

Supporting Information

Visible Light-Induced Selective Aerobic Oxidative Transposition of Vinyl Halides Using a Tetrahalogenoferrate (III) Complex Catalyst

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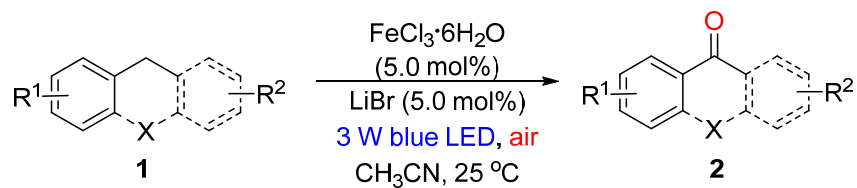
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1. General information

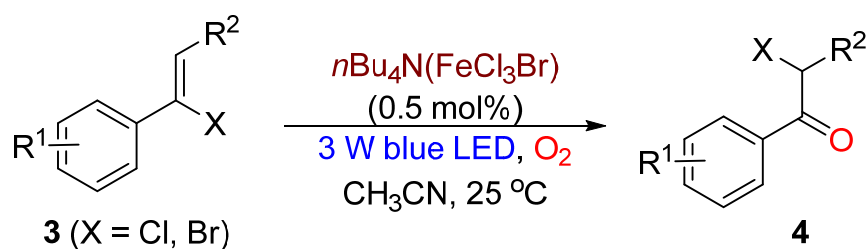
Column chromatography was carried out with *silica gel* 60 (particle size 0.040–0.063 mm, 230–400 mesh) and commercially available solvents. Thin-layer chromatography (TLC) was conducted on aluminum sheets coated with *silica gel* 60 F254 with visualization by a UV lamp (254 or 360 nm).

^1H and ^{13}C NMR spectra were recorded at 300 and 75 MHz at 25 °C with a 300 MHz instrument. Chemical shifts are reported in parts per million (ppm), using the residual solvent signal as an internal standard: CDCl_3 (^1H NMR: δ 7.26, singlet; ^{13}C NMR: δ 77.0, triplet). Multiplicities were given as: *s* (singlet), *d* (doublet), *t* (triplet), *q* (quartet), *quintet*, *m* (multiplets), *dd* (doublet of doublets), *dt* (doublet of triplets), and *br* (broad). Coupling constants (*J*) were recorded in Hertz (Hz). The number of proton atoms (*n*) for a given resonance was indicated by *nH*. The number of carbon atoms (*n*) for a given resonance was indicated by *nC*. HRMS was reported in units of mass of charge ratio (*m/z*). Mass samples were dissolved in DCM and MeOH (HPLC grade) unless otherwise stated. Electrochemical measurements were carried out by cyclic voltammetry (CV). The cyclic voltammetry was performed with an Autolab potentiostat by Echochemie under nitrogen atmosphere in a one-compartment electrolysis cell consisting of a platinum wire working electrode, a platinum wire counter electrode, and a quasi Ag/AgCl reference electrode. Cyclic voltammograms were monitored at scan rates of either 100 $\text{mV}\cdot\text{s}^{-1}$ or 50 $\text{mV}\cdot\text{s}^{-1}$ and recorded in distilled acetonitrile. The concentration of the complex was maintained at 0.5 mM or less and each solution contained 0.1 M of tetrabutylammonium hexafluorophosphate (TBAP) as the electrolyte.

