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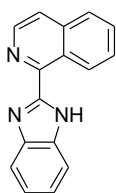
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1. Chemistry

All chemicals were purchased from Sigma Aldrich, Merck, A K Scientific, or Ark Pharm Inc. BODIPY®630/650-SE was purchased from Life Technologies. Reactions were carried out at room temperature (rt) unless otherwise stated. Thin layer chromatography (t.l.c.) was performed on 0.2 mm aluminium-backed silica gel plates 60 F₂₅₄ and visualised under UV light at $\lambda = 254$ and 365 nm, and with ninhydrin and/or KMnO₄ dip. Flash silica gel column chromatography was performed using 40-63 μm silica. An Agilent 1260 Infinity system was used for reverse phase high-performance liquid chromatography (RP-HPLC), with a YMC C8 5 μm (150 x 10 mm) semi-preparative or YMC C8 5 μm (150 x 4.6 mm) analytical column. RP-HPLC solvents were A: H₂O (0.05% TFA) and B: 9:1 MeCN:H₂O (0.05% TFA). Analytical RP-HPLC retention times are reported using following method - 5% solvent B 1 min, gradient of 5-95% solvent B 1-27 mins, 95% solvent B 27-28 mins, gradient of 95-5% solvent B 28-30 mins, 5% solvent B 30-34 mins. TFA salts of RP-HPLC purified compounds were neutralised using an Amberlyst A21 ion exchange resin before biological testing. Analytical RP-HPLC was used to confirm purity (> 95%) at 254 and 380 nm for all compounds biologically tested. High resolution electrospray ionization mass spectra (HRMS-ESI) were recorded on a Bruker microTOF mass spectrometer. NMR spectra were obtained on a Varian 400-MR or Varian 500 MHz AR Premium Shielded spectrometer. Chemical shifts are listed in ppm (δ), calibrated using residual undeuterated solvent as the internal standard, and coupling constants (J) are recorded in hertz (Hz). Note - not all magnetically non-equivalent carbons were observed in ¹³C NMR spectrum for all compounds.

Synthesis of 1-(1*H*-Benzimidazol-2-yl)isoquinoline (1)

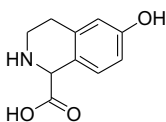


Compound was synthesised according to a previously reported literature synthesis of **1**.¹ ¹H NMR spectroscopy data in DMSO-*d*₆ was similar to this literature report¹. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.22 – 7.36 (m, 2H, ArH benzimidazole), 7.59 – 7.66 (m, 1H, ArH benzimidazole), 7.80 – 7.91 (m, 3H, ArH benzimidazole and ArH benzimidazole), 7.96 – 8.03 (m, 1H, ArH isoquinoline), 8.03 – 8.13 (m, 1H, ArH isoquinoline), 8.70 (d, 1H, *J* = 5.5, ArH isoquinoline), 10.09 – 10.13 (m, 1H, ArH isoquinoline), 13.22 (s, 1H, NH). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 112.05, 119.72, 121.94, 122.34, 123.63, 125.95, 127.21, 127.82, 128.61, 130.61, 134.02, 136.77, 141.64, 144.07, 146.64, 151.17.

¹H NMR (400 MHz, Methanol-*d*₄) δ 7.28 – 7.40 (m, 2H, ArH benzimidazole), 7.65 (s, 1H, ArH benzimidazole), 7.75 – 7.87 (m, 3H, ArH benzimidazole and ArH isoquinoline), 7.90 (d, 1H, *J* = 5.9 Hz, ArH isoquinoline), 7.97 – 8.05 (m, 1H, ArH isoquinoline), 8.64 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 9.45 – 9.54 (m, 1H, ArH isoquinoline). ¹³C NMR (101 MHz, Methanol-*d*₄) δ 123.62, 127.91, 128.25, 128.71, 129.59, 131.88, 138.67, 142.83, 149.04, 152.34. HRMS calculated for C₁₆H₁₂N₃ (M + H)⁺, 246.1026; found, 246.1029. Analytical RP-HPLC R_t = 12.93 min.

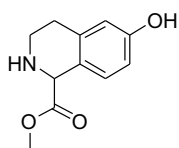
1.1 Experimental details for scheme 1

6-Hydroxy-1,2,3,4-tetrahydroisoquinoline-1-carboxylic acid (5)



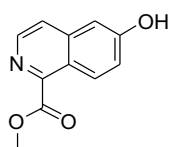
2-(3-Methoxyphenyl)ethan-1-amine was converted to **4** according to literature.² ¹H NMR (400 MHz, DMSO-*d*₆) δ 2.74 – 2.81 (m, 2H, CH₂), 2.95 – 3.04 (m, 2H, CH₂), 6.60 – 6.75 (m, 3H, ArH), 7.03 – 7.17 (m, 1H, ArH), 7.80 (br s). Amine **4** was then cyclised to give **5** according to literature.³ ¹H NMR (400 MHz, DMSO-*d*₆) δ 2.66 – 2.79 (m, 1H, CH^{*}HCH₂), 2.79 – 2.93 (m, 1H, CHH^{*}CH₂), 3.01 – 3.14 (m, 1H, CH₂CH^{*}H), 3.26 – 3.34 (m, 1H, CH₂CHH^{*}), 4.33 (s, 1H, CHCO₂H), 6.49 (d, *J* = 2.5 Hz, 1H, ArH), 6.60 (dd, *J* = 2.6 Hz, 1H, 8.5, ArH), 7.50 (d, *J* = 8.5 Hz, 1H, ArH), 8.80 (br m, 2H, NH, OH), 9.37 (br s, 1H, OH). (* designates diastereotopic protons). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 25.25, 57.82, 113.57, 113.91, 121.19, 129.06, 132.69, 155.88, 167.15. HRMS calculated for C₁₀H₁₀NO₃ (M - H)⁻, 192.0652; found, 192.0666.

Methyl 6-hydroxy-1,2,3,4-tetrahydroisoquinoline-1-carboxylate (**6**)



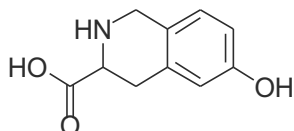
SOCl₂ (7.5 mL, 102.95 mmol) was added dropwise to a 0 °C suspension of **5** (6.63 g, 34.32 mmol) in MeOH (150 mL), then the mixture was warmed to rt and refluxed for 15 h. Solvent was removed under reduced pressure and the resulting solid was dissolved in THF (30 mL) and neutralised with Et₃N (15 mL). The reaction mixture was filtered to remove Et₃N.HCl and the filtrate evaporated to give **6** (7.11 g, 34.31 mmol, yield quantitative) as a yellowish solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 2.66 – 2.79 (m, 2H, CH₂*CH₂), 2.93 – 3.02 (m, 1H, CH₂CH*H), 3.21 – 3.30 (m, 1H, CH₂CHH*), 3.74 (s, 3H, OCH₃), 4.60 (s, 1H, CHCO₂CH₃), 6.54 (d, *J* = 2.6 Hz, 1H, ArH), 6.60 (dd, *J* = 2.6, 8.4 Hz, 1H, ArH), 7.13 (d, *J* = 8.46 Hz, 1H, ArH). (* designates diastereotopic protons). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 29.49, 41.18, 52.65, 58.98, 114.60, 116.22, 123.45, 129.77, 137.57, 157.64, 174.89. HRMS calculated for C₁₁H₁₄NO₃ (M + H)⁺, 208.0973; found, 208.0975. The synthesis of **6** has previously been reported by Ma *et al*⁴ but without spectroscopic data of **6**.

Methyl 6-hydroxyisoquinoline-1-carboxylate (**7**)



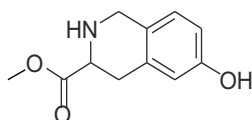
To a solution of **6** (0.17 g, 0.83 mmol) in 1,4-dioxane:THF (10mL 1:1, *v:v*) at 45 °C was added DDQ (0.38 g, 1.67 mmol) and the reaction was stirred vigorously at 45 °C for 5 h with the mouth of the flask open to the atmosphere to allow mixing of air. 1,4-Dioxane (10 mL) was added and the reaction mixture filtered, the filtrate was diluted with EtOAc and washed three times with sat aq. NaHCO₃ solution. The organic washings were combined, washed once with water, brine solution, then dried over MgSO₄, concentrated under reduced pressure and purified by silica gel flash column chromatography (30 to 50% EtOAc/hexane) to provide **7** (84 mg, 0.413 mmol, yield 49 %) as an off-white solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 4.04 (s, 3H, OCH₃), 7.16 (d, *J* = 2.5 Hz, 1H, ArH), 7.28 (dd, 1H, *J* = 2.5, 9.3 Hz, ArH), 7.73 (d, 1H, *J* = 5.7 Hz, ArH), 8.30 (d, 1H, *J* = 5.7 Hz, ArH), 8.52 (d, 1H, *J* = 9.3 Hz, ArH). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 53.24, 108.72, 122.70, 122.89, 124.25, 129.49, 141.07, 141.83, 149.28, 161.18, 167.61. HRMS calculated for C₁₁H₁₀NO₃ (M + H)⁺, 204.0655; found, 204.0647. To our knowledge, isoquinoline **7** has only been reported once before in the literature in a Japanese patent⁵ but no spectroscopic data was provided.

6-Hydroxy-1,2,3,4-tetrahydroisoquinoline-3-carboxylic acid (**9**)



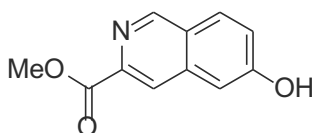
(+/-)-*m*-Tyrosine was converted to **9** according to literature.⁶ ¹H NMR (400 MHz, deuterium oxide[#]) δ 2.73 (dd, 1H, *J* = 11.0, 16.4 Hz, CH^{*}HCHNH), 2.92 (dd, 1H, *J* = 4.5, 16.5 Hz, CHH^{*}CHNH), 3.38 (dd, 1H, *J* = 4.5, 11.0 Hz, CH), 3.73 – 3.9 6 (m, 2H, NHCH₂^{*}), 6.40 – 6.51 (m, 2H), 6.87 (d, 1H, *J* = 8.3 Hz) (* designates diastereotopic protons). ¹³C NMR (101 MHz, deuterium oxide[#]) δ 32.03, 45.96, 58.01, 117.30, 118.14, 120.55, 127.30, 135.26, 164.38, 181.23. [#]Due to insolubility of compound in MeOD-*d*₄, DMSO-*d*₆ and deuterium oxide, NMR was obtained by dissolving 31 mg of **9** in a solution of deuterium oxide (20 mg KOH in 0.8 mL deuterium oxide). HRMS calculated for C₁₀H₁₀NO₃ (M - H)⁻, 192.0651; found, 192.0666.

Methyl 6-hydroxy-1,2,3,4-tetrahydroisoquinoline-3-carboxylate (**10**)



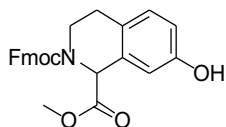
To a solution of **9** (2.0 g, 10.35 mmol) in MeOH (50 mL) was added H₂SO₄ (96%, 1 mL) and the reaction refluxed for 12 h, cooled, neutralised with NaHCO₃ solution and extracted with EtOAc (3 × 100 mL). The EtOAc layer was washed with brine solution (50 mL), dried over MgSO₄ and the solvent removed under reduced pressure to give **10** (2.05 g, 9.90 mmol, 96% yield) as an off-white solid. ¹H NMR (400 MHz, CDCl₃) δ 2.80 – 3.03 (m, 2H, NHCHCH₂^{*}), 3.68 – 3.74 (m, 1H, NHCH), 3.76 (s, 3H, OCH₃), 3.93 – 4.10 (m, 2H, NHCH₂^{*}), 4.68 (br s, 2H, OH, NH), 6.48 (d, 1H, *J* = 2.6 Hz, ArH), 6.60 (dd, 1H, *J* = 2.6, 8.3 Hz, ArH), 6.85 (d, 1H, *J* = 8.3 Hz, ArH). * designates diastereotopic protons. ¹³C NMR (101 MHz, CDCl₃) δ 31.48, 46.66, 52.45, 55.73, 114.30, 115.62, 125.78, 127.36, 134.12, 154.91, 173.44. HRMS calculated for C₁₁H₁₄NO₃ (M + H)⁺, 208.0968; found, 208.0950. The synthesis of methyl ester **10** from **9** has previously been reported⁷⁻¹¹ however none of these reports include spectroscopic data for **10**.

Methyl 6-hydroxyisoquinoline-3-carboxylate (**11**)



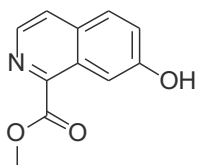
Following the procedure described for **7**, a mixture of **10** (1.2 g, 5.79 mmol) and DDQ (2.63 g, 11.58 mmol) gave **11** (0.6g, 2.95 mmol, 52% yield) as an off-white solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 3.99 (s, 3H, OCH₃), 7.23 (d, *J* = 2.3 Hz, 1H, ArH), 7.34 (dd, *J* = 8.9, 2.4 Hz, 1H, ArH), 8.01 (d, *J* = 8.9 Hz, 1H, ArH), 8.37 (s, 1H, ArH), 9.04 (s, 1H, ArH). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 53.01, 109.88, 123.57, 123.97, 126.30, 131.11, 139.25, 141.89, 152.66, 162.04, 167.28. HRMS calculated for C₁₁H₉NNaO₃ (M + Na)⁺, 226.0475; found, 226.0462. The synthesis of **11** from **10** has previously been reported¹¹ but without spectroscopic data of **11**.

2-(9H-Fluoren-9-yl)methyl 1-methyl 7-hydroxy-1,2,3,4-tetrahydroisoquinoline-1,2-dicarboxylate (**13**)



4-(2-aminoethyl)phenol was converted to (9H-fluoren-9-yl)methyl N-[2-(4-hydroxyphenyl)ethyl]carbamate **12** according to modified literature procedure.³ ¹H NMR (400 MHz, MeOD-*d*₄) δ 2.67 (t, 2H, $J = 7.3$ Hz, NHCH₂CH₂), 3.27 (t, 2H, $J = 7.3$ Hz, NHCH₂CH₂), 4.18 (t, 1H, $J = 6.8$ Hz, CH Fmoc), 4.32 (d, 2H, $J = 6.9$ Hz, CH₂ Fmoc), 6.63 – 6.73 (m, 2H, ArH), 7.00 (d, 2H, $J = 8.1$ Hz, ArH), 7.25 – 7.35 (m, 2H, ArH Fmoc), 7.35 – 7.44 (m, 2H, ArH Fmoc), 7.62 (d, 2H, $J = 7.5$ Hz, ArH Fmoc), 7.79 (d, 2H, $J = 7.5$ Hz, ArH Fmoc). HRMS calculated for C₂₃H₂₁NNaO₃ (M + Na)⁺, 382.1414; found, 382.1387. To a solution of **12** (19.7 g, 54.85 mmol) in AcOH/H₂SO₄ (200 mL, 3:1 v:v) was added a solution of glyoxylic acid monohydrate (5.5 g, 92.1 mmol) and stirred for 24 h. The reaction mixture was poured slowly into ice/water (caution: exothermic reaction) and the precipitate formed was separated and dried under vacuum oven (60°C, 300 mbar) for 12 h to give a pinky white solid. This solid was dissolved in DCM and washed with water, brine, dried over MgSO₄ and evaporated under reduced pressure to give 2-[(9H-fluoren-9-yl)methoxy]carbonyl-7-hydroxy-1,2,3,4-tetrahydroisoquinoline-1-carboxylic acid (previously reported by Maillard *et al*³) as a pinky white solid (18.4 g) which was used in the next reaction without further purification. To a solution of 2-[(9H-fluoren-9-yl)methoxy]carbonyl-7-hydroxy-1,2,3,4-tetrahydroisoquinoline-1-carboxylic acid (18.2 g, 43.81 mmol) in MeOH (200 mL) at 0° C was added dropwise SOCl₂ (6.4 mL, 87.62 mmol). The reaction mixture was then heated to 60° C and stirred for 12 h. The reaction mixture was cooled to rt then concentrated under reduced pressure to give a syrupy residue. This residue was dissolved in EtOAc, washed with sat aq. NaHCO₃ solution, brine solution, dried over MgSO₄ and purified by silica gel flash column chromatography (30 to 40% EtOAc/hexane) to give **13** (4.3 g, 10.01 mmol, 18% yield from **12**) as a white foamy solid. Rt and high temperature NMR spectra revealed the presence of two rotamers in an 2:3 ratio (labelled rotamer A and B). ¹H NMR (400 MHz, DMSO-*d*₆) δ 2.56 – 2.79 (m, 2H, CH₂CH₂NFmoc rotamer A and B), 3.45 – 3.72 (m, 4H, OCH₃, CHCO₂CH₃ rotamer A and B), 4.23 – 4.51 (m, 2H, CH₂CH₂NFmoc rotamer A and B), 5.13 – 5.45 (m, 1H, CHFmoc rotamer A and B) 6.61 – 6.71 (m, 1H, ArH rotamer A and B), 6.81 (dd, $J = 2.5$, 37.4 Hz, 1H, ArH rotamer A and B), 7.00 (t, $J = 8.5$, 8.5 Hz, 1H, ArH rotamer A and B), 7.26 – 7.38 (m, 2H, ArH rotamer A and B), 7.38 – 7.48 (m, 2H, ArH rotamer A and B), 7.57 – 7.71 (m, 2H, ArH rotamer A and B), 7.85 – 7.95 (m, 2H, ArH rotamer A and B), 9.39 – 9.46 (m, 1H, OH rotamer A and B). HRMS calculated for C₂₆H₂₃NNaO₅ (M + Na)⁺, 452.1468; found, 452.1443.

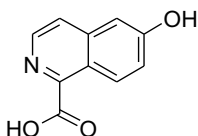
Methyl 7-hydroxyisoquinoline-1-carboxylate (**14**)



According to a modified literature procedure¹² - tetrahydroisoquinoline **13** (4.1 g, 9.55 mmol), DMSO (20 mL) and MeOH (20 mL) were stirred at 60 °C for 12 h, diluted with EtOAc, filtered, the filtrate washed with water and dried over MgSO₄. The solvent was evaporated under reduced pressure to give the Fmoc-deprotected product methyl 7-hydroxy-1,2,3,4-tetrahydroisoquinoline-1-carboxylate, which was then aromatised according to the procedure for **7**, using DDQ (4.4 g, 19.09 mmol) and 1,4-dioxane:THF (1:1 v:v, 100 mL). The crude material was purified by silica gel flash column chromatography (30 to 50% EtOAc/hexane) to provide **14** (1.2 g, 5.90 mmol, 61% yield) as an off-white solid. ¹H NMR (500 MHz, MeOD-*d*₄) δ 4.03 (s, 3H, OCH₃), 7.39 (dd, 1H, *J* = 2.4 Hz, 8.9, ArH), 7.84 – 7.90 (m, 2H, ArH), 8.01 – 8.06 (m, 1H, ArH), 8.31 (d, 1H, *J* = 5.5 Hz, ArH). ¹³C NMR (126 MHz, MeOD-*d*₄) δ 53.11, 107.81, 125.21, 125.86, 129.76, 130.23, 133.68, 139.16, 147.02, 159.46, 167.59. HRMS calculated for C₁₁H₉NNaO₃ (M + Na)⁺, 226.0475; found, 226.0481.

