Simple and efficient Fmoc removal in ionic liquid.

Maria Luisa Di Gioia, *^a Antonio De Nino,^b Loredana Maiuolo,^b Monica Nardi, ^{b,c} Fabrizio Olivito^d and Antonio Procopio^d

^aDipartimento di Farmacia e Scienze della Salute e della Nutrizione, Università della Calabria, Arcavacata di Rende (CS), 87036, Italy

^b Dipartimento di Chimica, Università della Calabria, Cubo 12C, 87036, Arcavacata di Rende (CS), Italy

^c Dipartimento di Agraria, Università Telematica San Raffaele, Via di Val Cannuta, 00166,247, Rome, Italy

^dDipartimento di Scienze della Salute, Università Magna Graecia, Viale Europa, 88100, Germaneto (CZ), Italia.

Supplementary Material

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Experimental Section

Commercially available reagents were purchased from Sigma-Aldrich Chemical Co. (Milano, Italy) and used as supplied unless stated otherwise. All syntheses were carried out in atmospheric conditions. ¹H NMR spectra were recorded at 300 MHz, while ¹³C NMR spectra were measured at 75 MHz. Spectral analysis was performed at 293 K on diluted solutions of each compound by using CDCl₃ as the solvent. Chemical shifts (δ) are reported in ppm and referenced to CDCl₃ (7.25 ppm for ¹H and 77.0 ppm for ¹³C spectra). Coupling costants (J) are reported in Hertz (Hz). Reaction mixtures were monitored by thin layer chromatography (TLC) using Merck Silica gel 60-F₂₅₄ precoated glass plates, and UV light (254 nm) or 0.2% ninhydrin in ethanol and charring as visualizing agent. Evaporation of solvents was performed at reduced pressure using a rotary vacuum evaporator. Chiral GC analysis were carried out using a Thermo Gas Chromatograph instrument. Chiral GC analyses of enantiomeric compounds Ac-DL-AlaOMe and Ac-L-AlaOMe were performed by using a 25 m × 0.25 mm, diethyl tertbutyldimethylisilyl- β -cyclodextrine chiral capillary column.

The GC-MS Shimadzu workstation is constituted by a GC 2010 (provided of a 30 m-QUADREX 007–5MS capillary column, operating in "split" mode, 1 ml min–1 flow of He as carrier gas) and a 2010 quadrupole mass-detector. LC-MS analysis were carried using an Agilent 6540 UHD Accurate - Mass Q-TOF LC–MS (Agilent, Santa Clara, CA) fitted with a electrospray ionisation source (Dual AJS ESI) operating in positive ion mode. Chromatographic separation was achieved using a C18 RP analytical column (Poroshell 120, SB-C18, 50×2.1 mm, 2.7μ m) at 30°C with a elution gradient from 5% to 95% of B over 13 min., A being H₂O (0.1% FA) and B CH₃CN (0.1% FA). Flow rate was 0.4 ml/min.

N-Fmoc amines **1a–i** and *N*-Fmoc α -amino acid methyl esters **1j–u** were prepared according to previously published protocol.^{14b} Spectral data of **1a–p** agreed with those already reported for the same compounds prepared as previously reported.^{14b}

Spectroscopic data (1q-1u)

N-(9-Fluorenylmethoxycarbonyl) glycine *t*-butyl ester (1q): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 7.68- 7.52 (m, 4 H, Ar*H*), 7.32-7.22 (m, 4H, Ar*H*), 6.38 (br s, 1H, N*H*), 3.97 (t, *J* = 6.0 Hz, 1 H, C*H*_{Fmoc}), 3.82 (d, *J* = 5.1 Hz, 2 H, C*H*_{2Fmoc}), 3.23 (s, 2 H, C*H*₂), 1.38 (s, 9H, C(C*H*₃)₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 171.8, 8, 156.1, 145.9, 141.9, 128.9, 127.5, 124.8, 120.2, 81.4, 52.7, 42.4, 31.7, 28.3.

HRMS (ESI) for $(C_{21}H_{13}NO_4) + Na^+$: calcd 376.1525, found 376.1518 (M+Na)+

