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## **SUPPORTING INFORMATION**

# **Continuous Flow Synthesis of Hydroxy Lactones from Alkenoic Acids**

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#### **General methods**

Unless otherwise noted, chemicals were obtained from commercial suppliers and used without further purification. NMR spectra were recorded on a Bruker AC 400 MHz spectrometer in the indicated solvent. Chemical shifts are reported in parts per million (ppm) and are relative to  $CDCl_3$ (7.26 ppm and 77.0 ppm) or to  $d^6$ -DMSO (2.49 ppm and 39.7 ppm). The abbreviations used are as follows: s, singlet; brs, broad singlet; d, doublet; dd, double doublet; ddd, doublet of doublet of doublet; t, triplet; g, guartet; guint, guintet; m, multiplet; brm, broad multiplet. With the exception of compounds 5f and 5l, all the synthesised substrates, the selenium catalyst and the products have been previously reported and their spectroscopic data obtained in this study are consistent with the literature data.<sup>1-10</sup> For new compounds **5f** and **5l**, high resolution mass spectra (HRMS) were recorded on Agilent 6540-UHD Accurate Mass Q-TOF LC/MS instrument and mass spectrum of the analyte extract ion chromatogram (EIC) was acquired in positive full scan mode (ESI+). A Prevail C18 column (250 x 4.6 mm i.d., 5µm) was used as analytical column for HPLC analysis with DAD detector. A mixture of H<sub>2</sub>O/CH<sub>3</sub>CN (60:40, v/v) was used as the eluent with a flow rate of 1 mL min<sup>-1</sup>. IR spectra were recorded on a Jasco FT/IR 410 instrument (KBr). All flow experiments were performed using a commercially available modular Vapourtec R serie equipped with two-loop injection systems (PTFE, 1/16" OD x 0.02" ID, 2 mL each), two integrated HPLC pumps (R2+ pumping module), two external HPLC pumps (Jasco PU-980), a mesoreactor coil (PFA, 1/16" OD x 0.02" ID, 10 mL), two back pressure regulators (BPR, 100 psi, PEEK, 1/16" OD, 1/4"-28), an in-line liquid-liquid separator SEP-10 (Zaiput, Cambridge, MA – USA), tubular mesoreactors (Omnifit Labware DIBA HIT column,  $L \times ID$  6.6 mm x 100 mm and 6.6 mm x 150 mm), 3-way hexagonal valves (PTFE, 1/4"-28, 1-4 mm OD, 1.5 mm ID, DIBA), 6-way connector (PTFE, 1/4"-28, 0.5-4 mm OD, 1.5 mm ID, DIBA) and a fraction collector. Thin-layer chromatography was performed using glass plates coated with silica gel 60 F-254. Spots were visualized by UV detector ( $\lambda$ : 254 nm) and/or by staining and worming with potassium permanganate. When required, flash chromatographic purifications were performed using Biotage Isolera™ Prime.

#### Synthesis of phenylseleninic acid (2)<sup>1</sup>



To a solution of  $(PhSe)_2$  (2.5 g, 8.01 mmol) in  $CH_2Cl_2$  (6 mL),  $H_2O_2$  (30% wt, 3.3 mL, 31.49 mmol) was added dropwise in 30 min at 0-5 °C. The yellow solution was vigorously stirred at this temperature for 8 h. The white suspension thus obtained

was filtered off and the solid was washed with cold H<sub>2</sub>O (4 mL) and cold CH<sub>2</sub>Cl<sub>2</sub> (7 mL) to afford

phenylseleninic acid (**2**) (2.63 g, 13.91 mmol, 87%) as white solid (m.p.: 124-126 °C, Lit.:<sup>1</sup> 124-125 °C). <sup>1</sup>H-NMR (*d*<sup>6</sup>-DMSO, 400 MHz): δ 7.58-7.60 (m, 3H), 7.81-7.83 (m, 2H). <sup>13</sup>C-NMR (*d*<sup>6</sup>-DMSO, 100.6 MHz): δ 126.3, 129.4, 132.1, 149.2. <sup>77</sup>Se-NMR (*d*<sup>6</sup>-DMSO, 76.3 MHz): δ 1180.40.

