

Supporting Information

Eco-friendly reactions in PEG-400: highly efficient and green approach for stereoselective access to multisubstituted 3,4-dihydro-2(1*H*)-quinazolines using 2-aminobenzylamines

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X-ray Crystallographic Studies

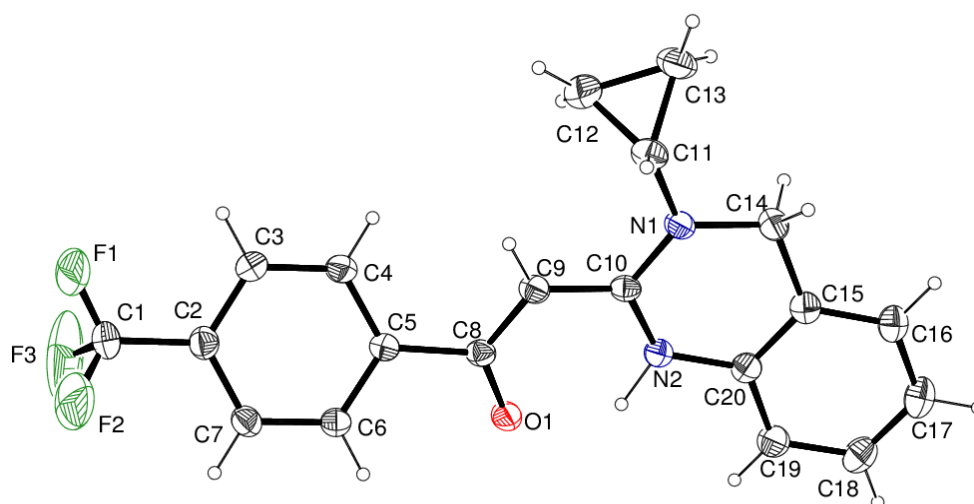


Figure 1. ORTEP Structure of compound 4o

The intensity data for 4o was collected on an Oxford CCD diffractometer with an Xcalibur sapphire diffraction measurement Mo-K α radiation ($\lambda = 0.71073 \text{ \AA}$) at 293(2) K¹. A multi-scan correction was applied. The structure was solved by the direct methods using SIR-92 and refined by full-matrix least-squares refinement techniques on F^2 using SHELXL97². The hydrogen atoms were placed into the calculated positions and included in the last cycles of the refinement. All calculations were done using Wingx software package³.

Table1. Crystal data collection and structure refinement parameters for **4o**

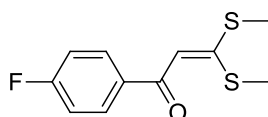
Empirical formula	C ₂₀ H ₁₇ F ₃ N ₂ O
Formula weight	358.36
Temperature	293(2) K
Wavelength	0.71073 Å
Crystal system	triclinic
Space group	<i>P</i> -1
<i>a</i>	7.2030(7) Å
<i>b</i>	10.0784(8) Å
<i>c</i>	13.335(2) Å
α	108.279(9)°
β	92.593(10)°
γ	107.656(8)°
Volume	865.30(17) Å ³
<i>Z</i>	2
Density (calculated)	1.375 Mg/m ³
Absorption coefficient	0.108 mm ⁻¹
<i>F</i> (000)	372
Crystal size	0.22 x 0.19 x 0.18 mm ³
Theta range for data collection	3.10 to 25.00°
Index ranges	-8 ≤ <i>h</i> ≤ 6, -11 ≤ <i>k</i> ≤ 11, -15 ≤ <i>l</i> ≤ 15
Reflections collected	5952
Independent reflections	3048 [<i>R</i> (int) = 0.0143]
Completeness to theta = 25.00°	99.9 %
Absorption correction	Multi-scan
Max. and min. transmission	0.9808 and 0.9767
Refinement method	Full-matrix least-squares on <i>F</i> ²
Data / restraints / parameters	3048 / 0 / 236
Goodness-of-fit on <i>F</i> ²	1.051
Final <i>R</i> indices [<i>I</i> >2sigma(<i>I</i>)] ^{a, b}	<i>R</i> ₁ = 0.0852, <i>wR</i> ₂ = 0.2504
<i>R</i> indices (all data)	<i>R</i> ₁ = 0.1057, <i>wR</i> ₂ = 0.2691
Extinction coefficient	0.034(9)
Largest diff. peak and hole	0.826 and -0.576 e.Å ⁻³

$$^aR = \sum(|F_o| - |F_c|) / \sum |F_o| ; ^b wR = \{ \sum [w(F_o^2 - F_c^2)] / \sum [w(F_o^2)] \}^{1/2}$$

General procedure for the synthesis of α -oxoketene dithioacetals (3a-c)

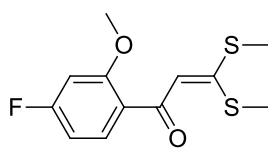
The procedure for the synthesis of α -oxoketene dithioacetals is previously reported.⁴ Compounds **2a-c** are previously reported.⁴ The structure and purity of known starting material was confirmed by comparison of physical and spectral data (¹H NMR, ¹³C NMR) with those reported in literature.

Analytical data of α -oxoketene dithioacetals



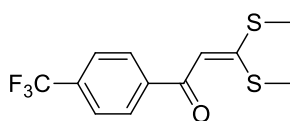
1-(4-Fluoro-phenyl)-3,3-bis-methylsulfanyl-propenone (3a) The

compound was obtained as a yellow crystalline solid, mp 83-85°C: IR (ν_{\max} cm⁻¹) (CHCl₃): 2919, 1597, 1470; ¹H NMR (400 MHz, CDCl₃) δ : 7.92-7.88 (m, 2H), 7.08 (t, J = 8.79Hz, 2H), 6.68 (s, 1H, vinylic CH), 2.51 (s, 6H, 2 x SCH₃); ¹³C NMR (100 MHz, CDCl₃) δ : 184.05, 166.9, 166.1, 163.6, 135.3, 129.8, 115.3, 108.9 (vinylic CH), 17.3 (-SCH₃), 15.0 (-SCH₃); HRMS (ESI) (M+H)⁺ Calcd for C₁₁H₁₁FOS₂: 243.0313, found 243.0306.



1-(4-Fluoro-2-methoxy-phenyl)-3,3-bis-methylsulfanyl-propenone (3b)

The compound was obtained as a pale yellow solid, mp 96-98°C: IR (ν_{\max} cm⁻¹) (CHCl₃): 2920, 1592, 1471; ¹H NMR (400 MHz, CDCl₃) δ : 7.77 (dd, J = 8.79, 7.32 Hz, 1H), 6.84 (s, 1H, vinylic CH), 6.68 (td, J = 8.79, 8.05 Hz, 2.2, 1H), 6.62 (dd, J = 10.98, 2.2 Hz, 1H), 3.8 (s, 3H, Ar-OCH₃), 2.48 (s, 3H, (-SCH₃), 2.47 (s, 3H, (-SCH₃)); ¹³C NMR (100 MHz, CDCl₃) δ : 184.3, 166.5, 164.0, 158.8, 132.6, 125.7, 114.4, 107.7 (vinylic CH), 99.4, 55.7 (Ar-OCH₃), 17.2(-SCH₃), 15.0(-SCH₃).



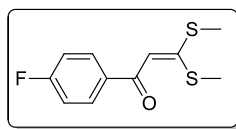
3,3-Bis-methylsulfanyl-1-(4-trifluoromethyl-phenyl)-propenone (3c)

The compound was obtained as a dark yellow solid, mp 84-86°C: IR (ν_{\max} cm⁻¹) (CHCl₃): 2992, 1480, 1323; ¹H NMR (400 MHz, CDCl₃) δ : 7.98 (d, J = 8.05 Hz, 2H), 7.67 (d, J = 8.05 Hz, 2H), 6.7 (s, 1H, vinylic CH), 2.56 (s, 3H, -SCH₃), 2.53 (s, 3H, -SCH₃); ¹³C NMR (100 MHz, CDCl₃) δ : 184.1, 168.8, 142.0, 127.9, 125.4, 108.6 (vinylic CH), 17.3(-SCH₃), 15.0(-SCH₃); HRMS (ESI) (M+H)⁺ Calcd for C₁₂H₁₁F₃OS₂: 293.0281, found 293.0273.

References:

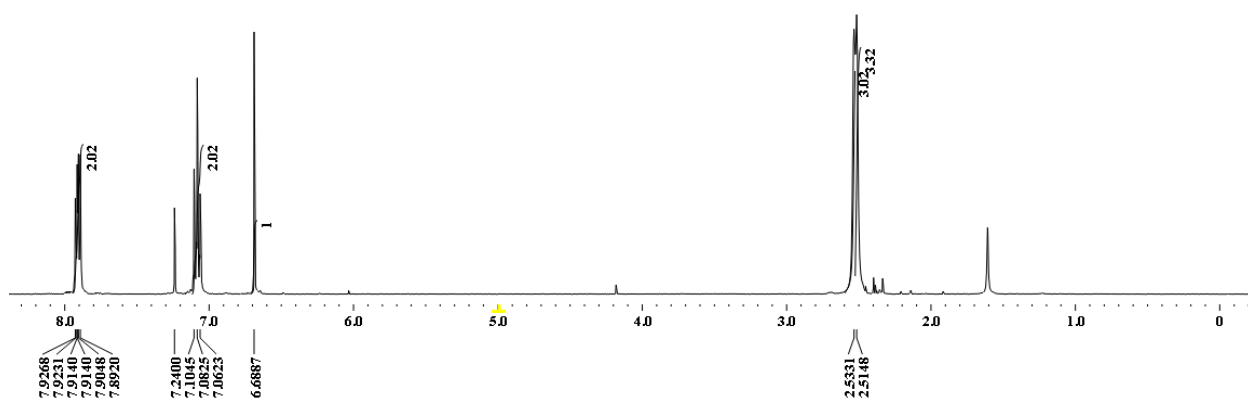
1. *CrysAlisPro*, v. 1.171.33.49b, Oxford Diffraction Ltd., Abingdon, UK, **2009**.
2. G. M. Sheldrick, *Acta Cryst.*, **2008**, A64, 112-122.
3. L. J. Farrugia, WinGX Version 1.80.05, *An integrated system of Windows Programs for the Solution, Refinement and Analysis of Single Crystal X-Ray Diffraction Data*; Department of Chemistry, University of Glasgow (**1997-2009**).
4. N. Sharma, T.S. Chundawat, S. C. Mahopatra, S. Bhagat, *RSC Advances*, **2013**, 3, 16336-16339.

COPIES OF ^1H NMR & ^{13}C NMR DATA

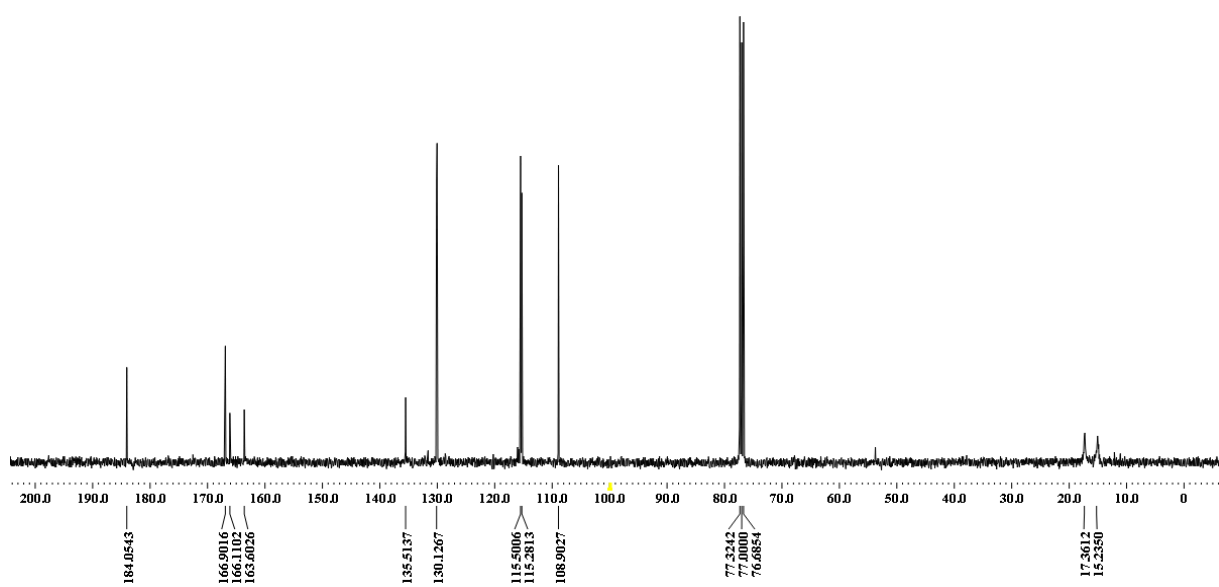


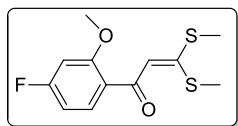
3a

¹H-NMR in CDCl₃ (400MHz)



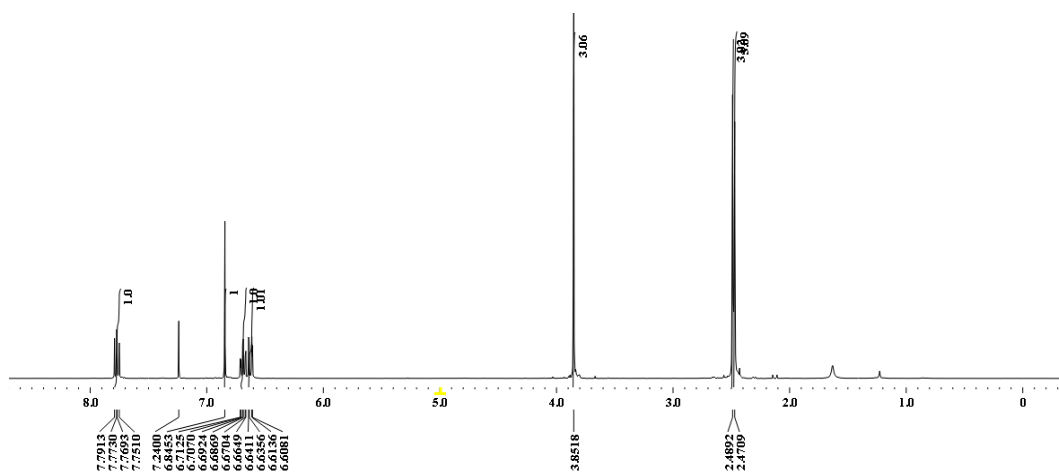
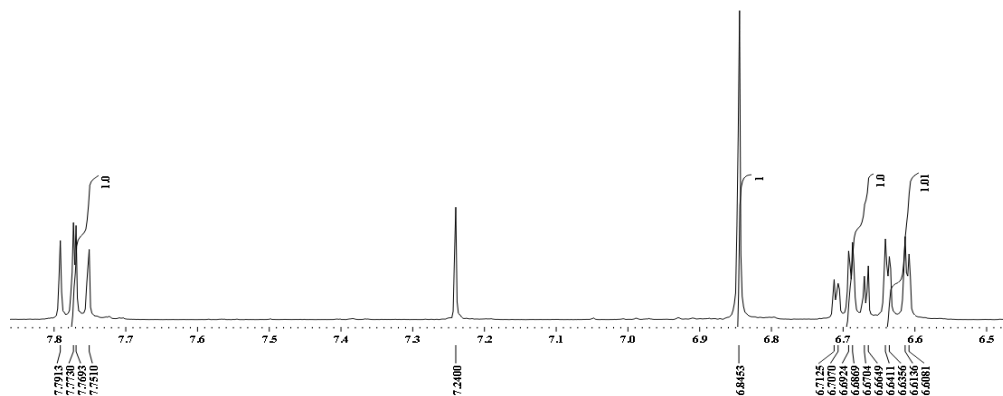
¹³C NMR in CDCl₃ (100MHz)



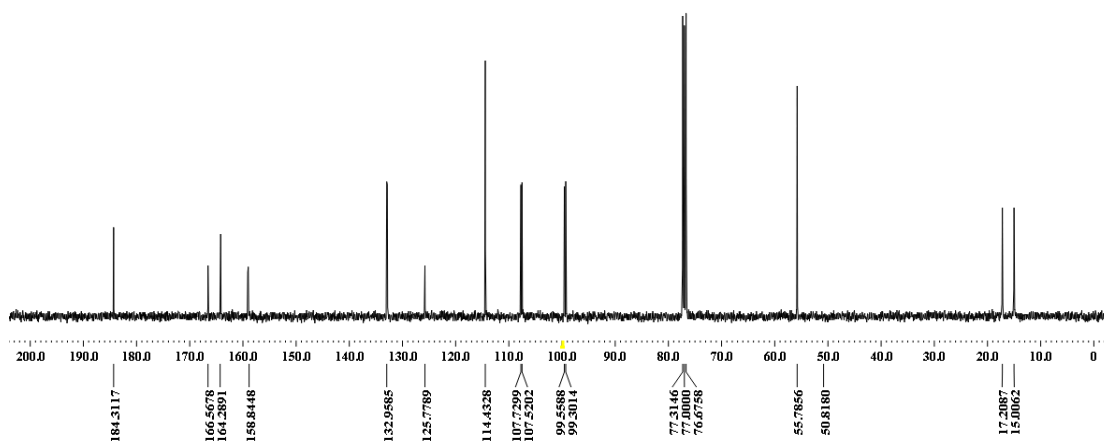


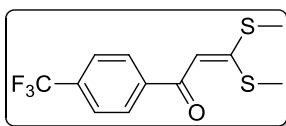
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¹H-NMR in CDCl₃ (400MHz)



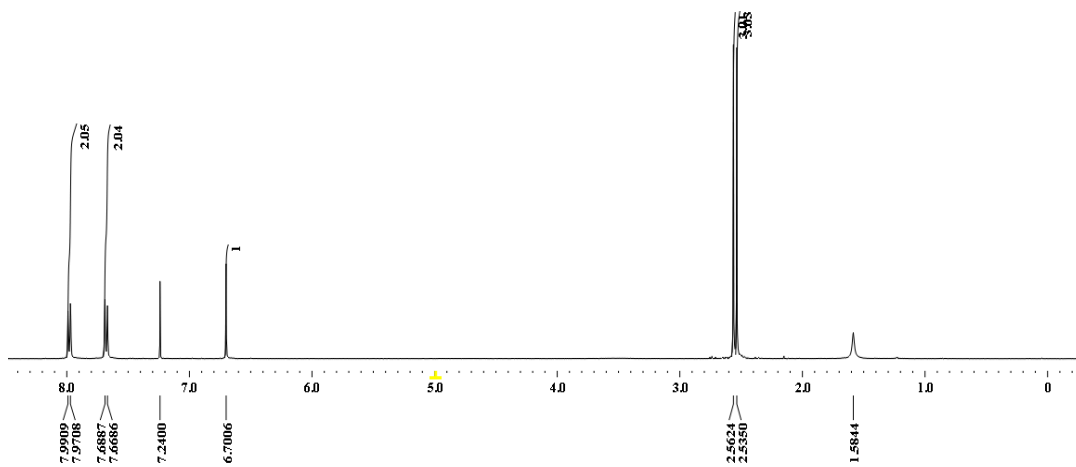
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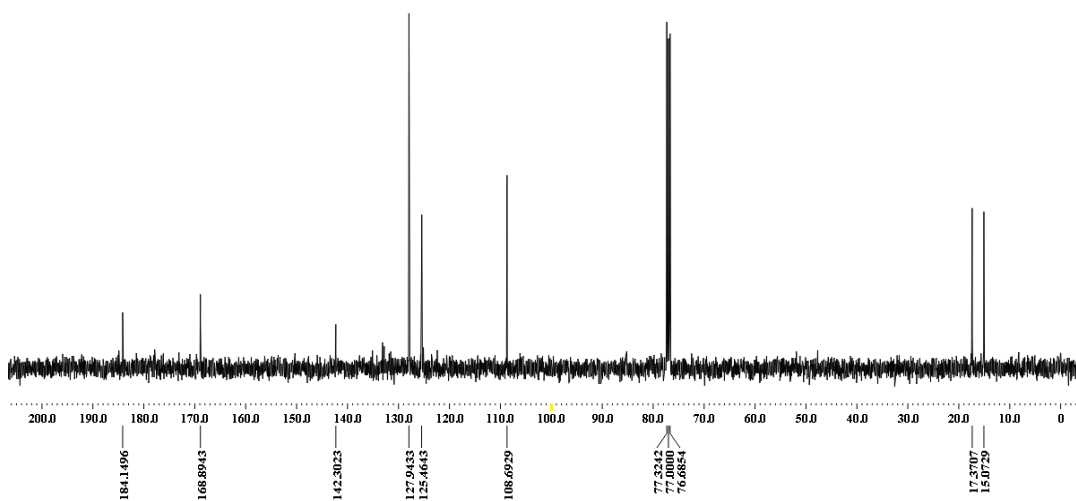


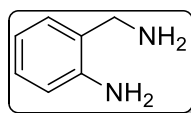
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¹H-NMR in CDCl₃ (400MHz)



