

Visible-Light Promoted Arene C-H/C-X Lactonization via Carboxylic Radical Aromatic Substitution

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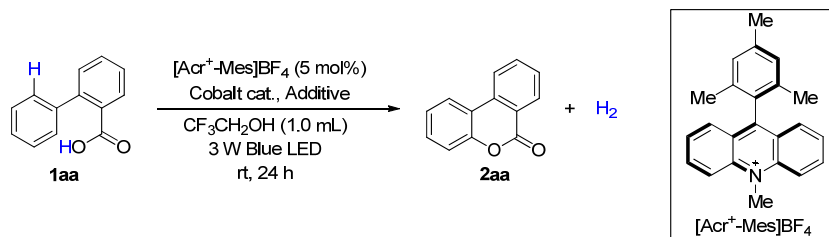
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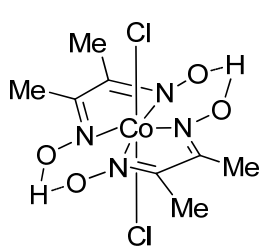
General information: Commercial reagents were used as received, unless otherwise indicated. Carboxylic acid **1**¹⁻³, especially substituted phenylacrylic acids (*Z/E*)⁴, cobalt catalysts⁵⁻⁸ and [Acr⁺-Mes]⁹ were prepared according to literature precedent. Nuclear magnetic resonance (NMR) spectra were recorded using Bruker AV-400 and AV-500 spectrometers (400 MHz and 500 MHz for ¹H NMR, 100 MHz and 125 MHz for ¹³C NMR, 377 MHz for ¹⁹F NMR). Tetramethylsilane (TMS) served as the internal standard for ¹H NMR, CDCl₃ and CD₃CN served as the internal standard for ¹³C NMR and ¹⁹F NMR. The following abbreviations were used to express the multiplicities: s = singlet; d = doublet; t = triplet; q = quartet; m = multiplet; br = broad. HRMS was recorded on a commercial instrument (ESI Source).

Reaction Condition Optimization

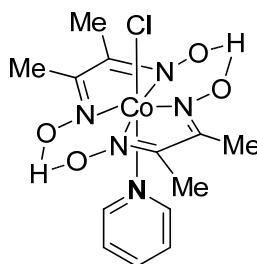
Table S1. Reaction condition optimization.



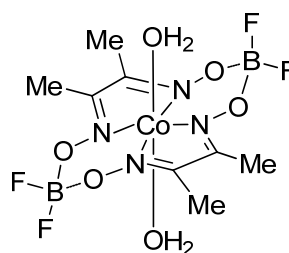
Entry	Cobalt catalyst	Additive	H ₂ ¹	Yield ²
1	NO	NO	trace	22%
2	Co(dmgh) ₂ ClPy (3 mol %) ³	NO	31%	60%
3	Co(dmghBF ₂) ₂ (H ₂ O) ₂ (3 mol %) ³	NO	33%	50%
4	Co(dmgh) ₂ Cl ₂ (3 mol %) ³	NO	28%	49%
5	Co(dmgh) ₂ ClPy (3 mol %)	2,6-Lutidine (100 mol %)	99%	99%
6	Co(dmgh) ₂ ClPy (3 mol %)	2,6-Lutidine (20 mol %)	99%	99%
7	Co(dmgh) ₂ ClPy (3 mol %)	Cs ₂ CO ₃ (20 mol %)	76%	83%
8	Co(dmgh) ₂ ClPy (3 mol %)	DBU (20 mol %)	90%	95%
9	Co(dmgh) ₂ ClPy (1 mol %) +[Acr ⁺ -Mes]BF ₄ (2 mol %)	2,6-Lutidine (20 mol %)	67%	59%



[Co(dmgh)₂Cl₂]



[Co(dmgh)₂ClPy]



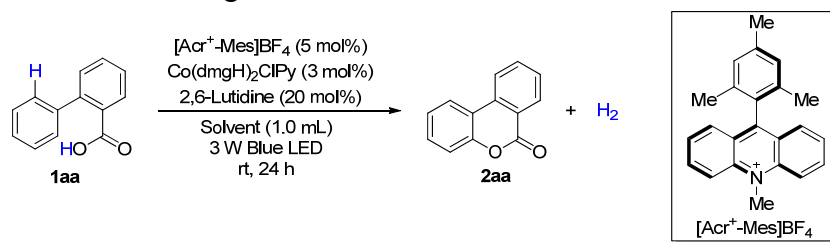
[Co(dmghBF₂)₂(H₂O)₂]

¹Determined by gas chromatography using methane as an internal standard.

²Determined by ¹H NMR analysis using 1, 3, 5-trimethoxybenzene as an internal standard.

³CH₃CN served as the solvent.

Table S2. Solvent screening.



Entry	Solvent	H ₂ ²	Yield ³
1	CH ₃ CN	85%	78%
2	CH ₃ CN:H ₂ O=1:1	Trace	Trace
3	DCM	Trace	35%
4	DCE	Trace	44%
5	CF ₃ CH ₂ OH	99%	99%
6	THF	Trace	23%
7	DMF	Trace	Trace
8	Et ₂ O	Trace	31%
9	EtOAc	Trace	22%
10	Acetone	Trace	22%
11	Toluene	Trace	35%

¹Determined by gas chromatography using methane as an internal standard.

²Determined by ¹H NMR analysis using 1, 3, 5-trimethoxybenzene as an internal standard.