N-(9-Fluorenylmethoxycarbonyl) phenylalanine benzyl ester (1r): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 7.78 (d, *J*= 7.2 Hz, 2 H, Ar*H*), 7.62 (d, *J*= 7.2 Hz, 2H, Ar*H*), 7.35- 7.17 (m, 12 H, Ar*H*), 7.00-6.97 (m, 2H, Ar*H* + N*H*), 5.12 (dd, *J*= 12.0 Hz, *J* = 9.6 Hz, 1 H, α -C*H*), 4.82- 4.79 (m, 2 H, OC*H*₂Ph), 4.14 (m, 1 H, C*H*_{Fmoc}), 4.10-4.04 (m, 2 H, C*H*_{2Fmoc}), 3.01-3.05 (m, 2 H, C*H*₂Ph) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 172.1, 8, 158.6, 144.2, 141.6, 137.0, 136.0, 135.9, 129.4, 128.6, 128.5, 128.4, 127.6, 126.9, 124.7, 120.0. HRMS (ESI) for (C₃₁H₂₇NO₄)+ Na]⁺ : calcd 500.1838, found 500.1824 (M+Na)⁺

N-(9-Fluorenylmethoxycarbonyl) tyrosine (*O*-*t*-butyl) *t*-butyl ester (1s): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 7.75 (d, 2 H, *J*= 7.5, Ar*H*), 7.58 (d, 2H, *J* = 7.4, Ar*H*), 7.42-7.31 (m, 4H, Ar*H*), 7.11-7.03 (m, 2 H, Ar*H*), 6.93-6.88 (m, 2H, Ar*H*), 5.30 (d, 1H, *J* = 8.4 Hz, N*H*), 4.54-4.39 (m, 3 H, α-C*H* + C*H*_{2Fmoc}), 4.34 (t, 1H, *J* = 8.6, C*H*_{Fmoc}), 3.04 (m, 1 H, β-C*H*₂), 2.80 (m, 1 H, β-C*H*₂), 1.40 (s, 9H, C(C*H*₃)₃), 1.39 (s, 9H, C(C*H*₃)₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 170.6, 155.5, 154.2, 144.5, 143.9, 134.6, 129.1, 127.7, 127.0, 124.3, 120.3, 119.7, 82.6, 81.1, 66.9, 55.2, 47.3, 37.9, 28.8, 27.9. HRMS (ESI) for (C₃₂H₃₇NO₅)+ H]⁺ : calcd 516.2750, found 516.2664, 538.2570 (M+Na)⁺.

N-(9-Fluorenylmethoxycarbonyl) glutamic acid (*O-tert*-butyl) methyl ester (1t): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 7.69 (d, *J* = 7.5 Hz, 2 H, Ar*H*), 7.52 (d, *J*= 4.8 Hz, 2H, Ar*H*), 7.35-7.21 (m, 4H, Ar*H*), 5.42 (d, *J*= 8.1 Hz, 1 H, N*H*), 4.74-4.38 (m., 3 H, α-C*H*+ C*H*_{2Fmoc}), 3.68 (s, 3H, OC*H*₃), 2.32–2.22 (m, 2 H, β-C*H*₂), 2.13 (m, 1H, γ-C*H*), 2.06 (m, 1H, γ-C*H*), 1.37 (s, 9 H, C(C*H*₃)₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 171.4, 170.9, 154.9, 142.7, 140.3, 126.6, 126.0, 124.0, 118.9, 79.8, 66.0, 52.4, 51.4, 46.1, 30.4, 27.0, 26.2 HRMS (ESI) for (C₂₅H₂₉NO₆)+ H]⁺: calcd 440,2073, found 440.2135, 462.1891 (M+ Na)⁺.

N-(9-Fluorenylmethoxycarbonyl) lysine (*N*-Boc) methyl ester (1u): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 7.53 (d, *J* = 6.9 Hz, 2 H, Ar*H*), 7.33 (d, *J* = 7.5 Hz, 2H, Ar*H*), 7.26-7.19 (m, 4H, Ar*H*), 5.39 (d, *J* = 7.8 Hz, 1 H, N*H*), 4.53 (m, 1 H, N*H*), 4.34-4.27 (m, 3 H, α -C*H* + C*H*_{2Fmoc}), 4.17 (t, *J* = 6.9 Hz, 1H, C*H*_{Fmoc}), 3.67 (s, 3H,

OCH₃), 3.04- 3.02 (m, 2 H, ε-CH₂), 1.77 (m, 1H, β-CH), 1.63 (m, 1H, β-CH), 1.42-1.16 (m, 4H, γ-CH₂ + δ-CH₂), 1.36 (s, 9 H, C(CH₃)₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 172.9, 156.1, 155.9, 143.7, 141.3, 127.6, 127.0, 125.1, 119.9, 80.0, 66.9, 53.7, 52.4, 47.1, 40.2, 32.1, 29.6, 28.4, 22.3.

HRMS (ESI) for $[(C_{27}H_{34}N_2O_6 + H]^+$: calcd 483,2495 found 483.2484, 505.2306 $(M+Na)^+$.

General procedure for the *N*-Fmoc removal of amines 1a-i in [Bmim][BF₄].

