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Supporting Information

for

A New Approach to Arylhydrazides via the Reaction of Mitsunobu

Reagent with Arynes: Further Application to Access Diverse Nitrogen-

containing Heterocycles in One Pot

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List of content

1. General ·····	••••••S2
2. General Experimental Procedure	••••••S2
3. Characterization Data of the Products	S3-S6
4. NMR spectra ·····	······S7-S26

1. General

All isolated compounds were characterized on Bruker 400 and JEOL 400 MHz spectrometers in the CDCl₃, CD₃OD or (CD₃)₂CO. Chemical shifts are reported as δ values relative to internal chloroform (δ 7.26 for ¹H NMR and 77.16 for ¹³C NMR), methanol (δ 3.31 for ¹H NMR) and acetone (δ 2.05 for ¹H NMR and 29.84 for ¹³C NMR). High resolution mass spectra (HRMS) were obtained on a 4G mass spectrometer by using electrospray ionization (ESI) analyzed by quadrupole time-of-flight (QTof). All melting points were measured with the samples after column chromatography and uncorrected. Column chromatography was performed on silica gel. Anhydrous THF, PhMe were distilled over sodium benzophenone ketyl under Ar. All other solvents and reagents were used as obtained from commercial sources without further purification.

2. General Experimental Procedure

2.1 General Procedure for the Preparation of Arylhydrazines 3a-3j

To a solution of benzyne precursor **1a** (149 mg, 0.500 mmol) and diisopropyl azodicarboxylate (**2a**, 212 mg, 1.05 mmol) in CH₃CN (2.5 mL) were added 18-Crown-6 (330 mg, 1.25 mmol) and Ph₃P (275 mg, 1.05 mmol) at rt. After 10 mins, CH₃CN (2.5 mL, containing H₂O 4.32 g/L) and CsF (152 mg, 1.00 mmol) were added successively. The mixture was heated to 50 °C and kept for 4 h. The solvent was removed under reduced pressure and the residue was diluted with EtOAc (50 mL) and washed with H₂O three times. The organic layer was dried with MgSO₄, filtered, and concentrated *in vacuo*. The resulting mixture was purified by flash chromatography to respectively give **3a** (116 mg, 83%) as a light yellow oil and **4a** as a light yellow oil (4 mg, 3%).

Other Arylhydrazines were prepared following the similar method: 1 (0.5 mmol), azodicarboxylate/PPh₃ (2.1 equiv.), CsF (2.0 equiv.), 18-Crown-6 (2.5 equiv.), H₂O (1.2 equiv.), solvent (5 mL), 50 °C.

2.2 General Procedure for the Preparation of Heterocycles 5a-5c

5a & 5c To a solution of benzyne precursor **1a** (149 mg, 0.500 mmol) and azodicarboxylate **2c** (242 mg, 1.05 mmol) in CH₃CN (2.5 mL) were added 18-Crown-6 (330 mg, 1.25 mmol) and Ph₃P (275 mg, 1.05 mmol) at rt. After 10 min, CH₃CN (2.5 mL, containing H₂O 4.32 g/L) and CsF (152 mg, 1.00 mmol) were added successively. The mixture was heated to 50 °C and kept for 4 h. The solvent was removed under reduced pressure.

To a solution of the above residue and cycloheptanone (2.0 equiv.) or acetylacetone (2.0 equiv.) in AcOH (5.0 mL) was added ZnCl₂ (68 mg, 0.5 mmol) at rt. The mixture was heated to 130 °C and kept for 12 h. The solvent was removed under reduced pressure and the residue was diluted with EtOAc and washed with H₂O three times. The organic layer was dried with MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography to respectively give **5a** (52 mg, 56% for 3 steps) as a white solid or **5c** (55 mg, 64% for 3 steps) as a yellow oil.

5b 1-tetralone (2.0 equiv.) was used as the ketone. In accordance with the above operation, for the second step, the mixture was stirred under O_2 , heated at 130 °C and kept for 24 h. The solvent was removed under reduced pressure and the residue was diluted with EtOAc and washed with H₂O three times. The organic layer was dried with MgSO₄, filtered, and concentrated *in vacuo*. The residue was purified by flash chromatography to give **5b** (58 mg, 53% for 4 steps) as a white solid.