General procedure for visible-light promoted arene C-H/C-X lactonization: A 10 mL Pyrex tube equipped with a rubber septum and magnetic stir bar was charged with carboxylic acid **1** (0.1 mmol), [Acr⁺-Mes]BF₄ (5 mol %), Co(dmgh)₂ClPy (3 mol %) and 2,6-Lutidine (20 mol %) followed by 1.0 mL CF₃CH₂OH. The mixture was bubbled with a stream of argon for 15 min. The sample was then irradiated under 3 W blue LED at specified temperature, until completion as indicated by TLC. The mixture was directly loaded onto silica gel column and eluted with ethyl acetate/petroleum ether to give the target product.

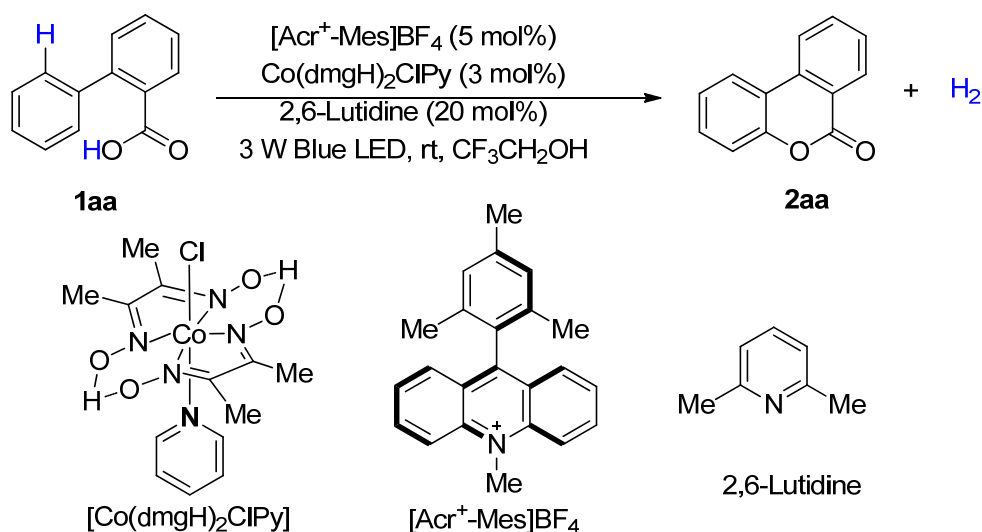
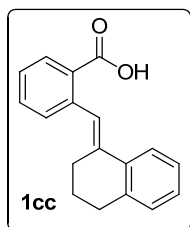


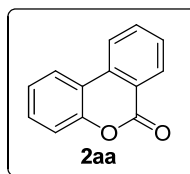
Figure S1. The reaction set-up (commercially available 3 W blue LEDs).



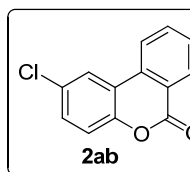
Figure S2. The reaction set-up under sunlight irradiation.



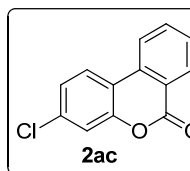
1cc: ^1H NMR (500 MHz, CDCl_3) δ 8.10 (d, J = 7.1 Hz, 1H), 7.84-7.68 (m, 1H), 7.54 (td, J = 7.6, 1.4 Hz, 1H), 7.49 (s, 1H), 7.36 (d, J = 7.5 Hz, 2H), 7.22-7.17 (m, 2H), 7.16-7.07 (m, 1H), 2.87 (t, J = 6.3 Hz, 2H), 2.61-2.48 (m, 2H), 1.84 (p, J = 6.3 Hz, 2H). ^{13}C NMR (125 MHz, CDCl_3) δ 171.9, 140.7, 137.8, 136.9, 136.2, 132.5, 131.6, 131.4, 129.3, 128.6, 127.5, 126.8, 126.3, 124.8, 123.1, 30.5, 28.0, 23.8. IR (KBr, cm^{-1}): 3564, 1669, 1649, 1481, 1453, 1404, 1303, 1259, 763, 719, 661. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{15}\text{O}_2^-$: 263.1078, found 263.1078.



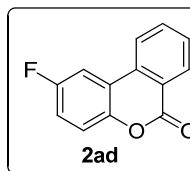
2aa: Prepared according to the general procedure outlined above and obtained as white solid (95% yield, 99% H_2). Analytical data were in agreement with literature values¹⁰: ^1H NMR (500 MHz, CDCl_3) δ 8.37 (dd, J = 7.9, 0.7 Hz, 1H), 8.08 (d, J = 8.1 Hz, 1H), 8.02 (dd, J = 7.9, 1.0 Hz, 1H), 7.84-7.77 (m, 1H), 7.56 (t, J = 7.6 Hz, 1H), 7.49-7.43 (m, 1H), 7.37-7.29 (m, 2H). ^{13}C NMR (125 MHz, CDCl_3) δ 161.2, 151.3, 134.9, 134.8, 130.6, 130.5, 128.9, 124.6, 122.8, 121.7, 121.3, 118.1, 117.8.



