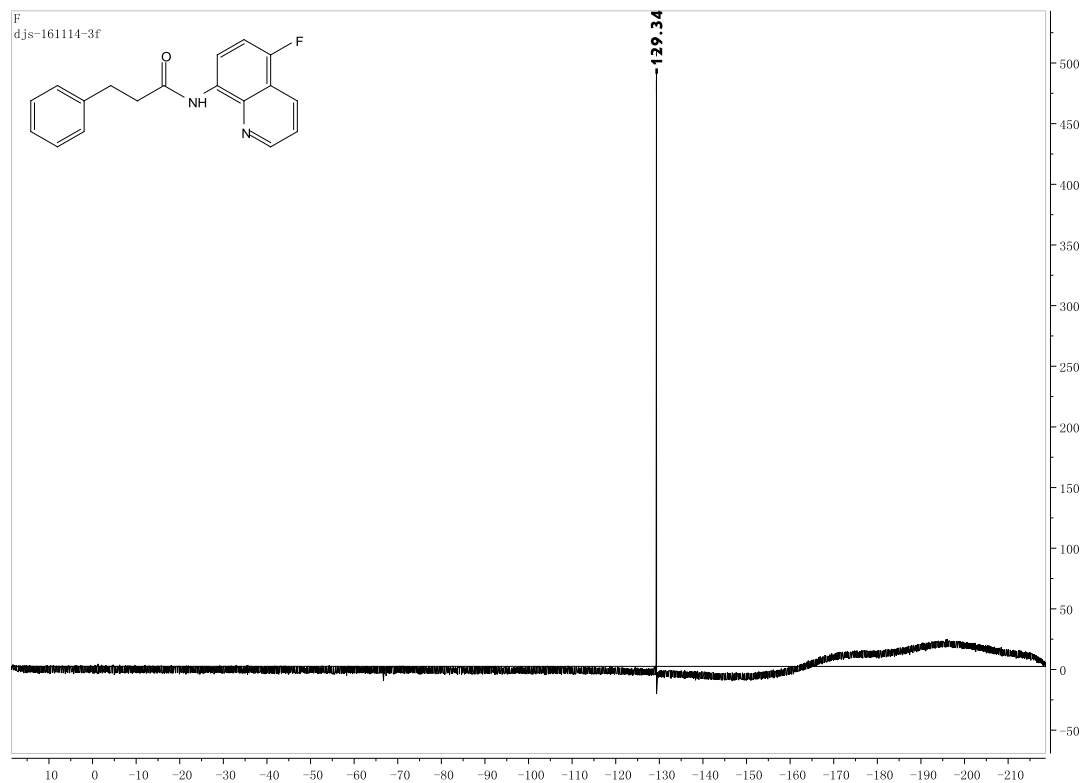
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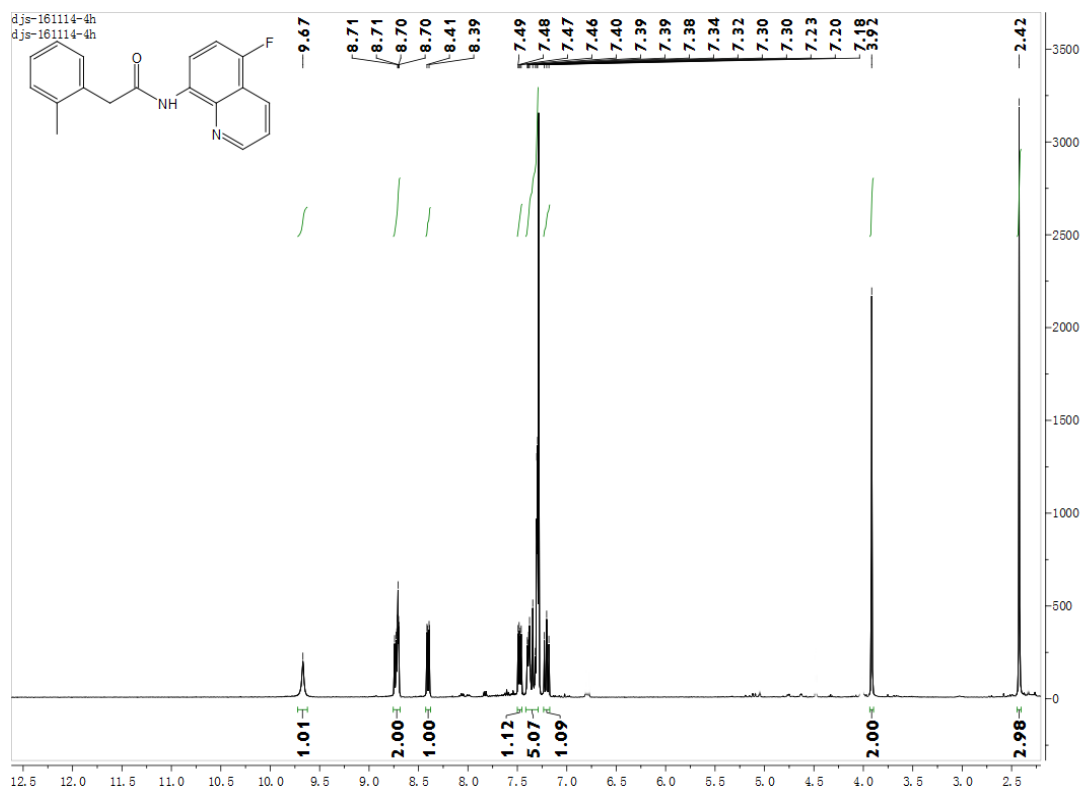


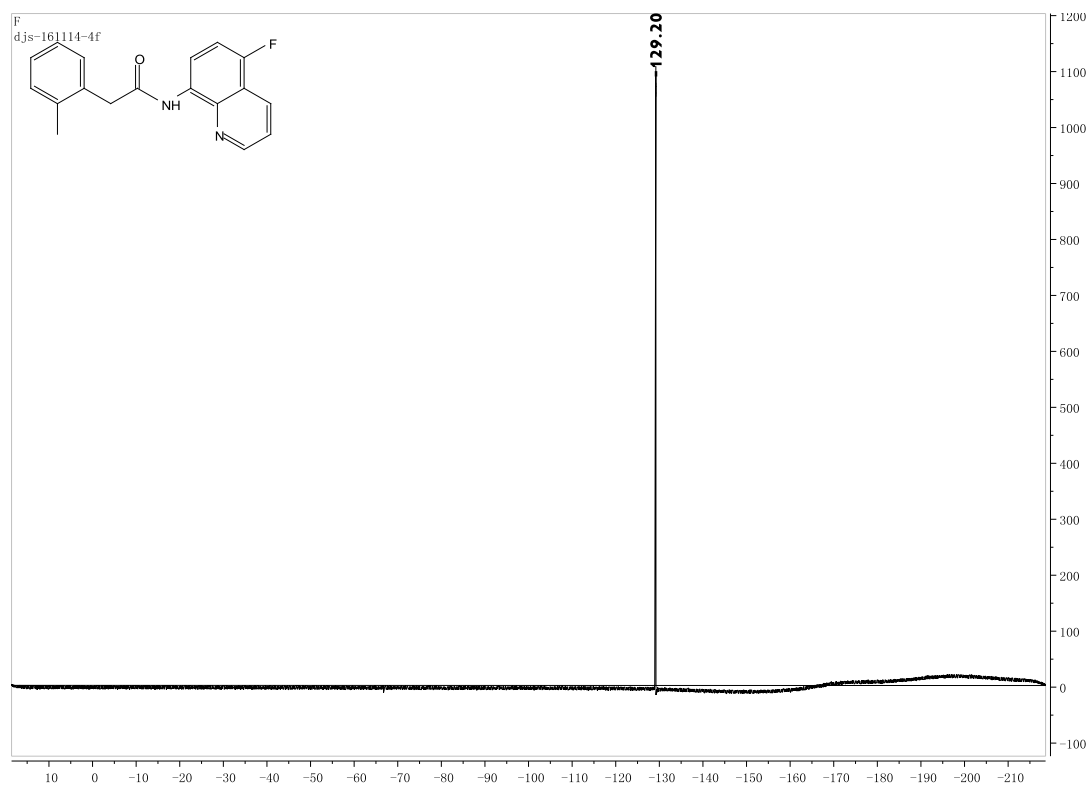
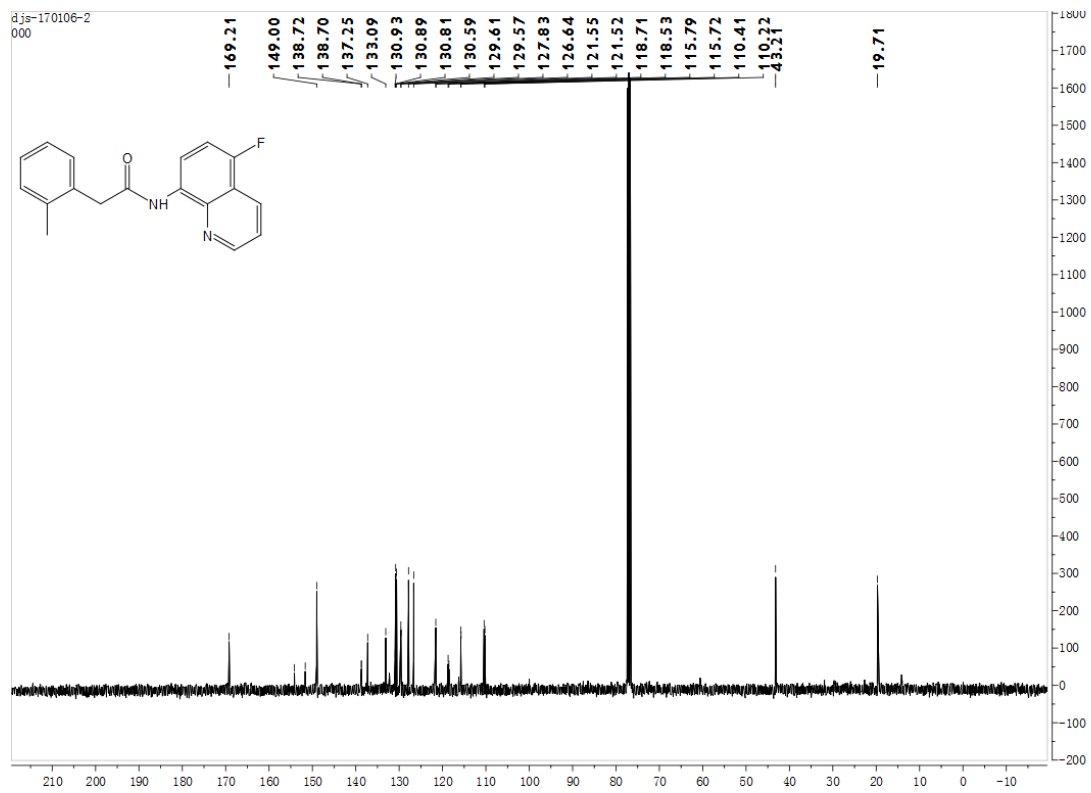
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