3. Characterization Data of the Products



4a (4 mg, Y = 3%, $R_f = 0.64$ (PE:EA = 5:1)) as a light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 8.03 (d, *J* = 6.4 Hz, 1H), 7.66 (d, *J* = 7.6 Hz, 1H), 7.50 (t, *J* = 7.6 Hz, 1H), 7.25 (t, *J* = 7.2 Hz, 1H), 5.36 (q, *J* = 6.0 Hz, 1H), 5.29 (q, *J* = 6.0 Hz, 1H), 1.49 (d, *J* = 6.4 Hz, 6H), 1.46 (d, *J* = 6.0 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 159.1, 150.7, 141.0, 129.7, 123.1, 120.2, 118.1, 114.8, 72.6, 71.6, 22.2, 22.1; ESI-HRMS *m/z* Calcd. for C₁₄H₁₈N₂O₃ + Na (M+Na): 285.1210, found 285.1214.



3a (116 mg, Y = 83%, $R_f = 0.32$ (PE:EA = 5:1) as a light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.41 (br s, 2H), 7.33-7.29 (m, 2H), 7.18-7.15 (m, 2H), 5.05-4.94 (m, 2H), 1.26 (d, *J* = 6.0 Hz, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 156.2, 154.5, 141.9, 128.6, 126.0, 124.0, 70.9, 70.1, 22.0, 22.0; ESI-HRMS *m/z* Calcd. for C₁₄H₂₀N₂O₄ + Na (M+Na): 303.1315, found 303.1314.

3b (97 mg, Y = 77%, $R_f = 0.16$ (PE:EA = 5:1)) as a light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.42 (d, J = 8.0 Hz, 2H), 7.35-7.31 (m, 2H), 7.20 (t, J = 7.2 Hz, 1H), 7.15 (br s, 1H), 4.28-4.18 (m, 4H), 1.27 (t, J = 6.8 Hz, 6H); ¹³C NMR (100 MHz, CDCl₃) δ 156.5, 155.1, 141.8, 128.8, 126.5, 124.4, 63.1, 62.4, 14.6; ESI-HRMS *m*/*z* Calcd. for $C_{12}H_{16}N_2O_4$ + Na (M+Na): 275.1002, found 275.1003.



3c (111 mg, Y = 72%, $R_f = 0.49$ (PE:EA = 5:1)) as a light yellow solid. m.p. 105-106 °C. ¹H NMR (400 MHz, CDCl₃) δ 7.40 (br s, 2H), 7.30 (t, *J* = 7.6 Hz, 2H), 7.14 (t, *J* = 7.2 Hz, 1H), 6.91-6.74 (m, 1H), 1.49 (s, 18H); ¹³C NMR (100 MHz, CDCl₃) δ 155.5, 153.7, 142.2, 128.5, 125.6, 123.8, 82.3, 81.5, 28.3, 28.2; ESI-HRMS *m*/*z* Calcd. for C₁₆H₂₄N₂O₄ + Na (M+Na): 331.1628, found 331.1629.



3d (133 mg, Y = 71%, $R_f = 0.27$ (PE:EA = 5:1)) as a light yellow oil. ¹H NMR (300 MHz, CD₃OD) δ 7.32-7.21 (br s, 15H), 5.18 (s, 2H), 5.12 (s, 2H); ¹H NMR (300 MHz, CDCl₃) δ 7.32-7.26 (br s, 16H), 5.21 (s, 2H), 5.18 (s, 2H); ¹³C NMR (100 MHz, CDCl₃) δ 156.3, 154.8, 141.5, 135.7, 135.6, 128.8, 128.7, 128.6, 128.6, 128.4, 128.3, 128.0, 126.6, 124.5, 68.5, 68.0; ESI-HRMS *m/z* Calcd. for C₂₂H₂₀N₂O₄ + Na (M+Na): 399.1315, found 399.1316.

3e (142 mg, Y = 83%, $R_f = 0.30$ (PE:EA =2:1)) as a yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 7.03 (br s, 1H), 6.95-6.93 (m, 2H), 6.81-6.79 (m, 1H), 5.04-4.96 (m, 2H), 3.86 (s, 3H), 3.85 (s, 3H), 1.28-1.25 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 156.1, 154.8, 148.6, 147.5, 135.2, 116.8, 110.7, 109.2, 70.6, 69.9, 56.0, 55.8, 22.0, 22.0; ESI-HRMS *m*/*z* Calcd. for C₁₆H₂₄N₂O₆ + Na (M+Na): 363.1527, found 363.1528.

$$\overbrace{O}^{CO_2'Pr}_{H}$$

3f (113 mg, Y = 70%, $R_f = 0.24$ (PE:EA = 5:1)) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 6.99 (br s, 1H), 6.94 (br s, 1H), 6.86 (br s, 1H), 6.74-6.72 (m, 1H), 5.95 (s, 2H), 5.03-4.93 (m, 2H), 1.27-1.23 (m, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 156.1, 154.7, 147.5, 146.1, 136.0, 118.4, 107.7, 106.9, 101.5, 70.8, 69.9, 22.0, 21.9; ESI-HRMS *m/z* Calcd. for C₁₅H₂₀N₂O₆ + Na (M+Na): 347.1214, found 347.1213.

