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Supporting Information for

Synthesis and Evaluation of Novel *O*²-Derived Diazeniumdiolates as Photochemical and Real-time Monitoring Nitric Oxide Delivery Agents

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1. General information

All reagents and solvents (analytical grade) were purchased commercially and used without further purification unless otherwise noted. Ultraviolet lighter (150W, 0.02 W/cm², TLS3009-X105) was used to supply UV light (365-400 nm), and visible lighter (20W, 0.05 W/cm², HL-2000) was used as visible light (\geq 410 nm) source. Melting points were determined on a X-4 precision micro-melting point detector. Infrared (IR) spectra (KBr) were recorded on a Nicolet iS10 FT-IR instrument (KBr pellet). ¹H NMR and ¹³C NMR spectra were recorded with a Bruker Avance 300 MHz spectrometer at 303 K, using TMS as an internal standard. MS spectra were recorded on a Mariner Mass Spectrum (ESI). High resolution mass measurement was performed on Agilent QTOF 6520 mass spectrometer with electron spray ionization (ESI) as the ion source. Analytical and preparative TLC was performed on silica gel (200-300 mesh) GF/UV 254 plates, and the chromatograms were visualized under UV light at 254 and 365 nm. Analytical reversed phase high performance liquid chromatography (RP-HPLC) was performed on an Shimadzu LC-20AT system, and monitored using a diode array UV/Vis detector. A Venusil MP C18 column $(250 \text{ mm} \times 4.6 \text{ mm} \times 5 \text{ }\mu\text{m})$ was used for purity analysis at a flow rate of 1 mL/min. Solutions after reactions and extractions were concentrated using a rotary evaporator operating at a reduced pressure of ca. 20 Torr. The excitation and emission spectrum were determined by Infinite® 200 Pro.

Preparation procedures and characterization data for compounds 1-4, 6, 10-12 2-bromo-1-(4-hydroxyphenyl) ethan-1-one (6).¹



To a solution of 4-hydroxyacetophenone (**5**, 1.36g, 10 mmol) in ethyl acetate (100 mL), CuBr₂ (3.5g, 15 mmol) was added slowly. The reaction mixture was stirred at room temperature for about 24h. The precipitate was removed by filtration. The obtained clear solution was washed with brine (50 mL \times 3), and dried over anhydrous Na₂SO₄. After filtration, the organic solution was concentrated, and the residue was purified by recrystalization with ethanol to yield of 2-bromo-1-(4-hydroxyphenyl) ethan-1-one (1.5 g, 69.7%). mp: 124-125 °C. ¹H NMR (DMSO_{d6}, 300 MHz): 4.79 (s, 2H, CH₂Br), 6.89 (dd, 2H, *J*=16, 9 Hz, 2ArH), 7.88 (dd, 2H, *J*=16, 9 Hz, 2ArH), 10.56 (s, 1H, OH). IR (KBr, cm⁻¹): 3271, 1676, 1222, 698. ESI-MS: [M + H]⁺, found 216 m/z C₈H₇BrO₂ requires 215.

O²-(p-Hydroxyphenacyl)-1-(N,N-diethylamino)diazen-1-ium-1,2-diolate (1).



To a solution of 1-(*N*,*N*-Diethylamino)diazen-1-ium-1,2-diolate (7)^{2,3} (155 mg, 1 mmol) in dry *N*,*N*-dimethylformamide (DMF, 2 mL) was added compound **6** (215 mg, 1 mmol) under an inert atmosphere at -10 °C. The obtained solution was allowed to stir at this condition for 2.5h. After completion of the reaction (as monitored by TLC), DMF was removed under vacuum. The resulting crimson oil was dissolved in dichloromethane (20 mL) and washed with brine (20 mL × 3). The organic layer was dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation to yield the crude product, which was purified on a silica column using 20 % ethyl acetate in petroleum ether to afford **2** as a white solid (59.8 mg, 22.4%). mp: 144-145 °C. ¹H NMR (300 MHz, CDCl₃): 1.09 (t, *J*=7.1 Hz, 6 H, CH₃ × 2), 3.15 (q, *J* = 7.1 Hz, 4 H, NCH₂ × 2), 5.37 (s, 2 H, OCH₂), 6.88-6.92 (m, 2H, 2ArH), 7.76-7.80 (m, 2H, 2ArH). ¹³C NMR (75 MHz, CDCl₃): δ 10.9, 48.2, 73.9, 115.4, 126.3, 130.1, 161.1, 191.2. IR (KBr, cm⁻¹): 3174, 1671, 1229, 1096. ESI-MS: [M - H]⁻, found 266.1 C₁₂H₁₇N₃O₄ requires 267.2. HRMS m/z calcd for C₁₂H₁₆N₃O₄ [M - H]⁻, 266.1141. found 266.1136. ppm error 1.9.