To a magnetically stirred mixture of *N*-Fmoc protected amines **1a-i** (1 mmol) and $[Bmim][BF_4]$ (1 mL), Et₃N (3 mmol) was added and the mixture was stirred at room temperature for 4-8 min. TLC monitored the reaction. Diethyl ether was added after the completion of reaction and the IL settled at the bottom. The supernatant was decanted off and the IL was washed with Et₂O (3 × 2 mL). The combined Et₂O extracts were acidified with an aqueous solution of 1N HCl and separated. The aqueous phase was then basified with sat. aq NaHCO₃ and finally extracted with diethyl ether. The organic phase was dried over Na₂SO₄ and filtered. The products were isolated after evaporation of the diethyl ether to yield the free amines **2a-i** in 80-93 % yields. Spectroscopic data showed full consistency of the spectra with the pure products.

General procedure for the *N*-Fmoc removal of amino acid methyl esters 1j-u in [Bmim][BF₄].

To a magnetically stirred mixture of *N*-Fmoc amino acid methyl esters **1**j-u (1 mmol) and $[Bmim][BF_4]$ (1 mL), Et₃N (3 mmol) was added and the mixture was stirred at room temperature for 8-15 min. TLC monitored the reaction. Diethyl ether was added after the completion of reaction and the IL settled at the bottom. The supernatant was decanted off and the IL was washed with Et₂O (3×2 mL). The combined Et₂O extracts were acidified with an aqueous solution of 1N HCl (for compounds **1q-u** bearing acid-sensitive protecting group a 5% aqueous solution of citric acid was used) and separated. The aqueous phase was then basified with sat. aq NaHCO3 and finally extracted with diethyl ether. The organic phase was dried over Na₂SO₄ and filtered. The products were isolated after evaporation of the diethyl ether to yield the free amino acid methyl ester 2j-u in 75-88% yields. Compounds 2j-u were acetylated in order to perform GC/MS analysis. N-Acetylation was achieved dissolving 2j-u in DCM (5 mL) and adding acetic anhydride (1 mL) and a 9% aqueous solution of NaHCO₃ (5 mL). The mixture was maintained under magnetic stirring at room temperature for 4 h. The organic layer was separated and the aqueous phase was extracted with three additional portions of DCM (3 x 10 mL). The combined organic layers were washed with a 9% aqueous solution of NaHCO₃, twice with aqueous HCl 1 N (or a 5% aqueous solution of citric acid), once with brine and finally dried (Na₂SO₄). The solvent was evaporated under reduced pressure to afford

the corresponding *N*-acetyl derivatives 3j-u as colourless oil in quantitative yield. Spectroscopic data for 3j-n compared to those reported in the literature.^{2a}

Spectroscopic data (3j-3u).

N-Acetyl alanine methyl ester (3j): ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 6.28$ (s, 1 H, N*H*), 4.58 (m, 1 H α -C*H*), 3.70 (s, 3 H, OC*H*₃), 2.02 (s, 3 H, C*H*₃CO), 1.40 (d, *J* = 7.2 Hz, 3 H, *CH*₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): $\delta = 173.7$, 169.5, 52.4, 48.0, 23.1, 18.6 ppm. GC/MS (EI): *m/z* (%) 145 (13) [(M)⁺], 102 (86 (70), 59 (5), 44 (100). HRMS (ESI) for ([C₆H₁₁NO₃] + Na)⁺ : calcd 168.0637, found 168.0630 [M+Na]⁺.

N-Acetyl valine methyl ester (3k): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 6.22 (d, 1 H, N*H*, *J* = 6.6 Hz), 4.55 (dd, *J* = 8.7, *J* = 5.1 Hz, 1 H, α-C*H*), 3.74 (s, 3 H, OC*H*₃), 2.13 (m, 1 H, C*H*(CH₃)₂), 2.01 (s, 3 H, C*H*₃CO), 0.93 (d, *J* = 6.9 Hz, 3 H, CH(*CH*₃)₂), 0.90 (d, *J* = 6.9 Hz, 3 H, CH(*CH*₃)₂) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ= 172.7, 170.2, 57.1, 52.1, 31.2, 23.1, 18.8, 17.8 ppm. GC/MS (CI): *m/z* (%) 214 (13) [(M + C₃H₅)⁺], 202 (16) [(M + C₂H₅)⁺], 174 (60) [(M + H)⁺], 156 (9), 142 (65), 132 (50), 114 (100), 101 (7). HRMS (ESI) for ([C₈H₁₅NO₃] + H)⁺ : calcd. 174.1130, found 174.1134 [M+H]⁺.