## Synthesis of alkenoic acids 3k-m<sup>2</sup>

To a solution of freshly distilled *N*,*N*-diisopropylamine (0.85 mL, 6 mmol) in dry THF (1 mL), 2.5 N solution of *n*-BuLi in hexane (2.4 mL, 6 mmol) was added dropwise in 15 min at -78 °C under argon atmosphere. After 1h, the mixture was warmed to -40 °C and, to this solution, a solution of the corresponding acid (1 mmol) in freshly distilled THF (2 mL) was added dropwise at -40 °C in 10 min. The resulting suspension was stirred for 1 h at r.t. and for 30 min at 60 °C. The suspension was cooled to 0 °C and allyliodide (5 mmol, 0.43 mL) was added dropwise. The mixture was stirred at 60 °C for 2 h, then poured into ice-cold water (20 mL) and extracted with Et<sub>2</sub>O (3 x 20 mL). The organic phase was acidified (pH 1) with 3N aqueous solution of HCl and extracted with Et<sub>2</sub>O (3 x 25 mL). The combined organic extracts were washed with brine (25 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude was purified by flash chromatography using Biotage Isolera<sup>m</sup> Prime (Cartridge: KP SIL 25 g, Eluent: *n*-hexane/EtOAc from 100:0 to 70:30, v/v).

## 2,2-diphenylpent-4-enoic acid (3k)<sup>2</sup>

 $\underbrace{CO_{2}H}_{Ph} \xrightarrow{CO_{2}H}_{Ph} = \frac{1}{3} \text{ (I65 mg, 0.65 mmol). White solid. }^{1}H-NMR (CDCl_{3}, 400 MHz):}{\delta 3.23 (d, J = 6.89 Hz, 2H, 3-CH_{2}), 4.97-5.01 (m, 2H, 5-CH_{2}), 5.62-5.69 (m, 1H, 4-CH), 7.31-7.37 (m, 10H, Ar). }^{13}C-NMR (CDCl_{3}, 100.6 MHz): \delta 42.3, 60.0, 118.4, 126.9, 127.7, 128.9, 133.7, 141.7, 180.4.$ 

## 1-allylcyclopentane-1-carboxylic acid (3I)<sup>2</sup>

CO<sub>2</sub>H Isolated yield: 68% (105 mg, 0.68 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): δ 1.57-1.69 (m, 6H, cyclopent), 2.09-2.13 (m, 2H, cyclopent), 2.39 (d, *J*= 7.16 Hz, 2H, CH<sub>2</sub>=CHCH<sub>2</sub>), 5.06-5.11 (m, 2H, CH<sub>2</sub>=CHCH<sub>2</sub>), 5.73-5.84 (m, 1H, CH<sub>2</sub>=CHCH<sub>2</sub>). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 24.9, 35.3, 42.4, 53.2, 117.5, 134.4, 184.4.

## 1-allylcyclohexane-1-carboxylic acid (3m)<sup>2</sup>

CO<sub>2</sub>H Isolated yield: 71% (120 mg, 0.71 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  1.25-1.31 (m, 3H, cyclohex), 1.38-1.46 (m, 2H, cyclohex), 1.56-1.60 (m,

3H, cyclohex), 2.04-2.07 (m, 2H, cyclohex), 2.30 (d, *J*= 7.23 Hz, 2H, CH<sub>2</sub>=CHCH<sub>2</sub>), 5.05-5.08 (m, 2H, CH<sub>2</sub>=CHCH<sub>2</sub>), 5.73-5.82 (m, 1H, CH<sub>2</sub>=CHCH<sub>2</sub>). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 22.8, 25.5, 33.2, 44.2, 46.9, 117.7, 133.1, 183.2.

#### Protocol and flow-set-up for reaction screening optimization

A solution of **3a** (0.6 mmol, 0.3 M, 2 mL) in EtOAc and a pre-mixed solution (ultrasound, 15 min) of **2** (0.1-0.5 equiv., 0.06-0.3 mmol, 0.03-0.15 M) and H<sub>2</sub>O<sub>2</sub> (30% wt, 5 equiv., 3 mmol, 1.5 M) in H<sub>2</sub>O/acetone (5:1 v/v, 2 mL) were injected into the loops and pumped with a flow rate of 0.1 or 0.15 mL min<sup>-1</sup> for each pump ( $\tau$ = 50 or 33 min). After the injection and switching of the valves into the loops, the solutions were mixed together in a T-junction and flowed through the reactor-coil thermostated (PFA, 1/16" OD x 0.02" ID, 10 mL) at 25 °C. The reactor output was collected, quenched with 10% (w/v) aqueous solution of NaHSO<sub>3</sub> (1 mL) and extracted with EtOAc (4 x 10 mL). The organic phase was washed with aqueous saturated solution of NaHCO<sub>3</sub> (20 mL), brine (20 mL), dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, concentrated under reduced pressure (<40 °C) and analyzed by <sup>1</sup>H-NMR.