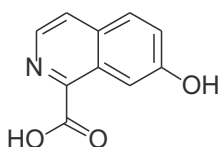
1.1 Experimental details for scheme 2

6-Hydroxyisoquinoline-1-carboxylic acid (**15**)



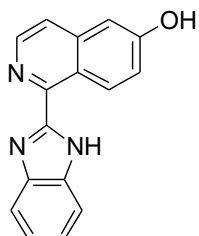
To a solution of **7** (2.6g, 12.79 mmol) in THF (20 mL) at 0°C was added a solution of LiOH (0.92 g, 38.38 mmol) in water (15 mL). The reaction mixture was warmed to rt and stirred for another 12 h. The solvent was evaporated under reduced pressure and the residue was acidified (pH 2.0-3.0) with 2.0 N aq. HCl, which gave a yellow precipitate that was collected by filtration, washed with 5.0 N aq. HCl (50 mL) and dried to give **15** (2.3 g, 12.15 mmol, 96% yield) as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.24 (d, 1H, *J* = 2.5 Hz, ArH), 7.33 (dd, 1H, *J* = 2.5, 9.2 Hz, ArH), 7.85 (dd, 1H, *J* = 0.8, 6.0 Hz, ArH), 8.33 (d, *J* = 5.8 Hz, 1H, ArH), 8.65 (d, 1H, *J* = 9.2 Hz, ArH), 10.78 (br s, 1H, OH). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 107.77, 120.06, 121.75, 122.26, 129.11, 139.05, 139.42, 149.60, 159.96, 165.99. HRMS calculated for C₁₀H₇NNaO₃ (M + Na)⁺, 212.0318; found, 212.0323.

7-Hydroxyisoquinoline-1-carboxylic acid (**16**)



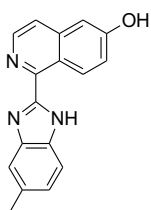
According to the procedure for **15**, a solution of **14** (1.0 g, 4.92 mmol) and LiOH (0.235 g, 9.84 mmol) gave **16** (0.65 g, 3.43 mmol, 70% yield) as yellowish solid. ¹H NMR (500 MHz, D₂O) δ 7.24 – 7.28 (m, 1H, ArH), 7.33 (dd, 1H, *J* = 2.4, 8.9 Hz, ArH), 7.68 (d, 1H, *J* = 5.8 Hz, ArH), 7.80 (d, 1H, *J* = 9.0 Hz, ArH), 8.09 (d, 1H, *J* = 5.7 Hz, ArH). ¹³C NMR (126 MHz, D₂O) δ 110.27, 123.79, 128.39, 128.88, 131.25, 133.19, 139.13, 158.22, 163.05, 177.87. HRMS calculated for C₁₀H₈NO₃ (M + H)⁺, 190.0499; found, 190.0494.

1-(1H-1,3-Benzodiazol-2-yl)isoquinolin-6-ol (**17**)



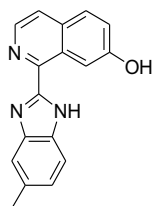
According to a modified literature procedure,¹ a mixture of **15** (0.1 g, 0.49 mmol), *o*-phenylenediamine (0.08 g, 0.74 mmol) and PPA (polyphosphoric acid, ≥83% phosphate as P₂O₅ basis) (~5 g) were heated at 250 °C for 6 h. The resulting viscous black liquid was slowly basified (pH = 8.0 - 10.0) with KOH at 0 °C (caution: exothermic) and extracted with EtOAc. The EtOAc layer was washed with sat aq. NaHCO₃ solution, water, brine solution, dried over MgSO₄ and the residue was purified by silica gel flash column chromatography (10 to 50% EtOAc/hexane) to give **17** (27 mg, 0.10 mmol, 21% yield) as an off-white solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 7.19 (d, 1H, *J* = 2.5 Hz, ArH isoquinoline), 7.27 – 7.38 (m, 3H, ArH isoquinoline, benzimidazole), 7.55 – 7.88 (m, 3H, ArH isoquinoline, benzimidazole), 8.46 (d, 1H, *J* = 5.8 Hz, ArH isoquinoline), 9.25 (d, 1H, *J* = 9.3 Hz, ArH isoquinoline). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 101.42, 108.77, 122.12, 122.31, 123.03, 130.86, 141.11, 142.86, 148.67, 152.44, 160.97. HRMS calculated for C₁₆H₁₂N₃O (M + H)⁺, 262.0975; found, 262.0967. Analytical RP-HPLC R_t = 11.61 min.

1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-ol (**18**)



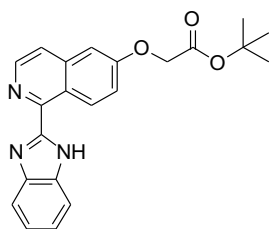
According to the procedure for **17**, a mixture of **15** (0.1 g, 0.49 mmol) and 3,4-diaminotoluene (0.07g, 0.59 mmol) were reacted to give **18** (24 mg, 0.08 mmol, 18% yield) as an off-white solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 2.48 (s, 3H, CH₃), 7.06 – 7.20 (m, 2H, ArH isoquinoline and ArH benzimidazole), 7.24 – 7.32 (m, 1H, ArH isoquinoline), 7.47 (br s, 1H, benzimidazole), 7.53 – 7.69 (m, 2H, ArH isoquinoline and ArH benzimidazole), 8.40 (d, 1H, *J* = 5.7 Hz, ArH isoquinoline), 9.20 (d, 1H, *J* = 9.3 Hz, ArH isoquinoline). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 21.81, 108.74, 121.98, 122.16, 122.94, 125.91, 130.87, 134.42, 141.02, 142.76, 148.70, 152.04, 160.82. HRMS calculated for C₁₇H₁₄N₃O (M + H)⁺, 276.1131; found, 276.1118. Analytical RP-HPLC R_t = 13.26 min.

1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-7-ol (**19**)



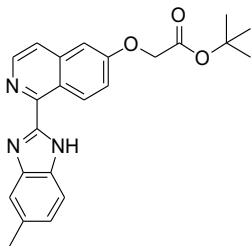
According to the procedure described for **17**, a mixture of **16** (0.2 g, 1.05 mmol) and 3,4-diaminotoluene (0.13 g, 1.05 mmol) were reacted to give **19** (72 mg, 0.26 mmol, 24% yield) as an off-white solid. ^1H NMR (500 MHz, MeOD- d_4) δ 2.51 (s, 3H, CH₃), 7.16 (d, 1H, J = 8.2 Hz, ArH benzimidazole), 7.40 (dd, 1H, J = 2.4, 8.9 Hz, ArH isoquinoline), 7.49 (br s, 1H, ArH benzimidazole), 7.60 (br s, 1H, ArH benzimidazole), 7.78 (d, 1H, J = 5.5 Hz, ArH isoquinoline), 7.88 (d, 1H, J = 8.9 Hz, ArH isoquinoline), 8.44 (d, 1H, J = 5.5 Hz, ArH isoquinoline), 8.74 (d, 1H, J = 2.4 Hz, ArH isoquinoline). ^{13}C NMR (126 MHz, MeOD- d_4) δ 21.82, 49.00, 109.34, 123.37, 124.61, 129.65, 129.98, 133.56, 140.13, 147.22, 158.93. HRMS calculated for C₁₇H₁₄N₃O (M + H)⁺, 276.1131; found, 276.1138. Analytical RP-HPLC R_t = 12.69 min.

tert-Butyl 2-([1-(1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy)acetate (**20**)



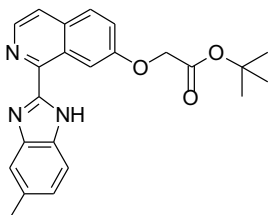
Isoquinoline **17** (45 mg, 0.172 mmol) and K₂CO₃ (48 mg, 0.034 mmol) in anhydrous THF (5 mL) were stirred at 60 °C for 20 min, followed by addition of a solution of *tert*-butyl bromoacetate (40 μL , 0.26 mmol) in anhydrous THF (1 mL). The reaction was stirred at 60 °C for 12 h. The solvent was removed under reduced pressure and the residue was partitioned between EtOAc and saturated aq. NH₄Cl, the EtOAc layer was washed further with brine solution, dried over MgSO₄ and concentrated under reduced pressure. The residue was purified by silica gel flash column chromatography (20 to 30% EtOAc/hexane) to give **20** (60 mg, 0.16 mmol, 93% yield) as an off-white solid. ^1H NMR (400 MHz, CDCl₃) δ 1.51 (s, 9H, C(CH₃)₃), 4.69 (s, 2H, CH₂), 7.03 (d, 1H, J = 2.6 Hz, ArH benzimidazole), 7.28 – 7.34 (m, 2H, ArH benzimidazole), 7.37 – 7.54 (m, 2H, ArH isoquinoline and ArH benzimidazole), 7.58 (d, 1H, J = 5.6 Hz, ArH isoquinoline), 7.94 (br s, 1H, ArH benzimidazole), 8.47 (d, 1H, J = 5.6 Hz, ArH isoquinoline), 10.17 (d, 1H, J = 9.5 Hz, ArH isoquinoline), 11.48 (br s, 1H, NH). ^{13}C NMR (101 MHz, CDCl₃) δ 28.20, 65.77, 82.98, 105.71, 111.28, 120.68, 121.41, 121.82, 122.63, 122.95, 124.24, 131.02, 133.35, 139.31, 141.95, 144.95, 146.40, 151.48, 159.28, 167.46. HRMS calculated for C₂₂H₂₂N₃O₃ (M + H)⁺, 376.1656; found, 376.1628.

***tert*-Butyl 2-[[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy]acetate (21)**



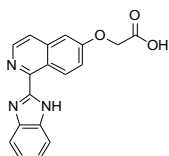
According to the procedure described for **20**, a mixture of **18** (150 mg, 0.54 mmol), K₂CO₃ (151 mg, 1.09 mmol) and *tert*-butyl bromoacetate (0.12 mL, 0.82 mmol) were reacted to give **21** (203 mg, 0.521 mmol, 96% yield) as an off-white solid. ¹H NMR (400 MHz, CDCl₃) δ 1.51 (s, 9H, C(CH₃)₃), 2.50 (s, 3H, CH₃), 4.69 (s, 2H, CH₂), 7.03 (d, 1H, *J* = 2.7 Hz, ArH isoquinoline), 7.10 – 7.17 (m, 1H, ArH benzimidazole), 7.32 (br s, 1H, ArH benzimidazole), 7.48 (dd, 1H, *J* = 2.7, 9.5 Hz, ArH isoquinoline), 7.57 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 7.64 – 7.89 (br s, 1H, ArH benzimidazole), 8.45 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 10.16 (d, 1H, *J* = 9.5 Hz, ArH isoquinoline), 11.24 (br s, 1H, NH). ¹³C NMR (101 MHz, CDCl₃) δ 21.94, 28.20, 65.78, 82.96, 105.71, 111.19, 120.20, 121.33, 121.63, 122.88, 124.56, 131.09, 133.85, 139.30, 141.89, 146.47, 151.14, 159.27, 167.47. HRMS calculated for C₂₃H₂₄N₃O₃ (M + Na)⁺, 390.1812; found, 390.1783.

***tert*-Butyl 2-[[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-7-yl]oxy]acetate (22)**



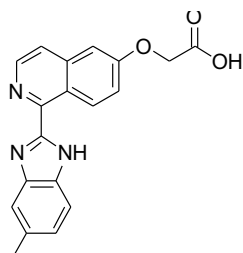
According to the procedure described for **20**, a mixture of **19** (45 mg, 0.16 mmol), K₂CO₃ (68 mg, 0.49 mmol) and *tert*-butyl bromoacetate (0.025 mL, 0.17 mmol) were reacted to give **22** (42 mg, 0.11 mmol, 66% yield) as an off-white solid. ¹H NMR (400 MHz, CDCl₃) δ 1.52 (s, 9H, C(CH₃)₃), 2.52 (s, 3H, CH₃), 4.88 (s, 2H, CH₂), 7.09 – 7.21 (m, 1H, ArH benzimidazole), 7.28 – 7.45 (m, 1H, ArH benzimidazole), 7.52 (dd, 1H, *J* = 2.6, 9.0 Hz, ArH isoquinoline), 7.66 (d, 1H, *J* = 5.4 Hz, ArH isoquinoline), 7.70 – 7.86 (m, 2H, ArH isoquinoline and ArH benzimidazole), 8.46 (d, 1H, *J* = 5.4 Hz, ArH isoquinoline), 9.67 – 9.75 (m, 1H, ArH isoquinoline), 10.78 (br s, 1H, NH). ¹³C NMR (101 MHz, CDCl₃) δ 22.05, 28.27, 65.77, 82.72, 107.06, 110.63*, 110.98*, 120.20*, 120.42*, 122.20, 124.08, 124.19*, 125.81*, 127.92, 128.68, 133.47, 134.34, 140.03, 143.24, 145.19, 151.39, 157.82, 167.83 (*designates carbons linked to broadened benzimidazole protons, as determined by HSQC, gHMBC experiment). HRMS calculated for C₂₃H₂₃N₃NaO₃ (M + Na)⁺, 412.1632; found, 412.1602.

2-[[1-(1H-1,3-Benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetic acid (**23**)



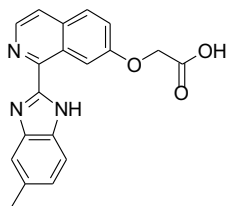
To a solution of **20** (50 mg, 0.13 mmol) in DCM (5 mL) at 0 °C was added trifluoroacetic acid (TFA) (1 mL, 13.32 mmol) and the mixture stirred for 5 h. The DCM and TFA were removed under reduced pressure to provide an oil which solidified to a yellow solid upon co-evaporation with CHCl₃/hexane. This solid was washed with cold MeOH to give **23** (27 mg, 0.08 mmol, 64% yield) as a yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 4.91 (s, 2H, CH₂), 7.19 – 7.35 (m, 2H, ArH benzimidazole), 7.42 (d, 1H, *J* = 2.7 Hz, ArH isoquinoline), 7.52 (dd, 1H, *J* = 2.7, 9.5 Hz, ArH isoquinoline), 7.61 (d, 1H, *J* = 7.8 Hz, ArH benzimidazole), 7.83 (d, 1H, *J* = 8.0 Hz, ArH benzimidazole), 7.87 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 8.59 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 10.04 (d, 1H, *J* = 9.5 Hz, ArH isoquinoline), 13.17 (br m, 2H). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 64.64, 105.98, 112.05, 119.66, 121.12, 121.59, 121.82, 121.93, 123.57, 129.87, 133.99, 138.90, 142.21, 144.02, 146.10, 151.25, 158.74, 169.65. HRMS calculated for C₁₈H₁₄N₃O₃ (M + H)⁺, 320.10297; found, 320.1008.

2-[[1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetic acid (**24**)



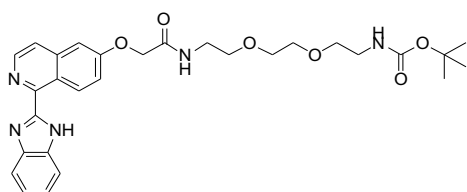
According to the procedure described for **23**, a mixture of **21** (196 mg, 0.503 mmol) and TFA gave **24** (172 mg, 0.51 mmol, quantitative yield) as a yellow solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 2.53 (s, 3H, CH₃), 4.90 (s, 2H, CH₂), 7.21 – 7.32 (m, 1H, ArH benzimidazole), 7.35 (d, 1H, *J* = 2.6 Hz, ArH isoquinoline or ArH benzimidazole), 7.48 (dd, 1H, *J* = 2.6, 9.4 Hz, ArH isoquinoline), 7.56 (s, 1H, ArH benzimidazole), 7.66 (d, 1H, *J* = 8.3 Hz, ArH isoquinoline or ArH benzimidazole), 7.85 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 8.56 (d, 1H, *J* = 5.7 Hz, ArH isoquinoline), 9.00 (d, 1H, *J* = 9.4 Hz, ArH isoquinoline). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 21.79, 65.95, 107.39, 115.10, 115.56, 123.63, 123.70, 124.56, 128.61, 128.81, 133.62, 135.34, 137.94, 140.84, 143.65, 144.56, 149.01, 161.33, 171.68. HRMS calculated for C₁₉H₁₅N₃NaO₃ (M + Na)⁺, 356.1006; found, 356.0997. Analytical RP-HPLC R_t = 13.00 min.

2-{{1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-7-yl}oxy}acetic acid (25)



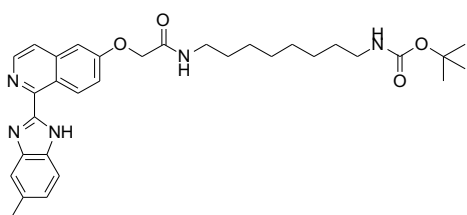
According to the procedure described for **23**, a mixture of **22** (350 mg, 0.90 mmol) and TFA gave **25** (179 mg, 0.54 mmol, 60% yield) as a yellow solid. This compound was used as such for next reaction without further purification.

tert-Butyl N-(2-{2-[2-(2-{{1-(1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl}oxy}acetamido)ethoxy]ethoxy}ethyl)carbamate (26)



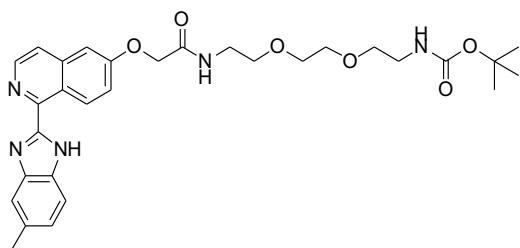
To a solution of **23** (25 mg, 0.08 mmol) and HATU (31 mg, 0.08 mmol) in anhydrous DMF (1 mL), was added DIPEA (30 μ L) and the mixture was stirred for 15 min. A solution of *tert*-butyl {2-[2-(2-aminoethoxy)ethoxy]ethyl}carbamate (prepared according to literature procedure¹³) (40 mg, 0.16 mmol) in DMF (1 mL) was then added and the mixture stirred for 4 h. The solvent was removed under reduced pressure and the resulting residue partitioned between EtOAc and water. The EtOAc layer was washed three times with NH₄Cl solution, once with water and brine solution, dried over MgSO₄ and the solvent evaporated. The residue was purified by silica gel flash column chromatography (50% EtOAc/hexane to 2% MeOH/EtOAc) to provide **26** as pale yellow oil (23 mg, 0.04 mmol, 55% yield). ¹H NMR (500 MHz, CDCl₃) δ 1.41 (s, 9H, C(CH₃)₃), 3.28 (q, 2H, J = 5.4 Hz, CONHCH₂ or CH₂NHBoc), 3.44 – 3.70 (m, 10H, CH₂ PEG), 4.61 (s, 2H, OCH₂CO), 5.06 (s, 1H, NH), 7.05 (s, 1H, ArH isoquinoline), 7.12 (s, 1H, NH), 7.29 – 7.44 (m, 3H, ArH isoquinoline or ArH benzimidazole), 7.62 (d, 1H, J = 5.6 Hz, ArH isoquinoline), 7.73 – 7.83 (m, 2H, ArH isoquinoline or ArH benzimidazole), 8.36 – 8.49 (m, 1H, ArH isoquinoline), 9.77 (d, 1H, J = 9.2 Hz, ArH isoquinoline). ¹³C NMR (126 MHz, CDCl₃) δ 28.54, 29.83, 39.15, 40.44, 67.30, 69.70, 70.24, 70.29, 70.38, 79.53, 106.11, 115.86, 121.64, 122.46, 122.70, 124.41, 130.57, 139.46, 141.55, 145.01, 149.70, 156.20, 158.78, 167.71. HRMS calculated for C₂₉H₃₅N₅NaO₆ (M + Na)⁺, 572.2480; found, 572.2456.

tert-Butyl N-[8-(2-{{1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl}oxy}acetamido)octyl]carbamate (27)



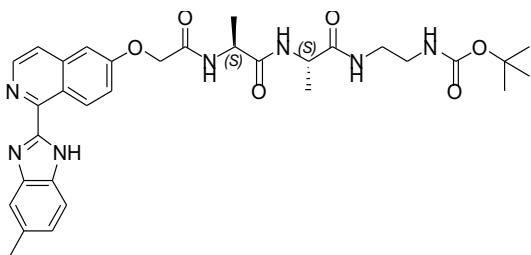
According to the procedure described for **26**, a mixture of **24** (32 mg, 0.09 mmol), *tert*-butyl (8-aminoethyl)carbamate (prepared according to literature procedure¹⁴) (70 mg, 0.29 mmol), HATU (38 mg, 0.10 mmol) and DIPEA (30 μ L, 0.19 mmol) gave a residue that was purified by silica gel flash column chromatography (20 to 50% EtOAc/hexane) to give **27** (37 mg, 0.06 mmol, yield 69%) as a clear oil. ¹H NMR (400 MHz, CDCl₃) δ 1.08 – 1.32 (m, 8H, CONHCH₂CH₂(CH₂)₄), 1.44 (s, 13H, 4H of CONHCH₂CH₂(CH₂)₄CH₂ and 9H of C(CH₃)₃), 2.50 (s, 3H, CH₃), 3.05 (q, 2H, *J* = 6.8 Hz, CONHCH₂ or CH₂NHBoc), 3.35 (q, 2H, *J* = 6.8 Hz, CONHCH₂ or CH₂NHBoc), 4.53 (t, 1H, *J* = 6.0 Hz, NH), 4.66 (s, 2H, OCH₂CO), 6.59 (t, 1H, *J* = 6.0 Hz, NH), 7.09 (d, 1H, *J* = 2.7 Hz, ArH isoquinoline), 7.11 – 7.18 (m, 1H, ArH benzimidazole), 7.43 (dd, 1H, *J* = 2.6, 9.5 Hz, ArH isoquinoline), 7.50 – 7.75 (m, 3H, ArH isoquinoline and ArH benzimidazole), 8.48 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 10.18 (d, 1H, *J* = 9.4 Hz, ArH isoquinoline), 11.16 (br s, 1H, NH benzimidazole). ¹³C NMR (101 MHz, CDCl₃) δ 21.94, 26.79, 26.88, 28.58, 29.25, 29.27, 29.63, 30.11, 39.26, 40.68, 67.39, 79.15, 105.92, 120.96, 121.68, 122.97, 124.79, 131.38, 139.28, 142.27, 146.49, 151.01, 156.14, 158.33, 167.41. HRMS calculated for C₃₂H₄₂N₅O₄ (M + H)⁺, 560.3231; found, 560.3196.