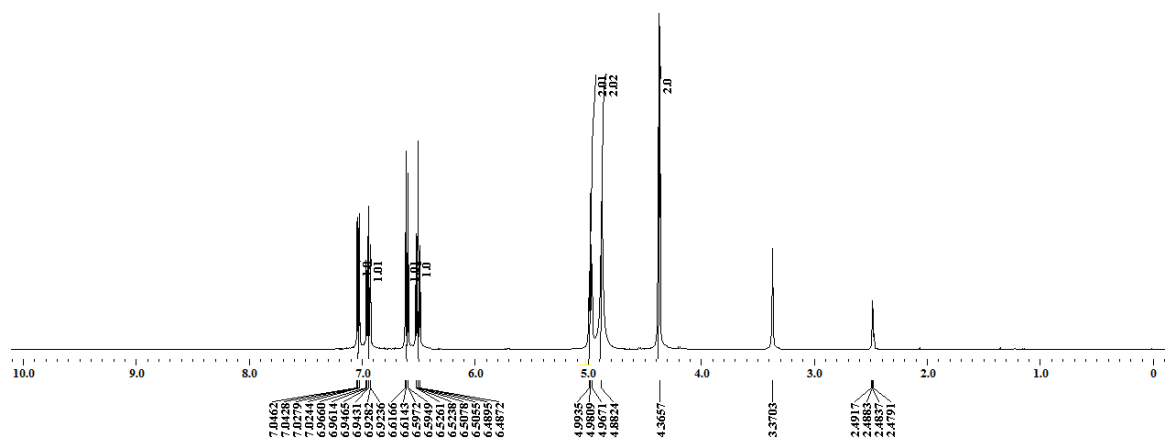
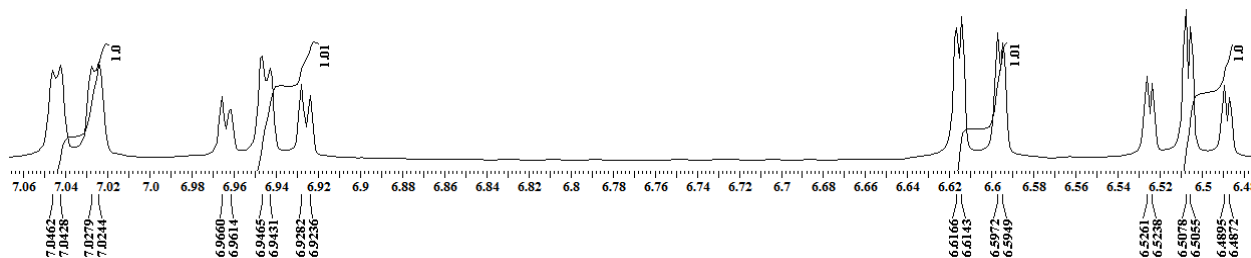
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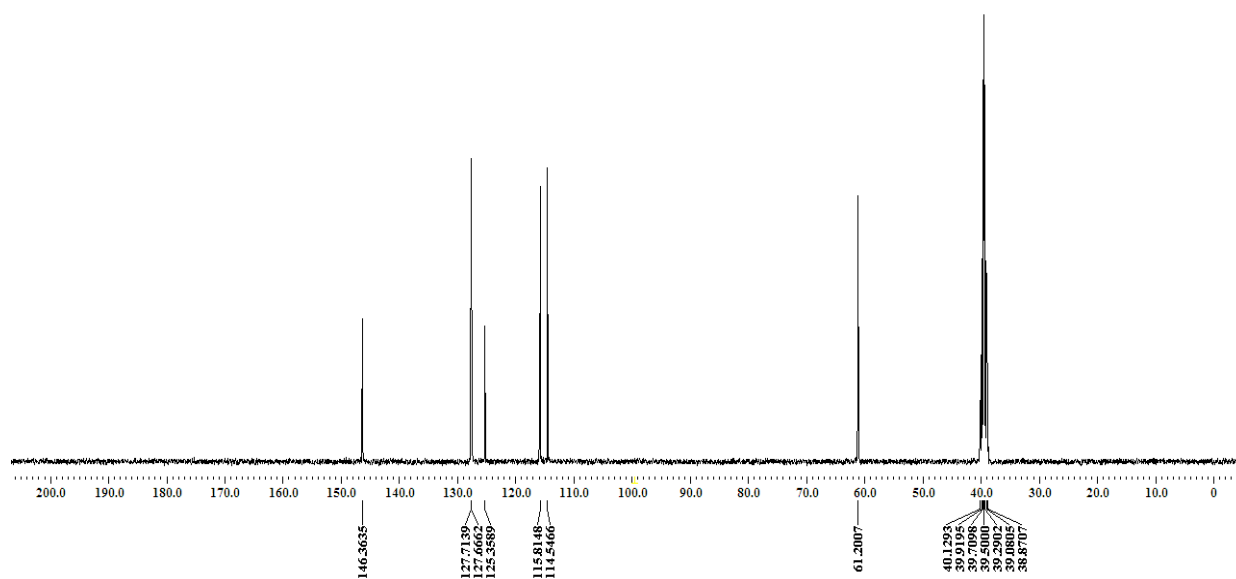


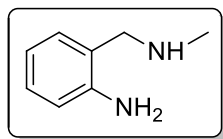
2a

^1H NMR in DMSO- d_6 (400Mz)



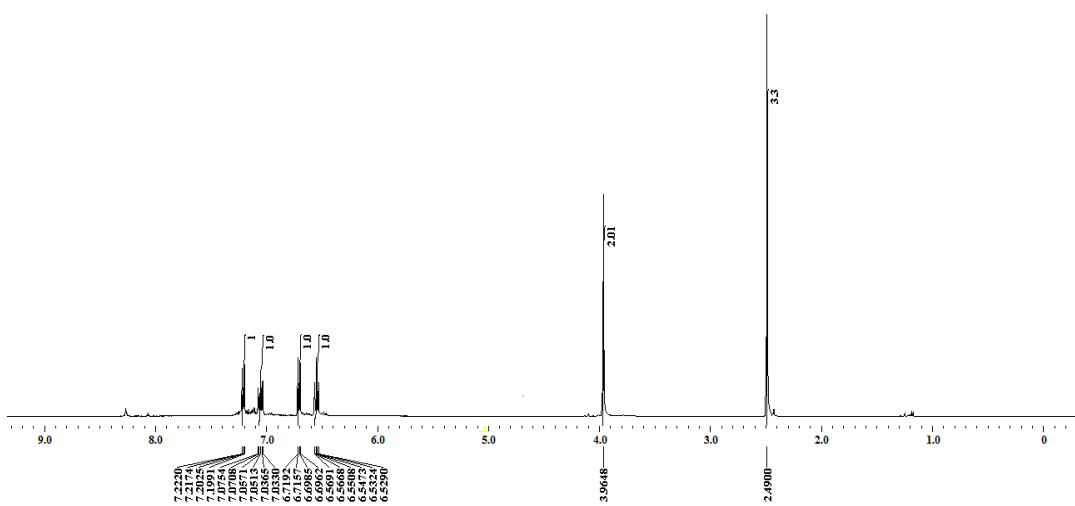
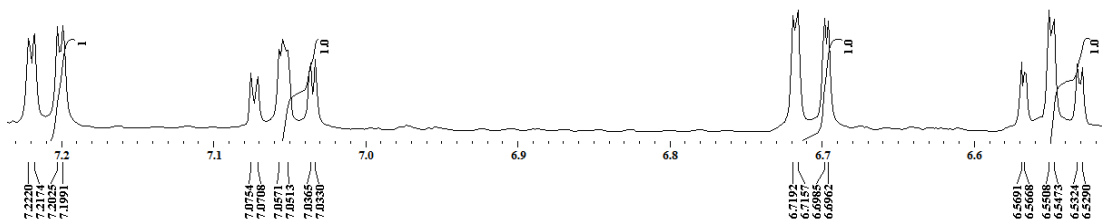
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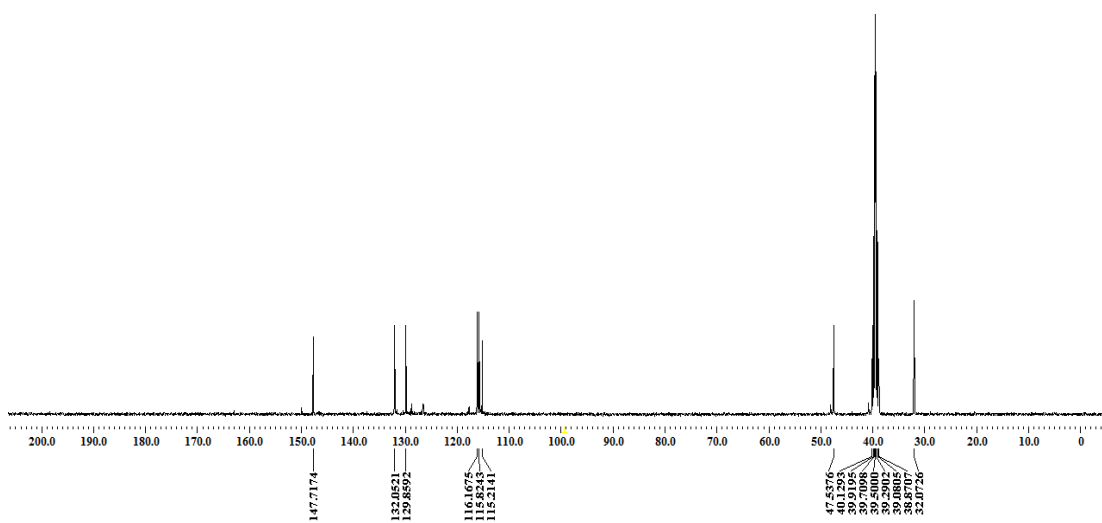


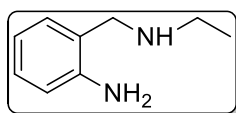
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¹H NMR in DMSO-d₆ (400Mz)



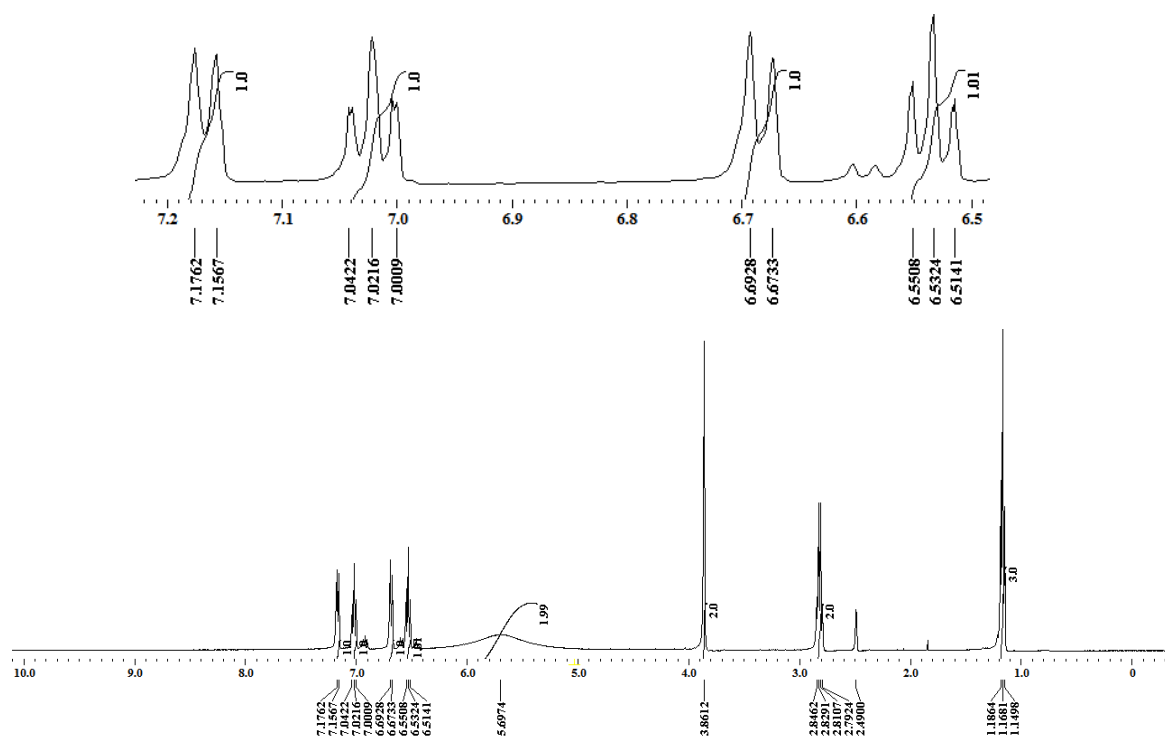
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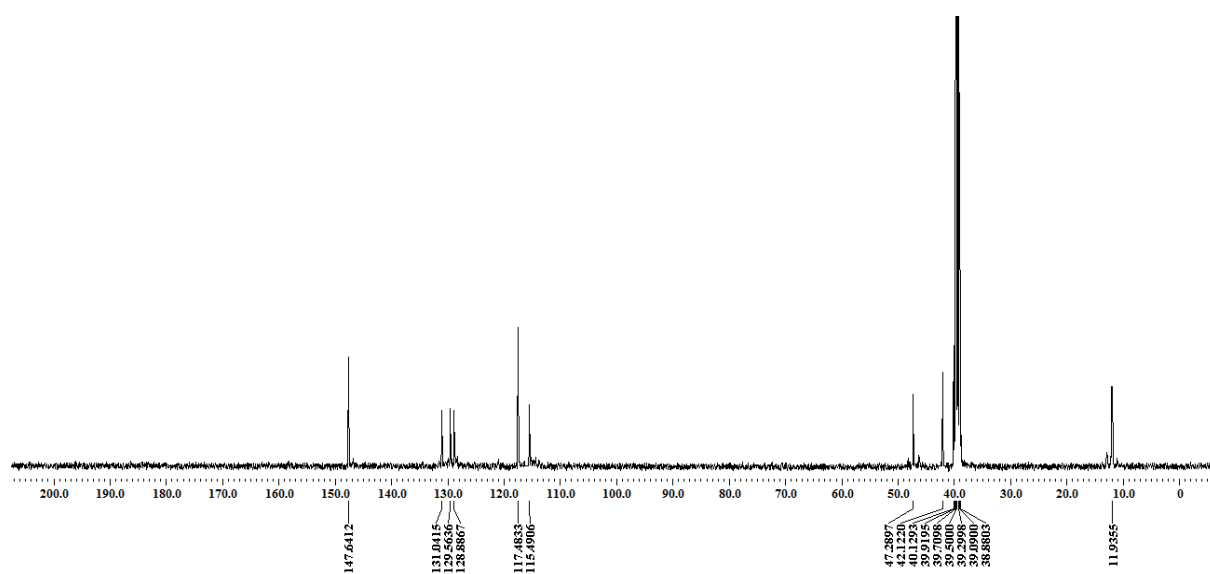


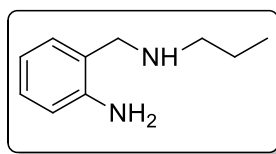
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¹H NMR in DMSO-d₆ (400Mz)



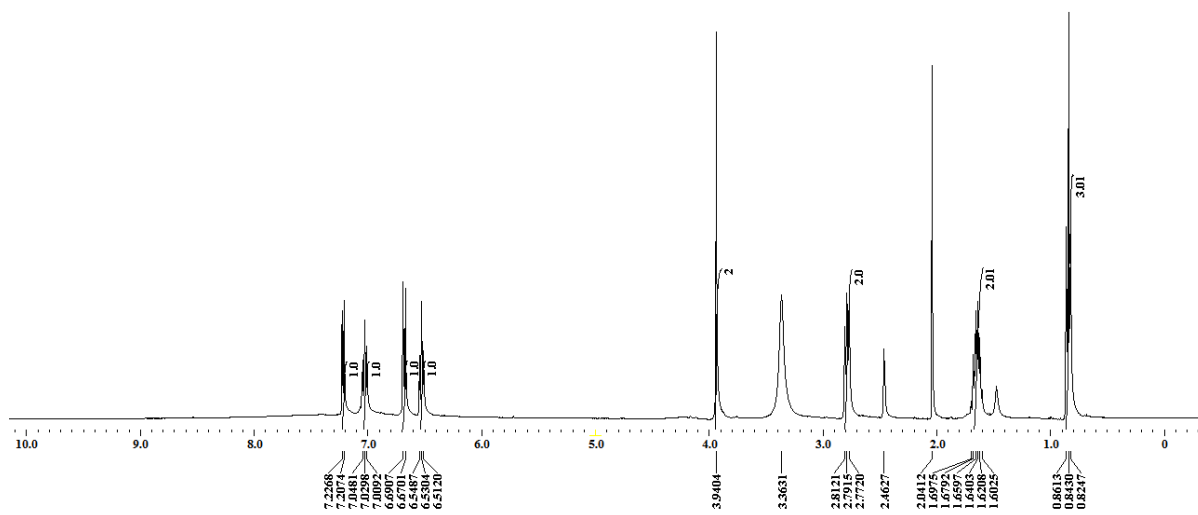
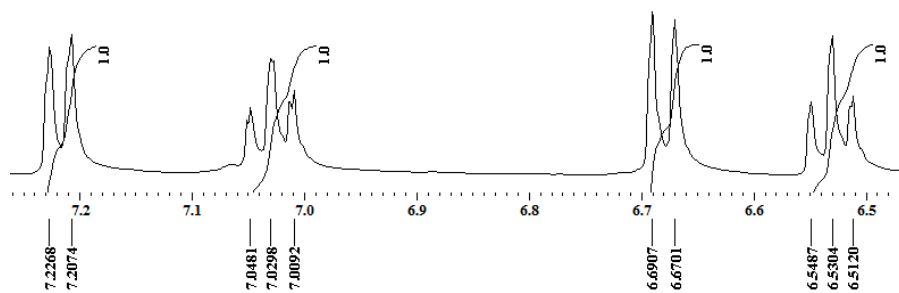
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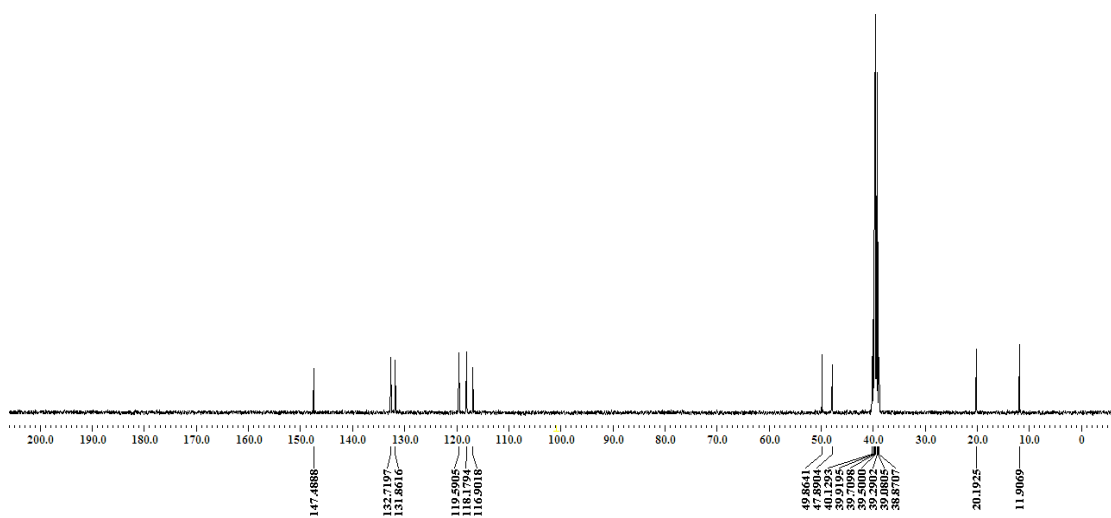


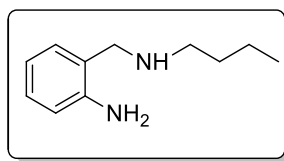
2d

¹H NMR in DMSO-d₆ (400Mz)



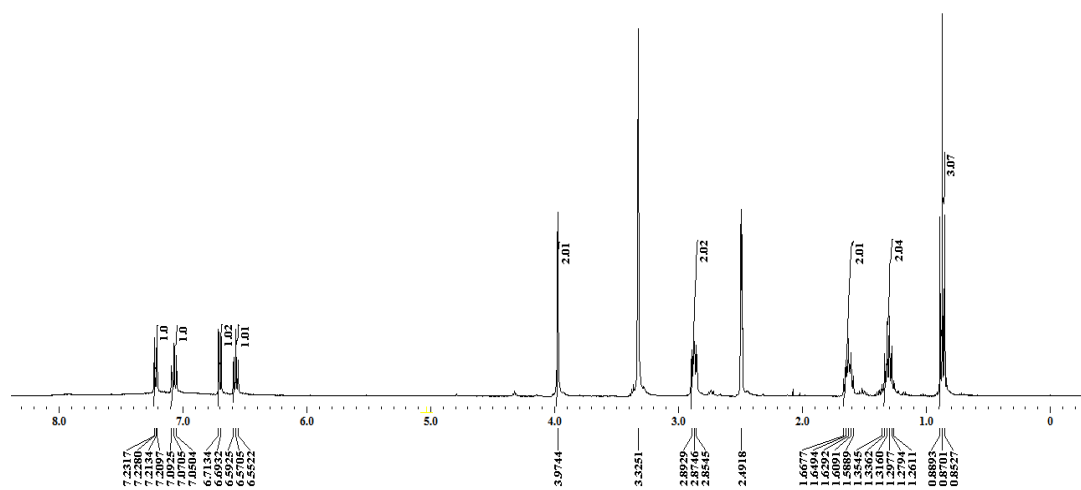
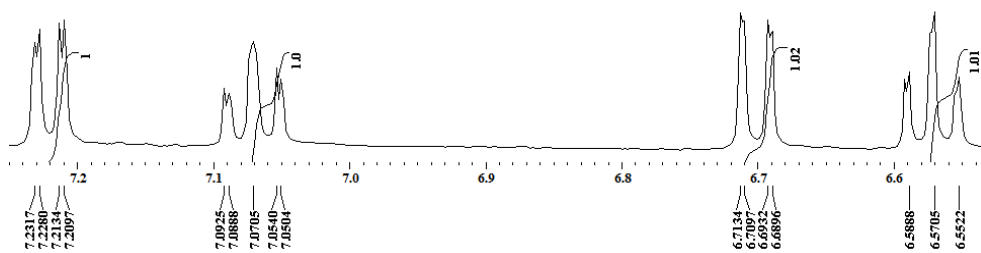
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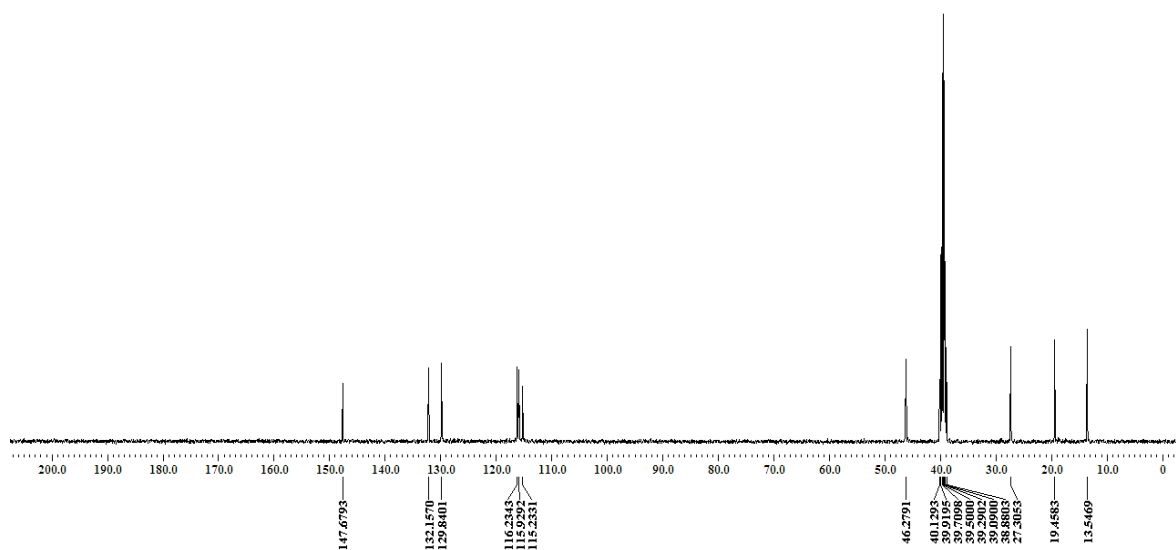


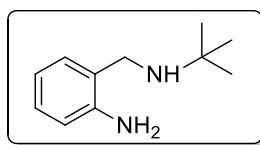
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^1H NMR in DMSO- d_6 (400Mz)