2. General experimental procedure for the synthesis of 2

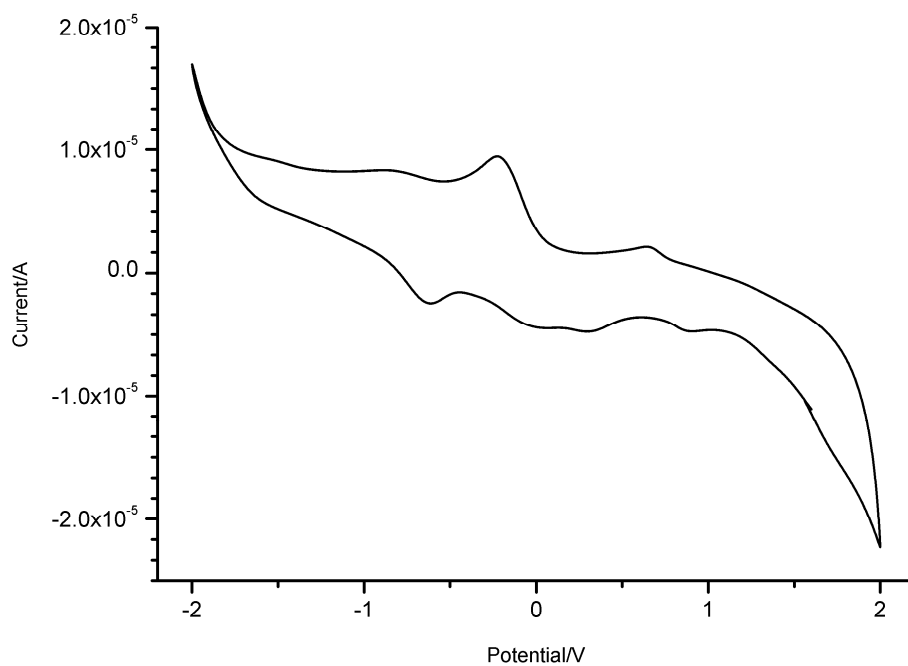


To a stirred solution of $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ (2.7 mg, 0.01 mmol, 0.05 equiv) and LiBr (0.87 mg, 0.01 mmol, 0.05 equiv) in 2.0 mL of MeCN (HPLC grade) was added compound **1** (0.2 mmol, 1.0 equiv). Then the reaction mixture was stirred at 25 °C under an ambient atmosphere (the temperature was maintained in an incubator) and irradiated by a 3 W blue LED ($\lambda = 450\text{--}455$ nm). The reaction was monitored by TLC or GC analysis. Upon complete consumption of **1**, the solvent was removed *in vacuo*. The reaction mixture was purified by flash chromatography on *silica gel*, followed by gradient elution with petroleum ether to petroleum ether/ethyl acetate (100/1 ratio), then concentrated the solvent *in vacuo* to afford products **2a-r**.

3. General experimental procedure for the synthesis of 4

FeCl₃·6H₂O (0.27 mg, 0.001 mmol, 0.005 equiv) and TBAB (0.32 mg, 0.001 mmol, 0.005 equiv) were added in 2.0 mL of MeCN (HPLC grade) and the reaction mixture was stirred for 10 minutes at 25 °C (the temperature was maintained in an incubator) under an ambient atmosphere with an oxygen balloon. To the stirred solution, compound **3** was then added and irradiated by a 3 W blue LED ($\lambda = 450\text{--}455$ nm). The reaction was monitored by TLC or GC analysis. Upon complete consumption of **3**, the solvent was removed *in vacuo*. The reaction mixture was purified by flash chromatography on *silica gel*, followed by gradient elution with petroleum ether to petroleum ether/ethyl acetate (100/1 ratio), then concentrated the solvent *in vacuo* to afford products **4a-v** and **5**.