***tert*-Butyl N-(2-{2-[2-(2-{[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)ethoxy]-ethoxy}ethyl)carbamate (**28**)**



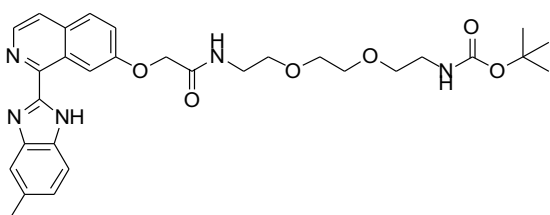
According to the procedure described for **26**, a mixture of **24** (60 mg, 0.18 mmol), *tert*-butyl {2-[2-(2-aminoethoxy)ethoxy] ethyl} carbamate (134 mg, 0.54 mmol), HATU (72 mg, 0.19 mmol) and DIPEA (70 μ L, 0.36 mmol) gave a residue that was purified by silica gel flash column chromatography (50% EtOAc/hexane to 4% MeOH/EtOAc) to give **28** (52 mg, 0.09 mmol, yield 52%) as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 1.42 (s, 9H, C(CH₃)₃), 2.48 (s, 3H, CH₃), 3.25 – 3.32 (m, 2H, CONHCH₂ or CH₂NHBoc), 3.45 – 3.63 (m, 10H, CH₂ peg), 4.65 (s, 2H, OCH₂CO), 5.06 (br s, 1H, NH), 6.97 – 7.18 (m, 3H, NH, ArH benzimidazole, ArH isoquinoline), 7.28 – 7.79 (m, 4H, ArH benzimidazole, ArH isoquinoline), 8.46 (d, 1H, *J* = 5.6 Hz, ArH isoquinoline), 10.15 (d, 1H, *J* = 9.4 Hz, ArH isoquinoline), 11.32 (br s, 1H, NH benzimidazole). ¹³C NMR (101 MHz, CDCl₃) δ 21.90, 28.51, 39.03, 40.39, 67.34, 69.78, 70.19, 70.30, 70.35, 79.40, 105.90, 120.92, 121.63, 122.93, 124.96, 131.31, 133.71, 139.23, 142.18, 146.56, 151.00, 156.08, 158.33, 167.61. HRMS calculated for C₃₀H₃₈N₅O₆ (M + H)⁺, 564.2817; found, 564.2798. Analytical RP-HPLC R_t = 16.42 min.

***tert*-Butyl N-{2-[2-(2-[2-(2-[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)propanamido]propanamido]ethyl}carbamate (29)**



Fmoc-Ala-Ala-CH₂-CH₂-NH-Boc was prepared in 3 steps according to standard step-wise solution-phase peptide synthesis using Fmoc-*L*-Ala-OH, *N*-Boc-ethylenediamine, HBTU, HOBT, DIPEA and diethylamine. This was then Fmoc-deprotected using diethylamine to afford Ala-Ala-CH₂-CH₂-NH-Boc as white solid. According to the procedure described for **26**, a mixture of **24** (45 mg, 0.13 mmol), Ala-Ala-CH₂-CH₂-NH-Boc (122 mg, 0.40 mmol), HATU (54 mg, 0.14 mmol), DIPEA (50 μL, 0.27 mmol) gave a residue that was purified by silica gel flash column chromatography (70% EtOAc/hexane to 4% MeOH/EtOAc) to give **29** (24 mg, 0.04 mmol, 29% yield) as a pale yellow solid. ¹H NMR (500 MHz, MeOD-*d*₄) δ 1.29 – 1.48 (m, 15H, 2×3H of CH₃ belonging to alanines and 9H of C(CH₃)₃), 2.50 (s, 3H, CH₃ benzimidazole), 3.10 – 3.18 (m, 2H, CH₂CH₂NHBoc or CH₂CH₂NHBoc), 3.20 – 3.28 (m, 2H, CH₂CH₂NHBoc or CH₂CH₂NHBoc), 4.25 – 4.35 (m, 1H, CHCONH), 4.48 (q, 1H, *J* = 7.1 Hz, CHCONH), 4.78 (d, 2H, *J* = 3.0 Hz, OCH₂CO), 7.16 (d, 1H, *J* = 8.4 Hz, ArH benzimidazole), 7.32 – 7.36 (m, 1H, ArH isoquinoline), 7.46 – 7.54 (m, 2H, ArH benzimidazole, ArH isoquinoline), 7.60 (br s, 1H, ArH benzimidazole), 7.78 (d, 1H, *J* = 5.69 Hz ArH isoquinoline), 8.52 (d, 1H, *J* = 1.6, 5.6 Hz, ArH isoquinoline), 9.42 (d, 1H, *J* = 9.4 Hz, ArH isoquinoline). ¹³C NMR (126 MHz, MeOD-*d*₄) δ 18.00, 21.83, 28.75, 40.65, 40.79, 50.51, 50.67, 68.05, 80.14, 107.21, 107.22, 122.18, 122.21, 122.83, 123.91, 125.75, 130.91, 131.05, 140.70, 143.45, 148.59, 152.01, 158.49, 160.39, 170.43, 174.47, 175.07. HRMS calculated for C₃₂H₄₀N₇O₆ (M + H)⁺, 618.3035; found, 618.3078.

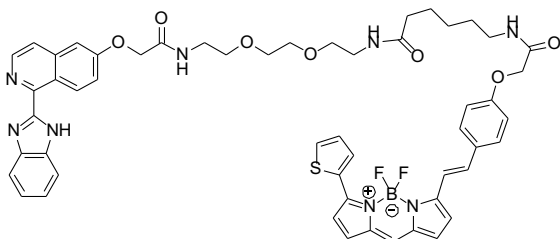
***Tert*-Butyl N-(2-{2-[2-(2-[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-7-yl]oxy}acetamido)-ethoxy]ethoxy}ethyl)carbamate (30)**



According to the procedure described for **26**, a mixture of **25** (20 mg, 0.06 mmol), *tert*-butyl {2-[2-(2-aminoethoxy)ethoxy]ethyl}carbamate (45 mg, 0.18 mmol), HATU (24 mg, 0.06 mmol), DIPEA (20 μL, 0.12 mmol) gave a residue that was purified by silica gel flash column chromatography (50% EtOAc/hexane to 3% MeOH/EtOAc) to give **30** (14 mg, 0.02 mmol, 43% yield) as a white solid. ¹H NMR (400 MHz, CDCl₃) δ 1.42 (s, 9H, C(CH₃)₃), 2.50 (s, 3H, CH₃), 3.27 (q, 2H, *J* = 5.5 Hz, m, 2H, CH₂NH), 3.41 – 3.67 (m, 10H, CH₂ peg), 4.85 (s, 2H, OCH₂CO), 5.04 (m, 1H, *J* = 6.7 Hz, NH), 7.14 (dd, 1H, *J* = 1.5, 8.3 Hz, ArH benzimidazole), 7.19 (br s, 1H, NH), 7.36 – 7.56 (m, 2H, ArH isoquinoline, ArH benzimidazole), 7.65 (d,

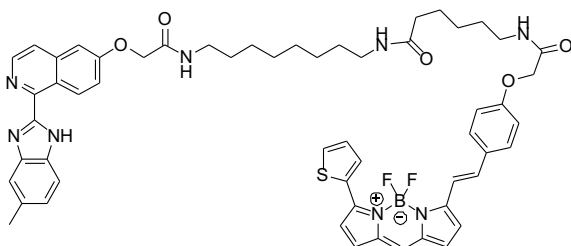
2H, $J = 5.5$ Hz, ArH isoquinoline, ArH benzimidazole), 7.80 (d, 1H, $J = 9.0$ Hz, ArH isoquinoline), 8.46 (d, 1H, $J = 5.4$ Hz, ArH isoquinoline), 9.73 (s, 1H, ArH isoquinoline). ^{13}C NMR (101 MHz, CDCl_3) δ 21.91, 28.53, 39.06, 40.41, 67.71, 69.94, 70.21, 70.30, 70.32, 79.46, 108.44, 122.14, 123.32, 125.20, 127.78, 128.89, 133.55, 140.06, 145.21, 150.80, 156.11, 157.05, 168.02. HRMS calculated for $\text{C}_{30}\text{H}_{38}\text{N}_5\text{O}_6$ ($\text{M} + \text{H}$) $^+$, 564.2817; found, 564.2769. Analytical RP-HPLC $R_t = 16.66$ min.

4-[(E)-2-{4-[(5-{2-[2-(2-{[1-(1H-1,3-Benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)ethoxy]ethoxy}ethyl)carbamoyl]pentyl}carbamoyl]methoxy]phenyl]ethenyl]-2,2-difluoro-12-(thiophen-2-yl)-1 lambda5,3-diaza-2 boratricyclo[7.3.0.0^{3,7}]dodeca-1(12),4,6,8,10-pentaen-1-ylum-2-uid (31)



To a solution of **26** (7 mg, 0.01 mmol) in DCM (2.0 mL) at 0 °C was added TFA (0.5 mL). The reaction mixture was warmed to rt and stirred for 2 h, volatiles were removed under reduced pressure to provide *N*-{2-[2-(2-aminoethoxy)ethoxy]ethyl}-2-{[1-(1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamide trifluoroacetate in quantitative yield. To a solution of this semiprep RP-HPLC purified trifluoroacetate salt (5.2 mg, 9.22 μmol) in DMF (200 μL), was added a solution of DIPEA (6.42 μL , 36.90 μmol , 4 equiv) in DMF (100 μL), followed by addition of solution of BOPIPY630/650-SE (1.7 mg, 2.52 μmol , 1 equiv) in DMF (700 μL). The reaction was stirred for 12 h with the exclusion of light then concentrated under reduced pressure and purified by RP-HPLC, freeze-dried, neutralised with Amberlyst A21 ion exchange resin to give **31** (2.1 mg, 2.11 μmol , 84% yield) as a blue solid. HRMS calculated for $\text{C}_{53}\text{H}_{53}\text{BF}_2\text{N}_8\text{NaO}_7\text{S}$ ($\text{M} + \text{Na}$) $^+$, 1017.3720; found, 1017.3773. Analytical RP-HPLC $R_t = 21.44$ min.

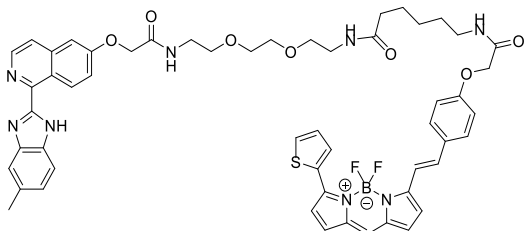
2- Fluoraniumyl-2-fluorescent-4-[(E)-2-(4-{[(5-{[8-(2-{[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}33acetamido)octyl]carbamoyl]pentyl}carbamoyl]methoxy}-phenyl)ethenyl]-12-(thiophen-2-yl)-1 lambda4,3-diaza-2-boratricyclo[7.3.0.0^{3,7}]dodeca-1(12),4,6,8,10-pentaen-2-uide (32)



According to the procedure for **31**, **27** (6 mg, 0.01 mmol) was treated with TFA to give *N*-(8-aminoethyl)-2-{[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamide trifluoroacetate salt in quantitative yield as yellowish solid. This trifluoroacetate salt (3.9 mg, 6.79 μmol) on reaction with BOPIPY630/650-SE

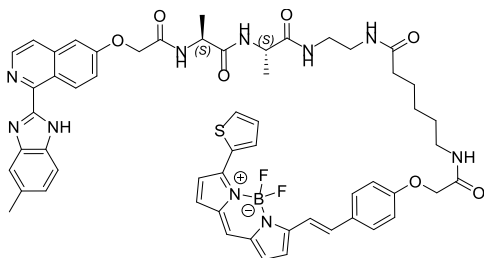
(1.2 mg, 1.9 μmol) provided **32** (0.8 mg, 0.83 μmol , 44% yield) as dark blue solid. HRMS calculated for $\text{C}_{56}\text{H}_{59}\text{BF}_2\text{N}_8\text{NaO}_5\text{S}$ ($\text{M} + \text{Na}$)⁺, 1027.4292; found, 1027.4322. Analytical RP-HPLC R_t = 22.85 min.

[12-(2-{4-[(5-[(2-[2-(2-([1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)ethoxy]ethoxy}ethyl)carbamoyl]pentyl}carbamoyl)methoxy]phenyl)ethenyl)-4-(thiophen-2-yl)-2-(λ^2 -fluoranidyl)-1 λ^4 -aza-3 λ^4 -aza-2 λ^1 -boratricyclo[7.3.0.0^{3,7}]dodeca-3,5,7,9,11-pentaene-2,2,2-triium-1-id-2-yl]- λ^2 -fluoranide (33)



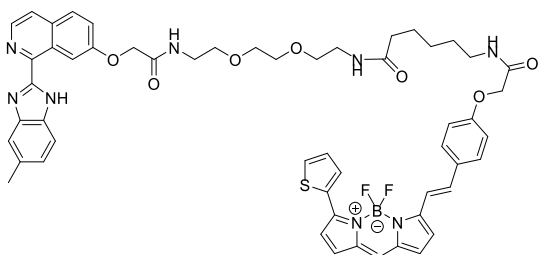
According to the procedure for **31**, **28** (10 mg, 0.02 mmol) was treated with TFA to give *N*-{2-[2-(2-aminoethoxy)ethoxy]ethyl}-2-[[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamide trifluoroacetate salt in quantitative yield as yellowish solid. This trifluoroacetate salt (5.1 mg, 8.83 μmol) on reaction with BOPIPY630/650-SE (1.2 mg, 1.9 μmol) provided **33** (1.0 mg, 0.99 μmol , 54% yield) as dark blue solid. HRMS calculated for $\text{C}_{54}\text{H}_{55}\text{BF}_2\text{N}_8\text{NaO}_7\text{S}$ ($\text{M} + \text{Na}$)⁺, 1031.3877; found, 1031.3959. Analytical RP-HPLC R_t = 21.63 min.

(12-{2-[4-[(5-[(2-[(2S)-2-[(2S)-2-(2-([1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)propanamido]propanamido]ethyl}carbamoyl]pentyl}carbamoyl)methoxy]phenyl)ethenyl)-4-(thiophen-2-yl)-2-(λ^2 -fluoranidyl)-1 λ^4 -aza-3 λ^4 -aza-2 λ^1 -boratricyclo[7.3.0.0^{3,7}]dodeca-3,5,7,9,11-pentaene-2,2,2-triium-1-id-2-yl]- λ^2 -fluoranide (34)



According to the procedure for **31**, **29** (15 mg, 0.02 mmol) was treated with TFA to give (2S)-*N*-[(1S)-1-[(2-aminoethyl)carbamoyl]ethyl]-2-(2-[[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)propanamide trifluoroacetate salt in quantitative yield as yellowish solid. This trifluoroacetate salt (5.5 mg, 8.7 μmol) on reaction with BOPIPY630/650-SE (1.2 mg, 1.9 μmol) provided **34** (0.63 mg, 0.59 μmol , 33% yield) as dark blue solid. HRMS calculated for $\text{C}_{56}\text{H}_{57}\text{BF}_2\text{N}_{10}\text{NaO}_7\text{S}$ ($\text{M} + \text{Na}$)⁺, 1085.4095; found, 1085.4162. Analytical RP-HPLC R_t = 21.05 min.

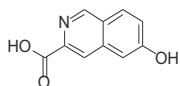
[12-(2-{4-[(5-{2-[2-(2-{1-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-7-yl]oxy}acetamido)ethoxy]ethoxy}ethyl)carbamoyl]pentyl}carbamoyl)methoxy]phenyl}ethenyl)-4-(thiophen-2-yl)-2-(λ^2 -fluoranidyl)-1 λ^4 -aza-3 λ^4 -aza-2 λ^1 -boratricyclo[7.3.0.0^{3,7}]dodeca-3,5,7,9,11-pentaene-2,2,2-triium-1-id-2-yl]- λ^2 -fluoranide (35)



According to the procedure for **31**, **30** (7 mg, 0.01 mmol) was treated with TFA to give N-{2-[2-(2-aminoethoxy)ethoxy]ethyl}-2-[[1-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-7-yl]oxy} trifluoroacetate salt in quantitative yield as yellowish solid. This trifluoroacetate salt **2.32** (5.5 mg, 9.52 μ mol) on reaction with BOPIPY630/650-SE (1.2 mg, 1.9 μ mol) provided **2.38** (0.9 mg, 0.89 μ mol, 49% yield) as dark blue solid. HRMS calculated for C₅₄H₅₅BF₂N₈NaO₇S (M + Na)⁺, 1031.3877; found, 1031.3969. Analytical RP-HPLC R_t = 21.68 min.

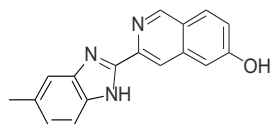
1.3 Experimental details for scheme 3

6-Hydroxyisoquinoline-3-carboxylic acid (36)



Following the procedure described for **15**, a mixture of **11** (0.6 g, 2.95 mmol) and LiOH (0.212 g, 8.86 mmol) gave **36** (0.5 g, 2.64 mmol, 90% yield) as yellow solid. ¹H NMR (400 MHz, DMSO-*d*₆) δ 7.55 – 7.63 (m, 2H, ArH), 8.39 (d, 1H, *J* = 8.7 Hz, ArH), 8.66 (s, 1H, ArH), 9.46 (s, 1H, ArH), 11.79 (br s, 1H, OH). ¹³C NMR (101 MHz, DMSO-*d*₆) δ 109.88, 122.88, 123.65, 123.92, 132.49, 135.02, 139.40, 148.32, 163.39, 163.77. HRMS calculated for C₁₀H₈NO₃ (M + H)⁺, 190.0499; found, 190.0512.