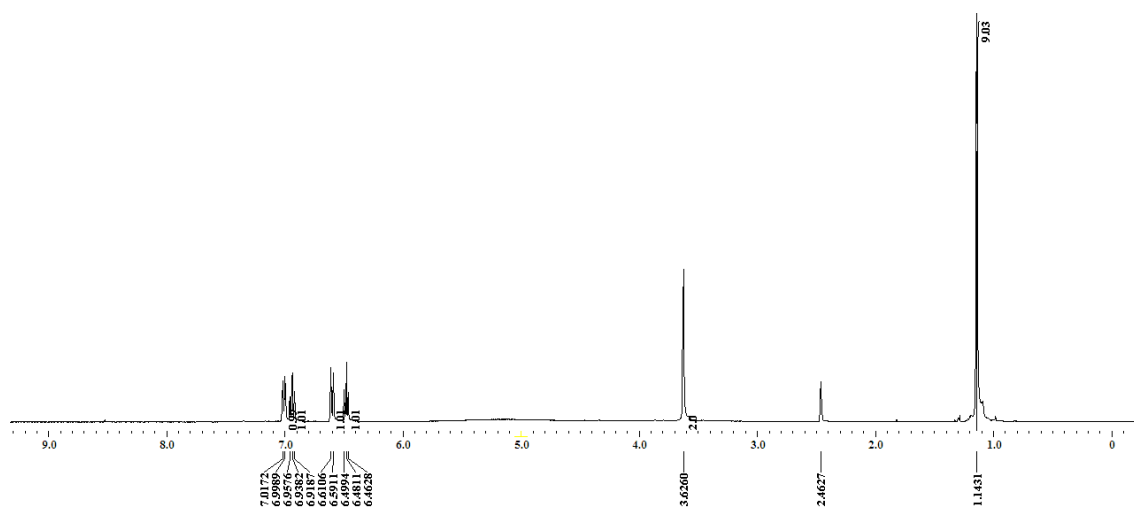
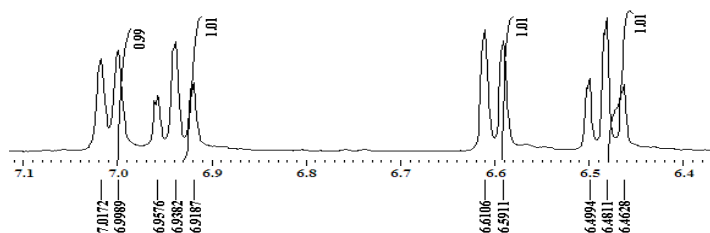
^{13}C NMR in DMSO- d_6 (100Mz)



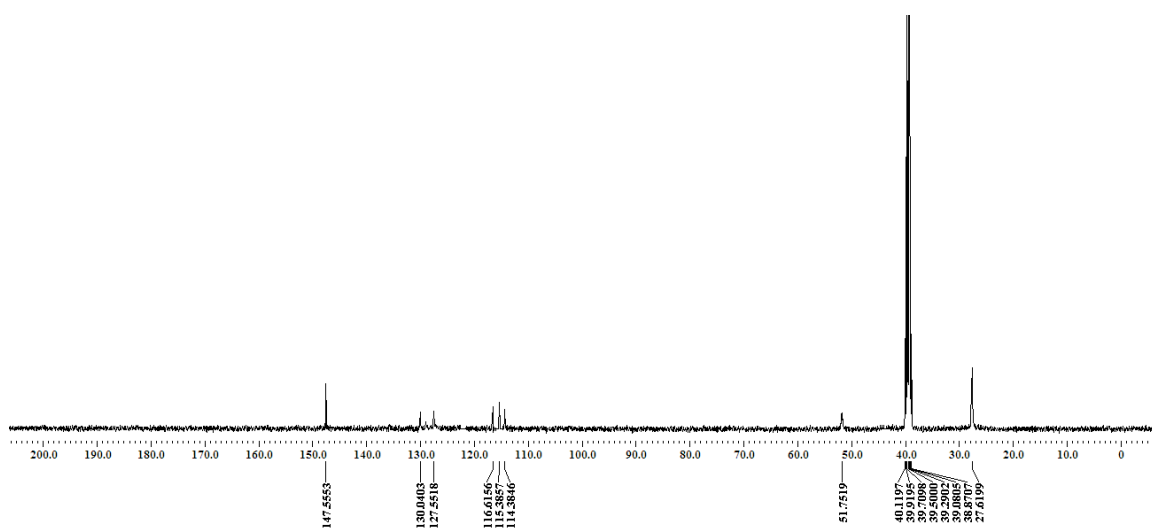


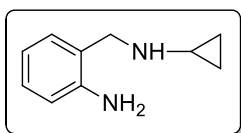
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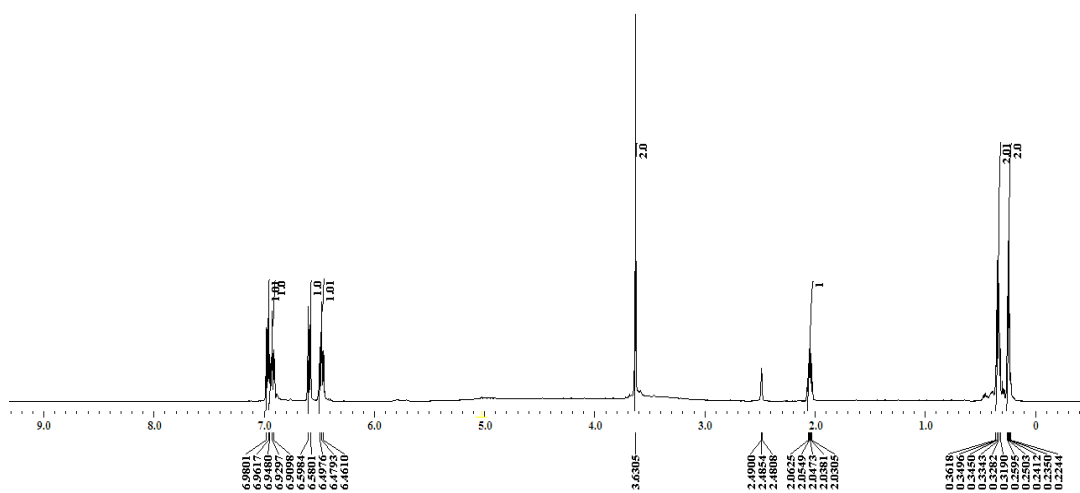
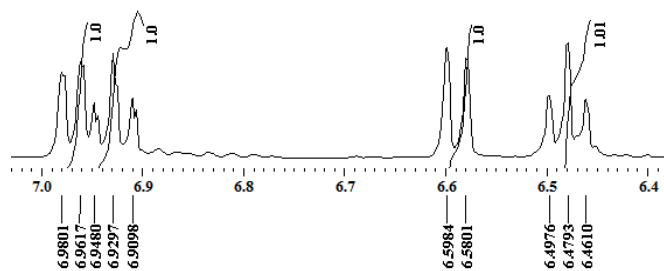
^{13}C NMR in DMSO- d_6 (100Mz)



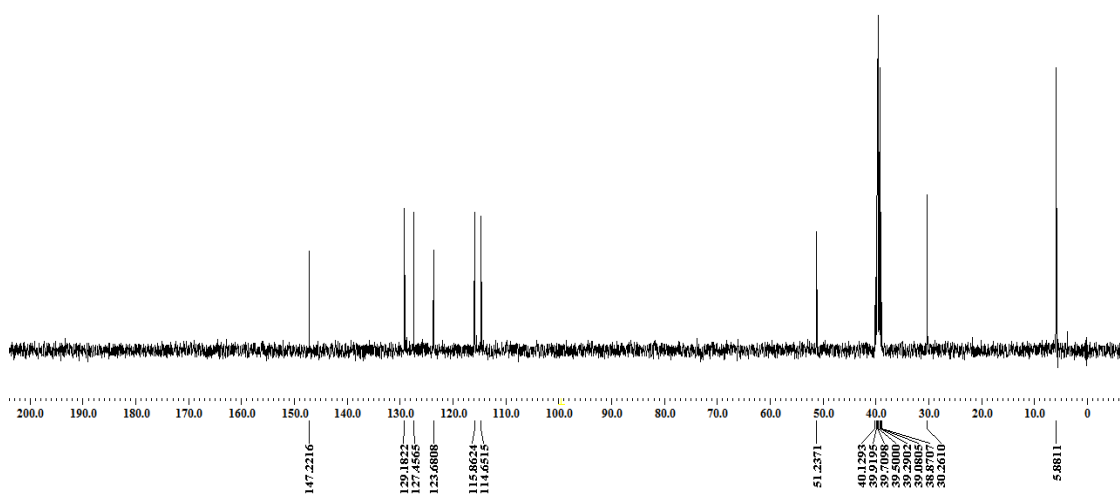


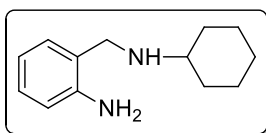
2h

¹H NMR in DMSO-d₆ (400Mz)



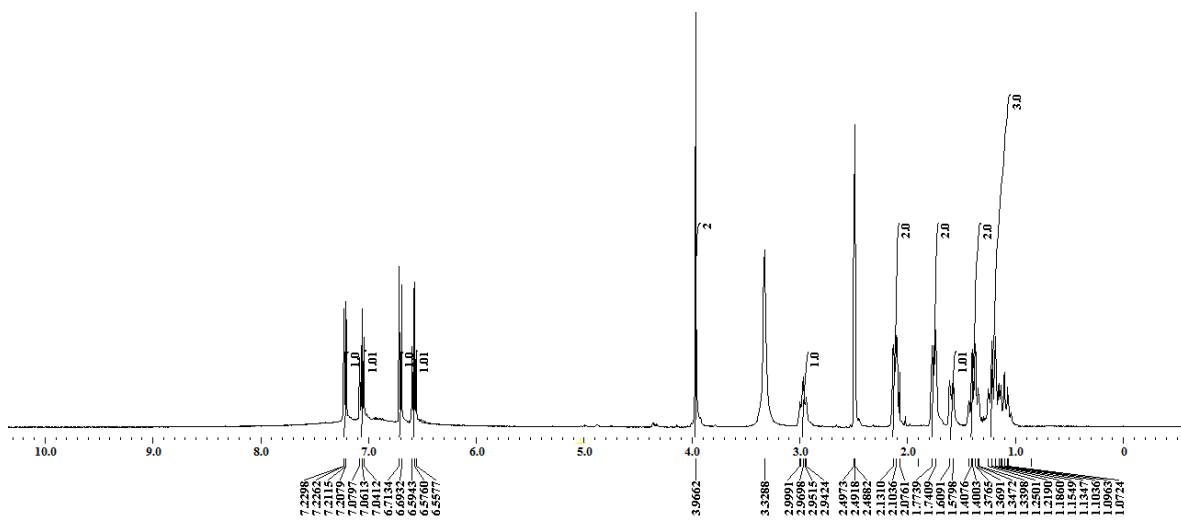
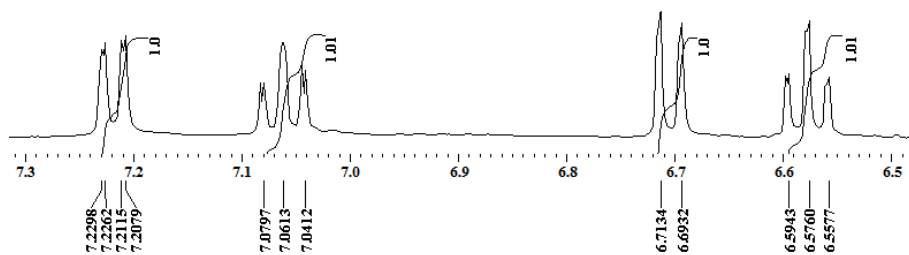
¹³C NMR in DMSO-d₆ (100Mz)



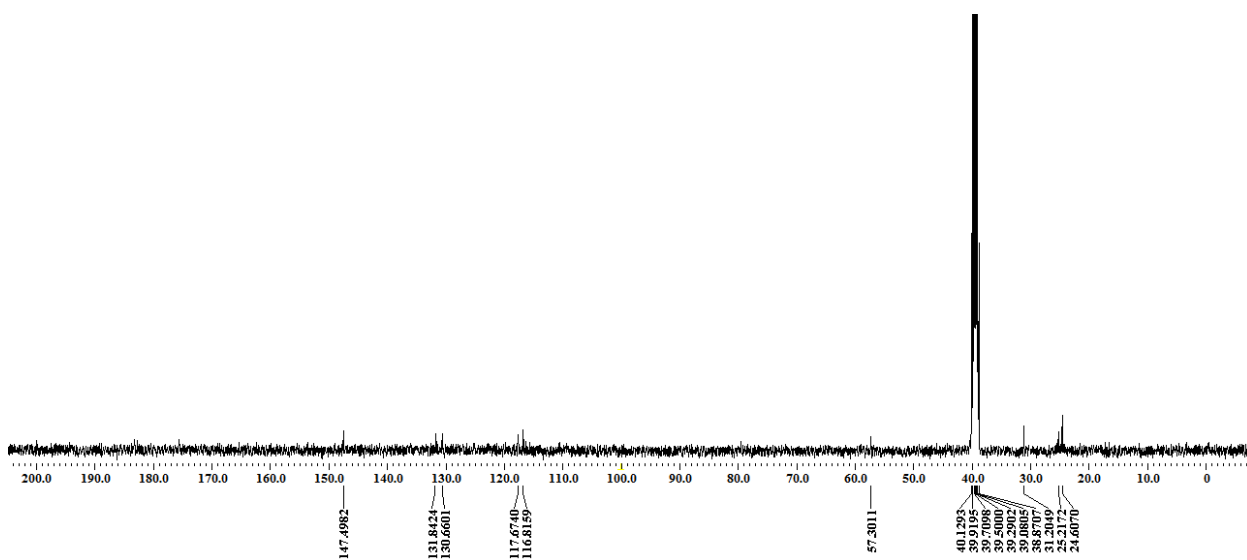


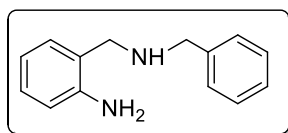
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^1H NMR in DMSO- d_6 (400Mz)



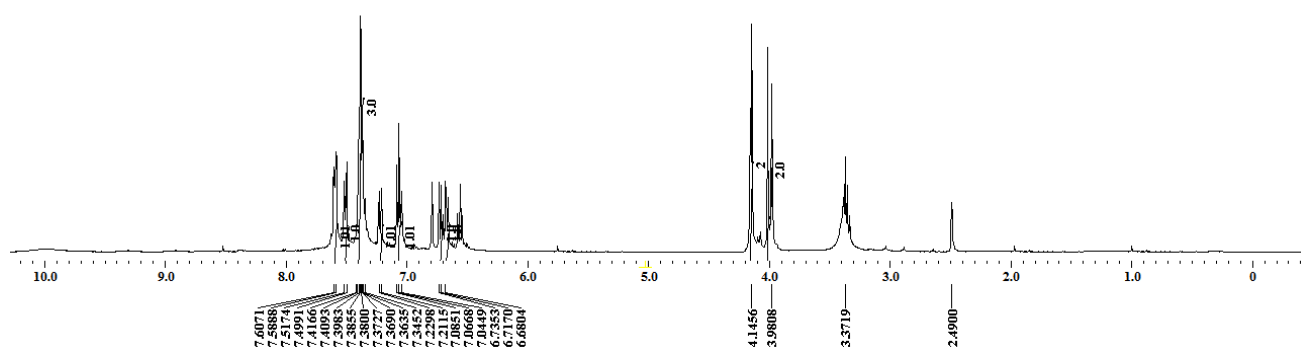
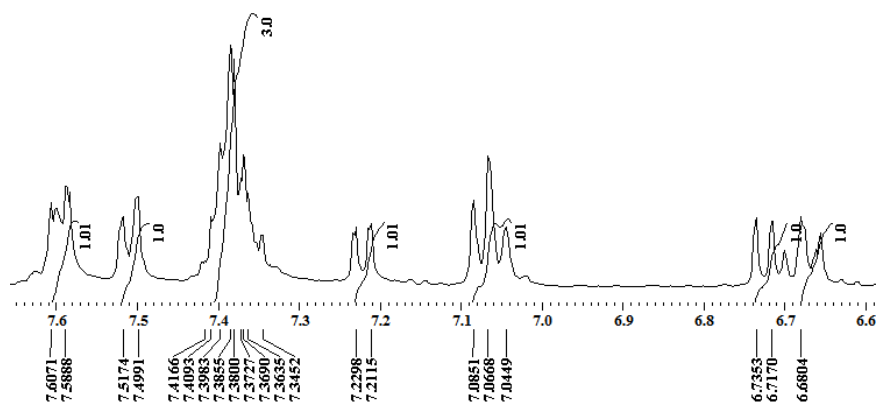
^{13}C NMR in DMSO- d_6 (100Mz)

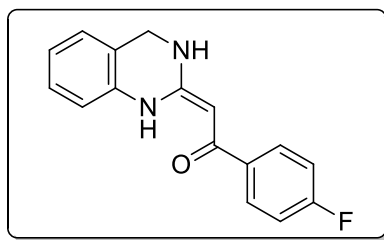




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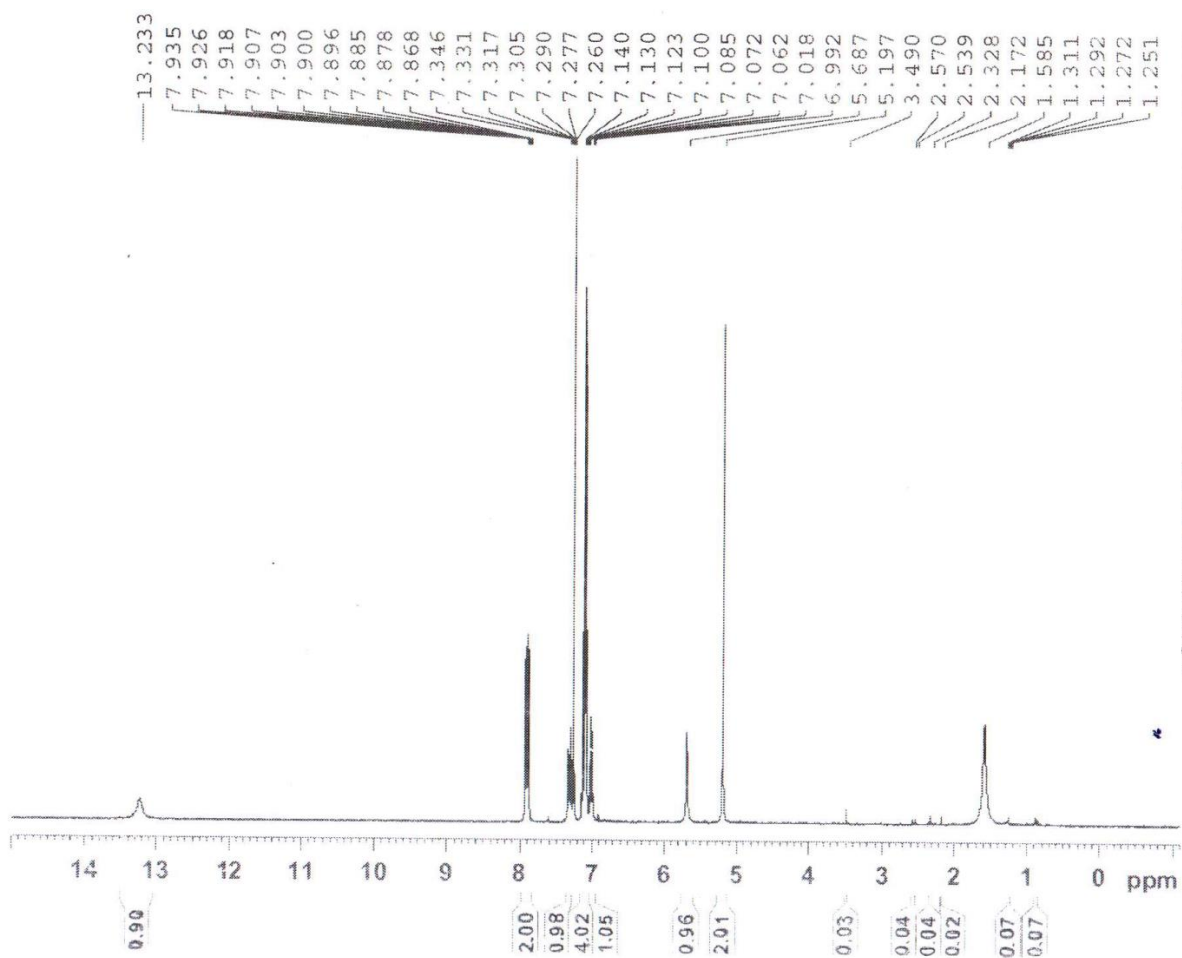
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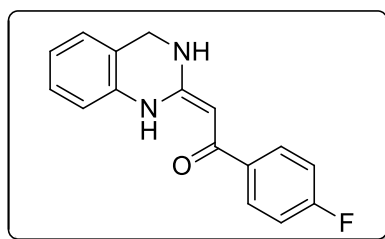




4a

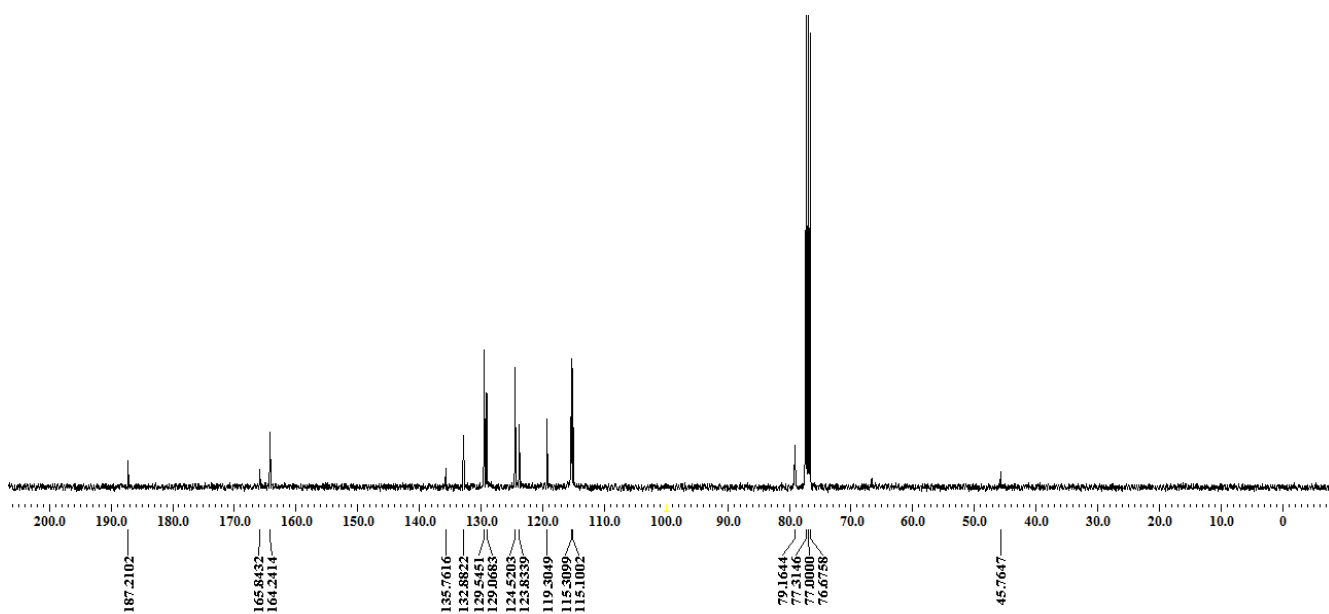
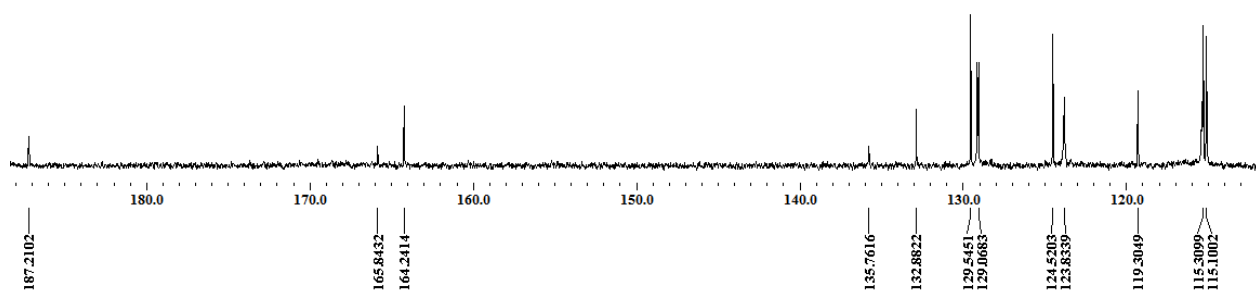
¹H-NMR in CDCl₃ (400MHz)