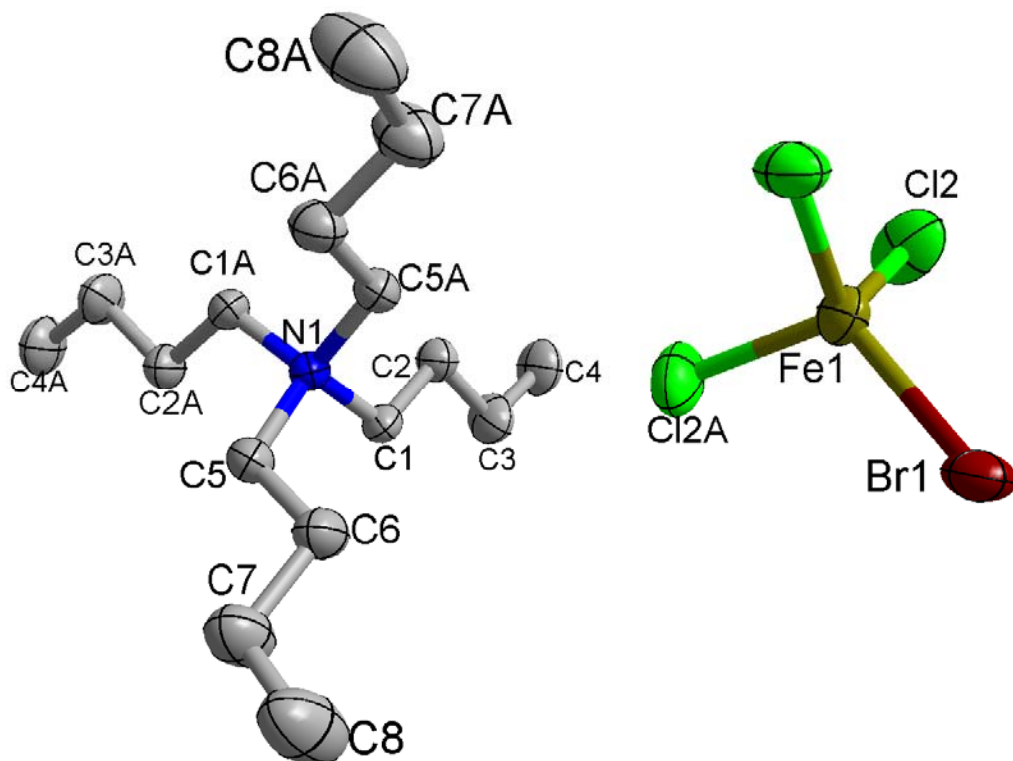
4. The measurement of redox potentials TBA(FeCl₃Br)



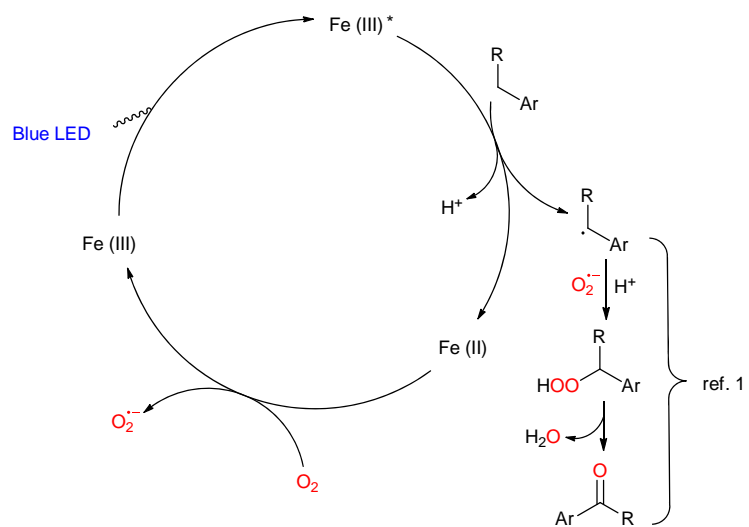
$E_{1/2(\text{red1})} = -0.295 \text{ V}$, $E_{1/2(\text{ox1})} = -0.642 \text{ V}$. $E_{1/2(\text{ox1})}/E_{1/2(\text{red1})}$ are half-wave potentials of the first oxidation/reduction measured in CH₃CN versus SCE

5. X-Ray crystallography of TBA(FeCl₃Br)

TBA(FeCl₃Br): CCDC 1525407

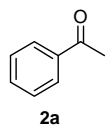


Displacement ellipsoids are drawn at the 30% probability level. (Solvent: ethyl acetate)

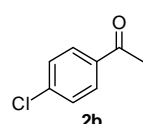
6. Proposed mechanism for oxygenation of benzylic sp^3 C-H

1. H. Yi, C. Bian, X. Hu, L. Niu, A. Lei, *Chem. Commun.*, 2015, **51**, 14046.

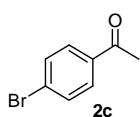
7. Characterization data



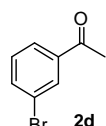
Colorless oil; 93% yield (from **1a**, 22.3 mg); 92% yield (from **1s**, 22.1 mg) ^1H NMR (300 MHz, CDCl_3) δ 8.04–7.88 (m, 2H), 7.54 (ddd, $J = 6.5, 3.8, 1.2$ Hz, 1H), 7.43 (dd, $J = 10.3, 4.6$ Hz, 2H), 2.58 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 198.1, 136.9, 133.0, 128.5, 128.2, 26.5.