O²-(p-hydroxyphenacyl)-1-(N,N-morpholine)diazen-1-ium-1,2-diolate (2).



The title compound was prepared from the reaction of **6** with 1-(*N*,*N*-morpholine)diazen-1ium-1,2-diolate **8** by using the same method as compound **1**, as a white solid (90 mg, 32%). mp: 166-167 °C. ¹H NMR (DMSO_{*d6*}, 300 MHz): 3.28 (t, *J* = 4.8 Hz, 4H, NCH₂ × 2), 3.71 (t, *J* = 4.8 Hz, 4H, OCH₂ × 2), 5.53 (s, 2H, OCH₂), 6.87 (d, *J* = 8.7 Hz, 2H, 2ArH), 7.83 (d, *J* = 8.7 Hz, 2H, 2ArH), 10.51 (s, 1H, OH). ¹³C NMR (DMSO_{*d6*}, 75 MHz): δ 51.6, 74.6, 115.9, 126.2, 130.9, 163.1, 192.2. IR (KBr, cm-1): 3173, 1676, 1243, 1112,1090. ESI-MS: [M-H]⁻, found 280.1. C₁₂H₁₅N₃O₅ requires 281.1. HRMS m/z calcd for C₁₂H₁₄N₃O₅ [M-H]⁻ : 280.0933. found 280.0939. ppm error 2.1.

5-(2-bromoacetyl)-2-hydroxybenzaldehyde (10).⁴



To a suspension of aluminum chloride (AlCl₃) (1.06 g, 8 mmol) in 60 mL of dichloromethane (DCM), bromoacetyl bromide (480 mg, 2.4 mmol) was added slowly at 10 °C. The temperature of the mixture was brought to 30 °C and the mixture was stirred for an hour. Then a solution of salicylaldehyde (9) (245 mg, 2 mmol) in DCM (10 mL) was added slowly. The obtained mixture was stirred at rt for 15h. Then the reaction was quenched by adding cool water (50 mL). The DCM layer was separated and the aqueous layer was extracted with DCM (30 mL × 2), and dried over anhydrous Na₂SO₄. Removal of solvent generated crude slurry, which was added n-heptane (20 mL) and stirred for 15 min. The precipitate was filtrated, washed with n-heptane and dried at 50 °C, to yield compound **10** as a yellow solid (192 mg; 60%). mp 57-58 °C. ¹H NMR (300 MHz, CDCl₃): 4.40 (s, 2H, CH₂Br), 7.10 (d, *J* = 8.8 Hz, 1H, ArH), 8.17 (1H, dd, *J* = 8.8, 2.2 Hz, ArH), 8.30 (d, J = 2.2 Hz, 1H, ArH), 9.99 (s, 1H, OH), 11.53 (s, 1H, CHO). IR (KBr, cm⁻¹): 3250, 2877, 1725, 1681, 1197, 662. ESI-MS: [M+Cl]⁻, found 278 m/z C₉H₈BrO₃ requires 243.1.

*O*²-(5-(2-acetoxy)-2-hydroxybenzaldehyde)-1-(*N*,*N*-diethylamino)diazen-1-ium-1,2-diolate (11).