2ab: Prepared according to the general procedure outlined above and obtained as white solid (78% yield, p:o = 7:1, 86% H_2). Analytical data were in agreement with literature values¹⁰: ^1H NMR (500 MHz, CDCl_3) δ 8.39 (dd, J = 7.9, 0.8 Hz, 1H), 8.03 (d, J = 8.1 Hz, 1H), 7.98 (d, J = 2.3 Hz, 1H), 7.84 (dt, 1H), 7.65-7.60 (m, 1H), 7.41 (dd, J = 8.8, 2.4 Hz, 1H), 7.29 (d, J = 8.8 Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 160.7, 149.8, 135.2, 133.7, 130.8, 130.5, 130.2, 129.7, 122.7, 121.9, 121.3, 119.5, 119.3.

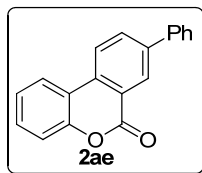


2ac: Prepared according to the general procedure outlined above and obtained as white solid (92% yield, 99% H_2). Analytical data were in agreement with literature values¹⁰: ^1H NMR (500 MHz, CDCl_3) δ 8.38 (dd, J = 7.9, 0.6 Hz, 1H), 8.06 (d, J = 8.1 Hz, 1H), 7.97 (d, J = 8.5 Hz, 1H), 7.87-7.80 (m, 1H), 7.60 (t, J = 7.6 Hz, 1H), 7.36 (d, J = 2.0 Hz, 1H), 7.31 (dd, J = 8.5, 2.0 Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 160.7, 151.6, 136.1, 135.2, 134.1, 130.9, 129.3, 125.2, 123.9, 121.8, 121.0, 118.1, 116.8.

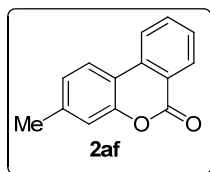


2ad: Prepared according to the general procedure outlined above and obtained

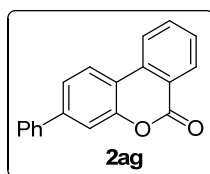
as white solid (88% yield, p:o = 11:1, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.40 (dd, *J* = 7.9, 0.8 Hz, 1H), 8.02 (d, *J* = 8.0 Hz, 1H), 7.89-7.82 (m, 1H), 7.70 (dd, *J* = 9.1, 2.9 Hz, 1H), 7.63 (t, *J* = 7.6 Hz, 1H), 7.34 (dd, *J* = 9.0, 4.7 Hz, 1H), 7.19 (ddd, *J* = 9.0, 7.8, 2.9 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 160.9, 160.4, 158.4, 147.5, 147.5, 135.1, 134.0, 134.0, 130.8, 129.7, 122.0, 121.3, 119.4, 119.4, 119.3, 117.9, 117.7, 109.0, 108.8.



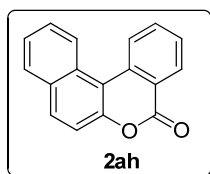
2ae: Prepared according to the general procedure outlined above and obtained as white solid (89% yield, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.57 (d, *J* = 1.8 Hz, 1H), 8.11 (d, *J* = 8.3 Hz, 1H), 8.04-7.98 (m, 2H), 7.65 (d, *J* = 7.4 Hz, 2H), 7.51-7.37 (m, 4H), 7.31 (t, *J* = 8.6 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 161.3, 151.2, 141.7, 138.9, 133.5, 133.5, 130.4, 129.2, 128.5, 128.4, 127.1, 124.7, 122.8, 122.4, 121.6, 117.9, 117.8.



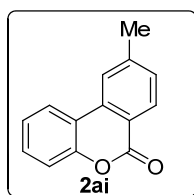
2af: Prepared according to the general procedure outlined above and obtained as white solid (98% yield, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.33 (d, *J* = 7.9 Hz, 1H), 8.01 (d, *J* = 8.1 Hz, 1H), 7.86 (d, *J* = 8.5 Hz, 1H), 7.78-7.74 (m, 1H), 7.51 (t, *J* = 7.6 Hz, 1H), 7.10 (d, *J* = 4.3 Hz, 2H), 2.41 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.4, 151.3, 141.3, 135.0, 134.8, 130.5, 128.4, 125.7, 122.5, 121.5, 120.9, 117.9, 115.4, 21.5.



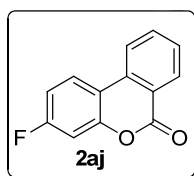
2ag: Prepared according to the general procedure outlined above and obtained as white solid (95% yield, 99% H₂). Analytical data were in agreement with literature values¹¹: ¹H NMR (500 MHz, CDCl₃) δ 8.37 (d, *J* = 7.9 Hz, 1H), 8.06 (dd, *J* = 14.3, 8.1 Hz, 2H), 7.79 (t, *J* = 7.7 Hz, 1H), 7.62 (d, *J* = 7.6 Hz, 2H), 7.56-7.52 (m, 3H), 7.47 (t, *J* = 7.6 Hz, 2H), 7.39 (t, *J* = 7.3 Hz, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 161.3, 151.7, 143.5, 139.3, 135.0, 134.7, 130.7, 129.1, 128.9, 128.4, 127.1, 123.4, 123.3, 121.8, 121.2, 117.0, 115.9.