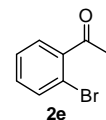
Colorless oil; 71% yield (21.9 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.94–7.79 (m, 2H), 7.48–7.32 (m, 2H), 2.57 (d, $J = 1.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.8, 139.4, 135.3, 129.6, 128.8, 26.5.



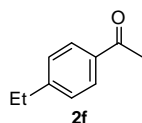
White solid; 67% yield (26.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.87–7.71 (m, 2H), 7.63–7.49 (m, 2H), 2.55 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.9, 135.7, 131.8, 129.7, 128.2, 26.4.



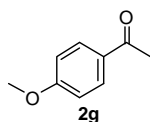
Colorless oil; 78% yield (31.0 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.07 (t, $J = 1.7$ Hz, 1H), 7.86 (d, $J = 7.8$ Hz, 1H), 7.72–7.64 (m, 1H), 7.34 (t, $J = 7.9$ Hz, 1H), 2.58 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.6, 138.7, 135.9, 131.3, 130.2, 126.8, 122.9, 26.6.



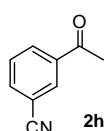
Colorless oil; 72% yield (28.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.60 (d, $J = 7.8$ Hz, 1H), 7.46 (d, $J = 7.4$ Hz, 1H), 7.36 (t, $J = 7.4$ Hz, 1H), 7.33–7.27 (m, 1H), 2.62 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 201.2, 141.2, 133.7, 131.7, 128.8, 127.3, 118.7, 30.2.



Colorless oil; 68% yield (20.2 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.94 (d, $J = 8.0$ Hz, 2H), 7.34 (d, $J = 7.7$ Hz, 2H), 2.76 (q, $J = 7.6$ Hz, 2H), 2.63 (s, 3H), 1.31 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 197.9, 150.1, 134.9, 128.5, 128.1, 28.9, 26.6, 15.2.

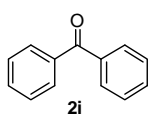


Pale yellow oil; 74% yield (22.2 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.94 (d, $J = 7.6$ Hz, 2H), 6.93 (d, $J = 7.8$ Hz, 2H), 3.87 (s, 3H), 2.56 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 196.8, 163.5, 130.6, 130.3, 113.7, 55.5, 26.3.

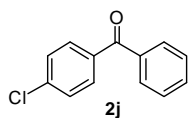


Colorless oil; 35% yield (10.2 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.22 (s, 1H), 8.17 (d, $J = 7.9$ Hz, 1H), 7.83 (d, $J = 7.7$ Hz, 1H), 7.61 (t, $J = 7.8$ Hz, 1H), 2.63 (s, 3H); ^{13}C NMR

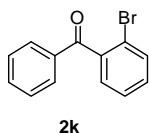
(75 MHz, CDCl₃) δ 195.8, 137.6, 135.9, 132.2, 131.9, 129.6, 117.9, 113.0, 26.5.



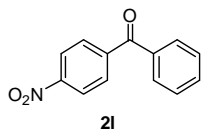
White solid; 76% yield (27.7 mg); ¹H NMR (300 MHz, CDCl₃) δ 7.91–7.73 (m, 4H), 7.64–7.53 (m, 2H), 7.47 (t, *J* = 7.6 Hz, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 196.5, 137.4, 132.3, 129.9, 128.1.



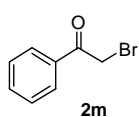
White solid; 79% yield (34.2 mg); ¹H NMR (300 MHz, CDCl₃) δ 7.87–7.70 (m, 4H), 7.60 (t, *J* = 7.2 Hz, 1H), 7.48 (dd, *J* = 12.1, 7.9 Hz, 4H); ¹³C NMR (75 MHz, CDCl₃) δ 195.5, 138.9, 137.2, 135.8, 132.6, 131.4, 129.9, 128.6, 128.4.