3g (The regioselectivity was confirmed through comparison of the ¹³C-NMR of **3g** with analogous *m*-methoxylarylhydrazide. See **Fig. S35**) (94 mg, Y = 60%, R_f = 0.25 (PE:EA = 5:1)) as a light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.20 (t, *J* = 8.0 Hz, 1H), 7.09-7.02 (m, 3H), 6.72 (d, *J* = 7.6 Hz, 1H), 5.02-4.97 (m, 2H), 3.77 (s, 3H), 1.26 (d, *J* = 5.6 Hz, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 159.8, 156.1, 154.3, 143.0, 129.2, 116.0, 111.8, 109.7, 70.9, 70.1, 55.4, 22.0, 22.0; ESI-HRMS *m/z* Calcd. for C₁₅H₂₂N₂O₅ + Na (M+Na): 333.1421, found 333.1420.

$$\underbrace{\overset{CO_2 i Pr}{\overset{}_{H}}}_{H} \overset{CO_2 i Pr}{\overset{}_{H}}$$

3h (131 mg, Y = 85%, $R_f = 0.33$ (PE:EA = 5:1)) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.20 (br s, 1H), 7.12 (br s, 2H), 7.08-7.06 (m, 1H), 5.05-4.96 (m, 2H), 2.23 (s, 3H), 2.22 (s, 3H), 1.26 (d, *J* = 6.4 Hz, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 156.1, 154.7, 139.6, 136.8, 134.7, 129.7, 125.5, 121.8, 70.7, 69.9, 22.0, 22.0, 19.9, 19.3; ESI-HRMS *m*/*z* Calcd. for C₁₆H₂₄N₂O₄ + Na (M+Na): 331.1628, found 331.1629.

3i (141 mg, Y = 88%, $R_f = 0.32$ (PE:EA = 5:1)) as a light yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.28 (br s, 1H), 7.18 (br s, 1H), 7.15 (br s, 2H), 5.03-4.96 (m, 2H), 2.87 (q, *J* = 7.2 Hz, 4H), 2.10-2.02 (m, 2H), 1.26 (d, *J* = 6.4 Hz, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 156.1, 154.8, 144.7, 142.4, 140.1, 124.1, 122.7, 120.8, 70.6, 69.8, 32.9, 32.4, 25.6, 22.0, 22.0; ESI-HRMS *m/z* Calcd. for C₁₇H₂₄N₂O₄ + Na (M+Na): 343.1628, found 343.1630.

3j (67 mg, Y = 42%, $R_f = 0.29$ (PE:EA = 5:1)) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.35 (br s, 1H), 7.18 (br s, 1H), 7.13-7.06 (m, 1H), 6.87-6.75 (m, 1H), 5.05-4.97 (m, 2H), 1.28 (d, J =

6.4 Hz, 12H); ¹³C NMR (75 MHz, CDCl₃) δ 156.1, 154.1, 151.4 (*J* = 13.5Hz), 148.1 (*J* = 13.6 Hz), 138.3 (dd, *J* = 7.5, 3.0 Hz), 119.5, 116.8 (*J* = 18.0 Hz), 113.5, 71.5, 70.6, 22.0, 22.0; ESI-HRMS *m*/*z* Calcd. for C₁₄H₁₈F₂N₂O₄ + Na (M+Na): 339.1127, found 339.1128.



3k (The regioselectivity was confirmed through comparison of the ¹³C-NMR of **3k** with analogous *m*-alkoxylarylhydrazide like **3g**) (108 mg, Y = 62%, $R_f = 0.22$ (PE:EA = 5:1)) as a light yellow oil. ¹H NMR (300 MHz, CDCl₃) δ 7.22 (d, *J* = 8.4 Hz, 1H), 7.08 (br s, 2H), 6.85-6.64 (m, 2H), 5.06-4.96 (m, 2H), 4.63 (q, *J* = 2.1 Hz, 2H), 1.86 (t, *J* = 2.1 Hz, 3H), 1.28 (d, *J* = 6.0 Hz, 12H); ¹³C NMR (100 MHz, CDCl₃) δ 158.1, 156.1, 154.3, 143.0, 129.3, 116.5, 112.4, 110.7, 84.0, 74.0, 71.1, 70.2, 56.7, 22.1, 22.1, 3.8; ESI-HRMS *m/z* Calcd. for C₁₈H₂₄N₂O₅+ Na (M+Na): 371.1577, found 371.1574.