N-Acetyl leucine methyl ester (3l). ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 6.50$ (d, J = 7.68 Hz, 1H, NH), 4.52 (m, 1 H, α -CH), 3.69 (s, 3 H, OCH₃), 1.99 (s, 3 H, CH₃CO), 1.62-1.40 (m, 3 H, CH₂CH), 1.40–1.26 (m, 1 H, CH₂), 0.85-0.87 (m, 6 H,

CH(*CH*₃)₂) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 173.8, 170.1, 52.1, 50.6, 41.3, 24.7, 22.8, 22.7, 21.8 ppm. GC/MS (CI): *m/z* (%) 228 (20) [(M + C₃H₅)⁺], 216 (35) [(M + C₂H₅)⁺], 188 (100) [(M + H)⁺], 170 (5), 156 (60), 146 (55), 128 (88), 86 (9).

HRMS (ESI) for $([C_9H_{17}NO_3] + Na)^+$: calcd. 210.1106, found 210.1103 [M+Na]⁺.

N-Acetyl isoleucine methyl ester (3m): ¹H NMR (300 MHz, CDCl₃, 25 °C): δ = 6.06 (s, 1 H, *N*H), 4.54 (d, *J* = 8.7 Hz, *J* = 4.8 Hz, 1 H, α-C*H*), 3.67 (s, 3 H, OC*H*₃), 1.96 (s, 3H, C*H*₃CO), 1.78 (m, 1 H, β-C*H*), 1.35 (ddd, *J* = 7.5 Hz, *J* = 4.8 Hz, *J* = 4.8 Hz, 1H, C*H*₂), 1.12 (m, 1 H, C*H*₂), 0.85 (s, 3 H, C*H*₃), 0.82 (s, 3 H, C*H*₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 172.7, 169.8, 56.4, 51.9, 37.9, 25.2, 23.2, 15.3, 11.5 ppm. GC/MS (CI): *m/z* (%) 228 (51) [(M + C₃H₅)⁺], 216 (32) [(M + C₂H₅)⁺], 188 (100) [(M + H)⁺], 157 (76), 146 (83), 128 (68), 102 (2). HRMS (ESI) for ([C₉H₁₇NO₃] + H)⁺ 188.1287, found 188.1279 [M+H]⁺, 210.1099 [M+Na]⁺.

N-Acetyl phenylalanine methyl ester (3n): ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 7.32-7.24$ (m, 3 H, Ar*H*), 7.09 (dd, J = 7.9 Hz, J = 1.8 Hz, 2H, Ar*H*), 6.02 (d, J = 6.6 Hz, 1 H, *NH*,), 4.88 (dt, J = 7.8 Hz, J = 5.8 Hz, 1 H, α-*CH*), 3.72 (s, 3 H, OC*H*₃), 3.14 (dd, J = 13.83 Hz, J = 5.7 Hz, 1H, β-*CH*), 3.07 (dd, J = 13.83 Hz, J = 5.7 Hz, 1H, β-*CH*), 1.99 (s, 3 H, *CH*₃CO) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): $\delta = 172.0$, 169.5, 135.8, 129.2, 128.5, 127.1, 53.1, 52.2, 37.8, 23.0 ppm. HRMS (ESI) for ([C₁₂H₁₅NO₃] + Na)⁺ 244.0950, found 244.0941 [M+Na]⁺.

N-Acetyl *N*-methyl valine methyl ester (**3o**) (two rotamers): ¹H NMR (300 MHz, CDCl₃, 25 °C), (two rotamers): $\delta = 4.88$ (d, 1 H, *J*= 10.5 Hz, α-*CH*), 3.69 and 3.65 (2s, 3 H, OC*H*₃), 2.94 and 2.82 (2s, 3 H, *N*-*CH*₃), 2.16 and 2.12 (2s, 3 H, *CH*₃CO), 2.27–2.19 (m, 1 H, β-*CH*), 0.95 and 0.94 (d, *J* = 6.6 Hz, 3 H, CH(*CH*₃)₂), 0.86 and 0.82 (d, *J* = 6.9 Hz, 3 H, CH(*CH*₃)₂] ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 170.86, 169.58, 66.21, 60.32, 51.01, 31.27, 26.73,21.06, 19.00, 18.82 ppm. GC/MS (CI): *m/z* (%) 228 (51) [(M + C₃H₅)⁺], 216 (32) [(M + C₂H₅)⁺], 188 (100) [(M + H)⁺], 157 (76), 146 (83), 128 (68), 102 (2).

