

## Supporting Information I

### 1 Purity determination of Neurons

To observe the purity of neurons 7 d after incubation, cortical neurons in culture plates were fixed by 4% paraformaldehyde for 20 min, blocked in antibody buffer with 20% goat serum, permeabilized with 0.2 % Triton X-100<sup>TM</sup>and incubated in mouse anti-MAP2 antibody(Ab11267; 1:200; Abcam, Cambridge, UK) for 24 h at 4 °C, incubated with FITC-labeled goat anti-rabbit IgG for 1.5 h at room temperature. Sections were washed in PBS after each step and finally visualized by laser scanning confocal microscopy (CLSM, Tokyo, Japan)

### 2 Oxygen-glucose deprivation (OGD)

After washing twice, control group were immersed in controlled salt solution (CSS, 120 mMNaCl, 5.4 mM KCl, 0.8 mM MgCl<sub>2</sub>×6 H<sub>2</sub>O, 1.8 mM CaCl<sub>2</sub>×2 H<sub>2</sub>O, 25 Mm Tris, 15 mM D-glucose, PH=7.2) and incubated at 37 °C in an incubator supplemented with 5 % CO<sub>2</sub> for 3 h, 6 h, 9 h, 12 h. OGD group were immersed in deoxygenated controlled salt solution (CSSO, prepare for CSS, but substitute 15 mM D-glucose. Bubble the amount of solution needed for each experiment with N<sub>2</sub> for at least 10 min in the anaerobic chamber) and incubated at oxygen-free N<sub>2</sub>/CO<sub>2</sub> (95%/5%) gas at 37 °C for 3 h, 6 h, 9 h, 12 h. Percentage of cell death was determined by LDH assay. A time interval, at which 50 % of cell mortality was observed, was utilized for further analysis.

### 3 Evaluation of edaravone protection effect against OGD

To determine the effect of edaravone, cultured cortical neurons were treated with 1 µM, 0.1 µM, 0.01 µM, 0.001 µM EDA for 6 h of OGD. The cell viability was quantitatively assessed by the measurement of LDH released into the bathing medium.

### 3 Results

At 7 d after plating, the cultures were used for the experiments. Confirmed by the use of mouse anti-MAP2 staining, the majority of cultured cells showed MAP2 immunoreactivity (Figure S1), which indicated that these cells were neurons and the purity was calculated over 87%. Later on, the cells further showed full of thick and projecting interweaving network. This stage was the best time for testing

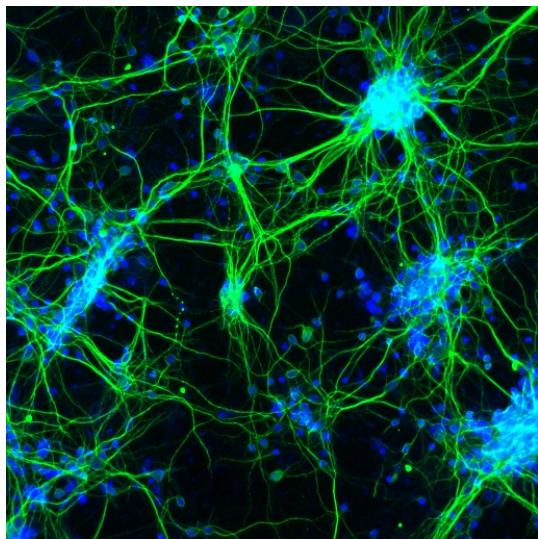


Figure S1. immunofluorescence staining of primary cortical neurons. MAP2 (green) and DAPI (blue). The purity of cortical neurons was calculated as over 87% ( $\times 200$ ).

As shown in figure S2, LDH assay has shown time-dependent increase in cell mortality with OGD duration. At 3 and 6 h, 29.6 % and 48.5 % cell deaths, respectively, were observed, whereas at 9 h and 12 h exposure, the cell mortality had increased to 62.0 % and 72.7 %, respectively, in comparison to non-OGD control. Hence, 6 h OGD exposure was carried forward for evaluating neuroprotective efficacy of the molecules.

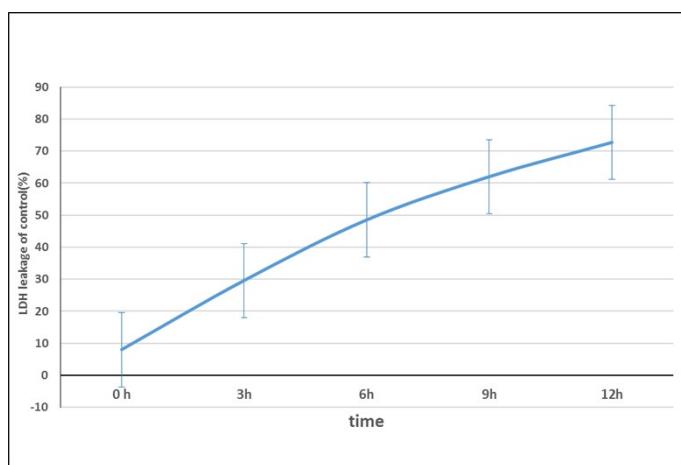


Figure S2. The extent of LDH release of OGD duration. Line chart represent mean, n>3.