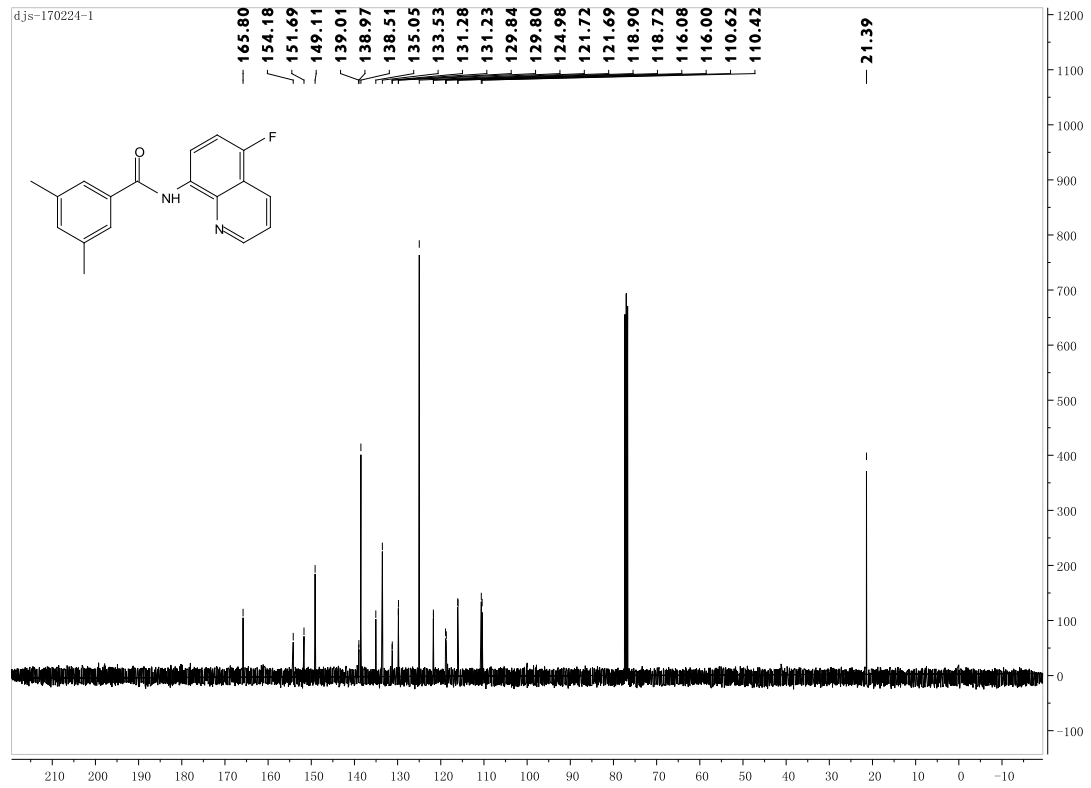
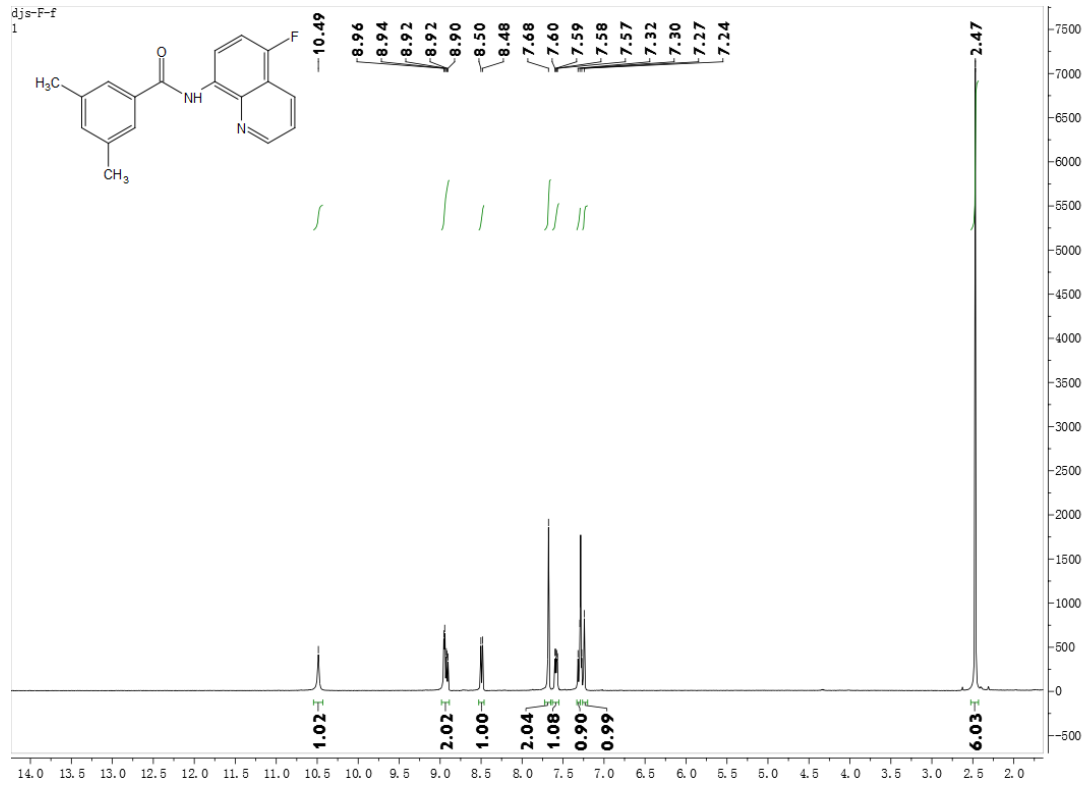


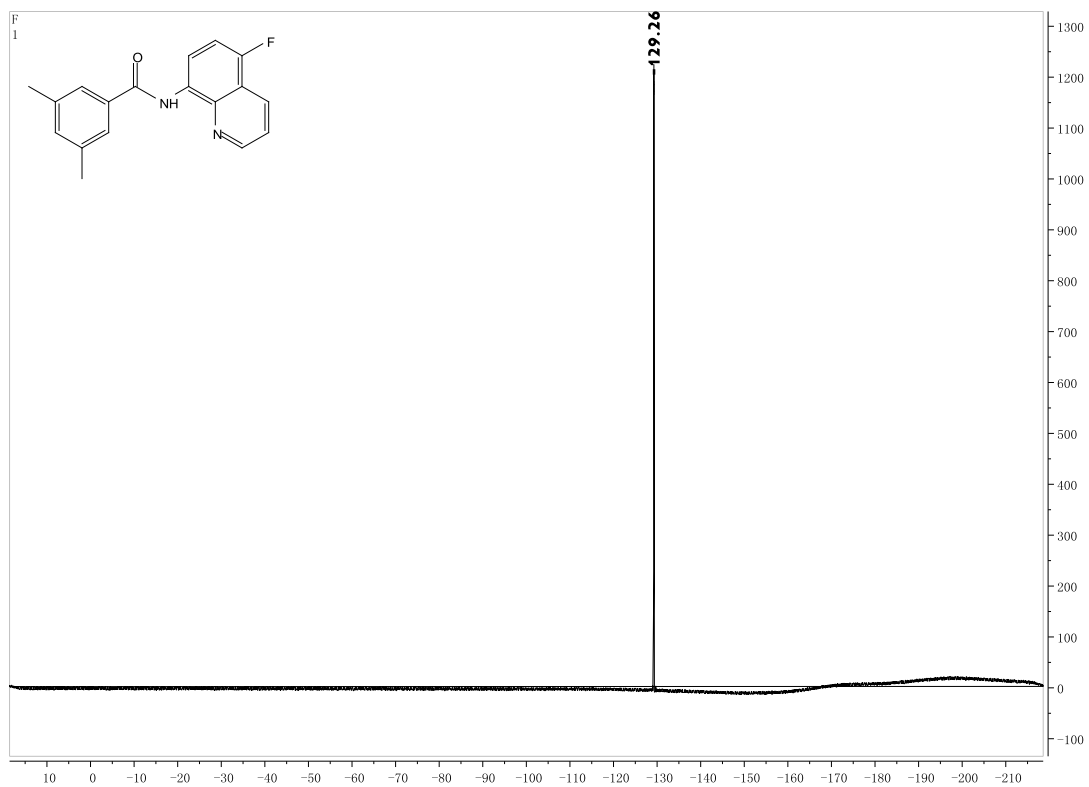
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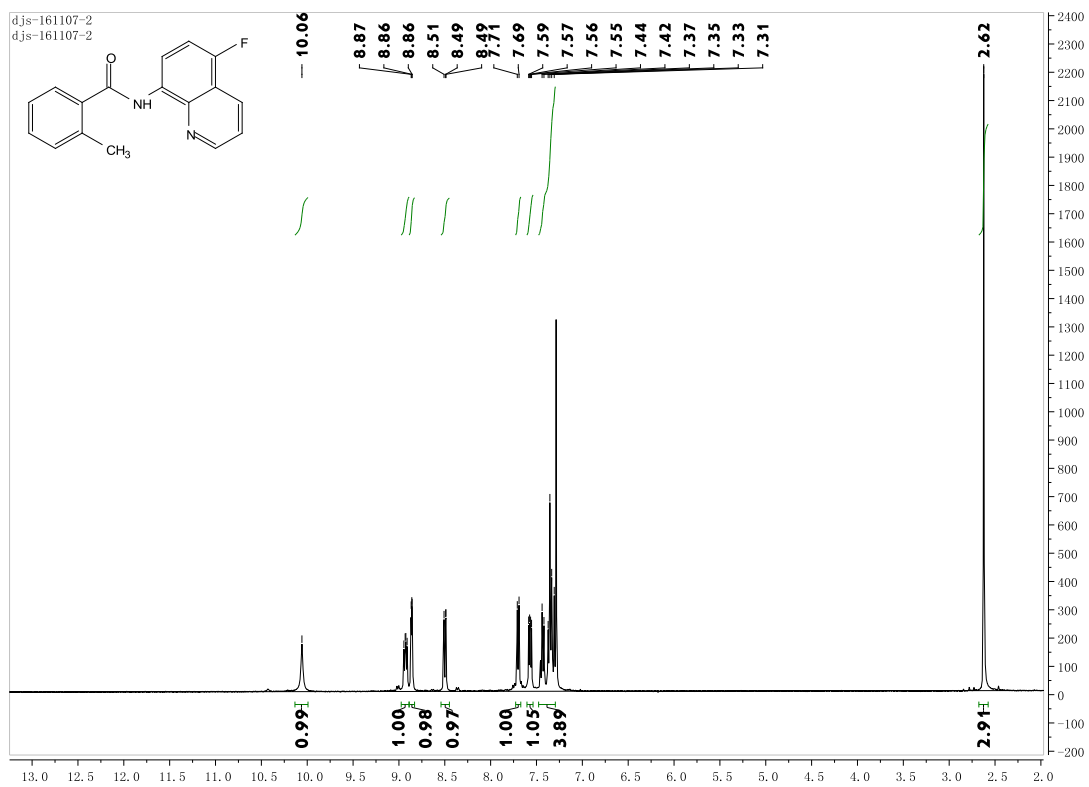


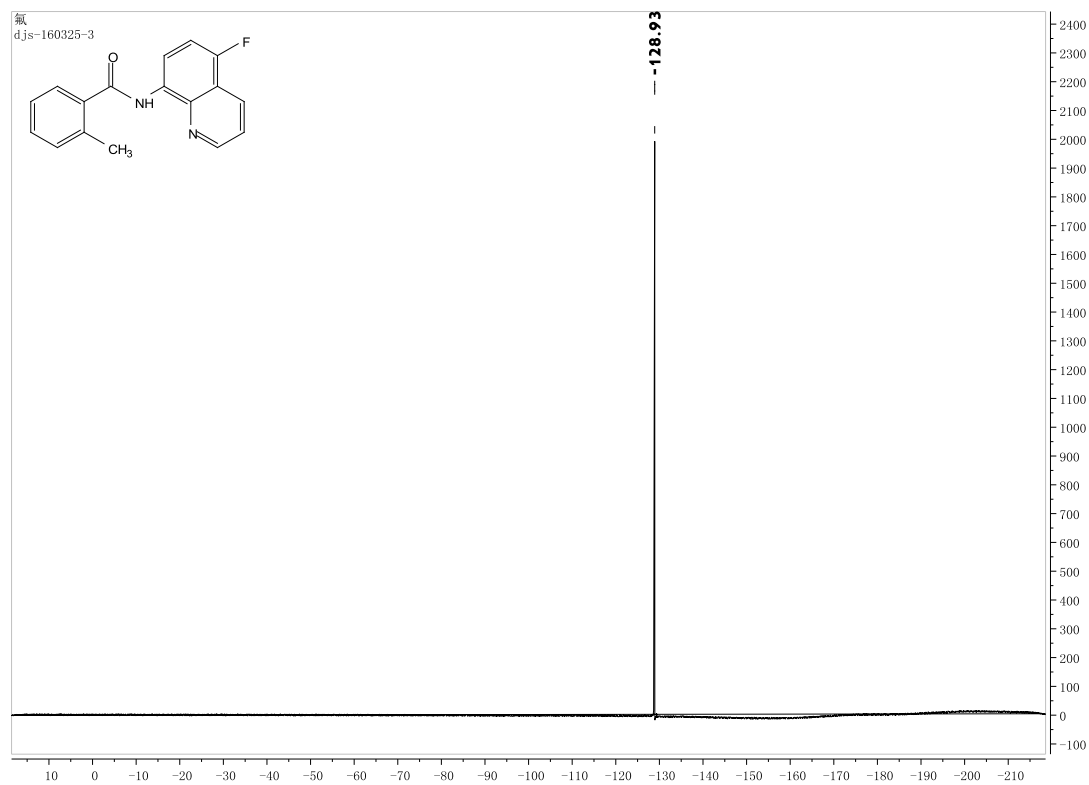