N-Acetyl *N*-methyl isoleucine methyl ester (**3**p). ¹H NMR (300 MHz, CDCl₃, 25 °C), (two rotamers): $\delta = 4.99$ (d, J = 10.5 Hz, 1 H, α -CH), 3.71 and 3.67 (2s, 3 H, OCH₃), 2.95 and 2.83 (2s, 3 H, N-CH₃), 2.14 and 2.10 (2s, 3 H, CH₃CO), 2.00–1.90 (m, 1 H, β -CH), 1.40–1.26 (m, 1 H, CH₂), 1.10–1.00 (m, 1 H, CH₂), 0.97 and 0.96 (2d, J = 6.9 Hz, 3 H, CH₃) ppm.

N-Acetyl glycine *t*-butyl ester (3q). ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 3.92$ (d, *J*= 5.1 Hz, 2 H, α -C*H*), 2.04 (s, 3 H, C*H*₃), 1.47 (s, 9 H, C(C*H*₃)₃), ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): $\delta = 170.3$, 169.3, 82.3, 42.1, 28.0, 22.9 ppm. HRMS (ESI) for [(C₈H₁₅NO₃) + H]⁺ 174.1130, found 174.1122 [M+H]⁺, 196.0945 [M+Na]⁺.

N-Acetyl phenylalanine benzyl ester (3r). ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 7.35-7.21$ (m, 8 H, Ar*H*), 7.00 – 6.97 (m, 2 H, Ar*H*), 6.02 (d, J = 6.9 Hz, 1H, NH), 5.14-5.13 (m, 2 H, COOC*H*₂), 4.93 (m, 1 H, α -C*H*), 3.12-3.09 (m, 2 H, C*H*₂Ph), 1.96 (s, 3 H, C*H*₃CO) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): $\delta = 171.7$, 169.9,

135.8, 135.2, 129.4, 128.8, 128.7, 128.6, 127.2, 67.4, 53.3, 37.9, 23.2 ppm. HRMS (ESI) for $(C_{18}H_{19}NO_3) + H]^+$: calcd 298.1443, found 298.1437 $[M + H]^+$, 320.1254 $[M + Na]^+$.

N-Acetyl tyrosine (*O*-*t*-butyl) *t*-butyl ester (3s). ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 7.31$ (d, J = 8.4 Hz, 2 H, Ar*H*), $\delta = 7.90$ (d, J = 8.4 Hz, 2 H, Ar*H*), 6.04 (br s, s, 1 H, N*H*), 4.74 (dd, J = 6.6 Hz, J = 5.7 Hz, 1 H, α -C*H*), 3.09–3.02 (m, 2 H, β -C*H*), 2.02 (s, 3 H, C*H*₃CO), 1.38 (s, 9 H, C(C*H*₃)₃), 1.40 (s, 9 H, C(C*H*₃)₃),ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): $\delta = 170.9$, 169.3, 146.4, 131.1, 129.9, 124.0, 90.8, 82.4, 53.6, 37.5, 28.8, 27.9, 23.2 ppm. HRMS (ESI) for [(C₁₉H₂₉NO₄] + H)⁺ : calcd 336.2175, found 336.2133 [M + H]⁺, 358.1987 [M + Na]⁺.

N-Acetyl glutamic acid (*O-t*-butyl) methyl ester (3t): ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 6.34$ (d, J = 7.5 Hz, 1 H, NH), 4.74 (dt, J = 8.0 Hz, J = 5.1 Hz, 1 H, α-*CH*), 3.67 (s, 3H, OCH₃), 2.35–2.15 (m, 2 H, β-CH₂), 2.05 (ddd, J = 14.1 Hz, J = 7.3Hz, J = 2.2 Hz, 1H, γ-CH), 1.95 (s, 3 H, CH₃CO), 1.87 (ddd, J = 14.1 Hz, J = 6.1Hz, J = 1.3 Hz, 1H, γ-CH), 1.37 (s, 9 H, C(CH₃)₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): $\delta = 172.6$, 172.2, 170.0, 80.8, 52.4, 51.8, 31.4, 28.0, 27.2, 23.0 ppm. HRMS (ESI) for [(C₁₂H₂₁NO₅] + Na)⁺: calcd. 282.1317, found 282,1307 [M + Na]⁺.

N-Acetyl lysine (*N*-Boc) methyl ester (**3**u): ¹H NMR (300 MHz, CDCl₃, 25 °C): ¹H NMR (300 MHz, CDCl₃, 25 °C): $\delta = 6.48$ (d, J = 7.5 Hz, 1 H, *NH*), 4.73 (m, 1 H, N*H*), 4.50 (dd, J = 12.6 Hz, J = 7.5 Hz, 1 H, α -C*H*), 3.67 (s, 3H, OC*H*₃), 3.04- 2.99 (m, 2 H, ϵ -C*H*₂), 1.96 (s, 3 H, C*H*₃CO), 1.75 (m, 1H, β -C*H*), 1.58 (m, 1H, β -C*H*),

1.44-1.27 (m, 4H, γ -CH₂ + δ -CH₂), 1.36 (s, 9 H, C(CH₃)₃) ppm. ¹³C NMR (75 MHz, CDCl₃, 25 °C): δ = 172.1, 169.1, 155.2, 78.1, 51.3, 51.1, 38.9, 30.8, 28.6, 27.4, 21.9, 21.4 ppm. HRMS (ESI) for [(C₁₄H₂₆N₂O₅] + H)⁺ : calcd. 303.1920, found 303.1904 [M + H]⁺, 325.1737 [M + Na]⁺.



