

SUPPLEMENTARY INFORMATION

Analytical method for metabolites involved in biosynthesis of plant volatile compounds

Xiumin Fu^{a, #}, Ying Zhou^{a, #}, Lanting Zeng^a, Fang Dong^b, Xin Mei^a, Yinyin Liao^a, Naoharu Watanabe^c and Ziyin Yang^{a,*}

^a. Key Laboratory of South China Agricultural Plant Molecular Analysis and Genetic Improvement & Guangdong Provincial Key Laboratory of Applied Botany, South China Botanical Garden, Chinese Academy of Sciences, Xingke Road 723, Tianhe District, Guangzhou 510650, China

^b. Guangdong Food and Drug Vocational College, Longdongbei Road 321, Tianhe District, Guangzhou 510520, China

^c. Graduate School of Science and Technology, Shizuoka University, 3-5-1 Johoku, Naka-ku, Hamamatsu 432-8561, Japan

Co-first authors.

*Corresponding author. Ziyin Yang, Tel: +86-20-38072989, Email address: zyyang@scbg.ac.cn.

Supplemental Table S1 The methods for analysis of metabolites involved in biosynthesis of plant volatile compounds

Metabolites		Analysis methods	Advantage	Disadvantage	Ref			
① Metabolites involved in glycolysis and pentose phosphate pathway	G6P	Spectrophotometer analysis	● low cost and easy analysis	● imprecision	S1, S2, S3, S4, S5, S6			
	F6P							
	FBP	GC-MS	● high sensitivity	● requirement for derivatization				
	PEP							
	Pyr		● high reliability					
	3PG							
	2PG	LC-MS	● wide analytical range of metabolites	● lower reproducibility of retention times ● lower accurate quantification				
	DHAP							
	GAP							
② Metabolites involved in formation of volatile phenylpropanoids/benzenoids	3PG+2PG	CE-MS	● small quantity of sample ● without derivatization step ● high mass accuracy and resolution	● poor migration time reproducibility ● lack of reference libraries	S7, S8, S9, S10			
	1,3-PBG							
	organic acid							
	3-deoxy-arabino-heptulonate 7-phosphate							
	3-dehydroquinic acid							
	3-dehydroshikimic acid	GC-MS	● high sensitivity	● requirement for derivatization				
	shikimic acid							
	shikimate 3-phosphate							
	5-enolpyruvylshikimate-3-phosphate (EPSP)							
	chorismic acid							
	prephenic acid							

	phenylpyruvate <i>trans</i> -cinnamic acid <i>para</i> -coumaric acid 3-hydroxy-3-phenylpropionic acid amino acid L-phenylalanine phenolic acid conjugated coenzyme A <i>trans</i> -cinnamoyl CoA 3-hydroxy-3-phenylpropionyl CoA 3-oxo-3-phenylpropionyl CoA	HPLC-PDA	<ul style="list-style-type: none"> ● low cost and easy analysis ● direct analysis 	<ul style="list-style-type: none"> ● limited compounds detected ● poor separation
		HPLC-MS	<ul style="list-style-type: none"> ● direct analysis 	<ul style="list-style-type: none"> ● requirement for sample clean-up
		CE-MS	<ul style="list-style-type: none"> ● simple extraction process ● short analytical time ● small quantity of sample 	<ul style="list-style-type: none"> ● lower sensitivity
③ Metabolites involved in formation of volatile fatty acid derivatives	unsaturated fatty acid linoleic acid linolenic acid saturated fatty acid	GC	<ul style="list-style-type: none"> ● high sensitivity ● short analytical time 	<ul style="list-style-type: none"> ● lower accurate quantitative ● requirement for derivatization ● time-consuming
		HPLC	<ul style="list-style-type: none"> ● accurate quantitative analysis 	<ul style="list-style-type: none"> ● poor retention time reproducibility ● requirement for derivatization ● time-consuming
				S11, S12, S13, S14, S15

		LC-MS	● without derivatization step	● requirement for MS equipment	
	13-Hydroperoxylinolenic acid (12, 13S)-epoxylinolenic acid octadecanoid 12-oxo-phytodienoic acid Jasmonic acid	GC-MS	● high sensitivity	● requirement for derivatization	S16, S17, S18, S19
		LC-MS	● direct analysis	● unavailable of internal standards	
④ Glycosidically bound volatile compounds	Depend on plant species, such as glycosides constituting aglycons of the 3-hexenol, benzyl alcohol, 2-phenylethanol, methyl salicylate, geraniol and linalool	GC-MS	● easy to identify the structures of aglycone	● requirement for derivatization ● hard to identify the sugar residues	S20, S21, S22
⑤ Metabolites involved in formation of carotenoid derived aroma compounds	Phytoene Lycopene β -carotene δ -carotene	LC-PDA	● well separation ● fast detection of known carotenoid compounds	● hard to obtain internal standards ● more purification steps ● complex elution gradient program ● long analysis time	S23, S24
		UPLC-MS	● reduction in analysis time and mobile phase solvent	● poor separation ● requirement for MS equipment	