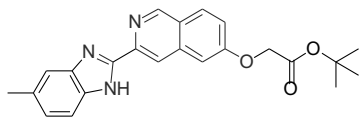
3-(6-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-ol (37)



Following the procedure described for **17**, a mixture of **36** (0.2 g, 1.05 mmol) and 3,4-diaminotoluene (0.13 g, 1.05 mmol) gave **37** (75 mg, 0.27 mmol, 26% yield) as off-white solid. ¹H NMR (400 MHz, MeOD-*d*₄) δ 2.49 (s, 3H, CH₃), 7.12 (d, 1H, *J* = 8.4 Hz, ArH isoquinoline or ArH benzimidazole), 7.20 – 7.30 (m, 2H, ArH isoquinoline and ArH benzimidazole), 7.44 (s, 1H, ArH benzimidazole), 7.54 (s, 1H, ArH benzimidazole), 7.98 (d, 1H, *J* = 8.8 Hz, ArH isoquinoline), 8.43 (s, 1H, ArH isoquinoline), 9.15 (s, 1H, ArH isoquinoline). ¹³C NMR (101 MHz, MeOD-*d*₄) δ 21.78, 109.11, 118.02, 122.20, 125.57, 131.01, 139.78,

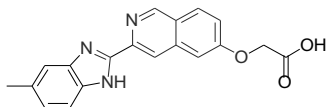
143.17, 152.99, 161.65. HRMS calculated for $C_{17}H_{14}N_3O$ ($M + H$)⁺, 276.1131; found, 276.1114. Analytical RP-HPLC $R_t = 13.83$ min.

***tert*-Butyl 2-{[3-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetate (38)**



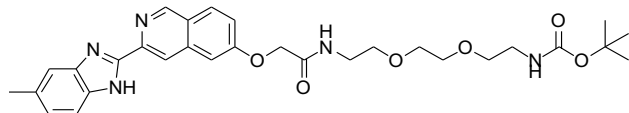
Following the procedure described for **20**, a mixture of **37** (18 mg, 0.06 mmol), K_2CO_3 (25 mg, 0.18 mmol) and *tert*-butyl bromoacetate (20 μ L, 0.13 mmol) gave **38** (24 mg, 0.06 mmol, 94% yield) as an off-white solid. 1H NMR (500 MHz, $CDCl_3$) δ 1.53 (s, 9H, $C(CH_3)_3$), 2.50 (s, 3H, CH_3), 4.68 (s, 2H, CH_2), 7.07 (d, 1H, $J = 2.5$ Hz, ArH isoquinoline or ArH benzimidazole), 7.09 – 7.16 (m, 1H, ArH isoquinoline or ArH benzimidazole), 7.34 (dd, 1H, $J = 2.5, 8.9$ Hz, ArH isoquinoline), 7.40 – 7.82 (s, 2H, ArH benzimidazole), 7.91 (d, 1H, $J = 8.9$ Hz, ArH isoquinoline), 8.68 (s, 1H, ArH isoquinoline), 9.10 (s, 1H, ArH isoquinoline), 10.74 (br s, 1H, NH). ^{13}C NMR (126 MHz, $CDCl_3$) δ 21.92, 28.25, 65.75, 83.14, 106.05, 107.48, 117.69, 121.32, 125.23, 129.84, 138.22, 142.54, 151.42, 159.93, 167.31. HRMS calculated for $C_{29}H_{34}N_3O_5$ ($M + H$)⁺, 504.2493; found, 504.2455.

2-{[3-(5-Methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetic acid (39)



Following the procedure described for **23**, a mixture of **38** (40 mg, 0.10 mmol) and TFA gave **39** in quantitative yield as a yellow solid. 1H NMR (500 MHz, $DMSO-d_6$) δ 2.43 (s, 3H, CH_3), 4.91 (s, 2H, CH_2), 7.06 (d, 1H, $J = 8.2$ Hz, ArH isoquinoline and/or ArH benzimidazole), 7.32 – 7.45 (m, 2H, ArH isoquinoline and/or ArH benzimidazole), 7.46 – 7.55 (m, 2H, ArH isoquinoline and/or ArH benzimidazole), 8.15 (d, 1H, $J = 8.9$ Hz, ArH isoquinoline), 8.66 (s, 1H, ArH isoquinoline), 9.33 (s, 1H, ArH isoquinoline). HRMS calculated for $C_{19}H_{14}N_3O_3$ ($M + H$)⁺, 332.1041; found, 332.1064.

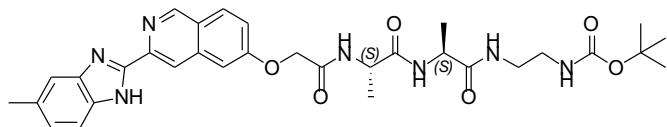
***tert*-Butyl N-(2-[2-[2-(2-{[3-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)-ethoxy]ethoxy]ethyl)carbamate (40)**



According to the procedure described for **26**, a mixture of **39** (17 mg, 0.05 mmol), *tert*-butyl {2-[2-(2-aminoethoxy)ethoxy] ethyl} carbamate (38 mg, 0.15 mmol), HATU (20 mg, 0.05 mmol) and DIPEA (20 μ L, 0.11 mmol) gave a residue that was purified by silica gel flash column chromatography (50% EtOAc/hexane to 10% MeOH/EtOAc) to give **40** (11 mg, 0.02 mmol, 39% yield) as an off-white solid. 1H NMR (400 MHz, $CDCl_3$) δ 1.42 (s, 9H, $C(CH_3)_3$), 2.50 (s, 3H, CH_3), 3.23 – 3.36 (m, 2H, $CONHCH_2$ or CH_2NHBoc), 3.47 – 3.65 (m, 10H, CH_2 peg), 4.66 (s, 2H, OCH_2), 5.04 (br s, 1H, NH), 7.05 (s, 1H, NH), 7.10 – 7.15 (m, 1H, ArH benzimidazole or ArH isoquinoline), 7.17 (d, 1H, $J = 2.4, 2.5$ Hz, ArH benzimidazole or ArH isoquinoline),

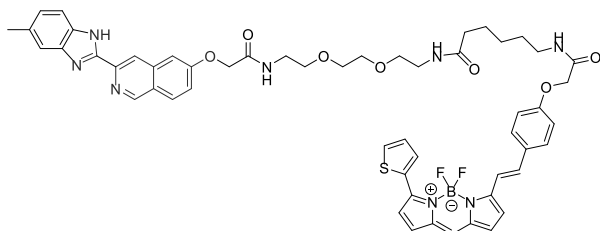
7.31 (dd, 1H, $J = 2.4, 8.9$ Hz, ArH isoquinoline), 7.37 – 7.49 (s, 1H, ArH benzimidazole), 7.50 – 7.70 (s, 1H, ArH benzimidazole), 7.94 (d, 1H, $J = 9.0$ Hz, ArH isoquinoline), 8.69 (s, 1H, ArH isoquinoline), 9.12 (s, 1H, ArH isoquinoline). ^{13}C NMR (101 MHz, CDCl_3) δ 21.91, 28.55, 39.09, 40.46, 67.44, 69.83, 70.35, 70.41, 79.51, 107.07, 118.46, 118.48, 120.82, 125.28, 125.48, 130.05, 133.97, 138.00, 150.33, 151.61, 156.14, 159.10, 167.43. HRMS calculated for $\text{C}_{30}\text{H}_{38}\text{N}_5\text{O}_6$ ($\text{M} + \text{H}$) $^+$, 564.2817; found, 564.2784. Analytical RP-HPLC $R_t = 16.81$ min.

***tert*-Butyl N-{2-[(2S)-2-[(2S)-2-(2-[[3-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)propanamido]propanamido]ethyl}carbamate (41)**



According to the procedure described for **26**, a mixture of **39** (20 mg, 0.06 mmol), Ala-Ala- $\text{CH}_2\text{-CH}_2\text{-NH-Boc}$ (54 mg, 0.18 mmol), HATU (24 mg, 0.06 mmol) and DIPEA (20 μL , 0.12 mmol) gave a residue that was purified by silica gel flash column chromatography (50% EtOAc/hexane to 12% MeOH/EtOAc) to give **41** (16 mg, 0.02 mmol, 37% yield) as an off-white solid ^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ 1.19 (d, 3H, $J = 7.1$ Hz, CH_3 Ala), 1.29 (d, 3H, $J = 7.1$ Hz, CH_3 Ala), 1.35 (s, 9H, $\text{C}(\text{CH}_3)_3$), 2.44 (s, 3H, CH_3 , benzimidazole), 2.91 – 2.99 (m, 2H, CH_2), 3.00 – 3.13 (m, 2H, CH_2), 4.18 – 4.26 (m, 1H, CH), 4.36 – 4.44 (m, 1H, CH), 4.72 – 4.82 (m, 2H, OCH_2), 6.74 (t, 1H, $J = 5.8$ Hz, NH), 7.05 – 7.09 (m, 1H, ArH benzimidazole), 7.42 (s, 1H, ArH benzimidazole), 7.45 (dd, 1H, $J = 2.4, 8.9$ Hz, ArH isoquinoline), 7.49 – 7.55 (m, 2H, ArH benzimidazole or ArH isoquinoline), 7.86 (t, 1H, $J = 5.7$ Hz, NH), 8.11 (d, 1H, $J = 7.5$ Hz, NH), 8.17 (d, 1H, $J = 9.0$ Hz, ArH isoquinoline), 8.36 (d, 1H, $J = 7.4$ Hz, NH), 8.67 (s, 1H, ArH isoquinoline), 9.34 (s, 1H, ArH isoquinoline). ^{13}C NMR (126 MHz, $\text{DMSO-}d_6$) δ 18.19, 18.21, 21.37, 28.21, 38.73, 39.52*, 48.09, 48.32, 66.85, 77.65, 106.67, 117.29, 120.94, 124.07, 124.49, 129.79, 137.57, 142.44, 150.73, 151.60, 155.58, 159.53, 166.84, 171.52, 172.11 (* underneath $\text{DMSO-}d_6$ peaks, identified through HSQC experiment). HRMS calculated for $\text{C}_{32}\text{H}_{40}\text{N}_7\text{O}_6$ ($\text{M} + \text{H}$) $^+$, 618.3035; found, 618.3073.

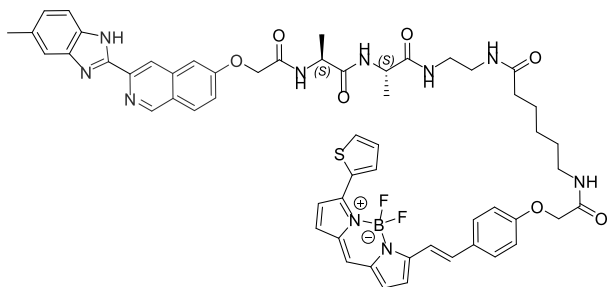
[12-(2-{4-[(5-[(2-2-2-2-[[3-(5-Methyl-1H-1,3-Benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)ethoxy]ethoxy]ethyl}carbamoyl]pentyl}carbamoyl)methoxy]phenyl]ethenyl)-4-(thiophen-2-yl)-2-(λ^2 -fluoranidyl)- $1\lambda^4$ -aza- $3\lambda^4$ -aza- $2\lambda^1$ -boratricyclo[7.3.0.0 3,7]dodeca-3,5,7,9,11-pentaene-2,2,2-triium-1-id-2-yl]- λ^2 -fluoranide (42)



According to the procedure for **31**, **40** (5.0 mg, 0.01 mmol) was treated with TFA to give *N*-{2-[2-(2-aminoethoxy)ethoxy]ethyl}-2-[[3-(5-methyl-1H-1,3-benzodiazol-2-yl)isoquinolin-6-

yl]oxy}acetamide trifluoroacetate salt in quantitative yield as yellowish solid, this trifluoroacetate salt (5.3 mg, 0.01 mmol) on reaction with BOPIPY630/650-SE (1.7 mg, 2.52 μ mol) provided **42** (2.2 mg, 2.18 μ mol, 86% yield) as dark blue solid. HRMS calculated for $C_{54}H_{55}BF_2N_8NaO_7S$ ($M + Na$)⁺, 1031.3877; found, 1031.3849. Analytical RP-HPLC R_t = 20.96 min.

(12-{2-[4-({5-({2-[(2S)-2-[(2S)-2-(2-{[3-(5-methyl-1H-1,3-Benzodiazol-2-yl)isoquinolin-6-yl]oxy}acetamido)propanamido]propanamido]ethyl}carbamoylethyl)carbamoylethyl)methoxy)phenyl]ethenyl}-4-(thiophen-2-yl)-2-(λ^2 -fluoranidyl)-1 λ^4 -aza-3 λ^4 -aza-2 λ^1 -boratricyclo[7.3.0.0^{3,7}]dodeca-3,5,7,9,11-pentaene-2,2,2-triium-1-id-2-yl)- λ^2 -fluoranide (43)



According to the procedure for **31**, **41** (9.0 mg, 0.01 mmol) was treated with TFA to give (2S)- N- [(1S)- 1- [(2- aminoethyl)carbamoylethyl]- 2- (2- {[3- (5- methyl- 1H- 1,3- benzodiazol- 2- yl)isoquinolin- 6- yl]oxy}acetamido)propanamide trifluoroacetate salt, this trifluoroacetate salt (9.2 mg, 0.01 mmol) on reaction with BOPIPY630/650-SE (1.7 mg, 2.52 μ mol) provided **43** (1.9 mg, 1.83 μ mol, 73% yield) as dark blue solid. HRMS calculated for $C_{56}H_{57}BF_2N_{10}NaO_7S$ ($M + Na$)⁺, 1085.4095; found, 1085.4166. Analytical RP-HPLC R_t = 20.51 min.

1.4 Excitation and emission spectra of fluorescent compounds

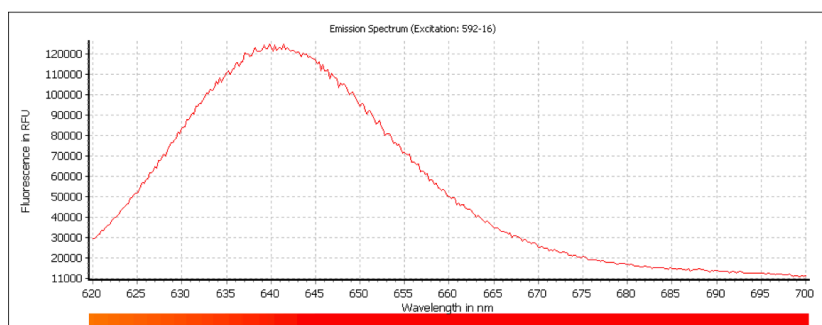
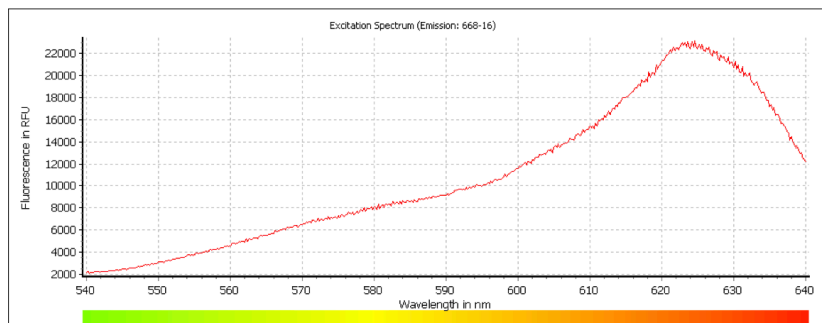
The excitation and emission spectra for each fluorescent compound (10 μ M in methanol) were measured in Griener Bio-One white 96 well flat bottom plates using a BMG LabTech CLARIOstar read type 'top optic' (software version 5.20 RS; firmware version 1.20) and analysed using BMG MARS (software version 3.10 RS). Alexa Flour 633 presets were used. Excitation wavelength 540 – 640 nm (1.0 stepwidth), excitation bandwidth 10 nm, emission wavelength 668 nm, emission bandwidth 16 nm, gain 1500, measured 0.2 sec. And excitation wavelength 592 nm excitation bandwidth 16 nm, emission wavelength 620 – 700 nm (0.2 stepwidth), emission bandwidth 10 nm, gain 1500, measured 0.2 sec.

Summary of results:

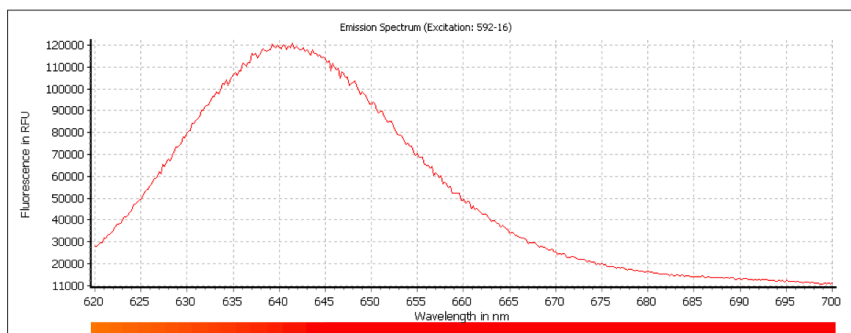
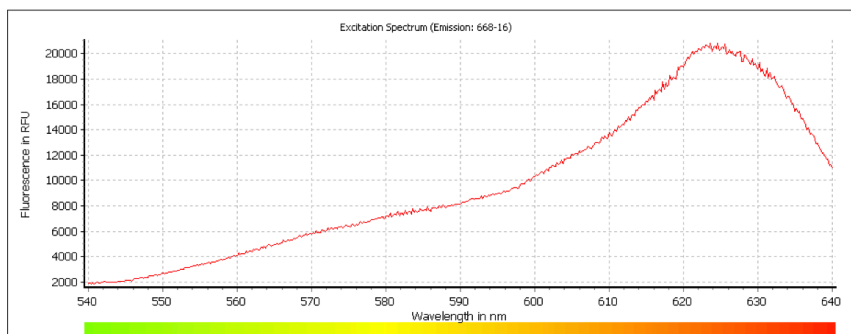
Fluorescent compound	Excitation (max) (nm)	Emission (max) (nm)
31	624	641
33	624	641
32	624	641
34	624	641
35	624	642
42	624	642
43	624	641

Representative spectra: Fluorescent ligands 31 and 33, from Figure 2

Compound 31 -0.2nm resolution



Compound 33 – 0.2nm resolution



2. Conformers of **18**

Benzimidazoles are well known for exhibiting tautomerism,¹⁵⁻²⁰ however the (benzimidazolyl)isoquinolinol scaffold has not been investigated as such. Since multiple sets of peaks were observed in initial NMR spectra of **17-19**, we sought to investigate this using **18** as a case study. Tautomers of **18** would constitute the chemical exchange of protons between N9 and N11 (Figure S1.A), usually with the participation of solvent molecules. Rotamer conformers could also be present, due to hindered rotation around a single bond; for **18** this would be around the C1-C10 bond connecting the benzimidazole and isoquinoline heterocycles (Figure S1.B). If it is conformers rather than discrete isomers that are present, then peak appearance and multiplicity in NMR and HPLC spectra should be sensitive to factors such as solvent (hydrogen bond acceptor or donor ability), temperature and concentration of the compound because the rate of conformer interconversion is likely affected by these factors. Also the atoms nearest to the point of difference between potential conformers should show the most non-equivalent NMR signals between conformers. Therefore, NMR and HPLC studies were carried out using **18** as a representative example from the compounds **17-19** to confirm the presence of interconvertible conformers versus discrete isomers/regioisomers.

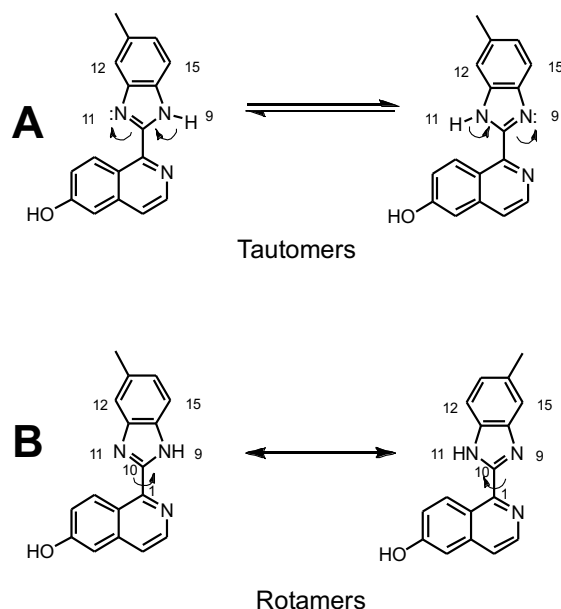


Figure S1: Interconversion of possible (A) tautomers or (B) rotamers of **18**.