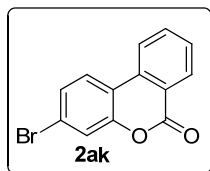
2ah: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.50 (dd, 1H), 8.39 (dd, *J* = 7.9, 0.8 Hz, 1H), 8.08 (d, *J* = 8.1 Hz, 1H), 7.94 (d, *J* = 8.8 Hz, 1H), 7.79 (dd, *J* = 10.7, 4.4 Hz, 2H), 7.67 (d, *J* = 8.7 Hz, 1H), 7.61-7.50 (m, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.2, 147.2, 135.4, 135.0, 134.3, 130.6, 128.6, 127.9, 127.7, 127.1, 124.5, 123.9, 122.3, 122.0, 121.2, 119.2, 113.0.



2ai: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H₂). Analytical data were in agreement with literature values¹¹: ¹H NMR (500 MHz, CDCl₃) δ 8.24 (d, *J* = 8.1 Hz, 1H), 8.01 (d, *J* = 7.9 Hz, 1H), 7.86 (s, 1H), 7.48-7.42 (m, 1H), 7.38-7.28 (m, 3H), 2.54 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.3, 151.5, 146.0, 134.8, 130.6, 130.4, 130.2, 124.5, 122.8, 121.9, 118.9, 118.1, 117.8, 22.4.

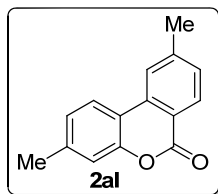


2aj: Prepared according to the general procedure outlined above and obtained as white solid (84% yield, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.34 (d, *J* = 7.9 Hz, 1H), 8.00 (t, *J* = 7.2 Hz, 2H), 7.85-7.78 (m, 1H), 7.55 (t, *J* = 7.6 Hz, 1H), 7.05 (tt, *J* = 8.9, 2.5 Hz, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 164.5, 162.5, 160.8, 152.3, 152.2, 135.2, 134.3, 130.7, 128.8, 124.5, 124.4, 121.6, 120.5, 114.7, 114.7, 112.6, 112.4, 105.2, 105.0.



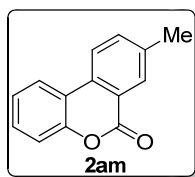
2ak: Prepared according to the general procedure outlined above and obtained

as colorless oil (97% yield, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.35 (d, *J* = 7.9 Hz, 1H), 8.03 (d, *J* = 8.1 Hz, 1H), 7.87 (d, *J* = 8.5 Hz, 1H), 7.82 (dd, *J* = 10.8, 4.5 Hz, 1H), 7.59 (t, *J* = 7.6 Hz, 1H), 7.48 (d, *J* = 1.8 Hz, 1H), 7.43 (dd, *J* = 8.5, 1.9 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 160.5, 151.5, 135.2, 134.1, 130.8, 129.4, 127.9, 124.0, 123.8, 121.7, 121.1, 120.9, 117.2.



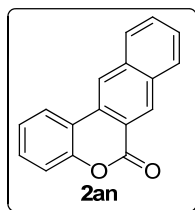
2al: Prepared according to the general procedure outlined above and obtained as white solid (95% yield, 99% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.22 (d, *J* = 8.1 Hz, 1H), 7.88-7.84 (m, 1H), 7.80 (s, 1H), 7.32 (d, *J* = 8.1 Hz, 1H), 7.10 (d, *J* = 7.3 Hz, 2H), 2.52 (s, 3H), 2.42 (s, 3H). ¹³C NMR (125 MHz, CDCl₃) δ 161.6, 151.5, 145.9, 141.2, 135.0, 130.6, 129.7, 125.6, 122.5, 121.6, 118.5, 117.9, 115.5, 22.4,

21.5.



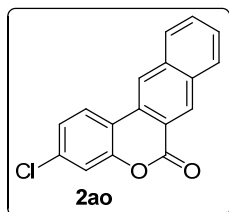
2am: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H₂). Analytical data were in agreement with literature values¹¹: ¹H NMR (500 MHz, CDCl₃) δ 8.17 (s, 1H), 8.02-7.96 (m, 2H), 7.61 (dd, *J* = 8.2, 1.3 Hz, 1H), 7.46-7.41 (m, 1H), 7.36-7.27 (m, 2H), 2.48 (s, 3H). ¹³C NMR (100 MHz, CDCl₃) δ 161.5, 151.1, 139.3, 136.1, 132.3, 130.4, 130.0,

124.6, 122.6, 121.8, 121.2, 118.3, 117.7, 21.4.



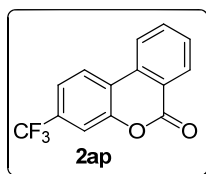
2an: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H₂). Analytical data were in agreement with literature values¹²: ¹H NMR (500 MHz, CDCl₃) δ 8.93 (s, 1H), 8.43 (s, 1H), 8.14 (dd, *J* = 7.9, 1.5 Hz, 1H), 7.96 (dd, *J* = 10.8, 8.3 Hz, 2H), 7.64 (ddd, *J* = 8.1, 6.6, 1.2 Hz, 1H), 7.55 (ddd, *J* = 8.0, 6.7, 1.2 Hz, 1H), 7.44 (td, *J* = 7.7, 1.5 Hz, 1H), 7.37-7.29 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 161.6, 150.9, 136.3, 132.9,

132.5, 130.2, 129.7, 129.7, 129.6, 128.2, 127.3, 124.7, 123.0, 120.8, 119.3, 118.4, 118.0.