¹H NMR spectra (1q-1u)







Sample 1r: N-(9-Fluorenylmethoxycarbonyl) phenyl alanine benzyl ester





Sample 1s: N-(9-Fluorenylmethoxycarbonyl) tyrosine (O-tert-butyl) t-butyl ester

ΗŇ



Sample 1t: *N*-(9-Fluorenylmethoxycarbonyl) glutamic acid (*O*-tert-butyl) methyl ester

































Sample 3k: N-Acetyl valine methyl ester









Sample 30 (two rotamers): N-Acetyl N-methyl valine methyl ester





Sample 3p (two rotamers): N-Acetyl N-methyl isoleucine methyl ester















¹³ C NMR spectra (1q-1u)

Sample 1q: N-(9-Fluorenylmethoxycarbonyl) glycine t-butyl ester





Sample 1r: N-(9-Fluorenylmethoxycarbonyl) phenylalanine benzyl ester





mdd 2 3 28.85 8 εe·Lε —— 6 82.74 50 92.88 — 8 26.95 ____ 2 8 81.13 8 100 110 120 70.01 - 129.06 - 129.06 28.421 130 99.45T -----140 74.641 150 9T.4ST TS.SST 160 170 69.071 ——

Sample 1s: N-(9-Fluorenylmethoxycarbonyl) tyrosine (O-tert-butyl) t-butyl ester















HRMS (ESI) (1q-1t)

Sample 1q: *N*-(9-Fluorenylmethoxycarbonyl) glycine t-butyl ester

Com	nound	Table

Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
Cpd 1: C21 H23 N O4	7,843	353,1625	70820	C21 H23 N O4	353,1627	-0,54	C21 H23 N O4	C21 H23 N O4

Compound Label	m/z,	RT	Algorithm	Mass
Cpd 1: C21 H23 N O4	376,1518	7,843	Find By Formula	353,1625

m/z	z	Abund	Formula	Ion
376,1518	1	70819,95	C21H23NO4	(M+Na)+
377,1546	1	16965,1	C21H23NO4	(M+Na)+
378,1576	1	2795,57	C21H23NO4	(M+Na)+
379,1629	1	509,69	C21H23NO4	(M+Na)+

Sample 1r: N-Fmoc phenylalanine benzyl ester

Compound Table

					Tgt	Diff		
Compound Label	RT	Mass	Abund	Formula	Mass	(ppm)	MFG Formula	DB Formula
Cpd 1: C31 H27 N O4	9,116	477,1936	9459	C31 H27 N O4	477,194	-0,88	C31 H27 N O4	C31 H27 N O4

	m/z	z	Abund	Formula	Ion				
	478,2043	1	152,58	C31H27NO4	(M+H)+				
	500,1829	1	9459,41	C31H27NO4	(M+Na)+				
	501,1856	1	3309,26	C31H27NO4	(M+Na)+				
	502,1894	1	678,8	C31H27NO4	(M+Na)+				

Sample 1s: N-Fmoc tyrosine (Otbutyl) tert butyl ester

Compound Table

Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (nnm)	MFG Formula	DB Formula
Cpd 1: C32 H37 N O5	10,448	515,2675	120636	C32 H37 N O5	515,2672	0,66	C32 H37 N O5	C32 H37 N O5

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C32 H37 N	538,257	10,448	Find By Formula	515,2675
05				

m/z		z	Abund	Formula	Ion
	516,2664	1	408,93	C32H37NO5	(M+H)+
	517,2693	1	138,79	C32H37NO5	(M+H)+
	538,257	1	120635,8	C32H37NO5	(M+Na)+
	539,2596	1	41841,45	C32H37NO5	(M+Na)+
	540,2624	1	8468,1	C32H37NO5	(M+Na)+
	541,2688	1	1445,46	C32H37NO5	(M+Na)+

Sample 1t: *N*-(9-Fluorenylmethoxycarbonyl) Glutamic acid (O-tert-butyl) methyl ester