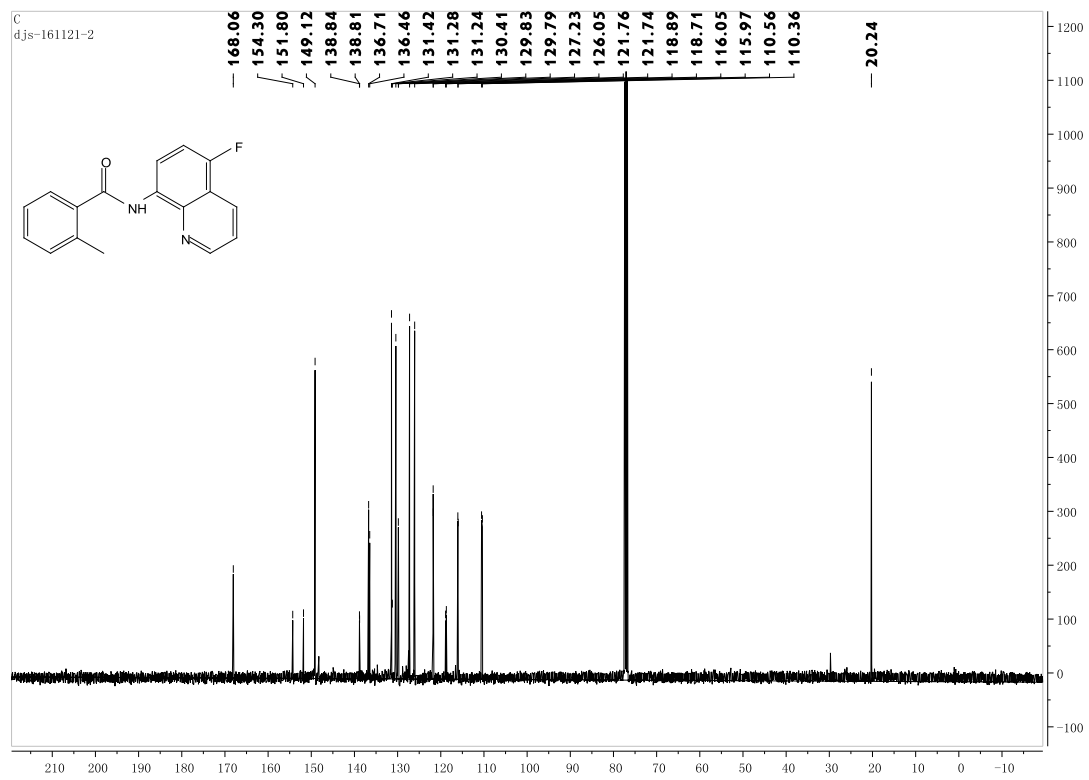
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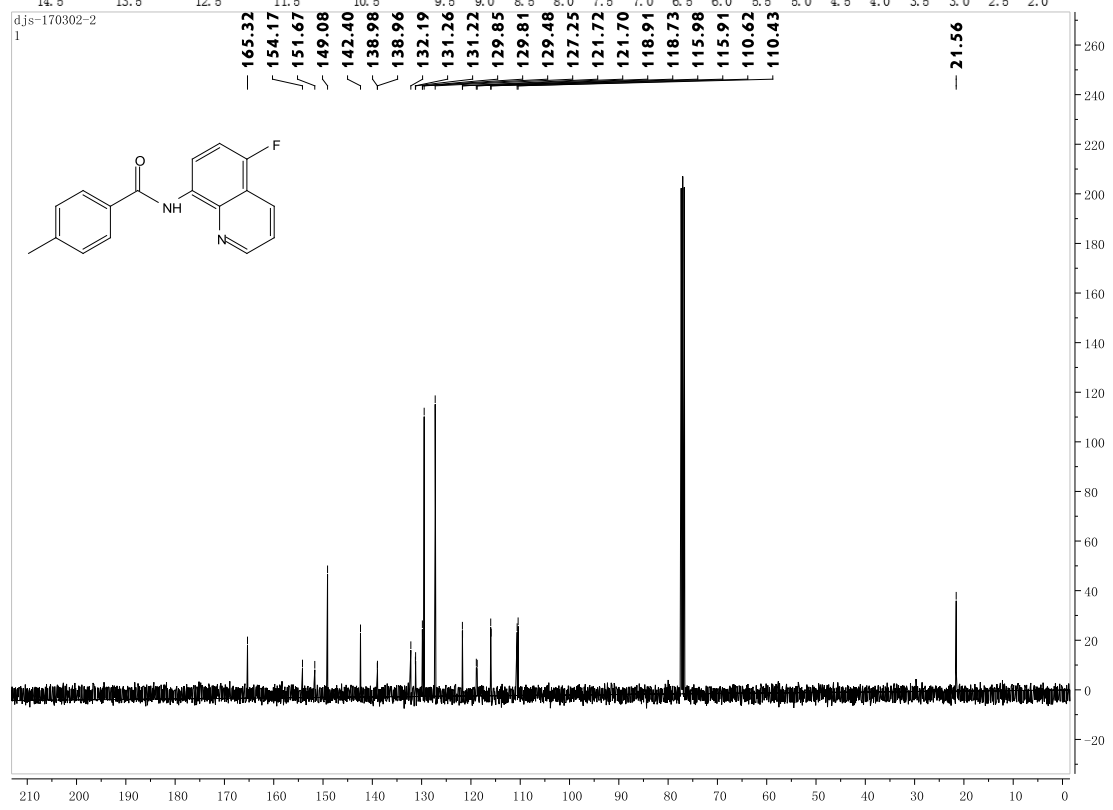
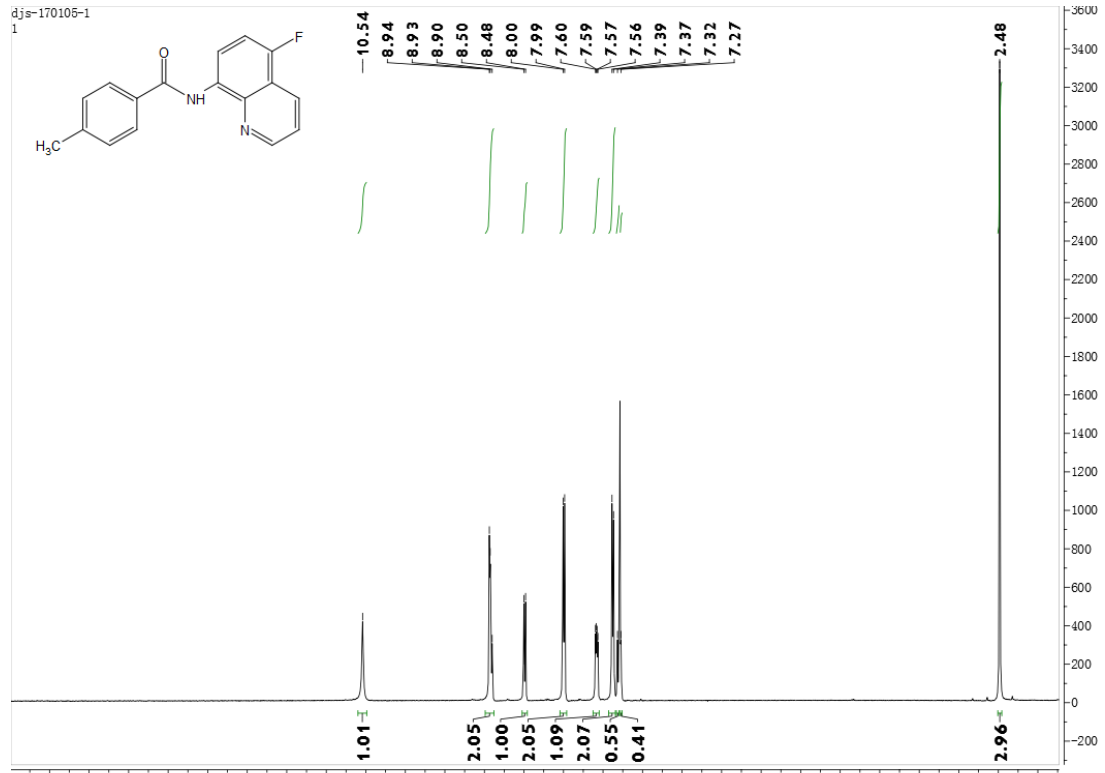


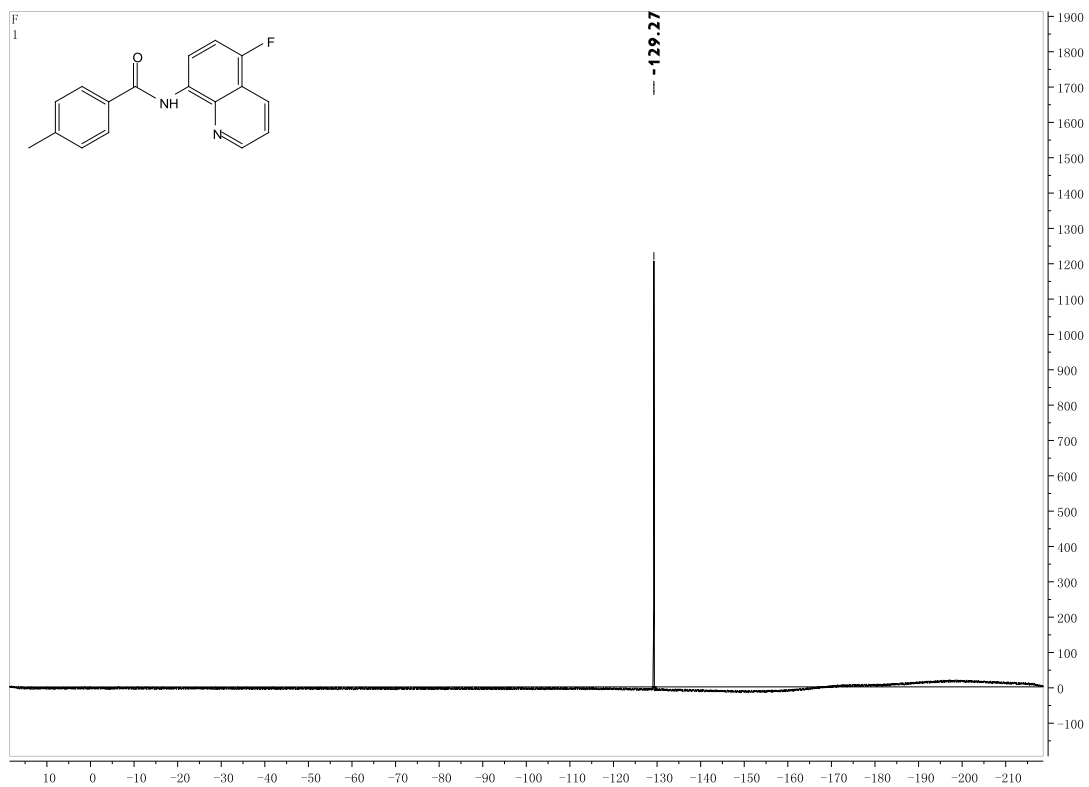
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