As shown in the Figure S3, the leakage rate of LDH has dramatically decreased compared with OGD group which implying the cortical neuron injury after EDA treatment with 0.1  $\mu$ M, 0.01  $\mu$ M, 0.001  $\mu$ M for 6 h of OGD. In addition, in the four doses of EDA-treated OGD groups, the dose of 0.01  $\mu$ M of EDA had the best neuroprotective efficacy. So, the dose of 0.01  $\mu$ M was chosen for the further analysis served as positive drugs.

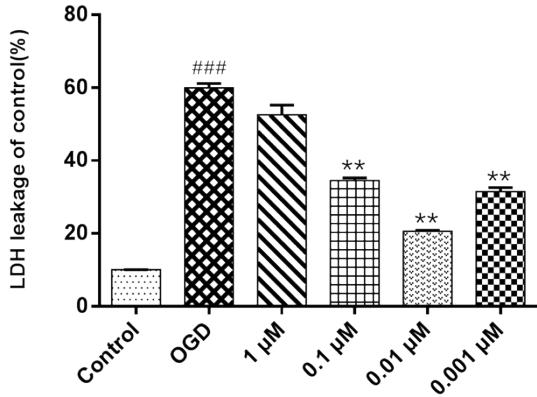


Figure S3. Effects of different concentrations of EDA on cell viability by LDH assay.  $\#P<0.05$ ,  $\#\#P<0.01$ ,  $\#\#\#P<0.001$  the OGD group versus the Control group;  $*P<0.05$ ,  $**P<0.01$ ,  $***P<0.001$  the DHI group (or positive group) versus the OGD group OGD. Histograms represent mean  $\pm$  SD, n>3.

## Supporting InformationII

In order to get a reliable result and improve the accuracy of lipid analysis, a target database including the accurately identified 98 PCs, 28 SMs and 41 Cers was constructed on the LC-MS platform in advance by the internal standard standards and using the relative data of accurate masses and MS/MS fragments based on the organic samples. The internal standard mixes including PCs, SMs and Cers were purchased from Avanti Polar Lipids. Cer/Sph Mixture I (<https://avantilipids.com/product/lm-6002/>) PC(12:0/13:0) (<https://avantilipids.com/product/lm-1000/>).

## Results

Separation for relative quantification of PCs, Cers, and SMs was performed by UltiMate<sup>TM</sup> 3000 Rapid Separation LC (RSLC) system (Thermo Scientific, USA). As stationary phase, an ACQUITY UPLC HSS T3 C18 column (1.8 $\mu$ M, 100x2.1mm, Waters<sup>TM</sup>, USA) was used suited for polar compound retention. The mobile phase consisted of solvent A (0.1% formic acid-water containing 2 mM ammonium formate) and solvent B (methanol) with a gradient elution (0–2 min, 20–30% B; 2–5 min, 30–45% B; 5–6.5 min, 45–55% B; 6.5–12 min, 55–65% B; 12–14 min, 65–85% B; 14–17.5 min, 85–100% B; 17.5–18 min, 100–100% B). The re-equilibration was 2 minutes with 20% B. The flow rate of the mobile phase was 0.3 mL/min. The column temperature was maintained at 45 °C and the sample manager temperature was set at 4 °C.

A Thermo Scientific<sup>TM</sup> Q Exactive hybrid quadrupole Orbitrap mass spectrometer equipped with a HESI-II probe was used in the positive electrospray ionization mode. The pos HESI-II spray voltages were 3.7 kV, the heated capillary temperature was 320 °C, the sheath gas pressure was 30 psi, the auxiliary gas setting was 10 psi, and the heated vaporizer temperature was 300 °C. The parameters of the full mass scan were as follows: resolution of 70,000, auto gain control target under  $1 \times 10^6$ , maximum isolation time of 50 ms, and  $m/z$  range of 150–1500.









38	GlcCer(d18:2/22:0)	LMSP0501AA37	C46H87NO8	10.03	[M+H]+, 782.6504
39	LacCer(d18:1/16:0)	LMSP0501AB03	C46H87NO13	8.58	[M+H]+, 862.625
40	LacCer(d18:1/24:0)	LMSP0501AB07	C54H103NO13	11.56	[M+H]+, 974.7502
41	LacCer(d18:1/24:1(15Z))	LMSP0501AB09	C54H101NO13	10.6	[M+H]+, 972.7346