To a solution of 1-(*N*,*N*-Diethylamino)diazen-1-ium-1,2-diolate (7) ^{2,3} (155 mg, 1 mmol) in dry tetrahydrofuran (THF, 3 mL) and dimethyl sulfoxide (DMSO, 0.5 mL) was added compound **10** (243mg, 1 mmol) under an inert atmosphere at -10 °C. The obtained solution was allowed to stir at this condition for 3.5h. After completion of the reaction (as monitored by TLC), DMF was removed under vacuum. The resulting crimson oil was dissolved in dichloromethane (20 mL) and washed with brine (20 mL × 3). The organic layer was dried over anhydrous Na₂SO₄ and concentrated by rotary evaporation to yield the crude product, which was purified on a silica column using 20 % ethyl acetate in petroleum ether to afford **11** as a white acicular crystal (77.3 mg, 26.2%). mp: 91-92 °C. ¹H NMR (300MHz, CDCl₃) 1.03 (t, *J* = 7.1 Hz, 6H, CH₃ × 2), 3.10 (q, *J* = 7.1 Hz, 4H, CH₂ × 2), 5.34 (s, 2H, OCH₂), 7.05 (d, 1H, *J* = 8.8 Hz, ArH), 8.11(dd, 2H, *J* = 8.8, 2.2 Hz, ArH), 8.27(d, *J* = 2.2 Hz, 1H, ArH), 9.95 (s, 1H, OH), 11.46 (s, 1H, CHO). ¹³C NMR (75 MHz, CDCl₃): δ 11.4, 48.5, 74.7, 118.4, 120.1, 126.9, 135.0, 136.4, 165.6, 191.0, 196.3. IR (KBr,

cm⁻¹): 3427, 2853, 1654, 1230, 1091. ESI-MS: $[M + Na]^+$, found 318.1 m/z C₁₃H₁₇N₃O₅ requires 295.1. HRMS m/z calcd for C₁₃H₁₇N₃O₅Na $[M + Na]^+$: 318.1066. found 318.1059. ppm error 2.1.

O²-(5-(2-acetoxy)-2-hydroxybenzaldehyde)-1-(N,N-morpholine)diazen-1-ium-1,2-diolate(12).



The title compound **12** was prepared from the reaction of **10** with 1-(*N*,*N*-morpholine)diazen-1-ium-1,2-diolate **8** by using the same method as compound **11**, as a white solid (102.6 mg, 33.2%). mp: 249-250 °C. ¹H NMR (300 MHz, CDCl₃) 3.34 (d, *J* = 4.4 Hz, 4H, NCH₂× 2), 3.73 (d, *J* = 4.4 Hz, 4H, OCH₂ × 2), 5.24 (s, 2H, OCH₂), 7.01(d, *J* = 8.8 Hz, 1H, ArH), 8.08 (d, *J* = 8.8 Hz, 1H, ArH), 8.25 (s, 1H, ArH), 9.90 (s, 1H, OH), 11.43 (s, 1H, CHO). ¹³C NMR (75 MHz, CDCl₃): δ 51.5, 65.5, 74.8, 118.5, 120.2, 126.8, 135.1, 136.5, 165.7, 191.1, 196.3. IR (KBr, cm⁻¹): 3447, 2857, 1693, 1225, 1180, 1103. ESI-MS: [M + Na]⁺, found 332.1 m/z C₁₃H₁₅N₃O₆ requires 309.1. HRMS m/z calcd for C₁₃H₁₅N₃O₆Na [M + Na]⁺, 332.0859 found 332.0865. ppm error 1.8.

O²-(p-hydroxyphenacyl-benzothiazole)-1-(N,N-diethylamino)diazen-1-ium-1,2-diolate(3).



To a solution of **11** (50 mg, 0.17 mmol) in DMSO (3 mL) was added 2-aminobenzothiol (21 mg, 0.17 mmol). The mixture was stirred at reflux for 2h. The reaction was quenched by adding cool water (20 mL), and then was extracted with ethyl acetate (10 mL × 3). Organic layer was collected, dried over anhydrous Na₂SO₄, and the solvent was removed to produce a brown solid residue, which was purified by column chromatography using 20 % ethyl acetate in petroleum ether to give **11** as a white solid (67.8 mg, 50%). mp: 217-218 °C. ¹H NMR (300MHz, CDCl₃) 1.00 (t, J = 7.1 Hz, 6H, CH₃ × 2), 3.03 (q, J = 7.1 Hz, 4H, CH₂ × 2), 5.34 (s, 2H, OCH₂), 7.06 (d, J = 8.7 Hz, 1H, ArH), 7.34-7.47 (m, 2H, ArH), 7.84-7.93 (m, 3H, ArH), 8.29 (d, J =1.9 Hz, 1H, ArH), 13.14 (s, 1H, OH). ¹³C NMR (75 MHz, CDC¹³): δ 11.4, 48.7, 74.6, 117, 118.2, 121.7, 122.3, 126.1, 126.4, 127, 129.4, 132.4, 132.7,162.4, 168.4, 191.1. IR (KBr, cm⁻¹): 3448, 1694, 1624, 1503, 1220, 1099. ESI-MS⁺: [M + H]⁺, found 401.1 m/z C₁₉H₂₀N₄O₄S requires 400.1 HRMS m/z calcd

for $C_{19}H_{21}N_4O_4S [M + H]^+$: 401.1284. found 401.1289. ppm error 1.2.