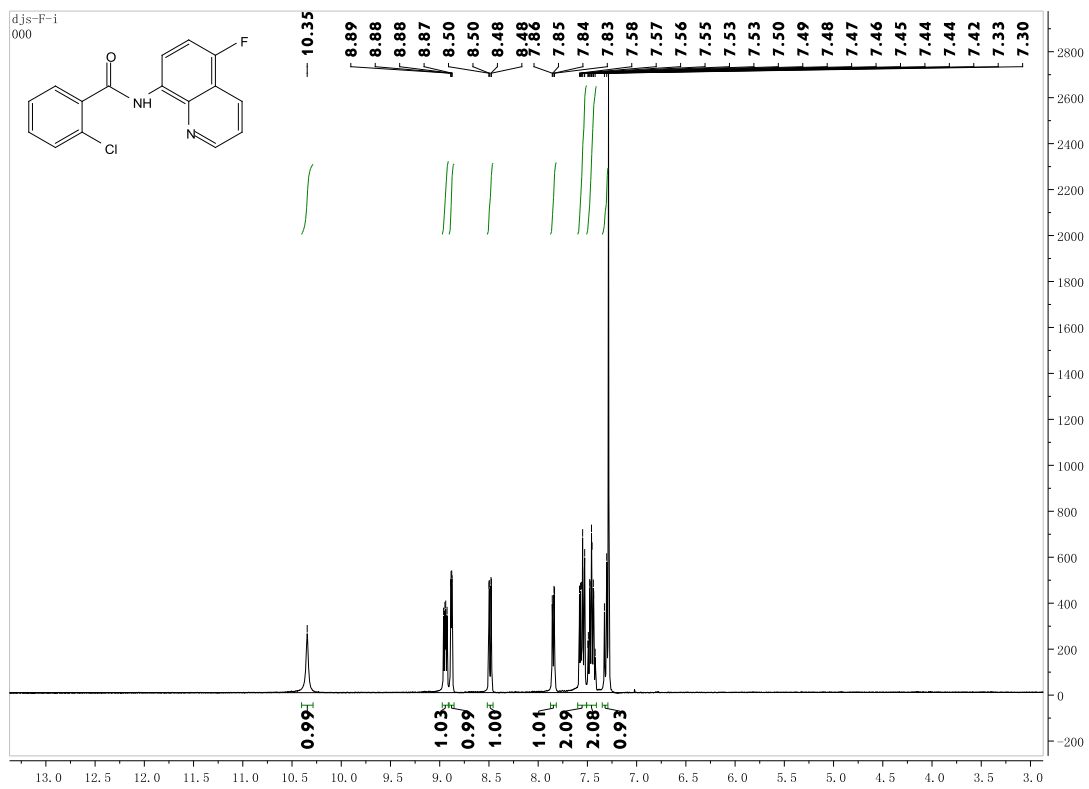


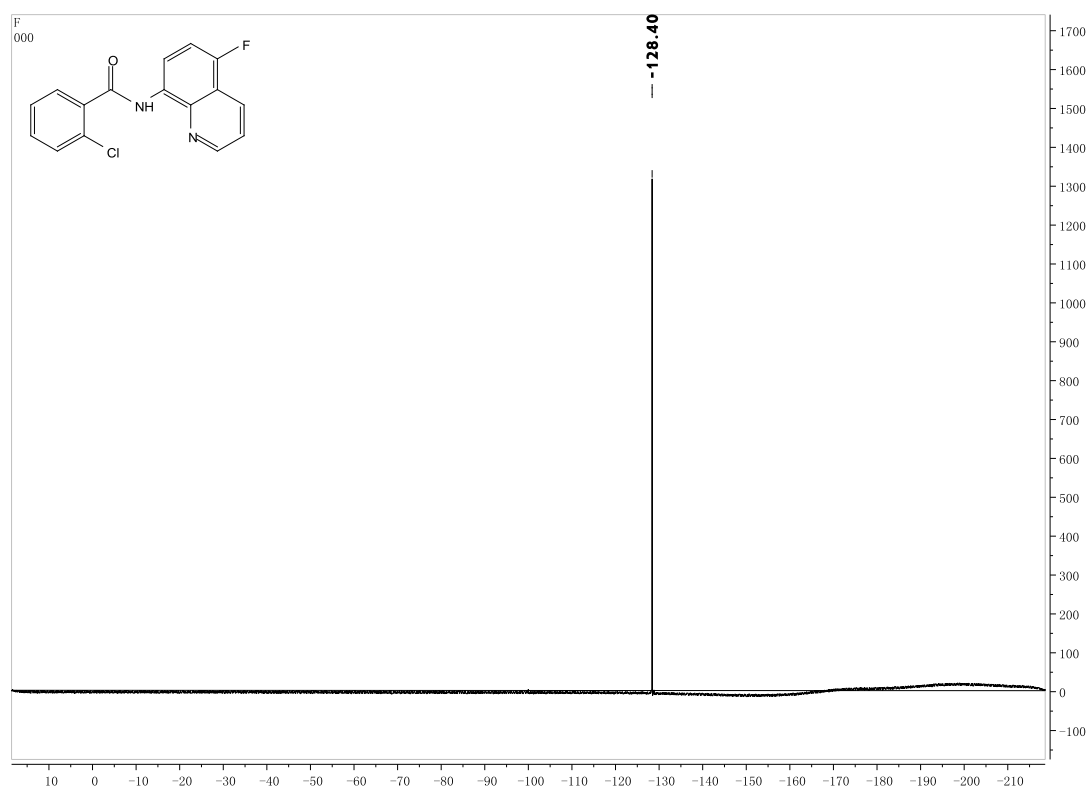
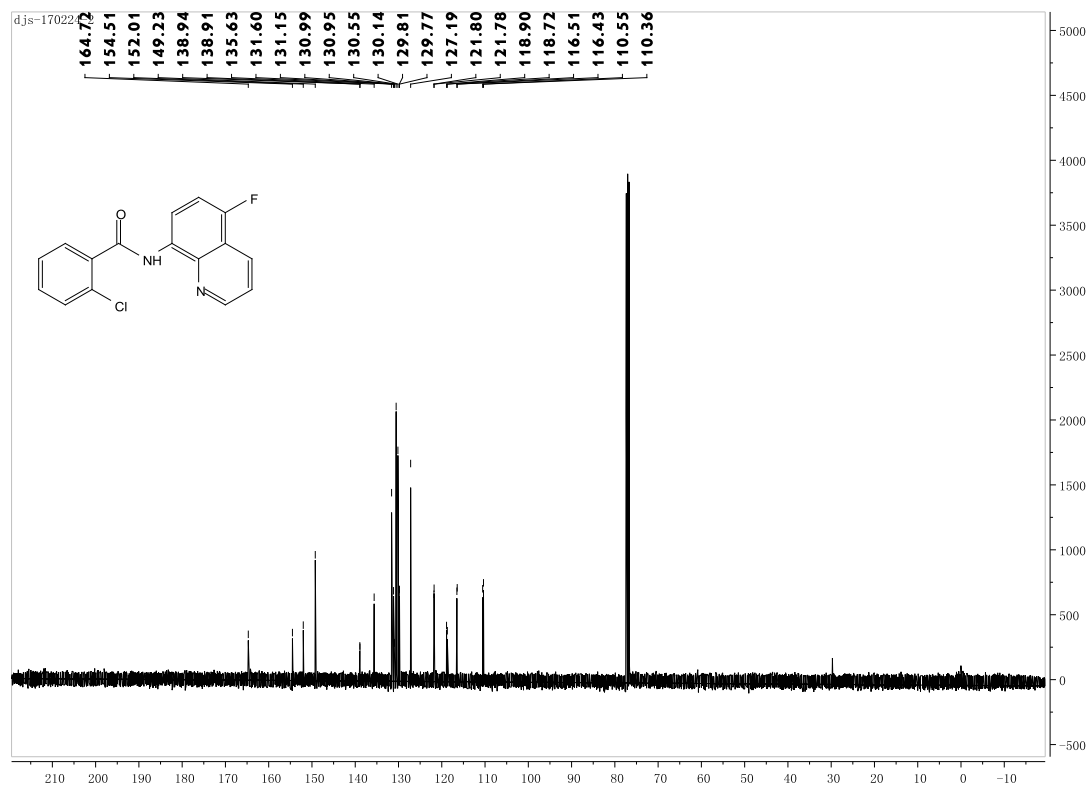
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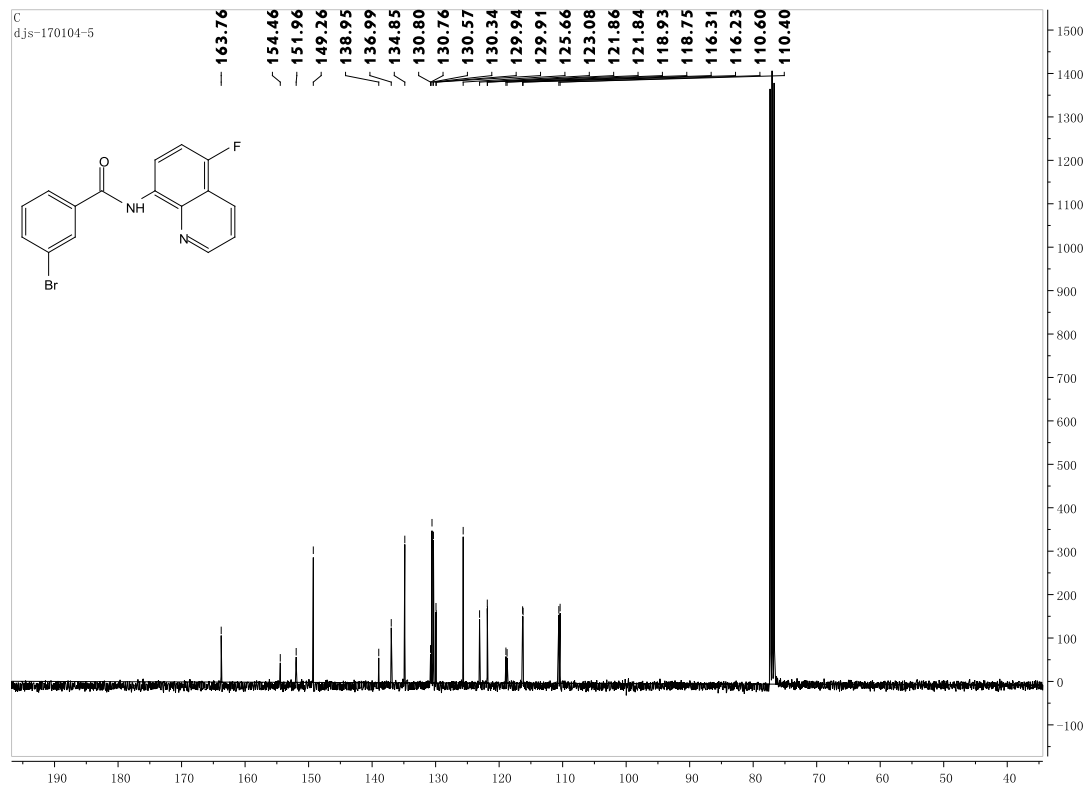
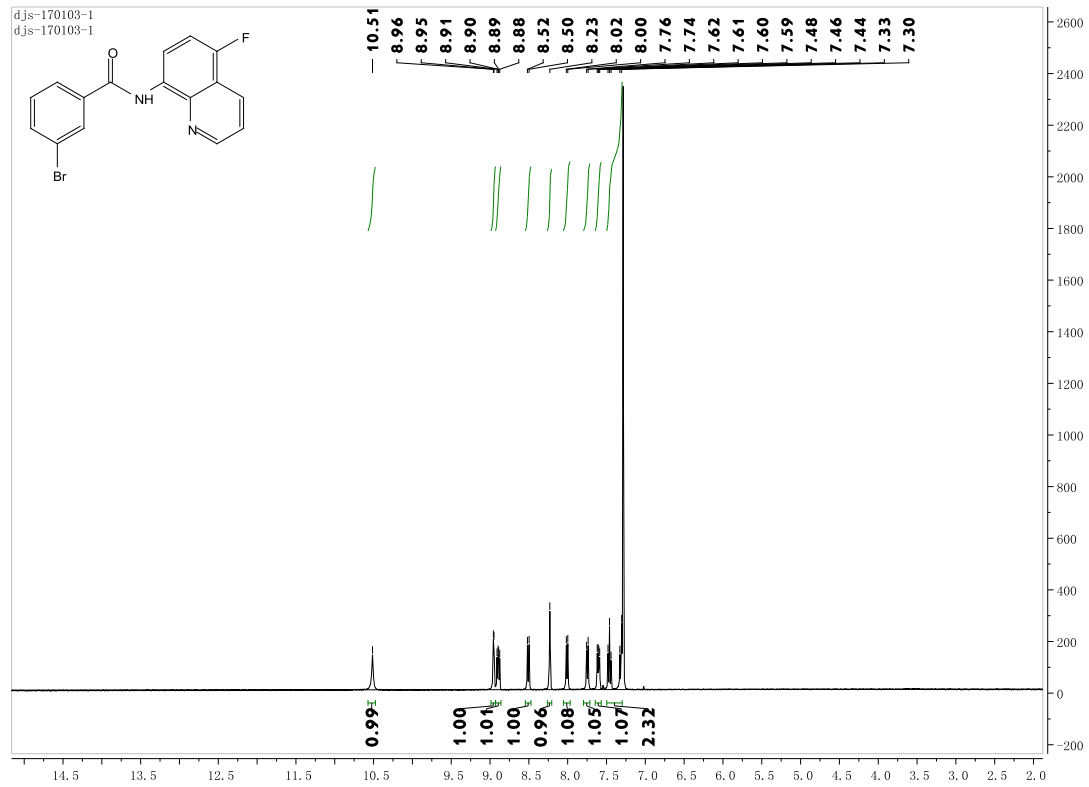