2.1 NMR studies

The ¹H NMR spectrum of **18** in MeOD-*d*₄ showed the protons belonging to the isoquinoline ring as sharp peaks while protons of the benzimidazole were broadened peaks (Figure S2.A). The ¹³C NMR spectrum of **18** in MeOD-*d*₄ showed carbons of the isoquinoline ring as sharp peaks compared to a low intensity or absence of benzimidazole carbon signals (Figure S2.B) (refer to Experimental section in paper for peak lists in MeOD-*d*₄). ¹H and ¹³C NMR spectra signals were assigned using gCOSY (Figure S3), HSQC (Figure S4) and gHMBC (Figure S5) spectra. However, more proton signals were seen in the ¹H NMR spectrum of **18** in DMSO-*d*₆ than observed in MeOD-*d*₄ (Figure S6.A) and correspondingly in the ¹³C NMR spectrum (Figure S6.B).

^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ 2.45 (s, 2.84H, 16-H), 7.07 (d, 0.62H, $J = 7.6$ Hz, 14-H), 7.12 (d, 0.41H, $J = 8.0$ Hz, 14-H), 7.21 (d, 1H, $J = 2.4$ Hz, 5-H), 7.33 – 7.40 (m, 1.53H, 7-H and 12-H), 7.47 (d, 0.41H, $J = 8.3$ Hz, 15-H), 7.60 (s, 0.41H, 12-H), 7.68 (d, 0.60H, $J = 8.3$ Hz, 15-H), 7.74 (d, 0.96H, $J = 5.6$ Hz, 4-H), 8.49 (d, 1.0H, $J = 5.6$ Hz, 3-H), 9.90 – 9.99 (m, 1.02H, 8-H), 10.53 (s, 1.53H, 17-H), 12.98 (s, 0.63H, 9-H), 12.99 (s, 0.40H, 9-H). (Additional ^1H NMR signals, than expected by magnetic equivalence for one chemical structure of **18**, are due to presence of another conformer)

^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ 21.33 (16-C), 21.48 (16-C), 107.63 (5-C), 111.52 (15-C), 111.59 (12-C), 119.18 (12-C and 15-C), 120.78 (4-C), 121.07 (7-C), 123.54 (14-C), 125.02 (14-C), 130.12 (8-C), 130.78 (13-C), 132.01 (11a-C or 15a-C), 132.86 (13-C), 134.21 (11a-C or 15a-C), 139.16 (8a-C or 4a-C), 141.68 (3-C), 142.24 (11a-C or 15a-C), 144.40 (11a-C or 15a-C), 146.19 (8a-C or 4a-C), 150.97 (10-C), 151.31 (1-C), 158.93 (6-C). (Additional ^{13}C NMR signals, than expected by magnetic equivalence for one chemical structure of **18**, are due to presence of another conformer)

The ^{13}C NMR spectrum of **18** in $\text{DMSO-}d_6$ revealed 23 aromatic carbon signals, however only 17 carbons are present in **18**. The HSCQ spectrum (Figure S8) of **18** showed 12 C-H correlations, however **18** has only nine carbons attached to at least one hydrogen. These HSQC correlations with data from gCOSY (Figure S7) and gHMBC spectra (Figure S9) revealed that additional three C-H correlations in HSQC spectrum and additional carbon signals in ^{13}C NMR spectrum were from atoms belonging to the benzimidazole moiety. Assignment of atoms to the benzimidazole moiety was guided by HSQC and gHMBC correlations to clearly identifiable H-16 protons.

Variable temperature ^1H NMR spectra

The rate of interconversion of conformers is dependent on temperature. Therefore, a complex NMR spectrum of two conformers simplifies at higher temperature as the rate of conversion of conformers increases and reaches beyond that observable on NMR timescale. ^1H NMR spectra carried out on **18** in $\text{DMSO-}d_6$ at three different temperatures (Figure S10) showed coalescence of proton signals as temperature increased (Figure S10), supporting the presence of conformers rather than discrete non-interchangeable isomers.

Addition of D_2O

One reason for the difference in complexity of the NMR spectra signals for **18** in $\text{MeOD-}d_4$ versus $\text{DMSO-}d_6$ could be the strong hydrogen bond accepting property of $\text{DMSO-}d_6$ compared to $\text{MeOD-}d_4$. Thus, if conformers are present, addition of D_2O to a solution of **18** dissolved in $\text{DMSO-}d_6$ should weaken a hydrogen bond between an NH of benzimidazole and $\text{DMSO-}d_6$ and thus increase rate of interconversion of tautomers. In agreement with this, less complex signals were observed in both ^1H and ^{13}C NMR spectra of **18** in $\text{DMSO-}d_6$ spiked with D_2O (Figure S11, Figure S12).

Tautomer or rotamer?

NMR spectra indicated it is highly likely tautomers are present in **18** because it is the benzimidazole atoms most affected by the change in solvent and temperature. If it was predominantly rotamers then the atoms belonging to isoquinoline would also be affected (FigureS1(B)). In MeOD-*d*₄, exchange of hydrogen between N9 and N11 exposes neighbouring atoms: 11a, 12, 13, 14, 15 and 15a to a fluctuating local chemical environment resulting in either broadening or absence of ¹H/¹³C NMR signals of their corresponding atoms (FigureS1(A)). Compound **18** in DMSO-*d*₆ presumably makes a strong hydrogen bond with the NH benzimidazole, thus slowing down the rate of exchange of hydrogen between N9 and N11 and consequently rate of interconversion of tautomers, thus leading to observation of both tautomers in NMR spectra time scale (FigureS1(A)). The multiplicity of 8-H in DMSO-*d*₆ is presumably due to its closeness to benzimidazole moiety exhibiting tautomerism.

2.2 RP-HPLC studies

Chromatograms from analytical HPLC of **17-19** revealed that peak shape was concentration dependent. Again using **18** as the model compound, at higher concentrations an elongated, square shoulder peak was observed (Figure S13). We hypothesised that the peculiar peak shape was either due to the presence of conformers, peak tailing, or the presence of chemical impurities/non-interconvertible isomers. To rule out that the shoulder peak is not due to chemical impurities/non-interconvertible isomers, further RP-HPLC experiments were done. Semi-preparative RP-HPLC purification of **18** was performed whereby the peak (Figure S14) was fractionated into 10 separate fractions. An analytical HPLC of each fraction was carried out (Figure S15 A), which showed broad peaks for fractions with high concentration and sharp peaks for samples with dilute concentration. An aliquot from each fraction was combined and analysed by analytical RP-HPLC at a dilute concentration, which showed a single peak (figure S15(B)), thus making the presence of non-interconvertible or discrete chemical impurities unlikely. To investigate the effect of temperature on the more concentrated analytical HPLC chromatogram of **18** (Figure S13), HPLC experiments were carried out at 25-40 °C using a column oven. However no significant change in chromatogram peak shape was observed in this temperature range (results not shown). A much higher temperature is likely needed to see any significant change in chromatogram of **18**, in alignment with ¹H NMR spectra results that showed little change from 25 °C to 45 °C but a much larger change at 60 °C. Higher temperature HPLC experiments were not carried out because of solvent and column incompatibility with higher temperatures.

2.3 Summary and Conclusions

NMR spectra and HPLC experiments indicated **18** is a mixture of interconvertible tautomers. The synthetic route to **18** made the presence of regioisomers very unlikely, and this was reinforced by the high temperature experiments since discrete regioisomers would not be interconvertible and the ratio would be the same at low and high temperature (which it was not). As the HRMS spectrum of **18** matched the expected molecular formula mass and no other parent peaks were observed the possibility of an impurity with different molecular formula was also ruled out.

2.4 NMR Spectra and HPLC chromatograms

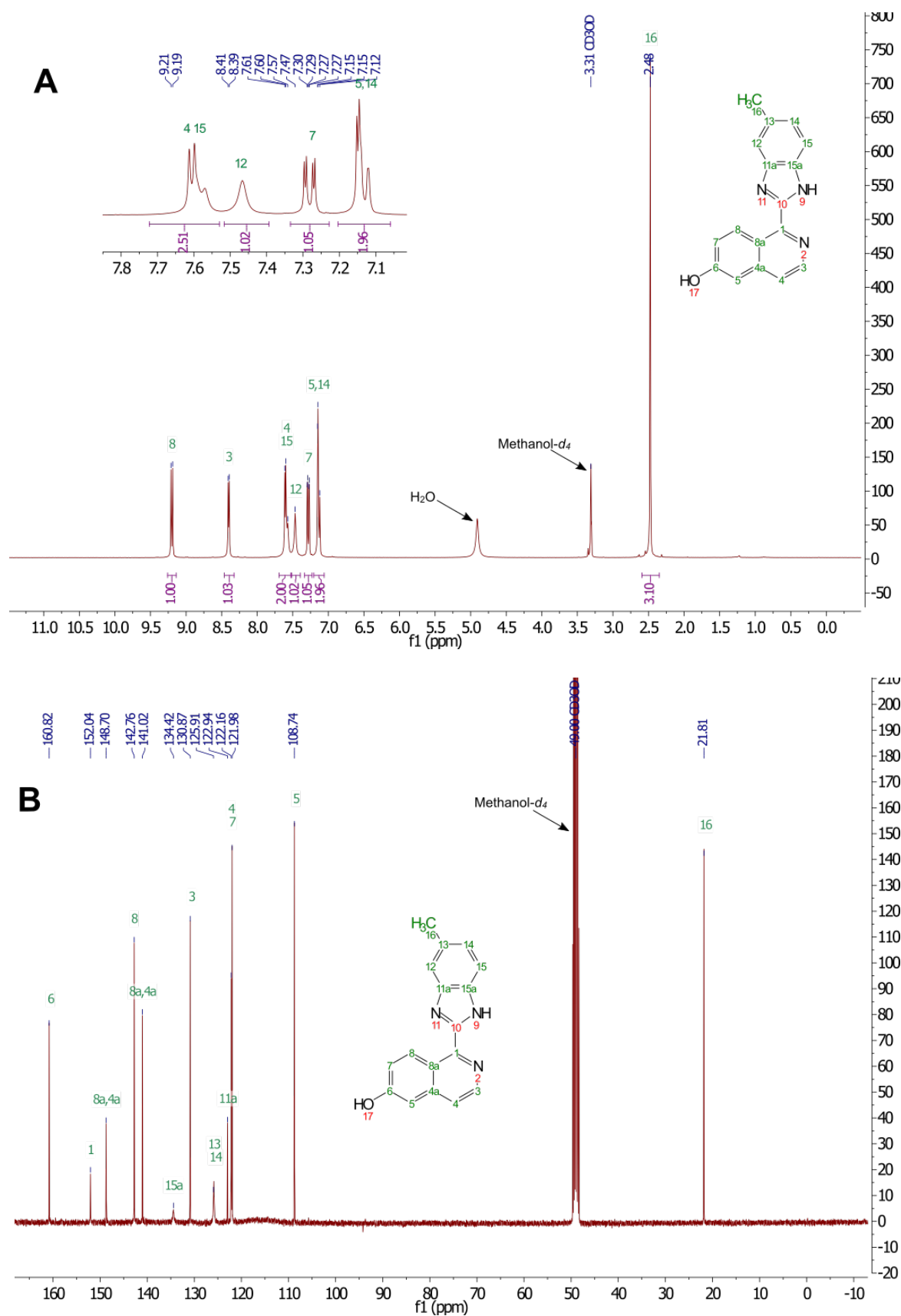


Figure S2. NMR spectra of **18** in MeOD- d_4 at 25 °C: (A) ^1H NMR (inset shows aromatic region) and (B) ^{13}C NMR spectrum.