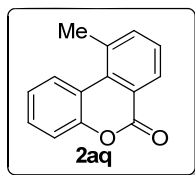
2ao: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H₂). ¹H NMR (500 MHz, CDCl₃) δ 8.90 (s, 1H), 8.39 (s, 1H), 8.06 (d, *J* = 8.3 Hz, 1H), 7.97 (t, *J* = 7.6 Hz, 2H), 7.68 (dd, *J* = 11.5, 4.4 Hz, 1H), 7.58 (t, *J* = 7.5 Hz, 1H), 7.34-7.27 (m, 2H). ¹³C NMR (125 MHz, CDCl₃) δ 160.96, 151.15, 136.28, 135.61, 133.16, 132.60, 129.96, 129.70, 128.89, 128.20, 127.55, 125.17, 123.98, 120.84, 118.79, 118.18,

117.10. IR (KBr, cm⁻¹): 1733, 1567, 1511, 1225, 765. HRMS (ESI) calcd for C₁₇H₉ClO₂Na⁺: 303.0183, found 303.0184.



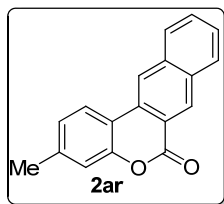
2ap: Prepared according to the general procedure outlined above and obtained as white solid (64% yield, 69% H₂). Analytical data were in agreement with literature values¹⁰: ¹H NMR (500 MHz, CDCl₃) δ 8.43 (dd, *J* = 7.9, 0.8 Hz, 1H), 8.17 (t, *J* = 9.0 Hz, 2H), 7.92-7.87 (m, 1H), 7.68 (t, *J* = 7.6 Hz, 1H), 7.62 (s, 1H), 7.59 (d, *J* = 8.3 Hz, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 160.3, 151.1, 135.3,

133.5, 132.5, 132.2, 130.9, 130.2, 124.8, 123.7, 122.3, 122.1, 121.8, 121.2, 121.2, 115.4, 115.3.

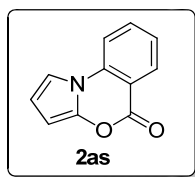


2aq: Prepared according to the general procedure outlined above and obtained as white solid (73% yield, 77% H₂). Analytical data were in agreement with literature values¹²: ¹H NMR (500 MHz, CDCl₃) δ 8.36 (dd, *J* = 7.9, 1.6 Hz, 1H), 8.29 (dd, *J* = 8.4, 1.5 Hz, 1H), 7.63 (dd, *J* = 7.5, 1.5 Hz, 1H), 7.51-7.42 (m, 2H), 7.39 (dd, *J* = 8.2, 1.6 Hz, 1H), 7.32 (ddd, *J* = 8.5, 7.1, 1.5 Hz, 1H), 2.89 (s, 3H).

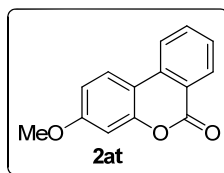
^{13}C NMR (125 MHz, CDCl_3) δ 161.8, 151.3, 139.2, 135.1, 133.6, 129.7, 129.2, 128.3, 127.3, 124.1, 122.9, 119.8, 118.0, 25.5.



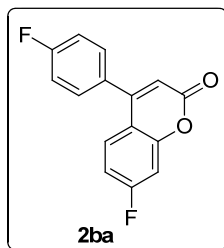
2ar: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H_2). ^1H NMR (500 MHz, CDCl_3) δ 8.93 (s, 1H), 8.40 (s, 1H), 8.02 (d, J = 8.1 Hz, 1H), 7.96 (dd, J = 14.9, 8.3 Hz, 2H), 7.64 (t, J = 7.5 Hz, 1H), 7.54 (t, J = 7.5 Hz, 1H), 7.14 (d, J = 8.1 Hz, 1H), 7.11 (s, 1H), 2.43 (s, 3H). ^{13}C NMR (125 MHz, CDCl_3) δ 161.84, 150.90, 141.05, 136.36, 132.87, 132.33, 130.01, 129.66, 129.60, 128.11, 127.04, 125.85, 122.76, 120.30, 119.18, 118.14, 115.75, 21.54. IR (KBr, cm^{-1}): 1726, 1587, 1533, 1165, 735. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{12}\text{O}_2\text{Na}^+$: 283.0730, found 283.0729.



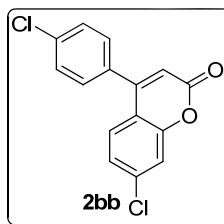
2as: Prepared according to the general procedure outlined above and obtained as white solid (63% yield, 78% H_2). Analytical data were in agreement with literature values¹³: ^1H NMR (400 MHz, CDCl_3) δ 8.22 (dd, J = 7.8, 1.6 Hz, 1H), 7.76 (ddd, J = 8.7, 7.4, 1.6 Hz, 1H), 7.49 (d, J = 8.3 Hz, 1H), 7.33 (td, J = 7.8, 1.0 Hz, 1H), 7.00 (dd, J = 3.5, 1.8 Hz, 1H), 6.40 (t, J = 3.6 Hz, 1H), 5.80 (dd, J = 3.7, 1.7 Hz, 1H). ^{13}C NMR (100 MHz, CDCl_3) δ 158.1, 140.0, 138.1, 136.8, 131.6, 124.9, 113.6, 111.4, 110.8, 106.8, 89.6.