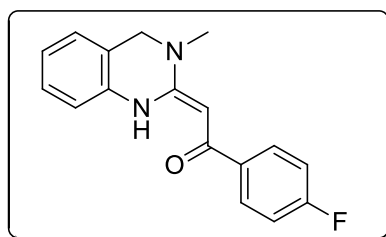




4a

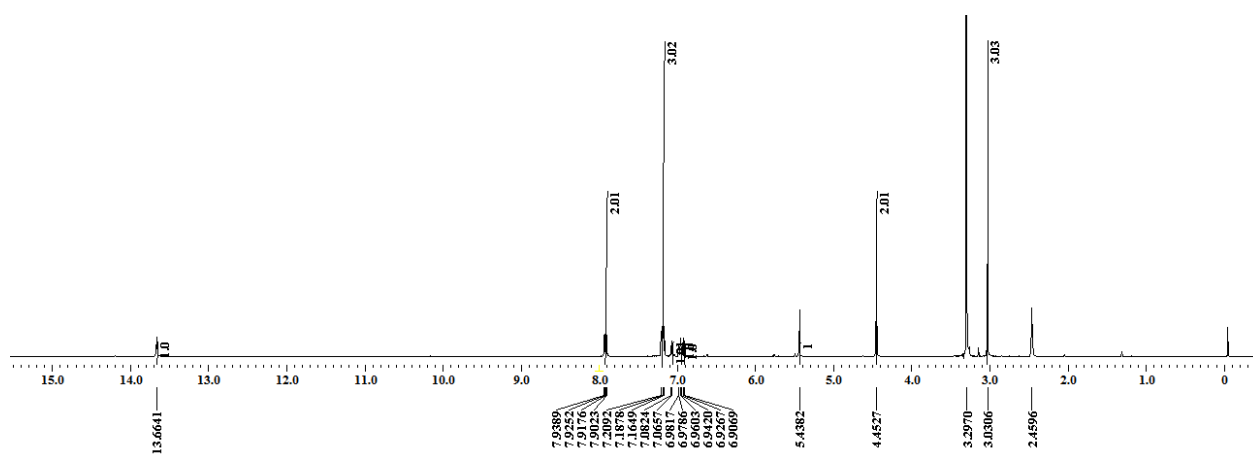
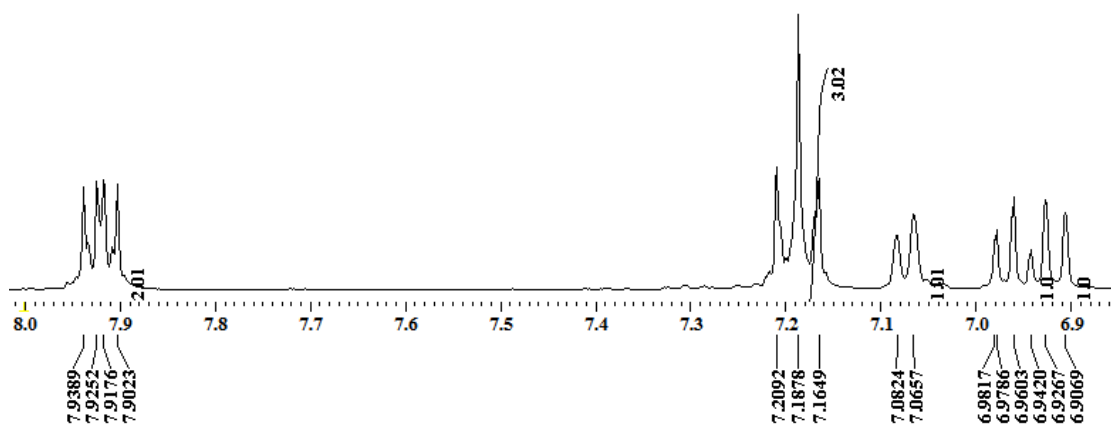
^{13}C NMR in CDCl_3 (100MHz)

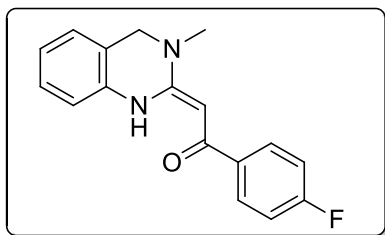




4b

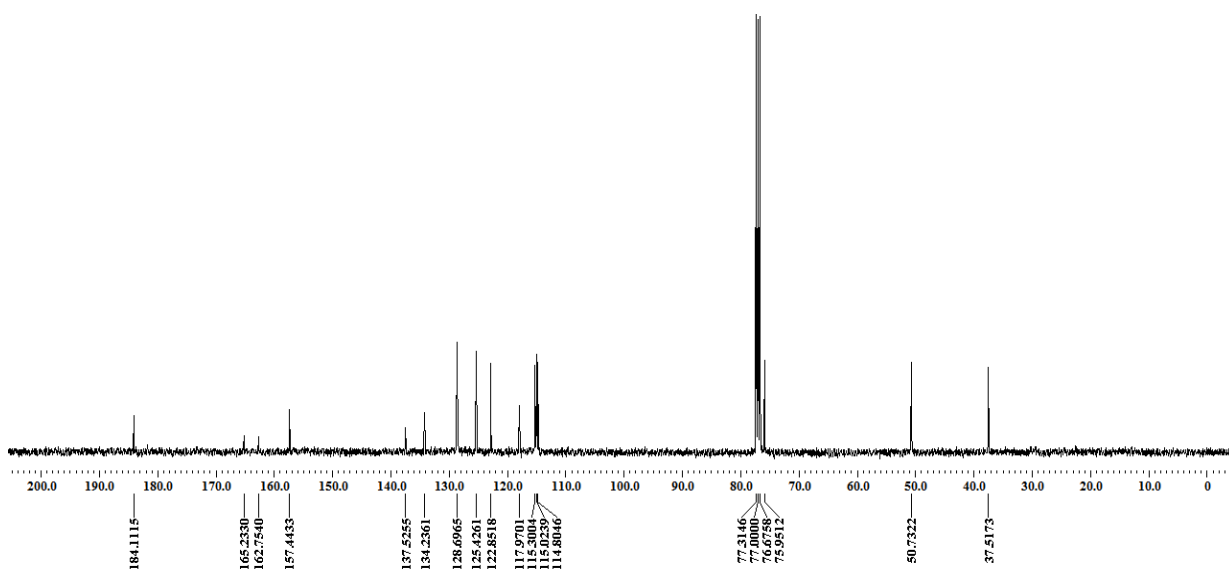
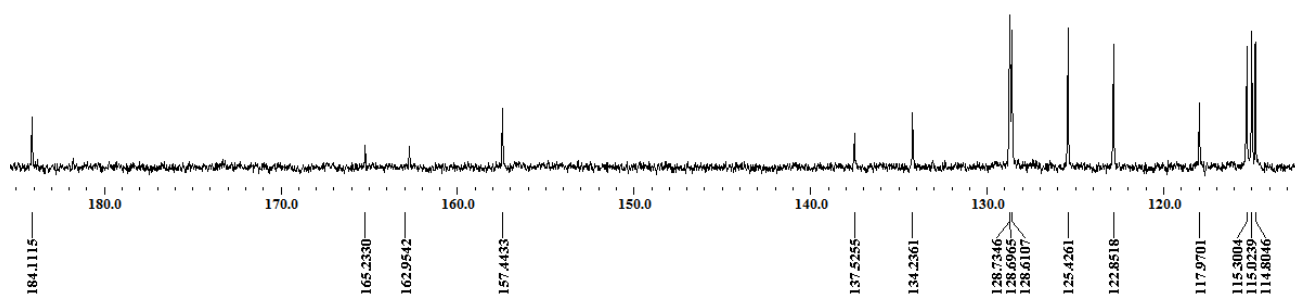
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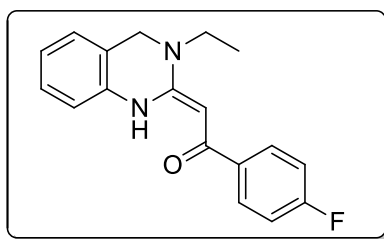




4b

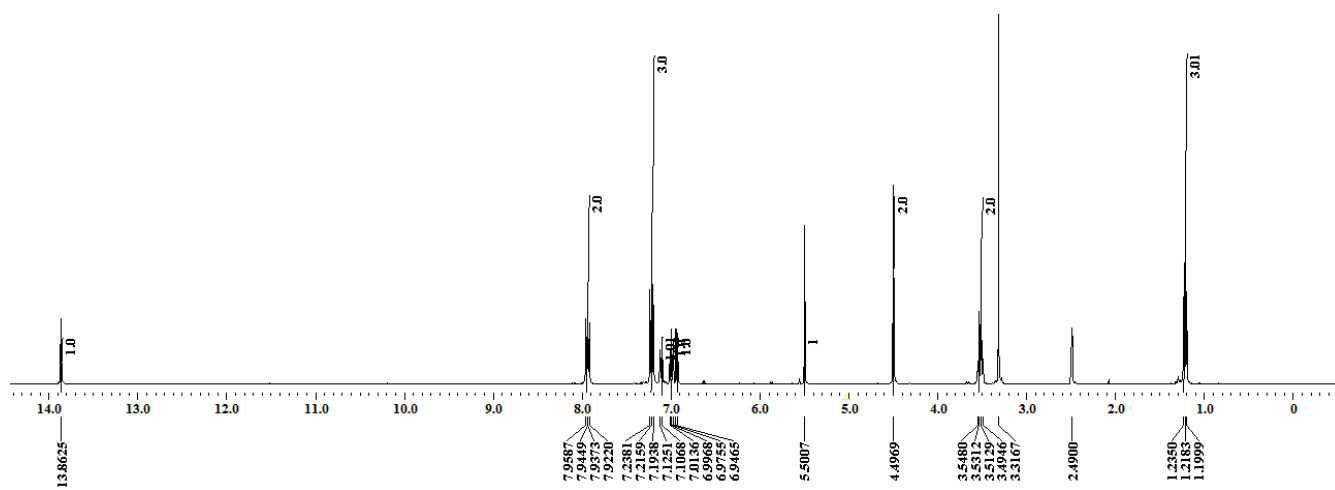
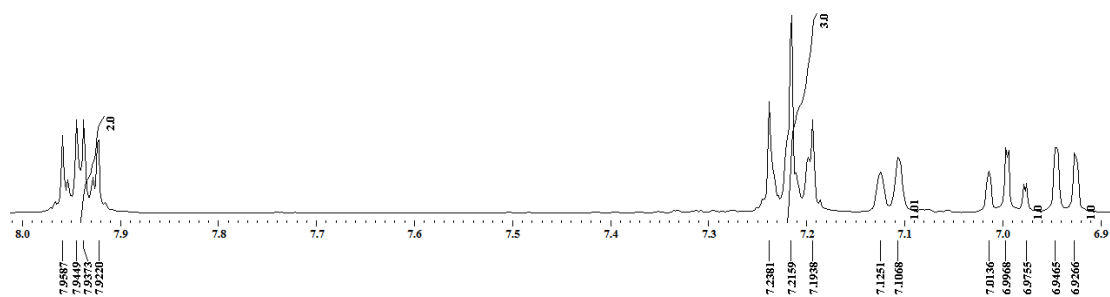
^{13}C NMR in CDCl_3 (100MHz)

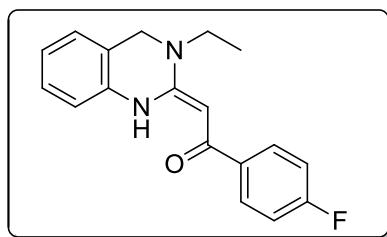




4c

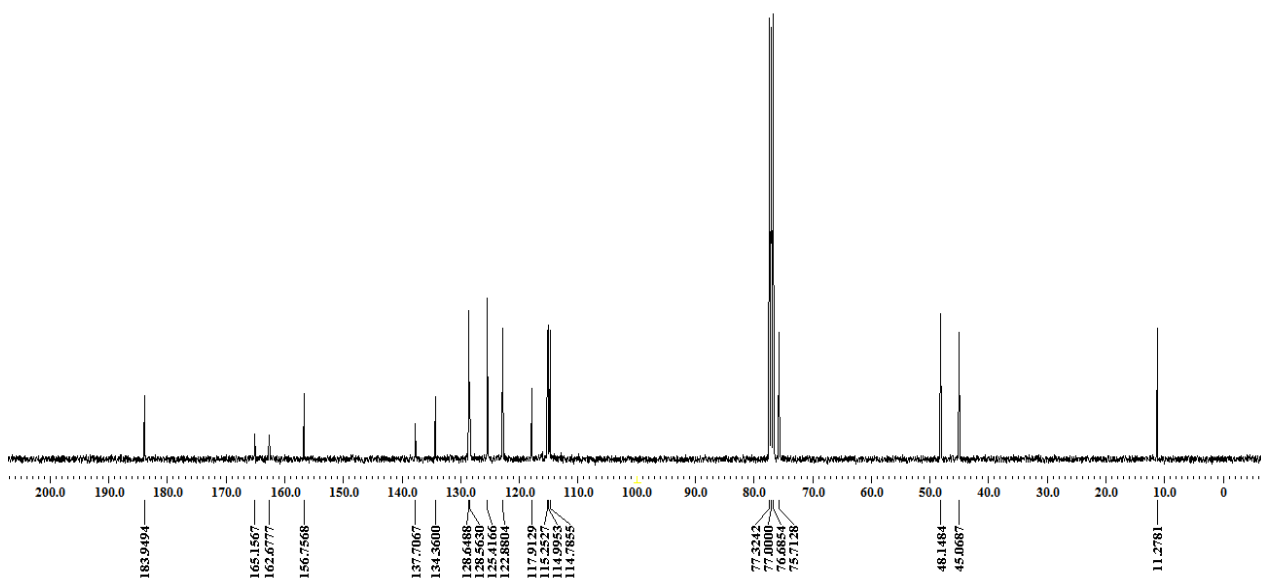
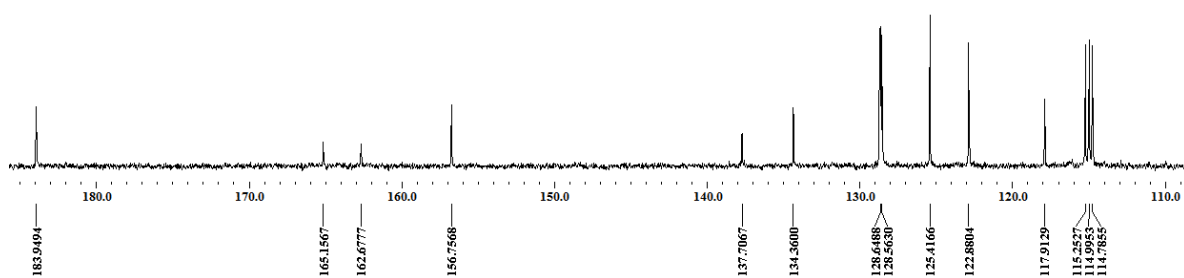
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

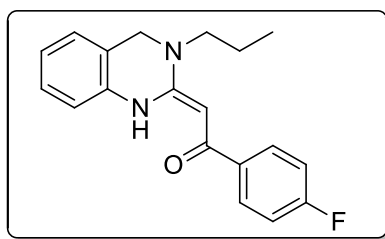




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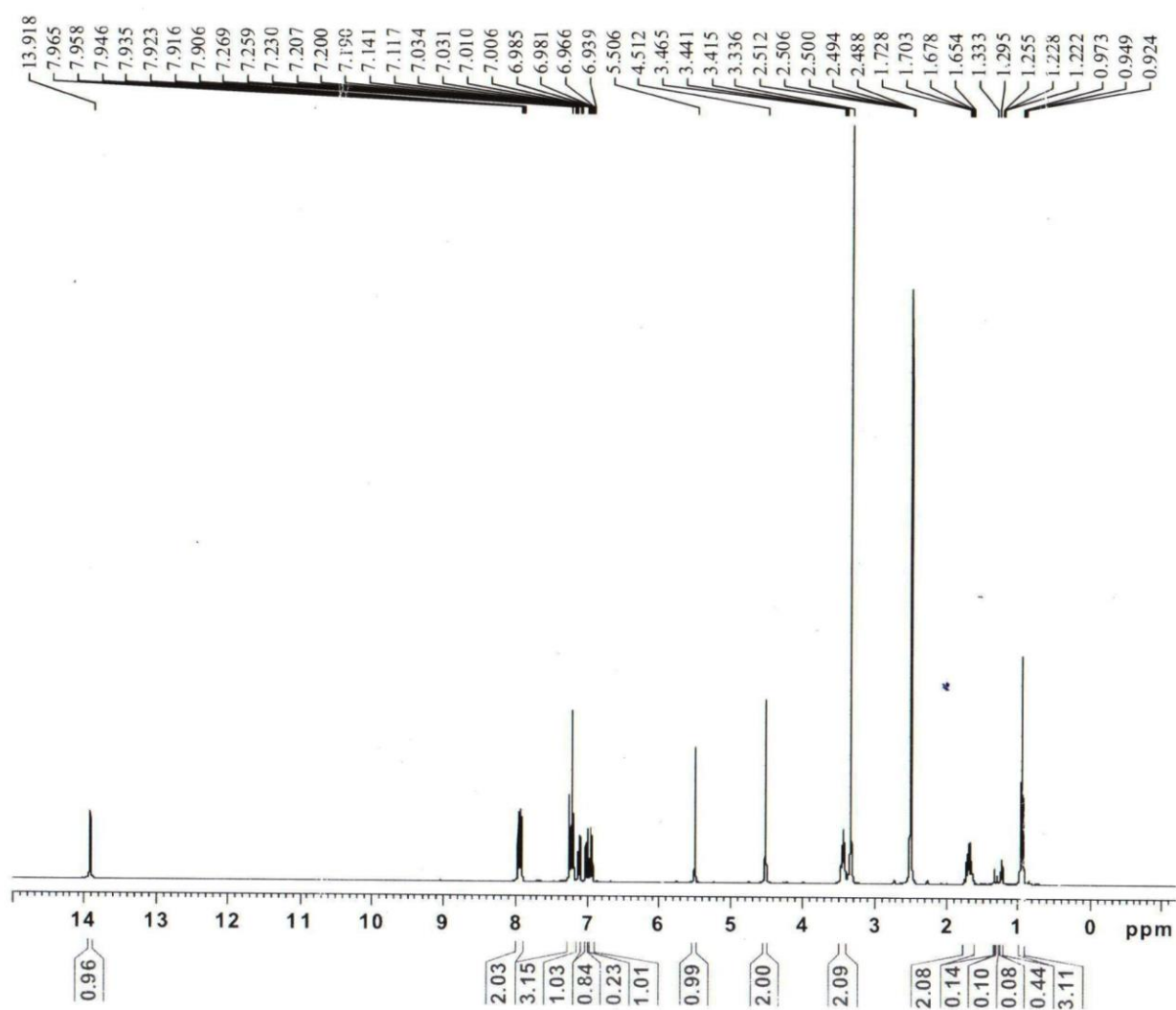
^{13}C -NMR in CDCl_3 (100MHz)

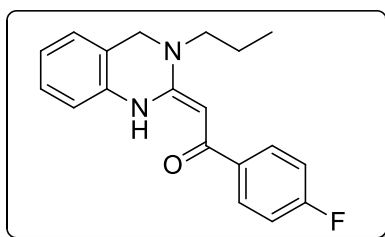




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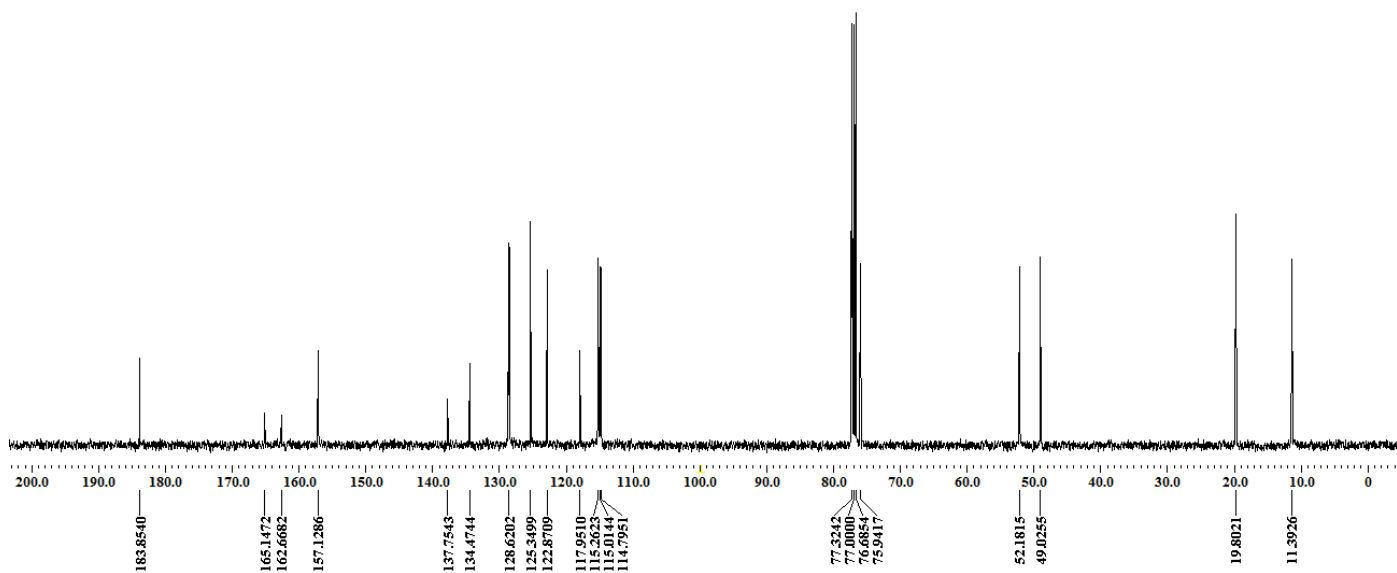
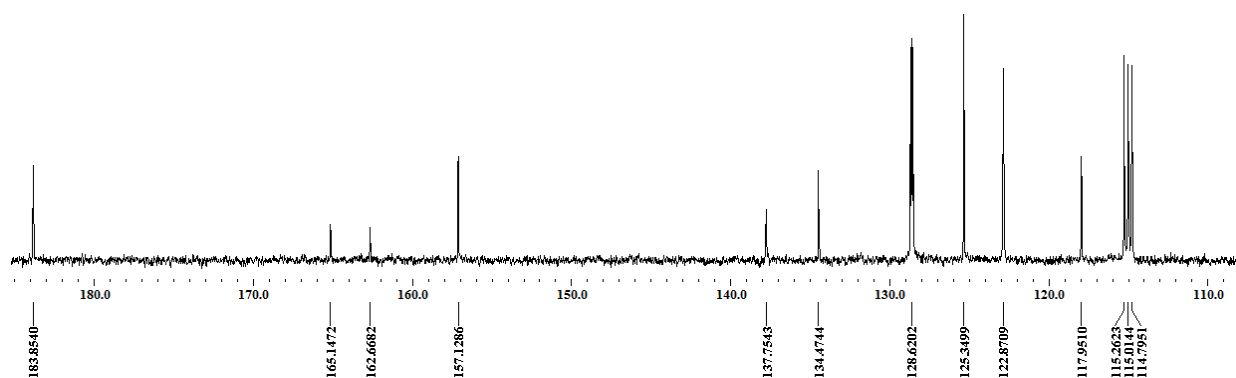
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