The title compound **4** was prepared from the reaction of **12** with 2-aminobenzothiol by using the same method as compound **3**, as a white solid (80.4 mg, 62.3%). mp: 289-290 °C. ¹H NMR (300 MHz, CDCl₃) 3.42 (t, J = 4.8 Hz, 4H, NCH₂ × 2), 3.80 (t, J = 4.8 Hz, 4H, OCH₂ × 2), 5.35 (s, 2H, OCH₂), 7.16 (d, J = 8.7 Hz, 1H, ArH), 7.43-7.56 (m, 2H, ArH), 7.79-8.02 (m, 3H, ArH), 8.40 (d, J = 1.9 Hz, 1H, ArH). ¹³C NMR (75 MHz, CDCl₃): δ 51.5, 65.6, 74.8, 117.1, 118.3, 121.7, 122.3, 126.1, 126.4, 127, 129.5, 132.5, 132.7, 151.4, 162.6, 168.4, 191.2. IR (KBr, cm⁻¹): 3422, 1695, 1654, 1498, 1226, 1114, 1022. ESI-MS, found [M - H]⁻, 413.1. m/z C₁₉H₁₈N₄O₅S requires 414.1 HRMS m/z calcd for C₁₉H₁₇N₄O₅S [M - H]⁻, 413.0920. found 413.0926. ppm error 1.45.

3. MTT Assay

Human lung cancer A549 cells, human colon carcinoma HCT116 cells, HT29 cells, human acute monocytic leukemia THP-1 cells, human leukemia HL60 cells, human liver carcinoma HepG2 cells, human normal colon epithelial CCD-18Co cells were purchased from American Tissue Culture Collection (ATCC, Rockville, MD, USA). Human normal liver LO2 cells were purchased from Shanghai Institute of Cell Research. All cells were cultured in Roswell Park Memorial Institute medium (RPMI-1640, Thermo, USA) and Fetal bovine serum (FBS, GIBCO, Invitrogen Corporation, NY, USA) at 37 °C under 5% CO₂ atmosphere. 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl- 2-H-tetrazolium bromide (MTT) were obtained from Sheng Xing Biotechnology Company.

Intermittent light exposure protocol for Table 1 in MS. Cells were incubated in a 96 well plate with a concentration of 10⁴ cells/well and cultured in 37 °C 5% CO₂ for 24 h. Then cells were respectively treated with or without test compounds containing EMEM, and intermittently exposed under ultraviolet light (365-400 nm, 0.02 W/cm², UV lamp TLS3009-X105) or under visible light (\geq 410 nm, 0.05 W/cm², HL-2000) for 20 mins at 0h, 1h, 2h, 4h, 6h, 8h, 12h, 24h, 36h time points, or under dark. Each concentration was repeated 3 times in parallel. After incubation for 44 h, MTT (20 µL, 5 mg/mL) was added into each well, and the cells were incubated for additional 4h. Then the medium was carefully removed. Dimethyl sulfoxide (150 µL/well) was added and the 96 well plate was oscillated gently until crystal dissolved. The absorbance at 570 nm was measured using a microplate reader. The cell viability was expressed as a percentage of OD570 and the corresponding IC₅₀ values are summarized in Table 1 in MS. It was found that the cancer cell viability was not affected by intermittent light exposure (Table S1).

	Without 1	Without 2	Without 3	Without 4
A549	0.26 ± 0.06	3.85 ± 0.98	0.68 ± 1.55	1.48 ± 2.32
HCT116	0.13 ± 0.04	1.15 ± 0.26	-1.32 ± 1.60	-1.68 ± 2.73
THP-1	1.37 ± 0.33	1.24 ± 0.26	1.13 ± 4.20	2.71 ± 2.62
HL60	1.50 ± 0.23	1.82 ± 0.52	-2.04 ± 2.02	0.85 ± 1.14

Table S1. The proliferation inhibitory rates (%) of intermittent ultraviolet light exposure (without treatment of 1 or 2) or intermittent visible light exposure (without treatment of 3 or 4) to A549, HCT116, THP-1, and HL-60 cells

Continuous visible light exposure protocol. HL-60 cells were incubated in a 96 well plate with a concentration of 10^4 cells/well and cultured in 37 °C 5% CO₂ for 24 h. Then cells were treated with the compound **3** at different concentrations containing EMEM for 4h in the dark.