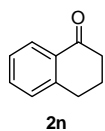
White solid; 90% yield (47.0 mg); ¹H NMR (300 MHz, CDCl₃) δ 7.85–7.78 (m, 2H), 7.66 (dd, *J* = 7.1, 2.2 Hz, 1H), 7.62–7.56 (m, 1H), 7.47 (dd, *J* = 10.6, 4.6 Hz, 2H), 7.41 (dd, *J* = 6.6, 1.9 Hz, 1H), 7.36 (td, *J* = 7.9, 1.9 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 195.9, 140.6, 136.1, 133.7, 133.2, 131.1, 130.2, 129.0, 128.6, 127.2, 119.5.



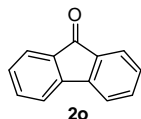
White solid; 54% yield (24.5mg); ¹H NMR (300 MHz, CDCl₃) δ 8.40 – 8.28 (m, 2H), 7.99 – 7.89 (m, 2H), 7.80 (dd, *J* = 5.2, 3.3 Hz, 2H), 7.66 (ddd, *J* = 7.4, 4.0, 1.2 Hz, 1H), 7.52 (dd, *J* = 10.5, 4.6 Hz, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 194.8, 149.8, 142.8, 136.2, 133.5, 130.7, 130.1, 128.7, 123.5.



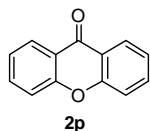
White solid; 81% yield (from **1m**, 32.2 mg); 83% yield (from **3a**, 33.0 mg) ¹H NMR (300 MHz, CDCl₃) δ 8.02–7.94 (m, 2H), 7.65–7.56 (m, 1H), 7.48 (dd, *J* = 10.4, 4.7 Hz, 2H), 4.46 (s, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 191.2, 133.9, 128.9, 128.8, 30.9.



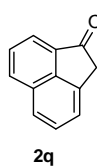
Colorless oil; 72% yield (21.0 mg); ¹H NMR (300 MHz, CDCl₃) δ 8.03 (d, *J* = 7.8 Hz, 1H), 7.46 (t, *J* = 7.4 Hz, 1H), 7.28 (dd, *J* = 18.9, 7.9 Hz, 2H), 2.96 (t, *J* = 6.0 Hz, 2H), 2.65 (t, *J* = 6.5 Hz, 2H), 2.28–1.98 (m, 2H); ¹³C NMR (75 MHz, CDCl₃) δ 198.4, 144.5, 133.4, 132.6, 128.8, 127.2, 126.6, 39.2, 29.7, 23.3.



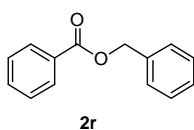
White solid; 88% yield (31.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.65 (d, $J = 7.3$ Hz, 2H), 7.54–7.43 (m, 4H), 7.33–7.25 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.9, 144.4, 134.7, 134.1, 129.0, 124.3, 120.3.



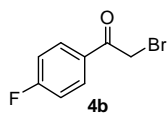
White solid; 99% yield (37.6 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.34 (d, $J = 7.9$ Hz, 2H), 7.73 (t, $J = 7.7$ Hz, 2H), 7.49 (d, $J = 8.4$ Hz, 2H), 7.38 (t, $J = 7.5$ Hz, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 177.2, 156.1, 134.8, 126.7, 123.9, 121.7, 117.9.



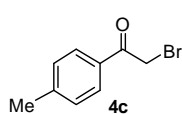
White solid; 52% yield (17.5 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.09 (d, $J = 8.1$ Hz, 1H), 7.96 (d, $J = 7.0$ Hz, 1H), 7.82 (d, $J = 8.3$ Hz, 1H), 7.71 (t, $J = 7.5$ Hz, 1H), 7.60 (t, $J = 7.6$ Hz, 1H), 7.46 (d, $J = 6.8$ Hz, 1H), 3.82 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 203.0, 142.9, 134.9, 134.6, 131.5, 130.9, 128.3, 128.0, 123.9, 121.4, 121.0, 42.0.