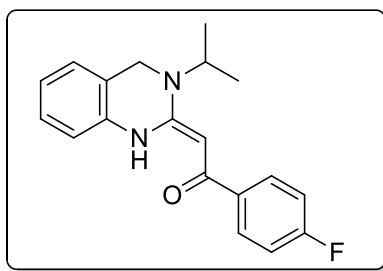




4d

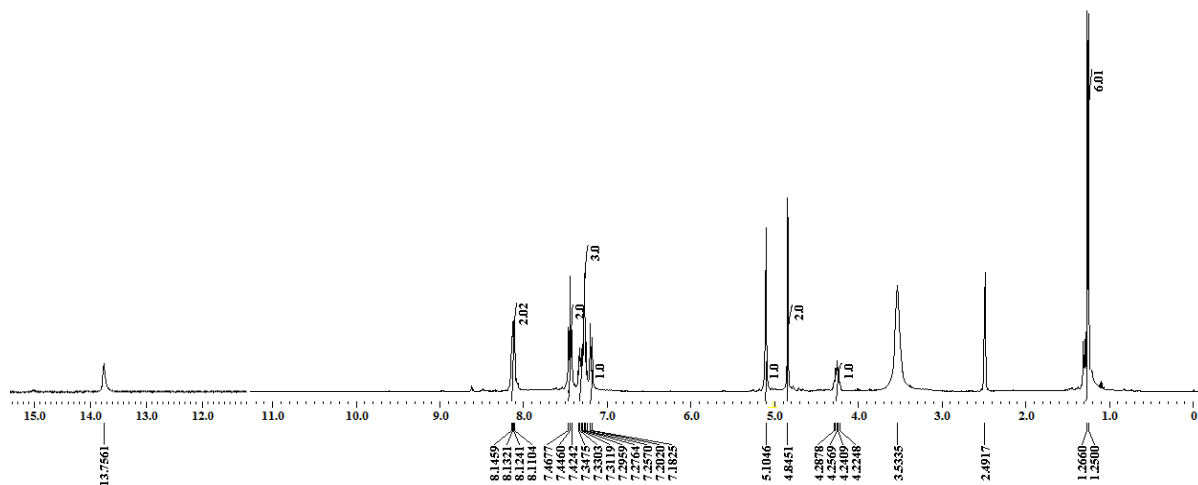
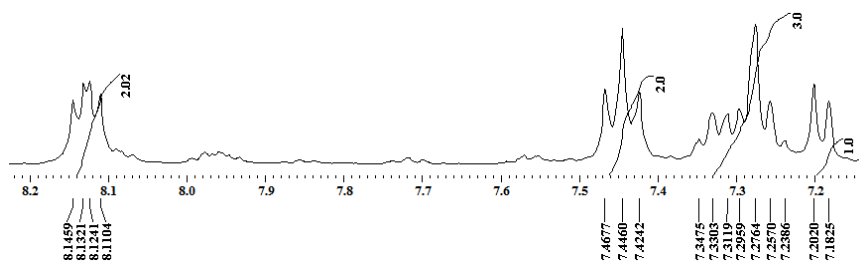
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

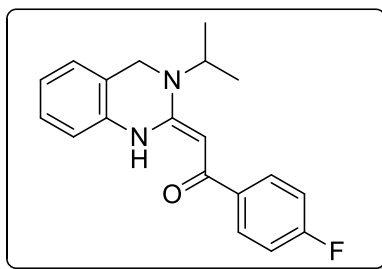




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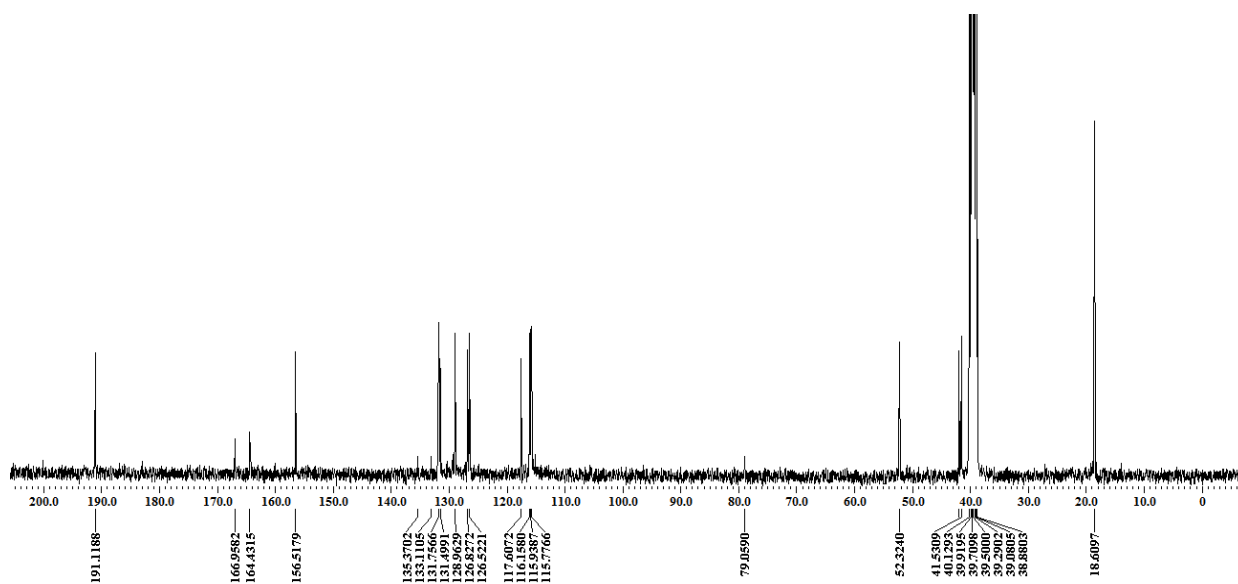
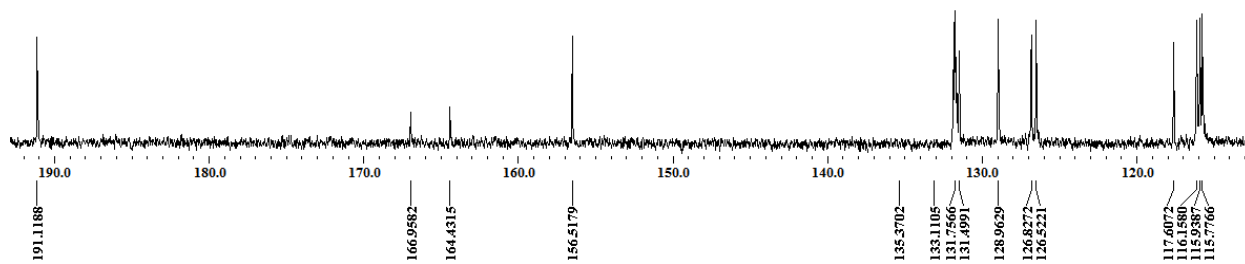
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

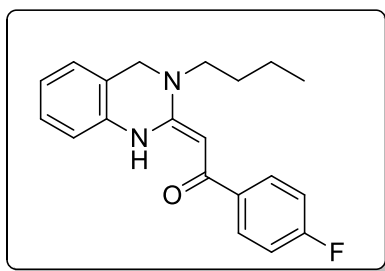




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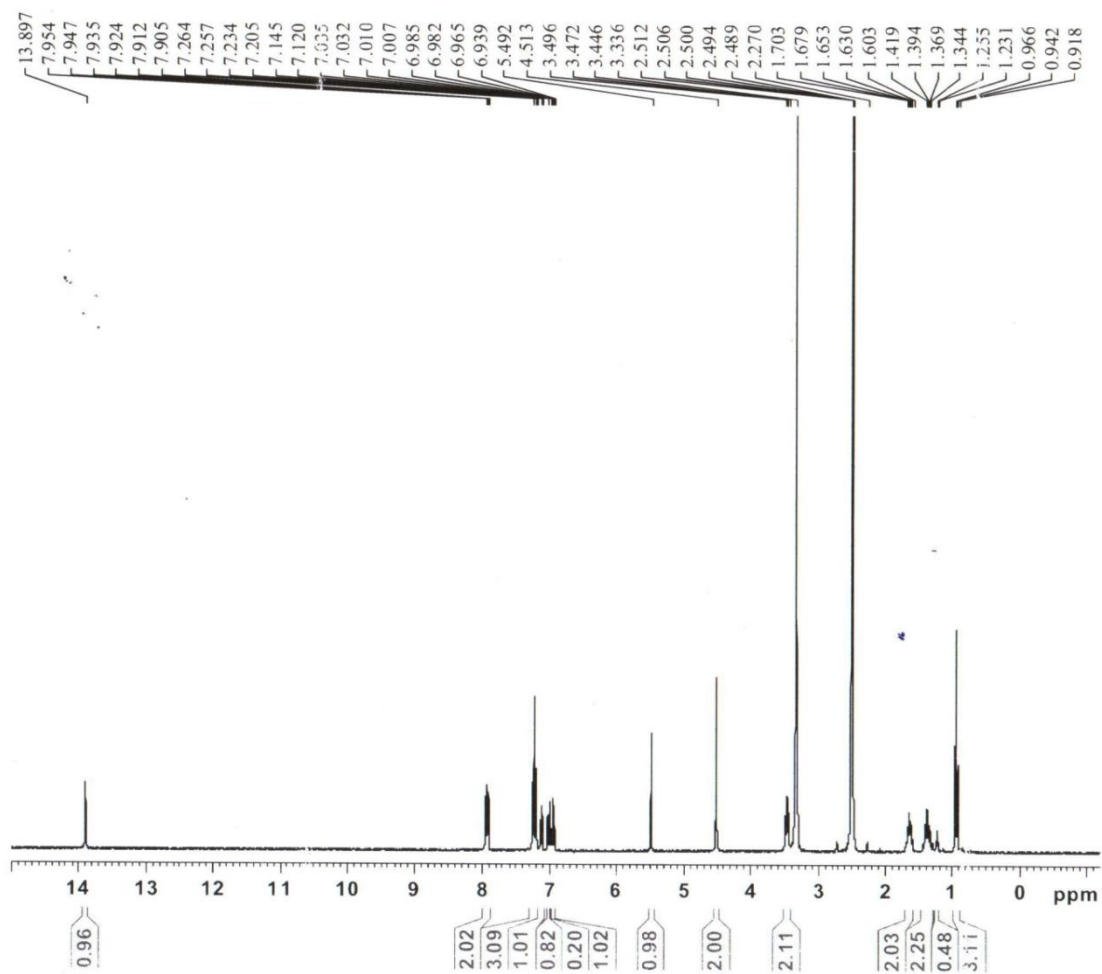
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

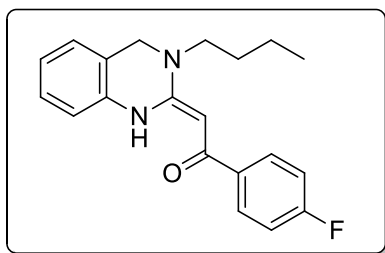




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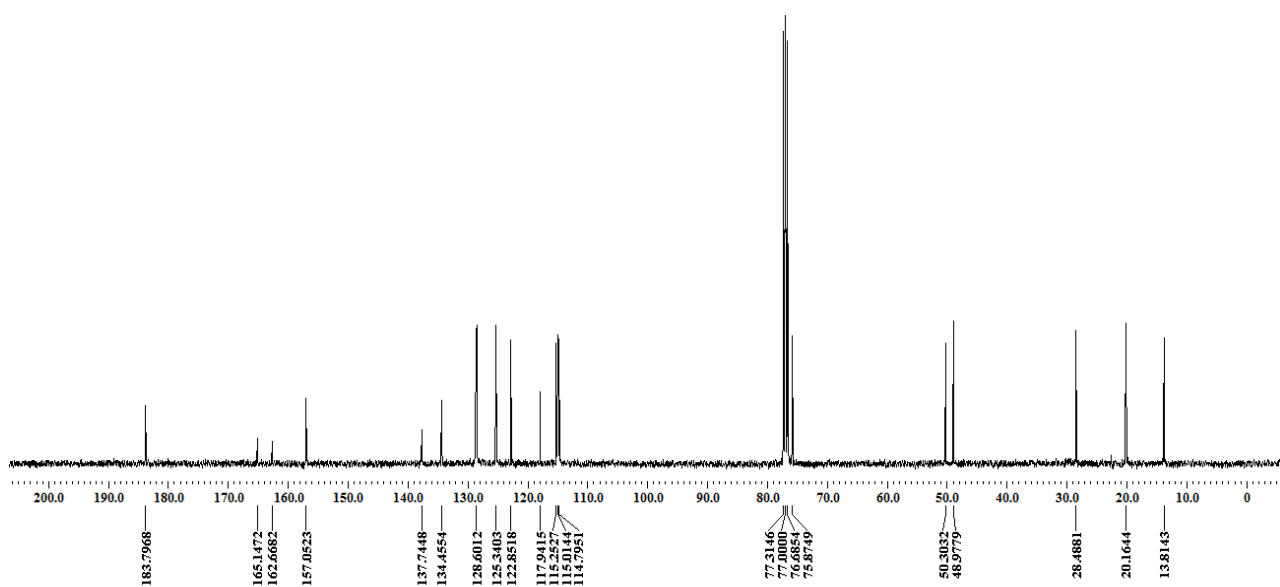
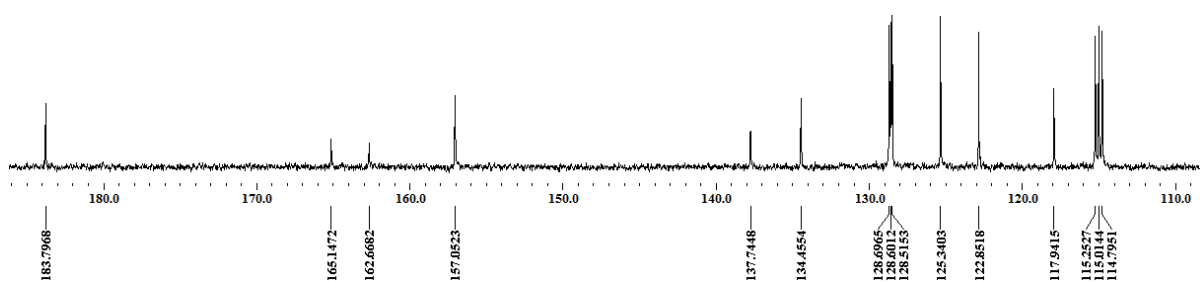
¹H-NMR in DMSO-d₆ (400MHz)

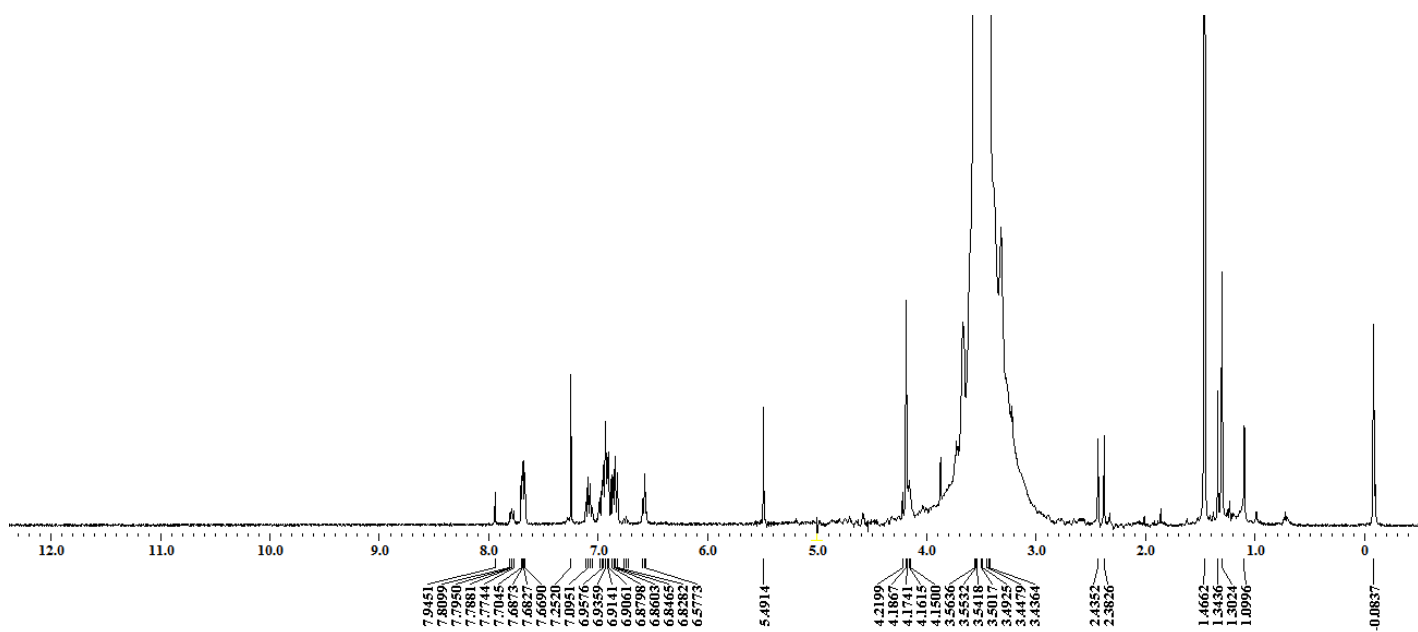
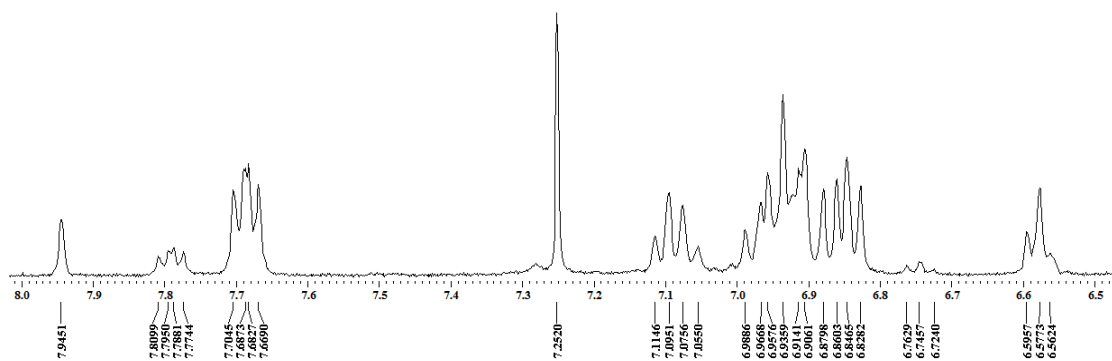
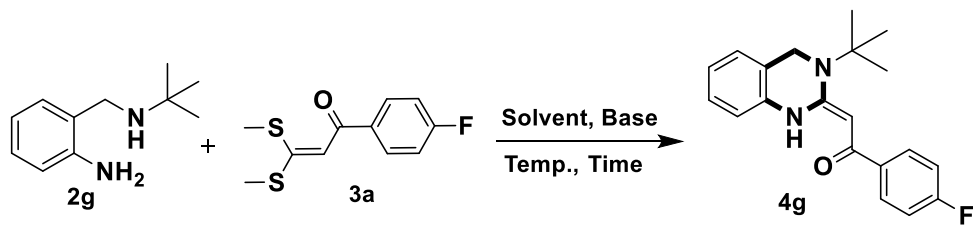


