Cav2.2 and Cav3.1 Calcium Channel Inhibitors from Valeriana jatamansi Jones

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Figure S1 Structures of compounds 14 and 15



Figure S2. ESIMS of velerivaltrate A (1).



Figure S3. HRESIMS of velerivaltrate A (1).



Figure S4. ¹H NMR (600 MHz, CDCl₃) of velerivaltrate A (1).



Figure S5. ¹³C NMR (100 MHz, CDCl₃) of velerivaltrate A (1).



Figure S6. HSQC (600 MHz, CDCl₃) of velerivaltrate A (1).



Figure S7. HMBC (600 MHz, CDCl₃) of velerivaltrate A (1).



Figure S8. ¹H-¹H COSY (600 MHz, CDCl₃) of velerivaltrate A (1).



Figure S9. ROESY (600 MHz, CDCl₃) of velerivaltrate A (1).



Figure S10. IR spectrum of velerivaltrate A (1).



Figure S11. UV spectrum of velerivaltrate A (1).

Optical rotation measurement

| Model | : P-1020 (A0 | 60460638) | | | | | | |
|-------|--------------|-----------|---------|------------------|-----------------------------|---|-----------------------------|---|
| No. | Sample | Mode | Data | Monitor Blank | Temp. Cell Temp Point | Date Comment Sample Name | Light Filter Operator | Cycle Time Integ Time |
| No.1 | 6 (1/3) | Sp.Rot | 59.7560 | 0.0049 0.0000 | 23.6 10.00 Cell | Tue Mar 17 20:16:57 2015 0.00082g/mL MeOH ZSJ-17A | Na 589nm | 2 sec 10 sec |
| No.2 | 6 (2/3) | Sp.Rot | 53.6590 | 0.0044 0.0000 | 23.6 10.00 Cell | Tue Mar 17 20:17:10 2015 0.00082g/mL MeOH ZSJ-17A | Na 589nm | 2 sec 10 sec |
| No.3 | 6 (3/3) | Sp.Rot | 57.3170 | 0.0047 0.0000 | 23.6 10.00 Cell | Tue Mar 17 20:17:24 2015 0.00082g/mL MeOH ZSJ-17A | Na 589nm | $ \begin{array}{c} 2 \sec \\ 10 \sec \\ +79, 1729 \end{array}^{\circ} $ |
| No.4 | 7 (1/3) | Sp.Rot | 64.6340 | 0.0053 0.0000 | 23.5 10.00 Cell | Tue Mar 17 20:20:53 2015 0.00082g/mL MeOH ZSJ-17A | Na 589nm | 2 sec / / 10 sec |
| No.5 | 7 (2/3) | Sp.Rot | 62.1950 | 0.0051 0.0000 | 23.5 10.00 Cell | Tue Mar 17 20:21:07 2015 0.00082g/mL MeOH ZSJ-17A | Na 589nm | 2 sec 10 sec |
| No.6 | 7 (3/3) | Sp.Rot | 59.7560 | 0.0049 0.0000 | 23.5 10.00 Cell | Tue Mar 17 20:21:20 2015 0.00082g/mL MeOH ZSJ-17A | Na 589nm | 2 sec 10 sec |

Figure S12. $[a]_D$ spectrum of velerivaltrate A (1).



Figure S13. ESIMS of velerivaltrate B (2).



Figure S14. HRESIMS of velerivaltrate B (2).



Figure S15. ¹H NMR (600 MHz, CDCl₃) of velerivaltrate B (2).



Figure S16. ¹³C NMR (100 MHz, CDCl₃) of velerivaltrate B (2).



Figure S17. HSQC (600 MHz, CDCl₃) of velerivaltrate B (2).



Figure S18. HMBC (600 MHz, CDCl₃) of velerivaltrate B (2).



Figure S19. ${}^{1}H$ - ${}^{1}H$ COSY (600 MHz, CDCl₃) of velerival trate B (2).



Figure S20. ROESY (600 MHz, CDCl₃) of velerivaltrate B (2).



Figure S21. IR spectrum of velerivaltrate B (2).



Figure S22. UV spectrum of velerivaltrate B (2).