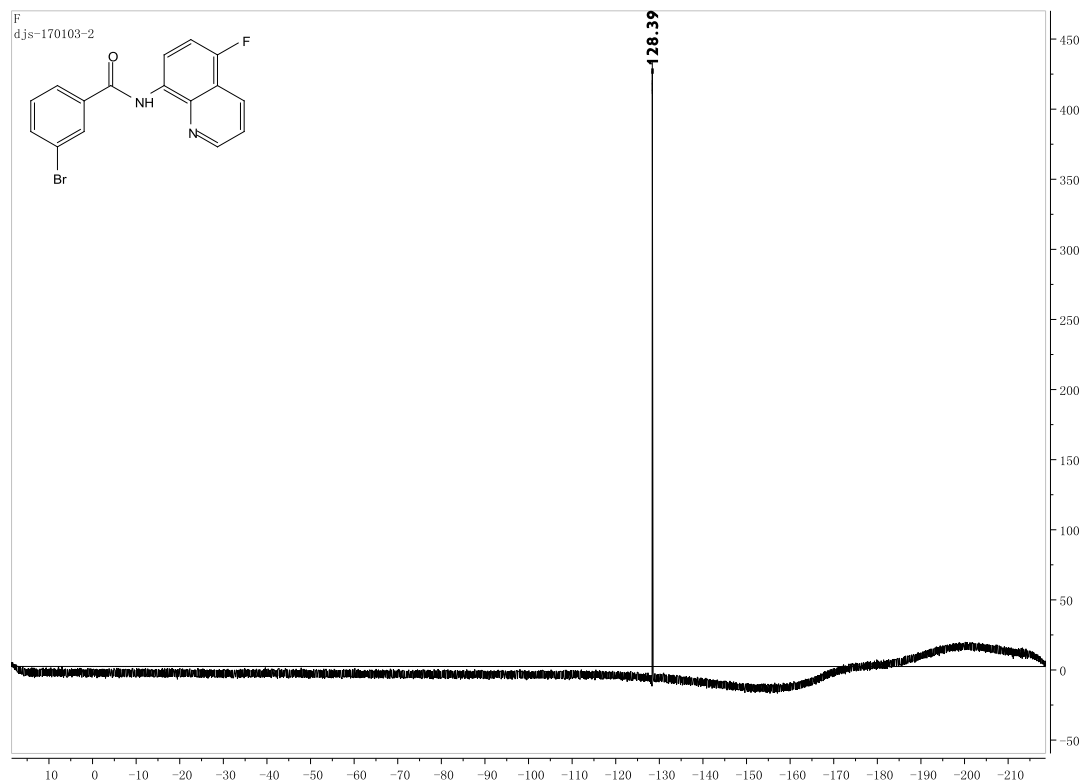
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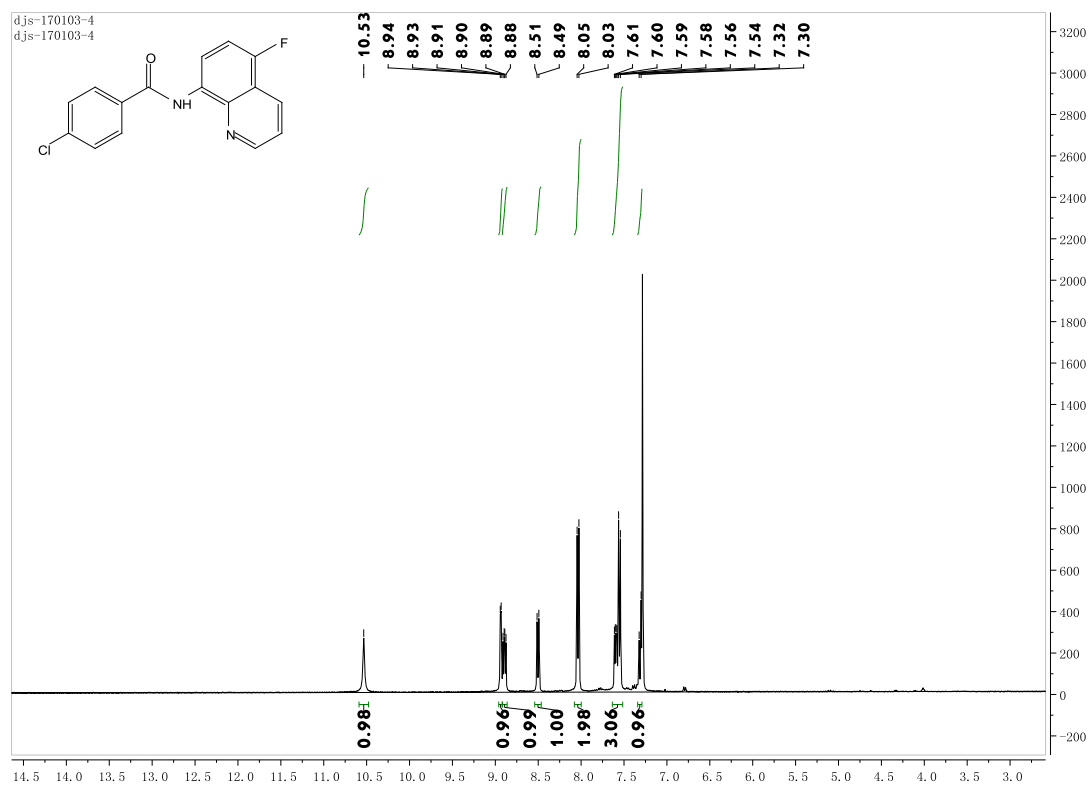


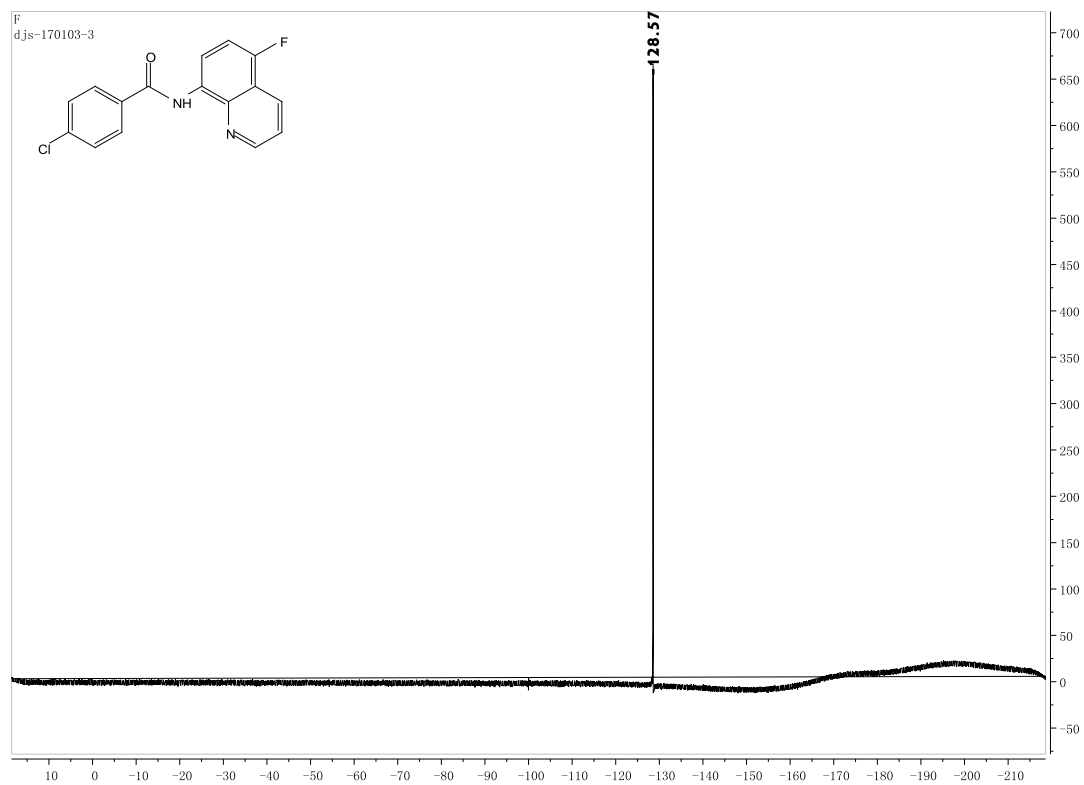
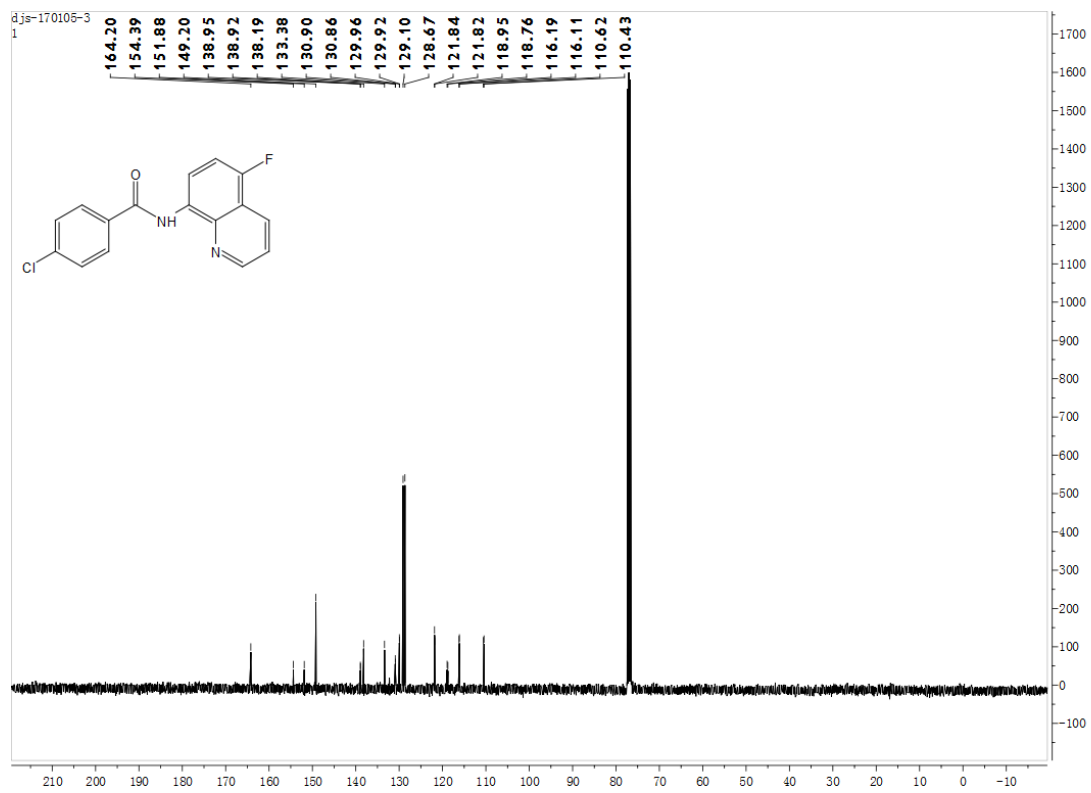