Then the cells were exposed by continuous visible light (\geq 410 nm, 0.05 W/cm², HL-2000) for 1h, and then kept in the dark for some hours. The total incubation time were 24, 48 or 72h. Each concentration was repeated 3 times in parallel.

Table S2. IC_{50} values (μ M) of **3** under intermittent or continuous visible light against HL-60 cells with different incubation time.

	The intermittent visible light		The continuous v	visible light
	Light	Dark	Light	Dark
24h	3.25 ± 0.15	679.1 ± 13.46	2.98 ± 0.17	706.5 ± 11.49
48h	0.64 ± 0.03	205 ± 2.6	0.59 ± 0.06	207.1 ± 3.47
72h	0.47 ± 0.05	131.7 ± 3.52	0.47 ± 0.08	136.0 ± 3.94

Table S3. IC_{50} values (μ M) of **3** under continuous visible light exposure or in dark against HT29, CCD-18Co, HepG2, and LO2 cells with total incubation time for 72h.

	HepG2	LO2	HT29	CCD-18Co
3 in the dark	202.3 ± 5.69	123.8 ± 7.64	163.5 ± 5.98	125 ± 6.61
3 under visible light	1.226 ± 0.051	124.2 ± 5.51	1.121 ± 0.09	95.38 ± 2.51

4. The NO release and photolytic decomposition kinetics studies of 1 and 3 in PBS

A solution of **1** (50 μ M) in PBS (50 mM, containing 5% DMSO) was exposed ultraviolet light (365-400 nm) for 360 mins (keeping the quartz cuvette 6 cm away from the light source, 0.02 W/cm²) or in dark at 37 °C. At 0, 5, 10, 30, 60, 120, 240, 360 min, 200 μ L of sample was take out. 150 μ L of sample was mixed with 50 μ L Griess reagent and incubated for addition 5 mins at 37 °C to determine the concentrations of NO₂⁻, while 50 μ L of sample was analyzed by RP-HPLC using Venusil MP C18 (250 mm × 4.6 mm × 5 μ m) at a flow rate of 1mL/min with a detection wavelength of 254 nm to determine the concentration of **1** remaining. The experiment was repeated three times and the results are shown in Figure 2A in the manuscript.

A solution of **3** (50 μ M) in PBS (50 mM, containing 5% DMSO) were exposed visible light (\geq 410 nm) for 30 mins (keeping the quartz cuvette 6 cm away from the light source, 0.05 W/cm²) or in dark at 37 °C. At 0, 5, 10, 15, 30 min, 200 μ L of sample was take out. 150 μ L of sample was mixed with 50 μ L Griess reagent and incubated for addition 5 mins at 37 °C to determine the concentrations of NO₂⁻, while 50 μ L of sample was analyzed by RP-HPLC using Venusil MP C18 (250 mm × 4.6 mm × 5 μ m) at a flow rate of 1mL/min with a detection wavelength of 254 nm to

determine the concentration of **3** remaining. The experiment was repeated three times and the results are shown in Figure 2B in the manuscript.

A solution of $1 (50 \ \mu\text{M})$ in PBS (50 mM, containing 5% DMSO) and a solution of $3 (50 \ \mu\text{M})$ in PBS (50 mM, containing 5% DMSO) were incubated in dark at 37 °C for 6h and 30 min, respectively. 50 μ L of samples were analyzed by HPLC to determine the stability of 1 and 3 in PBS in dark (Figure S1). It was found that compounds 1 and 3 were stable in dark in PBS solution.



Figure S1. (A) Overlay of HPLC chromatograms of **1** under the dark at 0 min, 6h. (B) Overlay of HPLC chromatograms of **3** under the dark at 0 min, 30 min.