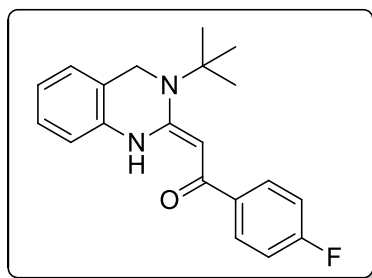


4f

$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

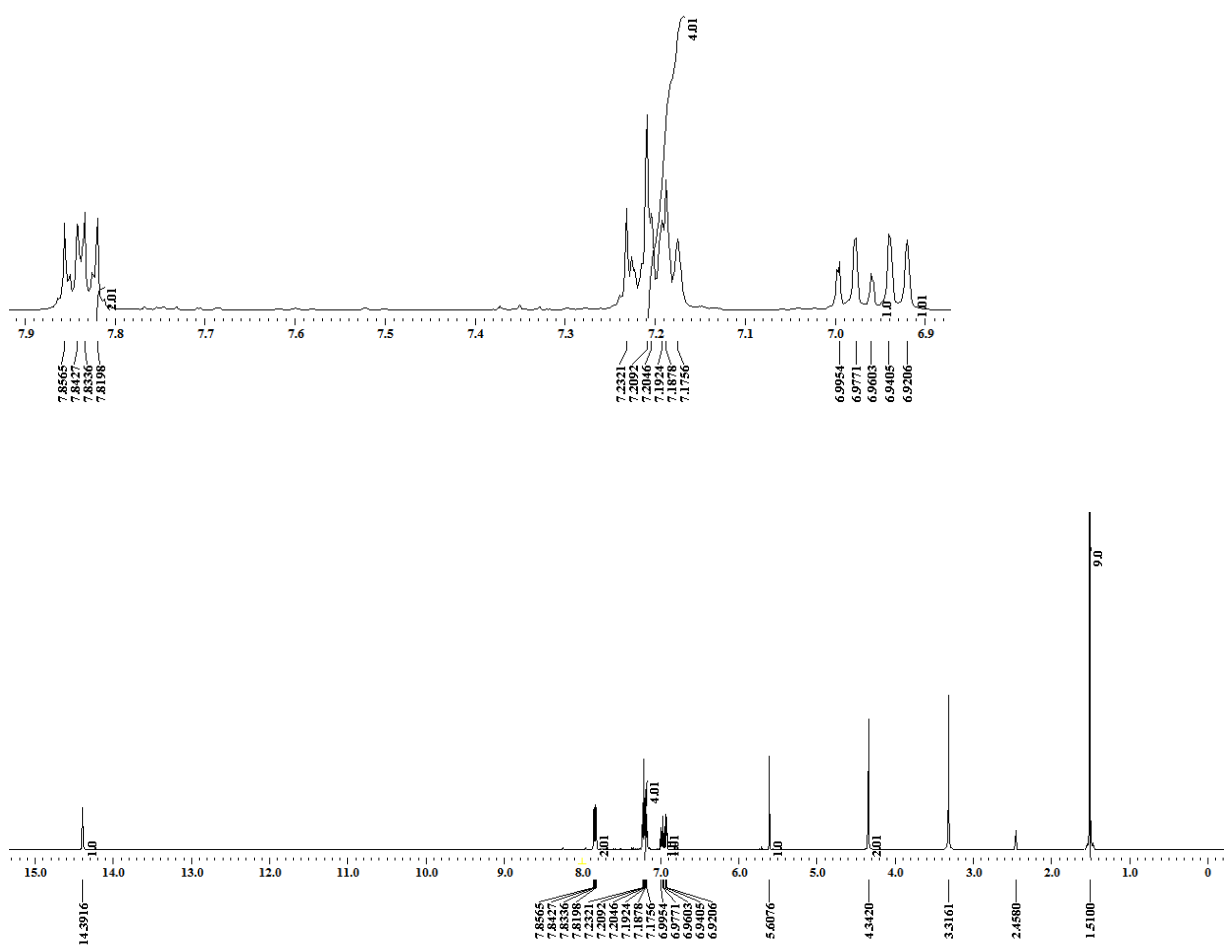


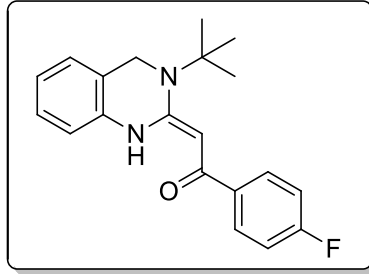




4g

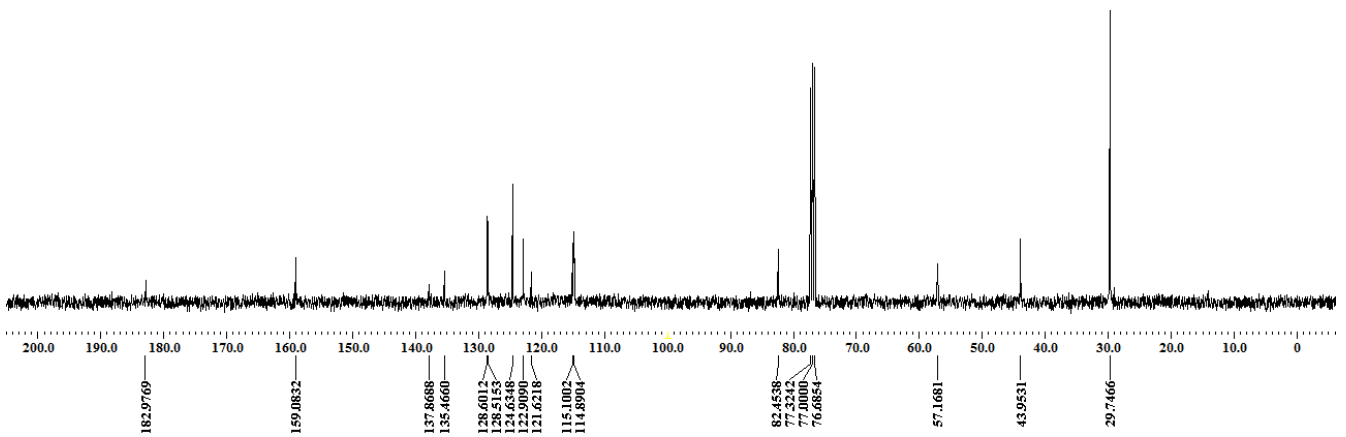
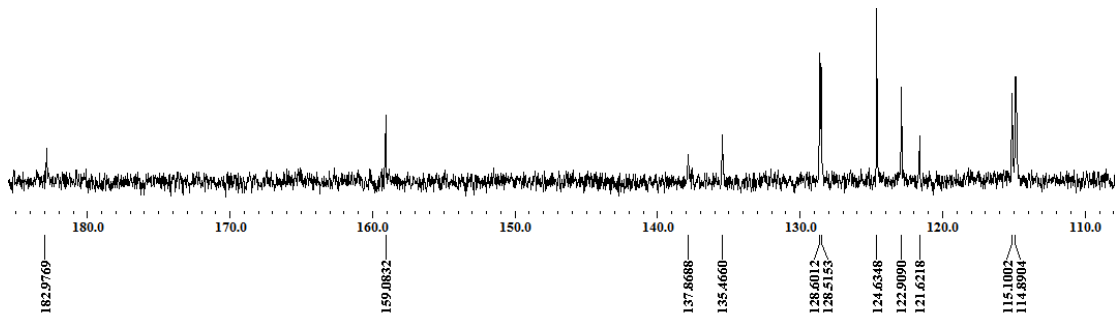
¹H-NMR in CDCl₃ (400MHz)

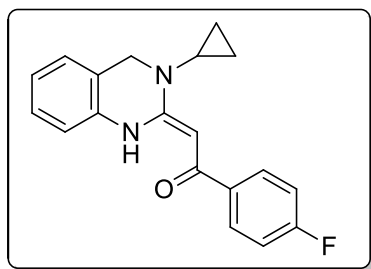




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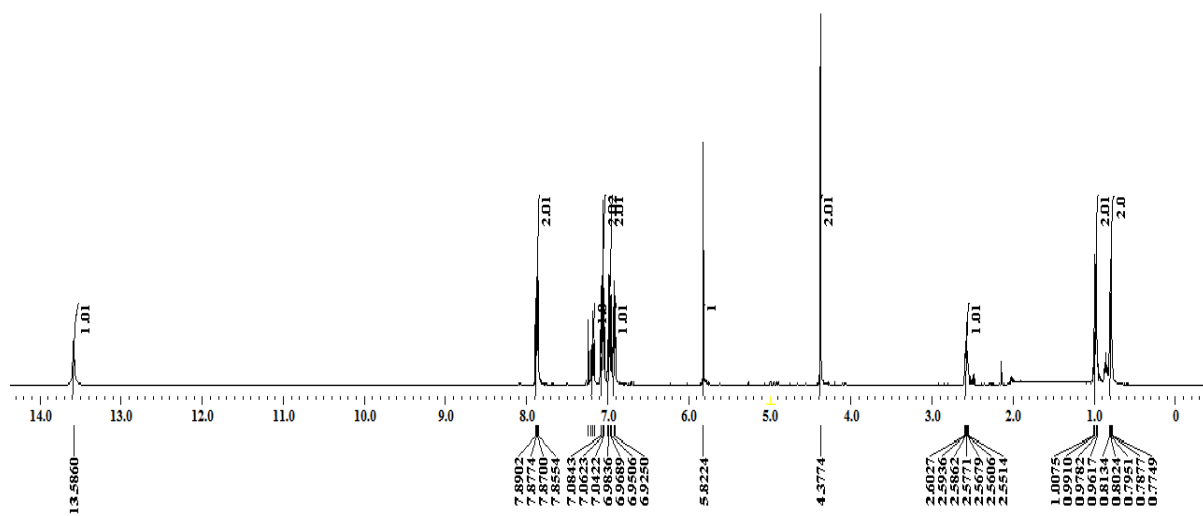
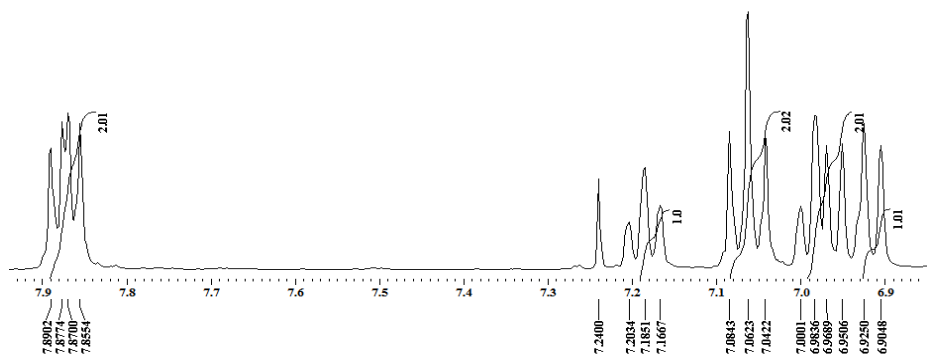
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

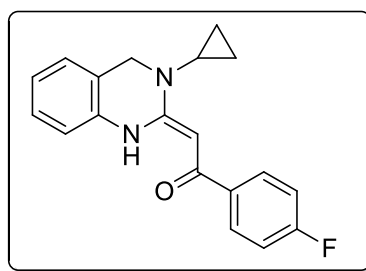




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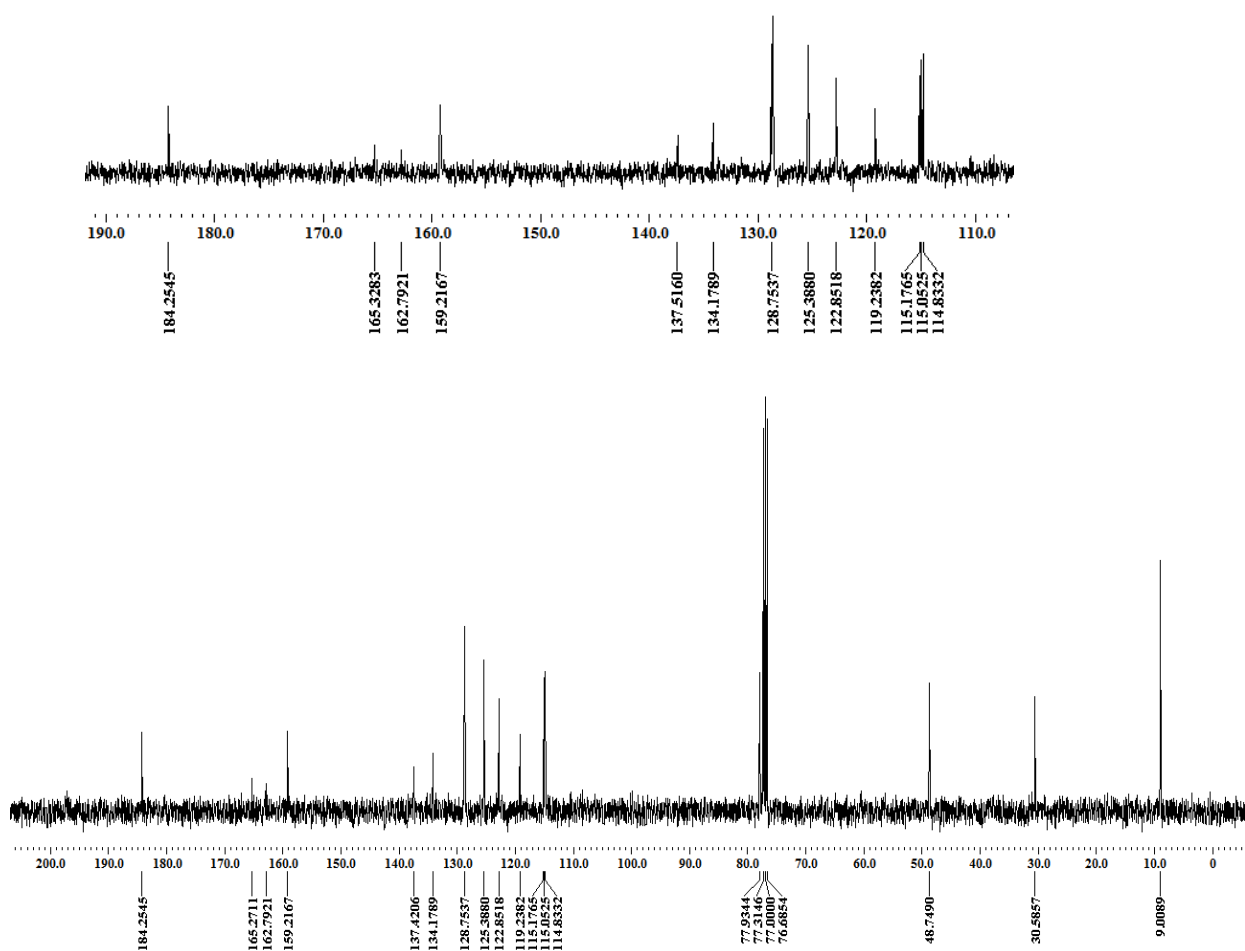
¹H-NMR in DMSO-d₆ (400MHz)

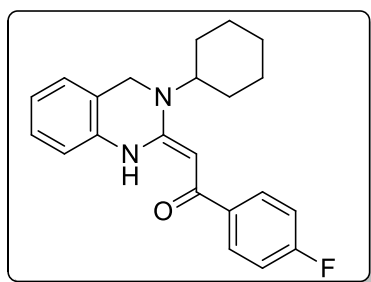




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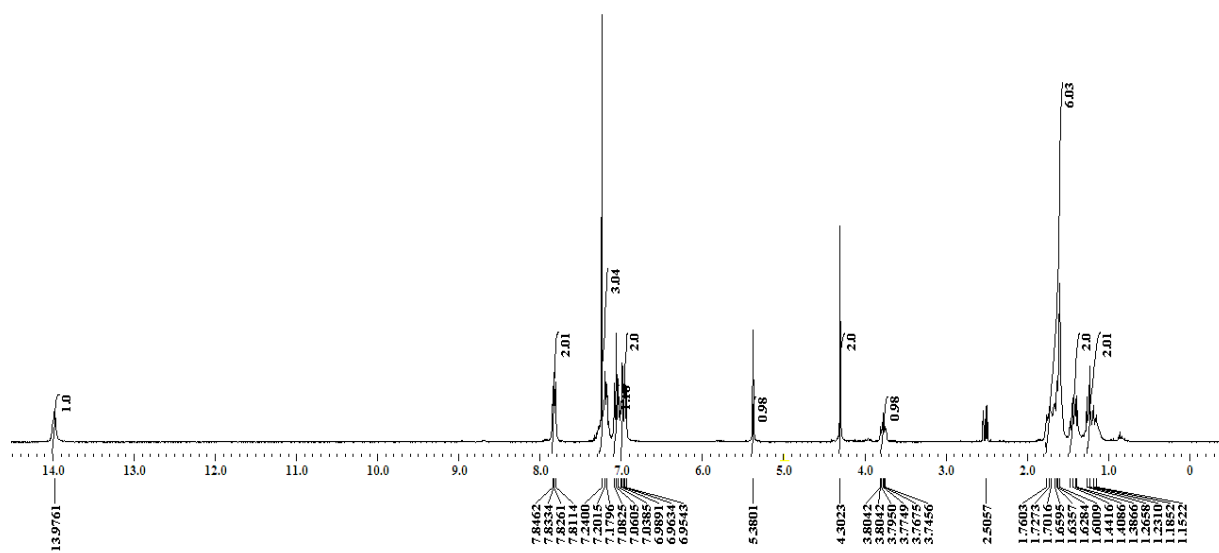
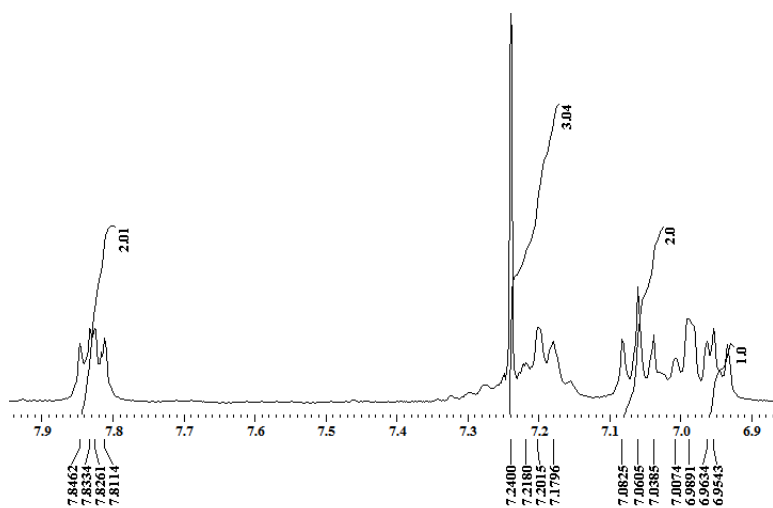
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

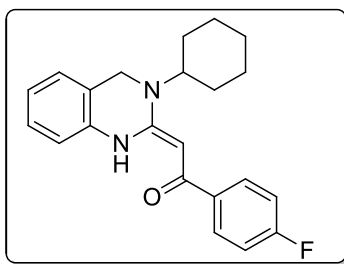




4i

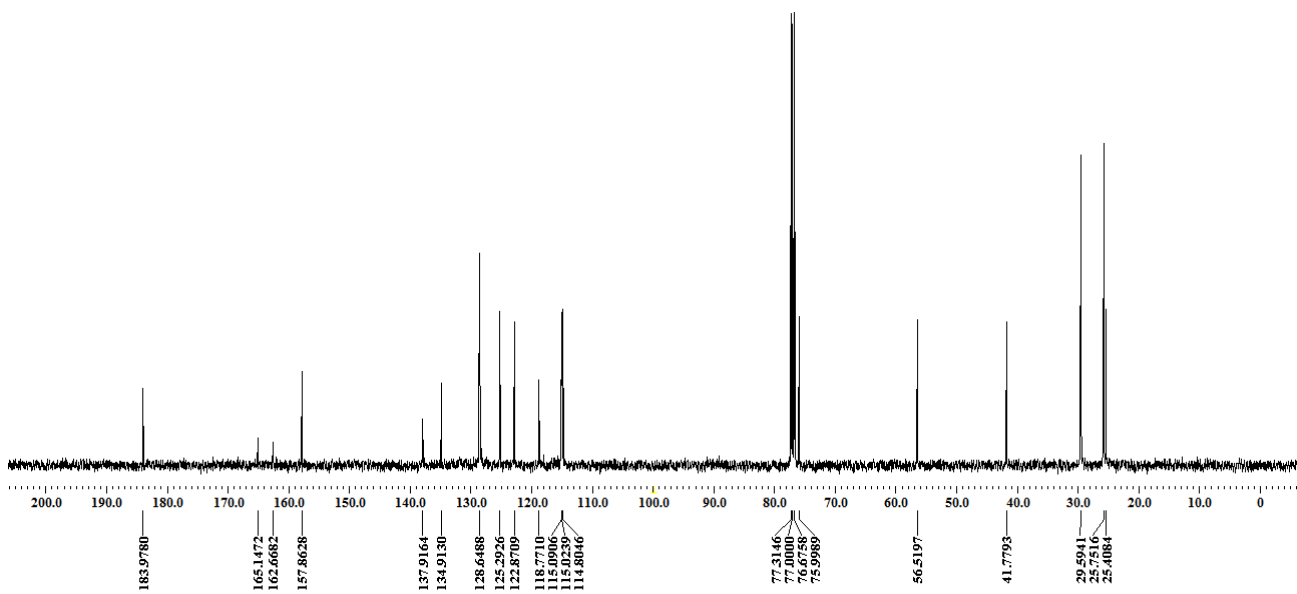
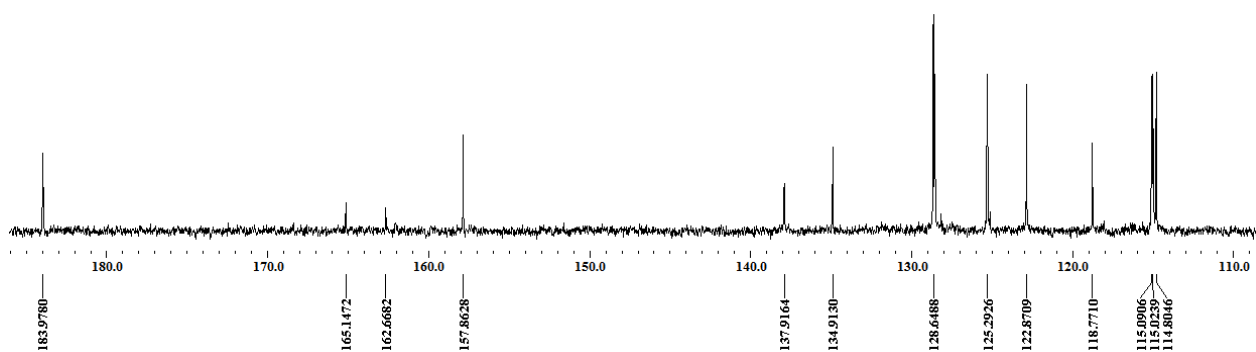
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

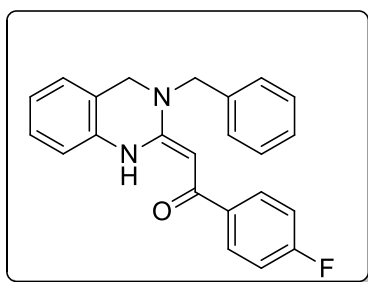




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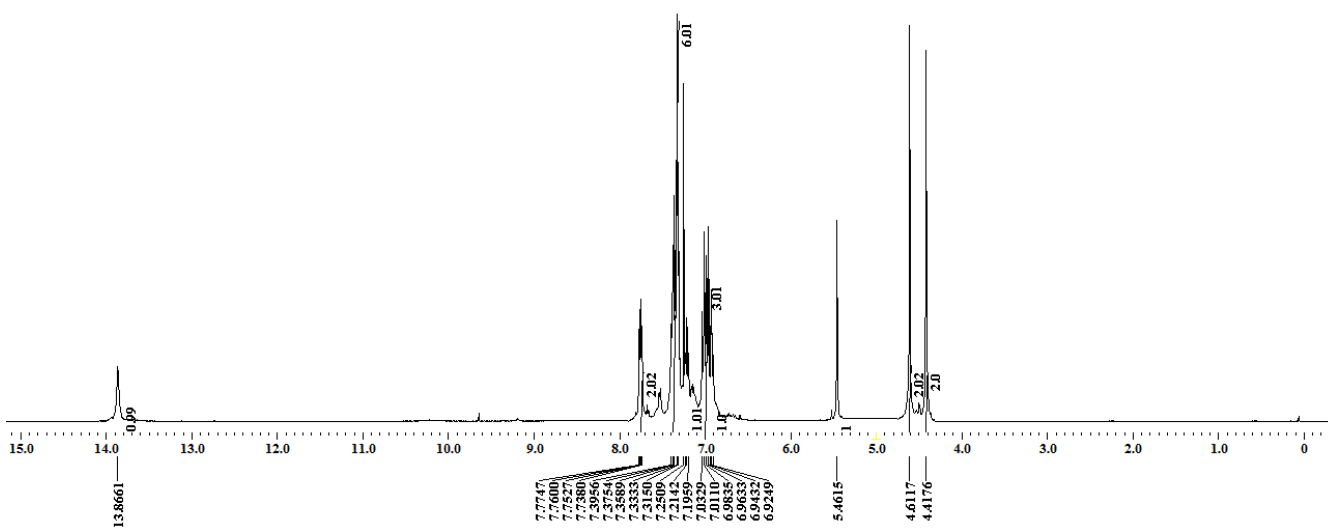
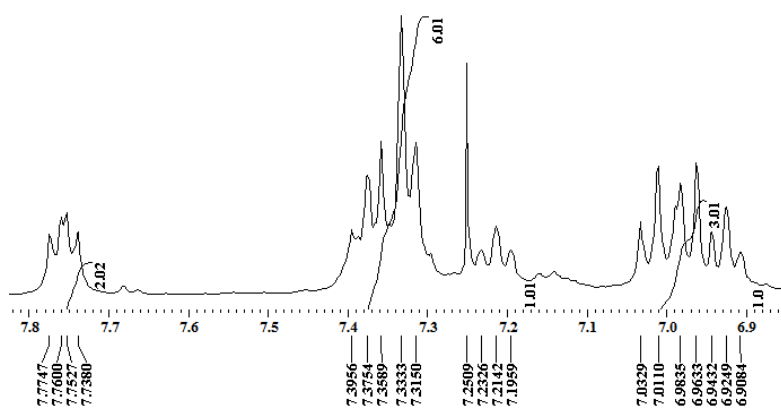
¹³C-NMR in CDCl₃ (100MHz)