Compound Table								
						Diff		DB
Compound Label	RT	Mass	Abund	Formula	Tgt Mass	(ppm)	MFG Formula	Formula
Cpd 1: C25 H29 N O6	9,72	439,1998	312258	C25 H29	439,1995	0,66	C25 H29 N O6	C25 H29
*				N 06	-			N 06

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C25 H29 N O6	462,1891	9,72	Find By Formula	439,1998

Pio opeccium				
m/z	z	Abund	Formula	Ion
440,2135	1	77,61	C25H29NO6	(M+H)+
462,1891	1	312257,66	C25H29NO6	(M+Na)+
463,192	1	79635,58	C25H29NO6	(M+Na)+
464,194	1	13154,34	C25H29NO6	(M+Na)+
465,1969	1	1764,08	C25H29NO6	(M+Na)+
466,2004	1	267,16	C25H29NO6	(M+Na)+

K

Sample 1u: N-Fmoc Lysine (N-Boc) methyl ester

Compound Table	9
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	DT				Tgt	Diff		
Compound Label	RT	Mass	Abund	Formula	Mass	(ppm)	MFG Formula	DB Formula
Cpd 1: C27 H34 N2 O6	8,04	482,2412	51924	C27 H34 N2 O6	482,2417	-1,08	C27 H34 N2 O6	C27 H34 N2 O6

Compound Label	m/z,	RT	Algorithm	Mass
Cpd 1: C27 H34 N2 O6	505,2306	8,04	Find By Formula	482,2412

m/z	z	Abund	Formula	Ion
483,2484	1	705,98	C27H34N2O6	(M+H)+
484,2512	1	232,07	C27H34N2O6	(M+H)+
485,2538	1	69,71	C27H34N2O6	(M+H)+
505,2306	1	51923,8	C27H34N2O6	(M+Na)+
506,2331	1	14876,35	C27H34N2O6	(M+Na)+
507,2357	1	2976,79	C27H34N2O6	(M+Na)+
508,2404	1	419,34	C27H34N2O6	(M+Na)+

HRMS (ESI) (3j-3u)

Sample 3j: *N*-Acetyl alanine methyl ester

Compound Table

Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
Cpd 1: C6 H11 N O3	1,094	145,0736	399	C6 H11 N O3	145,0739	-2,01	C6 H11 N O3	C6 H11 N O3

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C6 H11 N O3	168,063	1,094	Find By Formula	145,0736

m/z	z	Abund	Formula	Ion
168,063	1	399,23	C6H11NO3	(M+Na)+
169,0642	1	44,76	C6H11NO3	(M+Na)+

Sample 31: N-Acetyl leucine methyl ester

Compound Table								-
Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
Cpd 1: C9 H17 N O3	0,686	187,1211	809445	C9 H17 N O3	187,1208	1,2	C9 H17 N O3	C9 H17 N O3

MS Spectrum Peak List				
m/z	z	Abund	Formula	Ion
210,1103	1	809445,44	C9H17NO3	(M+Na)+
211,1133	1	75166,33	C9H17NO3	(M+Na)+
212,1156	1	9144,75	C9H17NO3	(M+Na)+
213,1172	1	1069,12	C9H17NO3	(M+Na)+

Sample 3m: N-Acetyl isoleucine methyl ester

Compound Table								
Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
Cpd 1: C9 H17 N O3	3,261	187,1208	9107	C9 H17 N O3	187,1208	-0,46	C9 H17 N O3	C9 H17 N O3

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C9 H17 N O3	188,1279	3,261	Find By Formula	187,1208

MS Spectrum Peak List										
m/z	z	Abund	Formula	Ion						
188,1279	1	9107,47	C9H17NO3	(M+H)+						
189,1313	1	1112,44	C9H17NO3	(M+H)+						
190,1322	1	147,65	C9H17NO3	(M+H)+						
210,1099	1	8663,41	C9H17NO3	(M+Na)+						
211,1133	1	994,68	C9H17NO3	(M+Na)+						
212,1178	1	942,35	C9H17NO3	(M+Na)+						

Sample 3n: N-Acetyl phenyl alanine methyl ester

(M+Na)+

Compound Table								
Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
Cpd 1: C12 H15 N O3	3,859	221,1049	13788	C12 H15 N O3	221,1052	-1,51	C12 H15 N O3	C12 H15 N O3

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C12 H15 N O3	244,0941	3,859	Find By Formula	221,1049

183,8 C12H15NO3

MS Spectrum Peak List m/z z Abund Formula Ion 1 13787,94 C12H15NO3 244,0941 (M+Na)+ 245,0975 1 1850,58 C12H15NO3 (M+Na)+