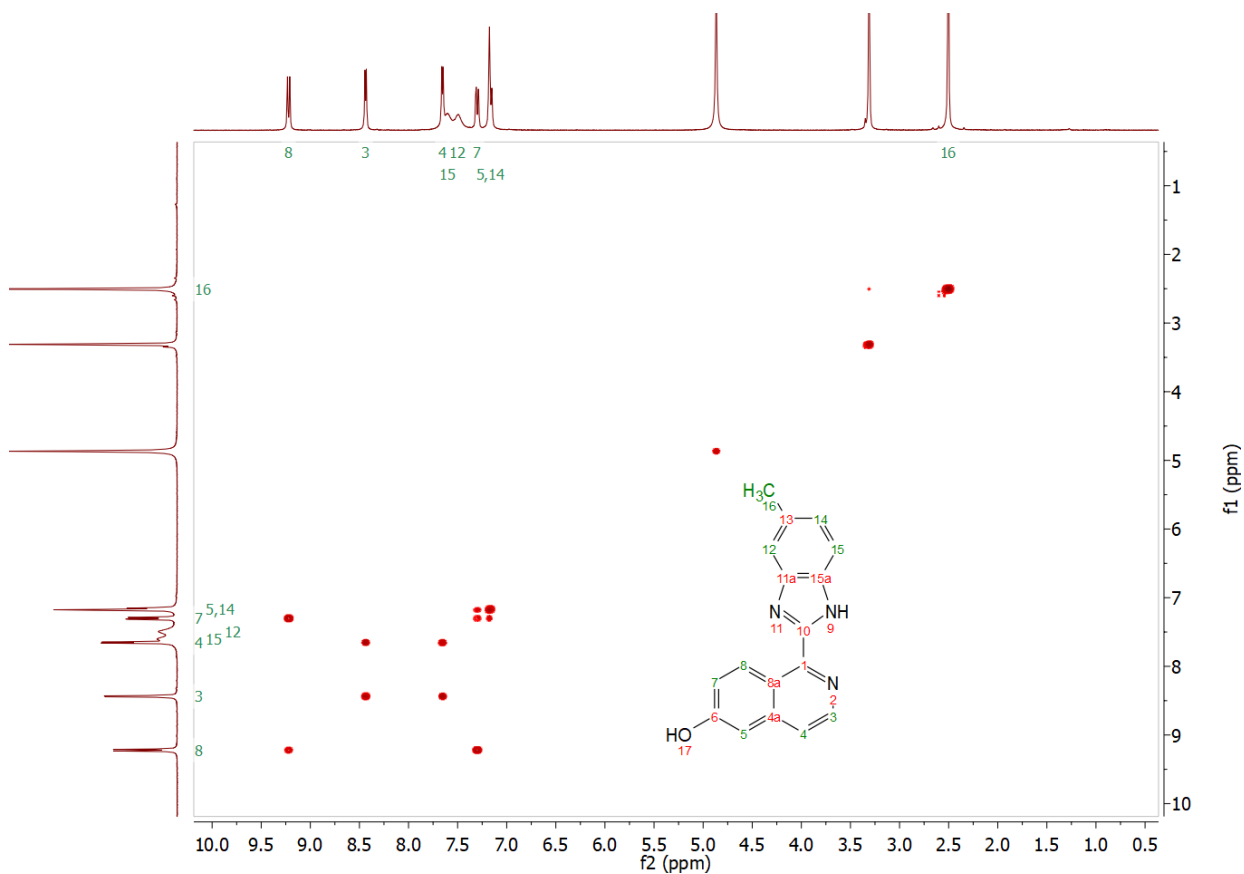


Figure S3. gCOSY spectrum of **18** in MeOD-*d*₄ at 25 °C

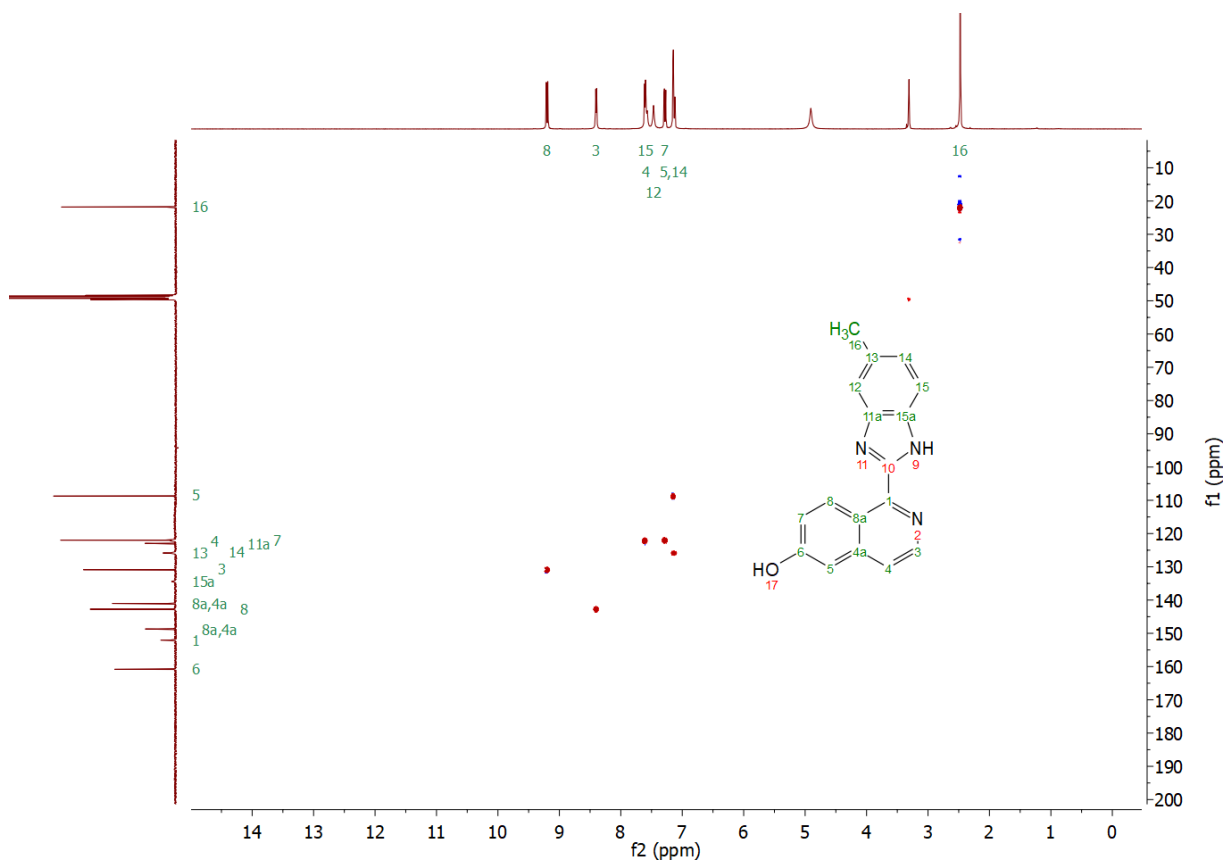


Figure S4. gHSQC spectrum of **18** in MeOD-*d*₄ at 25 °C

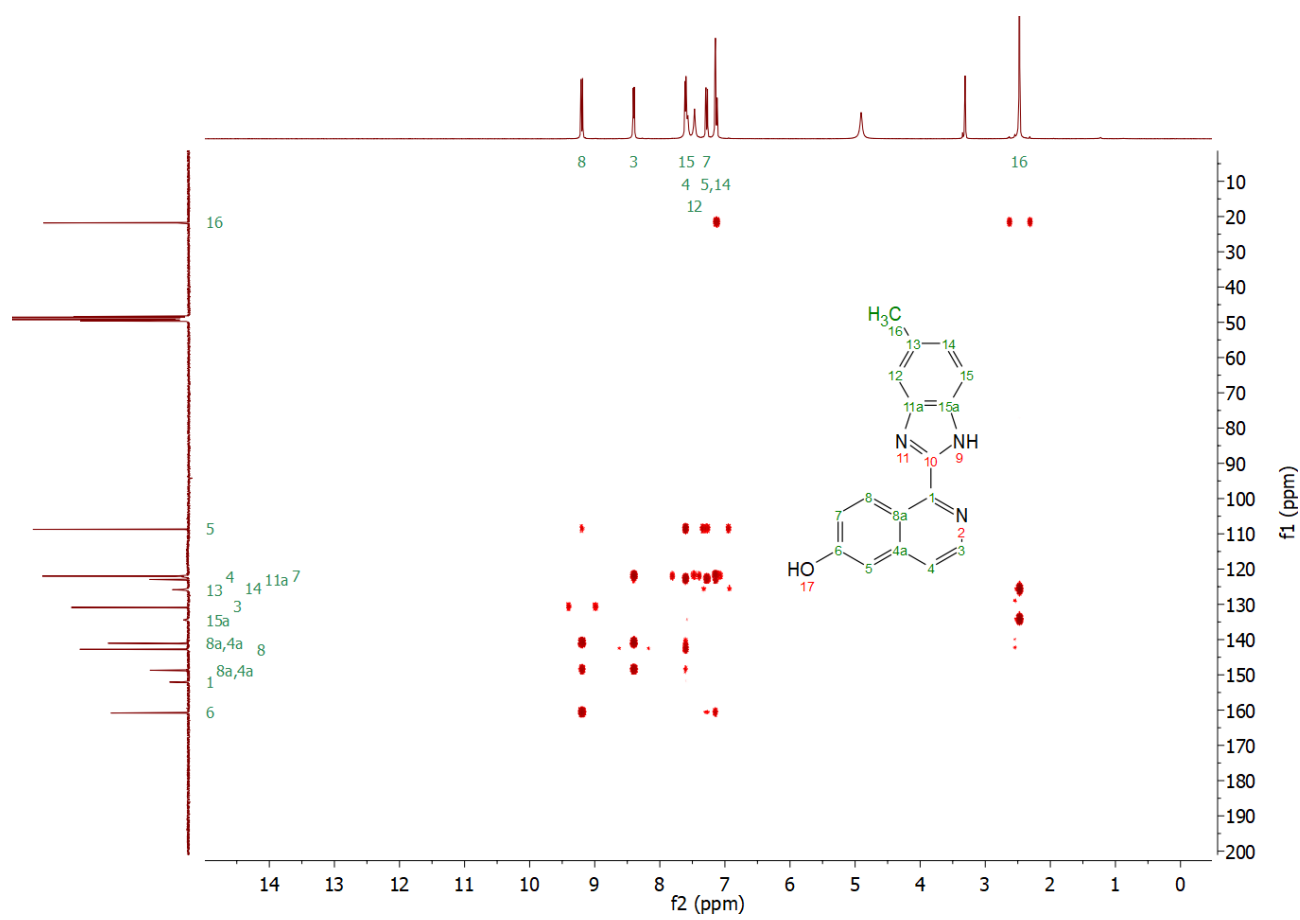


Figure S5. gHMBC spectrum of **18** in MeOD-*d*₄ at 25 °C

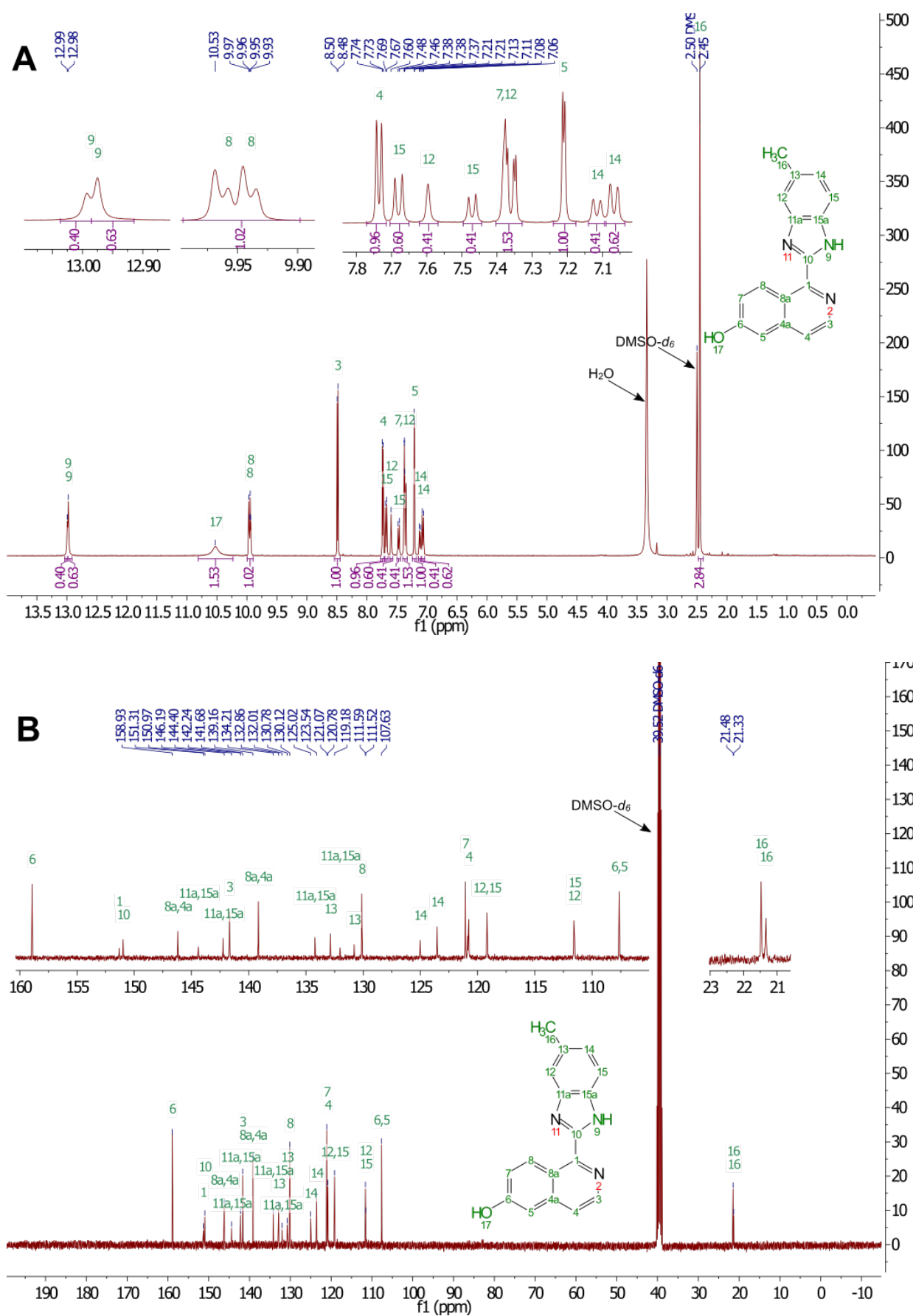


Figure S6. NMR spectra of **18** in DMSO-*d*₆ at 25 °C: (A) ¹H NMR (inset shows aromatic region) and (B) ¹³C spectrum (inset shows expanded view of ¹³C spectrum).

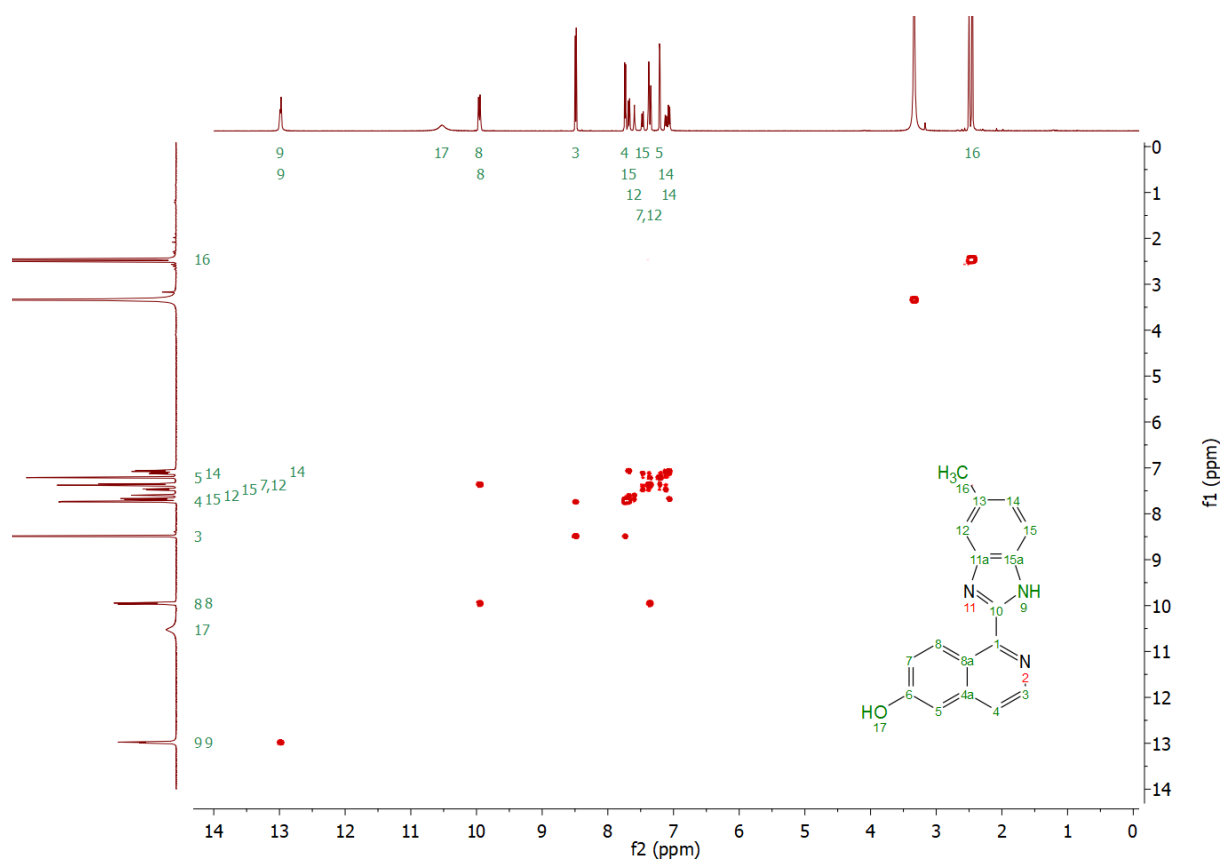


Figure S7. gCOSY spectrum of **18** in DMSO- d_6 at 25 °C

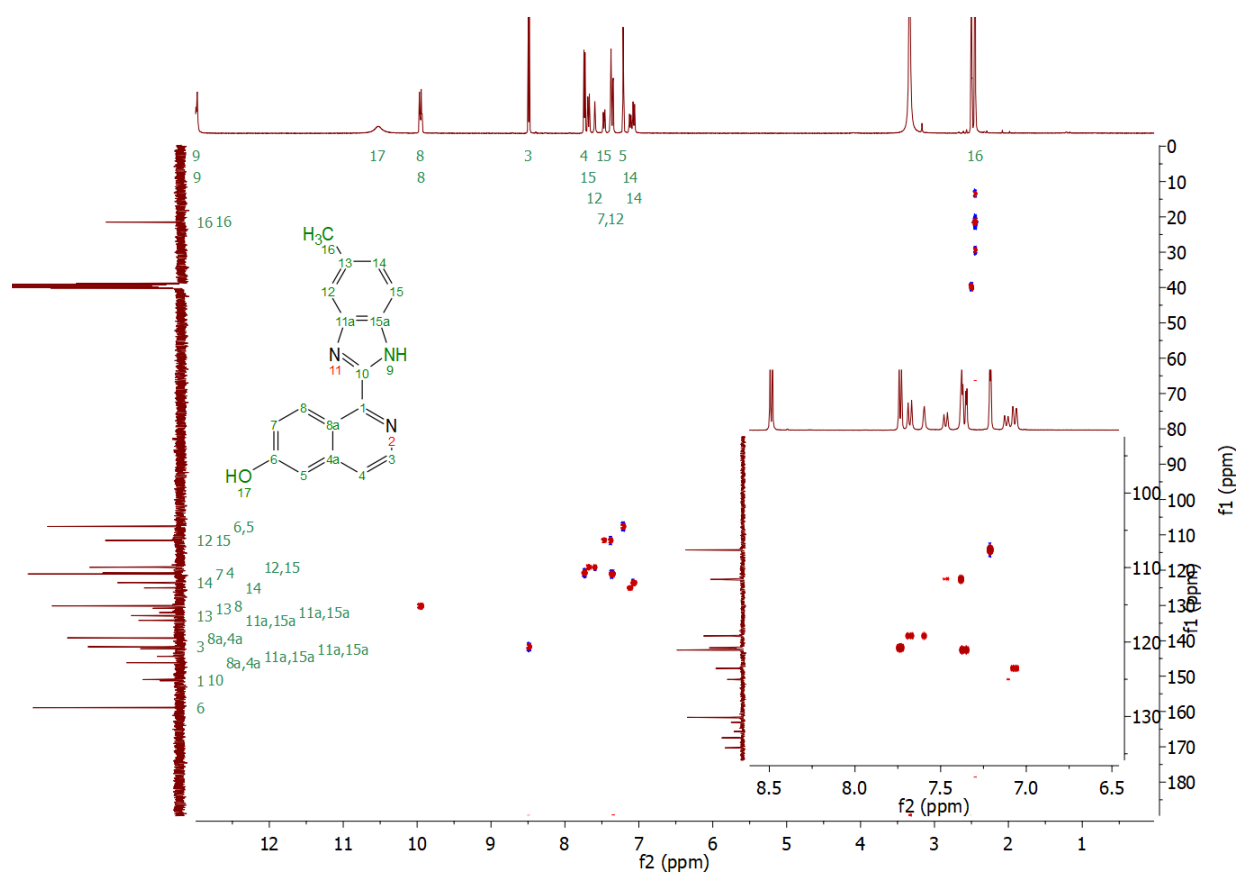


Figure S8. HSQC spectrum (inset shows aromatic region) of **18** in DMSO- d_6 at 25 °C

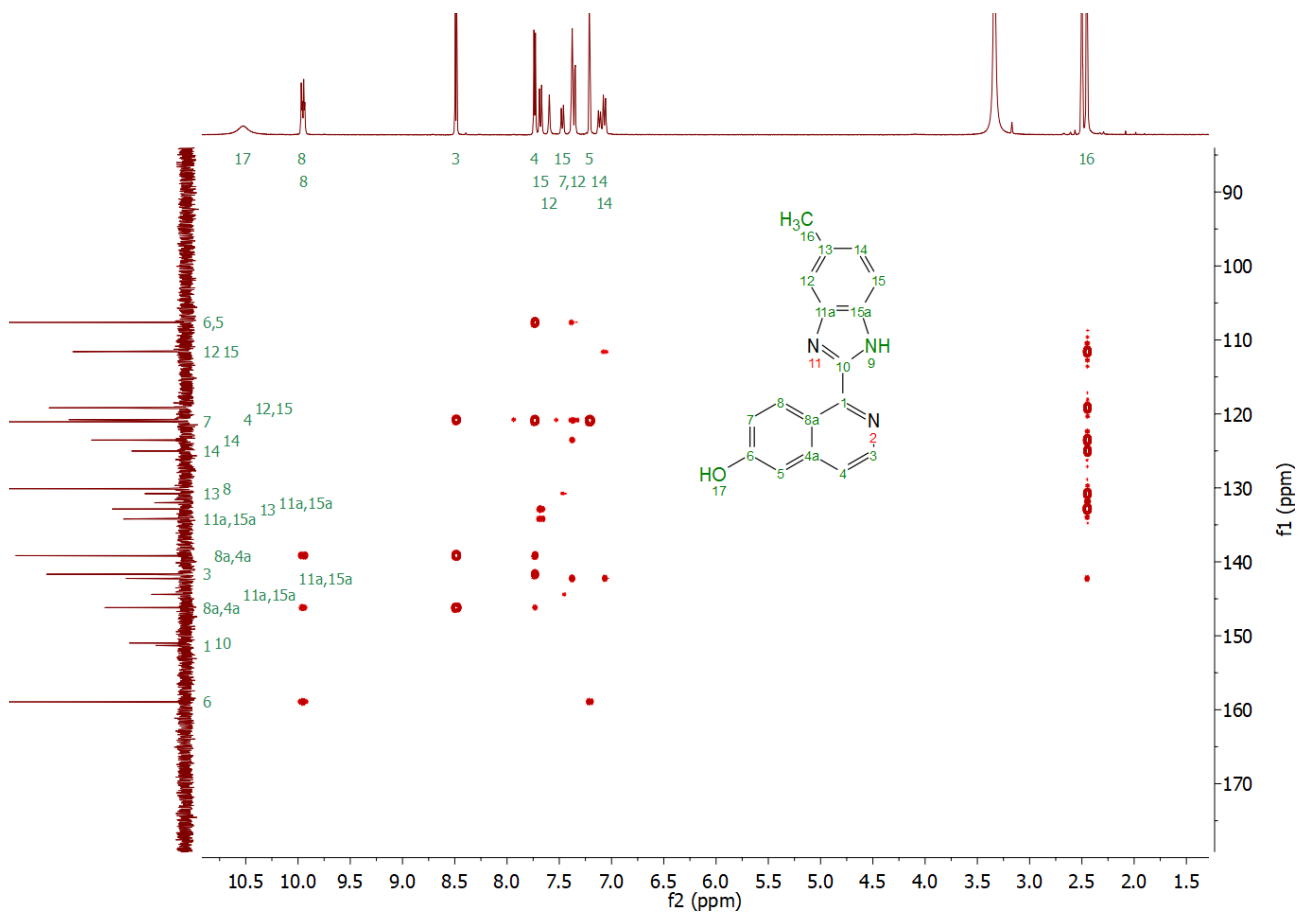


Figure S9. gHMBC spectrum of **18** in DMSO- d_6 at 25 °C

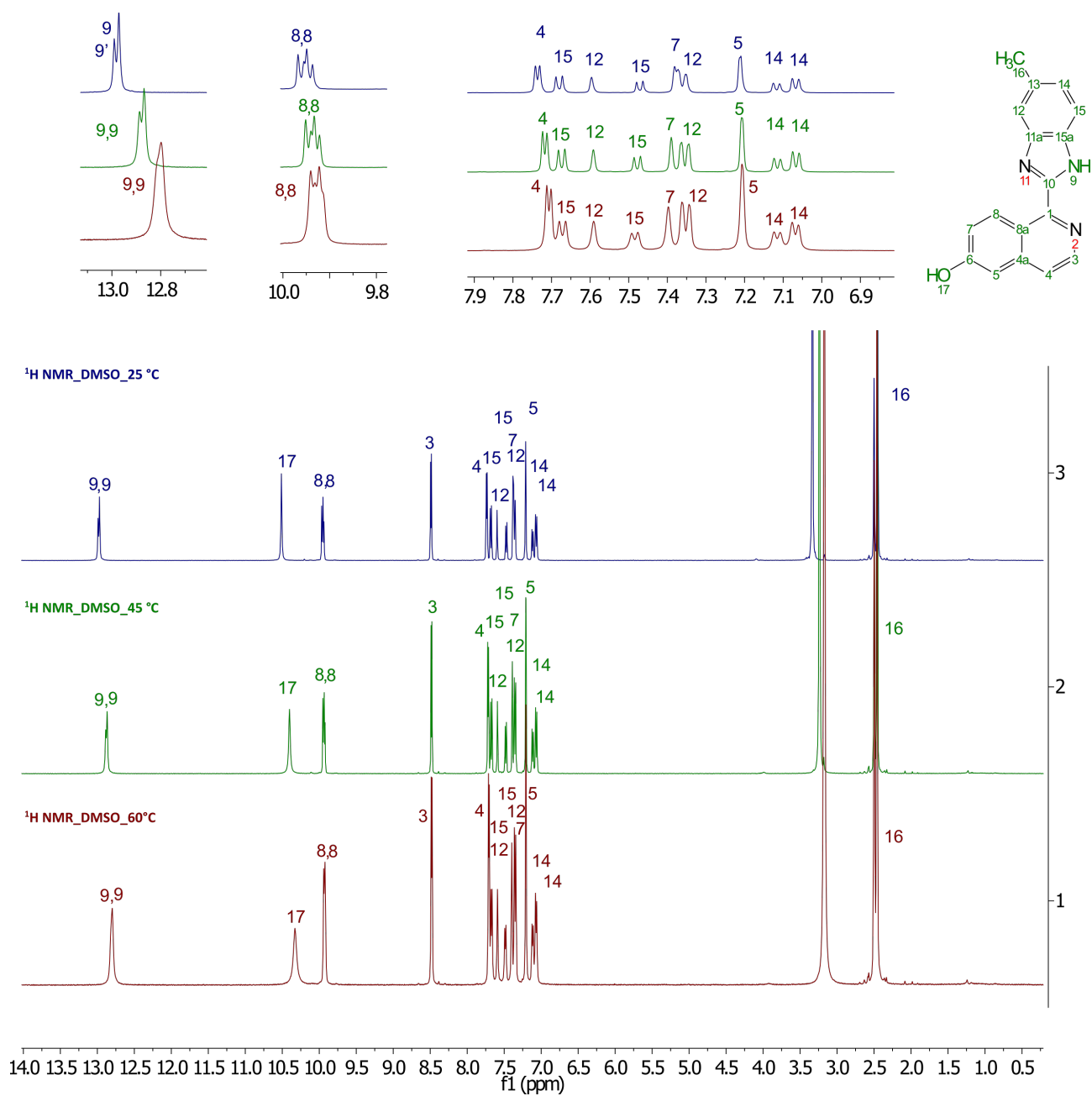


Figure S10: ^1H NMR experiments of **18** in $\text{DMSO}-d_6$ were carried at three different temperatures: Top (25 °C in blue), middle (45 °C in green), bottom (60 °C in red), inset shows aromatic region.