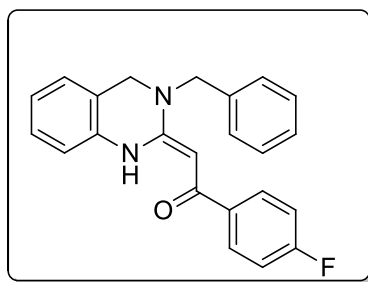




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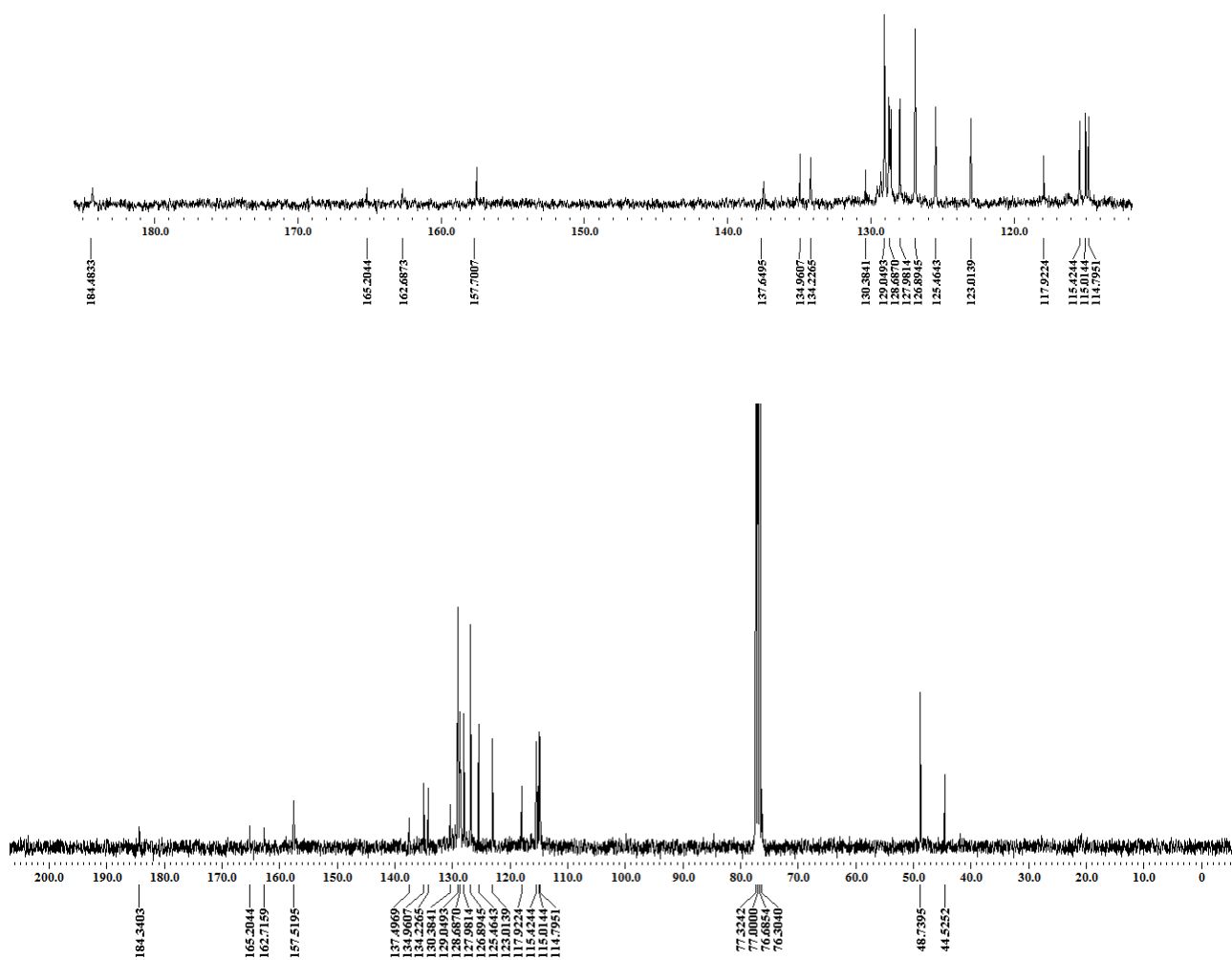
¹H-NMR in CDCl₃ (400MHz)

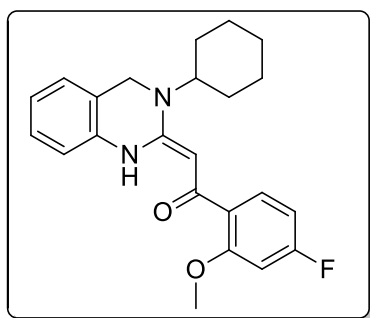




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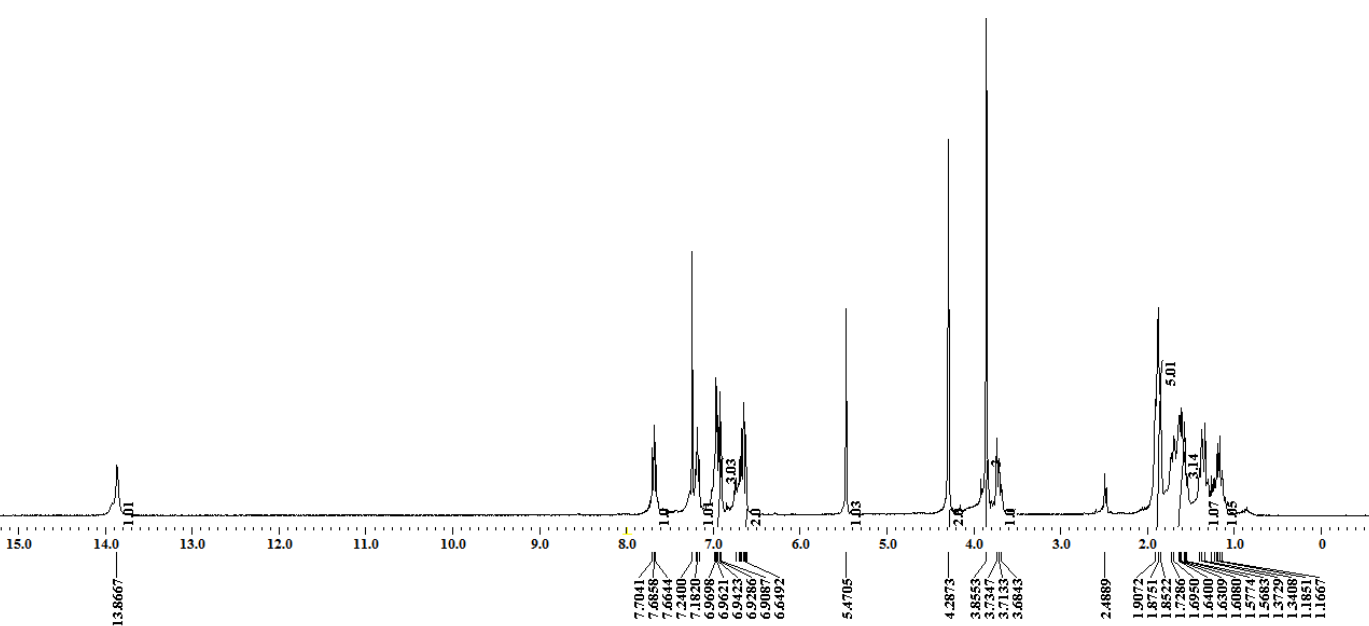
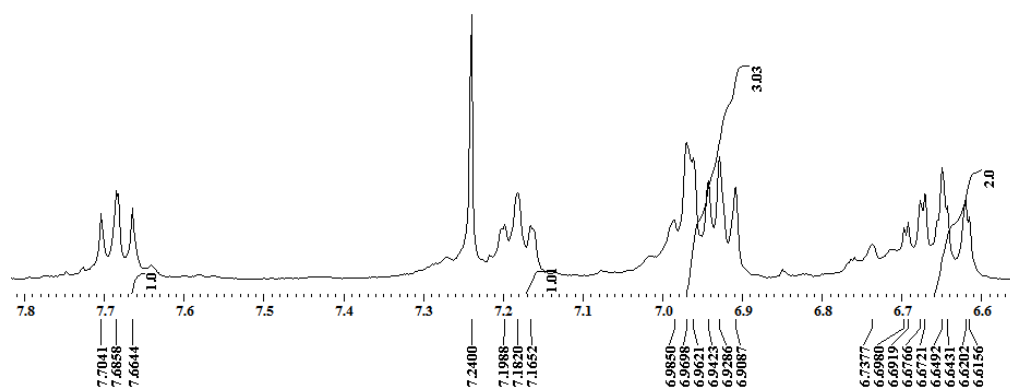
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

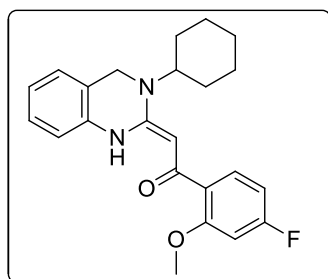




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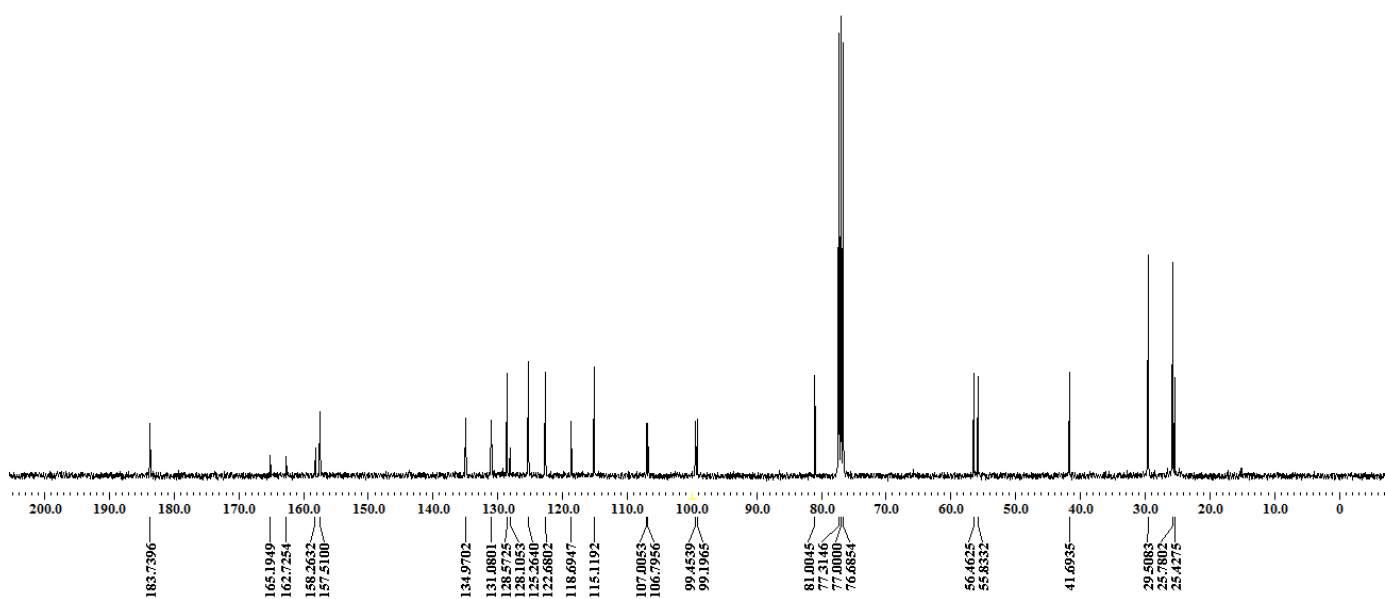
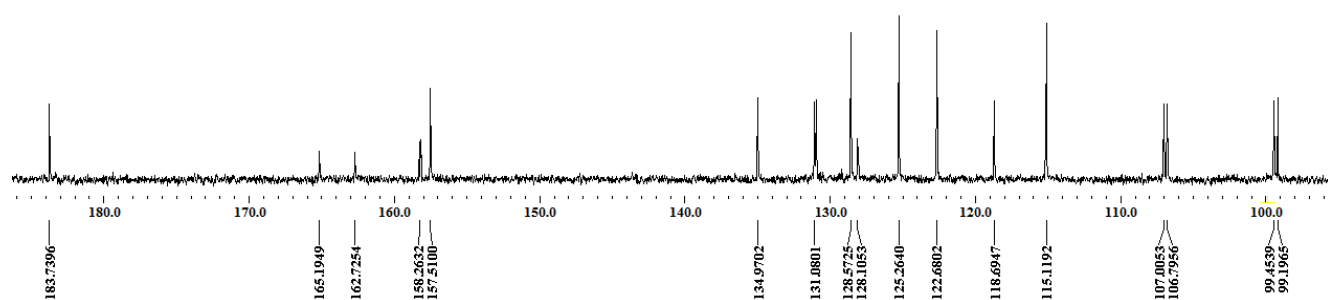
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

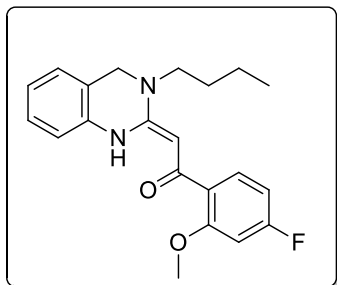




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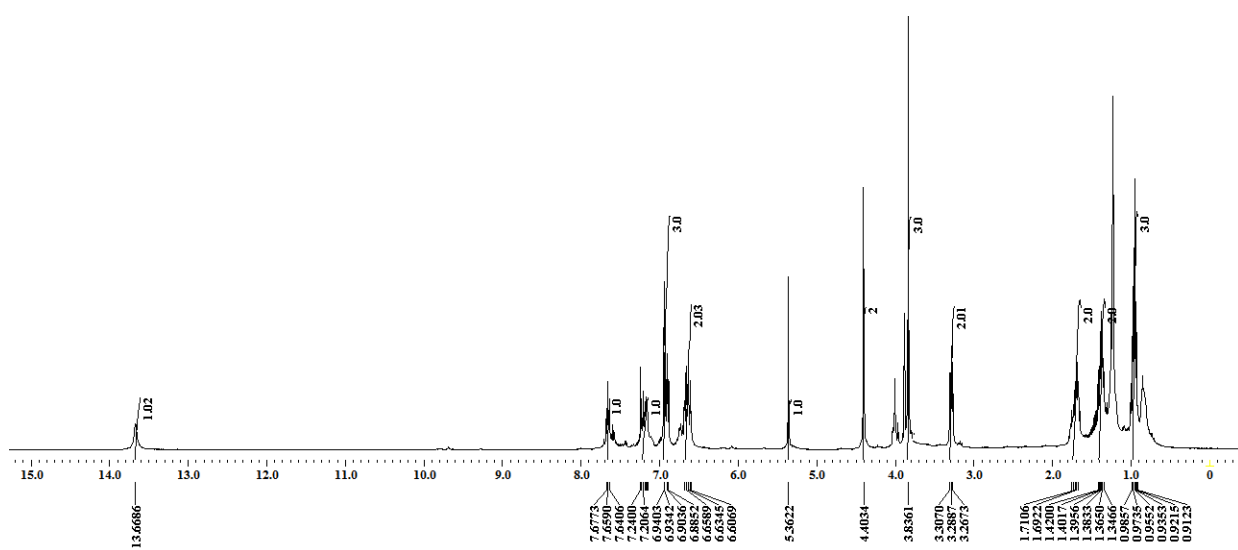
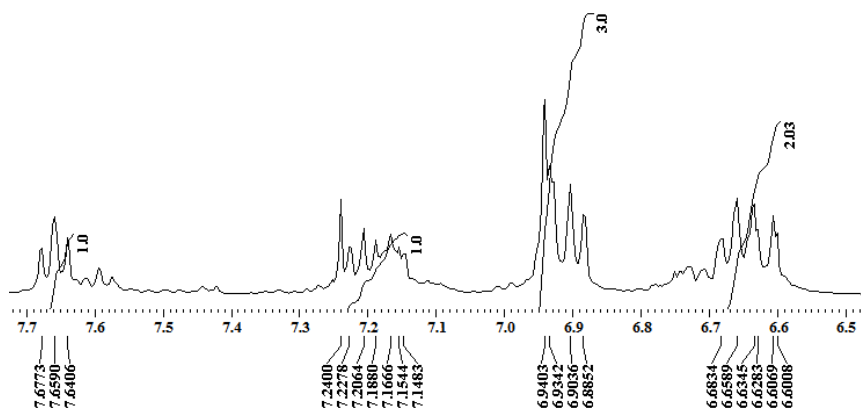
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

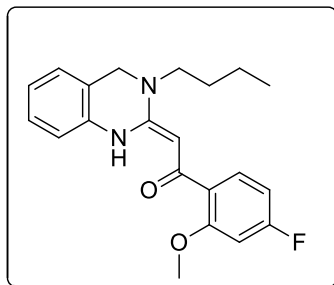




4I

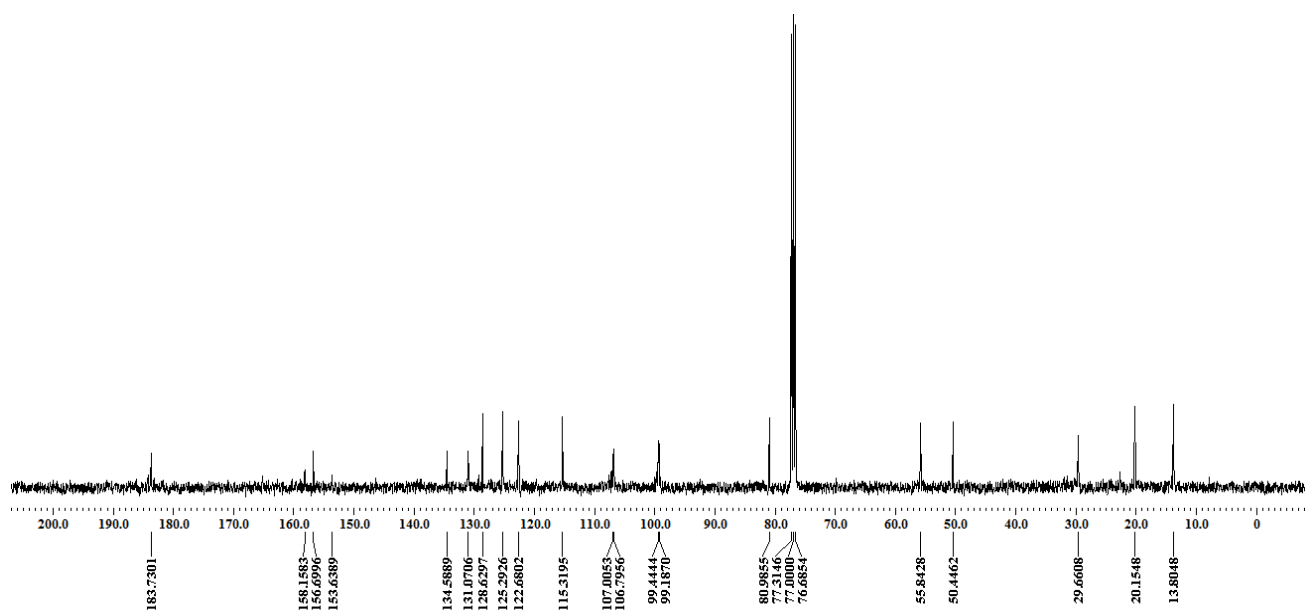
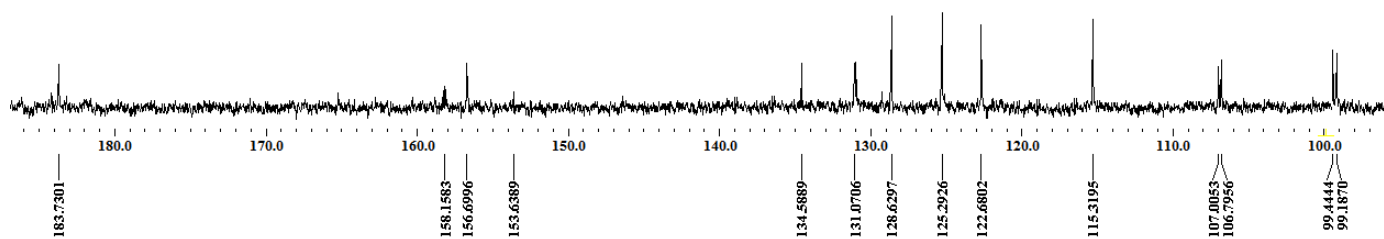
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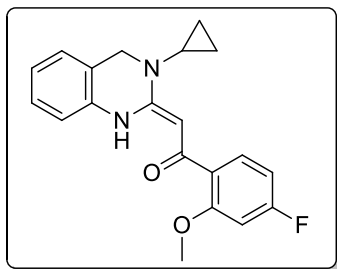




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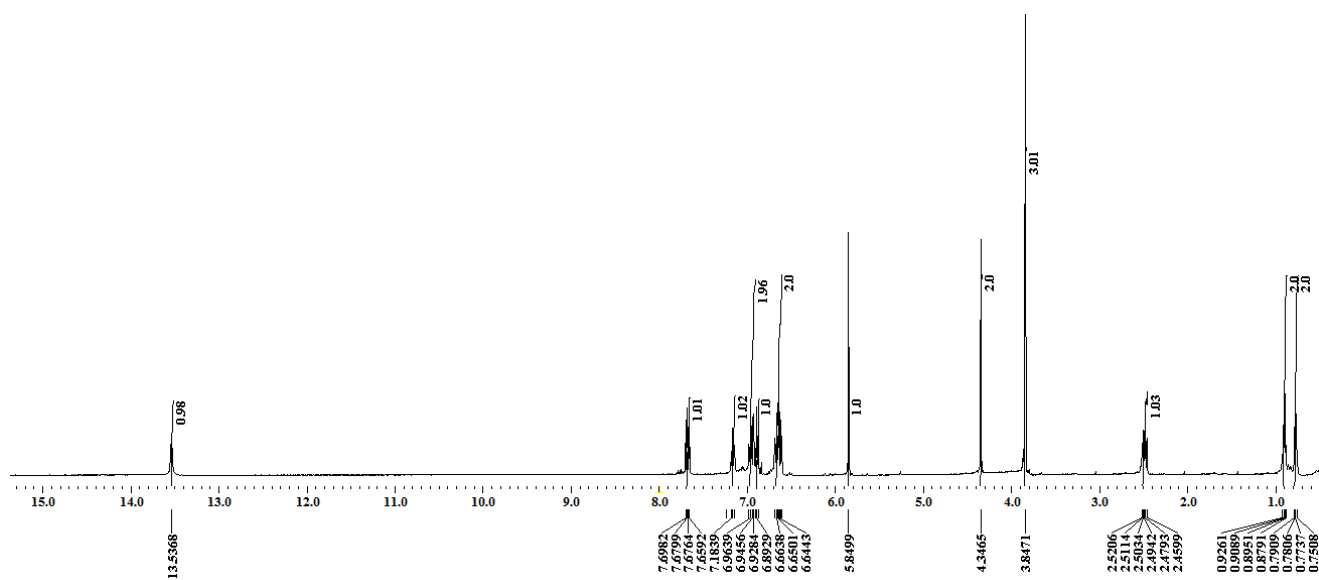
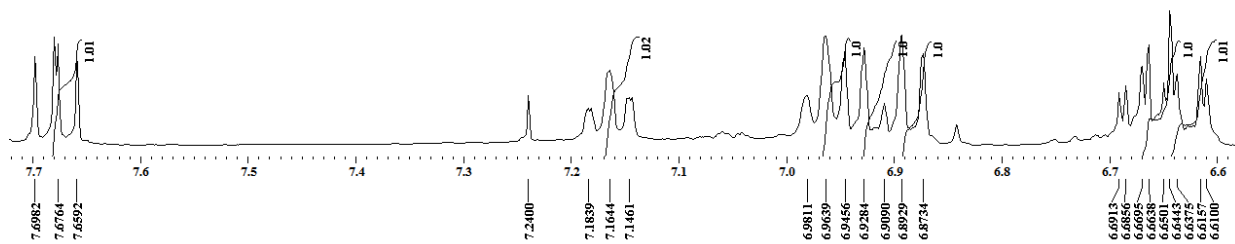
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