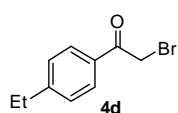
White solid; 86% yield (36.5 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.10 (d, $J = 7.4$ Hz, 2H), 7.57 (t, $J = 6.9$ Hz, 1H), 7.42 (dd, $J = 12.9, 8.8$ Hz, 7H), 5.39 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 166.4, 136.0, 133.0, 130.0, 129.7, 128.6, 128.3, 128.2, 128.1, 66.6.



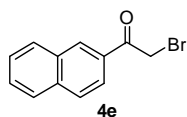
White solid; 82% yield (35.6 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.08–7.97 (m, 2H), 7.22–7.11 (m, 2H), 4.41 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 189.8, 167.8, 164.4, 131.8, 131.7, 130.3, 130.2, 116.2, 116.0, 30.4.



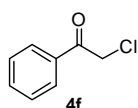
White solid; 83% yield (35.3 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.89 (d, $J = 8.3$ Hz, 2H), 7.29 (d, $J = 8.1$ Hz, 2H), 4.43 (s, 2H), 2.43 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 191.0, 145.0, 131.4, 129.5, 129.0, 30.9, 21.8.



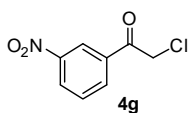
White solid; 86% yield (39.1 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.91 (d, $J = 8.3$ Hz, 2H), 7.32 (d, $J = 8.2$ Hz, 2H), 4.44 (s, 2H), 2.72 (q, $J = 7.6$ Hz, 2H), 1.27 (t, $J = 7.6$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 191.0, 151.1, 131.6, 129.2, 128.4, 31.0, 29.0, 15.1.



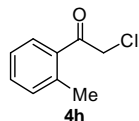
White solid; 76% yield (37.9 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.50 (s, 1H), 7.99 (dd, $J = 14.5, 8.3$ Hz, 2H), 7.94–7.84 (m, 2H), 7.70–7.49 (m, 2H), 4.58 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 191.2, 135.8, 132.3, 131.2, 130.9, 129.6, 129.0, 128.8, 127.8, 127.0, 124.1, 31.0.



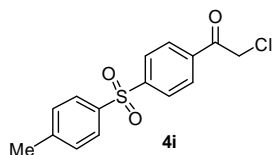
White solid; 80% yield (24.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.95 (d, $J = 7.3$ Hz, 2H), 7.62 (t, $J = 7.4$ Hz, 1H), 7.49 (t, $J = 7.6$ Hz, 2H), 4.72 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 191.0, 134.2, 134.0, 128.9, 128.5, 46.0.



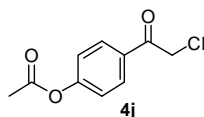
White solid; 72% yield (28.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.79 (t, $J = 1.9$ Hz, 1H), 8.48 (ddd, $J = 8.2, 2.1, 0.9$ Hz, 1H), 8.39–8.24 (m, 1H), 7.74 (t, $J = 8.0$ Hz, 1H), 4.73 (s, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 189.3, 148.5, 135.3, 134.1, 130.3, 128.2, 123.5, 45.4.



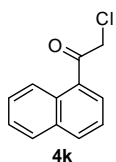
White solid; 73% yield (24.6 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.62 (d, $J = 7.4$ Hz, 1H), 7.44 (t, $J = 7.0$ Hz, 1H), 7.30 (d, $J = 7.4$ Hz, 2H), 4.64 (s, 2H), 2.53 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 194.2, 139.5, 134.5, 132.4, 128.6, 125.8, 47.9, 21.4.



Colorless oil; 54% yield (36.5 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.91 (d, $J = 8.3$ Hz, 2H), 7.71 (d, $J = 7.9$ Hz, 2H), 7.33 (d, $J = 7.9$ Hz, 2H), 7.13 (d, $J = 8.4$ Hz, 2H), 4.65 (s, 2H), 2.46 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 189.8, 153.6, 146.0, 132.8, 132.0, 130.4, 130.0, 128.5, 122.9, 45.7, 21.8.