### Protocol and flow-set-up for substrate scope

A solution of alkenoic acid **3a-m** (0.6 mmol, 0.3 M, 2 mL) in EtOAc and a pre-mixed solution (ultrasound, 15 min) of **2** (0.3 equiv., 0.18 mmol, 0.09 M),  $H_2O_2$  (30% wt, 5 equiv., 3 mmol, 1.5 M) and HCO<sub>2</sub>H (0.1 equiv., 0.06 mmol, 0.03 M) in H<sub>2</sub>O/acetone (5:1 v/v, 2 mL) were injected into the loops and pumped with a flow rate of 0.1 mL min<sup>-1</sup> for each pump ( $\tau$ = 50 min). After the injection and switching of the valves into the loops, the solutions were mixed together in a T-juction and flowed through the reactor-coil thermostated at 25 °C (PFA, 1/16" OD x 0.02" ID, 10 mL). The reactor output was collected and quenched with an in-line stream of NaHSO<sub>3</sub> (10% w/v aqueous solution, 1 mL). After in-line phase separation (liquid-liquid separator SEP-10, Zaiput, Cambridge, MA – USA), the organic phase was flowed with a flow rate of 0.5 mL min<sup>-1</sup> through a tubular mesoreactor (*Omnifit Labware DIBA HIT column*, L × ID 6.6 mm x 100 mm) packed with alternate layers of Amberlyst A21/silica (2:1 w/w, 4 layers, 300 mg for each layer, Fig. S1). The output was collected, dried over anhydrous Na<sub>2</sub>SO<sub>4</sub>, concentrated under reduce pressure (<40 °C) and directly characterized by <sup>1</sup>H-, <sup>13</sup>C-, and 2D-NMR analysis. The separation of the diastereomeric mixtures (Table 2, entries 5 and 8) was performed by automated flash chromatography (Biotage Isolera<sup>TM</sup> Prime, Cartridge: SNAP KP-SIL 10 g, Eluent: *n*-hexane/EtOAc, from 100:0 to 40:60, v/v).



Fig. S1. Omnifit Labware DIBA HIT column for catalyst scavenging.

## Compounds 5a-m characterization

#### 4-Hydroxy-5-methyl-dihydrofuran-2(3H)-one (5a)<sup>3</sup>

Isolated yield: 85% (59 mg, 0.51 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  1.38 (d, J= 6.62 Hz, 3H, 5-CHCH<sub>3</sub>), 2.24 (brs, 1H, OH), 2.53 (dd, J<sub>1</sub>= 3.71 Hz, J<sub>2</sub>= 17.72 Hz, 1H, 3-CH<sub>2(A)</sub>), 2.86 (dd, J<sub>1</sub>= 6.49 Hz, J<sub>2</sub>= 17.94 Hz, 1H, 3-CH<sub>2(B)</sub>), 4.23-4.27 (m, 1H, 4-CH), 4.51 (dq, J<sub>1</sub>= 2.94 Hz, J<sub>2</sub>= 6.62 Hz, 1H, 5-CH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz):  $\delta$  18.5, 37.3, 72.8, 84.0, 175.1.

### 5-Hydroxymethyl-dihydrofuran-2(3H)-one (5b)<sup>4</sup>

Isolated yield: 80% (56 mg, 0.48 mmol). Colorless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): OH  $\delta$  2.18-2.20 (m, 1H, 4-*CH*<sub>2(A)</sub>), 2.28-2.30 (m, 1H, 4-*CH*<sub>2(B)</sub>), 2.57-2.71 (m, 2H, 3-*CH*<sub>2</sub>), 3.67 (dd, J<sub>1</sub>= 4.57 Hz, J<sub>2</sub>= 12.54 Hz, 1H, 5-CH*CH*<sub>2(A)</sub>), 3.92 (dd, J<sub>1</sub>= 2.72 Hz,

*J*<sub>2</sub>= 12.54 Hz, 1H, 5-CH*CH*<sub>2(B)</sub>), 4.64-4.68 (m, 1H, 5-*CH*). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 23.1, 28.7, 64.0, 80.8, 177.8.