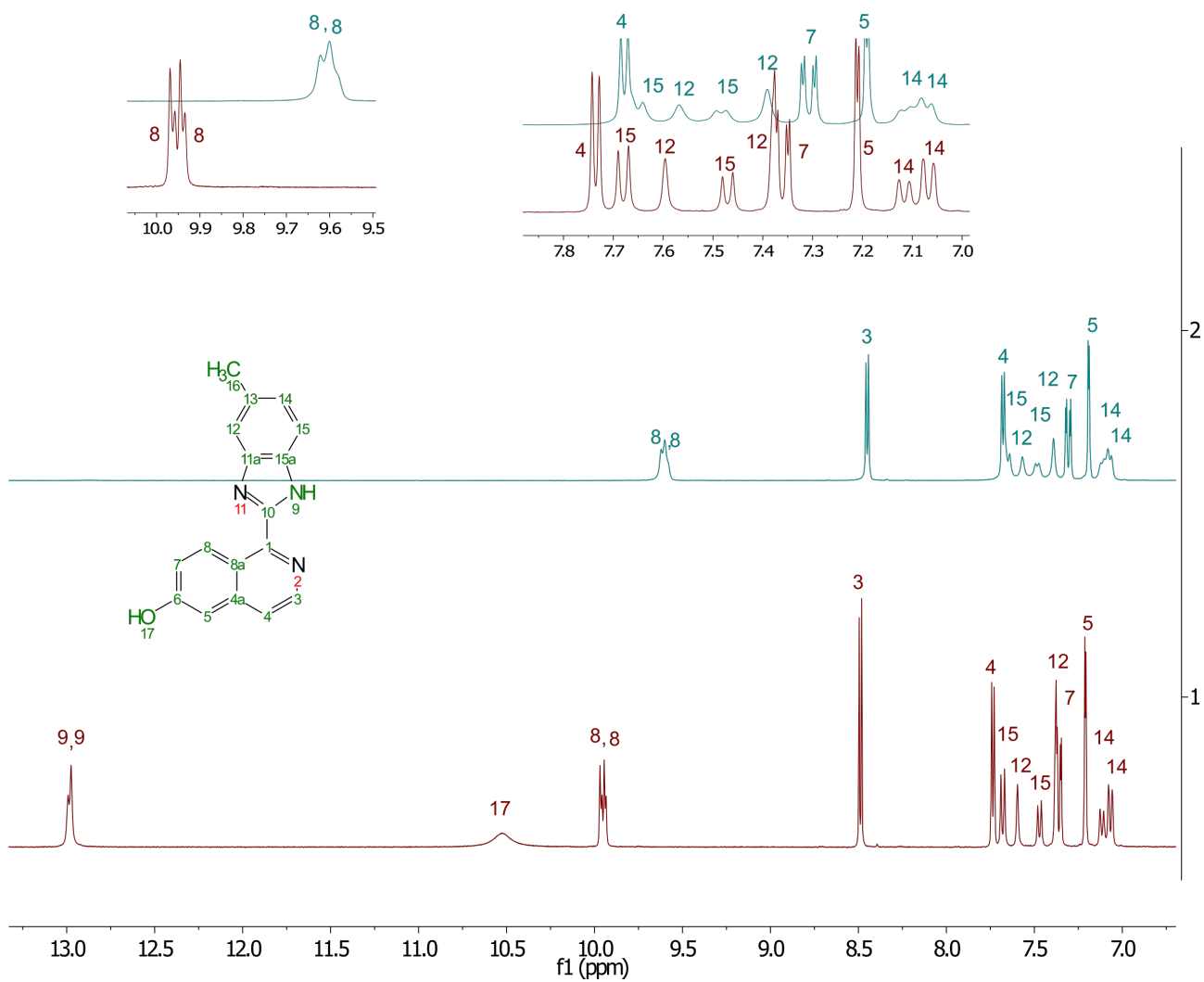


Figure S11: ¹H NMR spectrum of **18** in DMSO-*d*₆ (0.7 mL) at 25 °C (bottom), and after addition of 0.1 mL D₂O (Top).

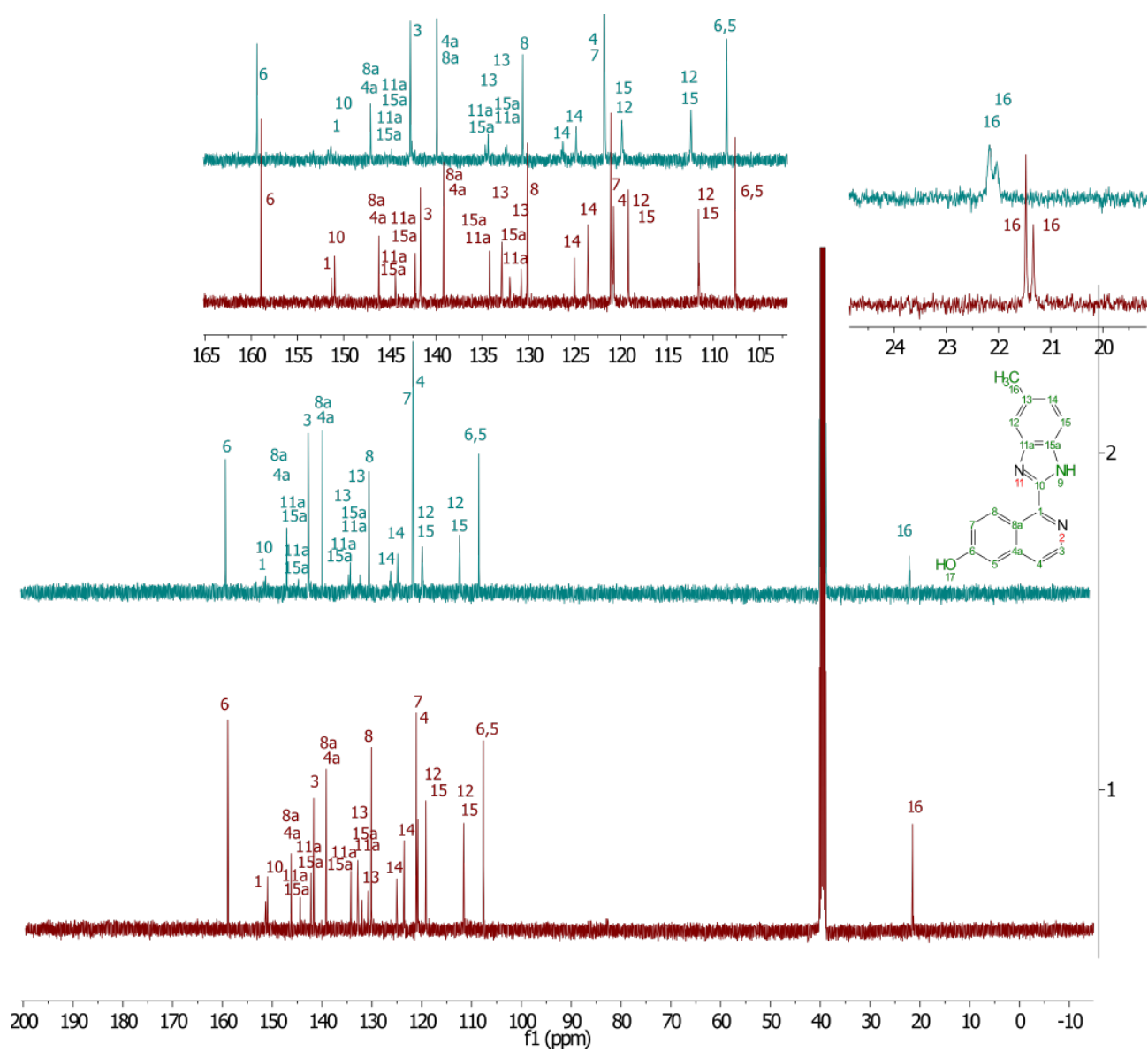


Figure S12: ^{13}C NMR spectrum of **18** in $\text{DMSO}-d_6$ (0.7 mL) at 25 °C (bottom), ^{13}C NMR spectrum after addition of 0.1 mL D_2O (Top).

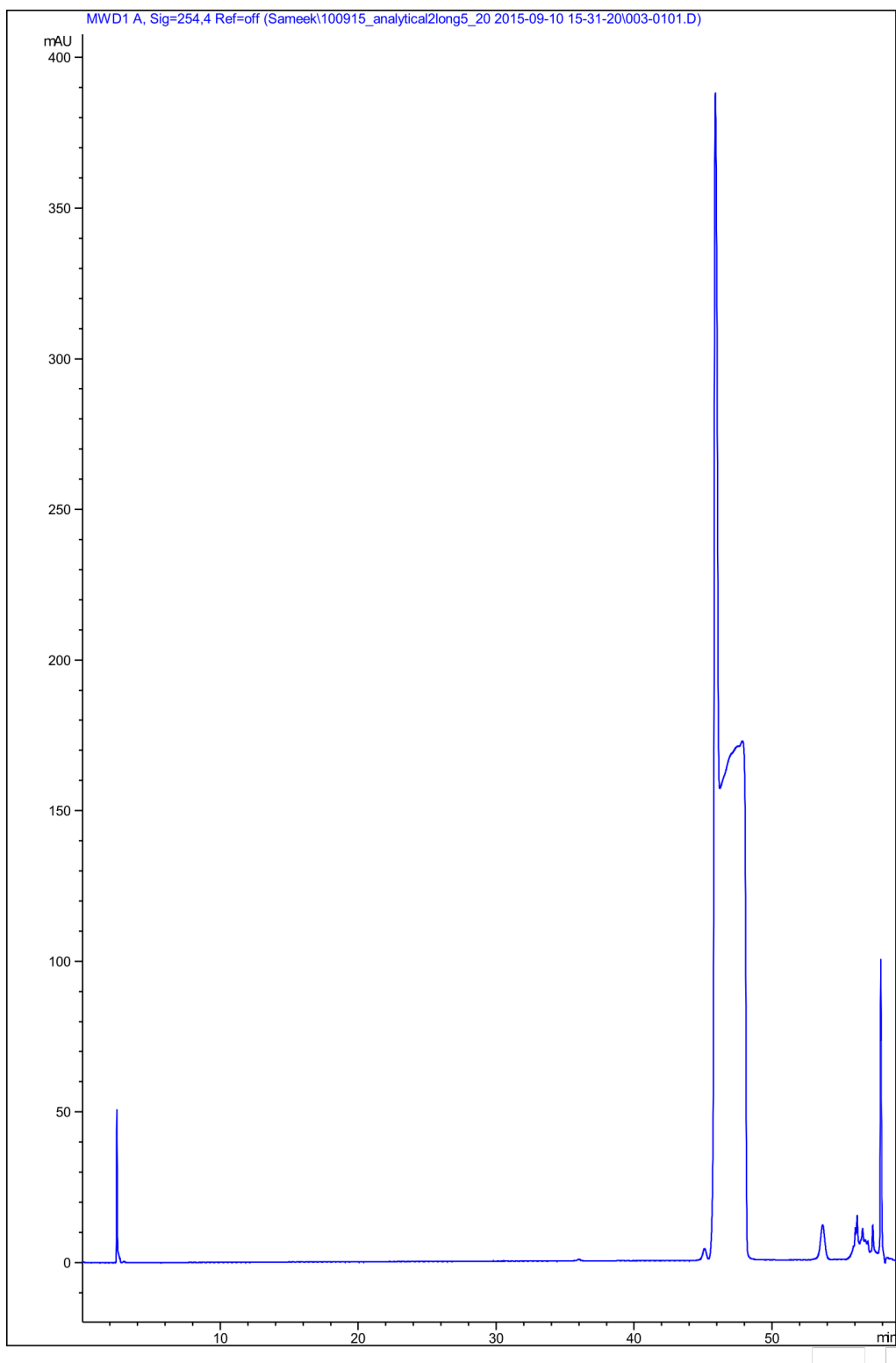


Figure S13: Analytical HPLC chromatogram of **18** at high concentration showed a shoulder peak.

The following Analytical RP-HPLC method was used for obtaining analytical HPLC chromatograms reported in Figure S13, Figure S15(A) and (B) - 5% solvent B 1 min, gradient of 5-20% solvent B 1-51 mins, 95% solvent B 52-53 mins, gradient of 95-5% solvent B 53-55 mins, 5% solvent B 55-59 mins.

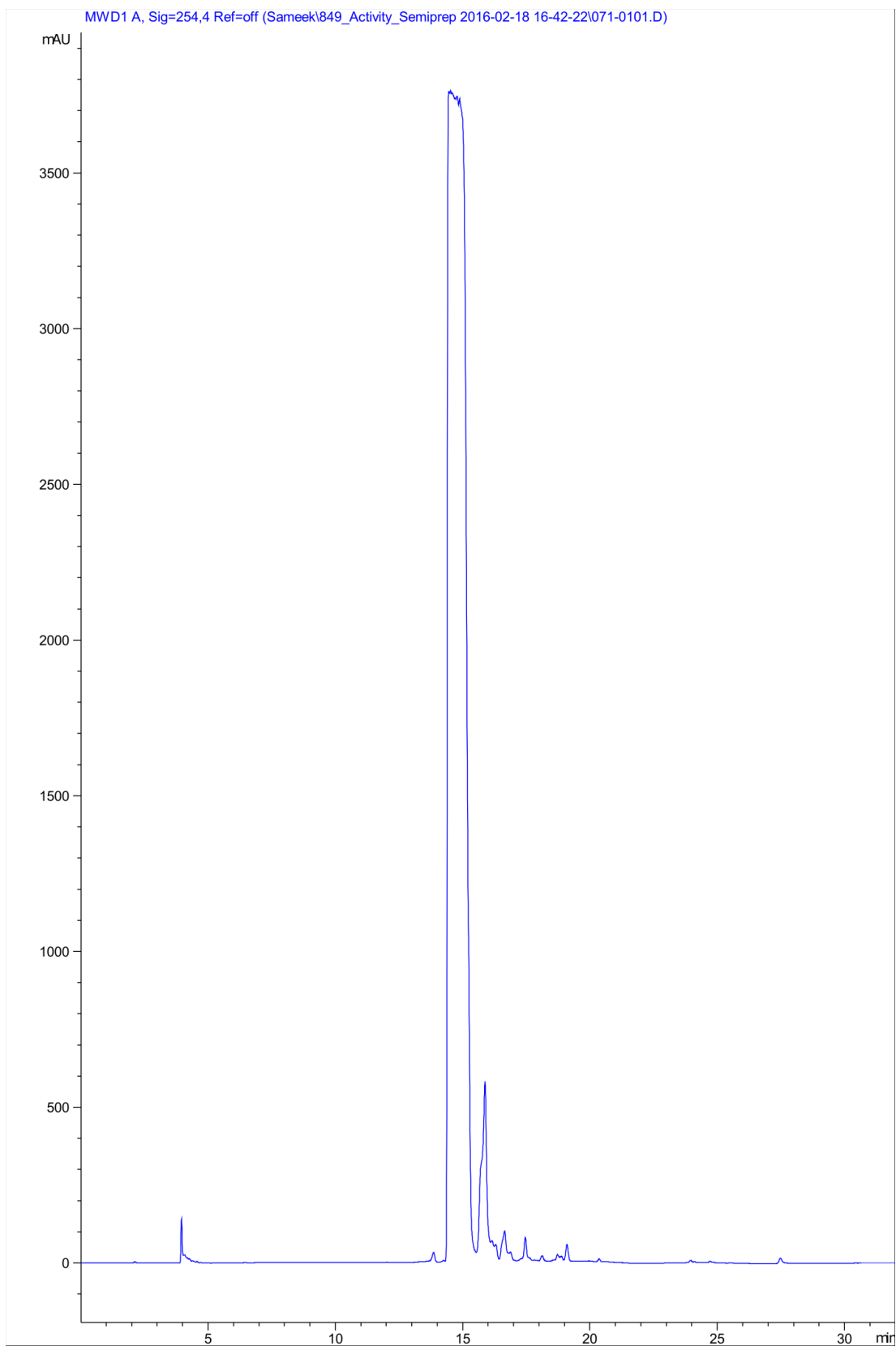


Figure S14: Semi-preparative HPLC chromatogram of **18**.