⑥ Metabolites involved in formation of volatile isoprenoids	DMAPP	CE	● consumption ● identification for unknown carotenoids	
	GPP FPP GGPP	HPLC-MS	● well separating isoprenoids ● direct analysis ● high sensitivity ● direct detection	● lower sensitivity S25, S26

References

- S1. Y. C. Du, A. Nose, K. Wasano and Y. Uchida, *Funct. Plant Biol.*, 1998, **25**, 253.
- S2. R. C. Leegood and R. T. Furbank, *Planta*, 1984, **162**, 450.
- S3. H. Usuda, M. Ku and G. Edwards, *Funct. Plant Biol.*, 1984, **11**, 509.
- S4. J. Liseć, N. Schauer, J. Kopka, L. Willmitzer and A. R. Fernie, *Nat. Protocols*, 2006, **1**, 387.
- S5. H. Yamakawa and M. Hakata, *Plant and Cell Physiol.*, 2010, **51**, 795.
- S6. B. Luo, K. Groenke, R. Takors, C. Wandrey and M. Oldiges, *J. Chromatogr. A*, 2007, **1147**, 153.
- S7. P. Oliveira, J. A. Pereira, P. B. Andrade, P. Valentão, R. M. Seabra and B. M. Silva, *Food Chem.*, 2008, **111**, 393.
- S8. Magnes, M. Suppan, T. R. Pieber, T. Moustafa, M. Trauner, G. Haemmerle and F. M. Sinner, *Anal. Chem.*, 2008, **80**, 5736.
- S9. Mardones, A. Hitschfeld, A. Contreras, K. Lepe, L. Gutiérrez and D. von Baer, *J. Chromatogr. A*, 2005, **1085**, 285.
- S10. W. Klampfl, W. Buchberger and P. R. Haddad, *J. Chromatogr. A*, 2000, **881**, 357.
- S11. B. Qi, T. Fraser, S. Mugford, G. Dobson, O. Sayanova, J. Butler, J. A. Napier, A. K. Stobart and C. M. Lazarus, *Nat. Biotech*, 2004, **22**, 739.
- S12. J. Browse, P. J. McCourt and C. R. Somerville, *Anal. Biochem.*, 1986, **152**, 141.
- S13. S.H. Chen and Y.-J. Chuang, *Anal. Chim. Acta*, 2002, **465**, 145.
- S14. T. Řezanka, *J. High. Resolut. Chromatogr.*, 2000, **23**, 338
- S15. A. Carrier and J. Parent, *J. Liq. Chromatogr. Rel. Technol.*, 2001, **24**, 97.

- S16. A. Müller, P. Düchting and E. W. Weiler, *Planta*, 2002, **216**, 44.
- S17. M. J. Mueller and W. Brodschelm, *Anal. Biochem.*, 1994, **218**, 425.
- S18. B. A. Stelmach, A. Müller, P. Hennig, D. Laudert, L. Andert and E. W. Weiler, *Phytochemistry*, 1998, **47**, 539.
- S19. M. O. Funk, R. Isaac and N. A. Porter, *Lipids*, 1976, **11**, 113.
- S20. Y. Zhou, F. Dong, A. Kunimasa, Y. Zhang, S. Cheng, J. Lu, L. Zhang, A. Murata, F. Mayer, P. Fleischmann, N. Watanabe and Z. Yang, *J. Agric. Food Chem.*, 2014, **62**, 8042.
- S21. C. C. Chyau, P. T. Ko, C. H. Chang and J. L. Mau, *Food Chem.*, 2003, **80**, 387.
- S22. Y. Z. Gunata, C. L. Bayonove, R. L. Baumes and R. E. Cordonnier, *J. Chromatogr. A*, 1985, **331**, 83.
- S23. P. D. Fraser, M. E. Pinto, D. E. Holloway and P. M. Bramley, *Plant J.*, 2000, **24**, 551.
- S24. B. Y. Hsu, Y. S. Pu, B. S. Inbaraj and B. H. Chen, *J. Chromatogr. B*, 2012, **899**, 36.
- S25. G. Nürenberg and D. A. Volmer, *Anal. Bioanal. Chem.*, 2012, **402**, 671.
- S26. L. Henneman, A. G. van Cruchten, S. W. Denis, M. W. Amolins, A. T. Placzek, R. A. Gibbs, W. Kulik and H. R. Waterham, *Anal. Biochem.*, 2008, **383**, 18.