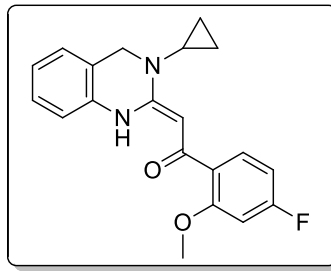




4m

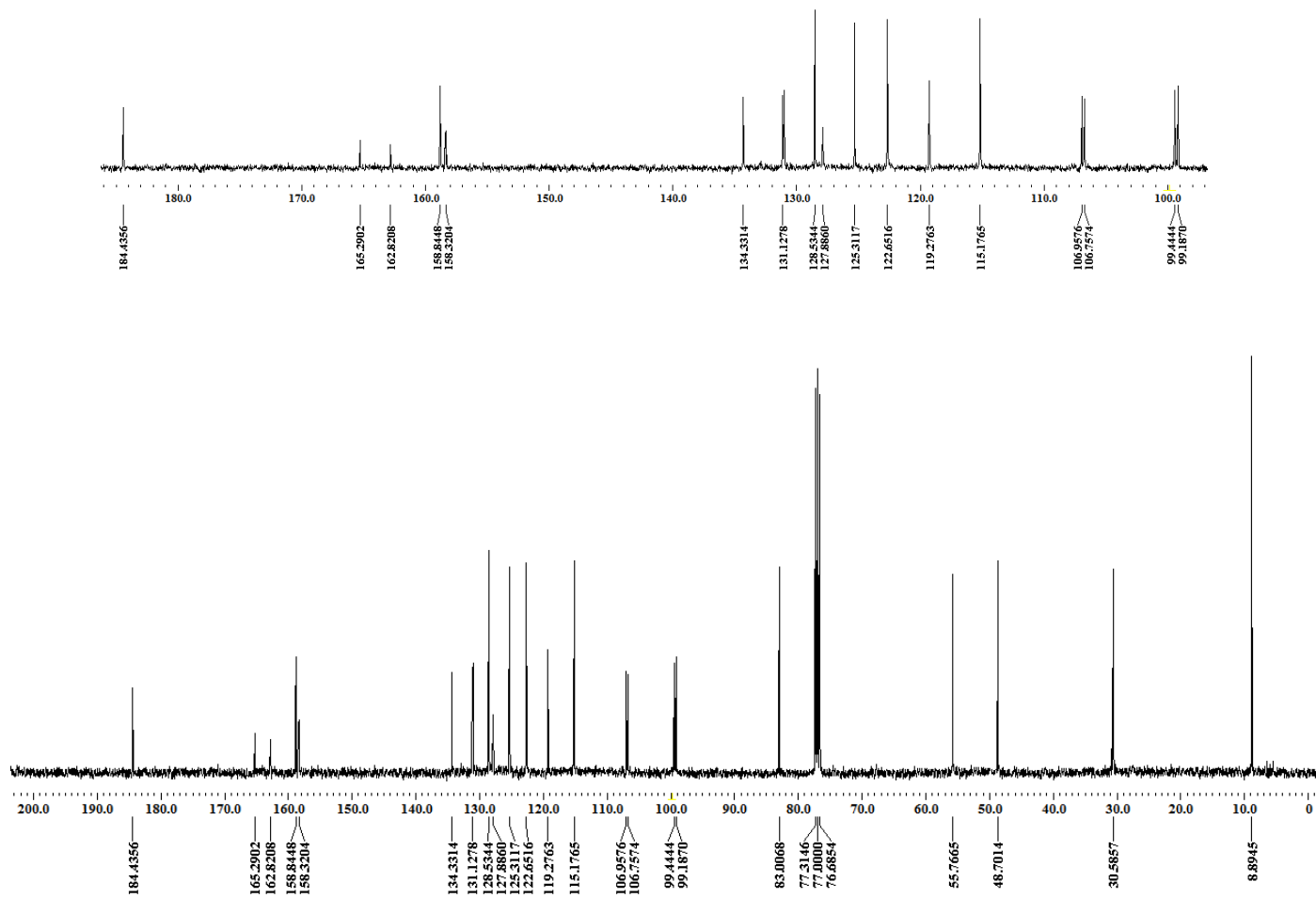
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

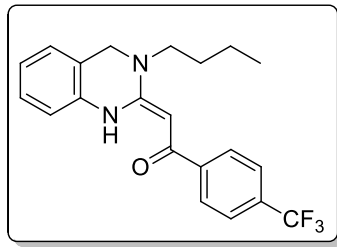




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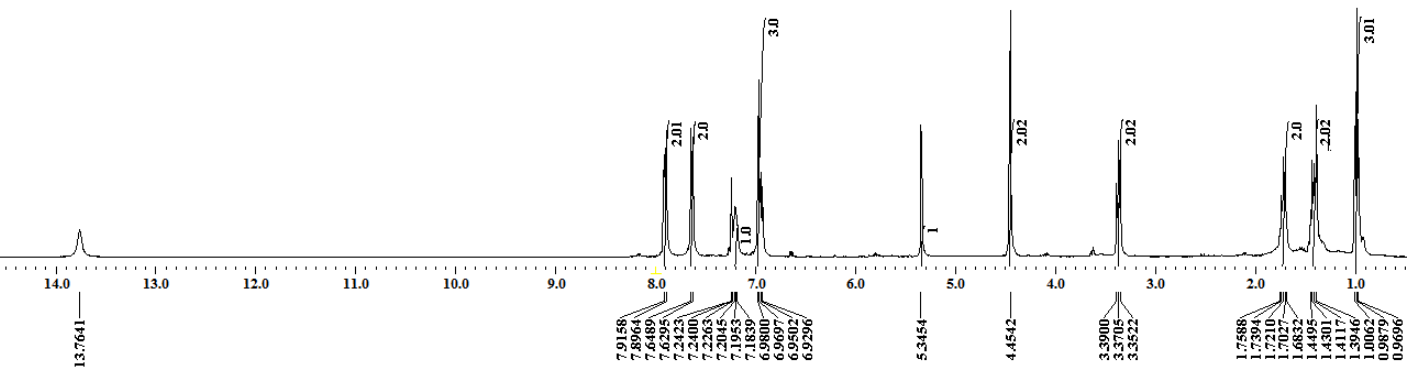
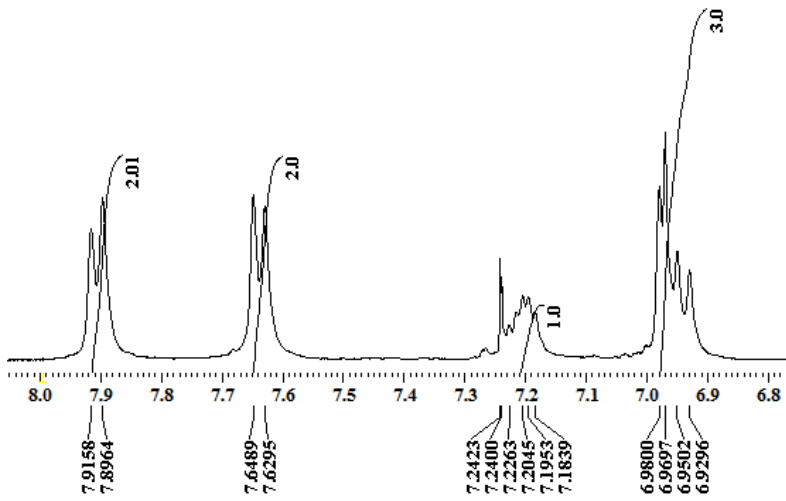
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

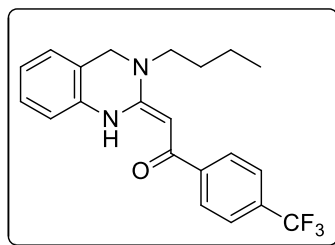




4n

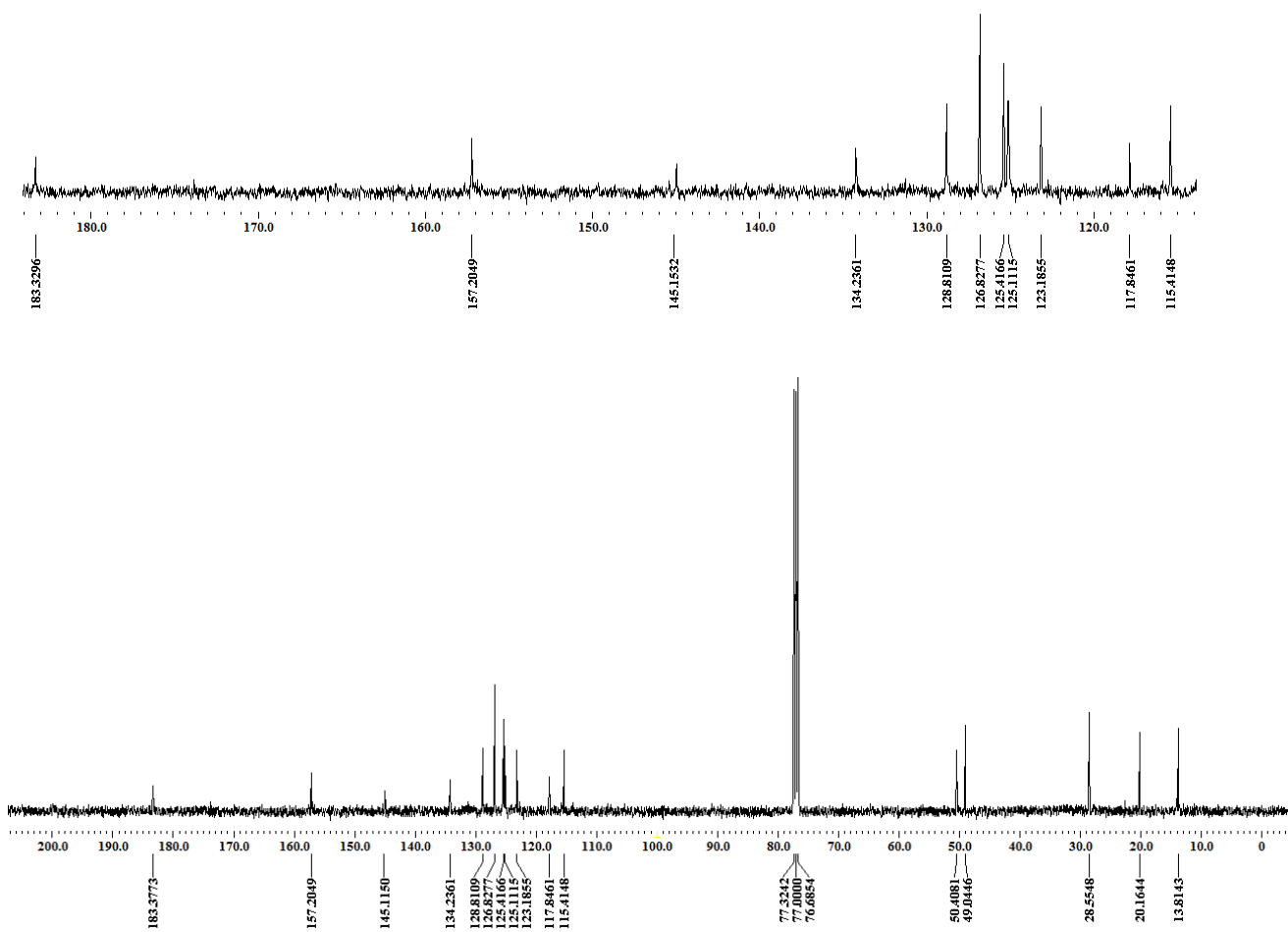
$^1\text{H-NMR}$ in CDCl_3 (400MHz)

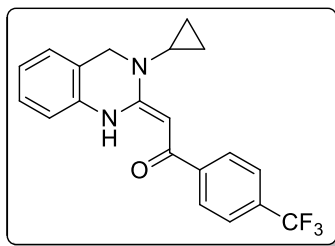




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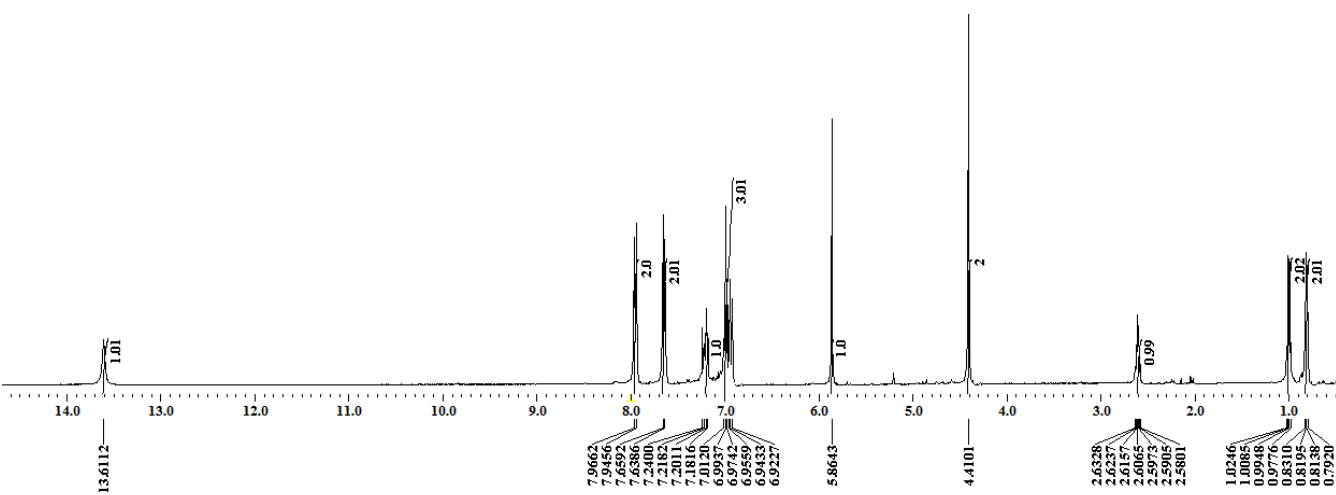
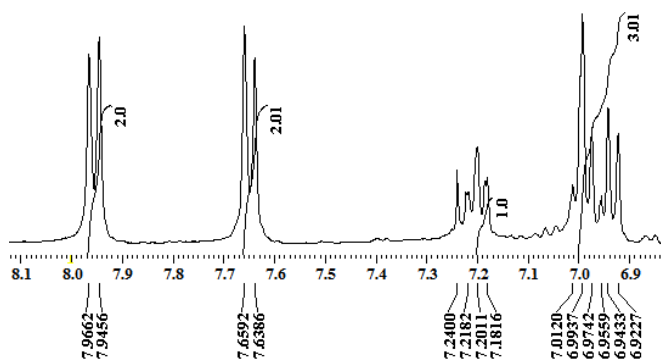
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

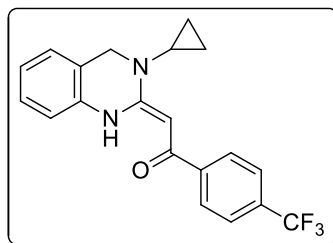




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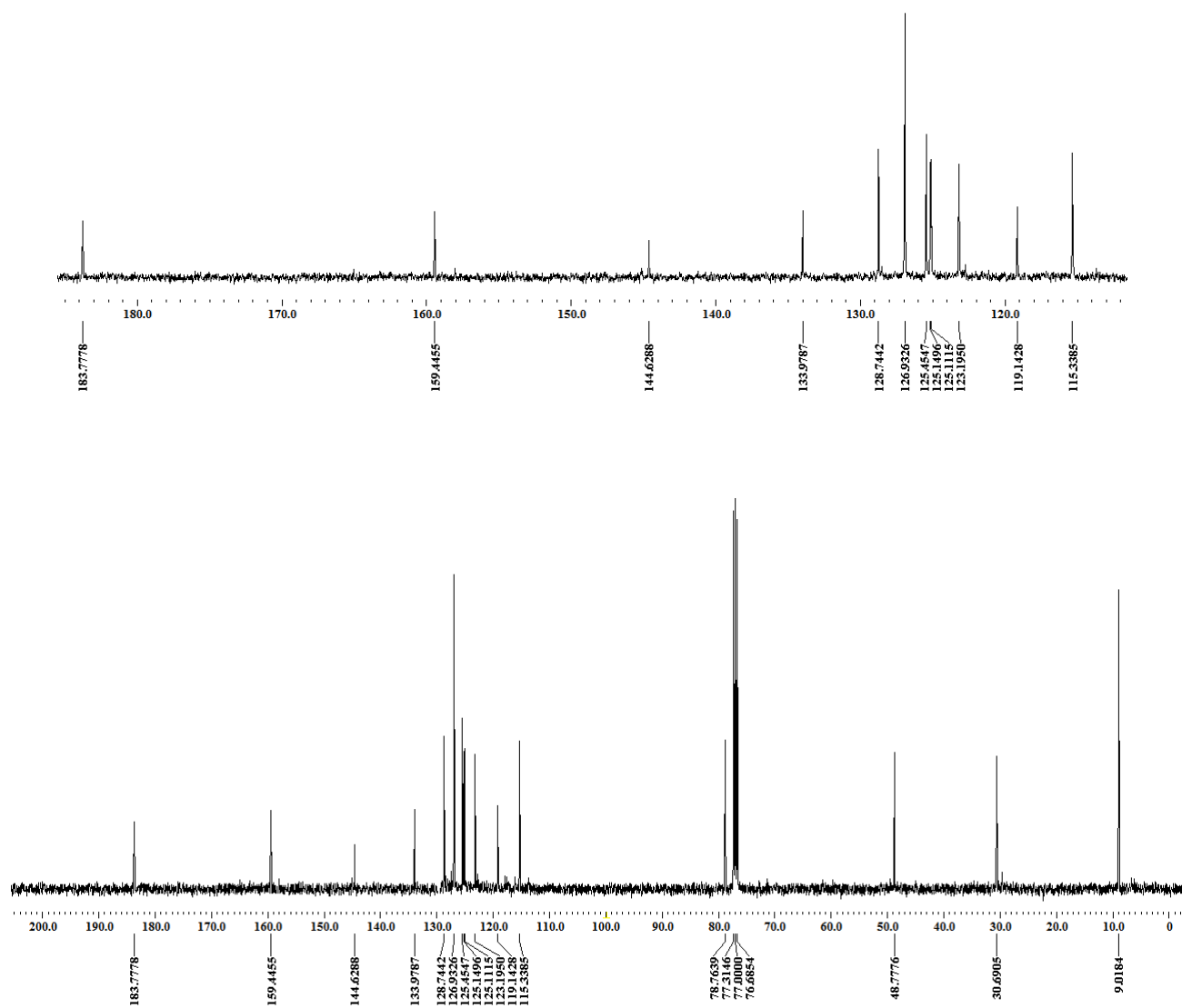
¹H-NMR in CDCl₃ (400MHz)

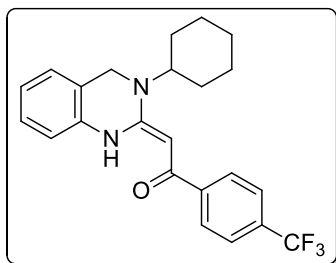




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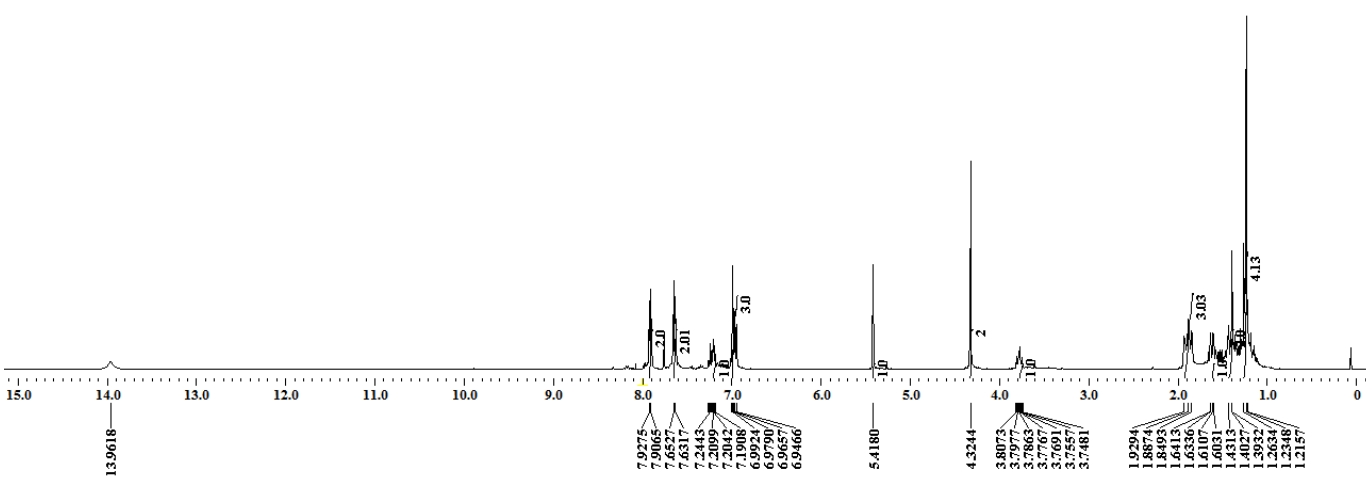
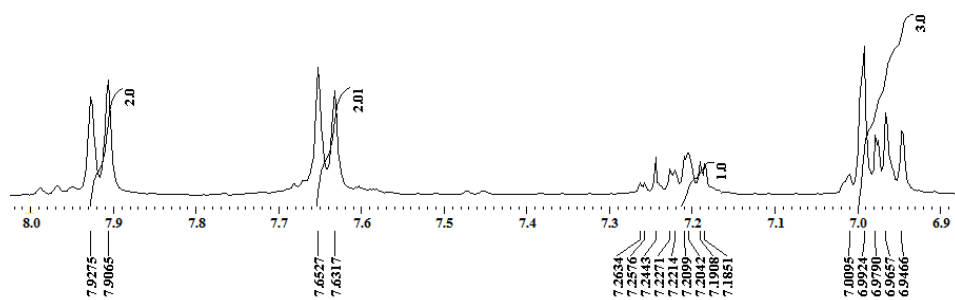
$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

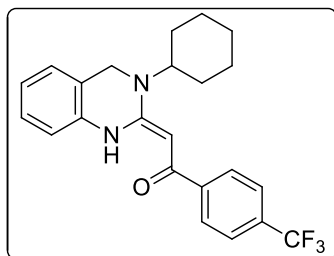




4p

¹H-NMR in CDCl₃ (400MHz)





4p

$^{13}\text{C-NMR}$ in CDCl_3 (100MHz)