5. NO release measurement in HL60 cells

DAF-FM DA (Beyotime) was used as a fluorescent indicator of intracellular NO release. HL-60 cells were treated with **1** (0.4 μ M) under intermittent ultraviolet light (365-400 nm, 0.02 W/cm²) exposure for 20 mins at 0, 1, 2, 4, 6, 8, 12, 24, 36h time points, or in the dark for total 48h. The cells were then collected and resuspended with DAF-FM DA (1.5 mL, 5 μ M) 37 °C for 20 mins in the dark. The cells were rinsed three times with PBS and then analyzed by fluorescence-activated cell sorting (FACS). NO production was measured with the flow cytometer with excitation and emission wavelengths of 495 and 515 nm, respectively. BD AccuriTM C6 flow cytometer was used, and 10,000 cells was gated for each sample. HL-60 cells were treated with **3** (0.6 μ M) under the same conditions as **1** except using visible light (\geq 410 nm, 0.05 W/cm²).



Figure S2. (A) and (B) HL-60 cells were treated with **1** (0.4 μ M) under intermittent ultraviolet light (365-400 nm, 0.02 W/cm²) exposure or in the dark for 48h. The cells were then collected and resuspended with DAF-FM DA (5 μ M) for 20 mins in the dark, and then analyzed by fluorescence-activated cell sorting (FACS). (C) and (D) HL-60 cells were treated with **3** (0.6 μ M) under the same conditions as (A and B) except using visible light (\geq 410 nm, 0.05 W/cm²). (E) Plot of applied gate in the flow cytometry.

6. The excitation and emission spectrum of compound 3

Compound **3** was dissolved in DMSO and diluted to 30 μ M under dark. The excitation (Ex) and emission (Em) spectra of **3** (30 μ M) in DMSO were recorded at 0 and 30 mins under visible light exposure (\geq 410 nm, 0.05 W/cm², HL-2000) by using Tecan Infinite® 200 Pro.



Figure S3. (A) The excitation spectra and (B) emission spectra of compound **3** (30 μ M in DMSO) under visible light exposure for 30 mins.

7. Confocal microscopy studies of HL60 cells internalization of 3

Real-time monitoring of NO release from **3** in HL60 cells under visible light (\geq 410 nm) irradiation was studied by a Zeiss LSM confocal scanning system. HL60 cells were treated with 0.3 µM of **3** under dark at 37 °C for 8h. The cells were washed twice with PBS, and incubated with DAF-FM-DA (5.0 µM) under dark for 20 min. After washing, compound **3**-treated cells were irradiated with visible light (\geq 410 nm, 0.05 W/cm², HL-2000, 20W) for 15 min. Druing the incubation, fluorescent images are taken every 5 minutes. For the compound **3**, excitation wavelength (Ex) was carried out with lasers at $\lambda = 546$ nm and emissions wavelength (Em) were recorded in 570 nm, respectively. For DAF-FM DA, Ex/Em: 495 nm/515 nm and for the decomposition product (benzothiazole-2-yl)-4-hydroxy phenylacetic acid, Ex/Em: 358 nm/461 nm. All images are normalized to have the same brightness to ensure that any changes in image brightness is due to biological/chemical phenomena. As to clearly distinguish the fluorescence colour changes, ZEN 2012 software, which comes with the microscope, was used to set Ex/Em: 546/570 nm in green, 358/461 nm in blue and 495/515 nm in red, respectively.



8. Absorbance spectra (200-500 nm) of 3 under visible light (\ge 410 nm) for 30 mins

Figure S4. The absorbance spectra (200-500 nm) of **3** (200 μ M) in PBS (10 % DMSO) at intervals of 2 mins under visible light (\geq 410 nm) for 30 mins.

9. NMR Spectra Scanning files





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8. FTIR of compounds 1-4,11,12





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11. HPLC assessment of compound purity

All tested compounds 1 - 4with a purity of > 97% were used for subsequent biological assays. We provided the spectra of HPLC assays as below.

Column: Venusil MP C18 ($250 \text{ mm} \times 4.6 \text{ mm} \times 5 \text{ um}$);

Mobile phase: The compound 1-2: Methanol-Water (50 to 50, v/v); 3-4: Acetonitrile-Water (65 to 35, v/v).

Wavelength: 254 nm;

Rate: 1 mL/min;

Temperature: 25 °C;

1,97.83%



2, 99.31%







12. Reference

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