31 (ratio of *o:m* = 1:4 according to the integrals of allylic hydrogens. The minor isomer was further confirmed through the transformation of E1cB eliminative cleavage of the N-N'-bond reported by Magnus (*Org. Lett.* **2009**, *11*, 5646), and the crude ¹H-NMR of which could be found in **Fig. S36**. It could be easily compared with known compounds. See **Fig. S37 and Fig. S38**) (112 mg, Y = 70%, R_f = 0.33 (PE:EA = 5:1)) as a yellow oil. ¹H NMR (300 MHz, (CD₃)₂CO) δ 8.83 (s, 0.80H), 8.50 (s, 0.20H), 7.59-7.20 (m, 4H), 7.07-6.97 (m, 1H), 6.04-5.90 (m, 1H), 5.24-4.83 (m, 4H), 3.51 (d, *J* = 6.0 Hz, 0.40H) 3.39 (d, *J* = 6.0 Hz, 1.6H), 1.42-1.09 (m, 12H); ¹³C NMR (75 MHz, (CD₃)₂CO) δ 156.8, 154.8, 143.6, 141.0, 138.2, 129.1, 127.6 (minor isomer), 126.3, 124.4, 121.9, 116.1, 40.6, 22.2, 22.2; ESI-HRMS *m/z* Calcd. for C₁₇H₂₄N₂O₄ + Na (M+Na): 343.1628, found 343.1622.

3m (The regioselectivity was readily observed from the ¹H-NMR of **3m**) (DIAD/PPh₃ (4.2 equiv.) were used instead, 50 mg, Y = 27%, $R_f = 0.37$ (PE:EA = 5:1)) as a yellow oil. ¹H NMR (300 MHz, (CD₃)₂CO) δ 8.71 (s, 1H), 7.39 (s, 1H), 7.18 (s, 1H), 7.05 (s, 1H), 4.85-4.74 (m, 2H), 2.19 (s, 3H), 1.13 (d, *J* = 3.9 Hz, 6H), 1.11 (d, *J* = 4.2 Hz, 6H); ¹³C NMR (75 MHz, (CD₃)₂CO) δ 156.8, 154.5, 144.7, 141.2, 129.3, 123.6, 123.0, 121.8, 71.1, 70.0, 22.2, 22.2, 21.2; ESI-HRMS *m/z* Calcd. for C₁₅H₂₁BrN₂O₄ + Na (M+Na): 395.0577, found 395.0575.



5a (52 mg, Y = 56%, $R_f = 0.66$ (PE:EA = 5:1)) as a white solid. m.p. 133-134 °C. ¹H NMR (300 MHz, CDCl₃) δ 7.68 (s, 1H), 7.48-7.45 (m, 1H), 7.27-7.24 (m, 1H), 7.09-7.06 (m, 2H), 2.85-2.80 (m, 4H), 1.90-1.87 (m, 2H), 1.80-1.77 (m, 4H); ¹³C NMR (100 MHz, CDCl₃) δ 137.5, 134.4, 129.4, 120.7, 119.1, 117.8, 113.8, 110.3, 31.9, 29.6, 28.8, 27.6, 24.8; ESI-HRMS *m/z* Calcd. for C₁₃H₁₅N + H (M+H): 186.1277, found 186.1278.



5b (58 mg, Y = 53%, $R_f = 0.51$ (PE:EA = 5:1)) as a white solid. m.p. 222-223 °C. ¹H NMR (400 MHz, (CD₃)₂CO) δ 11.28 (s, 1H), 8.49 (d, *J* = 8.4 Hz, 1H), 8.21 (d, *J* = 8.4 Hz, 1H), 8.18 (d, *J* = 8.0 Hz, 1H), 8.04 (d, *J* = 8.0 Hz, 1H), 7.68-7.59 (m, 3H), 7.56-7.52 (m, 1H), 7.41 (td, *J* = 7.2, 1.2 Hz, 1H), 7.28-7.24 (m, 1H); ¹³C NMR (100 MHz, (CD₃)₂CO) δ 140.1, 136.3, 133.4, 129.6, 126.3, 126.0, 125.5, 124.8, 122.5, 122.3, 120.5, 120.5, 120.3, 120.2, 118.9, 112.2; ESI-HRMS *m/z* Calcd. for C₁₆H₁₁N + H (M+H): 218.0964, found 218.0965.