## 6-Hydroxymethyl-tetrahydro-2H-pyran-2-one (5c)<sup>4</sup>

Isolated yield: 74% (57 mg, 0.44 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  1.71-1.96 (m, 4H, 3-*CH*<sub>2</sub> + 4-*CH*<sub>2</sub>), 2.42-2.48 (m, 1H, 5-*CH*<sub>2(A)</sub>), 2.57-2.62 (m, 1H, 5-*CH*<sub>2(B)</sub>), 3.10 (brs, 1H, OH), 3.65 (dd, J<sub>1</sub>= 5.45 Hz, J<sub>2</sub>= 12.30 Hz, 1H, 6-

CH*CH*<sub>2(A)</sub>), 3.76 (dd, *J*<sub>1</sub>= 3.17 Hz, *J*<sub>2</sub>= 12.30 Hz, 1H, 6-CH*CH*<sub>2(B)</sub>), 4.37-4.43 (m, 1H, 6-*CH*). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 18.2, 23.5, 29.5, 64.6, 81.2, 171.9.

### 4-Hydroxy-dihydrofuran-2(3H)-one (3-BHL) (5d)<sup>4</sup>



Isolated yield: 79% (48 mg, 0.47 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  2.37 (brs, 1H, OH), 2.54 (dd,  $J_1$ = 1.52 Hz,  $J_2$ = 17.86 Hz, 1H, 3- $CH_{2(A)}$ ), 2.77 (dd,  $J_1$ = 6.04 Hz,  $J_2$ = 17.95 Hz, 1H, 3- $CH_{2(B)}$ ), 4.32 (d, J= 10.26 Hz, 1H, 5- $CH_{2(A)}$ ), 4.44 (dd,  $J_1$ =

4.46 Hz, *J*<sub>2</sub>= 10.28 Hz, 1H, 5-*CH*<sub>2(B)</sub>), 4.69-4.72 (m, 1H, 4-*CH*). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 37.8, 67.4, 76.2, 176.8.

### 4-Hydroxy-5-phenyl-dihydrofuran-2(3H)-one (5e)<sup>5</sup>

OH Isolated yield: 71% (76 mg, 0.43 mmol). Pale yellow oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  2.60 (dd,  $J_1$ = 4.26 Hz,  $J_2$ = 17.81 Hz, 1H, 3-C $H_{2(A)}$ ), 2.87 (dd,  $J_1$ = 6.51 Hz,  $J_2$ = 17.82 Hz, 1H, 3-C $H_{2(B)}$ ), 3.14 (brs, 1H, OH), 4.49 (ddd,  $J_1$ = 3.20 Hz,  $J_2$ = 3.98 Hz,  $J_3$ = 6.42 Hz, 1H, 4-CH), 5.39 (d, J= 3.13 Hz, 1H, 5-CH), 7.27-7.42 (m, 5H, Ar). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz):  $\delta$  36.9, 74.4, 88.0, 125.0, 128.7, 128.9, 136.7, 175.4.

### 3a-Hydroxy-hexahydrobenzofuran-2(3H)-one (5f)

HO  $O = \left( \begin{array}{c} HO\\ H \end{array} \right)$ Isolated yield: 92% (86 mg, 0.55 mmol). Pale yellow oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  1.40-1.44 (m, 2H, 4-CH<sub>2</sub>), 1.60-1.71 (m, 4H, 5-CH<sub>2</sub> + 6-CH<sub>2</sub>), 1.92-2.14 (m, 2H, 7-CH<sub>2</sub>), 2.49-2.70 (m, 2H, 3-CH<sub>2</sub>), 4.31 (t, J= 6.59 Hz, 1H, 7a-CH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz):  $\delta$  20.8, 21.5, 28.4, 34.3, 41.8, 74.7, 83.9, 174.9. FT-IR (T%): v 2932.23 (20.01), 2857.99 (26.42), 2662.25 (64.91), 1759.24 (17.21), 1454.55 (27.02), 1353.78 (28.18), 1248.20 (28.05), 1213.49 (23.88), 1173.95 (21.15), 1062.59 (24.61), 1009.55 (20.05), 948.81 (28.47), 888.06 (28.57), 842.74 (34.29). HRMS (ESI+) m/z [M+H]<sup>+</sup> calcd for C<sub>8</sub>H<sub>12</sub>O<sub>3</sub> 156.07864; found: 156.07813;  $\Delta$ = 3.32 ppm; residence time (t<sub>R</sub>): 2.055 min.