Optical rotation measurement

| Model : No. | P-1020 (A0 Sample | 60460638) Mode | Data | Monitor Blank | Temp. Cell Temp Point | Date Comment Sample Name | Light Filter Operator | Cycle Time Integ Time |
|----------------|----------------------|-------------------|---------|------------------|-----------------------------|---|-----------------------------|--------------------------|
| No.1 | 13 (1/3) | Sp.Rot | 38.8060 | 0.0052 0.0000 | 23.3 10.00 Cell | Fri Mar 20 19:46:49 2015 0.00134g/mL MeOH ZSJ-17C | Na 589nm | 2 sec 10 sec |
| No.2 | 13 (2/3) | Sp.Rot | 35.8210 | 0.0048 0.0000 | 23.3 10.00 Cell | Fri Mar 20 19:47:02 2015 0.00134g/mL MeOH ZSJ-17C | Na 589nm | 2 sec 10 sec +32.0642 |
| No.3 | 13 (3/3) | Sp.Rot | 36.5670 | 0.0049 0.0000 | 23.3 10.00 Cell | Fri Mar 20 19:47:16 2015 0.00134g/mL MeOH ZSJ-17C | Na 589nm | 2 sec 10 sec |

Figure S23. $[a]_D$ spectrum of velerivaltrate B (2).







Figure S25. HRESIMS of velerivaltrate C (3).



Figure S26. ¹H NMR (600 MHz, CDCl₃) of velerivaltrate C (3).



Figure S27. ¹³C NMR (100 MHz, $CDCl_3$) of velerivaltrate C (3).



Figure S28. HSQC (600 MHz, CDCl₃) of velerivaltrate C (3).



Figure S29. HMBC (600 MHz, CDCl₃) of velerivaltrate C (3).



Figure S30. ¹H-¹H COSY (600 MHz, CDCl₃) of velerivaltrate C (3).



Figure S31. ROESY (600 MHz, $CDCl_3$) of velerival trate C (3).



Figure S32. IR spectrum of velerivaltrate C (3).



Figure S33. UV spectrum of velerivaltrate C (3).

Optical rotation measurement

| Model : No. | P-1020 (A06 Sample | 50460638) Mode | Data | Monitor Blank | Temp. Cell Temp Point | Date Comment Sample Name | Light Filter Operator | Cycle Time Integ Time |
|----------------|-----------------------|-------------------|---------|------------------|-----------------------------|--|-----------------------------|------------------------------------|
| No.1 | 5 (1/3) | Sp.Rot | 29.1080 | 0.0093 0.0000 | 23.6 10.00 Cell | Tue Mar 17 20:09:36 2015 0.00320g/mL MeOH ZSJ-15 | Na 589nm | 2 sec 10 sec |
| No.2 | 5 (2/3) | Sp.Rot | 29.4210 | 0.0094 0.0000 | 23.6 10.00 Cell | Tue Mar 17 20:09:50 2015 0.00320g/mL MeOH ZSJ-15 | Na 589nm | $\frac{2 \sec}{10 \sec} + 30.0469$ |
| No.3 | 5 (3/3) | Sp.Rot | 31.6120 | 0.0101 0.0000 | 23.6 10.00 Cell | Tue Mar 17 20:10:03 2015 0.00320g/mL MeOH ZSJ-15 | Na 589nm | 2 sec 10 sec |

Figure S34. $[a]_D$ spectrum of velerivaltrate C (3).

| compounds | concentration (μ M) | Inhibitory ratio (%) |
|---------------|--------------------------|------------------------|
| Blank control | 0.1% DMSO | 1.2±0.8 |
| 1 | 1 | 7.3±3.1.* |
| | 3 | 21.7±73* |
| | 10 | 55.9±3.3.* |
| | 30 | 59.9±5.2.* |
| | 60 | 64.3±3.9 ^{**} |
| 6 | 0.6 | 9.5±2.5.* |
| | 1 | 23.3±2.9.** |
| | 3 | 38.2±2.8 ^{**} |
| | 10 | 56.8±5.7.** |
| | 30 | 61.1±7.8 [*] |
| 7 | 0.3 | 2.5±1.4 |
| | 1 | 30.6±4.4.*** |
| | 3 | 51.4±8.4.*** |
| | 10 | 59.3±5.4.* |
| | 30 | 57.2±7.1.* |
| 11 | 0.3 | 3.6±2.10 |
| | 1 | 9.5±0.9 [*] |
| | 3 | 36.1±1.9.** |
| | 10 | 63.1±1.0 ^{**} |
| | 30 | 63.9±3.9 ^{**} |
| 12 | 0.6 | 3.9±2.7 |
| | 3 | 17.7±7.7* |
| | 10 | 37.0±6.9 ^{**} |
| | 30 | 61.8±6.7 ^{**} |
| | 60 | 47.9±1.7*** |

Table S1. Dose-related effects of compounds 1, 6, 7, 11, and 12 on peak currents of $Ca_v 2.2$.