1

246,0998

Sample 3q: N-Acetyl glycine t-butyl ester

RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
5,029	173,1051	5929	C8 H15 N O3	173,1052	-0,69	C8 H15 N O3	C8 H15 N O3
	RT 5,029	RT Mass 5,029 173,1051	RT Mass Abund 5,029 173,1051 5929	RT Mass Abund Formula 5,029 173,1051 5929 C8 H15 N O3	RT Mass Abund Formula Tgt Mass 5,029 173,1051 5929 C8 H15 N O3 173,1052	RT Mass Abund Formula Tgt Mass Diff (ppm) 5,029 173,1051 5929 C8 H15 N O3 173,1052 -0,69	RT Mass Abund Formula Tgt Mass Diff (ppm) MFG Formula 5,029 173,1051 5929 C8 H15 N O3 173,1052 -0,69 C8 H15 N O3

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C8 H15 N O3	174,1122	5,029	Find By Formula	173,1051

m/z	z	Abund	Formula	Ion					
174,1122	1	5929,13	C8H15NO3	(M+H)+					
175,1158	1	746,98	C8H15NO3	(M+H)+					
176,1146	1	151,98	C8H15NO3	(M+H)+					
196,0945	1	4083,82	C8H15NO3	(M+Na)+					
197,0974	1	428,3	C8H15NO3	(M+Na)+					

Sample 3r: N-Acetyl phenylalanine benzyl ester

Co	ompound Table								
							Diff		
	Compound Label	RT	Mass	Abund	Formula	Tgt Mass	(ppm)	MFG Formula	DB Formula
(Cpd 1: C18 H19 N O3	6,458	297,1363	16184	C18 H19 N O3	297,1365	-0,76	C18 H19 N O3	C18 H19 N O3
	Compound Label		m/z	RT	Algorithm		Mass		
	Cpd 1: C18 H19 N O3	3	320,1254	6,458	Find By For	nula	297,1363		

m/z	z	Abund	Formula	Ion
298,1437	1	1319,84	C18H19NO3	(M+H)+
299,15	1	291,36	C18H19NO3	(M+H)+
320,1254	1	16183,91	C18H19NO3	(M+Na)+
321,1288	1	3248,39	C18H19NO3	(M+Na)+
322,1318	1	464,67	C18H19NO3	(M+Na)+

Sample 3s: N-Acetyl tyrosine (O-t-butyl) t-butyl ester

Compound Table								
					Tgt	Diff		
Compound Label	RT	Mass	Abund	Formula	Mass	(ppm)	MFG Formula	DB Formula
Cpd 1: C19 H29 N O4	7,134	335,2101	1463	C19 H29 N O4	335,2097	1,44	C19 H29 N O4	C19 H29 N O4

m/z		z	Abund	Formula	Ion
	336,2133	1	86,93	C19H29NO4	(M+H)+
	358,1987	1	1463,28	C19H29NO4	(M+Na)+
	359,2054	1	333,28	C19H29NO4	(M+Na)+
	360,2119	1	72,39	C19H29NO4	(M+Na)+

Sample 3t: N-Acetyl glutamic acid (O-t-butyl) methyl ester

Compound Table								
Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
		11400	7104114	loinaia	11400	(PP)		2210
Cpd 1: C12 H21 N O5	4,215	259,1413	2038	C12 H21 N O5	259,142	-2,59	C12 H21 N O5	C12 H21 N O5

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C12 H21 N O5	282,1307	4,215	Find By Formula	259,1413

m/z		z	Abund	Formula	Ion
	282,1307	1	2038,26	C12H21NO5	(M+Na)+
	283,1332	1	299,12	C12H21NO5	(M+Na)+
	284,1343	1	86,64	C12H21NO5	(M+Na)+

Sample 3u: N-Acetyl Lysine (N-Boc) methyl ester

_Compound Table								
Compound Label	RT	Mass	Abund	Formula	Tgt Mass	Diff (ppm)	MFG Formula	DB Formula
Cpd 1: C14 H26 N2 O5	4,136	302,1845	906783	C14 H26 N2 O5	302,1842	1	C14 H26 N2 O5	C14 H26 N2 O5

Compound Label	m/z	RT	Algorithm	Mass
Cpd 1: C14 H26 N2 O5	325,1737	4,136	Find By Formula	302,1845

m/z	z	Abund	Formula	Ion
303,1904	1	3316,26	C14H26N2O5	(M+H)+
304,1956	1	449,96	C14H26N2O5	(M+H)+
305,1958	1	119,67	C14H26N2O5	(M+H)+
325,1737	1	906783,44	C14H26N2O5	(M+Na)+
326,1766	1	127692,35	C14H26N2O5	(M+Na)+
327,1784	1	16694,49	C14H26N2O5	(M+Na)+
328,1802	1	1730,82	C14H26N2O5	(M+Na)+