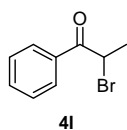


White solid; 87% yield (37.0 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.00 (d, $J = 7.8$ Hz, 2H), 7.23 (d, $J = 7.8$ Hz, 2H), 4.69 (s, 2H), 2.33 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 189.9, 168.7, 154.9, 131.7, 130.2, 122.1, 45.8, 21.1.

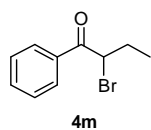


White solid; 74% yield (30.3 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.63 (d, $J = 8.5$ Hz, 1H), 8.05 (d, $J = 8.3$ Hz, 1H), 7.89 (t, $J = 7.4$ Hz, 2H), 7.69–7.47 (m, 3H), 4.78 (s, 2H);

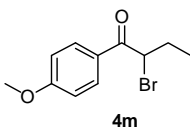
^{13}C NMR (75 MHz, CDCl_3) δ 194.4, 134.0, 133.9, 132.4, 130.4, 128.6, 128.5, 128.2, 126.8, 125.5, 124.2, 48.0.



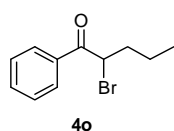
Pale yellow oil; 74% yield (36.2 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.03 (d, $J = 7.6$ Hz, 2H), 7.59 (t, $J = 6.9$ Hz, 1H), 7.49 (t, $J = 7.2$ Hz, 2H), 5.45–5.18 (m, 1H), 1.91 (d, $J = 6.5$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.3, 134.0, 133.6, 128.9, 128.7, 41.4, 20.1



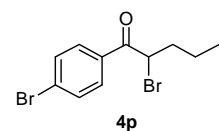
Pale yellow oil; 85% yield (38.6 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.02 (d, $J = 7.2$ Hz, 2H), 7.58 (d, $J = 7.2$ Hz, 1H), 7.49 (t, $J = 7.0$ Hz, 2H), 5.08 (dd, $J = 9.3, 4.3$ Hz, 1H), 2.36–2.05 (m, 2H), 1.09 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.2, 134.5, 133.6, 128.8, 128.7, 49.0, 26.9, 12.1.



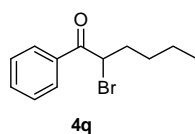
White solid; 81% yield (41.6 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.00 (d, $J = 7.5$ Hz, 2H), 6.95 (d, $J = 8.4$ Hz, 2H), 5.04 (t, $J = 6.8$ Hz, 1H), 3.87 (s, 3H), 2.33–1.99 (m, 2H), 1.07 (t, $J = 7.1$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 191.8, 163.9, 131.2, 127.2, 113.9, 55.5, 49.0, 27.0, 12.2.



Pale yellow oil; 90% yield (43.2 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.02 (d, $J = 7.5$ Hz, 2H), 7.58 (d, $J = 7.6$ Hz, 1H), 7.49 (t, $J = 7.2$ Hz, 2H), 5.16 (t, $J = 6.9$ Hz, 1H), 2.25–2.04 (m, 2H), 1.69–1.34 (m, 2H), 0.98 (t, $J = 7.2$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.3, 134.4, 133.6, 128.8, 128.7, 47.0, 35.4, 20.7, 13.6.

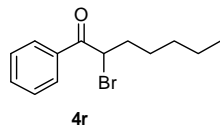


Pale yellow oil; 86% yield (49.0 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.87 (d, $J = 7.9$ Hz, 2H), 7.62 (d, $J = 8.1$ Hz, 2H), 5.07 (t, $J = 7.0$ Hz, 1H), 2.23–2.01 (m, 2H), 1.49 (m, 2H), 0.98 (t, $J = 7.3$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.2, 133.1, 132.1, 130.3, 128.9, 46.8, 35.2, 20.7, 13.6.

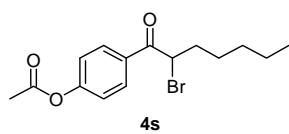


Pale yellow oil; 88% yield (45.0 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.02 (d, $J = 7.6$ Hz, 2H), 7.60 (t, $J = 7.1$ Hz, 1H), 7.49 (t, $J = 7.3$ Hz, 2H), 5.14 (t, $J = 6.8$ Hz,

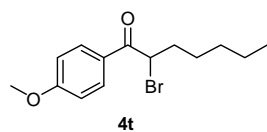
1H), 2.29 – 2.05 (m, 2H), 1.56 – 1.30 (m, 4H), 0.93 (d, $J = 5.7$ Hz, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.3, 134.4, 133.6, 128.8, 128.7, 47.2, 33.2, 29.6, 22.2, 13.9.