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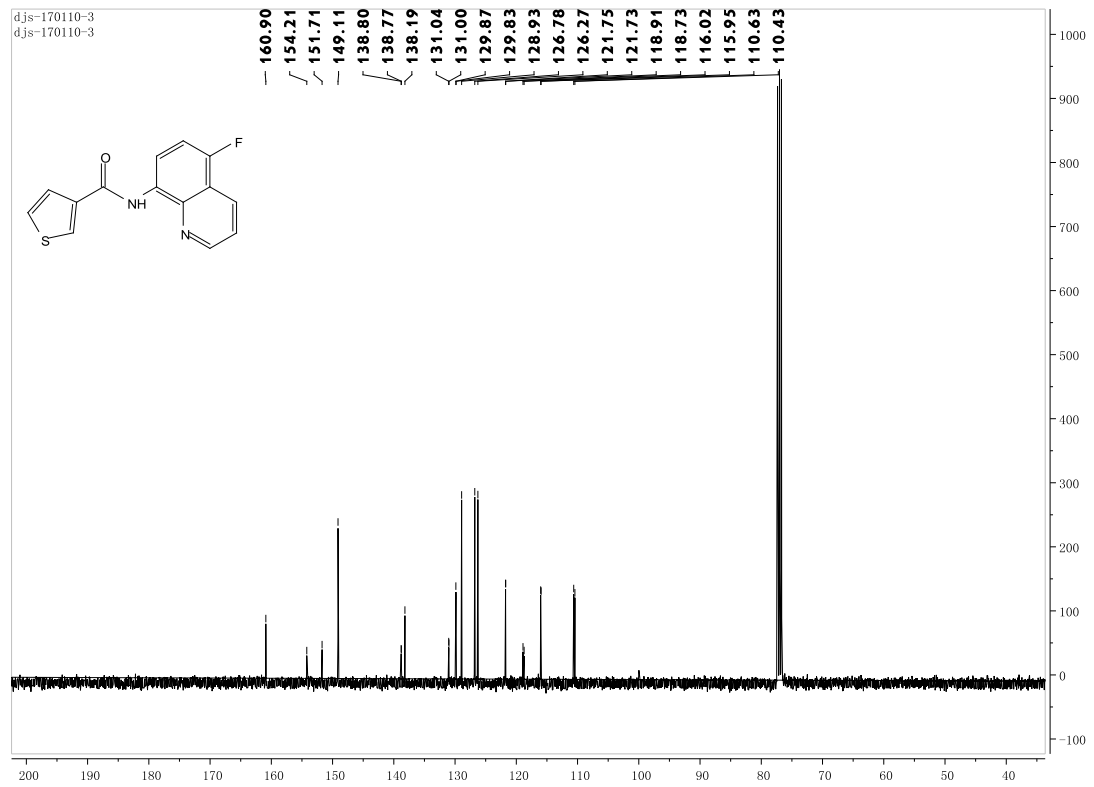
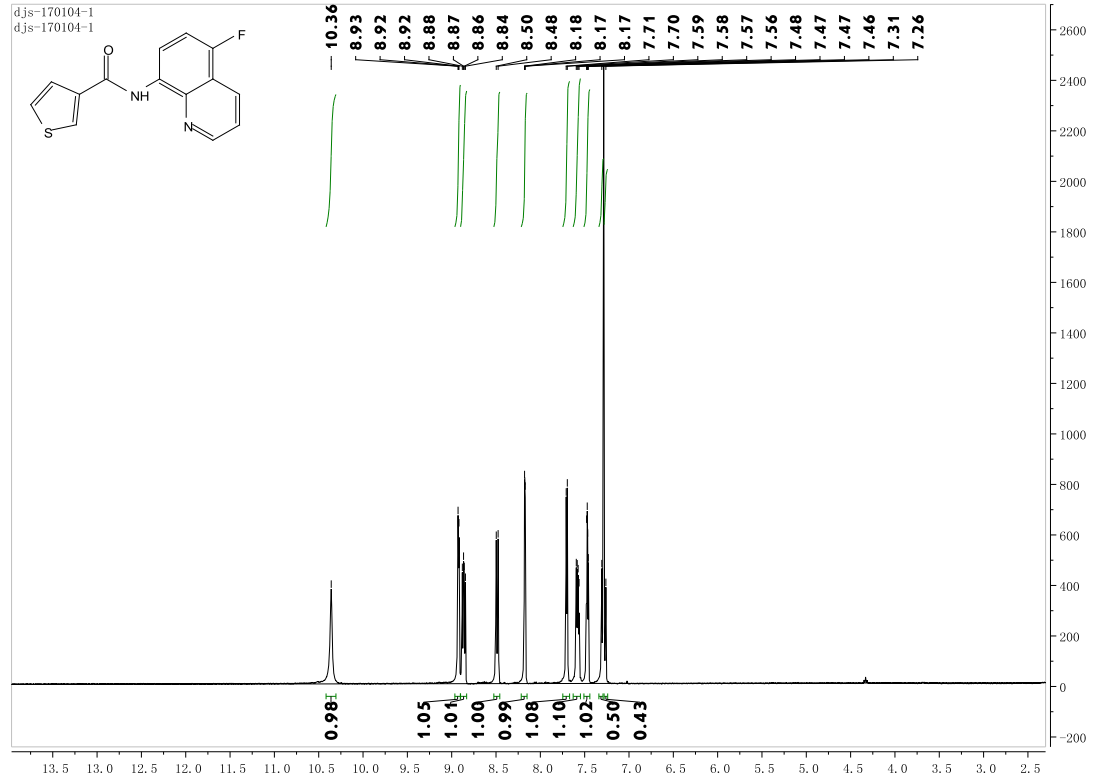


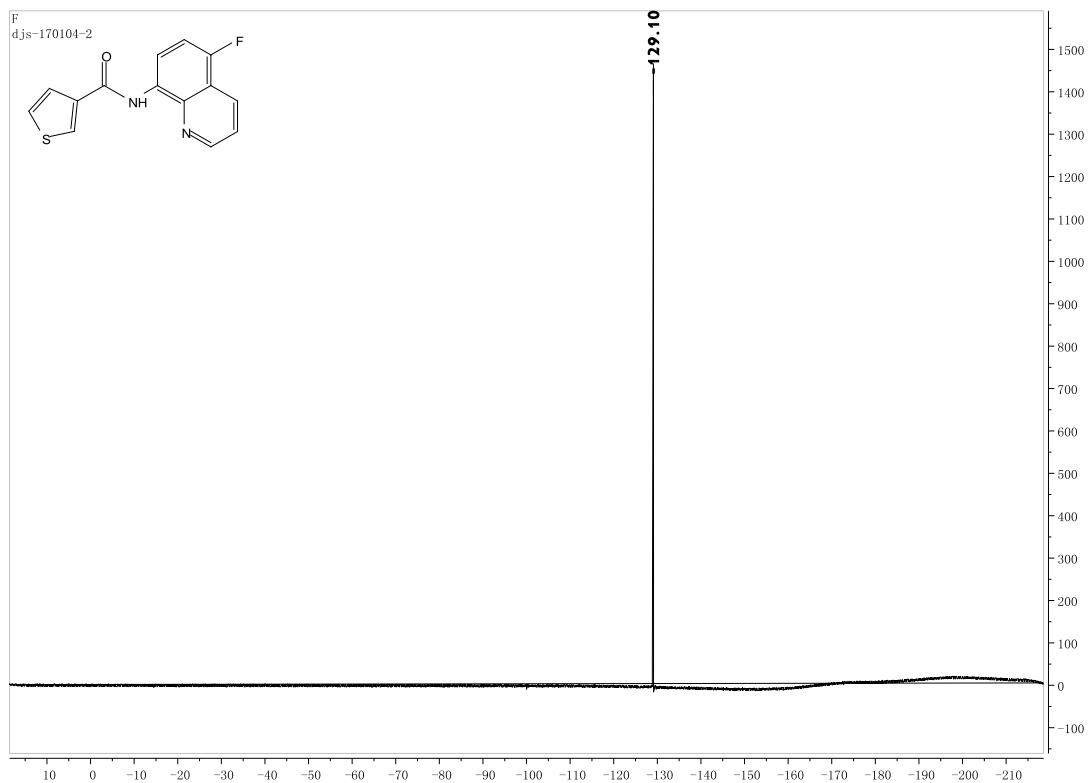
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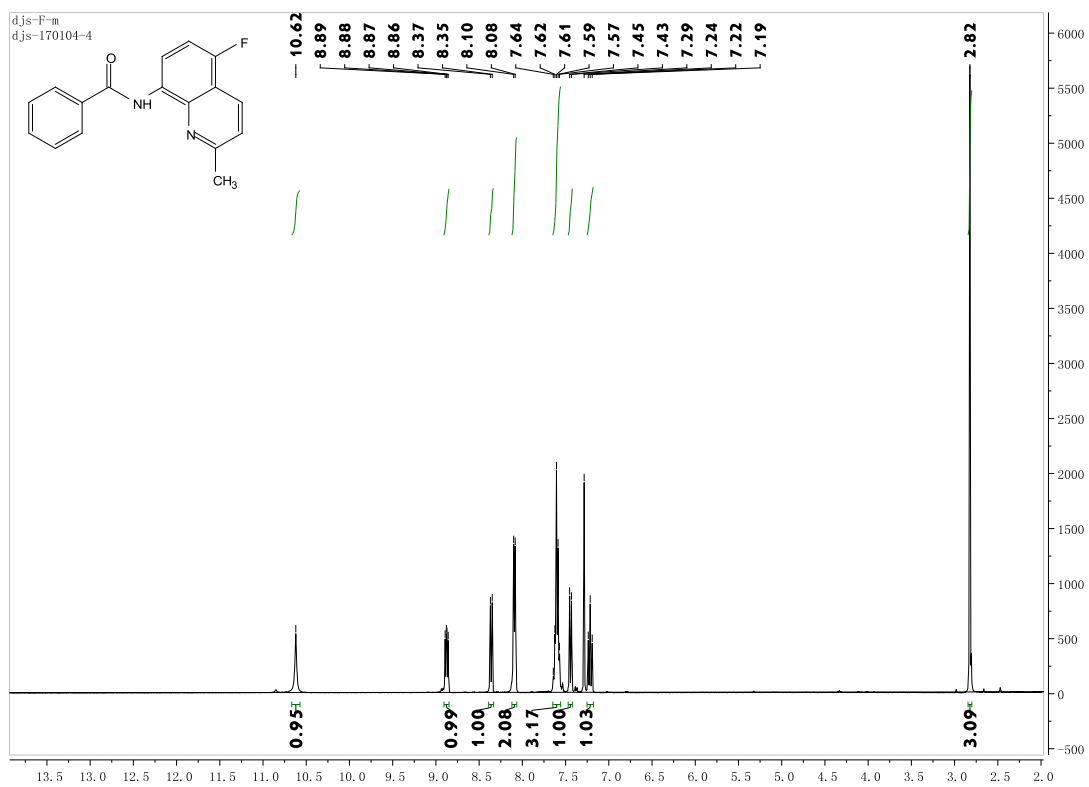


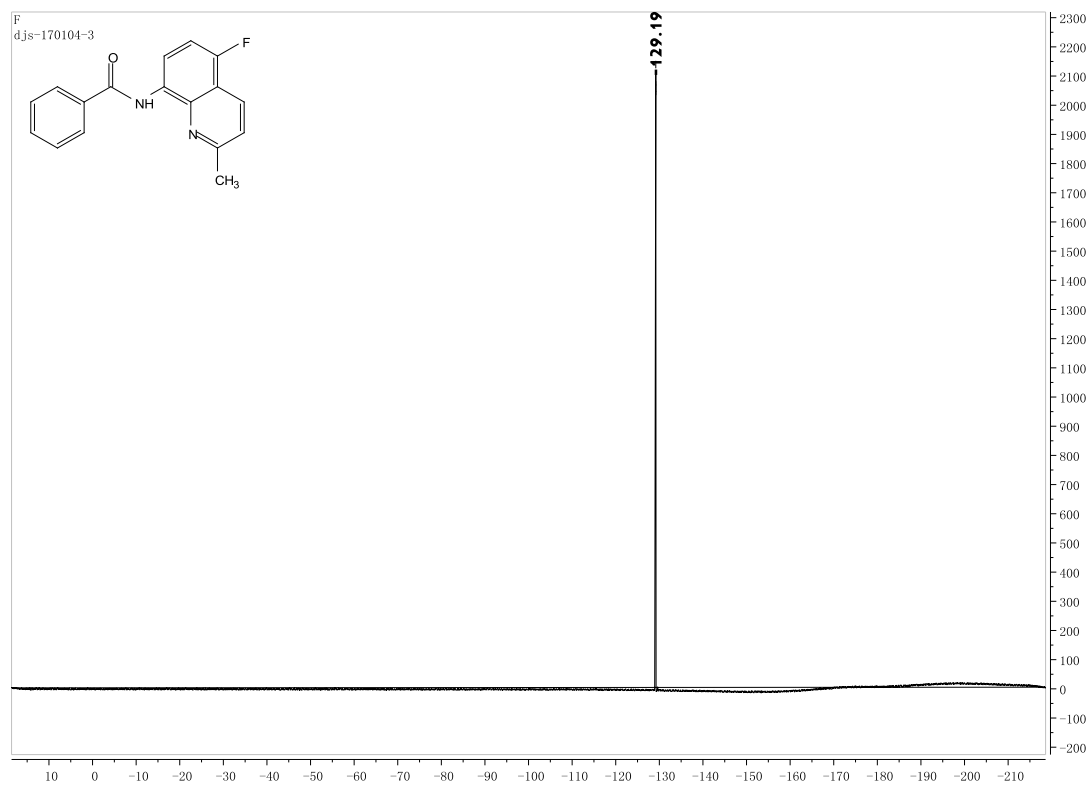