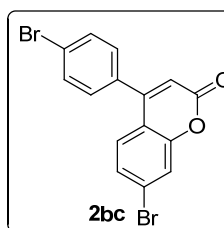
2at: Prepared according to the general procedure outlined above and obtained as white solid (72% yield, 81% H_2). Analytical data were in agreement with literature values¹⁰: ^1H NMR (400 MHz, CDCl_3) δ 8.37 (d, J = 7.9 Hz, 1H), 8.01 (d, J = 8.1 Hz, 1H), 7.96 (d, J = 8.8 Hz, 1H), 7.79 (t, 1H), 7.51 (t, J = 7.6 Hz, 1H), 7.03-6.70 (m, 2H), 3.89 (s, 3H). ^{13}C NMR (100 MHz, CDCl_3) δ 161.7, 161.6, 152.8, 135.4, 135.0, 130.7, 127.9, 123.9, 121.2, 120.2, 112.6, 111.3, 101.8, 55.9.



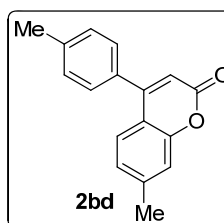
2ba: Prepared according to the general procedure outlined above and obtained as white solid (94% yield, 99% H_2). ^1H NMR (500 MHz, CDCl_3) δ 7.44 (dt, J = 8.6, 5.5 Hz, 3H), 7.29-7.21 (m, 2H), 7.13 (dd, J = 8.9, 2.5 Hz, 1H), 6.99 (td, J = 8.4, 2.6 Hz, 1H), 6.31 (s, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 165.7, 164.7, 163.7, 162.8, 160.3, 155.6, 155.5, 154.3, 131.2, 131.2, 130.5, 130.4, 129.2, 129.1, 128.7, 128.6, 116.5, 116.3, 115.8, 115.8, 115.5, 115.3, 114.3, 114.3, 112.6, 112.4, 105.1, 104.9. IR (KBr, cm^{-1}): 1725, 1589, 1529, 1479, 1445, 708, 694, 623. HRMS (ESI) calcd for $\text{C}_{15}\text{H}_8\text{F}_2\text{O}_2\text{Na}^+$: 281.0385, found 281.0385.



2bb: Prepared according to the general procedure outlined above and obtained as white solid (91% yield, 99% H_2). Analytical data were in agreement with literature values¹⁴: ^1H NMR (500 MHz, CDCl_3) δ 7.53 (d, J = 8.4 Hz, 2H), 7.43 (d, J = 2.1 Hz, 1H), 7.38 (dd, J = 8.5, 5.4 Hz, 3H), 7.22 (dd, J = 8.6, 2.1 Hz, 1H), 6.35 (s, 1H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.9, 154.6, 154.0, 138.3, 136.4, 133.3, 129.8, 129.5, 127.7, 125.0, 117.8, 117.5, 115.4.

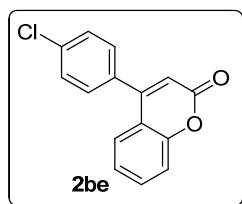


2bc: Prepared according to the general procedure outlined above and obtained as white solid (86% yield, 89% H_2). Analytical data were in agreement with literature values¹⁴: ^1H NMR (500 MHz, CDCl_3) δ 7.68 (d, J = 8.2 Hz, 2H), 7.59 (d, J = 1.9 Hz, 1H), 7.37 (dd, J = 8.6, 1.9 Hz, 1H), 7.30 (dd, J = 10.2, 8.4 Hz, 3H), 6.37 (s, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 159.8, 154.5, 154.1, 133.7, 132.5, 130.1, 127.9, 127.8, 126.3, 124.6, 120.8, 117.8, 115.5.

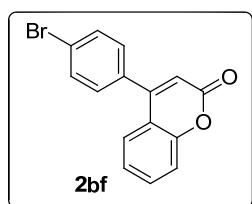


2bd: Prepared according to the general procedure outlined above and

obtained as white solid (88% yield, 91% H₂). Analytical data were in agreement with literature values¹⁴: ¹H NMR (500 MHz, CDCl₃) δ 7.40 (d, *J* = 8.1 Hz, 1H), 7.38-7.27 (m, 4H), 7.20 (d, *J* = 1.6 Hz, 1H), 7.03 (dd, *J* = 8.2, 1.6 Hz, 1H), 6.29 (s, 1H), 2.45 (s, 6H). ¹³C NMR (125 MHz, CDCl₃) δ 161.3, 155.9, 154.4, 143.2, 139.9, 132.6, 129.6, 128.5, 126.8, 125.4, 117.6, 116.8, 113.9, 21.7, 21.5.

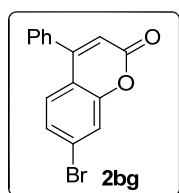


2be: Prepared according to the general procedure outlined above and obtained as white solid (99% yield, 99% H₂). Analytical data were in agreement with literature values¹⁴: ¹H NMR (500 MHz, CDCl₃) δ 7.57 (t, *J* = 7.8 Hz, 1H), 7.52 (d, *J* = 8.3 Hz, 2H), 7.48-7.36 (m, 4H), 7.25 (t, 1H), 6.36 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 160.6, 154.6, 154.3, 136.1, 133.7, 132.3, 129.9, 129.4, 126.8, 124.4, 118.8, 117.6, 115.5.