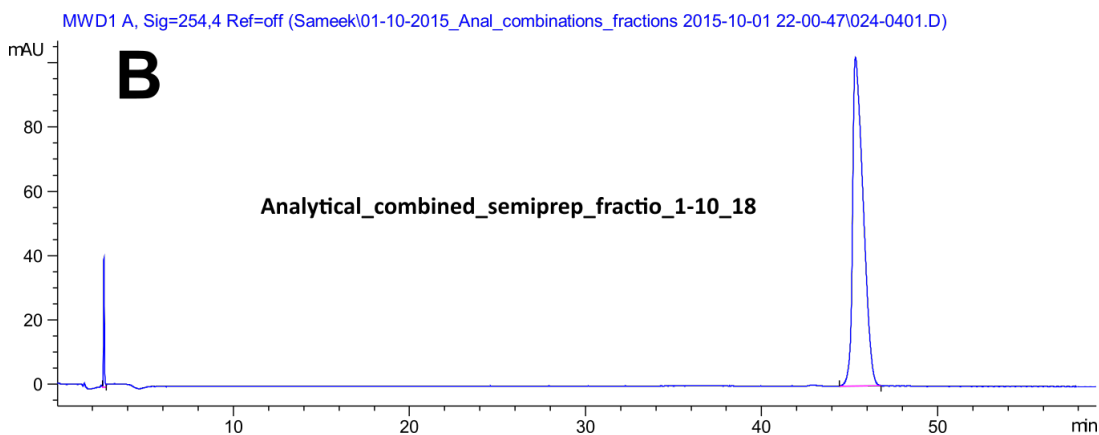
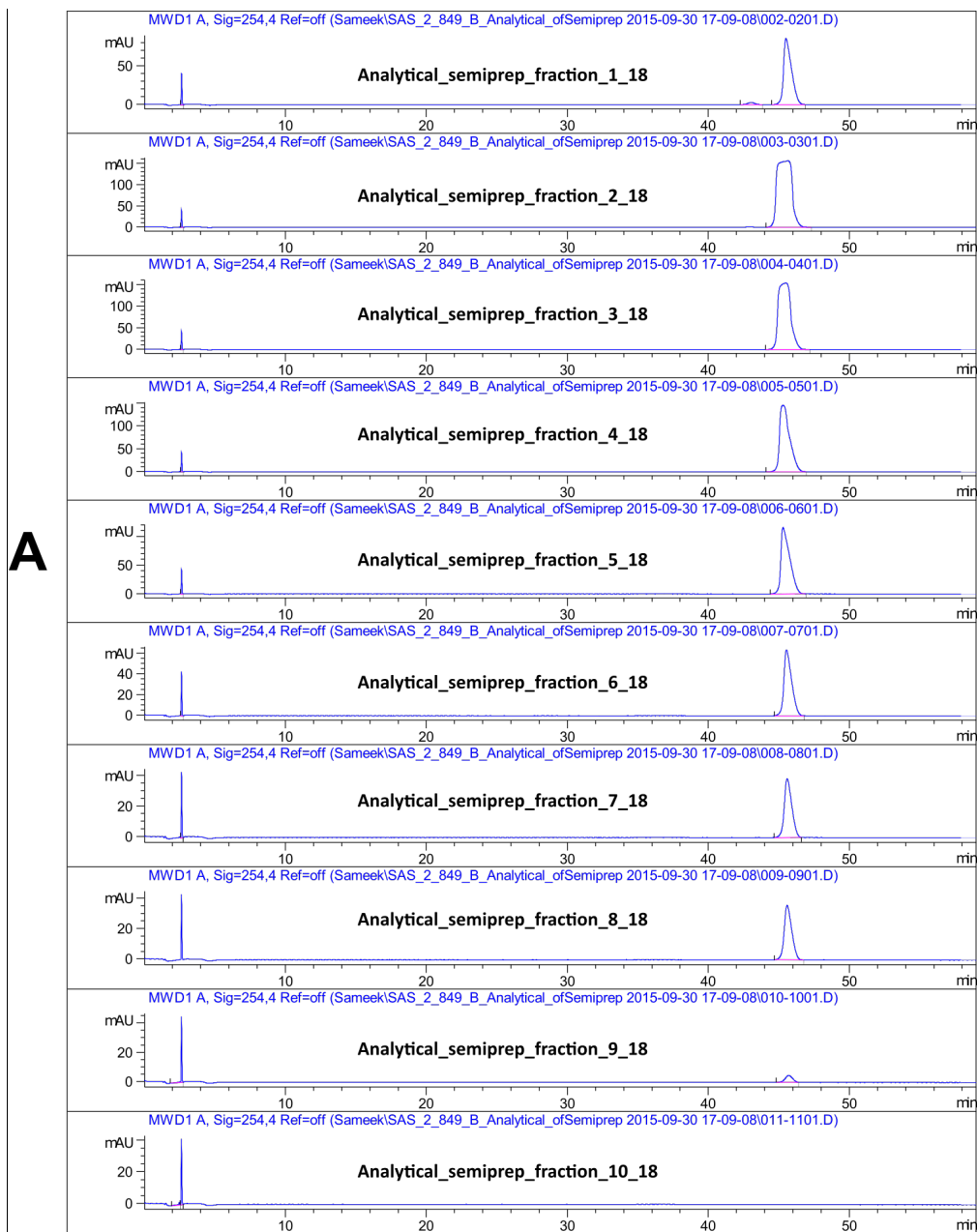


Figure S15: (A) Analytical HPLC chromatogram of 10 separate fractions collected from peak corresponding to **18** (14.5 to 15.5 min) in semi-preparative RP-HPLC chromatogram (Figure S14). **(B)** Analytical HPLC chromatogram of combined fractions.

3. Pharmacology

3.1 Material and Methods

NLuc-A₁AR and NLuc-A₃AR receptor constructs, stable cell line generation and cell culture were carried out as described by Stoddart *et al.*²¹ Briefly, the full length NLuc luciferase sequence (pNL1.1 vector; Promega Corporation) was amplified and fused in frame within pcDNA3.1 vector along with a short membrane signal sequence derived from the 5HT_{3A} receptor. Full length human adenosine A₁ or A₃ receptor subtypes (initiation methionine removed) were then fused to the 3' end of signal sequence-NLuc in pcDNA3.1. The resulting constructs were termed NLuc A₁AR and NLuc A₃AR respectively. Untagged A₁AR was obtained from cDNA.org.

HEK293 cells (NLuc A₁AR; American Type Culture Collection (ATCC)) or HEK293G cells (Glosensor™ cAMP HEK293 for NLuc A₃AR; Promega Corporation) were grown in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal calf serum at 37°C, 5% CO₂. Mixed population cell lines were generated following transfection with Lipofectamine (Life Technologies) and geneticin (G418; Thermo-Fisher Scientific) selection pressure. The NLuc A₃AR cell line was then dilution cloned to generate cell lines derived from a single cell

Bioluminescence resonance energy transfer (BRET) assays

The BRET A₁AR and A₃AR receptor-ligand binding assays were undertaken following the protocol described in Stoddart *et al.*²¹ In brief, saturation assays were performed on stably transfected cells that were seeded 24 h before experimentation in poly-D-lysine coated (2mg/ml; Sigma Aldrich) white Thermo Scientific Matrix 96-well microplates. The medium was removed from each well and replaced with HEPES Buffered Saline Solution (HBSS: 145 mmol/L NaCl, 5 mmol/L KCl, 1.7 mmol/L CaCl₂, 1 mmol/L MgSO₄, 10 mmol/L HEPES, 2 mmol/L sodium pyruvate, 1.5 mmol/L NaHCO₃, 10 mmol/L D-glucose, pH 7.2–7.45) with the required concentration of fluorescent ligand and competing ligand if present (1h at 37 °C). Non-specific binding was assessed in the absence and presence of 1μM DPCPX (NLuc-A₁AR) or 1μM MRS1220 (NLuc-A₃AR). For competition experiments, NLuc A₁AR cells were incubated with 25mM CA200645 in the presence or absence of increasing concentrations of unlabelled ligand (1h at 37 °C). For both saturation and competition assays, following ligand stimulation the NLuc substrate furimazine (Promega Corporation, USA) was added to give a final concentration of 10μM. Plates were left for 5min and then luminescence and fluorescence emissions were measured using a PHERAstar FS plate reader (BMG Labtech) at room temperature. Filtered light emissions were sequentially measured at 460 nm (80-nm bandpass; NLuc emission) and >610 nm (longpass; fluorescence emission) and the raw BRET ratio calculated by dividing the >610-nm emission by the 460-nm emission. Data was pooled from independent experiments (n=4-7) and for saturation experiments, data was baseline corrected (minus vehicle + furimazine BRET ratios) and expressed as fold increase in BRET ratios over basal. For saturation experiments, NLuc A₁AR data were fit using GraphPad Prism_v6 using a one site saturation binding fit (total and non specific binding) with shared

values for non specific binding. Specific binding for A₃AR was calculated by subtracting corrected BRET values obtained in the presence of competing unlabelled MRS1220 from those obtained for compound alone. Data were fit using Graph Prism_v6 using one site saturation binding. K_d values were estimated from each individual experiment, which were then averaged and expressed as mean ± standard error of the mean. For displacement experiments, data were pooled from 5 independent experiments. When possible, binding affinities (K_i) of unlabeled ligands were calculated using the Cheng-Prusoff equation, using K_d values derived from CA200645 saturation binding and are expressed as mean ± standard error of the mean.

High content fluorescence imaging

HEK293 cells were seeded 48 h before experimentation in poly-D-lysine coated (2mg/ml; Sigma Aldrich) white Thermo Scientific Matrix 96-well microplates and were transiently transfected with the A₁AR (untagged) construct 24 h before experimentation using Fugene (1:3 DNA:reagent ratio; 100ng cDNA/well). Additionally, HEK293 cells stably transfected with NanoLuc N-terminal, tagged A₁AR were seeded 24 h before experimentation in poly-D-lysine coated (2mg/ml; Sigma Aldrich) white Thermo Scientific Matrix 96-well microplates. On the day of experimentation, medium was removed from each well and replaced with HBSS. Cells were then incubated with increasing concentrations of the fluorescent ligand, CA200645 and incubated for 1hr at 37°C. Non-specific binding was defined using 1μM DPCPX. Plates were then washed with phosphate-buffered saline (PBS) and fixed using 3% paraformaldehyde in PBS. Plates were imaged at 4 sites per well using a IX Ultra Confocal Plate Reader (Molecular Devices, CA, USA) fitted with a 40x ELWD objective and using a long pass 650nm filter (60% laser power, 600 gain).²² Fluorescence intensity was quantified per well using a multi wavelength cell sorting algorithm (MetaXpress, Molecular Devices). Data were fit using Graph Prism_v6 using one site saturation binding. K_d values were estimated from each individual experiment (n=3/4), averaged and expressed as mean ± standard error of the mean.

3.2 Supplementary data

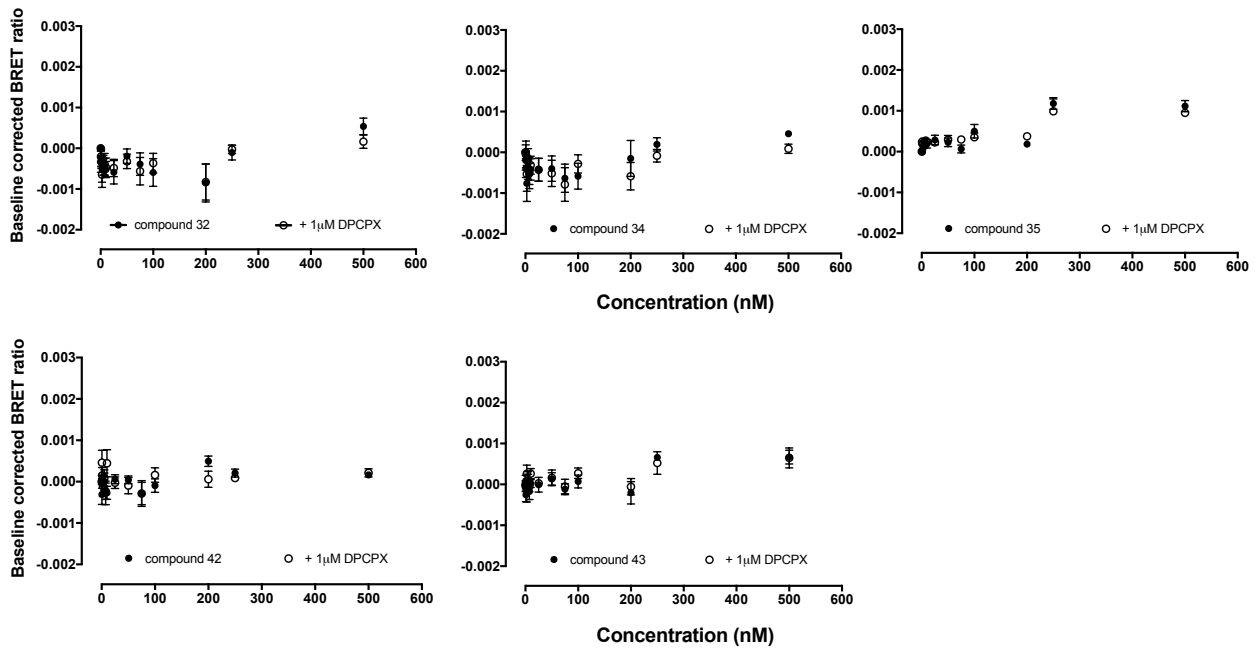


Figure S16. HEK293 cells stably transfected with *N*-terminally NLuc-labelled A₁AR were treated with increasing concentrations of fluorescent ligand and the BRET ratio measured after direct addition of the NLuc substrate furimazine (10 μM).²¹ Non-specific binding was assessed in the absence and presence of 1 μM DPCPX. Pooled raw BRET ratio's were baseline corrected (minus vehicle + furimazine BRET ratios) with data expressed as fold increase in BRET ratios over basal. Data represents five - seven independent experiments (in triplicate).

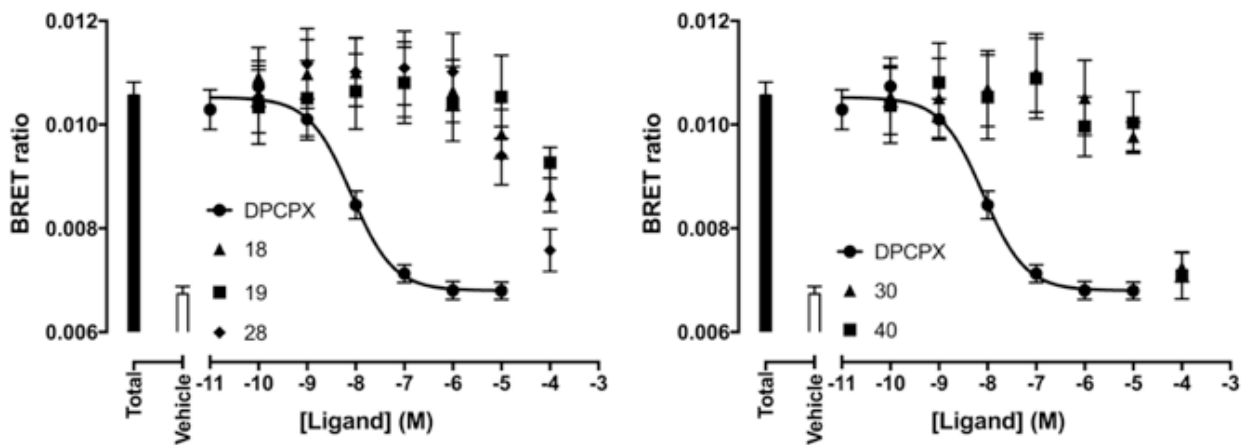


Figure S17. HEK293 cells stably expressing *N*-terminal NLuc tagged A1 were co-incubated with a fixed concentration of CA200645 (25nM) and increasing concentrations of unlabelled ligand (1hr at 37°C). The A₁AR selective antagonist DPCPX was included as a positive control. Total CA200645 binding and vehicle are shown by the black and white bars respectively. Data were pooled from five independent experiments and are expressed as mean ± S.E.M.

4. References

1. B. Cosimelli, S. Taliani, G. Greco, E. Novellino, A. Sala, E. Severi, F. Da Settimo, C. La Motta, I. Pugliesi, L. Antonioli, M. Fornai, R. Colucci, C. Blandizzi, S. Daniele, M. L. Trincavelli and C. Martini, *Chemmedchem*, 2011, **6**, 1909-1918.
2. J. J. Li, J. A. Tino, WO2001085695A1, 2001; Bristol-Myers Squibb Co.; Tetrahydroisoquinoline analogs useful as growth hormone secretagogues.
3. M. C. Maillard, F. A. Brookfield, S. M. Courtney, F. M. Eustache, M. J. Gemkow, R. K. Handel, L. C. Johnson, P. D. Johnson, M. A. Kerry, F. Krieger, M. Meniconi, I. Muñoz-Sanjuán, J. J. Palfrey, H. Park, S. Schaertl, M. G. Taylor, D. Weddell and C. Dominguez, *Biorg. Med. Chem.*, 2011, **19**, 5833-5851.
4. D. Ma, W. Wu, G. Yang, J. Li, J. Li and Q. Ye, *Bioorg. Med. Chem. Lett.*, 2004, **14**, 47-50.
5. Ishiguro, S.; Kawaguchi, N.; Nakakoshi, M.; Shimada, S.; Seya, M.; Nomoto, M.; Okue, M.; Tomizuka, H. JP07304744A, 1995; Snow Brand Milk Prod Co Ltd, Japan.; Preparation of isoquinoline derivatives as protease inhibitors.
6. P. L. Ornstein, M. B. Arnold, N. K. Augenstein and J. W. Paschal, *J. Org. Chem.*, 1991, **56**, 4388-4392.
7. A. M. M. Mjalli, D. Jones, D. R. Gohimmukkula, G. Huang, J. Zhu, M. Rao, R. C. Andrews, T. Ren, WO2006099379A2, 2006; Transtech Pharma, Inc.; Benzazole derivatives, compositions, and methods of use as b-secretase inhibitors.
8. Seiwert, S.; Beigelman, L.; Buckman, B.; Serebryany, V.; Stoycheva, A. D. WO2010045266A1, 2010; InterMune, Inc., USA.; Preparation of proline tripeptides and analogs as inhibitors of hepatitis C virus replication for treating hepatitis C infection and liver fibrosis.
9. Karanewsky, D. S.; Fotsing, J.; Tachdjian, C.; Arellano, M. WO2011106114A1, 2011; Senomyx, Inc., USA.; Identification of human T2R receptors that respond to bitter compounds that elicit the bitter taste in compositions, and the use thereof in assays to identify compounds that inhibit (block) bitter taste in compositions and use thereof.
10. E. F. Calderwood, M. Duffey, A. E. Gould, P. D. Greenspan, B. Kulkarni, M. J. Lamarche, R. S. Rowland, M. Tregay, T. J. Vos, WO2007067444A1, 2007; Millennium Pharmaceuticals, Inc.; Bicyclic compounds with kinase inhibitory activity.
11. A. Abeywardane, S. R. Brunette, M. J. Burke, T. M. Kirrane Jr, C. C. Man, D. R. Marshall, A. K. Padyana, H. Razavi, R. Sibley, K. L. L. Smith, R. J. Snow, R. J. Sorcek, H. Takahashi, S. J. Taylor, M. R. Turner, E. R. R. Young, Q. Zhang, Y. Zhang, R. M. Zindell, WO2014014874A1, 2014; Boehringer Ingelheim International GmbH; Pyrazole derivatives which inhibit leukotriene production.
12. S. Höck, R. Marti, R. Riedl and M. Simeunovic, *CHIMIA International Journal for Chemistry*, 2010, **64**, 200-202.

13. H. Y. Song, M. H. Ngai, Z. Y. Song, P. A. MacAry, J. Hobley and M. J. Lear, *Org. Biomol. Chem.*, 2009, **7**, 3400-3406.
14. M. A. Cinelli, B. Cordero, T. S. Dexheimer, Y. Pommier and M. Cushman, *Bioorg. Med. Chem.*, 2009, **17**, 7145-7155.
15. V. Sridharan, S. Saravanan, S. Muthusubramanian and S. Sivasubramanian, *Magn. Reson. Chem.*, 2005, **43**, 551-556.
16. H. Saito, Y. Tanaka and S. Nagata, *J. Am. Chem. Soc.*, 1973, **95**, 324-328.
17. I. S. H. Lee, E. H. Jeoung and C. K. Lee, *J. Heterocycl. Chem.*, 1996, **33**, 1711-1716.
18. R. M. Claramunt, C. Lopez, I. Alkorta, J. Elguero, R. Yang and S. Schulman, *Magn. Reson. Chem.*, 2004, **42**, 712-714.
19. R. Benassi, Lazzeret.P, Schenett.L, F. Taddei and Vivarell.P, *Tetrahedron Lett.*, 1971, 3299-3300.
20. E. P. Papadopoulos and U. Hollstein, *Organic Magnetic Resonance*, 1982, **19**, 188-191.
21. L. A. Stoddart, E. K. M. Johnstone, A. J. Wheal, J. Goulding, M. B. Robers, T. Machleidt, K. V. Wood, S. J. Hill and K. D. G. Pflieger, *Nat Meth*, 2015, **12**, 661-663.
22. M. A. Arruda, L. A. Stoddart, K. Gherbi, S. J. Briddon, B. Kellam and S. J. Hill, *Front. Pharmacol.*, 2017, **8**.