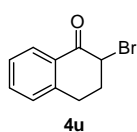
Pale yellow oil; 79% yield (42.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.02 (d, $J = 7.6$ Hz, 2H), 7.60 (t, $J = 7.1$ Hz, 1H), 7.49 (t, $J = 7.4$ Hz, 2H), 5.14 (t, $J = 7.0$ Hz, 1H), 2.29–2.01 (m, 2H), 1.60–1.29 (m, 6H), 0.89 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 193.3, 134.4, 133.6, 128.8, 128.7, 47.2, 33.4, 31.2, 27.2, 22.4, 13.9.



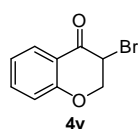
Pale yellow oil; 82% yield (42.7 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.05 (d, $J = 8.3$ Hz, 2H), 7.21 (d, $J = 8.3$ Hz, 2H), 5.09 (t, $J = 7.1$ Hz, 1H), 2.32 (s, 3H), 2.25–2.04 (m, 2H), 1.55–1.39 (m, 2H), 1.34 (s, 4H), 0.89 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 192.0, 168.7, 154.6, 132.0, 130.5, 121.9, 47.1, 33.4, 31.2, 27.1, 22.4, 21.1, 13.9.



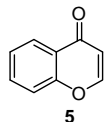
Pale yellow oil; 86% yield (51.5 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.00 (d, $J = 7.6$ Hz, 2H), 6.95 (d, $J = 8.4$ Hz, 2H), 5.10 (t, $J = 7.0$ Hz, 1H), 3.87 (s, 3H), 2.12 (tt, $J = 22.1, 10.9$ Hz, 2H), 1.47 (dd, $J = 18.7, 10.0$ Hz, 2H), 1.33 (s, 4H), 0.88 (s, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 191.9, 163.9, 131.2, 127.3, 113.9, 55.5, 47.2, 33.6, 31.3, 27.2, 22.4, 13.9.



Pale yellow oil; 58% yield (26.3 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.06 (d, $J = 7.8$ Hz, 1H), 7.49 (t, $J = 7.4$ Hz, 1H), 7.32 (t, $J = 7.5$ Hz, 1H), 7.25 (d, $J = 8.6$ Hz, 1H), 4.70 (s, 1H), 3.41–3.19 (m, 1H), 2.89 (d, $J = 17.1$ Hz, 1H), 2.63–2.22 (m, 2H); ^{13}C NMR (75 MHz, CDCl_3) δ 190.5, 143.0, 134.1, 129.9, 128.7, 128.6, 127.1, 50.4, 31.9, 26.1.



Pale yellow oil; 33% yield (15.0 mg); ^1H NMR (300 MHz, CDCl_3) δ 7.94 (d, $J = 7.9$ Hz, 1H), 7.54 (t, $J = 7.7$ Hz, 1H), 7.07 (dd, $J = 19.8, 8.1$ Hz, 2H), 4.78 – 4.38 (m, 3H); ^{13}C NMR (75 MHz, CDCl_3) δ 185.2, 160.6, 136.7, 128.2, 122.3, 118.7, 117.9, 71.2, 45.3.



White solid; 45% yield (13.2 mg); ^1H NMR (300 MHz, CDCl_3) δ 8.19 (d, $J = 7.9$ Hz, 1H), 7.84 (d, $J = 5.7$ Hz, 1H), 7.66 (t, $J = 7.7$ Hz, 1H), 7.41 (dd, $J = 18.0, 8.8$ Hz, 2H), 6.33 (d, $J = 5.6$ Hz, 1H); ^{13}C NMR (75 MHz, CDCl_3) δ 177.6, 156.4, 155.3, 133.7, 125.7, 125.2,

124.8, 118.1, 112.9.

8. NMR spectra