All the data were analyzed with two-tailed student T test and represented as mean \pm SEM (n=3) compared with the blank control, *P < 0.01, **P < 0.01.



Figure S35. Inhibitory effects of compounds **1-12** on $Ca_v 2.2$ at 30 μ M. Representative peak current of $Ca_v 2.2$ were evoked from a holding potential (HP) of -80 mV by 50 ms depolarization to + 20 mV at 3 s intervals in the absence or presence of the indicated compounds.



Figure S36. Inhibitory effects of compounds **1**, **4**, **5**, **6**, **7**, **9**, **10**, **11**, and **12** on $Ca_v 3.1$ at 30 μ M. Representative peak current of $Ca_v 3.1$ were evoked from a holding potential (HP) of -80 mV by 50 ms depolarization to -10 mV at 3 s intervals in the absence or presence of the indicated compounds.



Figure S37. Normalized current-voltage (I-V) curves of $Ca_v.1.2$ in the absence or presence of compounds 1-13. $Ca_v.1.2$ Currents were evoked from a holding potential (HP) of -80 mV by 50 ms depolarization from -30 mV to + 70 mV at 3 s intervals. All the data were analyzed with two-tailed student T test and represented as mean \pm SEM (n=3). *P<0.05, compared with the absence of the indicated compounds.



Figure S38. Normalized current-voltage (I-V) curves of Ca_v2.1 in the absence or presence of compounds **1-13**. Ca_v2.1 currents were evoked from a holding potential (HP) of -80 mV by 50 ms depolarization from -30 mV to + 70 mV at 3 s intervals. All the data were analyzed with tow-tailed student T test and represented as mean \pm SEM (n=3). **P*<0.05, compared with the absence of the indicated compounds.



Figure S39. Normalized current-voltage (I-V) curves of Ca_v2.2 in the absence or presence of compounds **1-13**. Ca_v2.2 currents were evoked from a holding potential (HP) of -80 mV by 50 ms depolarization from -30 mV to + 70 mV at 3 s intervals. All the data were analyzed with tow-tailed student T test and represented as mean \pm SEM (n=3). ***P*<0.01, **P*<0.05, compared with the absence of the indicated compounds.



Figure S40. Normalized current-voltage (I-V) curves of $Ca_v 3.1$ in the absence or presence of compounds **1-13**. $Ca_v 3.1$ currents were evoked from a holding potential (HP) of -80 mV by 50 ms depolarization from -50 mV to + 60 mV at 3 s intervals. All the data were analyzed with tow-tailed student T test and represented as mean \pm SEM (n=3). ***P*<0.01, **P*<0.05, compared with the absence of the indicated compounds.



Figure S41. Normalized current-voltage (I-V) curves of KNCH2 in the absence or presence of compounds 1-13. KCNH2 were pretreated by indicated compounds for 1 s and then currents were evoked from a holding potential (HP) of -80 mV by 3 s depolarization from -120 mV to + 40 mV at 30 s intervals. All the data were analyzed with tow-tailed student T test and represented as mean \pm SEM (n=3), compared with the absence of the indicated compounds.



Figure S42. The inhibitory effects of Mibefradil (the commercial T-type calcium channel inhibitor) and ω-Conotoxin MVIIA (the commercial N-type calcium channel inhibitor) on Ca_v3.1 and Ca_v2.2, respectively. (**A**) Representative Ca_v3.1 current trace evoked by 50-ms depolarizations to -10 mV at 3-s intervals from a holding potential (HP) of -80 mV in the absence and presence of mibefradil. Ca_v3.1 was expressed in *Xenopus* oocytes; (**B**) Dose-response relationship of mibefradil inhibition of Ca_v3.1. Data points represent mean±S.E.M. of three measurements at HP of -80 mV. Solid curve represents a fit to the Hill equation with an IC₅₀value of 10.4μ M and a Hill coefficient of 1.6; (**C**) Representative Ca_v2.2 current trace evoked by 50-ms depolarizations to +20 mV at 3-s intervals from a holding potential (HP) of -80 mV in the absence and presence of ω -Conotoxin MVIIA. Ca_v2.2 was expressed in *Xenopus* oocytes; (**D**) Normalized current-voltage (I-V) curves of Ca_v2.2 in the absence or presence of the indicated concentrations of ω -conotoxin MVIIA. Results were obtained in 3 cells.