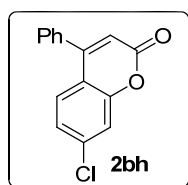


2bf: Prepared according to the general procedure outlined above and obtained as white solid (95% yield, 99% H₂). Analytical data were in agreement with literature values¹⁴: ¹H NMR (400 MHz, CDCl₃) δ 7.68 (d, *J* = 8.4 Hz, 2H), 7.57 (ddd, *J* = 8.6, 7.3, 1.6 Hz, 1H), 7.43 (td, *J* = 8.3, 1.4 Hz, 2H), 7.38-7.31 (m, 2H), 7.29-7.16 (m, 1H), 6.36 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 160.5, 154.6, 154.3, 134.2, 132.3, 132.3, 130.2, 126.8, 124.4,

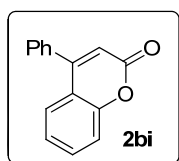
124.3, 118.8, 117.6, 115.5.



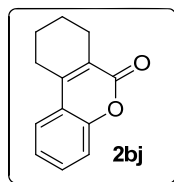
2bg: Prepared according to the general procedure outlined above and obtained as white solid (85% yield, 88% H₂). Analytical data were in agreement with literature values¹⁴: ¹H NMR (400 MHz, CDCl₃) δ 7.59 (t, *J* = 1.1 Hz, 1H), 7.56-7.51 (m, 3H), 7.46-7.40 (m, 2H), 7.36 (d, *J* = 1.2 Hz, 2H), 6.39 (s, 1H). ¹³C NMR (100 MHz, CDCl₃) δ 160.1, 155.3, 154.6, 134.9, 130.1, 129.2, 128.5, 128.2, 127.7, 126.1, 120.7, 118.2, 115.4.



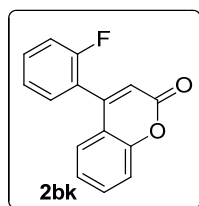
2bh: Prepared according to the general procedure outlined above and obtained as white solid (71% yield, 78% H₂). Analytical data were in agreement with literature values¹⁴: ¹H NMR (500 MHz, CDCl₃) δ 7.59-7.50 (m, 3H), 7.43 (dd, *J* = 9.2, 2.6 Hz, 4H), 7.21 (dd, *J* = 8.6, 2.1 Hz, 1H), 6.37 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 160.2, 155.2, 154.6, 138.0, 134.9, 130.1, 129.2, 128.5, 128.1, 124.9, 117.8, 117.7, 115.1.



2bi: Prepared according to the general procedure outlined above and obtained as white solid (95% yield, 99% H₂). Analytical data were in agreement with literature values¹⁴: ¹H NMR (500 MHz, CDCl₃) δ 7.61-7.40 (m, 8H), 7.24 (dt, *J* = 11.1, 4.1 Hz, 1H), 6.39 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 160.9, 155.8, 154.4, 135.4, 132.1, 129.8, 129.0, 128.6, 127.2, 124.3, 119.2, 117.5, 115.4.

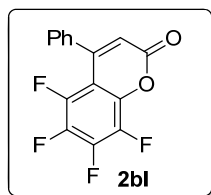


2bj: Prepared according to the general procedure outlined above and obtained as white solid (81% yield, 89% H₂). Analytical data were in agreement with literature values¹⁵: ¹H NMR (500 MHz, Chloroform-*d*) δ 7.56 (dd, *J* = 7.9, 1.4 Hz, 1H), 7.48-7.43 (m, 1H), 7.33-7.24 (m, 2H), 2.79 (ddd, *J* = 6.3, 4.1, 2.2 Hz, 2H), 2.62-2.56 (m, 2H), 1.91-1.76 (m, 4H). ¹³C NMR (126 MHz, CDCl₃) δ 161.9, 152.1, 147.1, 130.3, 124.1, 123.9, 123.2, 120.3, 116.8, 25.3, 24.2, 21.7, 21.5.

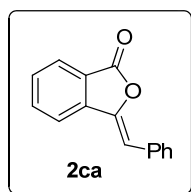


2bk: Prepared according to the general procedure outlined above and obtained as white solid (75% yield). Analytical data were in agreement with literature values¹⁶: ¹H NMR (400 MHz, CDCl₃) δ 7.67-7.48 (m, 2H), 7.46-7.31 (m, 3H), 7.31-7.20 (m, 3H), 6.43 (s, 1H). ¹³C NMR (125 MHz, CDCl₃) δ 160.6, 160.3,

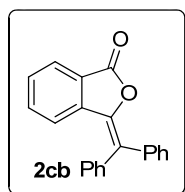
158.3, 154.0, 150.6, 132.2, 131.8, 131.8, 130.6, 130.6, 126.9, 124.9, 124.9, 124.4, 123.1, 122.9, 118.9, 117.4, 117.1, 117.1, 116.5, 116.4. ^{19}F NMR (377 MHz, CDCl_3) δ -112.3 (s).



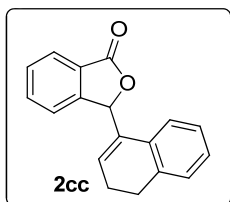
2bl: Prepared according to the general procedure outlined above and obtained as white solid (59% yield). ^1H NMR (500 MHz, CDCl_3) δ 7.49 (qd, J = 8.7, 7.8, 3.7 Hz, 3H), 7.37 (dt, J = 6.3, 2.0 Hz, 2H), 6.35 (s, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 157.4, 152.2, 152.2, 135.9, 135.9, 130.0, 128.6, 127.4, 127.4, 118.0, 118.0. ^{19}F NMR (377 MHz, Acetonitrile- d_3) δ -137.17 (ddd, J = 20.6, 10.4, 4.2 Hz), -153.42 (td, J = 20.4, 4.4 Hz), -161.42 (ddd, J = 20.1, 10.8, 1.8 Hz), -164.99 (td, J = 20.5, 1.8 Hz). IR (KBr, cm^{-1}): 1739, 1653, 1589, 1529, 1479, 1417, 1360, 1201, 1130, 1028, 1011, 880, 871, 856, 781, 766, 755, 708, 696, 611. HRMS (ESI) calcd for $\text{C}_{15}\text{H}_6\text{F}_4\text{O}_2\text{Na}^+$: 317.0196, found 317.0196.