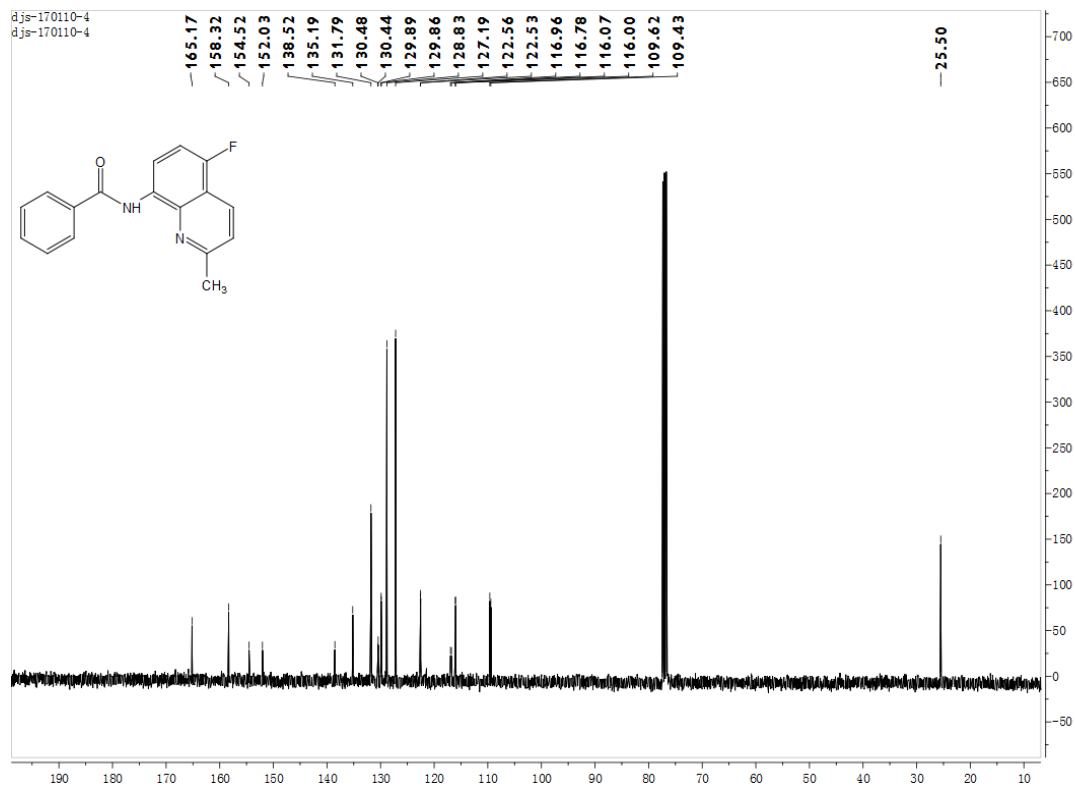
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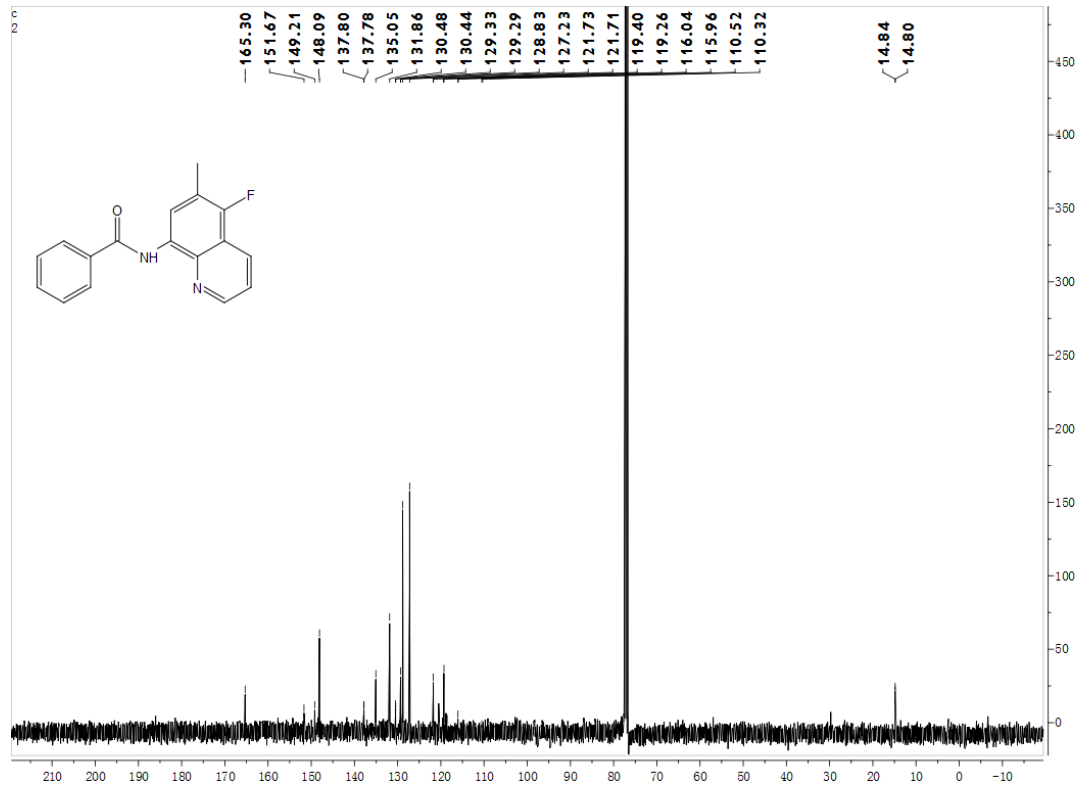
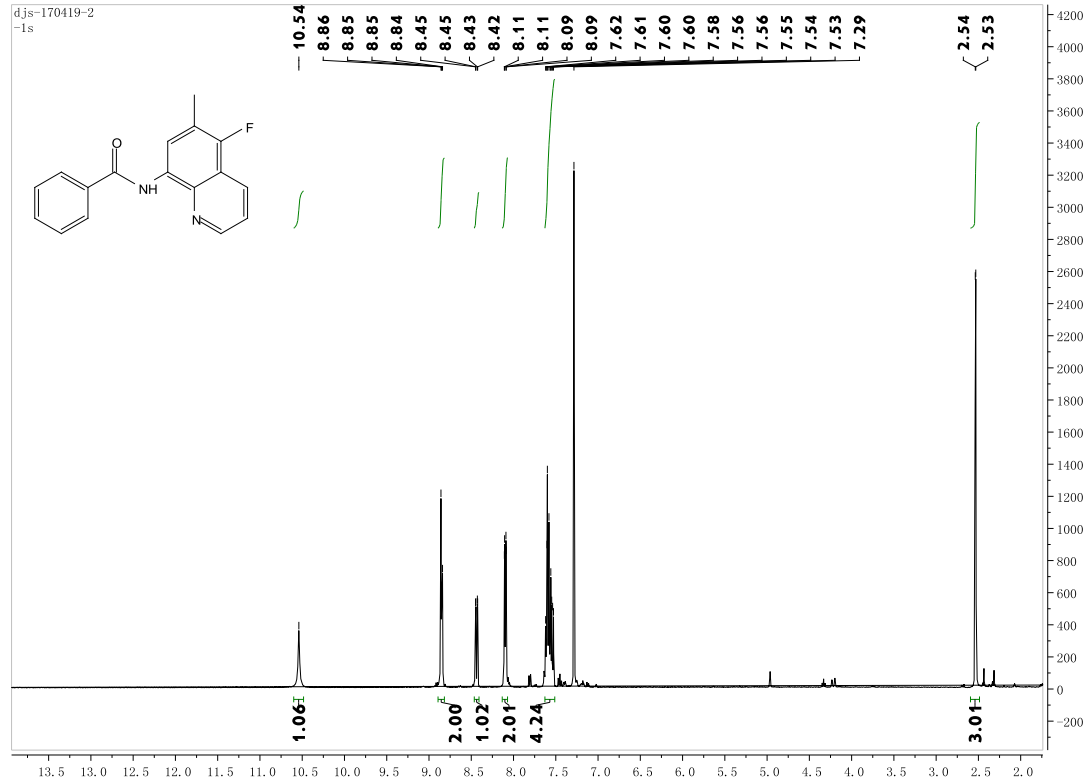


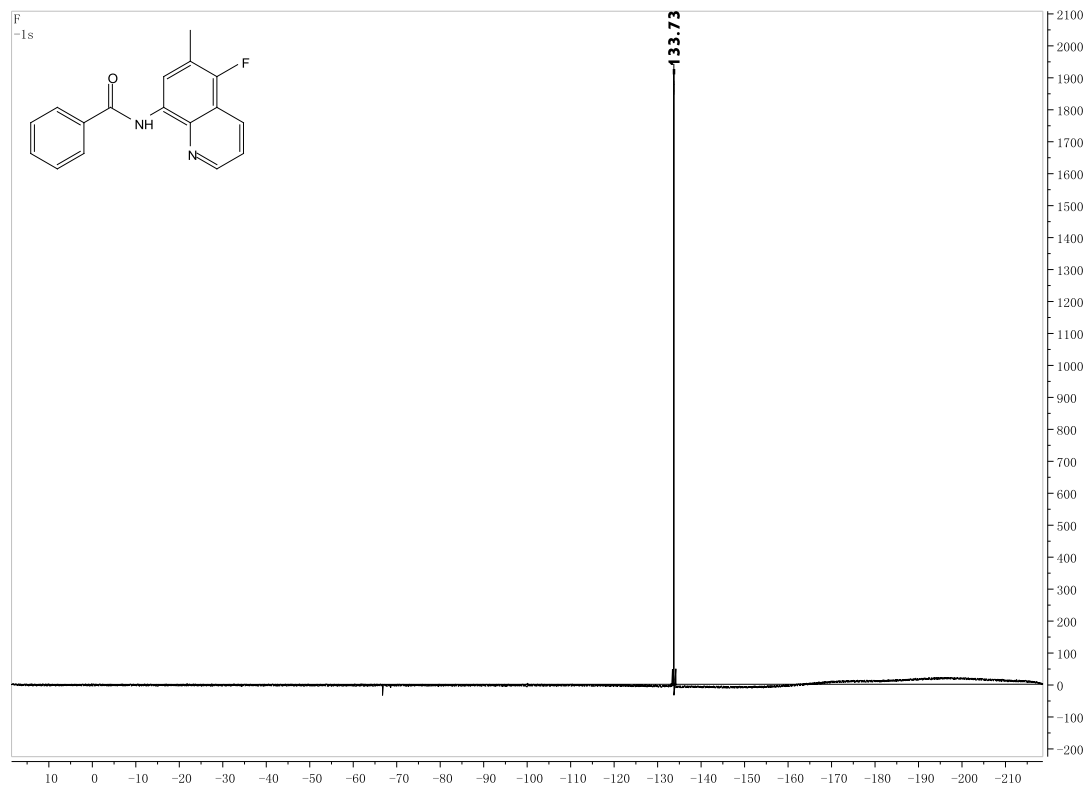