5c (55 mg, Y = 64%, $R_f = 0.53$ (PE:EA = 5:1)) as a yellow oil. ¹H NMR (400 MHz, CDCl₃) δ 7.43-7.42 (m, 4H), 7.34-7.32 (m, 1H), 5.99 (s, 1H), 2.30 (s, 3H), 2.29 (s, 3H); ¹³C NMR (100 MHz, CDCl₃) δ 149.0, 140.0, 139.4, 129.0, 127.3, 124.8, 107.0, 13.6, 12.4; ESI-HRMS *m/z* Calcd. C₁₁H₁₂N₂ + H (M+H): 173.1073, found 173.1074.

4. NMR spectra



Fig. S1. ¹H NMR of compound 4a (400 MHz, CDCl₃).



Fig. S2. ¹³C NMR of compound 4a (100 MHz, CDCl₃).



Fig. S3. ¹H NMR of compound 3a (400 MHz, CDCl₃).



Fig. S4. ¹³C NMR of compound 3a (100 MHz, CDCl₃).



Fig. S5. ¹H NMR of compound 3b (400 MHz, CDCl₃).



Fig. S6. ¹³C NMR of compound 3b (100 MHz, CDCl₃).



Fig. S7. ¹H NMR of compound 3c (400 MHz, CDCl₃).



Fig. S8. ¹³C NMR of compound 3c (100 MHz, CDCl₃).



Fig. S9'. ¹H NMR of compound 3d (300 MHz,CDCl₃).



Fig. S10. ¹³C NMR of compound 3d (100 MHz, CDCl₃).



Fig. S11. ¹H NMR of compound 3e (300 MHz, CDCl₃).



Fig. S12. ¹³C NMR of compound 3e (100 MHz, CDCl₃).



Fig. S13. ¹H NMR of compound 3f (400 MHz, CDCl₃).



Fig. S14. ¹³C NMR of compound 3f (100 MHz, CDCl₃).



Fig. S15. ¹H NMR of compound 3g (400 MHz, CDCl₃).



Fig. S16. ¹³C NMR of compound 3g (100 MHz, CDCl₃).



Fig. S17. ¹H NMR of compound **3h** (400 MHz, CDCl₃).



Fig. S18. ¹³C NMR of compound 3h (100 MHz, CDCl₃).



Fig. S19. ¹H NMR of compound 3i (400 MHz, CDCl₃).



Fig. S20. ¹³C NMR of compound 3i (100 MHz, CDCl₃).



Fig. S21. ¹H NMR of compound 3j (400 MHz, CDCl₃).



Fig. S23. ¹H NMR of compound 3k (300 MHz, CDCl₃).

Fig. S24. ¹³C NMR of compound 3k (100 MHz, CDCl₃).

Fig. S25. ¹H NMR of compound **31** (300 MHz, (CD₃)₂CO).

Fig. S27. ¹H NMR of compound **3m** (300 MHz, (CD₃)₂CO).

Fig. S29. ¹H NMR of compound 5a (300 MHz, CDCl₃).

Fig. S30. ¹³C NMR of compound 5a (100 MHz, CDCl₃).

Fig. S31. ¹H NMR of compound 5b (400 MHz, (CD₃)₂CO).

Fig. S32. ¹³C NMR of compound 5b (100 MHz, (CD₃)₂CO).

Fig. S33. ¹H NMR of compound 5c (400 MHz, CDCl₃).

Fig. S34. ¹³C NMR of compound 5c (100 MHz, CDCl₃).

Fig. S35. ¹³C NMR (100 MHz, CDCl₃): *δ* 159.4, 155.2, 153.3, 143.1, 126.8, 115.6, 111.0, 109.1, 82.0, 81.2, 55.0, 28.0, 27.9.

Fig. S37. ¹H NMR (400 MHz, CDCl₃) δ 7.25-7.15 (m, 3H), 6.88 (d, *J* = 7.2 Hz, 1H), 6.44 (s, 1H),

5.96 (ddt, *J* = 16.9, 10.1, 6.7 Hz, 1H), 5.17-4.97 (m, 2H), 3.37 (d, *J* = 6.7 Hz, 2H), 1.52 (s, 9H).

Fig. S38. ¹H NMR (400 MHz, CDCl₃) δ 7.81 (d, J = 7Hz, 1H), 7.26 (t, J = 8Hz, 1H), 7.17 (d, J = 7Hz, 1H), 7.07 (t, J = 7Hz, 1H), 6.48 (s (br), 1H), 5.99 (ddt, J = 17, 10, 6 Hz, 1H), 5.20-5.17 (m, 1H), 5.11-5.07 (m, 1H), 3.39 (d, J = 6 Hz, 2H), 1.54 (s, 9H).