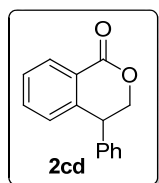
2ca: Prepared according to the general procedure outlined above and obtained as white solid (73% yield, 88% H_2). Analytical data were in agreement with literature values¹⁷: ^1H NMR (500 MHz, CDCl_3) δ 7.95 (d, J = 7.7 Hz, 1H), 7.86 (d, J = 8.0 Hz, 2H), 7.78 (d, J = 7.8 Hz, 1H), 7.73 (t, J = 7.5 Hz, 1H), 7.55 (t, J = 7.4 Hz, 1H), 7.42 (t, J = 7.6 Hz, 2H), 7.32 (t, J = 7.4 Hz, 1H), 6.43 (s, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 167.2, 144.7, 140.8, 134.6, 133.2, 130.3, 129.9, 128.9, 128.6, 125.7, 123.6, 120.0, 107.2.



2cb: Prepared according to the general procedure outlined above and obtained as white solid (81% yield, 92% H_2). Analytical data were in agreement with literature values¹⁸: ^1H NMR (500 MHz, Chloroform- d) δ 7.90 (d, J = 7.6 Hz, 1H), 7.60-7.54 (m, 2H), 7.52 (dd, J = 4.8, 1.9 Hz, 3H), 7.42 (t, J = 7.5 Hz, 1H), 7.40-7.27 (m, 6H), 6.30 (d, J = 8.0 Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 167.3, 142.6, 139.7, 137.7, 137.5, 134.0, 130.7, 130.6, 129.5, 129.5, 128.9, 128.3, 128.3, 125.4, 125.0, 125.0, 123.7.



2cc: Prepared according to the general procedure outlined above and obtained as white solid (85% yield, 93% H_2). ^1H NMR (400 MHz, CDCl_3) δ 8.08-7.89 (m, 1H), 7.65 (td, J = 7.5, 1.2 Hz, 1H), 7.56 (t, J = 7.5 Hz, 1H), 7.41 (ddd, J = 15.2, 7.1, 1.6 Hz, 2H), 7.29-7.09 (m, 3H), 6.44 (s, 1H), 6.12-5.91 (m, 1H), 2.76 (t, J = 8.0 Hz, 2H), 2.29 (dtd, J = 9.2, 4.9, 2.8 Hz, 2H). ^{13}C NMR (101 MHz, CDCl_3) δ 170.5, 148.6, 136.6, 134.0, 133.2, 132.8, 130.0, 129.4, 128.1, 127.8, 126.8, 126.6, 126.0, 123.4, 123.1, 80.8, 27.8, 23.1. IR (KBr, cm^{-1}): 2937, 2886, 2831, 1771, 1613, 1489, 1466, 1451, 1285, 1064, 954, 765, 737. HRMS (ESI) calcd for $\text{C}_{18}\text{H}_{14}\text{O}_2\text{Na}^+$: 285.0886, found 285.0886.



2cd: Prepared according to the general procedure outlined above and obtained as white solid (45% yield). Analytical data were in agreement with literature values¹⁹: ^1H NMR (500 MHz, CDCl_3) δ 8.19 (dd, J = 7.8, 1.3 Hz, 1H), 7.52 (td, J = 7.5, 1.4 Hz, 1H), 7.44 (t, J = 7.6 Hz, 1H), 7.36 (dt, J = 13.8, 6.8 Hz, 3H), 7.23-7.15 (m, 2H), 7.03 (d, J = 7.7 Hz, 1H), 4.78-4.51 (m, 2H), 4.39 (dd, J = 8.4, 4.9 Hz, 1H). ^{13}C NMR (125 MHz, CDCl_3) δ 165.1, 142.7, 138.3, 134.1, 130.7, 129.2, 128.8, 128.1, 128.0, 127.7, 125.2, 72.3, 43.8.

Electrochemical analysis and CV profiles

Electrochemical potentials were obtained with a standard set of conditions to main internal consistency. Cyclic voltammograms were collected with a Shanghai Chenhua CH1600D potentiostat. Samples were prepared with 0.05 mmol of substrate in 5 mL of 0.1 M Bu₄NPF₆ in CF₃CH₂OH. Measurements employed a 1.5 mm radius glassy carbon working electrode, platinum wire counter electrode, saturated KCl silver-silver chloride reference electrode, and a scan rate of 100 mV/s. The glassy carbon electrode was polished between each scan. Carboxylate salts were made by reaction of the corresponding acid with 1 equivalent of sodium hydroxide or TBA hydroxide bought in a solution of methanol. The solvent was then evaporated in vacuo. CV measurements were immediately taken once the salts were determined to be free of solvent. The obtained value was referenced to Ag/AgCl and converted to SCE by subtracting 0.045 V.

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NMR spectra:

