

Supporting Information for
Co(III)-Catalyzed *N*-Chloroamide-Directed C-H Activation for
3,4-Dihydroisoquinolone Synthesis

*Xiaolong Yu, Kehao Chen, Qi Wang, Wenjing Zhang, and Jin Zhu**

*Department of Polymer Science and Engineering, School of Chemistry and Chemical
Engineering, State Key Laboratory of Coordination Chemistry, Nanjing National
Laboratory of Microstructures, Collaborative Innovation Center of Chemistry for Life
Sciences,
Nanjing University, Nanjing 210023, China*

*Corresponding author. Email: jinz@nju.edu.cn; Phone: +86-25-89686291; Fax:
+86-25-83317761

1. General Information	2
2. Optimization of Reaction Conditions	3
3. Oxidizing Reactivity of N-Cl Bond Experiments.....	3
4. Mechanism Experiments	7
5. Deuterium Exchange Experiments	7
6. Kinetic Isotope Effect Experiment	8
7. Competition Experiment	10
8 Synthesis of Substrates.....	11
9. Preparation and Characterization Data of Products.....	12
10. References	30
11. NMR Spectra for New Compounds	31

1. General Information

All reactions were carried out under a dry nitrogen atmosphere. All commercial reagents were used without additional purification, unless otherwise stated. Anhydrous solvent was purchased from commercial sources and transferred under an argon atmosphere. Analytical thin layer chromatography (TLC) was performed on precoated silica gel 60 GF254 plates. Flash column chromatography was performed using Tsing dao silica gel (60, particle size 0.040 - 0.063 mm). Visualization on TLC was achieved by use of UV light (254 nm) or iodine. NMR spectra were recorded on a Bruker DPX 400 spectrometer at 400 MHz for ^1H NMR, 101 MHz for ^{13}C NMR in CDCl_3 or DMSO with tetramethylsilane (TMS) as internal standard. The chemical shifts are expressed in ppm and coupling constants are given in Hz. Data for ^1H NMR were recorded as follows: chemical shift (ppm), multiplicity (s, singlet; d, doublet; t, triplet; q, quarter; m, multiplet; br, broad), coupling constant (Hz), integration. Data for ^{13}C NMR were reported in terms of chemical shift (δ , ppm). High-resolution mass spectrometric data were obtained using Bruker Apex IV RTMS. Column conditions are reported in the experimental section below.

2. Optimization of Reaction Conditions

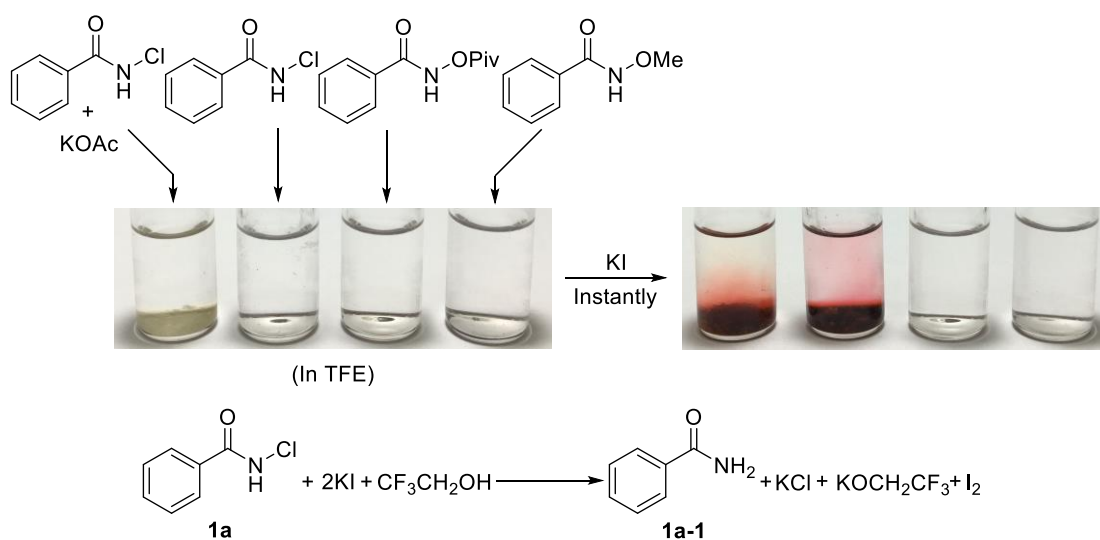
Table S1. Optimization of reaction conditions^a

$\text{1a} + \text{2a} \xrightarrow[\text{base (y mol \%)}]{\text{catalyst (x mol \%), AgOTf (20 mol \%), TFE, rt, 36 h}} \text{3aa}$

entry	catalyst (x mol %)	base (y mol %)	yield ^b (%)
1	[RuCp*Cl ₂] ₂ (5)	KOAc (120)	-
2	Co(acac) ₂ (10)	KOAc (120)	-
3	[CoCp*(CO)I ₂] (10)	KOAc (120)	58 ^c
4	[CoCp*(CO)I ₂] (10)	NaOAc (120)	65
5	[CoCp*(CO)I ₂] (10)	KO ^t Bu (120)	58
6	[CoCp*(CO)I ₂] (10)	KOAc (20)	11
7	[CoCp*(CO)I ₂] (10)	KOAc (50)	25

^aReaction conditions: **1a** (0.20 mmol), **2a** (0.30 mmol), TFE (1.0 mL), N₂. ^bIsolated yields are shown. ^cAir.

3. Oxidizing Reactivity of N-Cl Bond Experiments



To four 1.5 ml sample bottles were respectively added *N*-chlorobenzamide (0.2 mmol, 32 mg) and KOAc (0.24 mmol, 24 mg) in CF₃CH₂OH (1 mL) under air, *N*-chlorobenzamide (0.2 mmol, 32 mg) in CF₃CH₂OH (1 mL) under air, *N*-pivaloyloxybenzamide (0.2 mmol, 44 mg) in CF₃CH₂OH (1 mL) under air, and *N*-methoxybenzamide (0.2 mmol, 30 mg) in CF₃CH₂OH (1 mL) under air. And KI (0.4 mmol, 66.4 mg) was added respectively.

The high oxidizing power of *N*-chlorobenzamides is evidenced by upon mixing of *N*-chlorobenzamide (**1a**) with KI in trifluoroethanol (TFE; in the absence or presence of KOAc), an instant color change to reddish brown characteristic of the generation of I₂. The same color change is not observed by the replacement of **1a** with *N*-pivaloyloxybenzamide or *N*-methoxybenzamide, indicating that N-Cl bond is more oxidizing than N-O bond.

Cyclic voltammetry (CV) was carried out on a CHI 660D electrochemical workstation (Shanghai Chen Hua Instrument Co. Ltd., China) with a conventional three-electrode system composed of a glassy carbon electrode as the working electrode, a platinum wire auxiliary electrode, and a saturated calomel reference electrode. The higher oxidizing capability of N-Cl bond is also evidenced by cyclic voltammetry measurement.

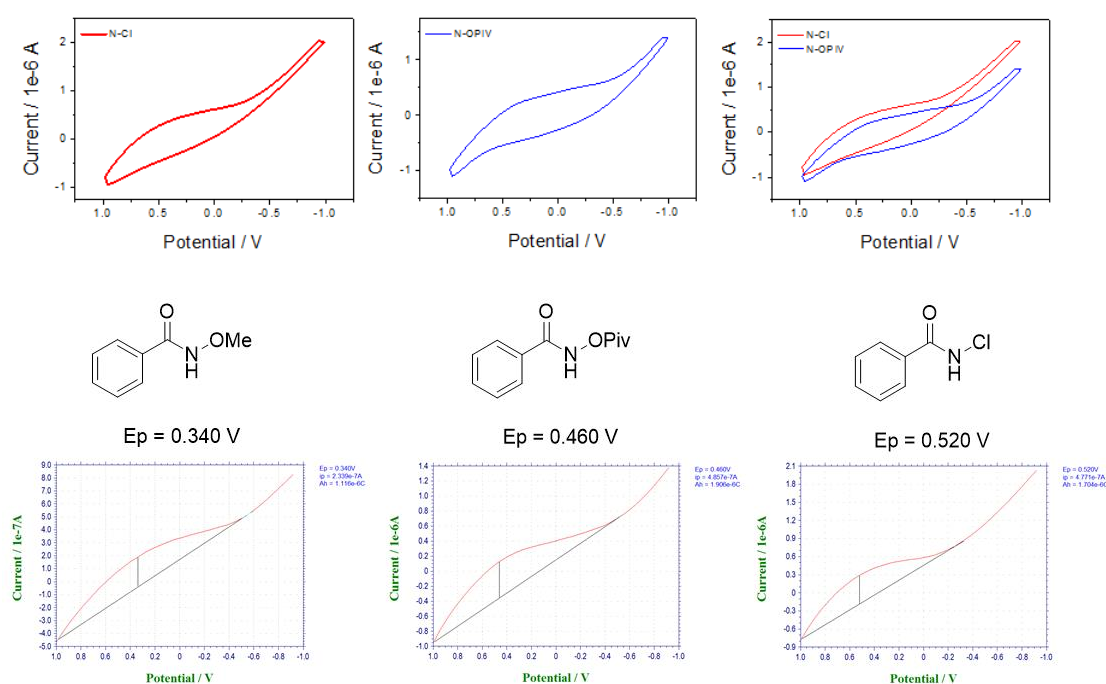
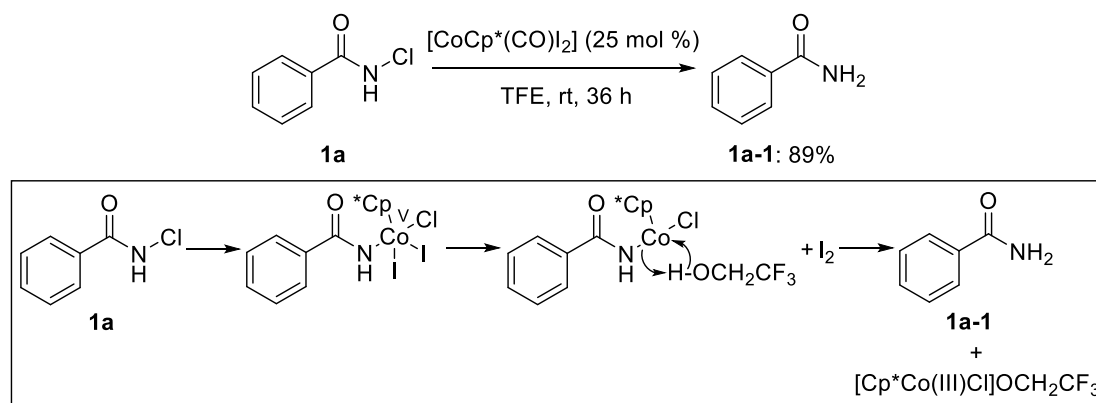


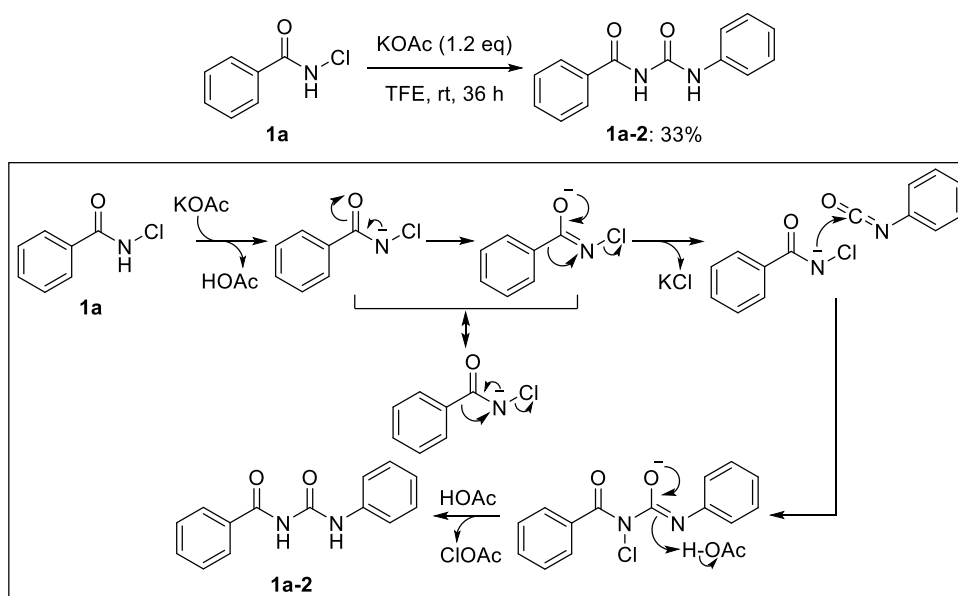
Figure S1. Cyclic voltammetry data (CF₃CH₂OH, 0.2 mmol/mL, 100 mV/s) for complexes **1a** (N-Cl), *N*-pivaloyloxybenzamide (N-OPiv) and *N*-methoxybenzamide



To a 13 × 150 mm test tube equipped with magnetic stir bar were added

N-chlorobenzamide (31.2 mg, 0.2 mmol), [CoCp*(CO)I₂] (25 mg, 25 mol %) under nitrogen. Then, 1 mL of CF₃CH₂OH was injected into the tube through a syringe. The resulting solution was stirred at rt for 36 h. The reaction mixture was concentrated in vacuo, the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 3:1). The corresponding product was obtained as a white solid (22 mg, 89%).

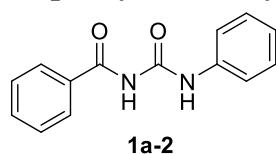
The oxidizing power of *N*-chlorobenzamides can translate to smooth conversion of **1a** to benzamide (**1a-1**, 89% yield) under [CoCp*(CO)I₂] catalysis at rt, with N-Cl bond oxidative addition and concomitant Co^{III}-to-Co^V transformation¹ as the critical step.



To a 13 × 150 mm test tube equipped with magnetic stir bar were added *N*-chlorobenzamide (31 mg, 0.2 mmol) and KOAc (24 mg, 120 mol %) under nitrogen. Into the tube was then injected 1 mL of CF₃CH₂OH via syringe. The resulting solution was stirred at rt for 36 h. The reaction mixture was concentrated in vacuo, the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 3:1). The corresponding product was obtained as a white solid (8 mg, 33%).

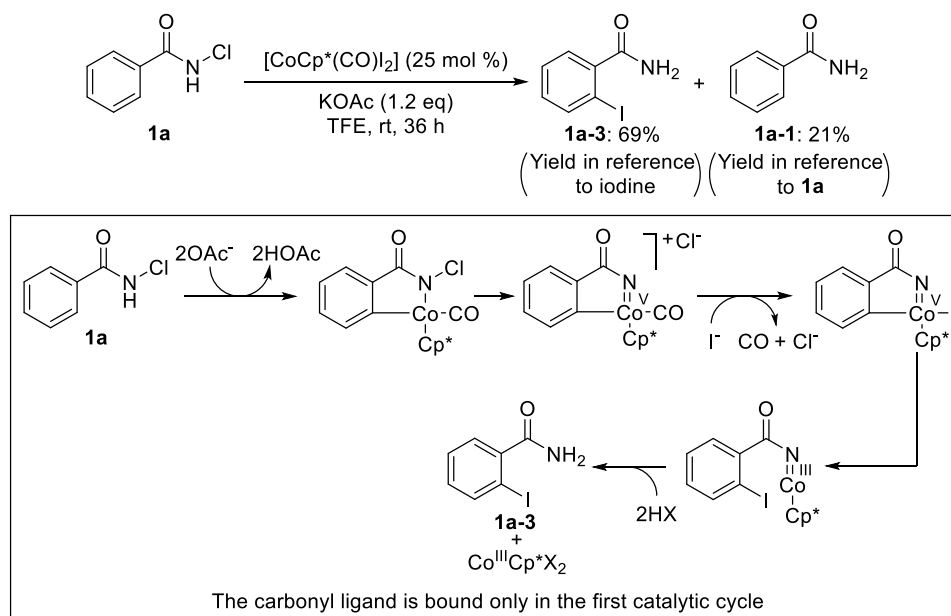
Switching of [CoCp*(CO)I₂] to KOAc results in the exhibition of nitrene-like reactivity and generation of group-migrated *N*-(phenylcarbamoyl)benzamide (**1a-2**) through initial deprotonation of **1a**. In fact, the group migration process can also be viewed as a formal oxidation of C atoms by the N-Cl bond.

N-(phenylcarbamoyl)benzamide (**1a-2**)²



^1H NMR (400 MHz, CDCl_3) δ 11.01 (s, 1H), 10.04 (s, 1H), 8.17 – 8.01 (m, 2H), 7.69 – 7.57 (m, 3H), 7.52 (dd, J = 10.6, 4.9 Hz, 2H), 7.41 – 7.32 (m, 2H), 7.20 – 7.10 (m, 1H).

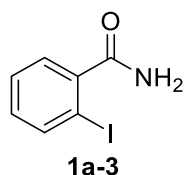
^{13}C NMR (101 MHz, CDCl_3) δ 168.68, 152.13, 137.24, 133.35, 132.16, 129.01, 128.86, 128.06, 124.45, 120.47.



To a 13×150 mm test tube equipped with magnetic stir bar were added N-chlorobenzamide (31 mg, 0.2 mmol), $[\text{CoCp}^*(\text{CO})\text{I}_2]$ (25 mg, 25 mol %) and KOAc (24 mg, 120 mol %) under nitrogen. Into the tube was then injected 1 mL of $\text{CF}_3\text{CH}_2\text{OH}$ via syringe. The resulting solution was stirred at rt for 36 h. The reaction mixture was concentrated in vacuo, the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 3:1). The corresponding products were obtained as white solids: 2-iodobenzamide (8.5 mg, 69%, the yield is calculated according to the amount of iodine ion) and benzamide (5 mg, 21%).

The combination of $[\text{CoCp}^*(\text{CO})\text{I}_2]$ with KOAc enables complete suppression of group migration pathway and elicitation of C-H activation capability for cobalt. Indeed, the formation of 2-iodobenzamide (**1a-3**) under this condition supports the following mechanistic sequence: N-H deprotonation, cobalt coordination, C-H activation, oxidative addition of N-Cl bond and simultaneous oxidation of Co^{III} to Co^{V} (in the form of Co^{V} -nitrenoid), I^- coordination, C-I reductive elimination and proto-demetalation to regenerate Co^{III} .

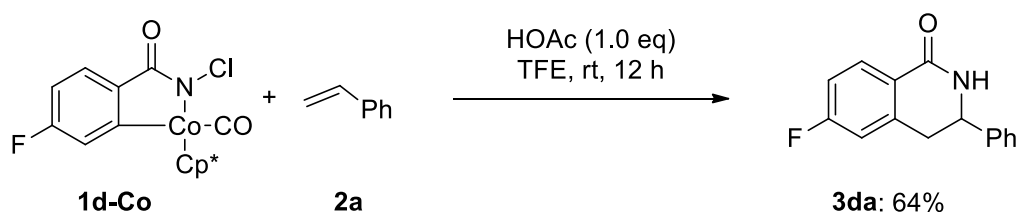
2-iodobenzamide (**1a-3**)



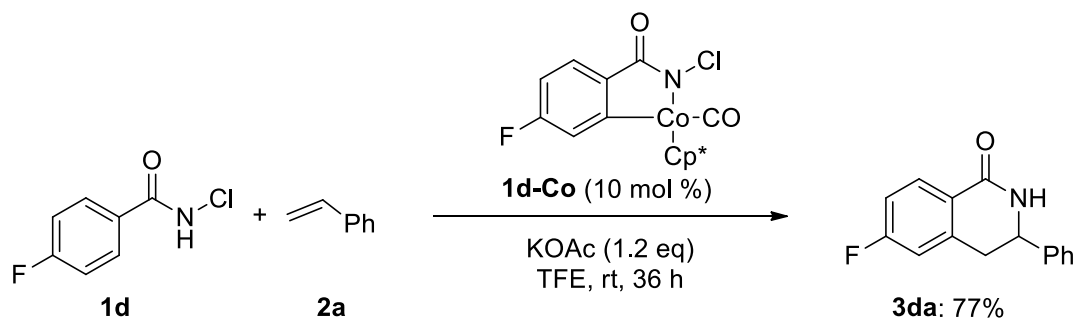
^1H NMR (400 MHz, DMSO) δ 7.87 (dd, J = 7.9, 0.9 Hz, 1H), 7.81 (s, 1H), 7.51 (s,

1H), 7.42 (td, $J = 7.5, 1.1$ Hz, 1H), 7.34 (dd, $J = 7.6, 1.7$ Hz, 1H), 7.19 – 7.11 (m, 1H).
 ^{13}C NMR (101 MHz, DMSO) δ 170.66, 143.11, 139.12, 130.54, 127.82 (d, $J = 15.0$ Hz), 93.09.
 HRMS (EI) m/z calcd. for $\text{C}_7\text{H}_6\text{INO} [\text{M}]^+$ 246.9494, found 246.9496.

4. Mechanism Experiments



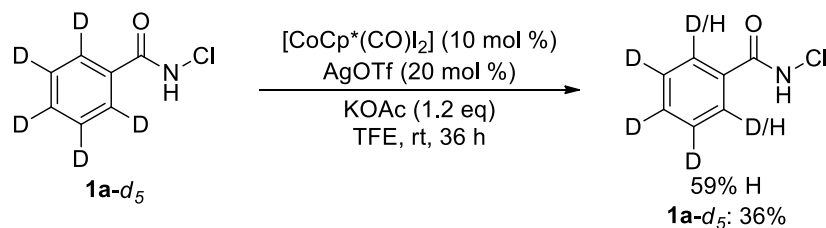
To a 13 × 150 mm test tube equipped with magnetic stir bar were added **1d-Co** (43 mg, 0.11 mmol), **2a** (18 mg, 150 mol %) and AcOH (7.0 mg, 100 mol %) in $\text{CF}_3\text{CH}_2\text{OH}$ (1 mL) under nitrogen. The resulting solution was stirred at rt for 20 h. The reaction was concentrated in vacuo, the residue was purified by column chromatography (silica gel, PE:EtOAc = 3:1). **3da** was obtained as white solid (17 mg, 64%).



To a 13 × 150 mm test tube equipped with magnetic stir bar were added **1d** (35 mg, 0.2 mmol), **1d-Co** (8 mg, 10 mol %), **2a** (32 mg, 150 mol %) and KOAc (24 mg, 120 mol %) in $\text{CF}_3\text{CH}_2\text{OH}$ (1 mL) under nitrogen. The resulting solution was stirred at rt for 36 h. The reaction was concentrated in vacuo, the residue was purified by column chromatography (silica gel, PE:EtOAc = 3:1). **3da** was obtained as white solid (37 mg, 77%).

5. Deuterium Exchange Experiments

General procedure for the catalytic deuterium exchange experiments:



To a 13 × 150 mm test tube equipped with magnetic stir bar were added *N*-chlorobenzamide **1a-d₅** (32 mg, 0.2 mmol), [CoCp*(CO)I₂] (10 mg, 10 mol %), KOAc (24 mg, 120 mol %) and AgOTf (10 mg, 20 mol %) under nitrogen. Then, 1 mL of CF₃CH₂OH was injected into the tube through a syringe. The resulting solution was stirred at rt for 36 h. The reaction mixture was concentrated in vacuo and the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 4:1). The reaction mixture was subjected to ¹H NMR measurement. **1a-d₅** was obtained as white solid (12 mg, 36%).

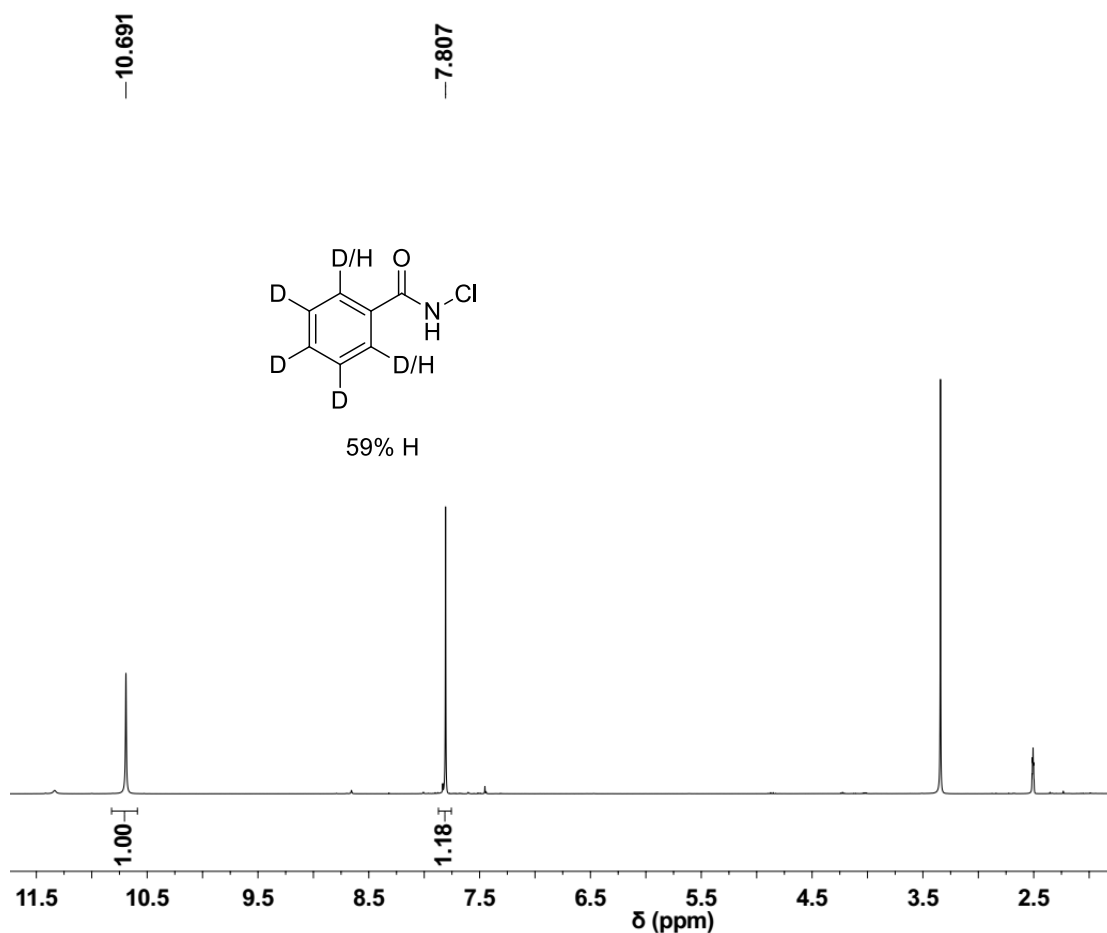
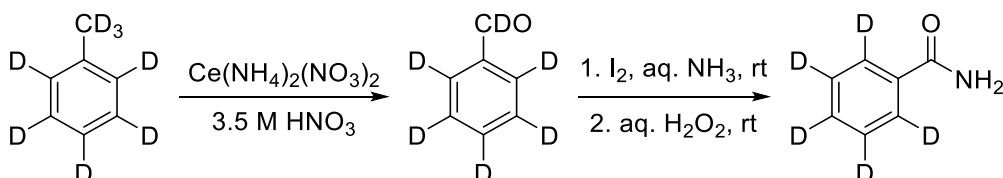
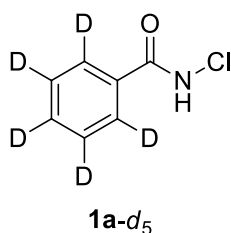


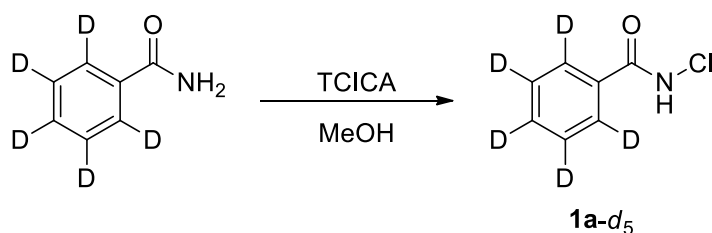
Figure S2. ¹H NMR spectrum for the product of equation above

6. Kinetic Isotope Effect Experiment

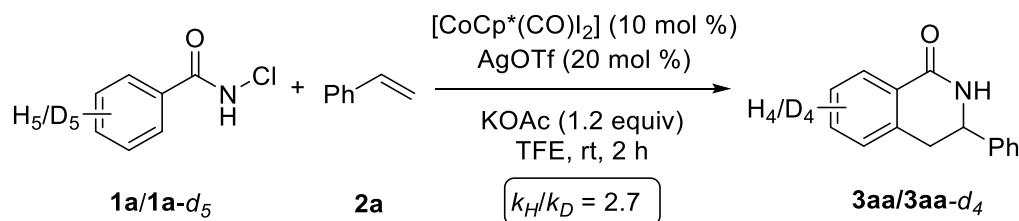
Preparation of deuterated 3-phenyl-1,2,4-oxadiazol-5(4*H*)-one:



To *d*₈-toluene (2.5 g, 25 mmol) in 3.5 M HNO₃ (50 mL) was added Ce(NH₄)₂(NO₃)₂ (54.8 g, 100 mmol) in 3.5 M HNO₃ (200 mL). After stirring for 2.5 h at 80 °C, the solution was cooled to room temperature, extracted with chloroform (3×50 mL), and the combined organic layers were washed with water until pH ≈ 7. The orange extract was dried over Na₂SO₄ and the solvent was removed in vacuum. The resulting red residue was distilled to give *d*₆-benzaldehyde, which was used without purification. A solution of the appropriate aldehyde (5 mmol) and iodine (5.5 mmol) in ammonia water (30 mL of 28% solution) and THF (5 mL) was stirred at room temperature for 1 h. The dark solution became colorless at the end of reaction. Then, aqueous H₂O₂ (3 mL of 35% solution) was added dropwise. The reaction mixture was stirred for 2-4 h and extracted with CH₂Cl₂. The organic phase was washed with brine, dried over Na₂SO₄, and concentrated in vacuo without purification.



In a 100 mL, round-bottom flask were added 30 mL methanol and *d*₅-benzamide (4 mmol), and the mixture was allowed to stir until the solid dissolved. TCICA (trichloroisocyanuric acid) (1.5 mmol), was added and a precipitate of cyanuric acid formed in 8 min. After stirring for 1 h, the mixture was vacuum filtered and the solid was washed with methylene chloride. The solvent was removed from the filtrate using a rotary evaporator to give crude solid product. Recrystallization from toluene gave *N*-chloro-amide (448 mg, 70%) as white solid. ¹H NMR (400 MHz, CDCl₃) δ 8.30 (s, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 167.51, 131.99 (t, *J* = 24.4 Hz), 131.61, 128.25 (t, *J* = 24.3 Hz), 127.29 (t, *J* = 24.5 Hz).



To a 13 × 150 mm test tube equipped with magnetic stir bar were added *N*-chlorobenzamide **1a** (31 mg, 0.2 mmol), *d*₅-*N*-chlorobenzamide **1a-d₅** (32 mg, 0.2 mmol), styrene (21 mg, 0.2 mmol), [CoCp*(CO)I₂] (10 mg, 10 mol %), KOAc (24 mg, 120 mol %) and AgOTf (10 mg, 20 mol %) under nitrogen. Into the tube was then injected 1 mL of CF₃CH₂OH via syringe. The resulting solution was stirred at rt for 2 h. The reaction mixture was concentrated in vacuo and the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 4:1). The corresponding product was obtained as an off-white solid (9 mg, 20%). The reaction mixture was subjected to ¹H NMR measurement.

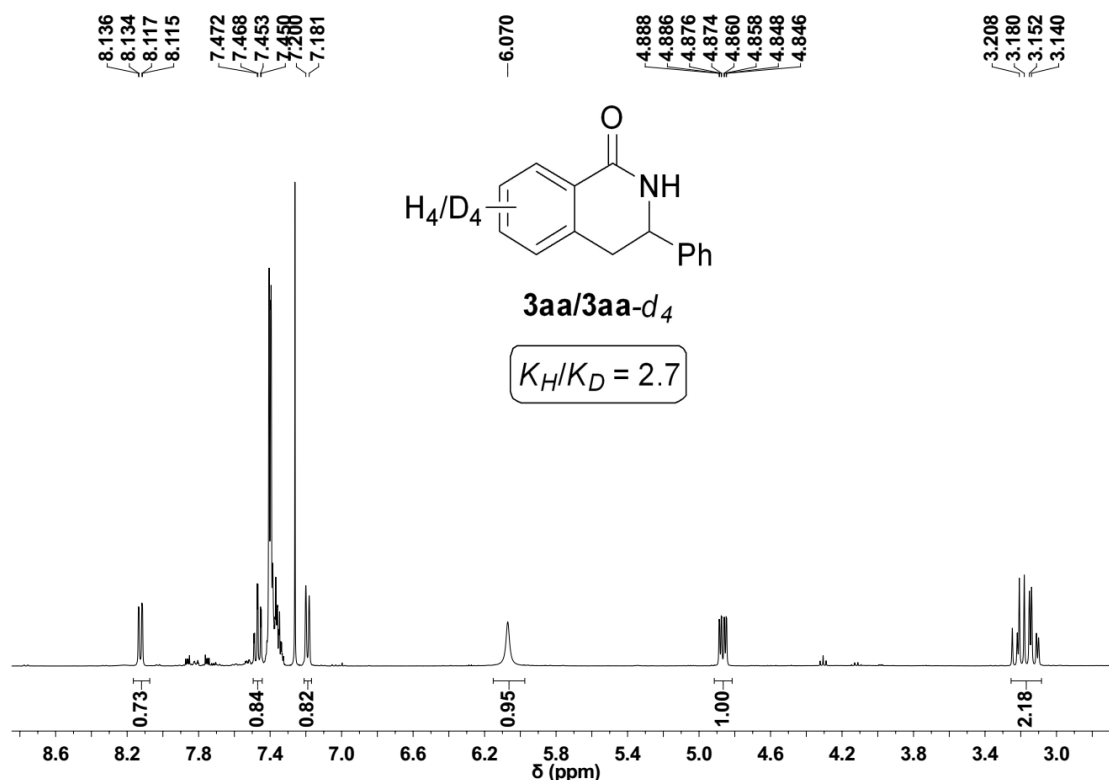
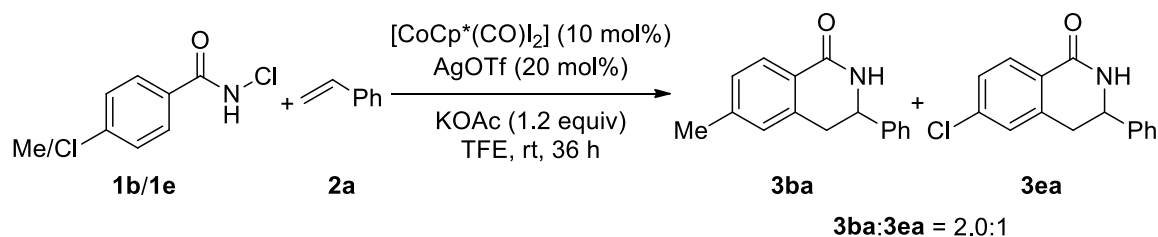


Figure S3. ¹H NMR spectrum for the mixture product of the equation above

7. Competition Experiment



To a 13 × 150 mm test tube equipped with magnetic stir bar were added *N*-chloro-4-methylbenzamide **1b** (35 mg, 0.2 mmol), *N*-chloro-4-chlorobenzamide **1e** (39 mg, 0.2 mmol), styrene (21 mg, 0.2 mmol), [CoCp*(CO)I₂] (10 mg, 10 mol %), KOAc (24 mg, 120 mol %) and AgOTf (10 mg, 20 mol %) under nitrogen. Then, 1 mL of CF₃CH₂OH was injected into the tube through a syringe. The resulting solution was stirred at rt for 36 h. The reaction mixture was concentrated in vacuo and the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 4:1). The reaction mixture was subjected to ¹H NMR measurement.

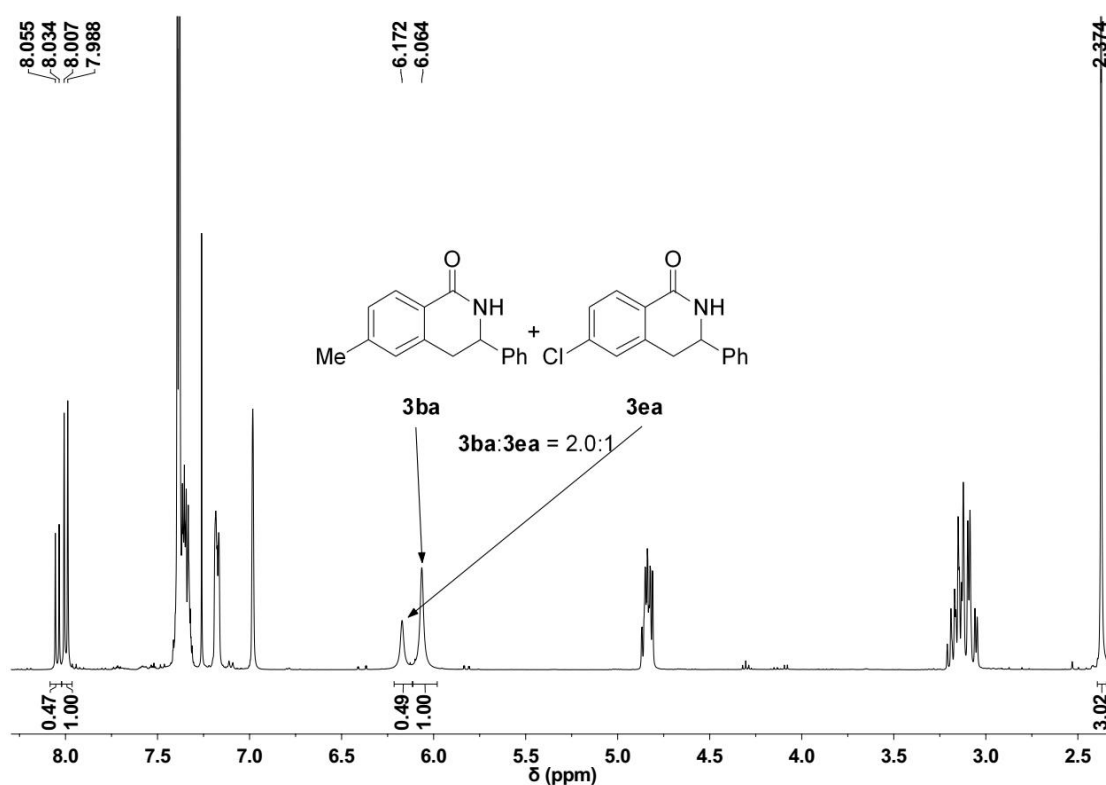
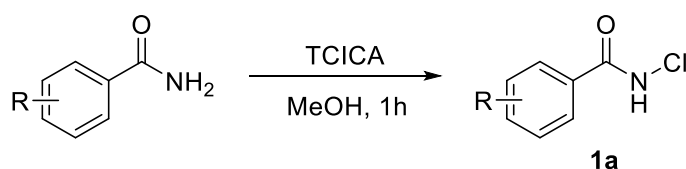


Figure S4. ¹H NMR spectrum for the products of equation above

8 Synthesis of Substrates

General procedure for the synthesis of substrates (1a-1j):

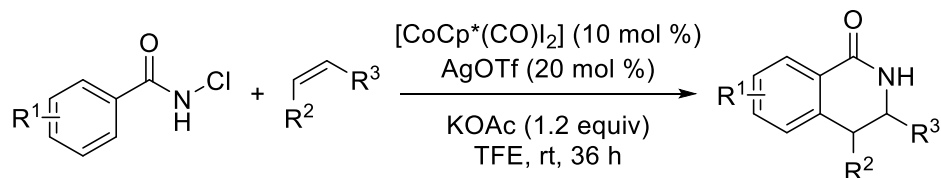


In a 100 mL, round-bottom flask were placed 20 mL of methanol and amide (20 mmol), and the mixture was allowed to stir until the solid dissolved. TCICA (trichloroisocyanuric acid) (7.3 mmol) was added and a precipitate of cyanuric acid formed in 8 min. After stirring for 1 h, the mixture was vacuum filtered and the solid was washed with methylene chloride. The solvent was removed from the filtrate using

a rotary evaporator to give crude solid product. Recrystallization from toluene gave *N*-chloroamide.

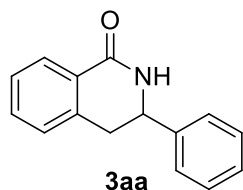
9. Preparation and Characterization Data of Products

General procedure for the synthesis of products



To a 13 × 150 mm test tube equipped with magnetic stir bar were added substituted *N*-chlorobenzamides (0.2 mmol), alkenes (0.3 mmol), [CoCp*(CO)I₂] (10 mg, 10 mol %), KOAc (24 mg, 120 mol %) and AgOTf (10 mg, 20 mol %) in CF₃CH₂OH (1 mL) under nitrogen. The resulting solution was stirred at rt for 36 h. The reaction mixture was concentrated in vacuo, the residue was dissolved in methylene chloride and purified by column chromatography (silica gel, PE:EtOAc = 3:1).

3-phenyl-3,4-dihydroisoquinolin-1(2*H*)-one (3aa)¹

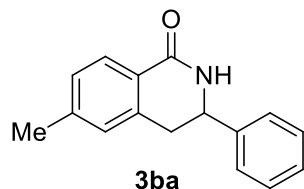


The corresponding product was obtained as an off-white solid (39 mg, 88%).

¹H NMR (400 MHz, CDCl₃) δ 8.11 (dd, *J* = 7.7, 1.2 Hz, 1H), 7.46 (td, *J* = 7.5, 1.5 Hz, 1H), 7.42 – 7.30 (m, 6H), 7.18 (d, *J* = 7.5 Hz, 1H), 6.18 (s, 1H), 6.18 (s, 1H), 4.85 (m, 1H), 3.16 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 166.36, 140.98, 137.58, 135.97, 132.53, 129.02, 128.41, 128.07, 127.35, 127.31, 126.45, 56.15, 37.47.

6-methyl-3-phenyl-3,4-dihydroisoquinolin-1(2*H*)-one (3ba)²

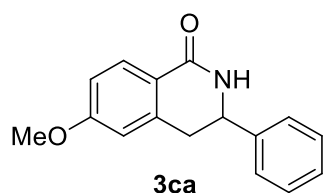


The corresponding product was obtained as an off-white solid (28 mg, 58%).

¹H NMR (400 MHz, CDCl₃) δ 8.00 (d, *J* = 7.9 Hz, 1H), 7.44 – 7.30 (m, 5H), 7.17 (d, *J* = 7.8 Hz, 1H), 6.98 (s, 1H), 6.08 (s, 1H), 4.86 – 4.79 (m, 1H), 3.11 (qd, *J* = 15.7, 7.9 Hz, 2H), 2.37 (s, 3H).

¹³C NMR (101 MHz, CDCl₃) δ 166.58, 143.16, 141.12, 137.57, 129.00, 128.36, 128.13, 127.97, 126.45, 125.74, 56.22, 37.48, 21.64.

6-methoxy-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3ca)²

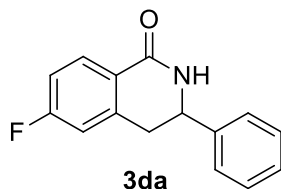


The corresponding product was obtained as an off-white solid (19 mg, 38%).

¹H NMR (400 MHz, CDCl₃) δ 8.06 (d, *J* = 8.6 Hz, 1H), 7.44 – 7.31 (m, 5H), 6.88 (dd, *J* = 8.6, 2.5 Hz, 1H), 6.66 (d, *J* = 2.4 Hz, 1H), 5.98 (s, 1H), 4.84 (dd, *J* = 10.8, 4.8 Hz, 1H), 3.84 (s, 3H), 3.12 (qd, *J* = 15.7, 7.8 Hz, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 166.46, 162.93, 141.04, 139.73, 130.24, 129.01, 128.39, 126.44, 121.14, 112.67, 112.48, 56.19, 55.44, 37.80.

6-fluoro-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3da)²

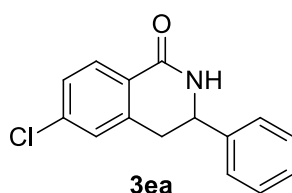


The corresponding product was obtained as an off-white solid (30 mg, 63%).

¹H NMR (400 MHz, CDCl₃) δ 8.11 (dd, *J* = 8.6, 5.8 Hz, 1H), 7.46 – 7.29 (m, 5H), 7.03 (td, *J* = 8.6, 2.5 Hz, 1H), 6.87 (dd, *J* = 8.8, 2.4 Hz, 1H), 6.26 (s, 1H), 4.86 (m, 1H), 3.15 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 165.63, 165.21 (d, *J* = 252.0 Hz), 140.60, 140.42 (d, *J* = 9.0 Hz), 130.89 (d, *J* = 10.0 Hz), 129.07, 128.52, 126.41, 124.83, 114.55 (d, *J* = 22.0 Hz), 114.23 (d, *J* = 22.0 Hz), 55.96, 37.40 (d, *J* = 1.0 Hz).

6-chloro-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3ea)²

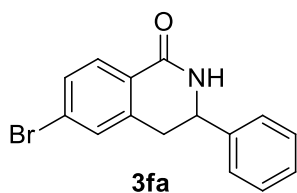


The corresponding product was obtained as an off-white solid (31 mg, 60%).

¹H NMR (400 MHz, CDCl₃) δ 8.01 (d, *J* = 8.3 Hz, 1H), 7.48 – 7.27 (m, 6H), 7.17 (d, *J* = 14.5 Hz, 1H), 6.46 – 6.18 (m, 1H), 4.87 (dd, *J* = 10.2, 5.2 Hz, 1H), 3.30 – 3.01 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 166.59, 140.55, 139.41, 138.64, 129.72, 129.10, 128.55, 127.68, 127.51, 126.41, 55.87, 37.14.

6-bromo-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3fa)²

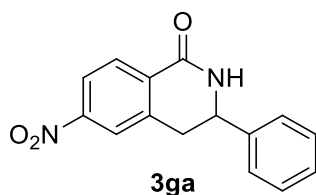


The corresponding product was obtained as an off-white solid (32 mg, 53%).

^1H NMR (400 MHz, CDCl_3) δ 7.95 (d, $J = 8.3$ Hz, 1H), 7.50 (dd, $J = 8.3, 1.8$ Hz, 1H), 7.44 – 7.28 (m, 6H), 6.29 (s, 1H), 4.85 (dd, $J = 10.3, 5.1$ Hz, 1H), 3.32 – 3.04 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.75, 140.51, 139.38, 130.66, 130.38, 129.80, 129.10, 128.56, 127.23, 126.40, 55.92, 37.06.

6-nitro-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3ga) ²

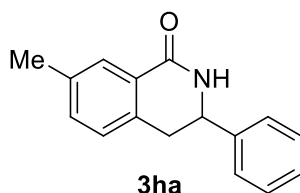


The corresponding product was obtained as an off-white solid (15 mg, 28%).

^1H NMR (400 MHz, CDCl_3) δ 8.28 (d, $J = 8.5$ Hz, 1H), 8.20 (dd, $J = 8.5, 2.2$ Hz, 1H), 8.07 (d, $J = 1.9$ Hz, 1H), 7.39 (qd, $J = 7.2, 3.3$ Hz, 5H), 6.45 (s, 1H), 4.93 (t, $J = 7.5$ Hz, 1H), 3.30 (d, $J = 7.7$ Hz, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 164.35, 150.10, 139.86, 139.08, 133.79, 129.58, 129.24, 128.83, 126.35, 122.61, 122.30, 55.73, 37.13.

7-methyl-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3ha) ²

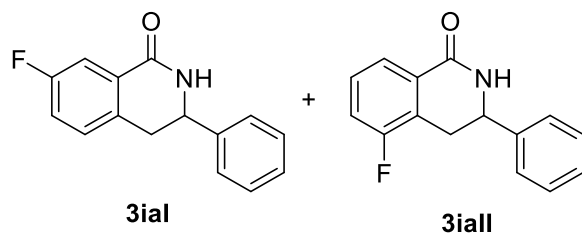


The corresponding product was obtained as an off-white solid (25 mg, 53%).

^1H NMR (400 MHz, CDCl_3) δ 7.93 (s, 1H), 7.43 – 7.30 (m, 5H), 7.29 – 7.24 (m, 1H), 7.07 (d, $J = 7.7$ Hz, 1H), 6.12 (s, 1H), 4.90 – 4.75 (m, 1H), 3.11 (m, 2H), 2.39 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.76, 141.08, 137.08, 134.75, 134.60, 133.32, 129.00, 128.43, 128.37, 128.19, 127.28, 126.45, 56.28, 37.11, 21.08.

7-fluoro-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3ia)



3ia (3ial:3iall = 1.4:1)

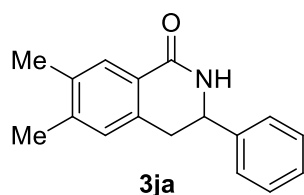
The corresponding product was obtained as an off-white solid (29 mg, 59%).

^1H NMR (400 MHz, CDCl_3) δ 7.92 (d, J = 7.3 Hz, 1H), 7.78 (dd, J = 8.7, 1.6 Hz, 0.74 \times 1H), 7.44 – 7.15 (m, 0.74 \times 7H + 7H), 6.33 (s, 0.74 \times 1H), 6.29 (s, 1H), 4.92 – 4.79 (m, 0.74 \times 1H + 1H), 3.37 – 2.98 (m, 0.74 \times 2H + 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.51, 163.22, 160.78, 160.29, 157.85, 140.63, 140.57, 133.22, 130.43, 129.15, 129.10, 129.06, 128.59, 128.52, 128.14, 128.06, 126.45, 126.43, 124.71, 124.53, 123.76, 123.73, 119.69, 119.47, 119.30, 119.08, 114.84, 114.61, 56.14, 55.60, 36.68, 29.85, 29.83.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{13}\text{FNO}$ $[\text{M}+\text{H}]^+$ 242.0976, found 242.0975.

6,7-dimethyl-3-phenyl-3,4-dihydroisoquinolin-1(2H)-one (3ja)



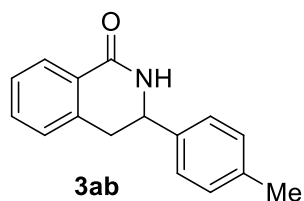
The corresponding product was obtained as an off-white solid (18 mg, 35%).

^1H NMR (400 MHz, CDCl_3) δ 7.87 (s, 1H), 7.42 – 7.30 (m, 5H), 6.95 (s, 1H), 6.01 (s, 1H), 4.83 (dd, J = 10.6, 5.0 Hz, 1H), 3.08 (qd, J = 15.6, 7.9 Hz, 2H), 2.29 (d, J = 5.2 Hz, 6H).

^{13}C NMR (101 MHz, CDCl_3) δ 141.94, 141.23, 135.75, 135.11, 128.98, 128.93, 128.60, 128.33, 126.45, 56.36, 37.03, 19.96, 19.39.

HRMS (ESI) m/z calcd. for $\text{C}_{17}\text{H}_{18}\text{NO}$ $[\text{M}+\text{H}]^+$ 252.1383, found 252.1383.

3-(p-tolyl)-3,4-dihydroisoquinolin-1(2H)-one (3ab)²

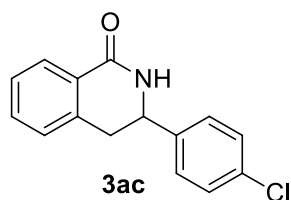


The corresponding product was obtained as light yellow solid (37 mg, 77%).

^1H NMR (400 MHz, CDCl_3) δ 8.10 (dd, J = 7.7, 1.0 Hz, 1H), 7.45 (td, J = 7.5, 1.4 Hz, 1H), 7.36 (t, J = 7.5 Hz, 1H), 7.27 (d, J = 11.9 Hz, 2H), 7.18 (dd, J = 7.4, 4.8 Hz, 3H), 6.19 (s, 1H), 4.81 (dd, J = 11.0, 4.7 Hz, 1H), 3.13 (m, 2H), 2.35 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.44, 138.20, 137.97, 137.72, 132.50, 129.65, 128.41, 128.06, 127.36, 127.26, 126.36, 55.88, 37.50, 21.13.

3-(4-chlorophenyl)-3,4-dihydroisoquinolin-1(2H)-one (3ac)³

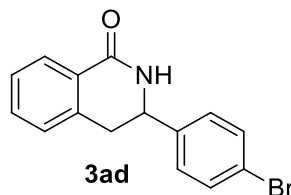


The corresponding product was obtained as light yellow solid (36 mg, 69%).

¹H NMR (400 MHz, CDCl₃) δ 8.16 – 8.03 (m, 1H), 7.45 (td, *J* = 7.5, 1.3 Hz, 1H), 7.41 – 7.28 (m, 5H), 7.16 (d, *J* = 7.4 Hz, 1H), 6.57 (s, 1H), 4.84 (dd, *J* = 8.5, 6.6 Hz, 1H), 3.22 – 3.09 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 166.48, 139.54, 137.16, 134.06, 132.65, 129.13, 128.06, 127.84, 127.40, 55.34, 37.21.

3-(4-bromophenyl)-3,4-dihydroisoquinolin-1(2H)-one (3ad)⁴

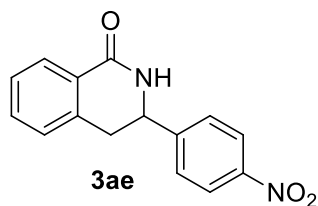


The corresponding product was obtained as light yellow solid (42 mg, 69%).

¹H NMR (400 MHz, CDCl₃) δ 8.08 (dd, *J* = 7.7, 1.1 Hz, 1H), 7.51 – 7.42 (m, 3H), 7.37 (t, *J* = 7.3 Hz, 1H), 7.30 – 7.23 (m, 2H), 7.16 (d, *J* = 7.3 Hz, 1H), 6.49 (s, 1H), 4.83 (dd, *J* = 8.4, 6.5 Hz, 1H), 3.23 – 3.03 (m, 2H).

¹³C NMR (101 MHz, CDCl₃) δ 166.41, 140.08, 137.11, 132.64, 132.09, 128.25, 128.16, 128.07, 127.42, 127.39, 122.16, 55.41, 37.18.

3-(4-nitrophenyl)-3,4-dihydroisoquinolin-1(2H)-one (3ae)⁵

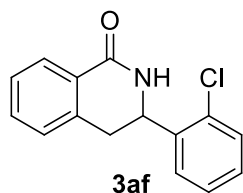


The corresponding product was obtained as light yellow solid (39 mg, 72%).

¹H NMR (400 MHz, DMSO) δ 8.51 (d, *J* = 2.6 Hz, 1H), 8.24 – 8.16 (m, 2H), 7.91 (dd, *J* = 7.7, 1.2 Hz, 1H), 7.62 (d, *J* = 8.7 Hz, 2H), 7.45 (td, *J* = 7.4, 1.4 Hz, 1H), 7.39 – 7.32 (m, 1H), 7.24 (d, *J* = 7.4 Hz, 1H), 5.02 (td, *J* = 6.1, 3.0 Hz, 1H), 3.44 – 3.37 (m, 1H), 3.14 (dd, *J* = 16.0, 6.6 Hz, 1H).

¹³C NMR (101 MHz, DMSO) δ 164.69, 149.95, 146.70, 136.73, 132.15, 128.77, 127.72, 126.99, 126.90, 123.48, 53.00, 35.10.

3-(2-chlorophenyl)-3,4-dihydroisoquinolin-1(2H)-one (3af)



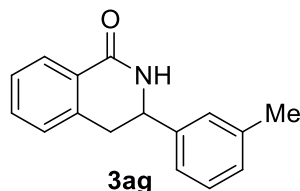
The corresponding product was obtained as light yellow solid (25 mg, 49%).

^1H NMR (400 MHz, CDCl_3) δ 8.13 (dd, $J = 7.7, 1.1$ Hz, 1H), 7.50 – 7.43 (m, 2H), 7.43 – 7.33 (m, 2H), 7.31 – 7.22 (m, 2H), 7.18 (d, $J = 7.5$ Hz, 1H), 6.26 (s, 1H), 5.36 (m, 1H), 3.36 (dd, $J = 15.8, 5.2$ Hz, 1H), 3.12 (dd, $J = 15.8, 9.0$ Hz, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.61, 138.26, 137.00, 132.74, 130.03, 129.27, 128.05, 127.60, 127.50, 127.40, 127.28, 52.11, 34.77.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{13}\text{ClNO}$ $[\text{M}+\text{H}]^+$ 258.0680, found 258.0680.

3-(*m*-tolyl)-3,4-dihydroisoquinolin-1(2*H*)-one (3ag)²

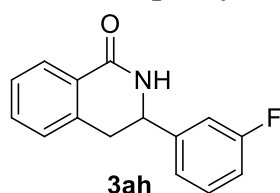


The corresponding product was obtained as an off-white solid (37 mg, 77%).

^1H NMR (400 MHz, DMSO) δ 8.27 (d, $J = 1.9$ Hz, 1H), 7.90 (dd, $J = 7.7, 1.2$ Hz, 1H), 7.45 (td, $J = 7.4, 1.5$ Hz, 1H), 7.34 (td, $J = 7.6, 1.1$ Hz, 1H), 7.26 – 7.14 (m, 3H), 7.08 (dd, $J = 16.8, 7.6$ Hz, 2H), 5.05 – 4.57 (m, 1H), 3.28 (dd, $J = 15.9, 5.4$ Hz, 1H), 3.08 (dd, $J = 15.9, 7.4$ Hz, 1H), 2.27 (s, 3H).

^{13}C NMR (101 MHz, DMSO) δ 165.30, 142.63, 137.88, 137.86, 132.46, 129.44, 128.70, 128.39, 128.18, 127.51, 127.32, 127.28, 123.91, 54.07, 36.25, 21.55.

3-(3-fluorophenyl)-3,4-dihydroisoquinolin-1(2*H*)-one (3ah)



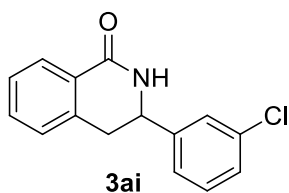
The corresponding product was obtained as an off-white solid (29 mg, 59%).

^1H NMR (400 MHz, CDCl_3) δ 8.10 (d, $J = 7.7$ Hz, 1H), 7.46 (m, 1H), 7.42 – 7.31 (m, 2H), 7.18 (d, $J = 7.6$ Hz, 2H), 7.11 (dd, $J = 9.6, 1.7$ Hz, 1H), 7.03 (dd, $J = 11.7, 5.0$ Hz, 1H), 6.50 (s, 1H), 4.94 – 4.80 (m, 1H), 3.26 – 3.10 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.44, 164.24, 161.78, 143.61, 143.54, 137.12, 132.69, 130.67, 130.58, 128.08, 127.44, 127.39, 122.07, 122.05, 115.40, 115.19, 113.64, 113.42, 55.53, 37.16.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{13}\text{FNO}$ $[\text{M}+\text{H}]^+$ 242.0976, found 242.0974.

3-(3-chlorophenyl)-3,4-dihydroisoquinolin-1(2*H*)-one (3ai)



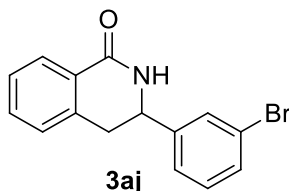
The corresponding product was obtained as light yellow solid (36 mg, 69%).

^1H NMR (400 MHz, DMSO) δ 8.39 (d, $J = 2.2$ Hz, 1H), 7.90 (dd, $J = 7.7, 1.2$ Hz, 1H), 7.46 (td, $J = 7.4, 1.4$ Hz, 1H), 7.42 – 7.22 (m, 6H), 4.89 – 4.81 (m, 1H), 3.31 (dd, $J = 16.0, 5.5$ Hz, 1H), 3.12 (dd, $J = 15.9, 7.1$ Hz, 1H).

^{13}C NMR (101 MHz, DMSO) δ 164.71, 144.76, 137.08, 132.98, 132.08, 130.21, 128.83, 127.73, 127.23, 126.89, 126.85, 126.42, 125.10, 52.99, 35.28.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{13}\text{ClNO}$ $[\text{M}+\text{H}]^+$ 258.0680, found 258.0680.

3-(3-bromophenyl)-3,4-dihydroisoquinolin-1(2H)-one (3aj)



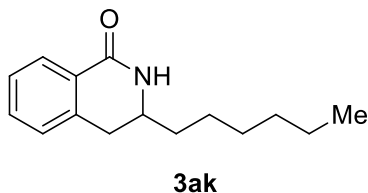
The corresponding product was obtained as an off-white solid (37 mg, 61%).

^1H NMR (400 MHz, CDCl_3) δ 8.09 (d, $J = 7.7$ Hz, 1H), 7.55 (s, 1H), 7.46 (t, $J = 7.4$ Hz, 2H), 7.37 (t, $J = 7.5$ Hz, 1H), 7.32 (d, $J = 7.7$ Hz, 1H), 7.24 (t, $J = 7.9$ Hz, 1H), 7.17 (d, $J = 7.5$ Hz, 1H), 6.60 (s, 1H), 4.83 (t, $J = 7.6$ Hz, 1H), 3.16 (d, $J = 7.7$ Hz, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.52, 143.27, 137.07, 132.72, 131.47, 130.58, 129.66, 128.10, 127.47, 127.38, 125.06, 122.99, 55.46, 37.18.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{13}\text{BrNO}$ $[\text{M}+\text{H}]^+$ 302.0175, found 302.0173.

3-hexyl-3,4-dihydroisoquinolin-1(2H)-one (3ak)



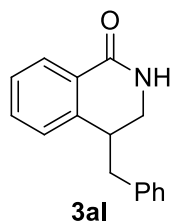
The corresponding product was obtained as an off-white solid (25 mg, 54%).

^1H NMR (400 MHz, DMSO) δ 7.93 (s, 1H), 7.86 – 7.80 (m, 1H), 7.46 (td, $J = 7.4, 1.4$ Hz, 1H), 7.37 – 7.26 (m, 2H), 3.57 – 3.47 (m, 1H), 3.00 (dd, $J = 15.7, 4.6$ Hz, 1H), 2.70 (dd, $J = 15.7, 9.0$ Hz, 1H), 1.62 – 1.48 (m, 1H), 1.47 – 1.23 (m, 9H), 0.86 (t, $J = 6.8$ Hz, 3H).

^{13}C NMR (101 MHz, DMSO) δ 164.88, 138.73, 132.21, 129.54, 128.22, 127.32, 127.08, 50.73, 35.07, 33.46, 31.68, 29.14, 25.33, 22.49, 14.42.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{22}\text{NO}$ $[\text{M}+\text{H}]^+$ 232.1696, found 232.1695.

3-benzyl-3,4-dihydroisoquinolin-1(2H)-one (3al)¹

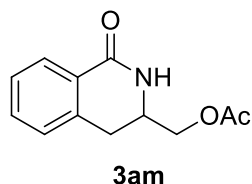


The corresponding product was obtained as an off-white solid (31 mg, 64%).

^1H NMR (400 MHz, CDCl_3) δ 8.06 (dd, $J = 7.7, 1.1$ Hz, 1H), 7.45 (td, $J = 7.5, 1.4$ Hz, 1H), 7.38 – 7.31 (m, 3H), 7.28 (dt, $J = 4.9, 2.0$ Hz, 1H), 7.23 – 7.16 (m, 3H), 6.03 (s, 1H), 4.15 – 3.77 (m, 1H), 3.32 – 2.58 (m, 4H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.12, 137.73, 136.60, 132.41, 129.21, 128.98, 128.64, 128.04, 127.54, 127.16, 52.48, 41.79, 34.15.

(1-oxo-1,2,3,4-tetrahydroisoquinolin-3-yl)methyl acetate (3am)



The corresponding product was obtained as an off-white solid (33 mg, 75%).

^1H NMR (400 MHz, CDCl_3) δ 8.06 (dd, $J = 7.7, 0.9$ Hz, 1H), 7.46 (td, $J = 7.5, 1.3$ Hz, 1H), 7.36 (t, $J = 7.4$ Hz, 1H), 7.22 (d, $J = 7.5$ Hz, 1H), 6.74 (s, 1H), 4.26 (dd, $J = 11.1, 4.4$ Hz, 1H), 4.10 (dd, $J = 11.1, 7.8$ Hz, 1H), 4.05 – 3.95 (m, 1H), 3.05 (dd, $J = 15.7, 4.9$ Hz, 1H), 2.91 (dd, $J = 15.7, 9.0$ Hz, 1H), 2.08 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 170.72, 166.07, 136.74, 132.55, 131.94, 128.60, 128.49, 128.05, 127.63, 127.35, 65.95, 49.93, 30.33, 20.74.

HRMS (ESI) m/z calcd. for $\text{C}_{12}\text{H}_{14}\text{NO}_3$ $[\text{M}+\text{H}]^+$ 220.0968, found 220.0967.

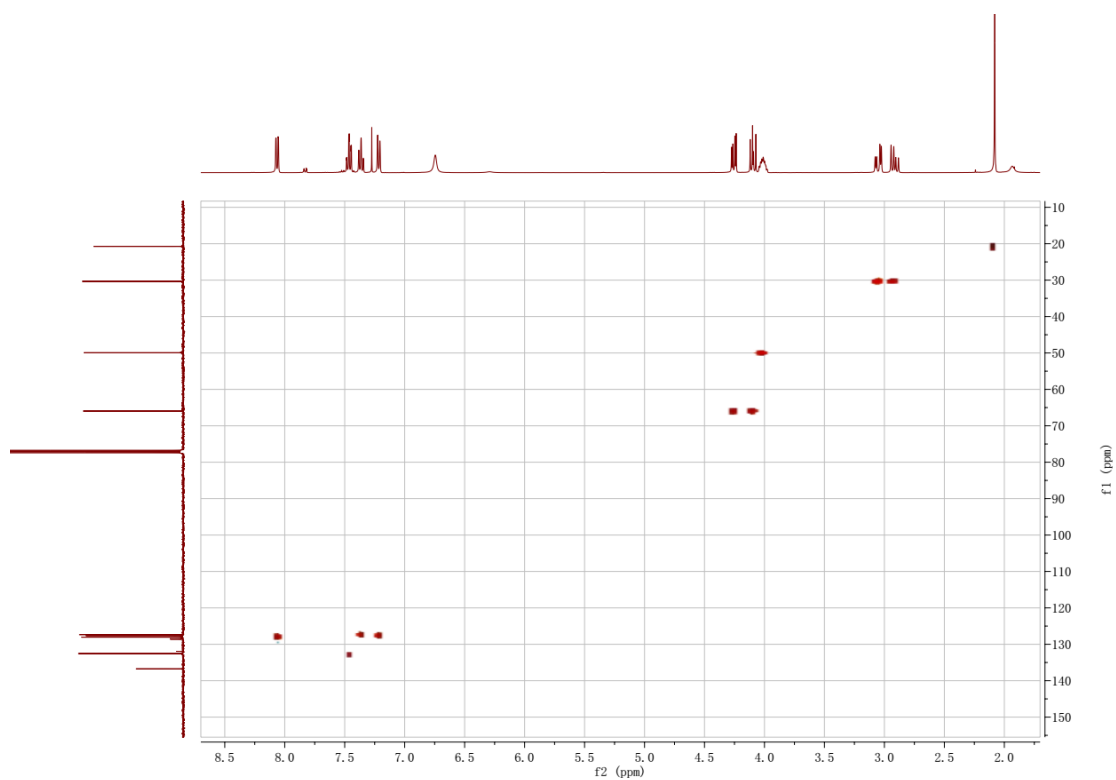


Figure S5. The ^1H – ^{13}C HSQC (400 MHz, CDCl_3) spectrum of **3am**

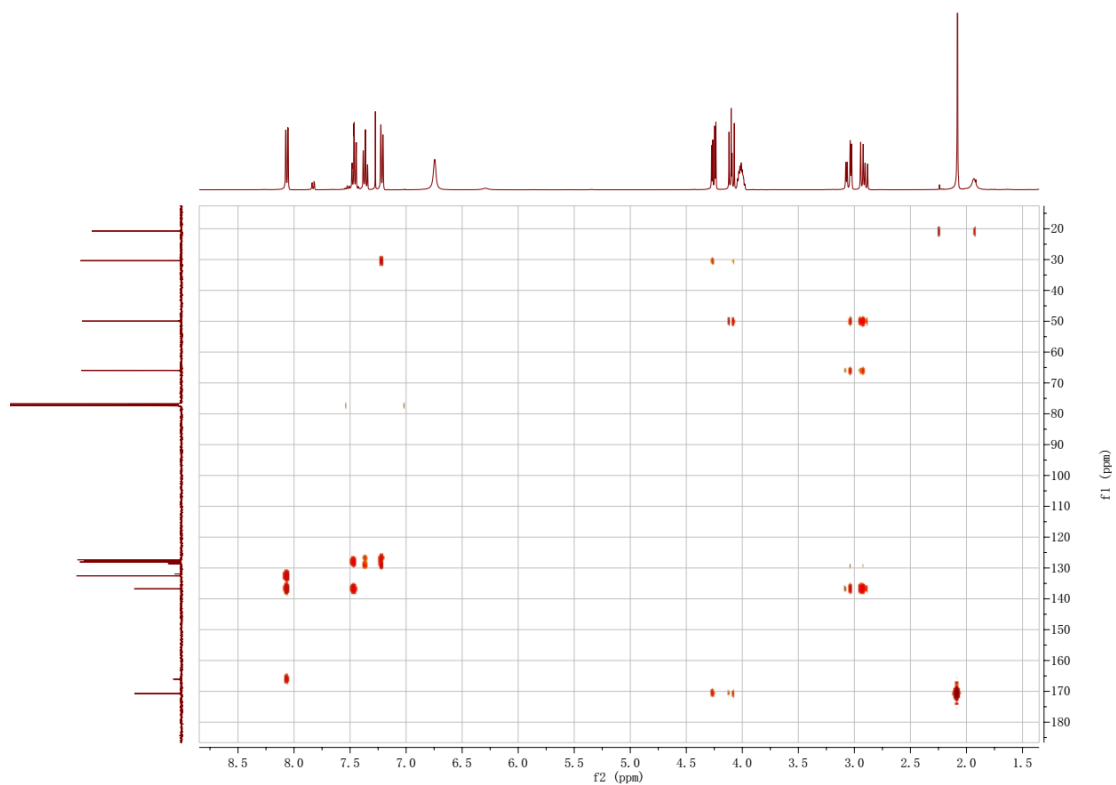
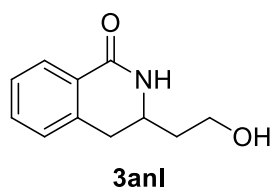


Figure S6. The ^1H – ^{13}C HMBC (400 MHz, CDCl_3) spectrum of **3am**

3-(2-hydroxyethyl)-3,4-dihydroisoquinolin-1(2*H*)-one (3anI)



The corresponding product was obtained as an off-white solid (22 mg, 58%).

^1H NMR (400 MHz, CDCl_3) δ 8.05 (dd, $J = 7.7, 1.3$ Hz, 1H), 7.47 (td, $J = 7.5, 1.5$ Hz, 1H), 7.36 (td, $J = 7.6, 1.2$ Hz, 1H), 7.29 – 7.25 (m, 1H), 6.64 (s, 1H), 3.81 – 3.70 (m, 2H), 3.65 (ddd, $J = 10.8, 6.8, 5.9$ Hz, 1H), 3.44 (ddd, $J = 12.6, 4.8, 2.8$ Hz, 1H), 3.13 (dt, $J = 7.3, 3.7$ Hz, 1H), 2.12 (s, 1H), 1.93 (dtd, $J = 7.4, 5.9, 3.6$ Hz, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.35, 142.59, 132.32, 128.19, 128.04, 127.25, 60.02, 44.26, 35.83, 34.38.

HRMS (ESI) m/z calcd. for $\text{C}_{11}\text{H}_{14}\text{NO}_2$ $[\text{M}+\text{H}]^+$ 192.1019, found 192.1018.

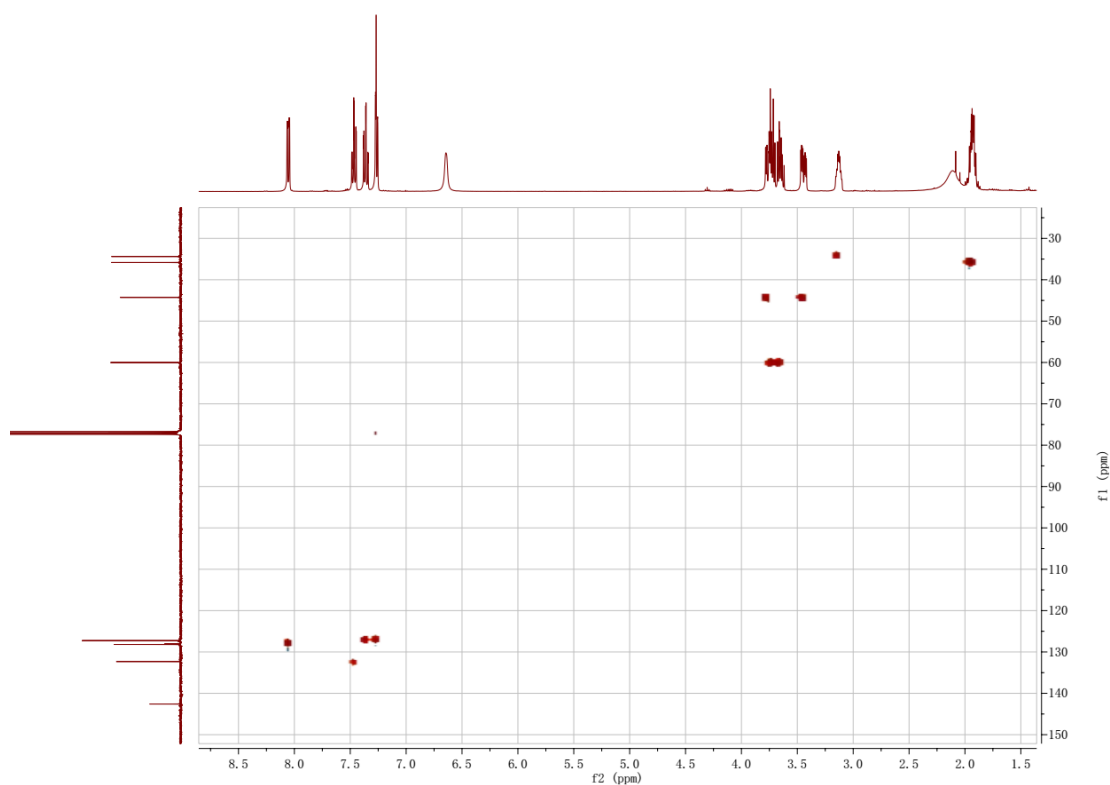


Figure S7. The ^1H – ^{13}C HSQC (400 MHz, CDCl_3) spectrum of **3anI**

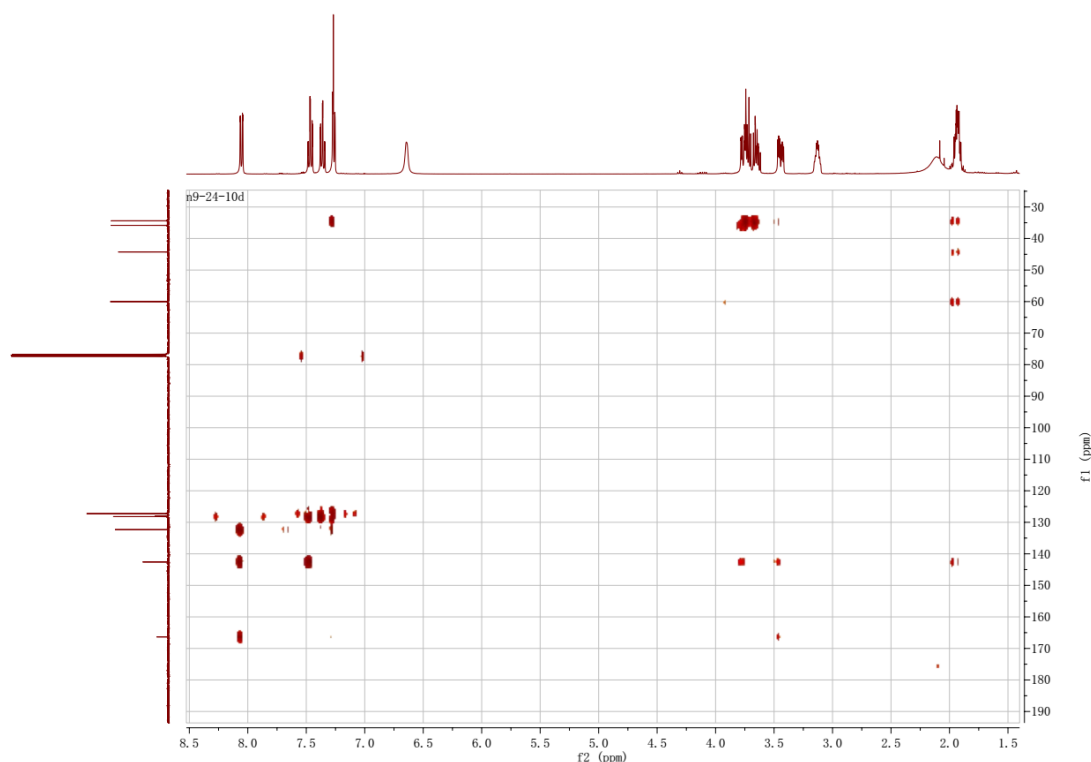
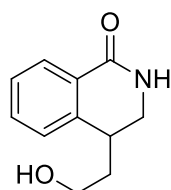


Figure S8. The ^1H – ^{13}C HMBC (400 MHz, CDCl_3) spectrum of **3anI**

4-(2-hydroxyethyl)-3,4-dihydroisoquinolin-1(2H)-one (3anII)



3anII

The corresponding product was obtained as an off-white solid (14 mg, 36%).

^1H NMR (400 MHz, CDCl_3) δ 8.06 – 7.98 (m, 1H), 7.61 (s, 1H), 7.44 (td, J = 7.5, 1.3 Hz, 1H), 7.33 (t, J = 7.5 Hz, 1H), 7.19 (d, J = 7.5 Hz, 1H), 4.04 – 3.91 (m, 2H), 3.88 – 3.80 (m, 1H), 3.05 (s, 1H), 2.98 – 2.84 (m, 2H), 1.92 (ddd, J = 13.3, 9.7, 4.6 Hz, 1H), 1.84 – 1.73 (m, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.63, 138.15, 132.38, 128.28, 127.86, 127.45, 127.09, 60.64, 51.10, 36.86, 34.67.

HRMS (ESI) m/z calcd. for $\text{C}_{11}\text{H}_{14}\text{NO}_2$ $[\text{M}+\text{H}]^+$ 192.1019, found 192.1018.

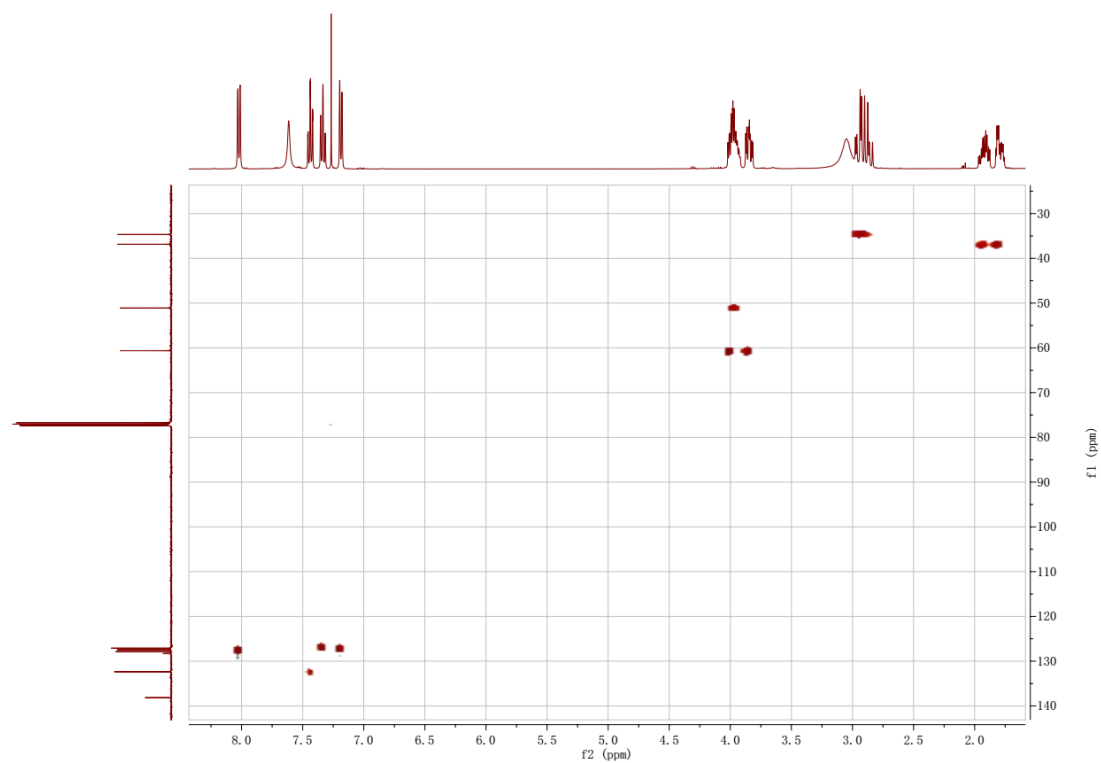


Figure S9. The ^1H – ^{13}C HSQC (400 MHz, CDCl_3) spectrum of **3anII**

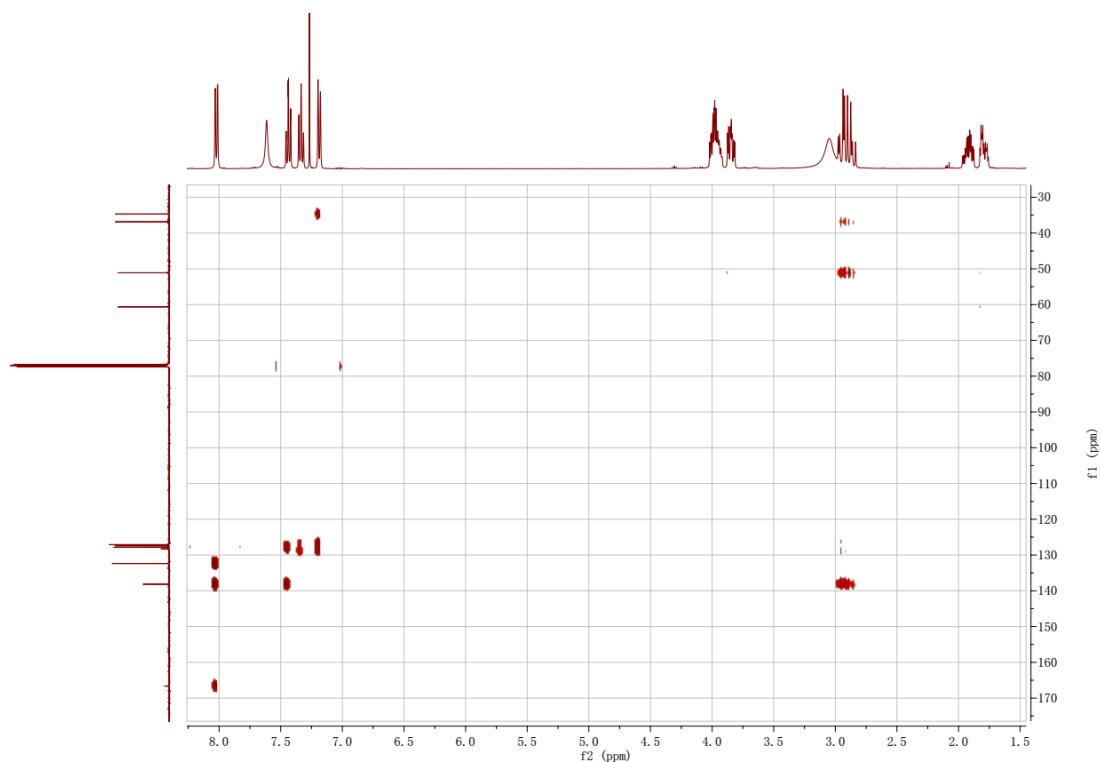
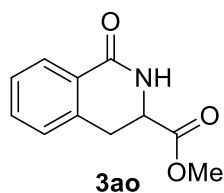


Figure S10. The ^1H – ^{13}C HMBC (400 MHz, CDCl_3) spectrum of **3anII**

methyl 1-oxo-1,2,3,4-tetrahydroisoquinoline-3-carboxylate (3ao)¹

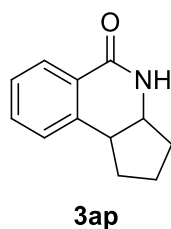


The corresponding product was obtained as an off-white solid (32 mg, 78%).

^1H NMR (400 MHz, CDCl_3) δ 8.08 (dd, $J = 7.7, 1.1$ Hz, 1H), 7.51 – 7.44 (m, 1H), 7.38 (t, $J = 7.5$ Hz, 1H), 7.24 (d, $J = 7.5$ Hz, 1H), 6.67 (s, 1H), 4.42 (m, 1H), 3.79 (s, 3H), 3.34 (dd, $J = 15.7, 5.2$ Hz, 1H), 3.22 (dd, $J = 15.7, 9.7$ Hz, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 170.89, 165.26, 136.21, 132.59, 128.42, 128.19, 127.61, 127.47, 53.08, 52.92, 31.15.

2,3,3a,4-tetrahydro-1H-cyclopenta[c]isoquinolin-5(9bH)-one (**3ap**)²

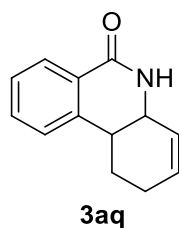


The corresponding product was obtained as an off-white solid (22 mg, 59%).

^1H NMR (400 MHz, CDCl_3) δ 8.09 (dd, $J = 7.7, 1.2$ Hz, 1H), 7.45 (td, $J = 7.5, 1.4$ Hz, 1H), 7.34 (td, $J = 7.6, 1.2$ Hz, 1H), 7.23 (d, $J = 7.5$ Hz, 1H), 5.74 (s, 1H), 4.21 (t, $J = 5.0$ Hz, 1H), 3.11 (td, $J = 9.2, 5.7$ Hz, 1H), 2.20 – 2.11 (m, 1H), 2.10 – 2.01 (m, 1H), 1.98 – 1.90 (m, 1H), 1.87 – 1.77 (m, 3H).

^{13}C NMR (101 MHz, DMSO) δ 164.08, 142.25, 132.28, 128.18, 127.64, 127.48, 127.00, 55.58, 42.38, 33.76, 33.41, 23.12.

1,4a,5,10b-tetrahydrophenanthridin-6(2H)-one (**3aq**)¹

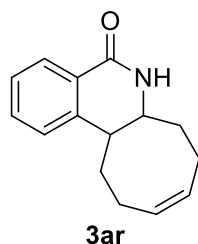


The corresponding product was obtained as an off-white solid (15 mg, 38%).

^1H NMR (400 MHz, CDCl_3) δ 8.08 (dd, $J = 7.7, 1.3$ Hz, 1H), 7.49 (td, $J = 7.5, 1.4$ Hz, 1H), 7.36 (td, $J = 7.6, 1.2$ Hz, 1H), 7.30 – 7.21 (m, 1H), 6.04 (dt, $J = 9.9, 3.7$ Hz, 1H), 5.88 (s, 1H), 5.82 – 5.75 (m, 1H), 4.28 (t, $J = 4.6$ Hz, 1H), 2.94 (dt, $J = 12.2, 3.8$ Hz, 1H), 2.31 – 2.13 (m, 2H), 2.06 – 1.85 (m, 1H), 1.76 – 1.62 (m, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 165.00, 142.79, 132.59, 132.45, 128.06, 127.50, 127.26, 127.14, 124.34, 48.00, 37.84, 25.20, 25.03.

(Z)-6,6a,7,8,12,12a-hexahydrocycloocta[c]isoquinolin-5(11H)-one (**3ar**)

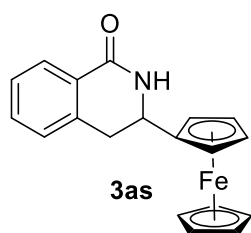


The corresponding product was obtained as an off-white solid (13 mg, 33%).

^1H NMR (400 MHz, CDCl_3) δ 8.04 (dd, $J = 7.7, 1.2$ Hz, 1H), 7.45 (td, $J = 7.5, 1.4$ Hz, 1H), 7.34 (td, $J = 7.6, 1.2$ Hz, 1H), 7.19 (d, $J = 7.5$ Hz, 1H), 6.25 (s, 1H), 5.78 – 5.53 (m, 2H), 4.08 – 3.83 (m, 1H), 3.25 (dt, $J = 11.9, 3.6$ Hz, 1H), 2.90 – 2.72 (m, 1H), 2.70 – 2.54 (m, 1H), 2.54 – 2.38 (m, 1H), 2.26 – 2.15 (m, 1H), 2.12 – 2.05 (m, 1H), 1.87 (m, 2H), 1.57 (m, 1H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.49, 144.96, 132.40, 130.37, 128.31, 128.00, 126.87, 126.67, 125.87, 55.72, 38.95, 29.71, 29.54, 27.52, 25.45.

HRMS (ESI) m/z calcd. for $\text{C}_{15}\text{H}_{18}\text{NO}$ $[\text{M}+\text{H}]^+$ 228.1383, found 228.1382.



The corresponding product was obtained as a brown solid (27 mg, 40%).

^1H NMR (400 MHz, CDCl_3) δ 8.11 (d, $J = 7.6$ Hz, 1H), 7.45 (dd, $J = 7.4, 6.6$ Hz, 1H), 7.37 (t, $J = 7.5$ Hz, 1H), 7.18 (d, $J = 7.4$ Hz, 1H), 6.28 (s, 1H), 4.52 (dd, $J = 10.3, 5.5$ Hz, 1H), 4.29 – 4.17 (m, 8H), 3.10 – 2.95 (m, 2H).

^{13}C NMR (101 MHz, CDCl_3) δ 166.01, 138.00, 134.19, 132.42, 128.56, 128.06, 127.25, 127.22, 123.51, 89.38, 68.66, 68.32, 68.31, 66.89, 65.20, 51.05, 37.65.

HRMS (ESI) m/z calcd. for $\text{C}_{19}\text{H}_{18}\text{FeNO}$ $[\text{M}+\text{H}]^+$ 332.0732, found 332.0732.

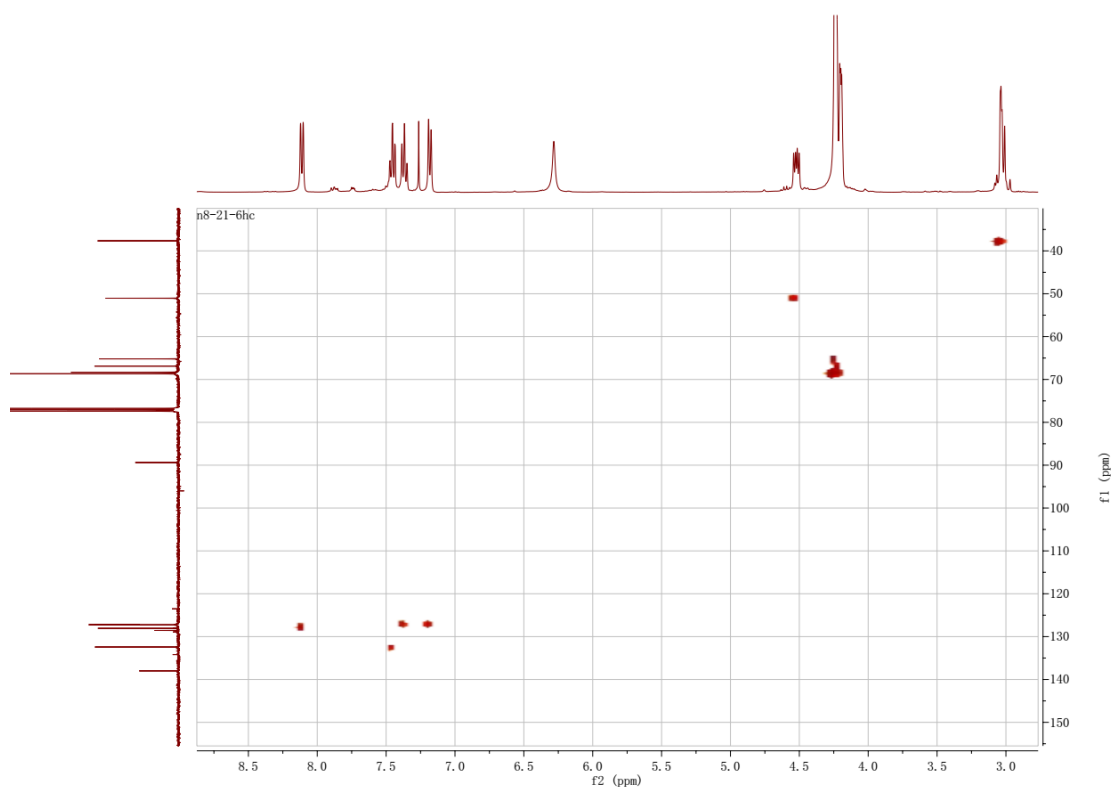


Figure S11. The ^1H – ^{13}C HSQC (400 MHz, CDCl_3) spectrum of **3as**

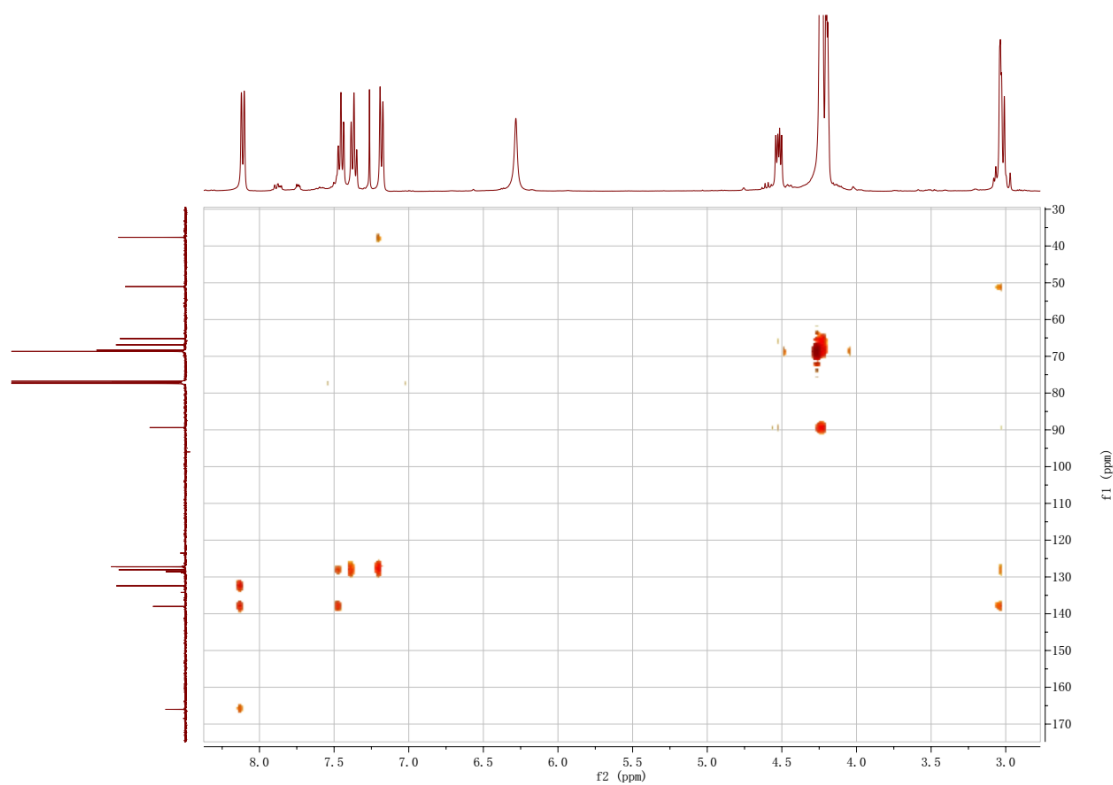
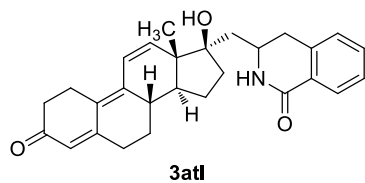


Figure S12. The ^1H – ^{13}C HMBC (400 MHz, CDCl_3) spectrum of **3as**



The corresponding product was obtained as a off-white solid (33 mg, 39%).

^1H NMR (400 MHz, CDCl_3) δ 8.01 (dd, $J = 9.4, 8.6$ Hz, 2H), 7.43 (td, $J = 7.5, 1.2$ Hz, 1H), 7.34 (t, $J = 7.5$ Hz, 1H), 7.16 (d, $J = 7.5$ Hz, 1H), 6.50 (s, 2H), 5.79 (s, 1H), 4.07 (dd, $J = 16.4, 6.7$ Hz, 1H), 3.15 (s, 1H), 2.94 – 2.79 (m, 3H), 2.72 (dd, $J = 15.5, 3.6$ Hz, 1H), 2.58 (dd, $J = 12.8, 2.0$ Hz, 2H), 2.48 (t, $J = 7.2$ Hz, 3H), 2.15 – 2.04 (m, 1H), 1.94 (m, 3H), 1.72 – 1.54 (m, 4H), 1.34 – 1.25 (m, 1H), 1.10 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 199.26, 166.12, 156.51, 141.87, 141.72, 138.35, 132.23, 128.48, 127.91, 127.15, 127.09, 124.09, 123.66, 83.20, 49.94, 48.86, 48.02, 41.58, 38.40, 36.70, 35.71, 34.27, 31.49, 27.16, 24.36, 23.16, 16.29.

HRMS (ESI) m/z calcd. for $\text{C}_{28}\text{H}_{32}\text{NO}_3$ $[\text{M}+\text{H}]^+$ 430.2377, found 430.2376.

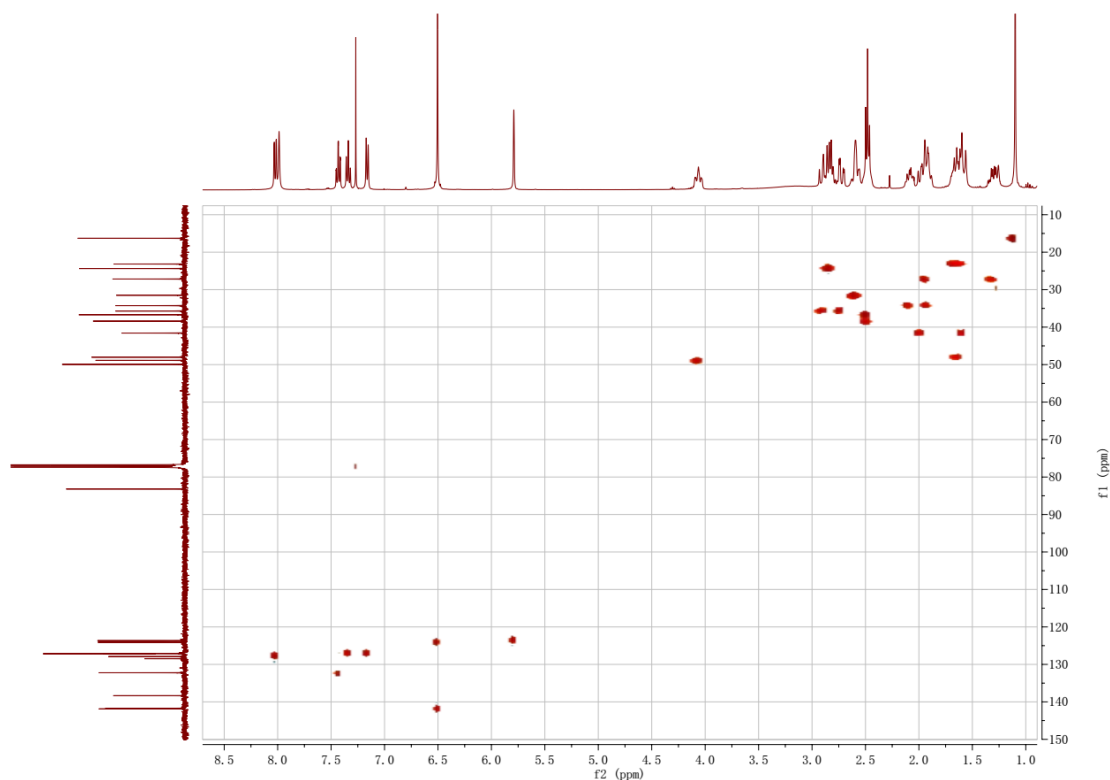
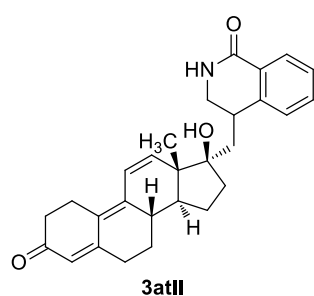


Figure S13. The ^1H – ^{13}C HSQC (400 MHz, CDCl_3) spectrum of **3atI**



Figure S14. The ^1H – ^{13}C HMBC (400 MHz, CDCl_3) spectrum of **3atlI**



The corresponding product was obtained as a off-white solid (29 mg, 33%).

^1H NMR (400 MHz, CDCl_3) δ 8.05 (dd, $J = 7.7, 1.1$ Hz, 1H), 7.73 (s, 1H), 7.44 (td, $J = 7.5, 1.4$ Hz, 1H), 7.34 (t, $J = 7.4$ Hz, 1H), 7.18 (d, $J = 7.4$ Hz, 1H), 6.42 (s, 2H), 5.78 (s, 1H), 4.16 (td, $J = 10.3, 5.6$ Hz, 1H), 2.91 (dd, $J = 15.6, 4.4$ Hz, 1H), 2.79 (dd, $J = 17.3, 9.1$ Hz, 3H), 2.55 (d, $J = 17.4$ Hz, 2H), 2.47 – 2.37 (m, 3H), 2.00 – 1.82 (m, 5H), 1.70 (dt, $J = 16.4, 7.4$ Hz, 2H), 1.54 – 1.44 (m, 1H), 1.35 – 1.26 (m, 1H), 1.07 (s, 3H).

^{13}C NMR (101 MHz, CDCl_3) δ 199.18, 166.16, 156.39, 141.43, 141.22, 138.33, 132.28, 128.45, 127.96, 127.39, 127.19, 127.04, 123.90, 123.69, 82.55, 49.62, 49.20, 47.40, 44.28, 39.60, 38.39, 36.64, 35.52, 31.43, 27.02, 24.30, 22.63, 17.03.

HRMS (ESI) m/z calcd. for $\text{C}_{28}\text{H}_{32}\text{NO}_3$ $[\text{M}+\text{H}]^+$ 430.2377, found 430.2376.

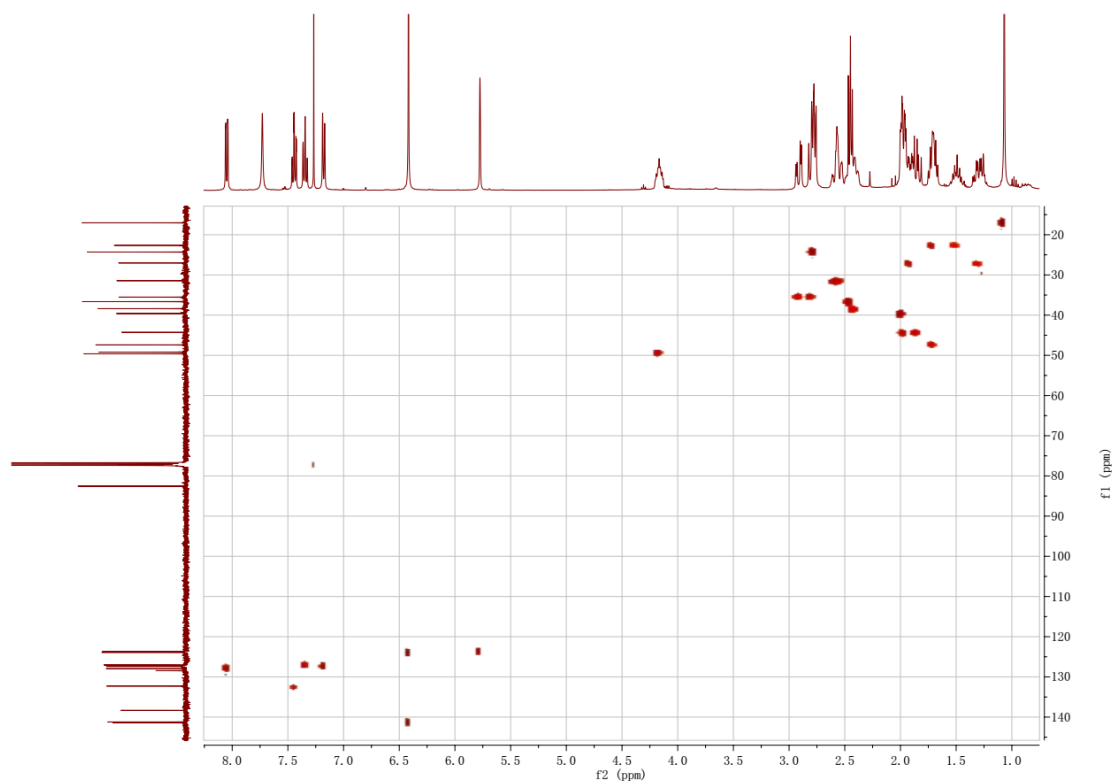


Figure S15. The ^1H – ^{13}C HSQC (400 MHz, CDCl_3) spectrum of **3atII**

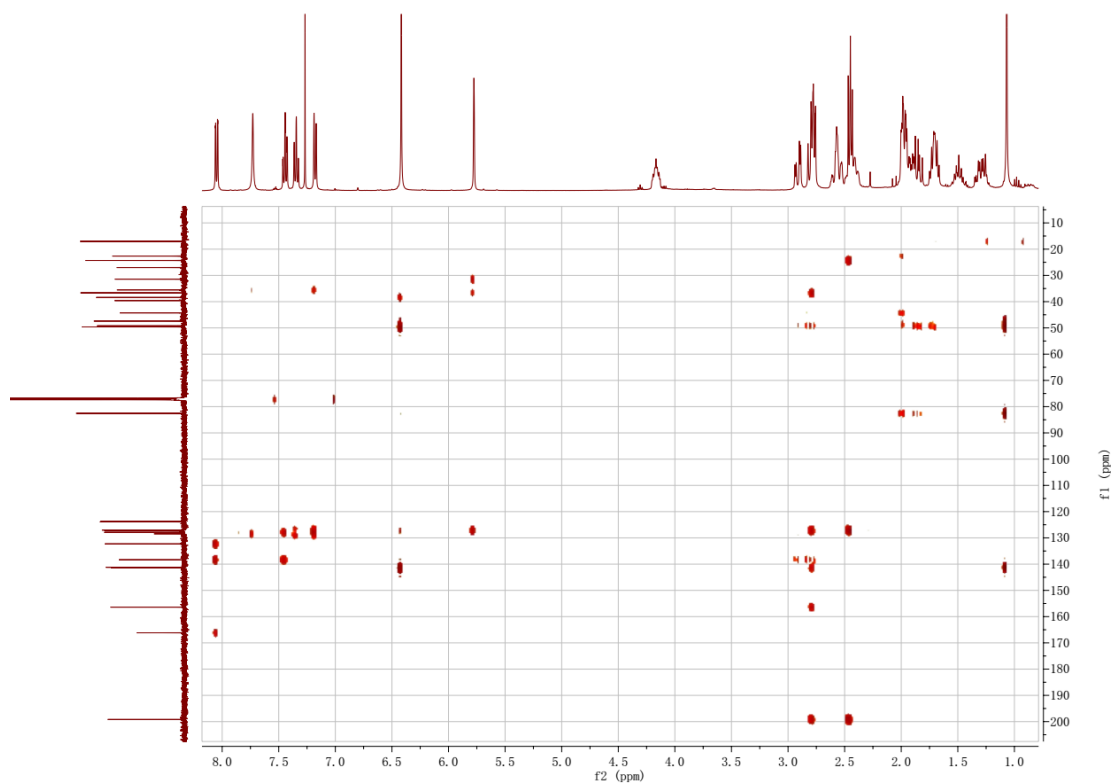
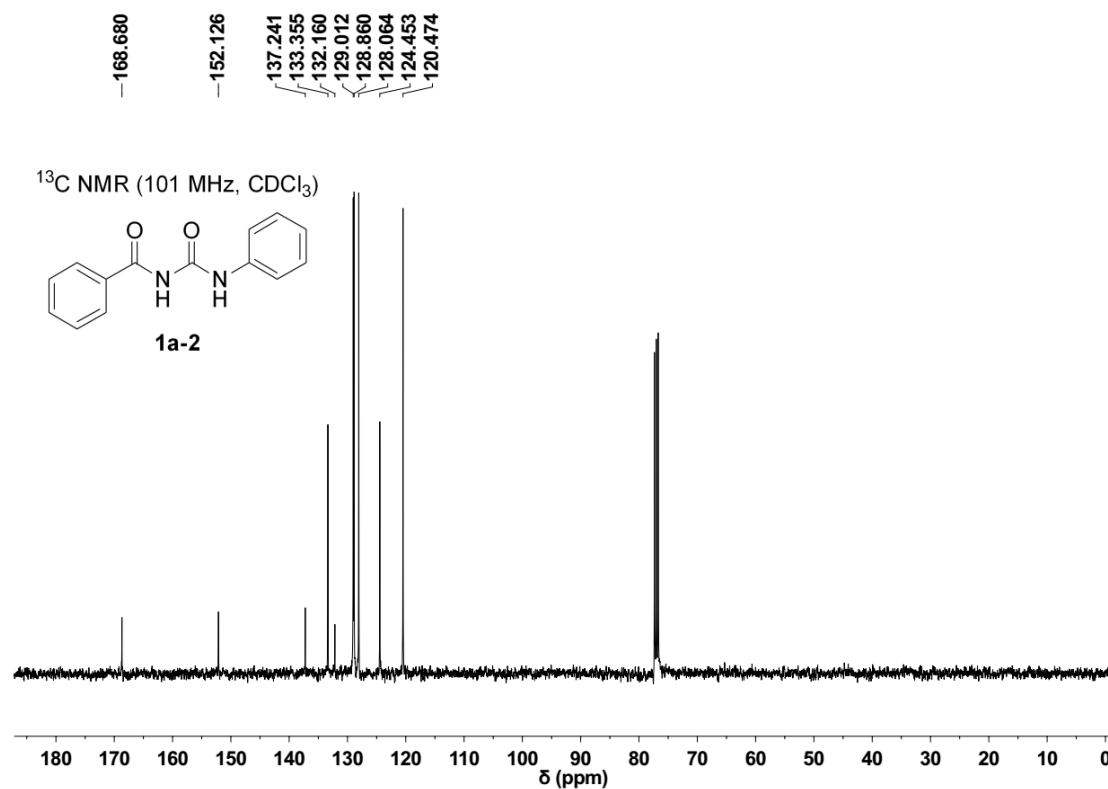
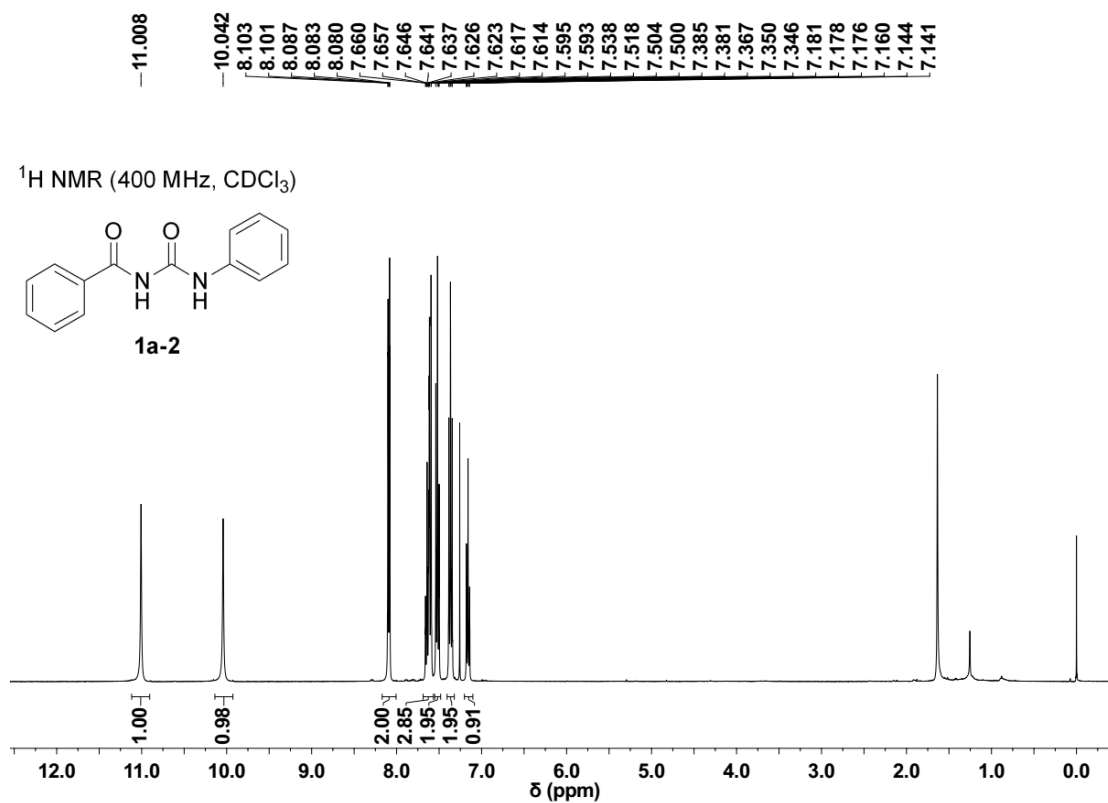


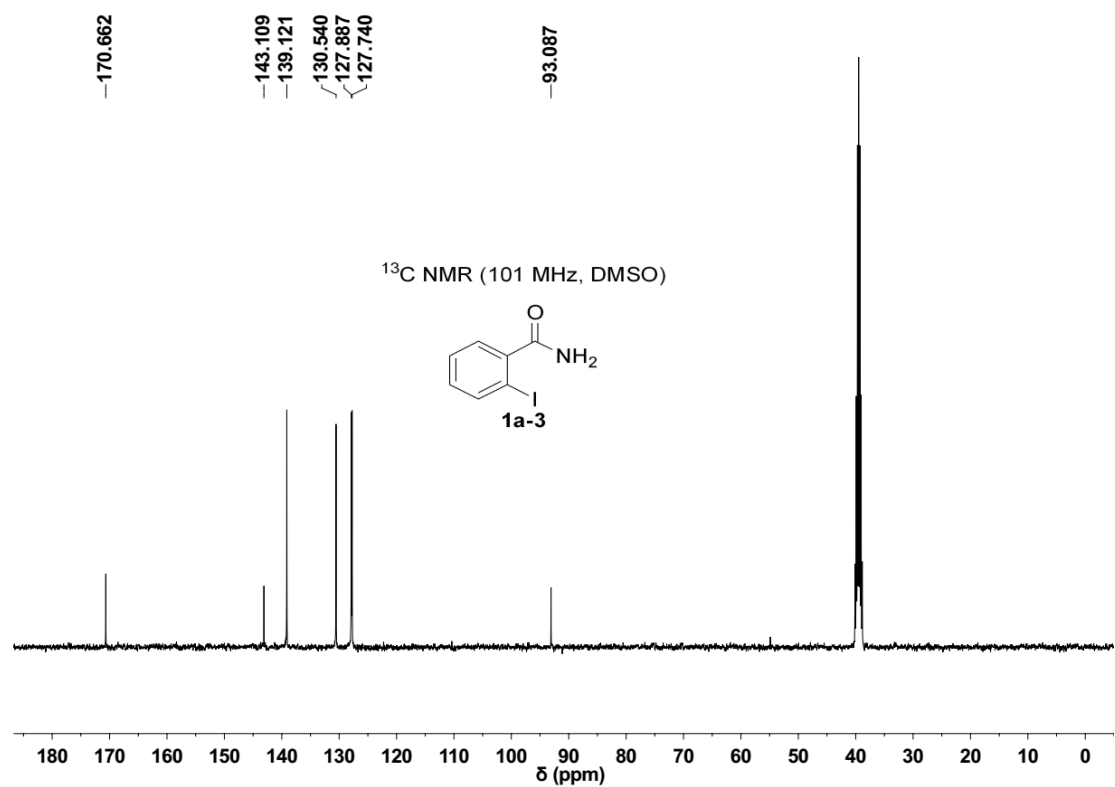
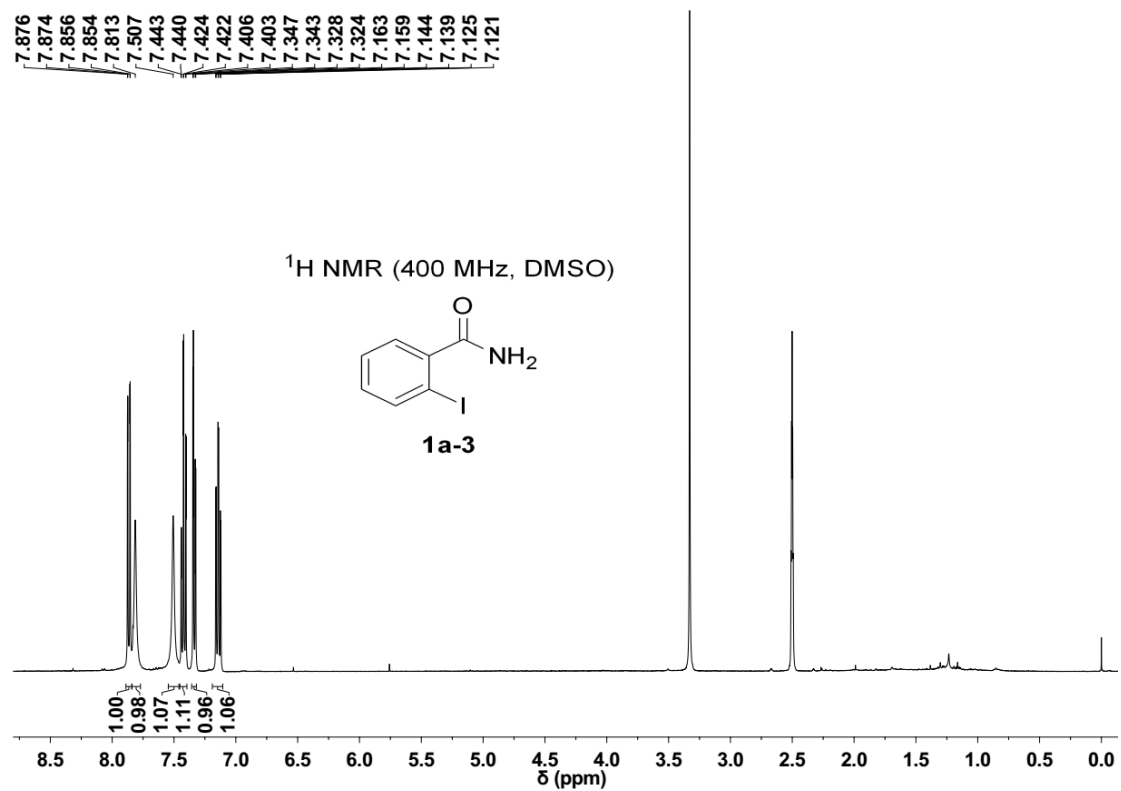
Figure S16. The ^1H – ^{13}C HMBC (400 MHz, CDCl_3) spectrum of **3atII**

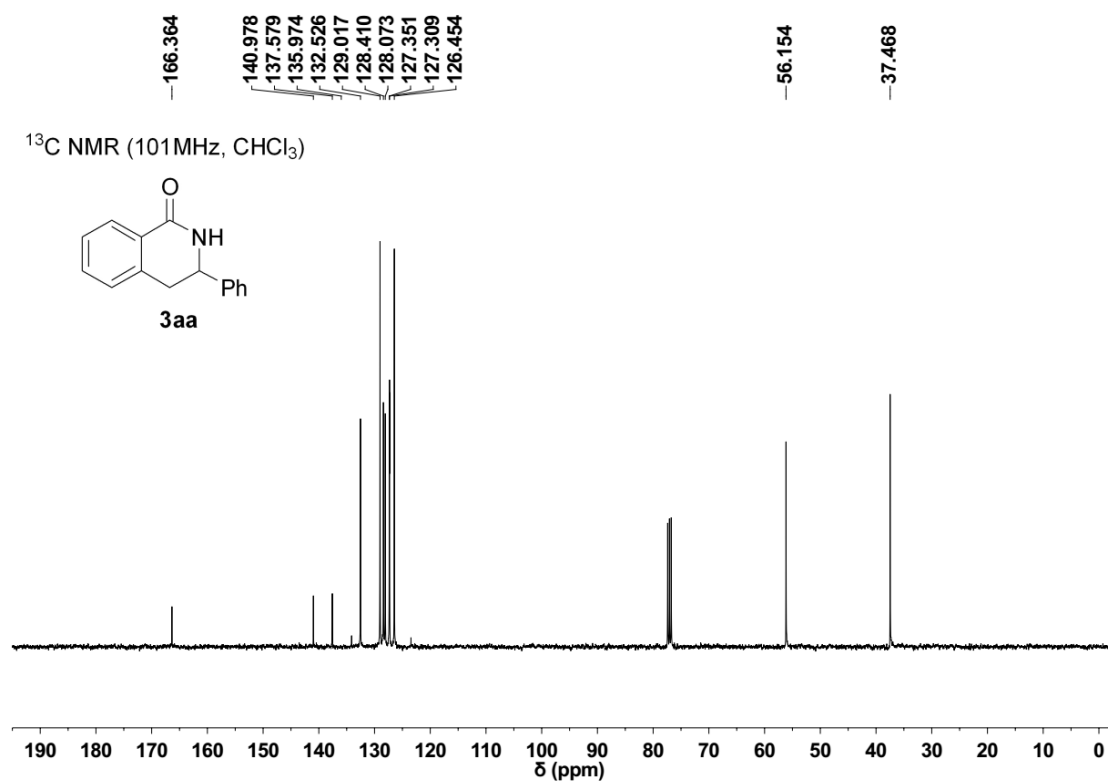
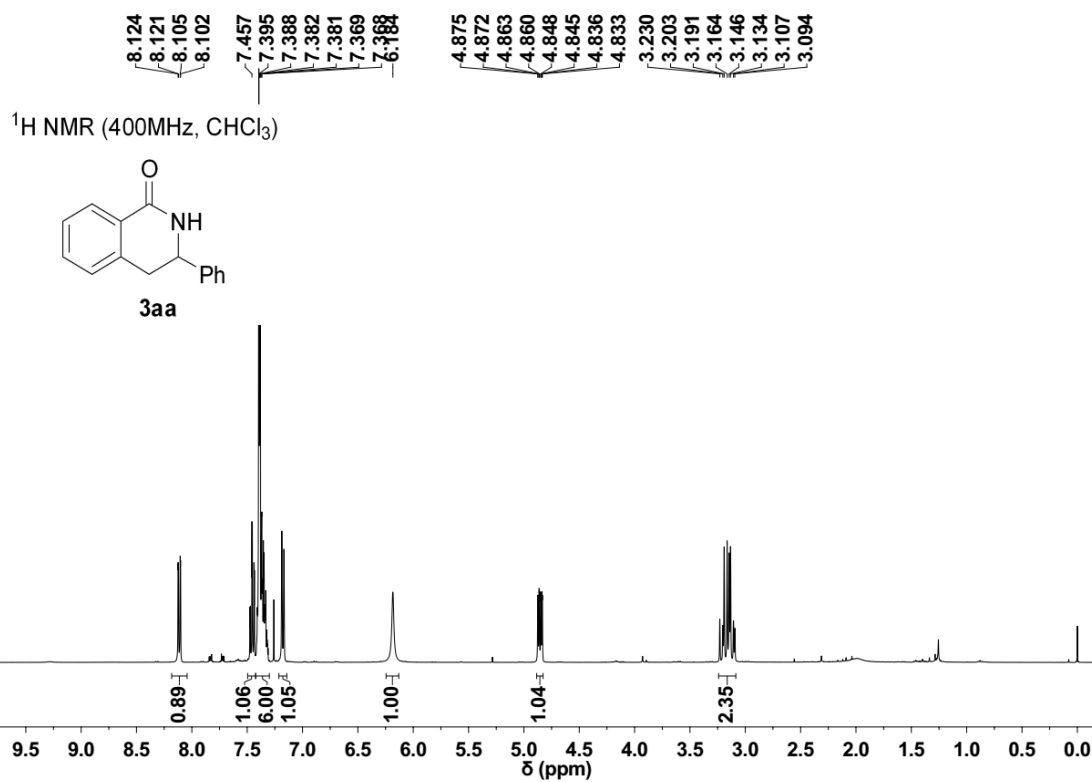
10. References

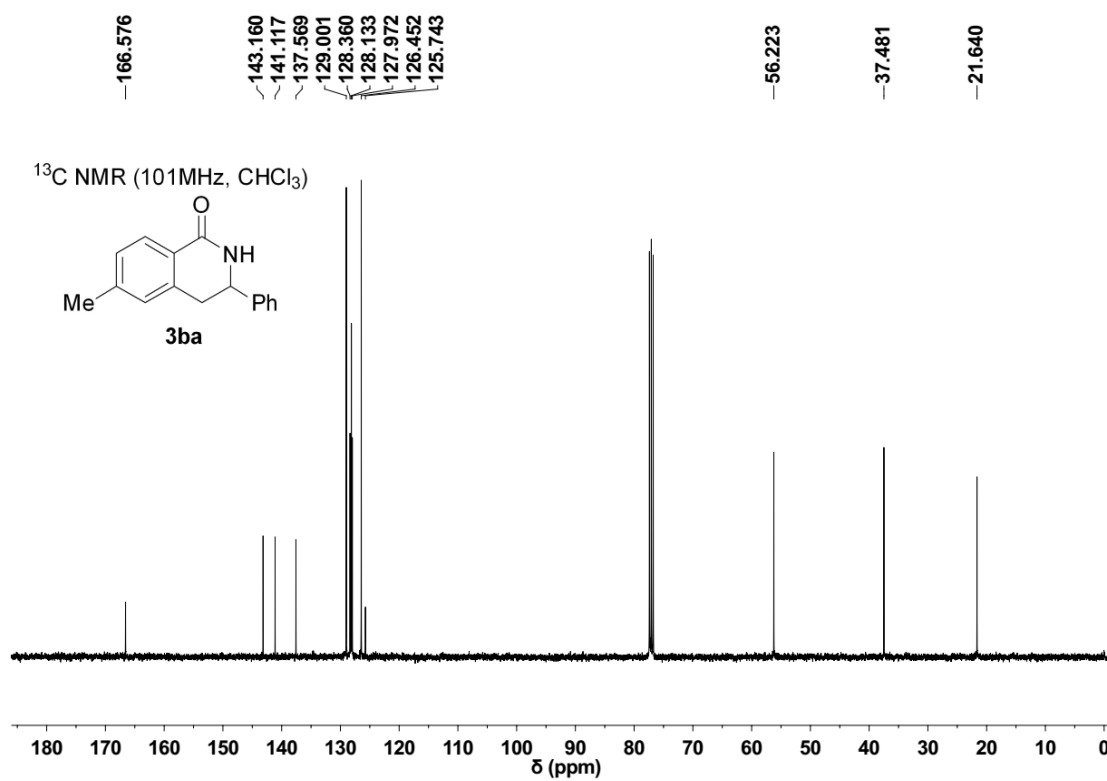
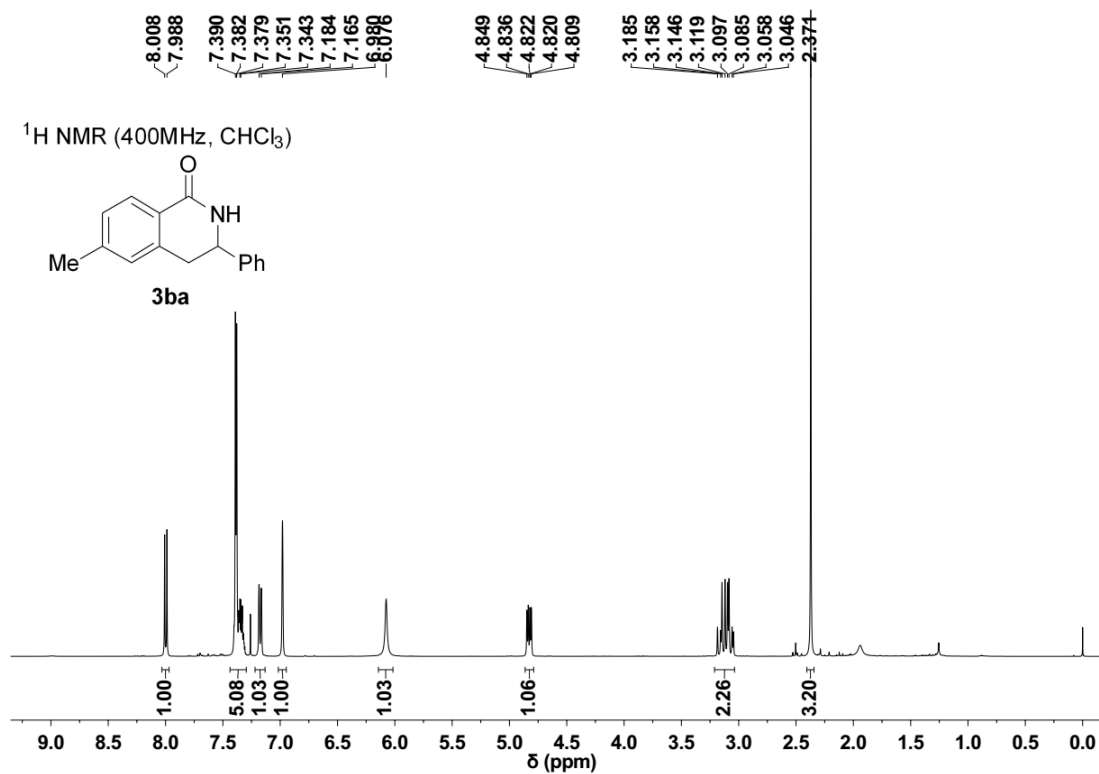
- [1] N. Guimond, S. I. Gorelsky and K. Fagnou, *J. Am. Chem. Soc.*, 2011, **133**, 6449.
- [2] B. Ye and N. Cramer, *Science*, **2012**, 338, 504.
- [3] S. Rakshit, C. Grohmann, T. Besset and F. Glorius, *J. Am. Chem. Soc.*, 2011, **133**, 2350.
- [4] B. Li, J. Ma, N. Wang, H. Feng, S. Xu and B. Wang, *Org. Lett.*, 2012, **14**, 736.
- [5] H. Liu, Z. An and J. He, *ACS Catal.*, 2014, **4**, 3543.

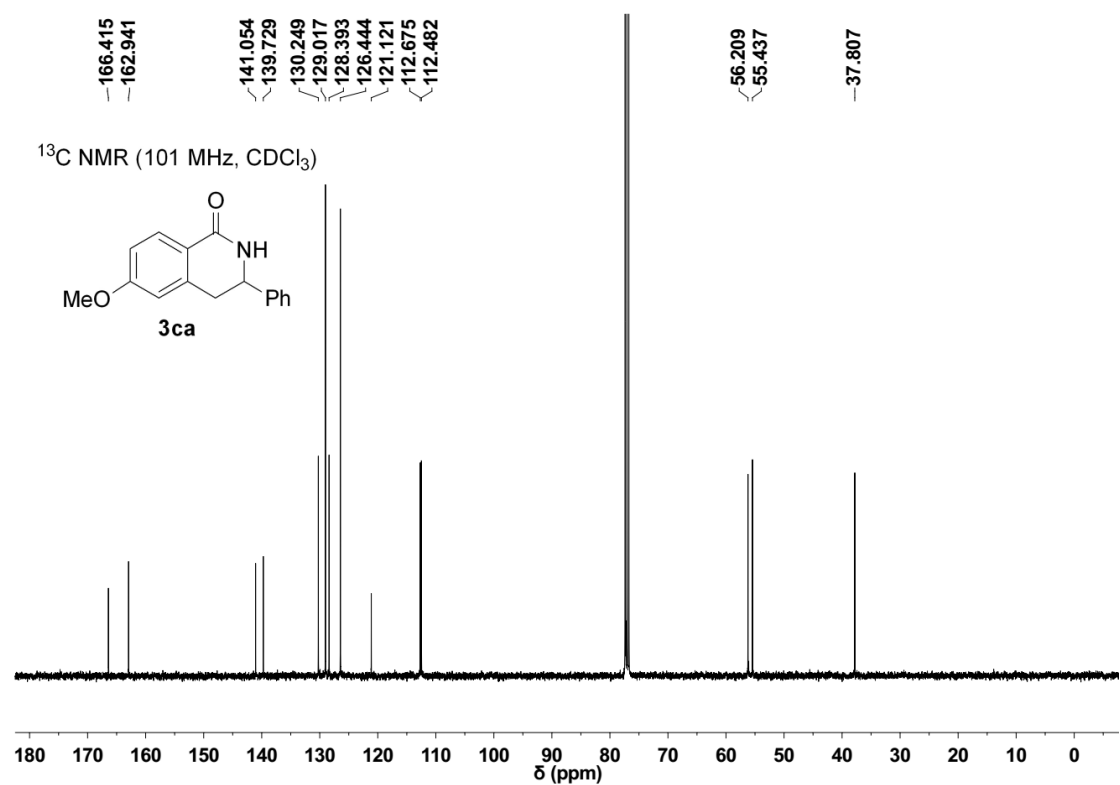
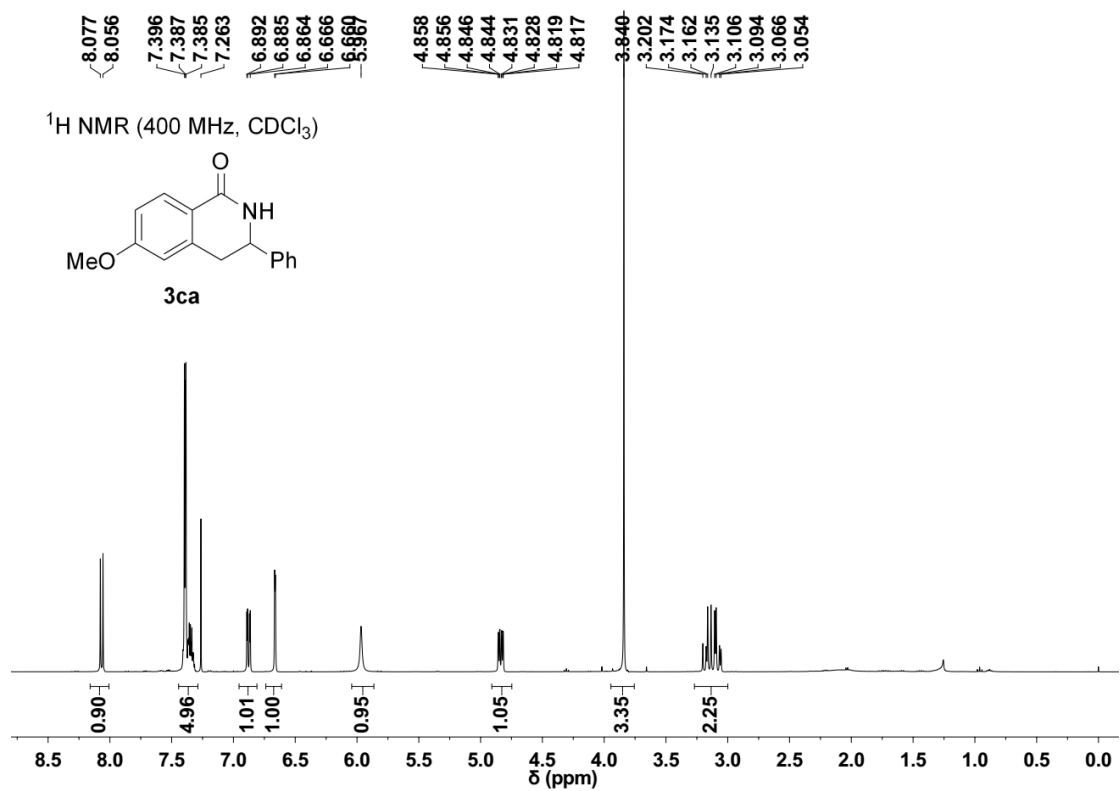
11. NMR Spectra for New Compounds

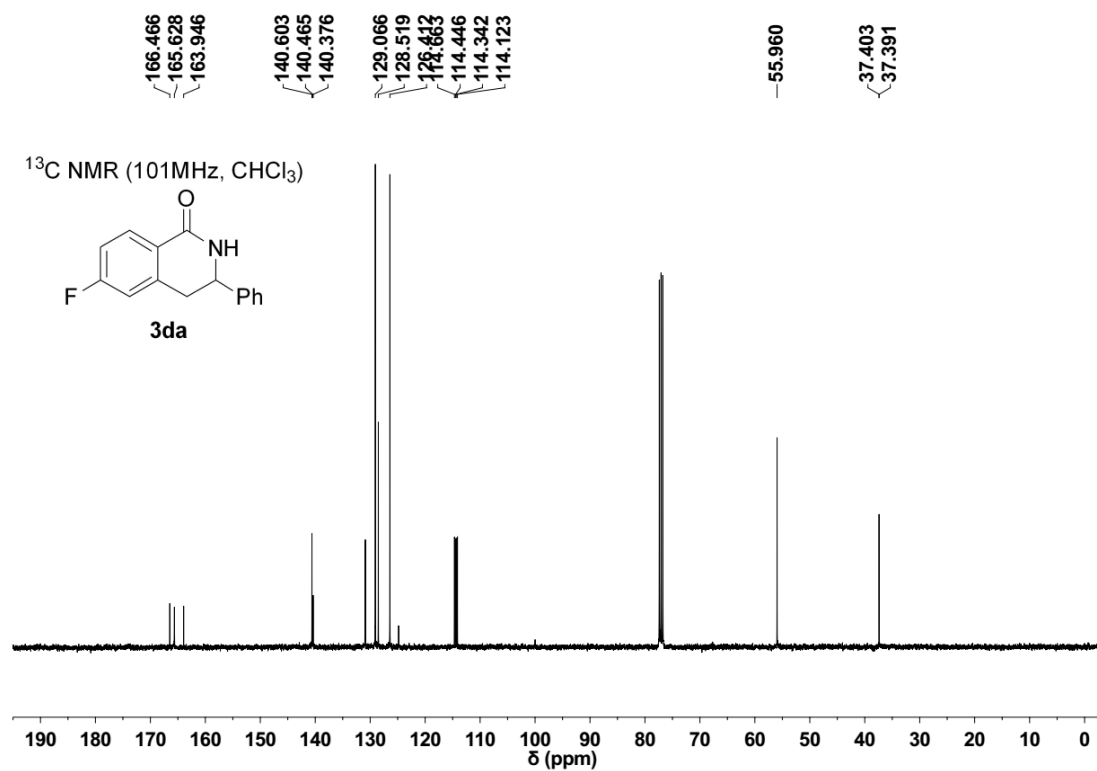
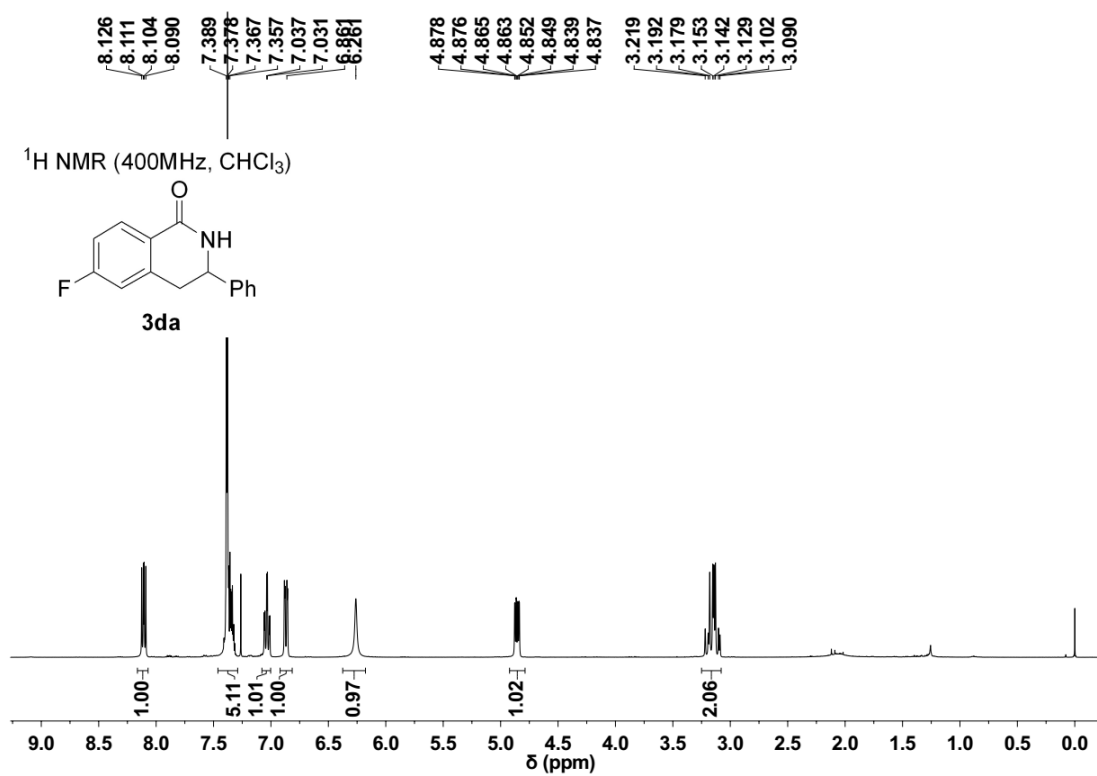


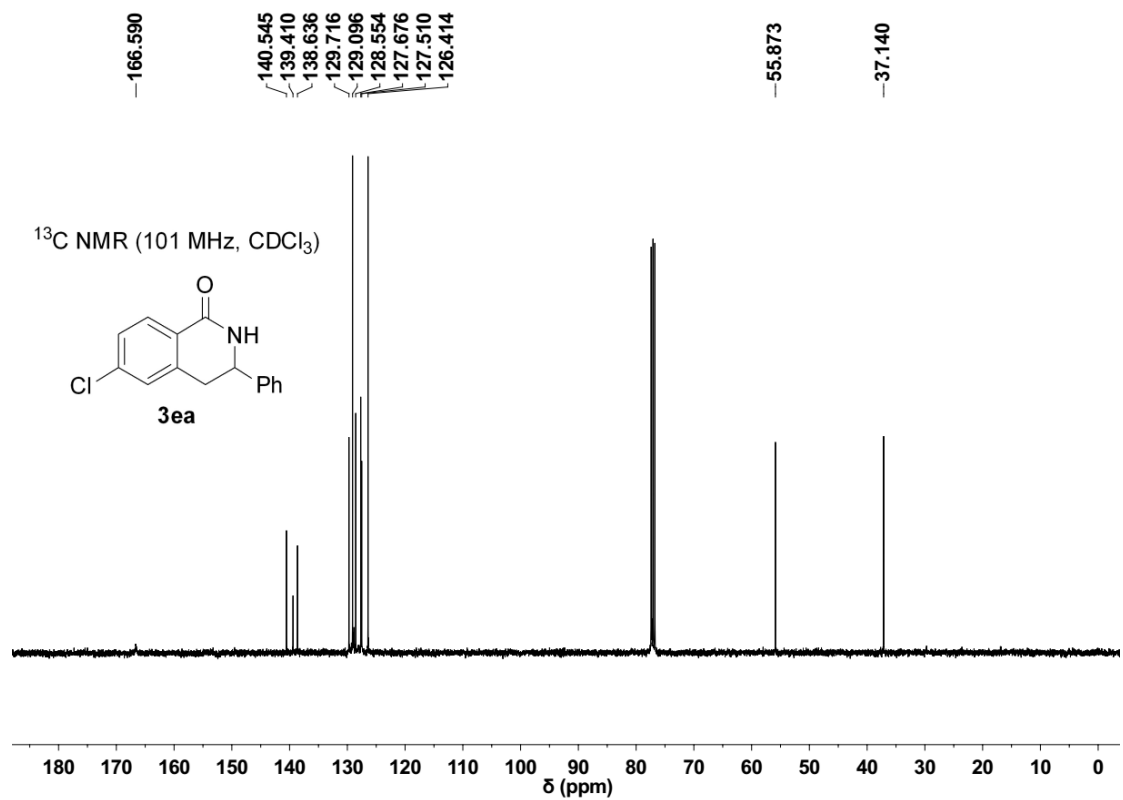
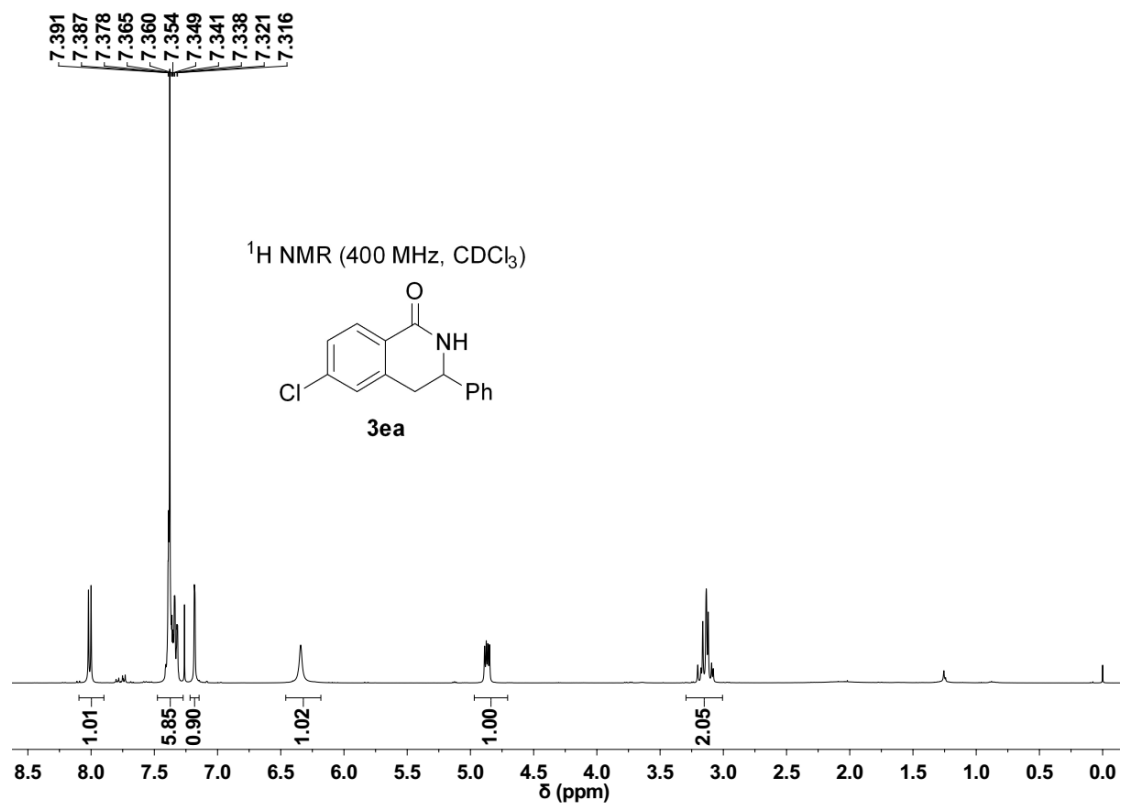


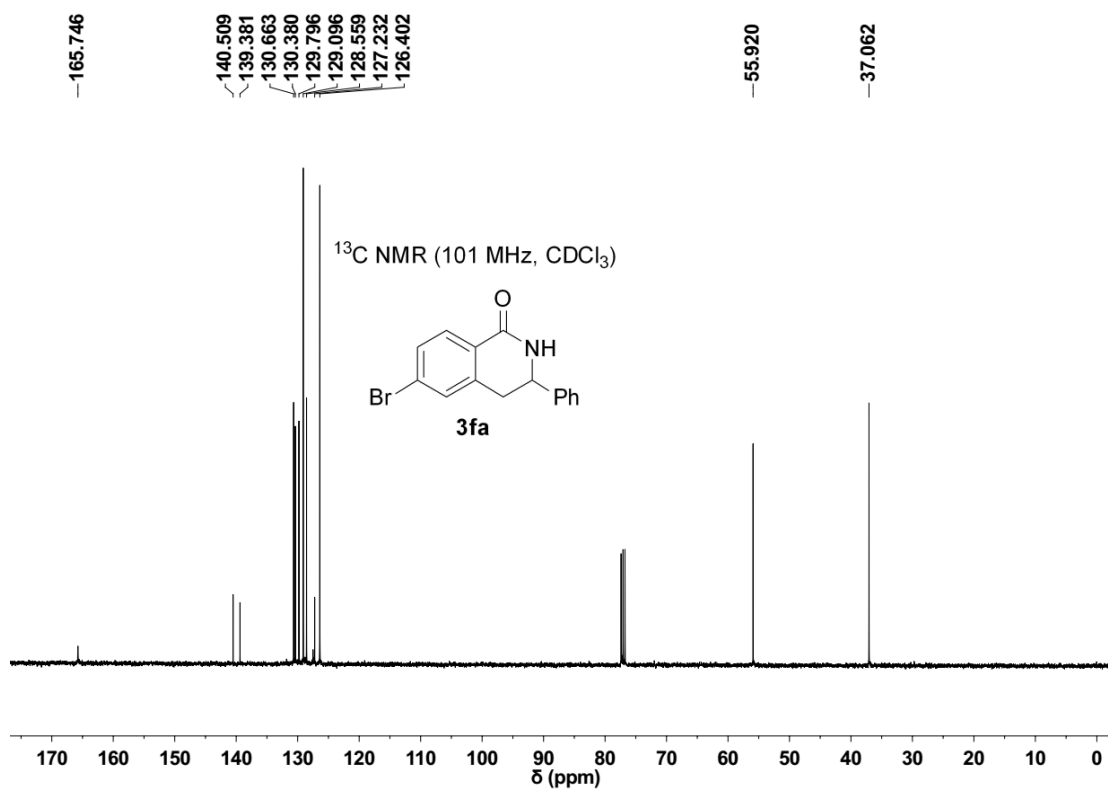
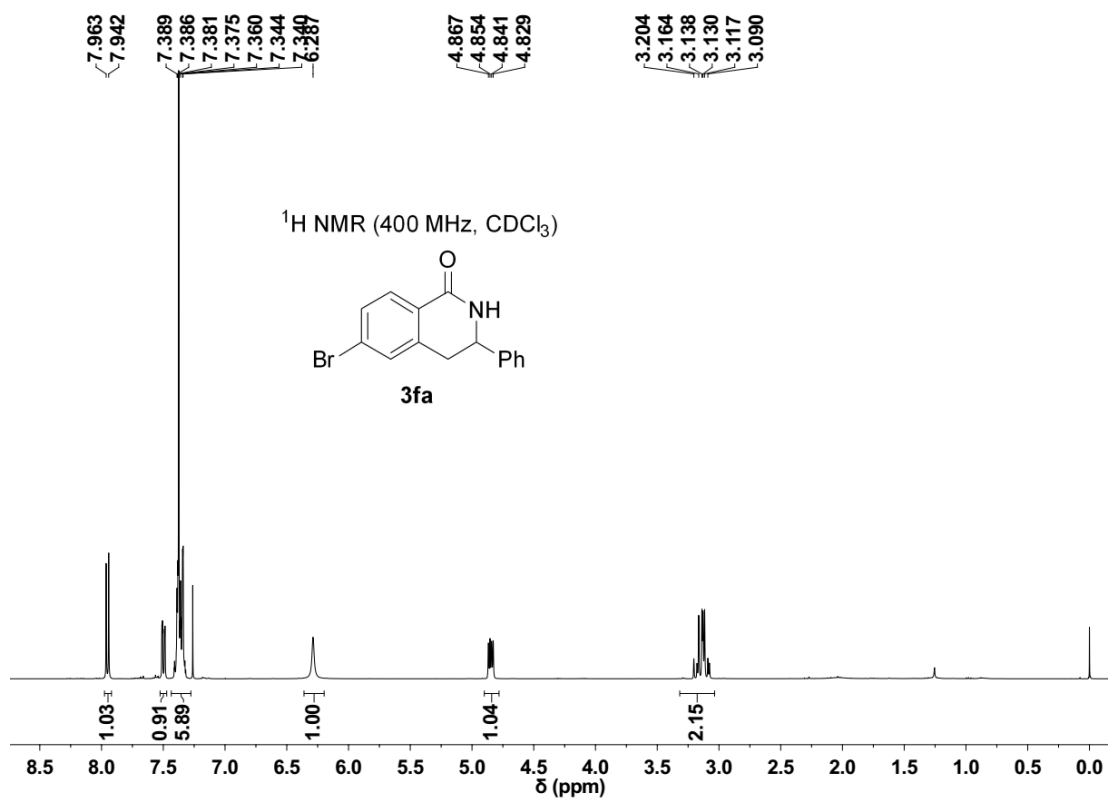


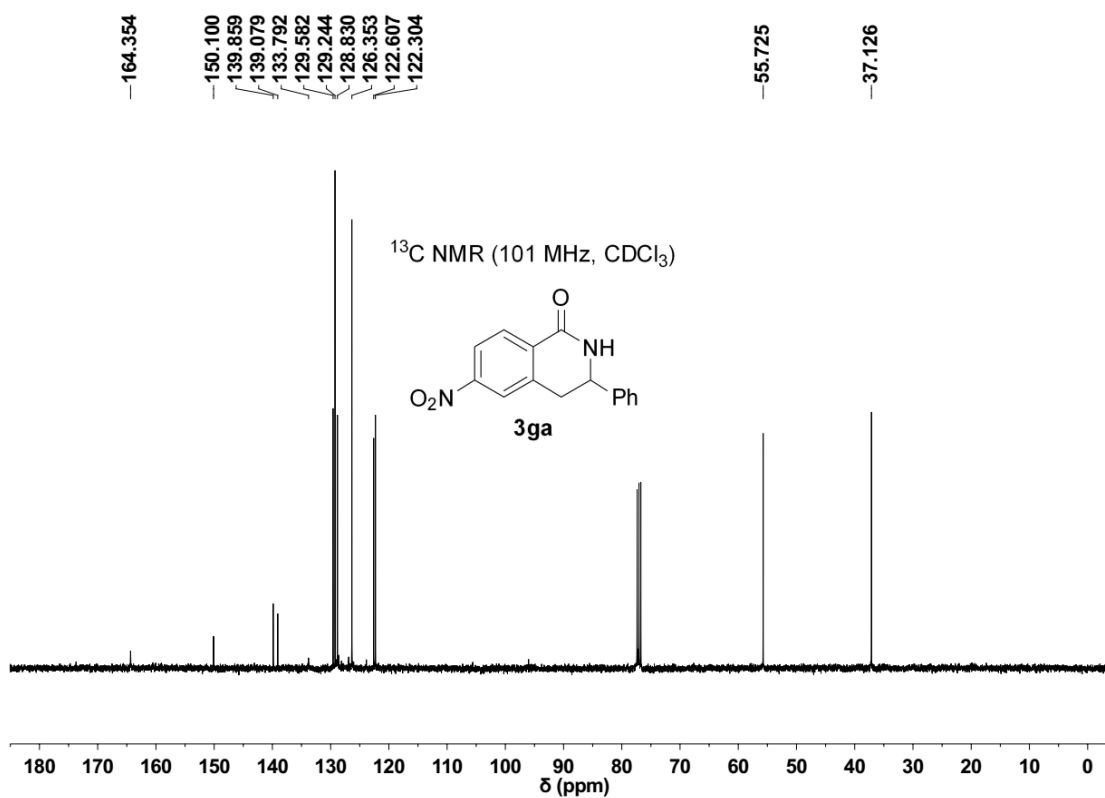
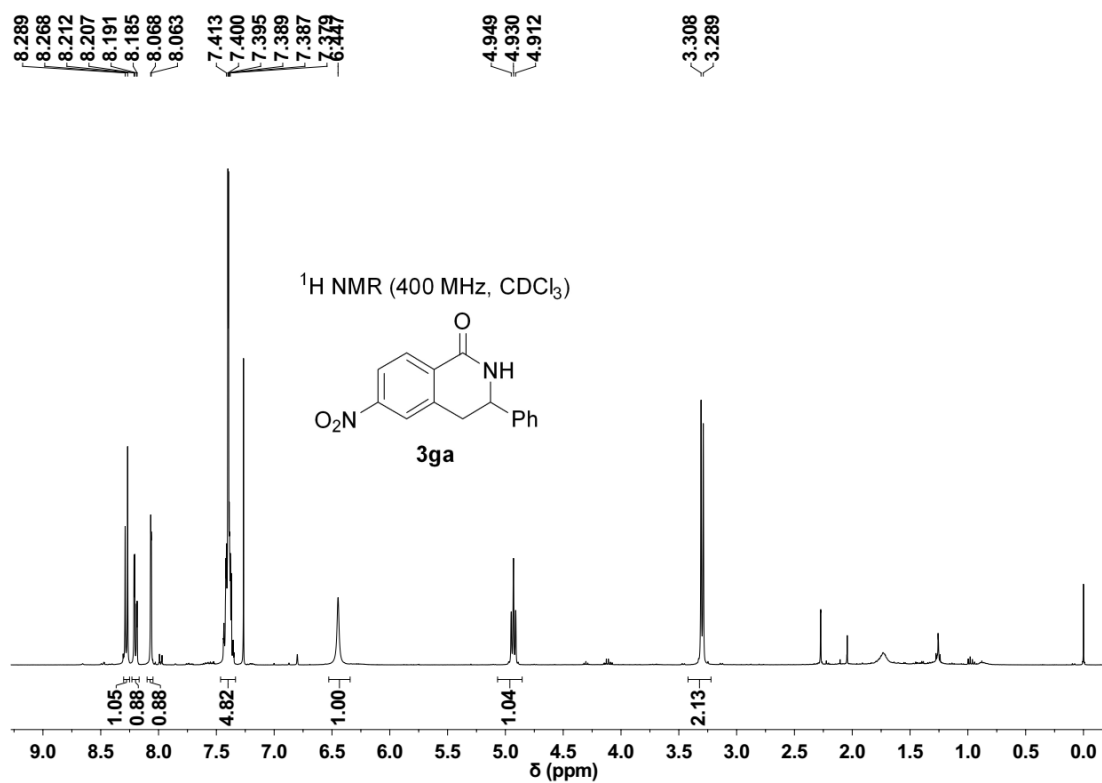


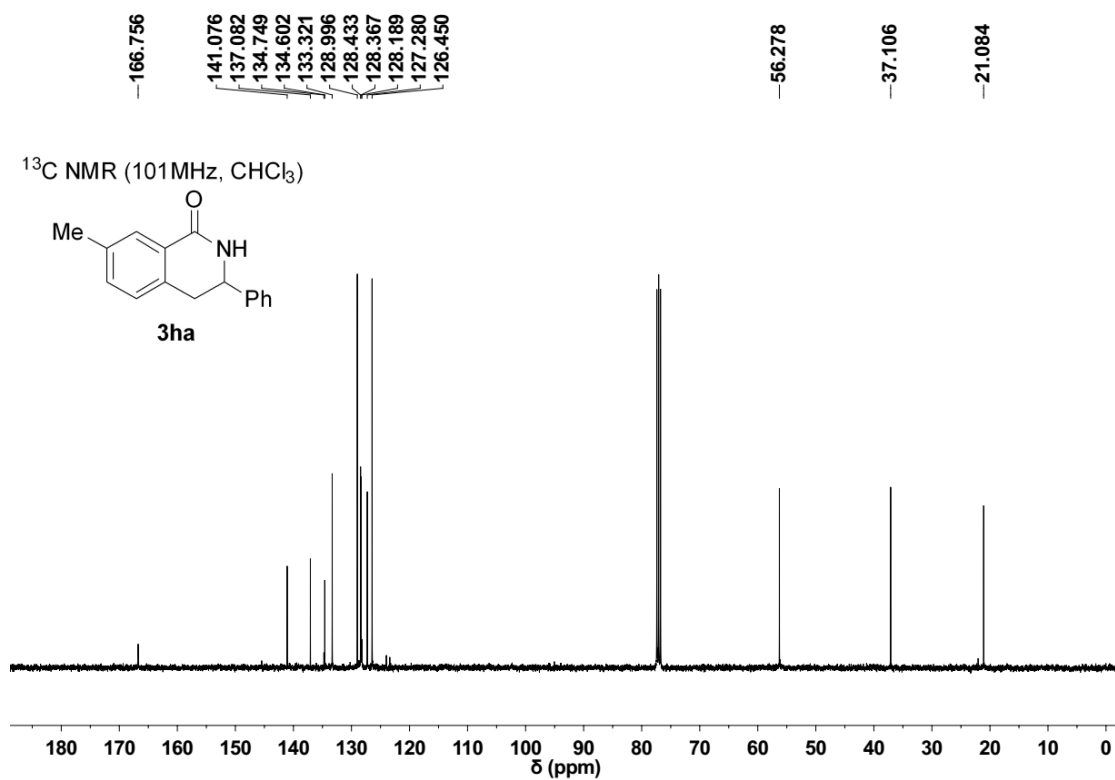
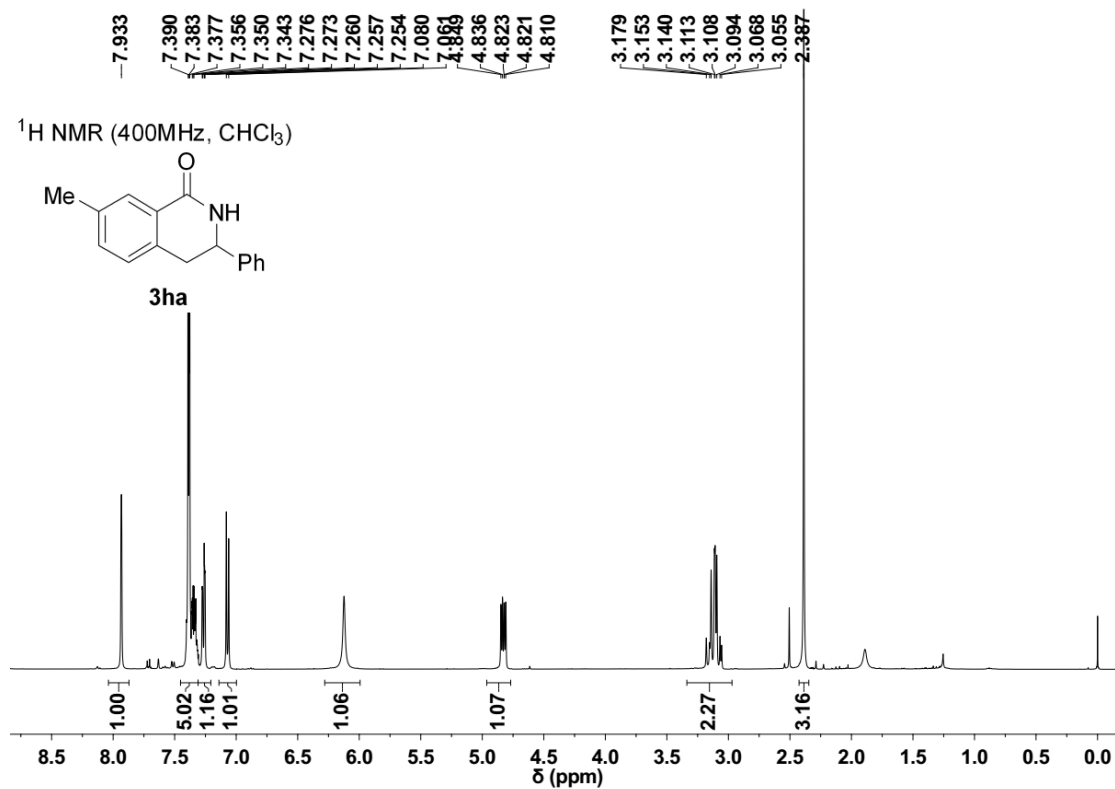


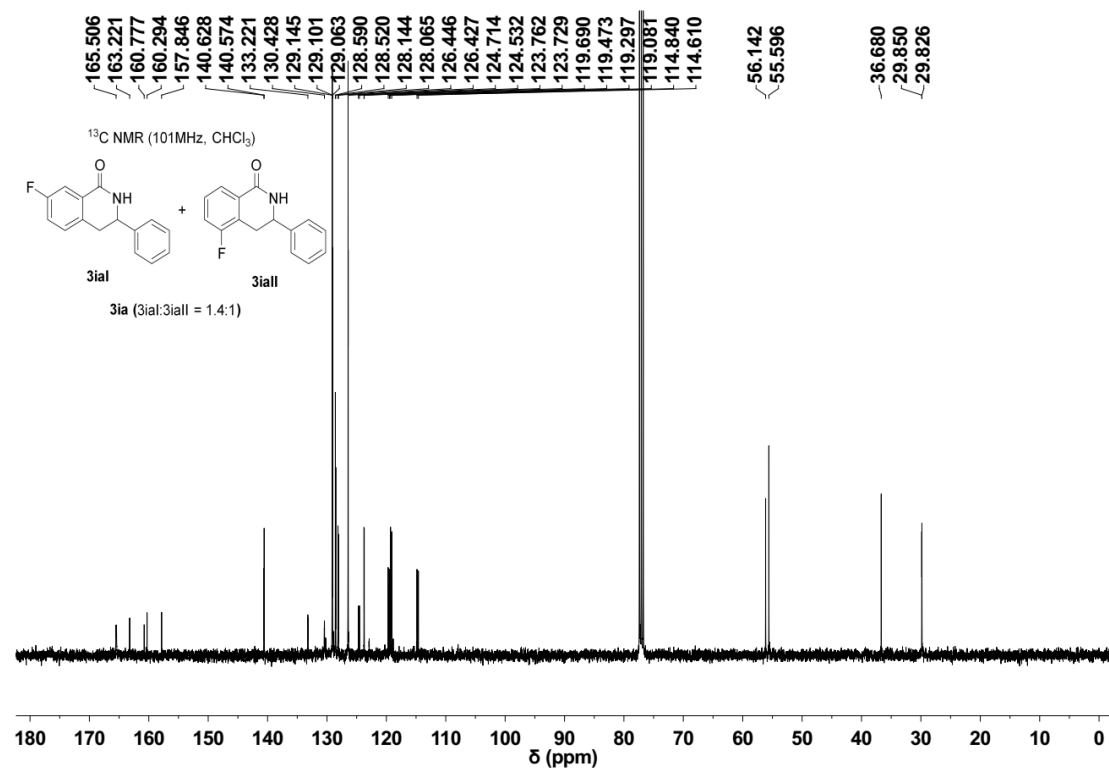
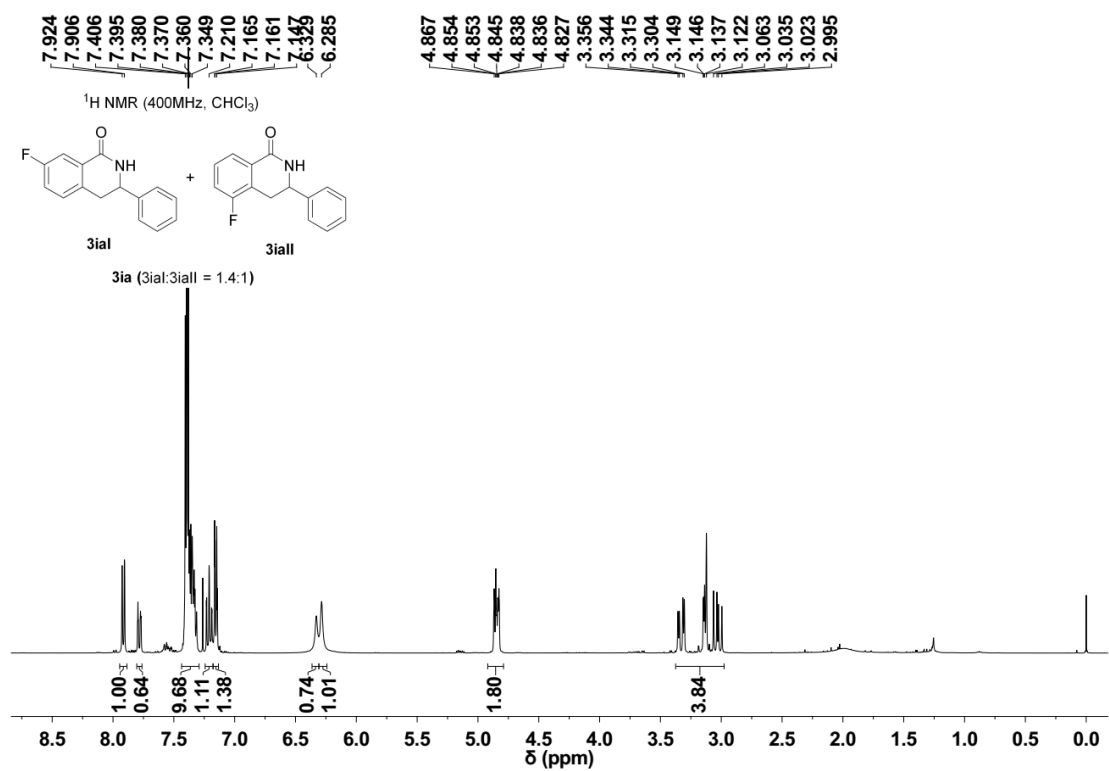


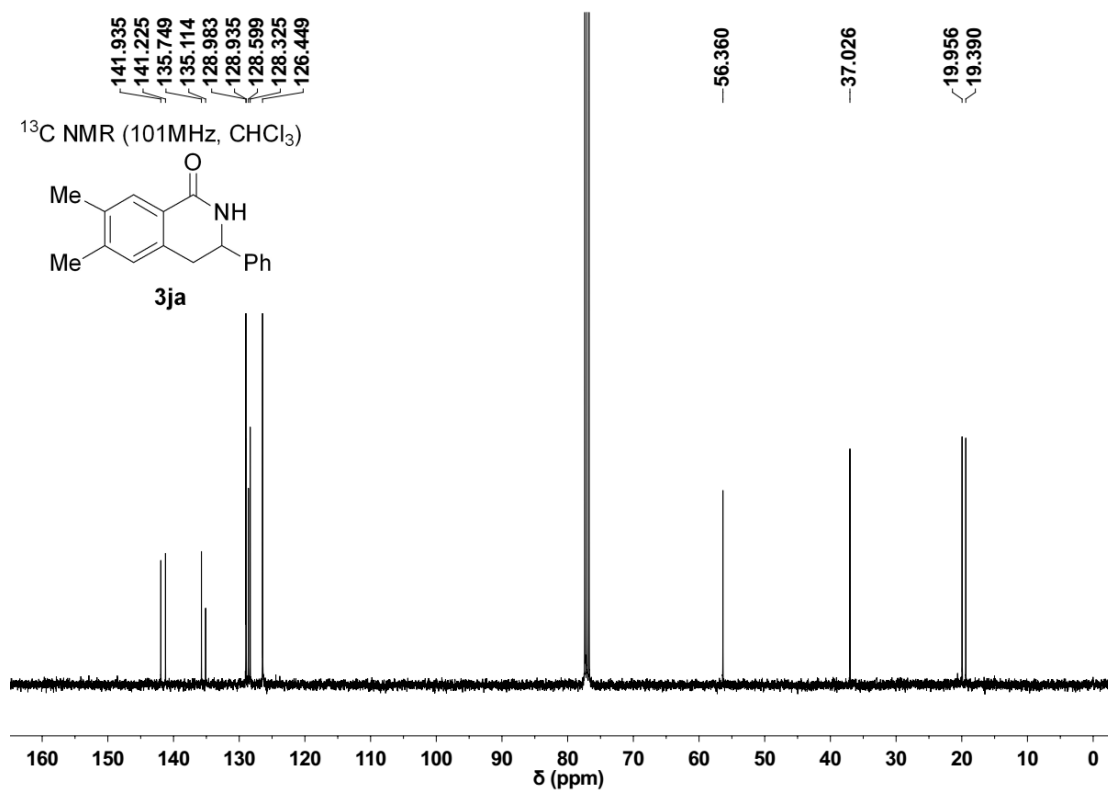
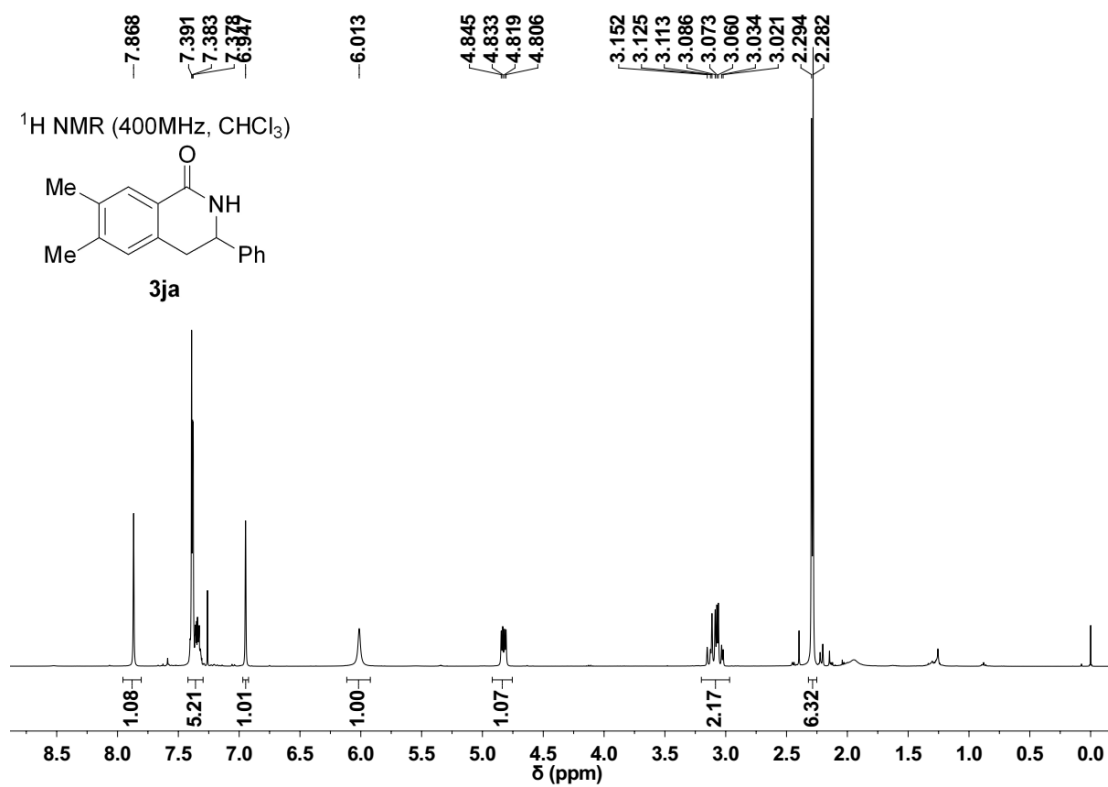


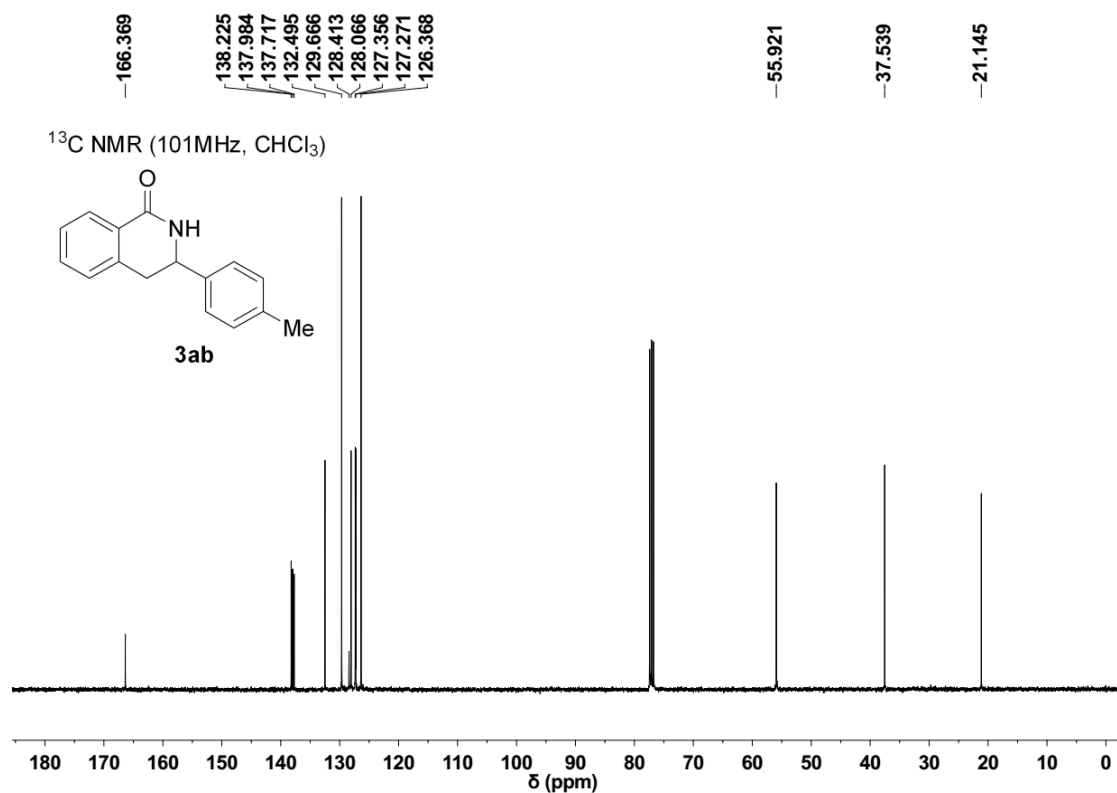
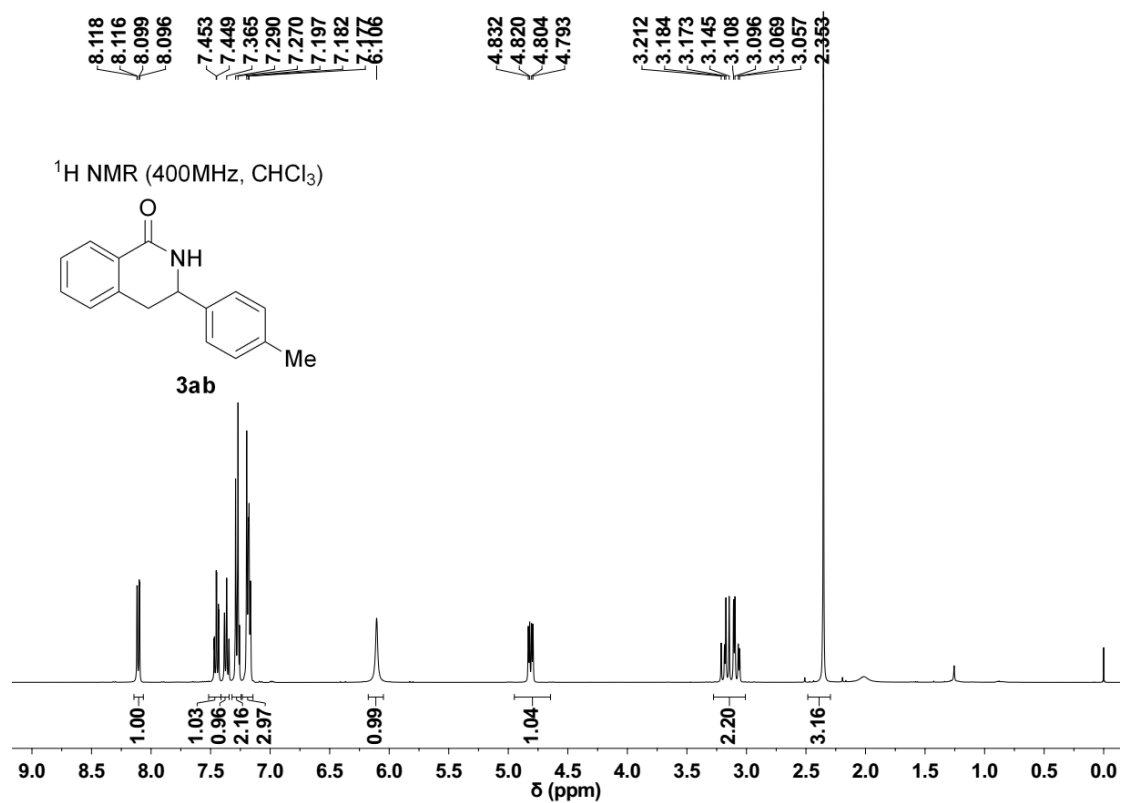


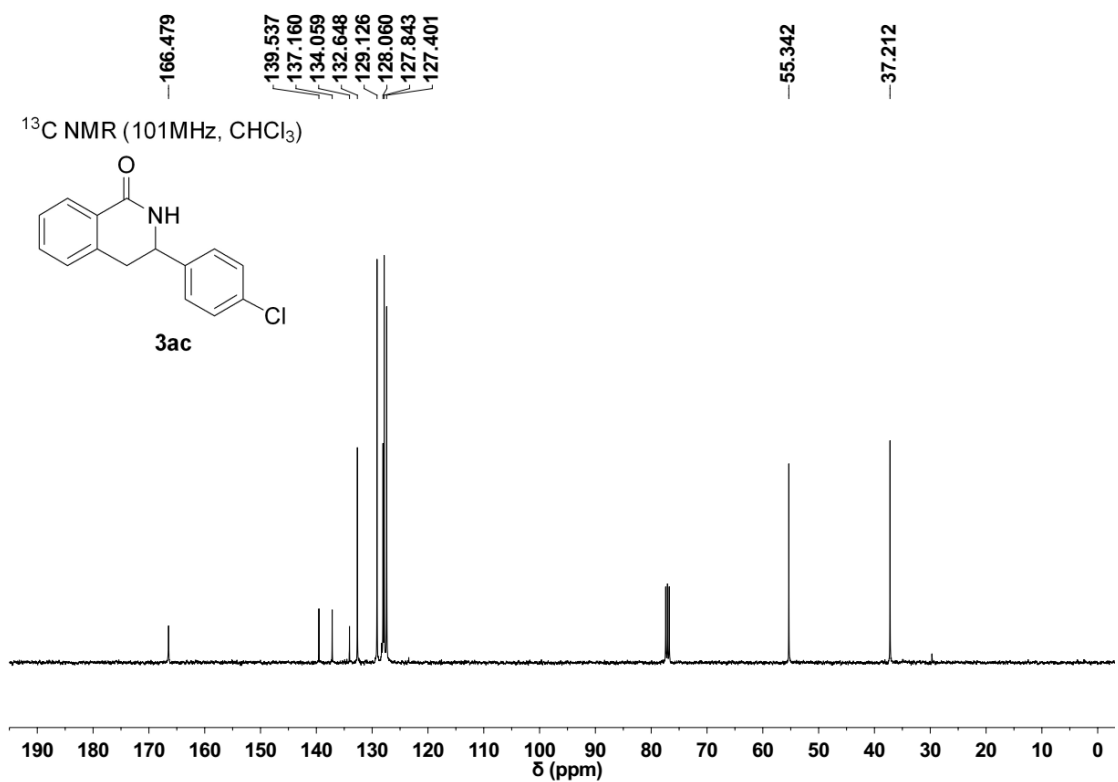
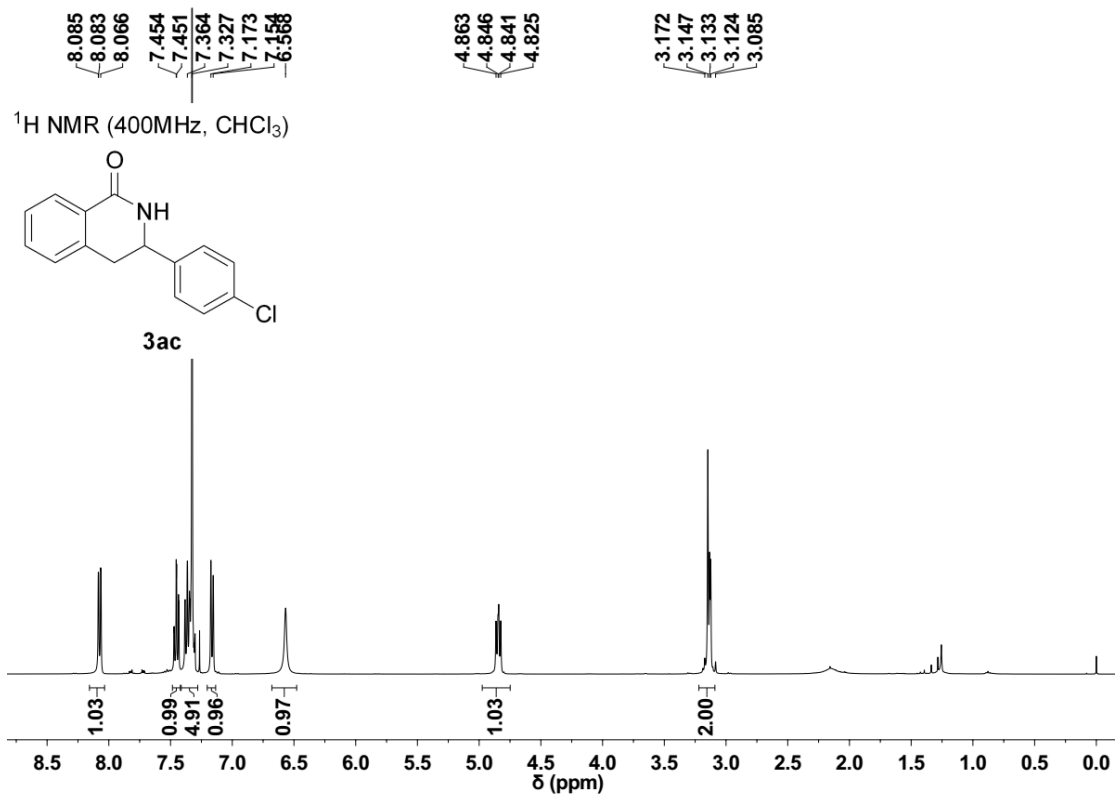


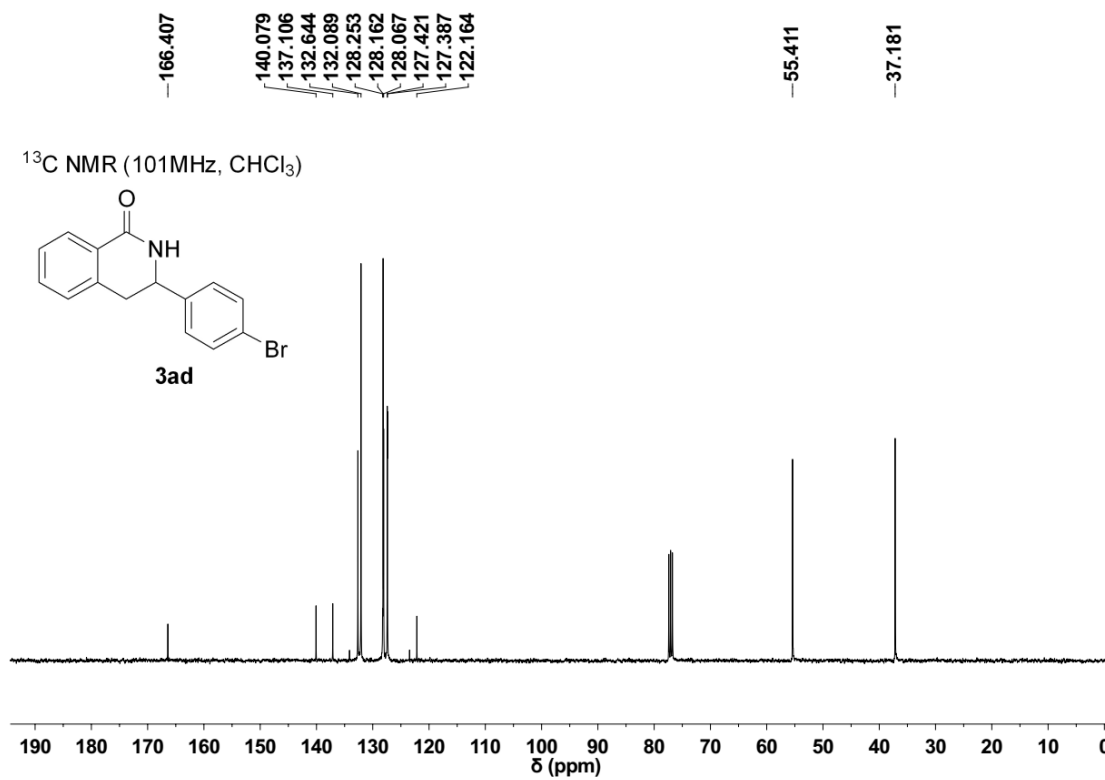
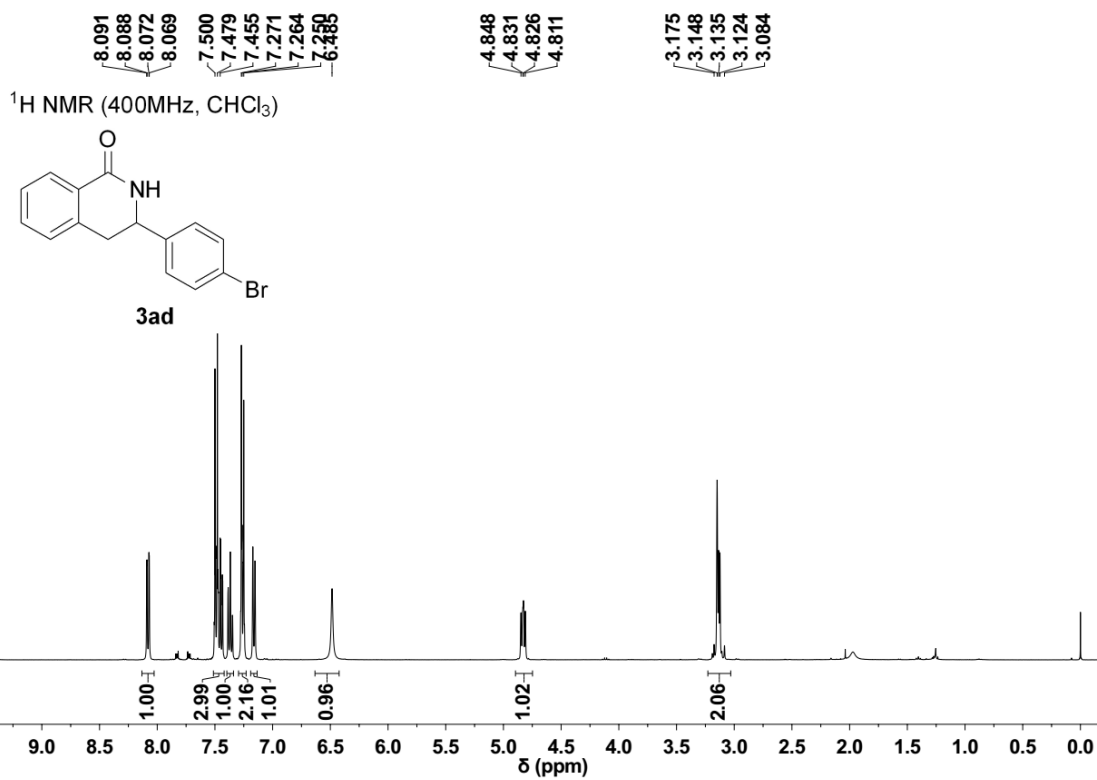


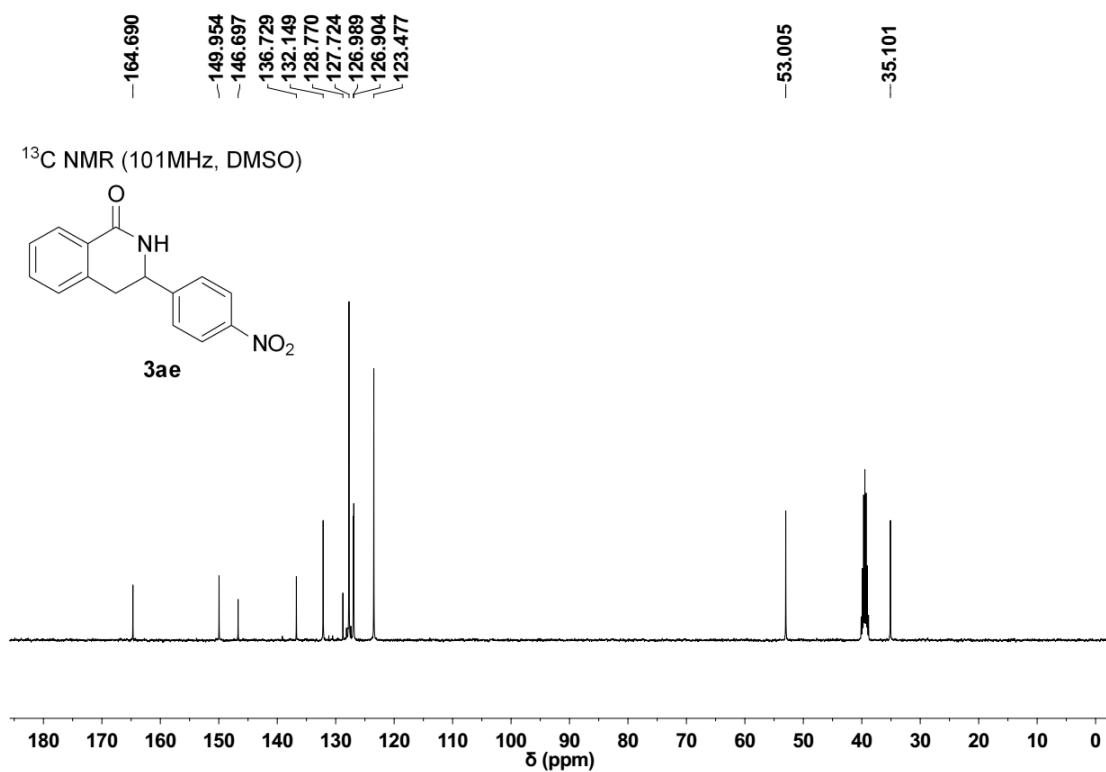
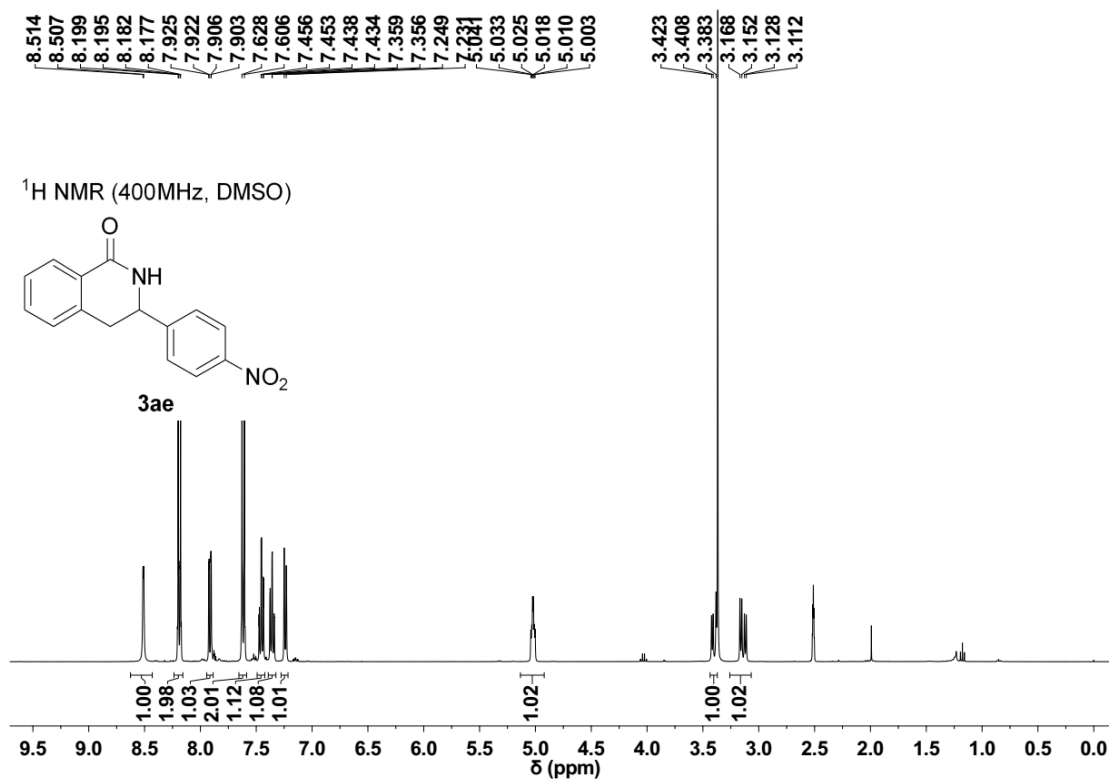


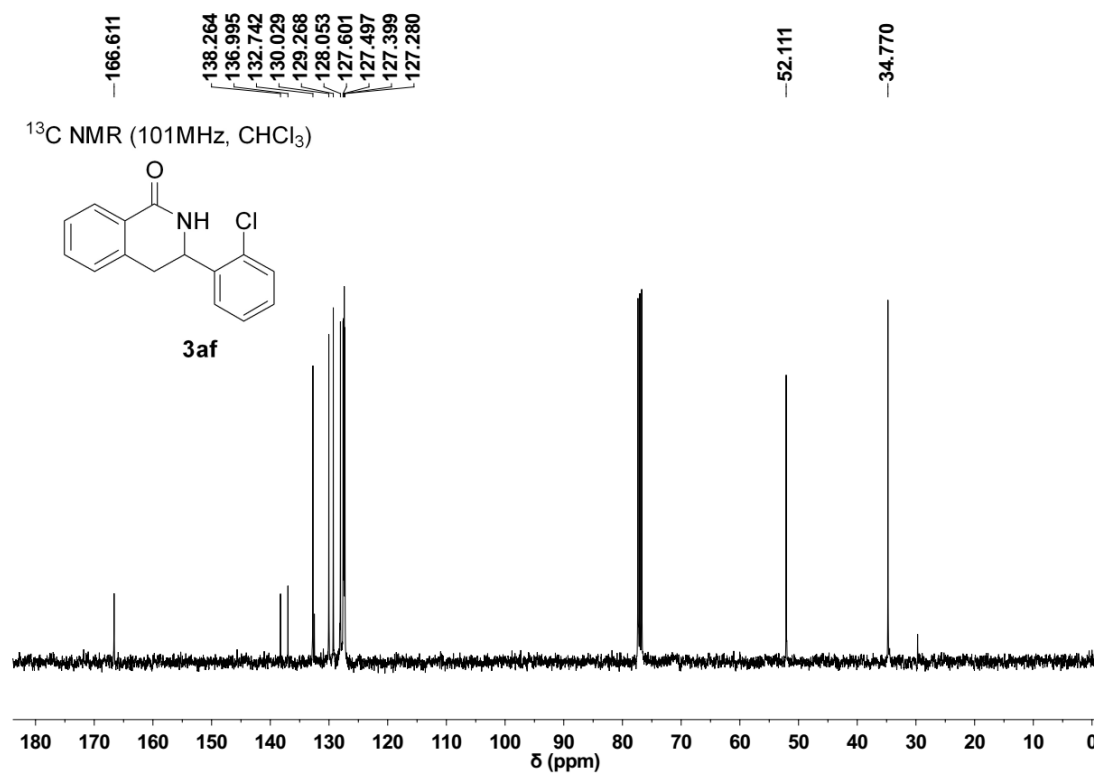
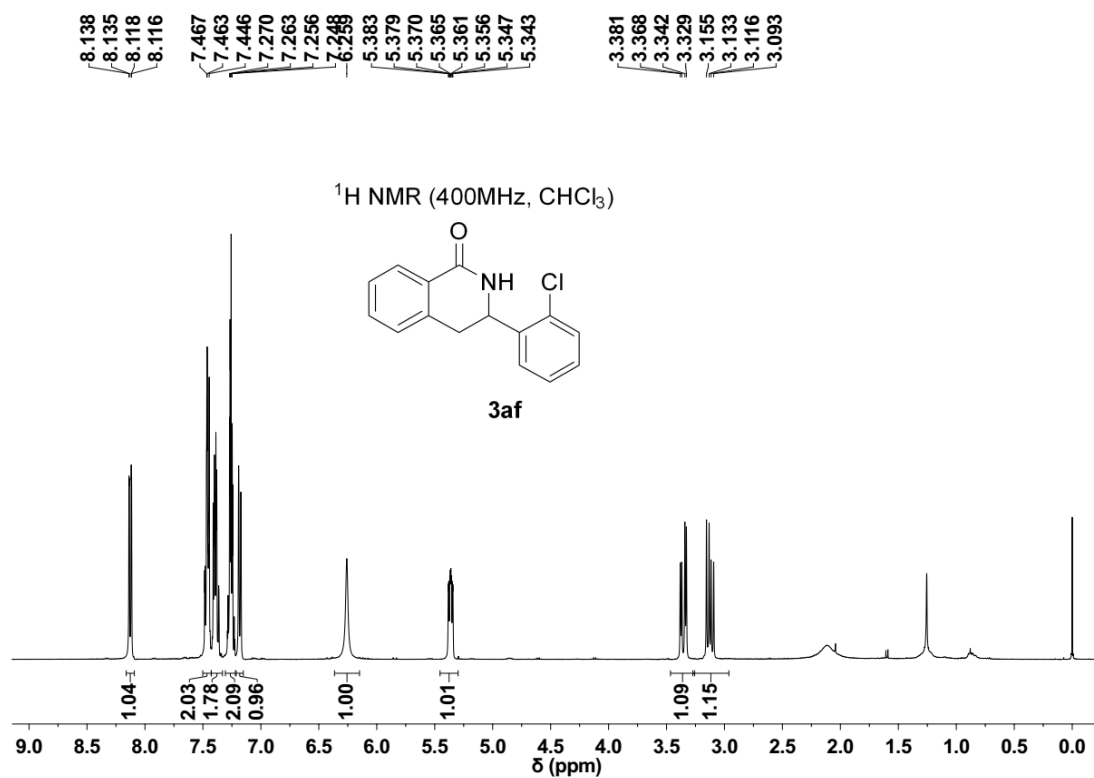


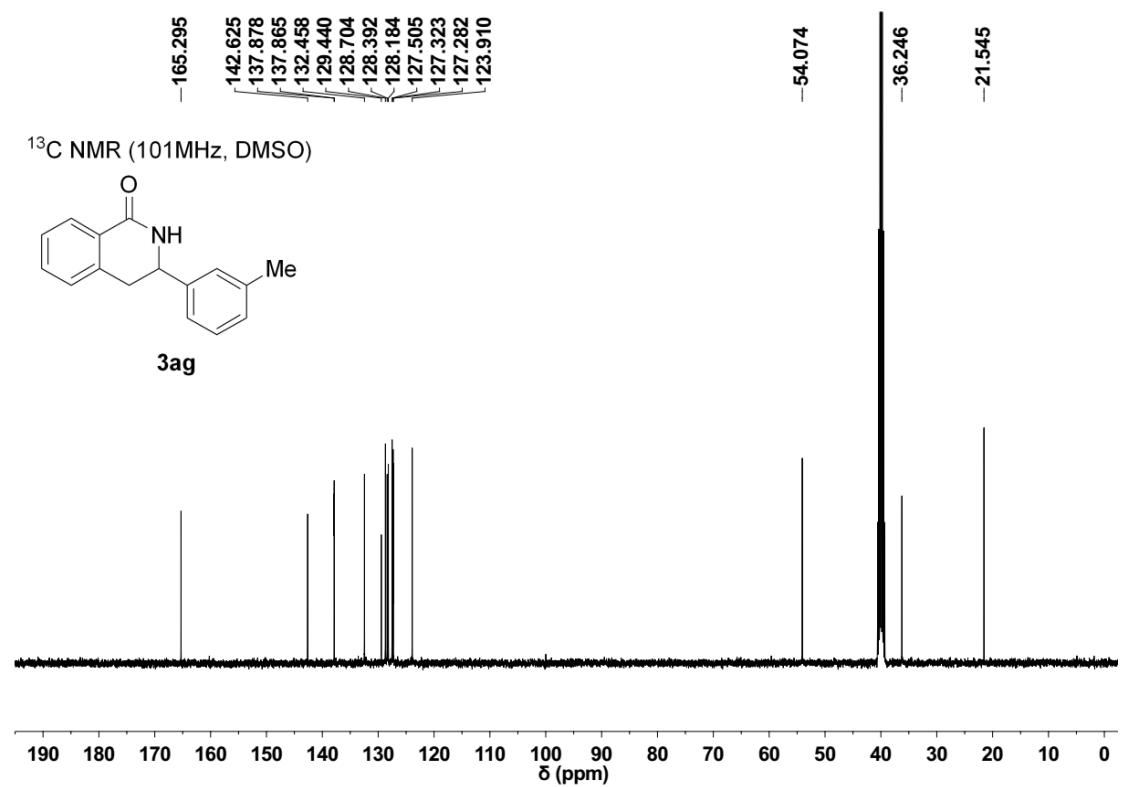
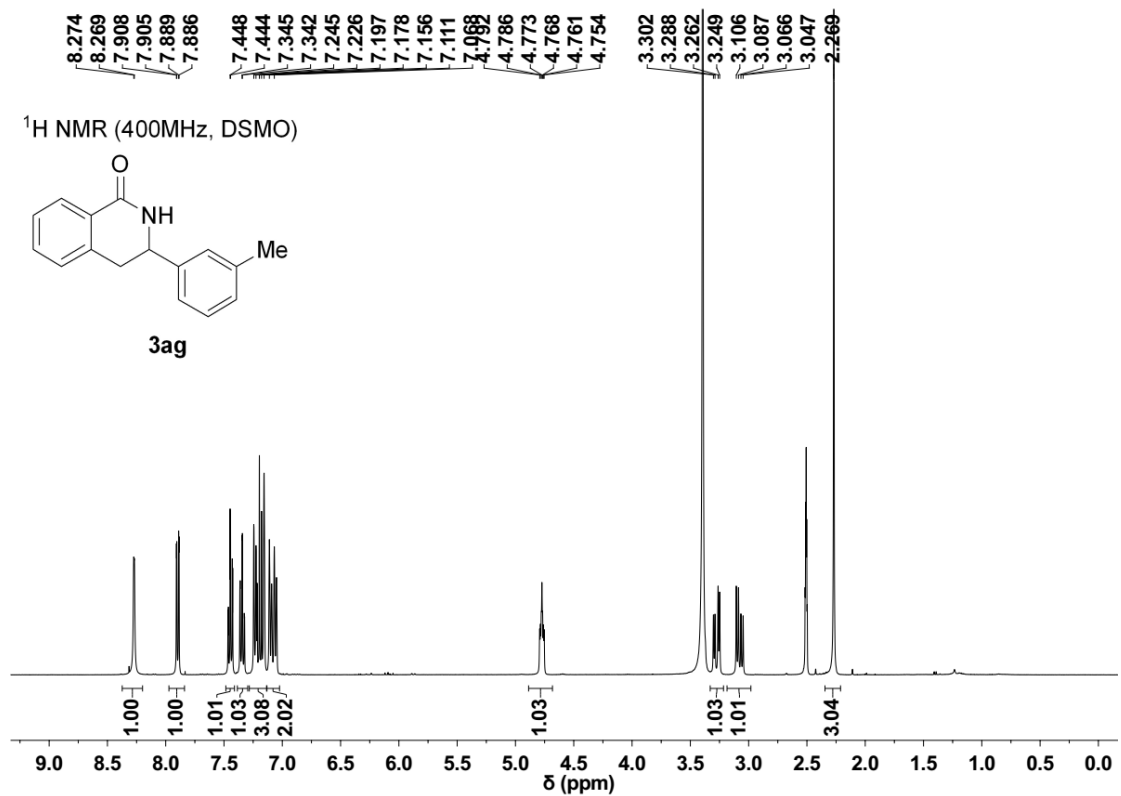


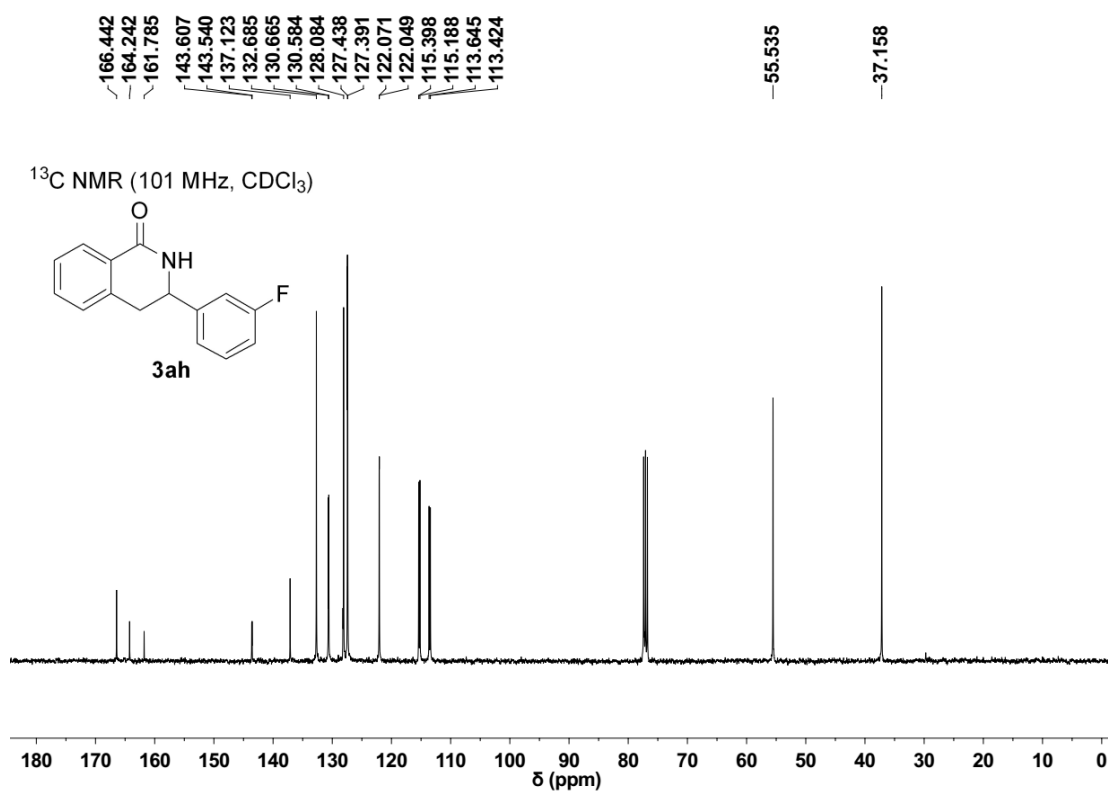
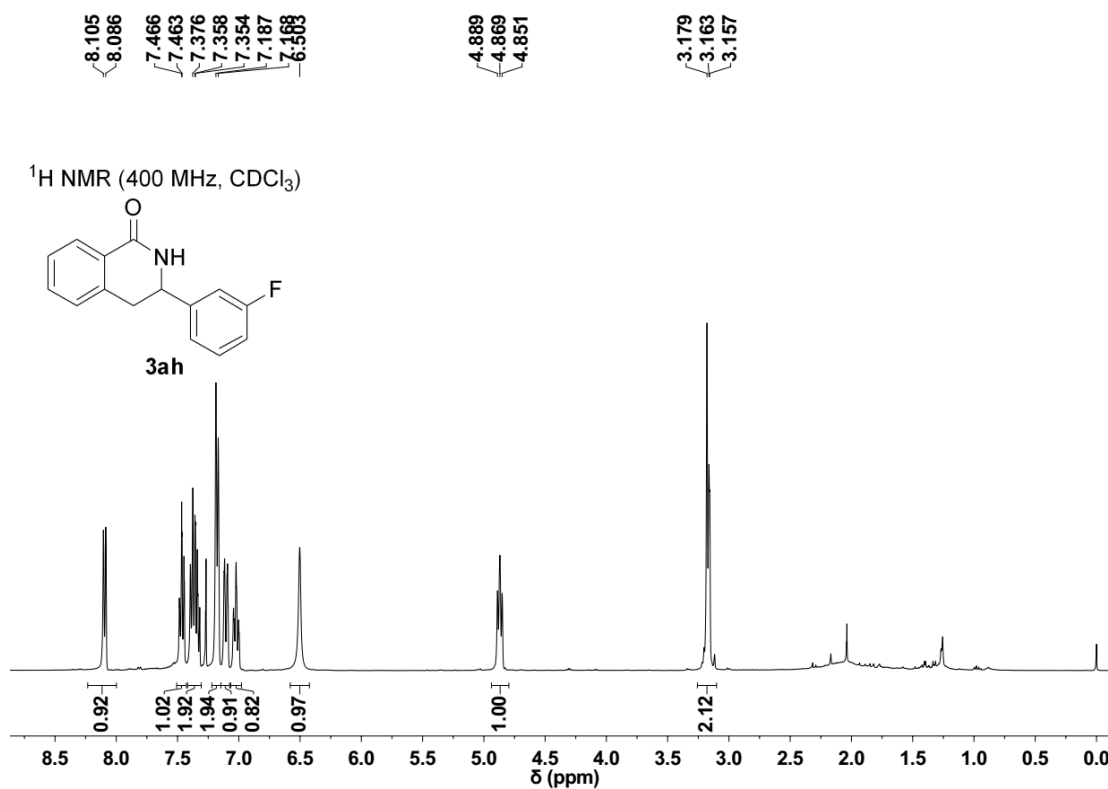


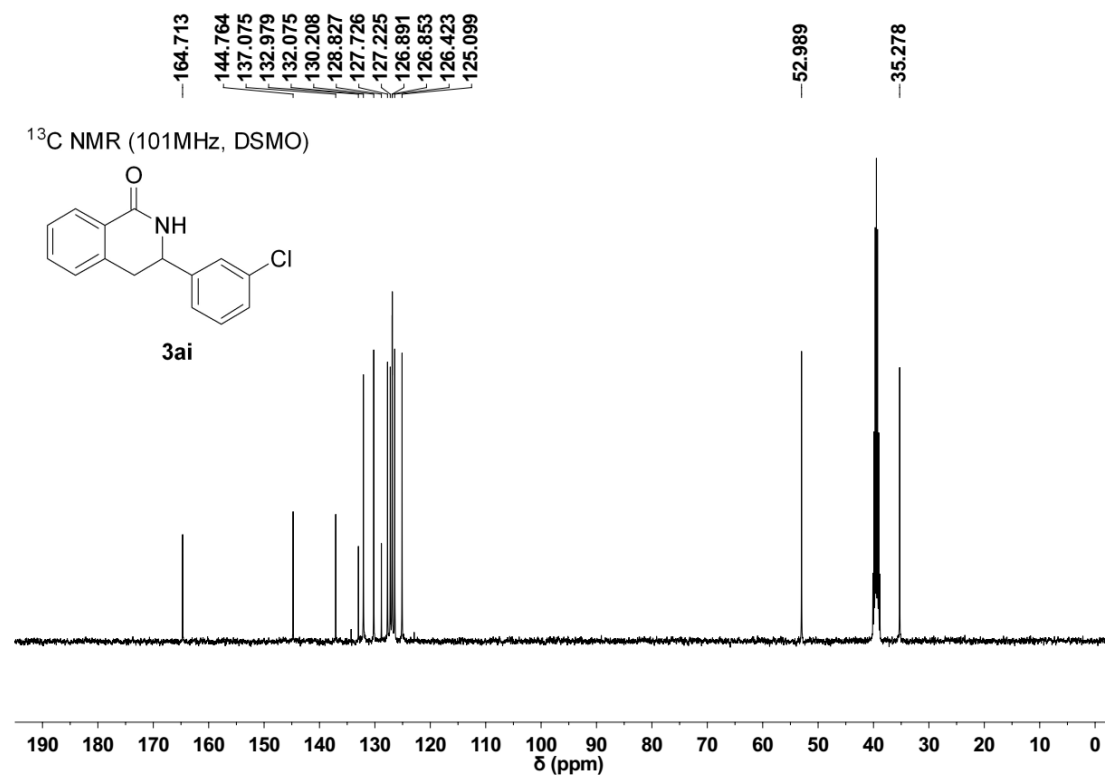
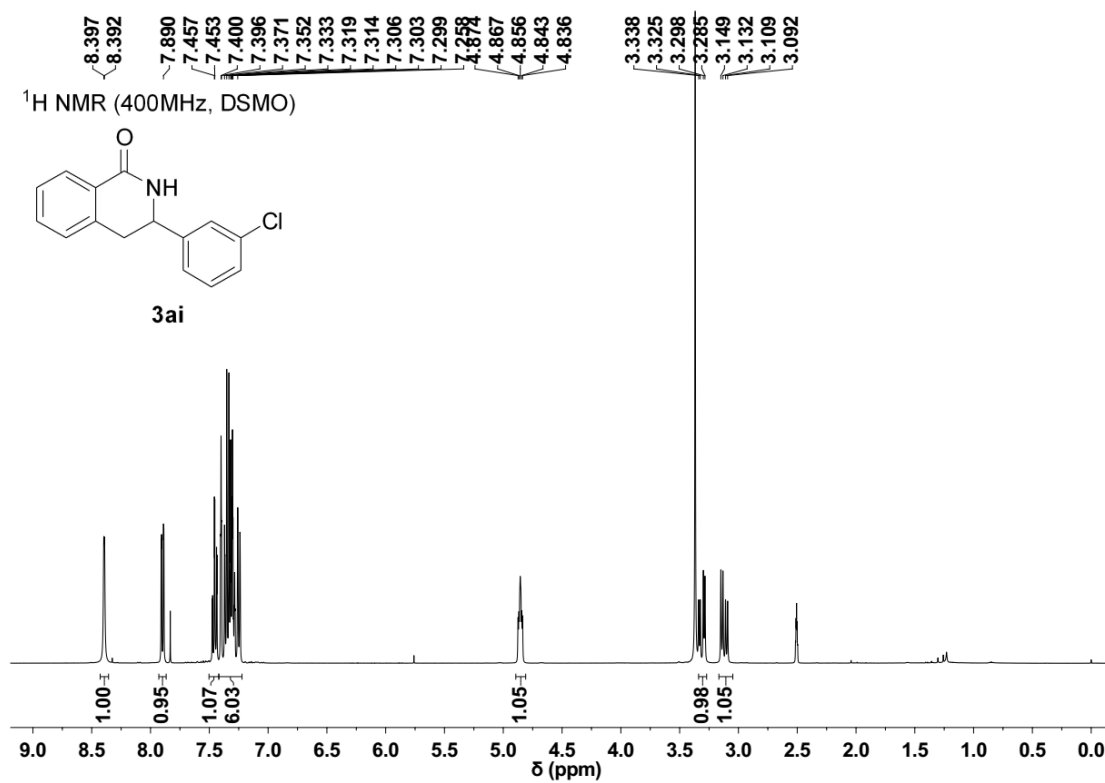


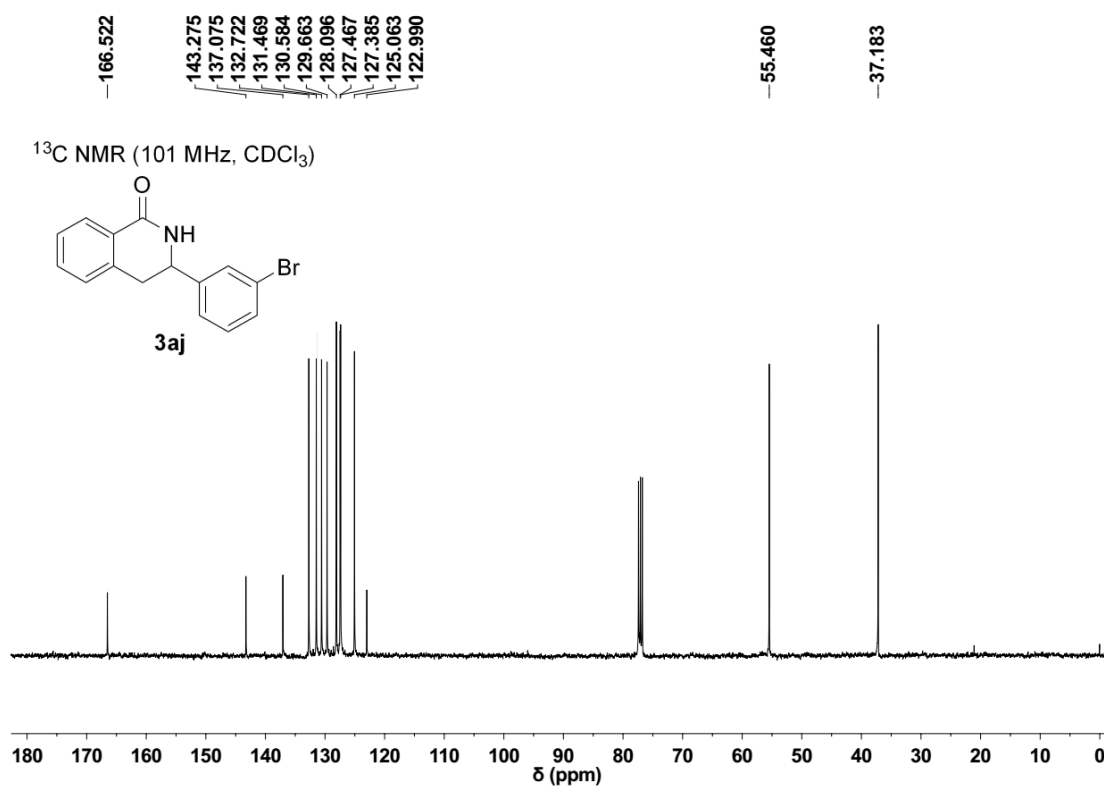
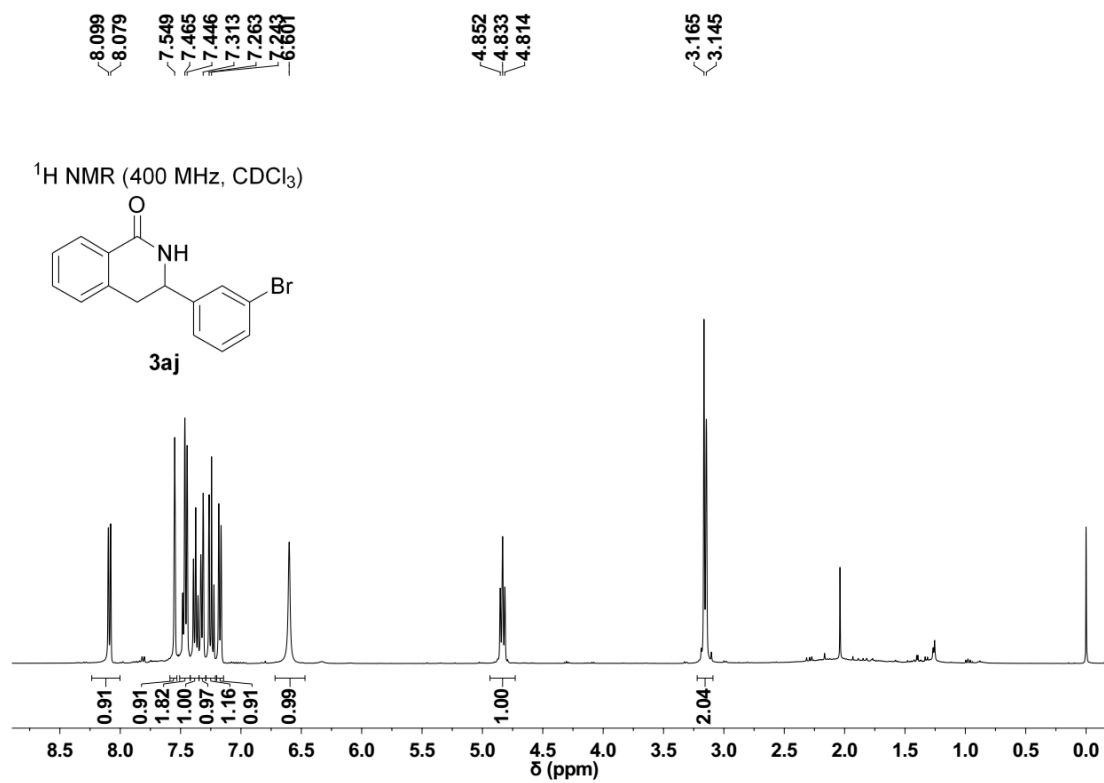


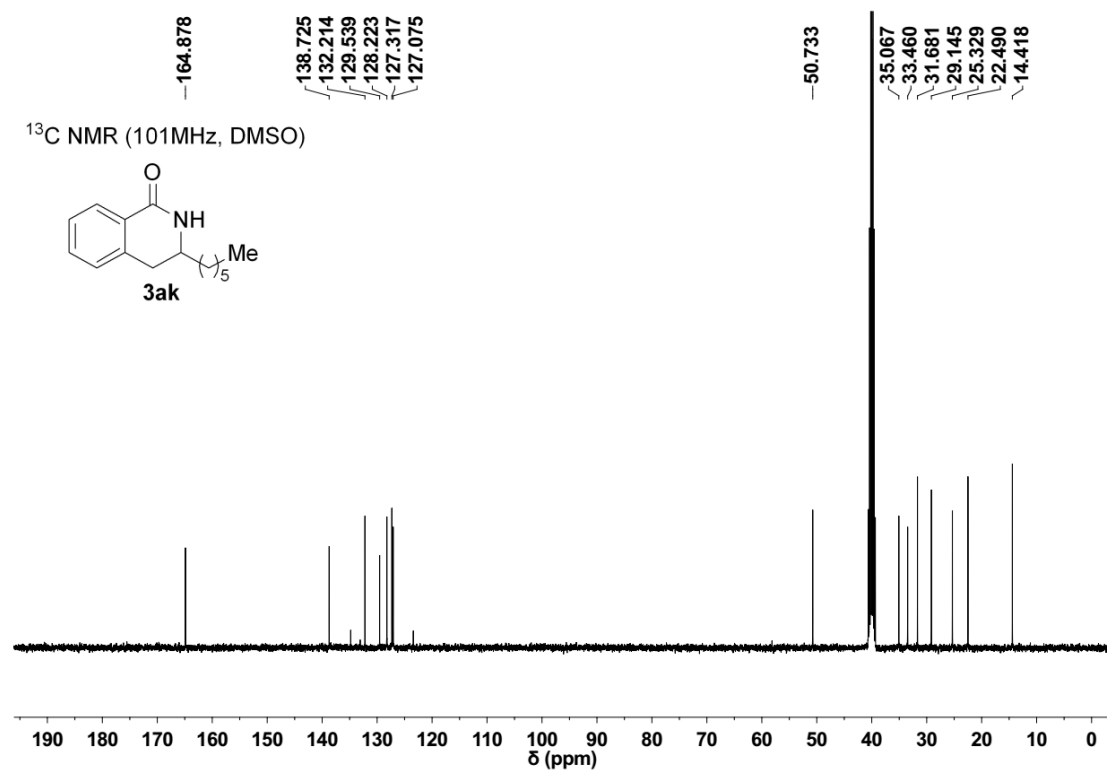
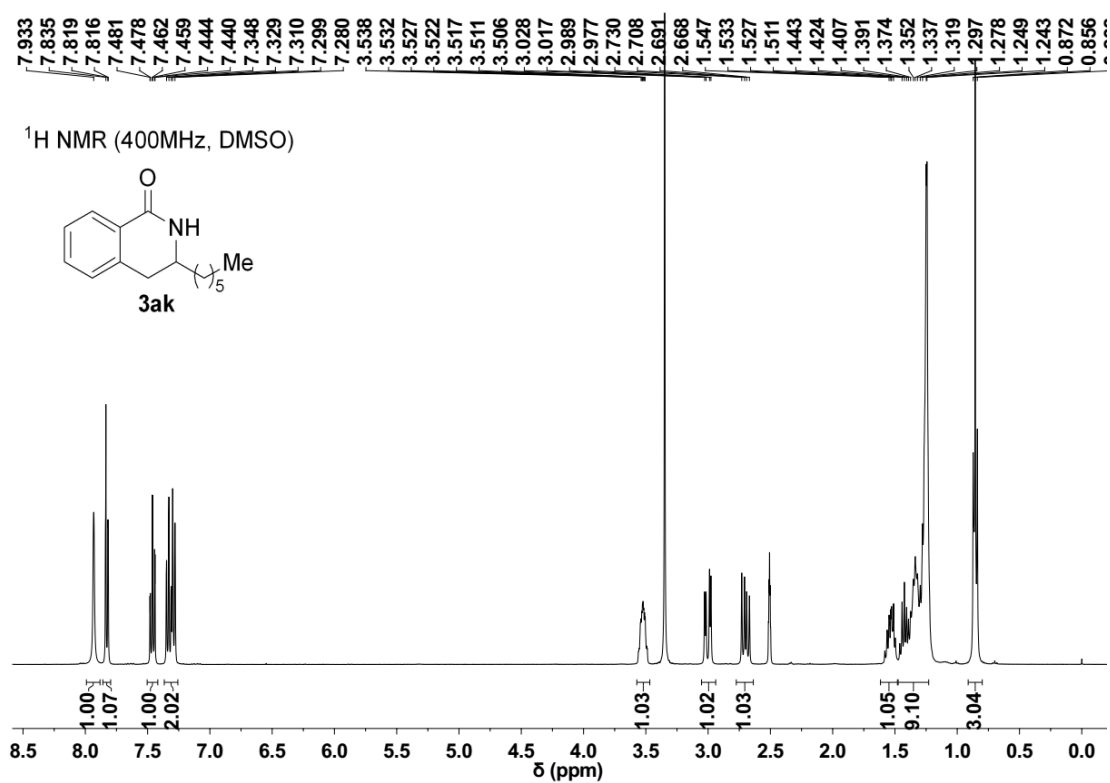


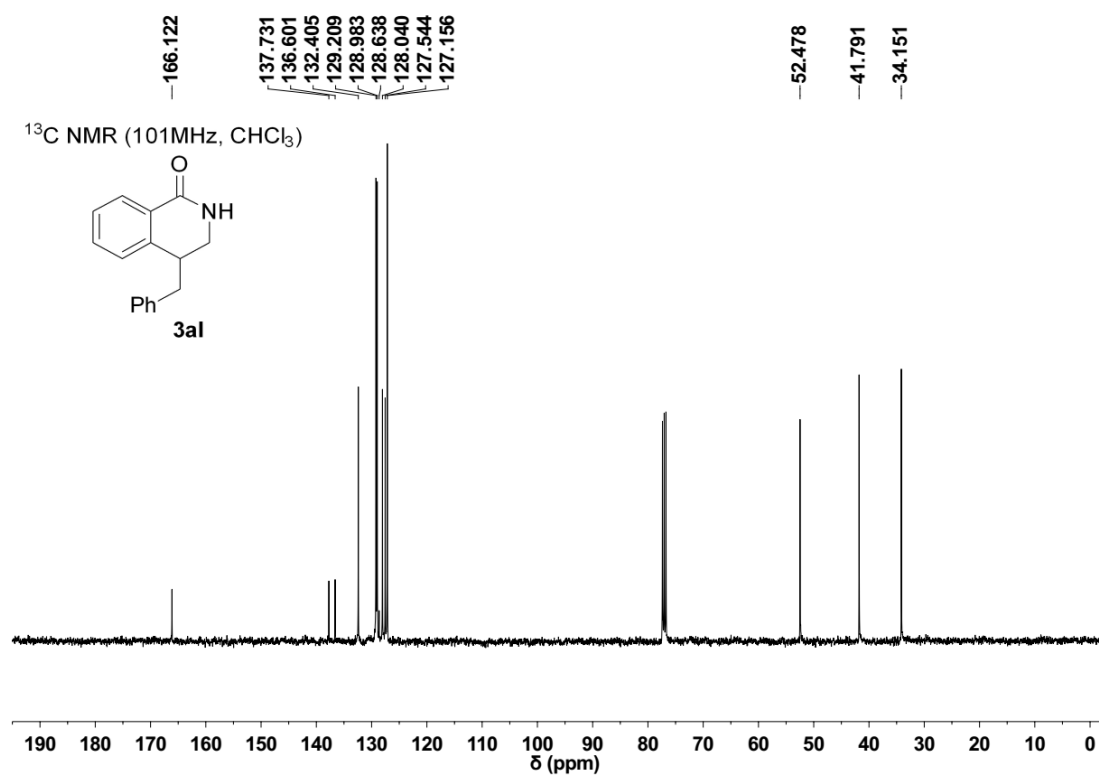
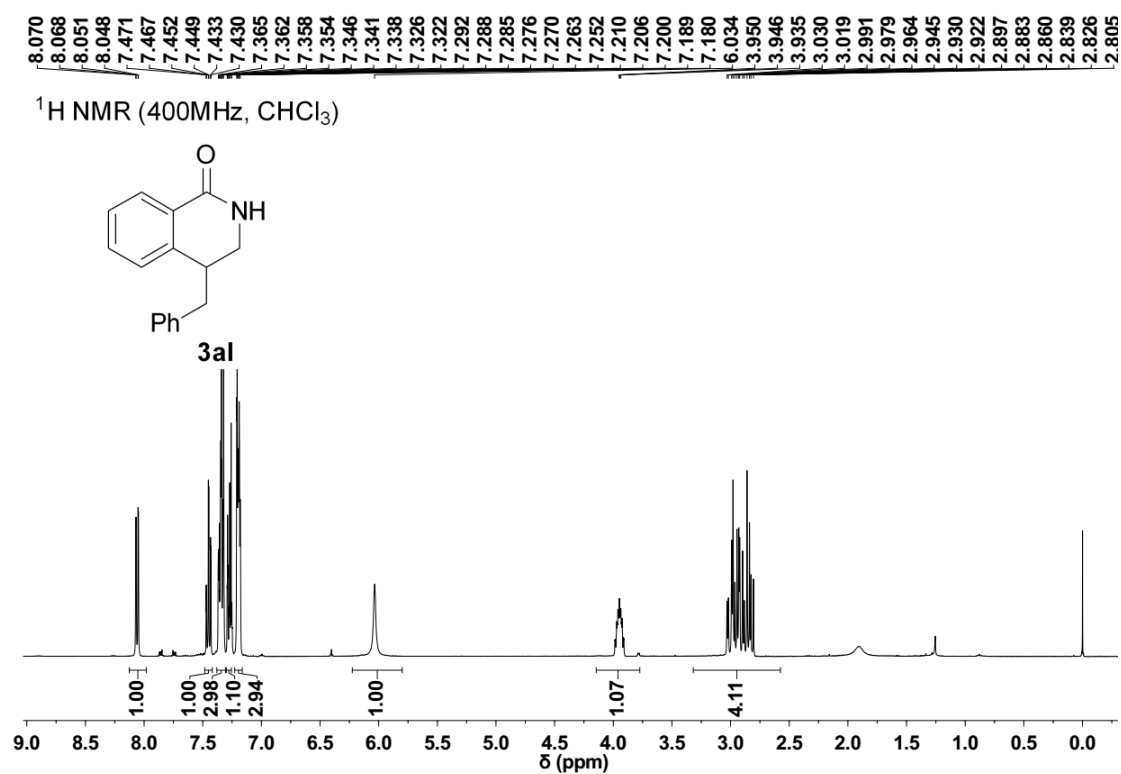


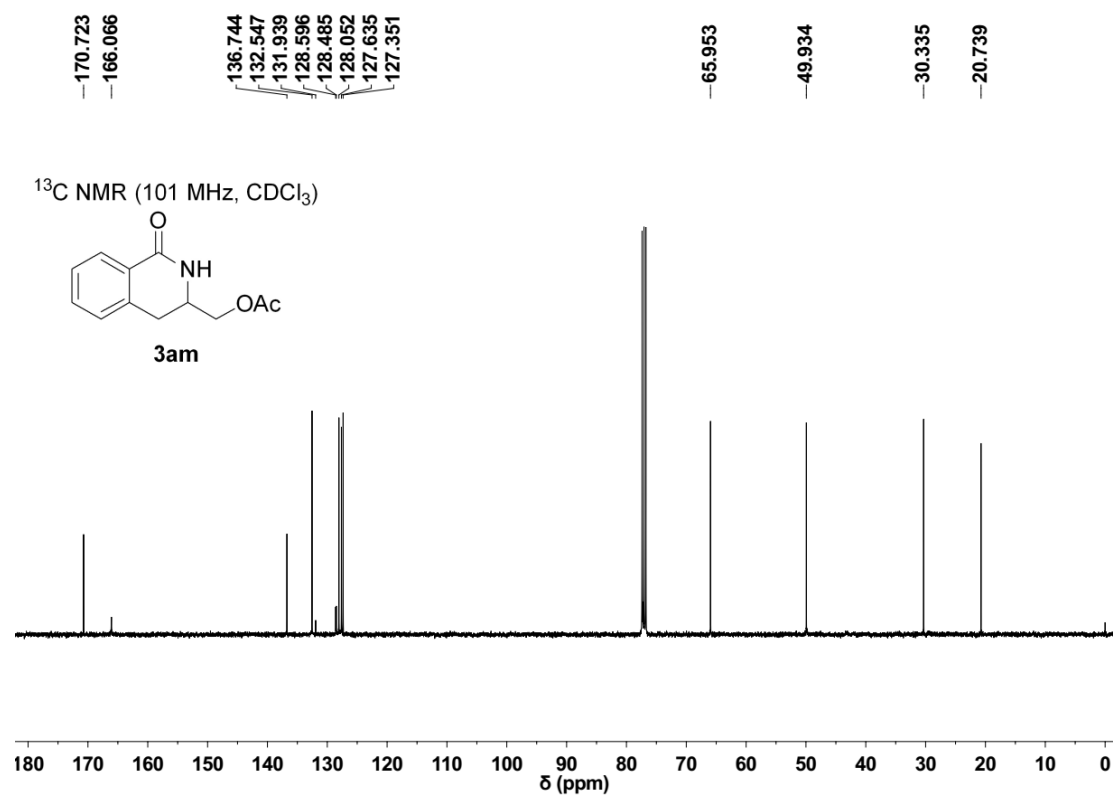
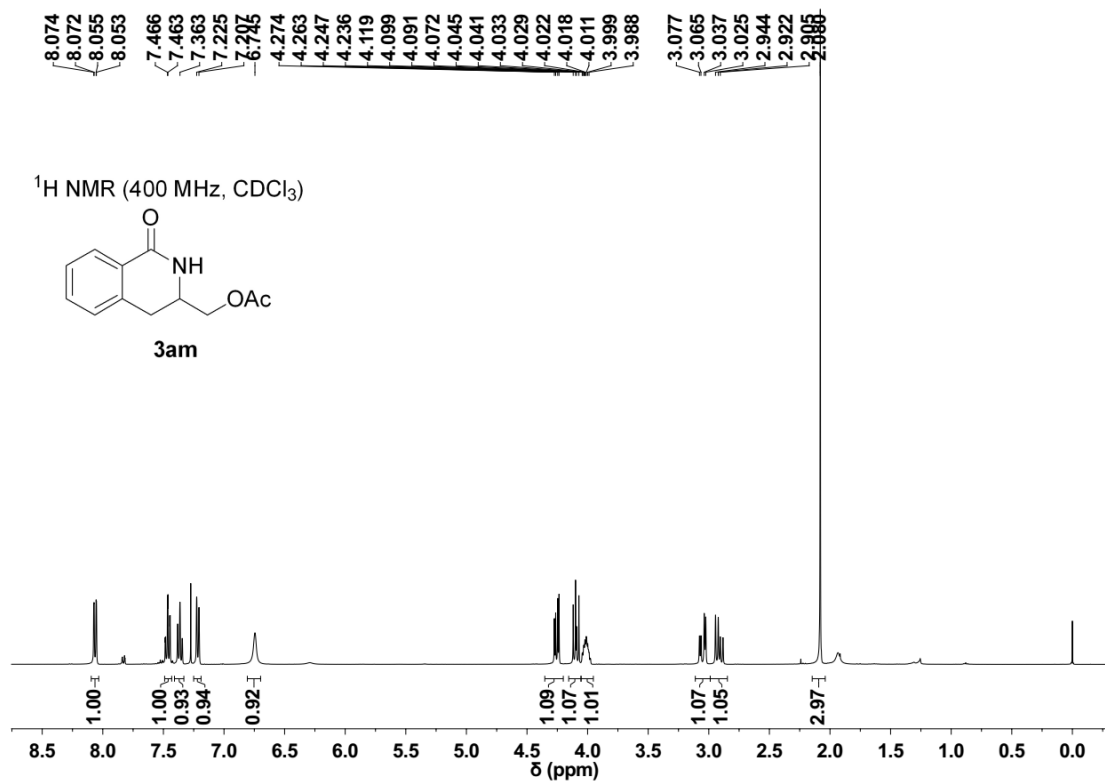


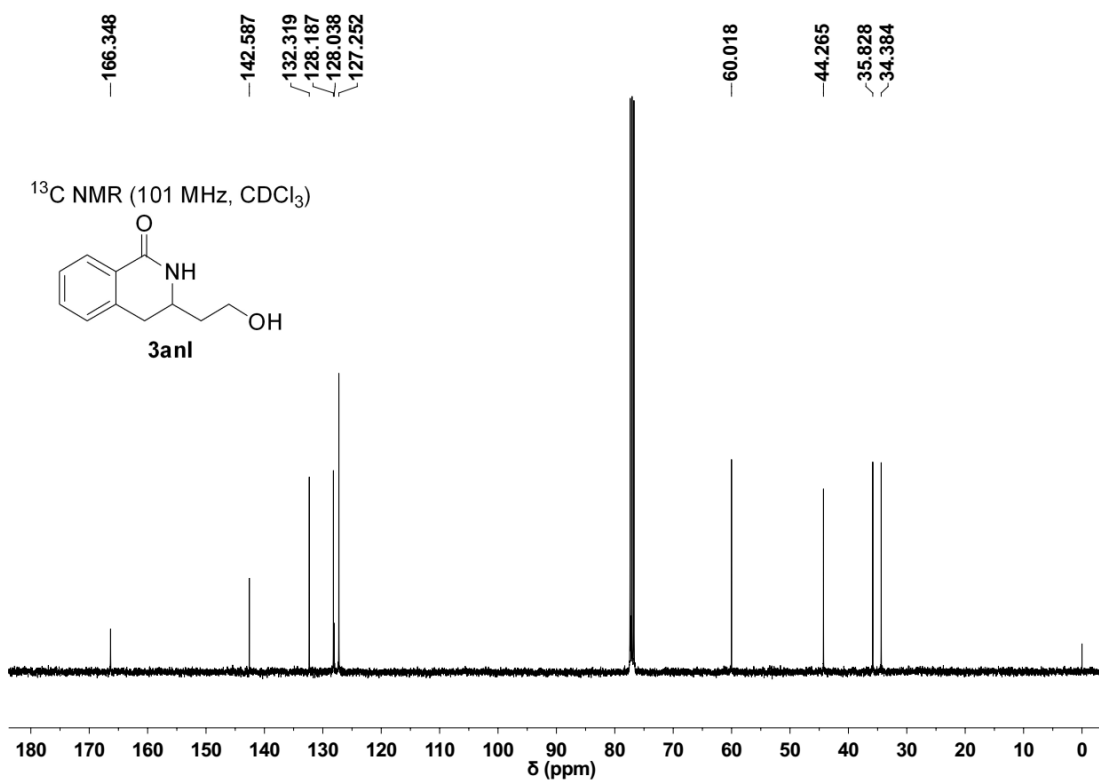
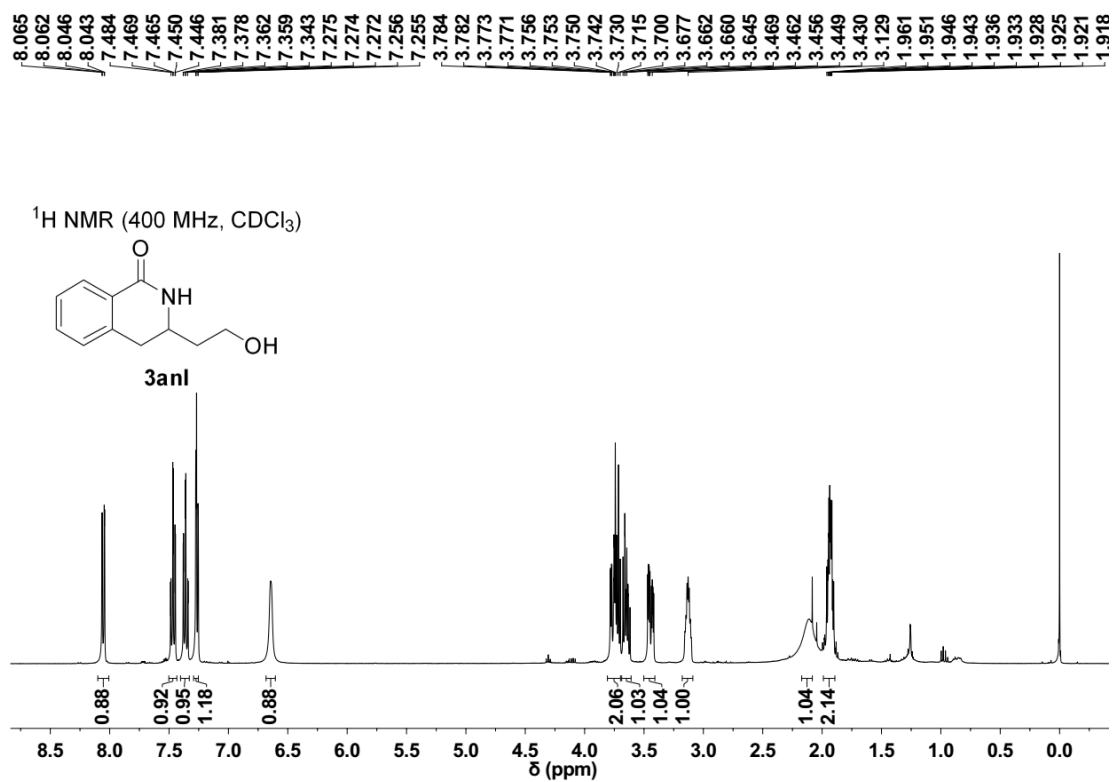






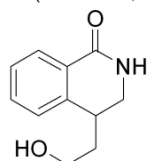




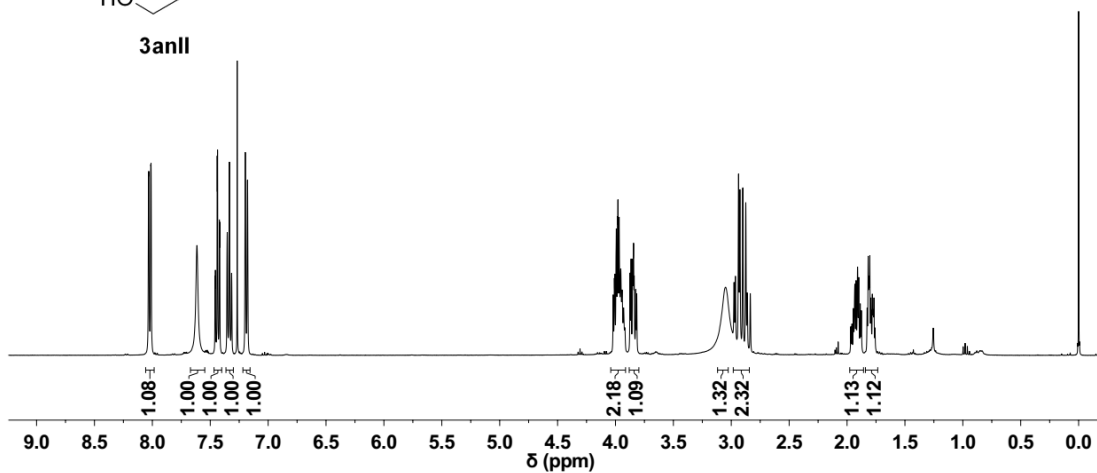


8.031 8.013 8.011 7.615 7.459 7.455 7.440 7.437 7.421 7.418 7.353 7.334 7.316 7.197 7.178 4.020 4.010 4.006 3.993 3.982 3.979 3.968 3.955 3.943 3.874 3.865 3.852 3.844 3.838 3.825 3.816 3.049 2.976 2.964 2.937 2.925 2.901 2.875 2.862 2.836 1.932 1.922 1.909 1.899 1.818 1.812 1.809 1.804 1.794 1.782

^1H NMR (400 MHz, CDCl_3)

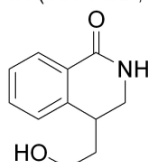


3anII

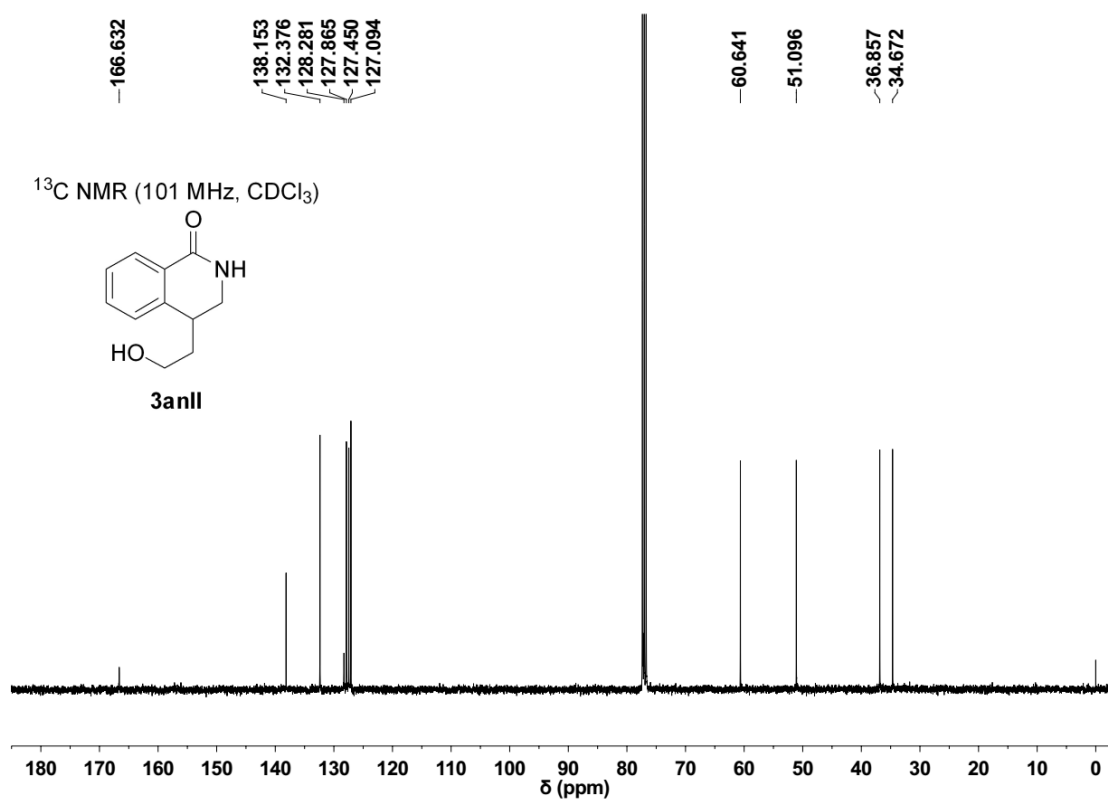


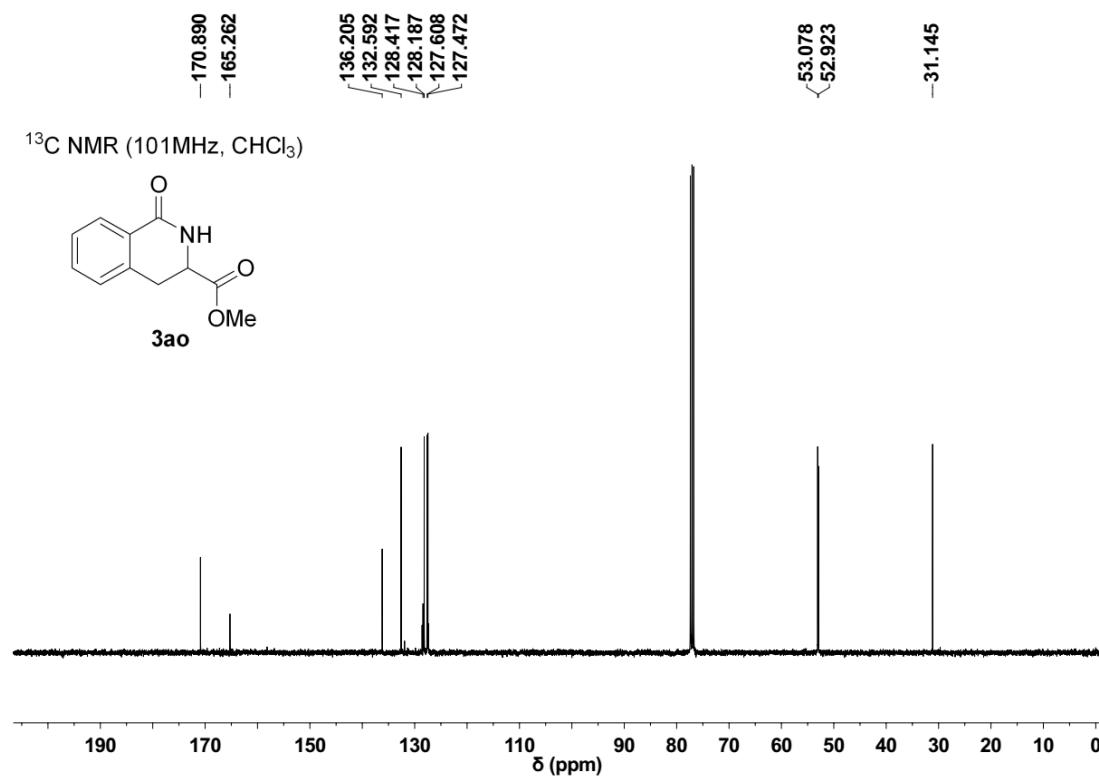
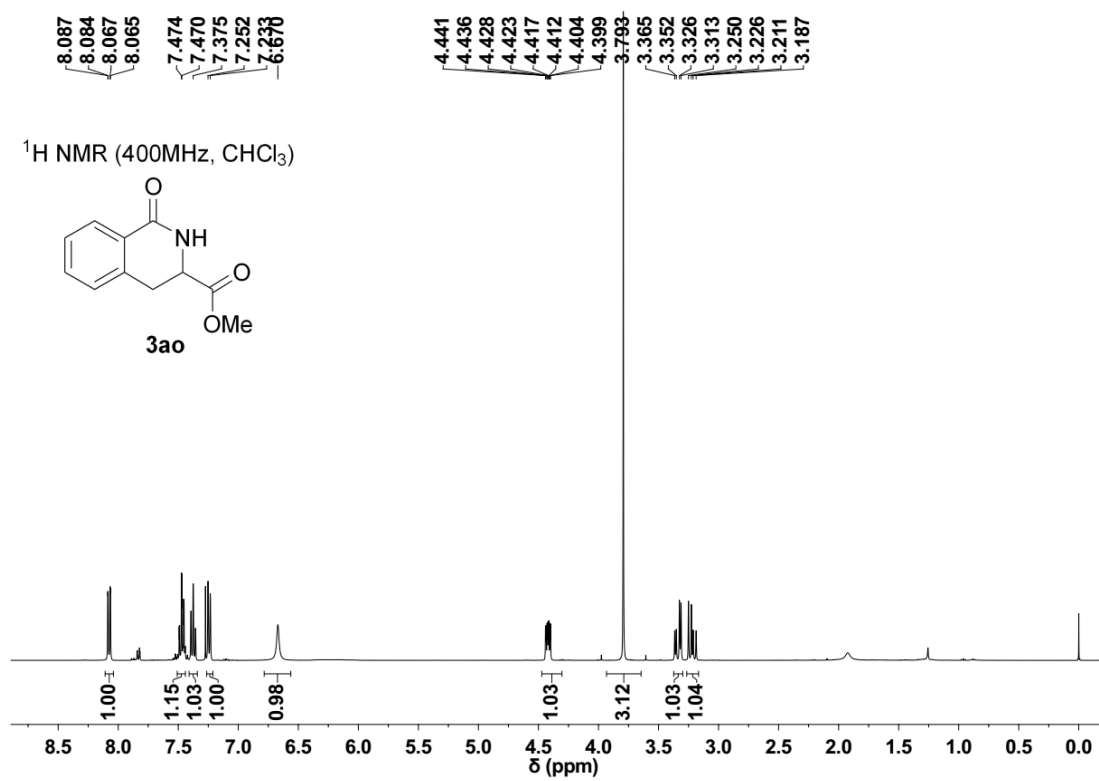
166.632 138.153 132.376 128.281 127.865 127.450 127.094 60.641 51.096 36.857 34.672

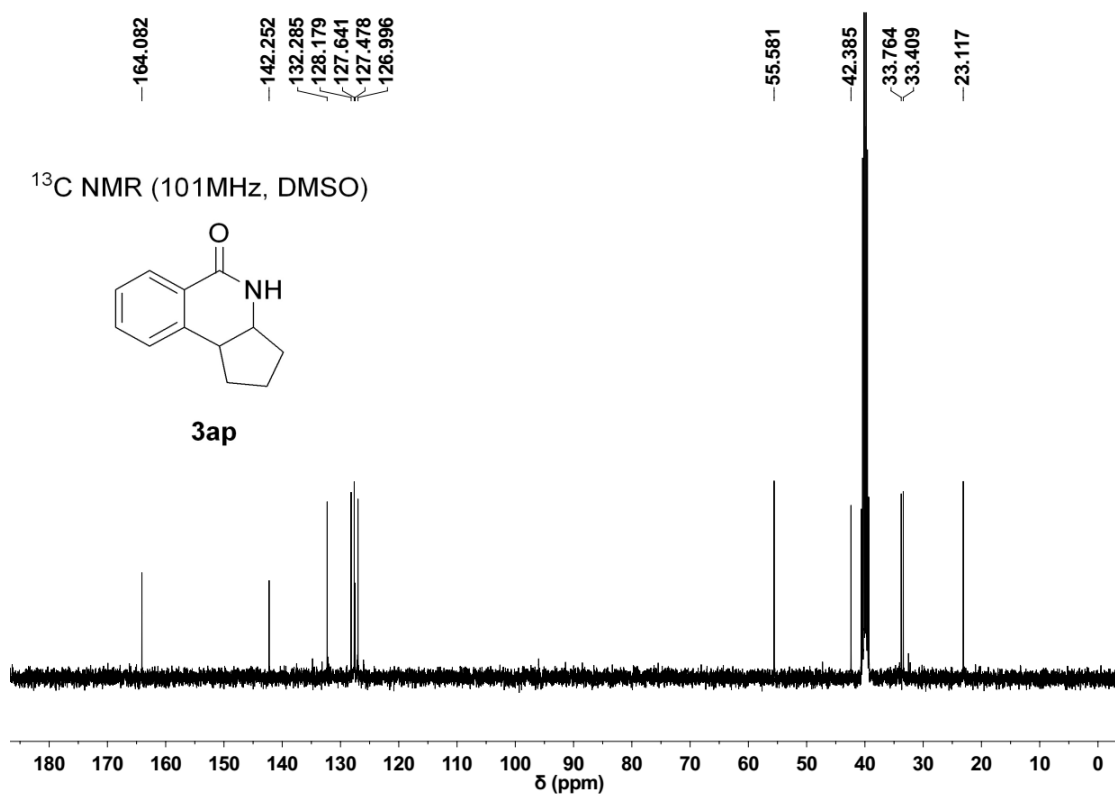
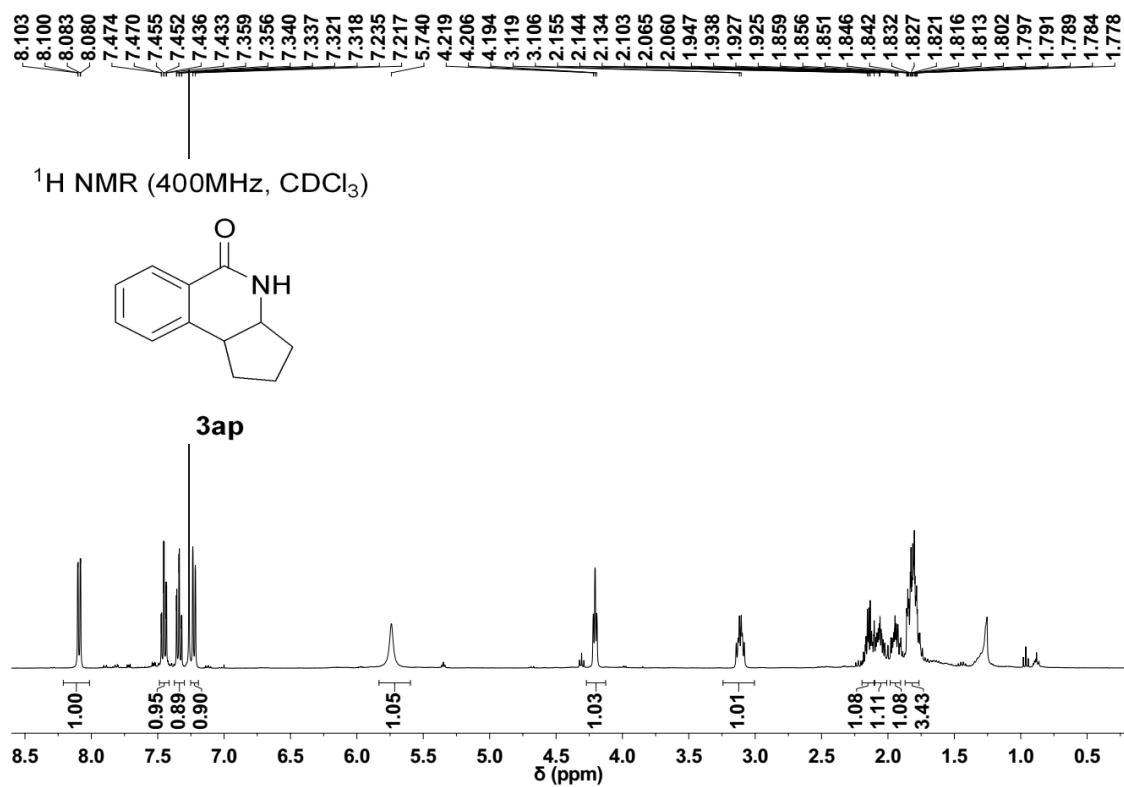
^{13}C NMR (101 MHz, CDCl_3)

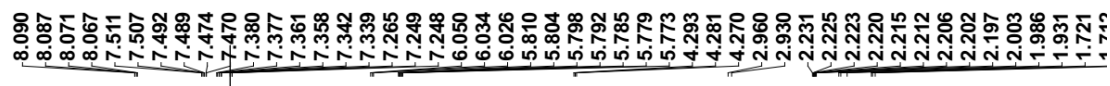


3anII

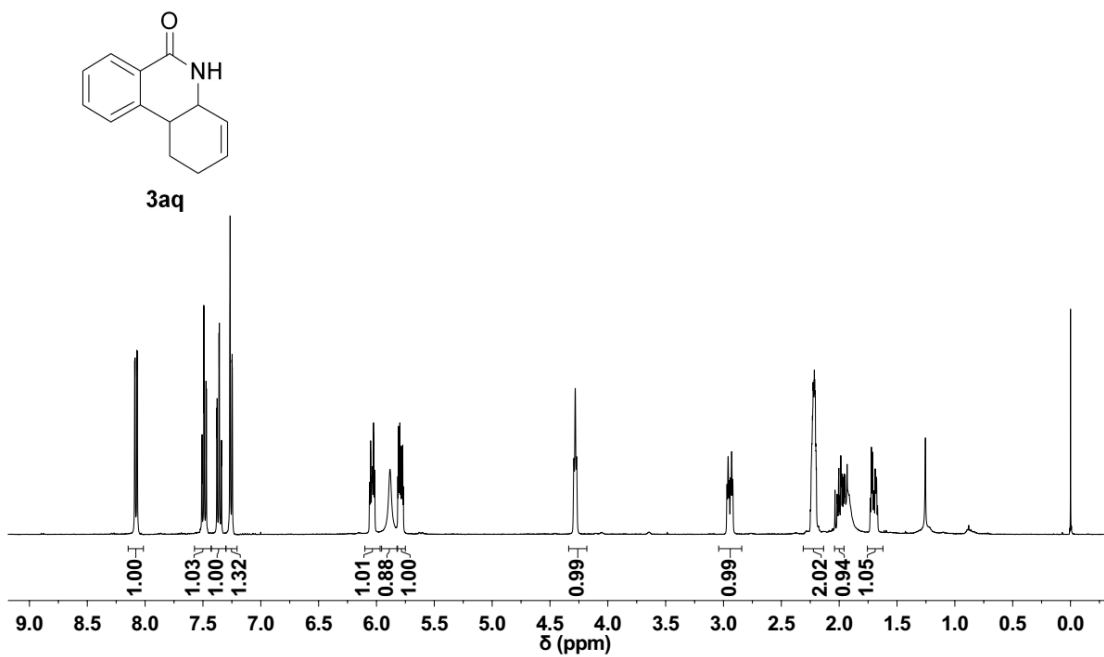








^1H NMR (400 MHz, CHCl_3)



^{13}C NMR (101 MHz, CHCl_3)