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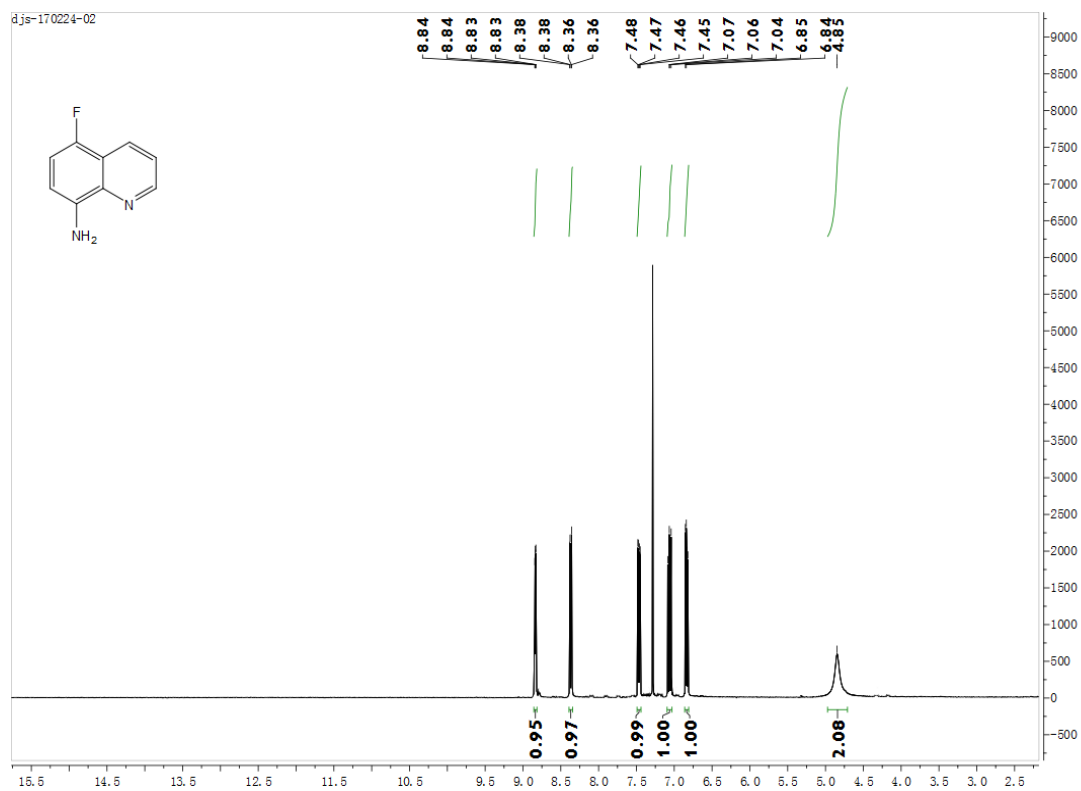


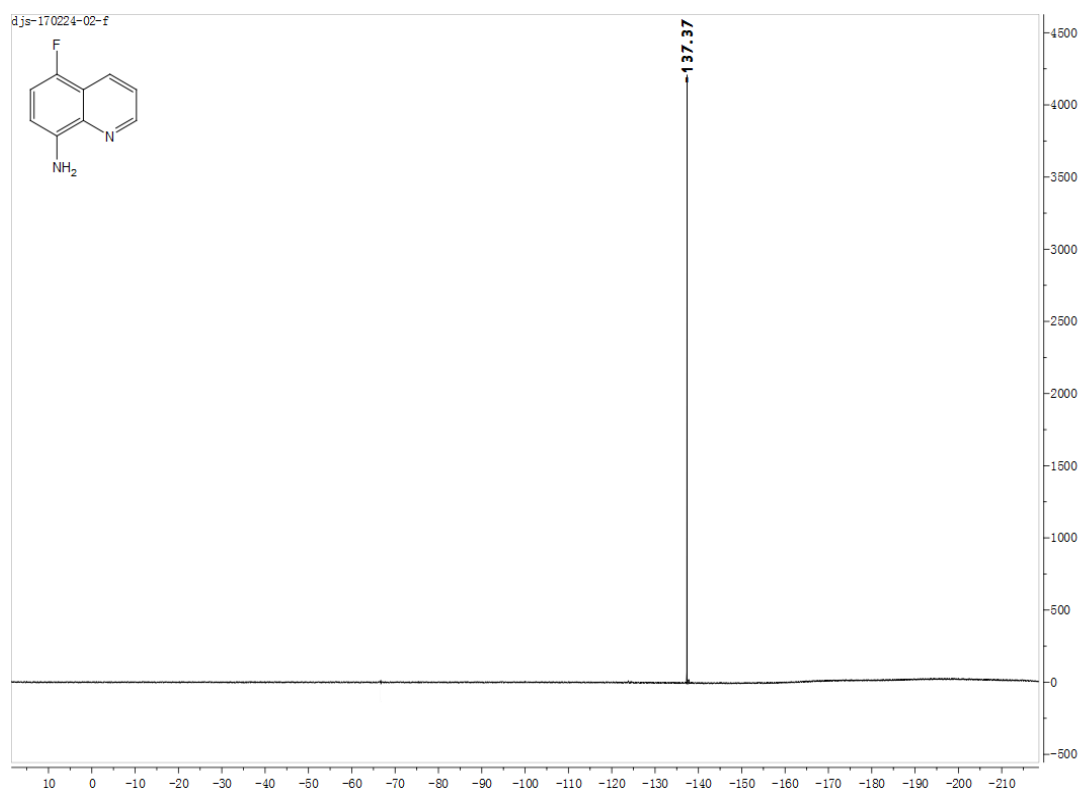
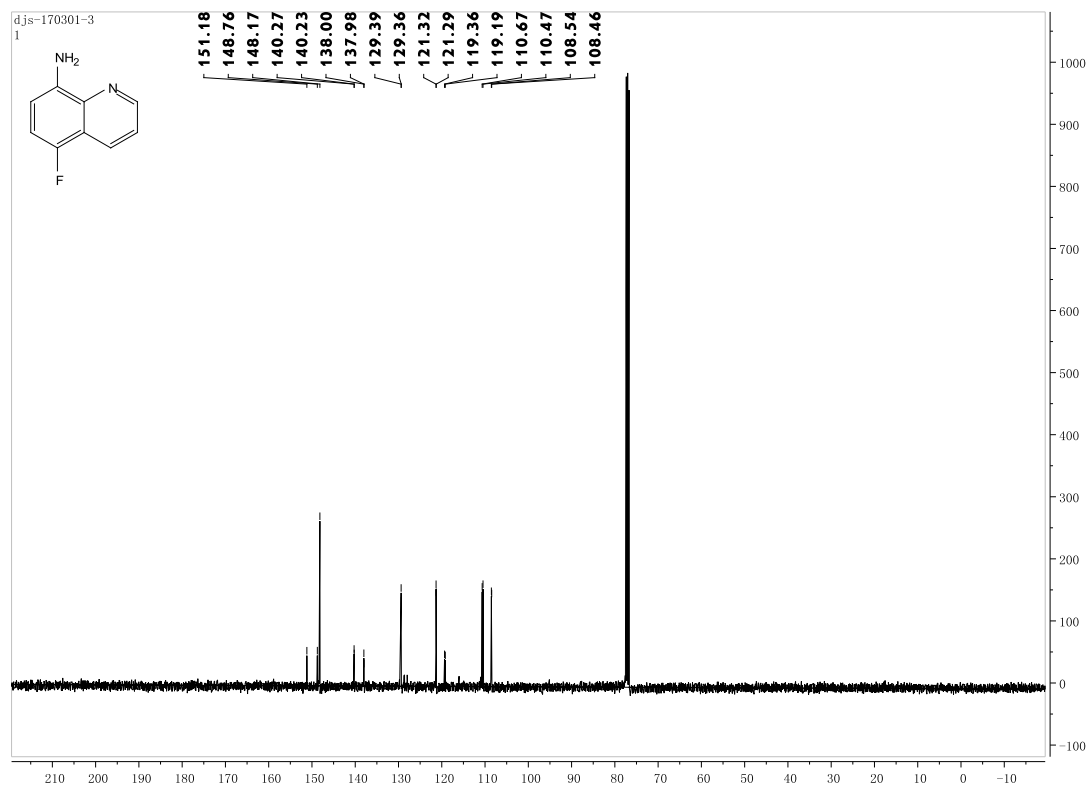
2p:





9:





8. Reference

1 Z. Wang, J. Z. Ni, Y. Kuninobu, and M. Kanai, *Angew. Chem.*, 2014, *126*, 3564-3567.