## 6-Hydroxy-hexahydro-2H-cyclopenta[b]furan-2-one (5g)<sup>4</sup>



Isolated yield: 76% (65 mg, 0.46 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  1.48-1.54 (m, 1H, 4-*CH*<sub>2(A)</sub>), 1.72-1.75 (m, 1H, 5-*CH*<sub>2(A)</sub>), 1.85-1.88 (m, 1H, 5-*CH*<sub>2(B)</sub>), 2.19-2.21 (m, 1H, 4-*CH*<sub>2(B)</sub>), 2.25 (dd, *J*<sub>1</sub>= 2.87 Hz, *J*<sub>2</sub>= 18.61 Hz, 1H, 3-*CH*<sub>2(A)</sub>), 2.59 (brs, 1H, *OH*), 2.81 (dd, *J*<sub>1</sub>= 10.38 Hz, *J*<sub>2</sub>= 18.56 Hz, 1H, 3-*CH*<sub>2(B)</sub>), 3.00-

3.08 (m, 1H, 3a-*CH*), 4.33 (brs, 1H, 6-*CH*), 4.72 (d, *J*= 6.90 Hz, 1H, 6a-*CH*). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 32.2, 33.0, 37.0, 37.6, 77.6, 91.8, 179.1.

#### 5-(1-hydroxyethyl)Dihydrofuran-2(3H)-one (5h, mixture anti:syn 80:20)<sup>6</sup>



Isolated yield: 78% (60 mg, 0.46 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): δ 1.18 (d, *J*= 6.46, 3H, *CH*<sub>3</sub>, *anti*), 1.25 (d, *J*= 6.08, 3H, *CH*<sub>3</sub>, *syn*), 2.02-2.08 (m, 2H, 4-*CH*<sub>2</sub>, *syn*), 2.12-2.30 (m,

2H, 4-CH<sub>2</sub>, anti), 2.47-2.64 (m, 4H, 2 x 3-CH<sub>2</sub>, anti + syn), 2.73 (brs, 2H, 2 x OH, anti + syn), 3.78 (quint, *J*= 6.12 Hz, 1H, CH(CH<sub>3</sub>)OH, syn), 4.10-4.12 (m, 1H, CH(CH<sub>3</sub>)OH, anti), 4.35 (q, *J*= 6.90 Hz, 1H, 5-CH, syn), 4.38-4.41 (m, 1H, 5-CH, anti). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 16.9 (anti), 17.7 (syn), 20.2 (anti), 23.3 (syn), 27.9 (anti + syn), 66.5 (anti), 69.0 (syn), 82.9 (anti), 83.5 (syn), 176.9 (syn), 177.0 (anti).

### 7-(hydroxymethyl)Oxepan-2-one (5i)<sup>7</sup>



Isolated yield: 67% (58 mg, 0.40 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  1.44-1.69 (m, 6H, 4-CH<sub>2</sub> + 5-CH<sub>2</sub> + 6-CH<sub>2</sub>), 2.38 (t, *J*= 6.98 Hz, 2H, 3- CH<sub>2</sub>), 3.47-3.51 (m, 1H, 7-CHCH<sub>2(B)</sub>), 3.62-6.67 (m, 1H, 7-CHCH<sub>2(A)</sub>), 3.83 (brs, 1H,

OH), 4.30-4.34 (m, 1H, 7-CH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 24.4, 25.0, 31.3, 33.8, 64.9, 78.2, 179.2.

## 5-(1-hydroxyhexyl)Dihydrofuran-2(3H)-one (5j)<sup>8</sup>



Isolated yield: 82% (92 mg, 0.49 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): δ 0.87 (t, *J*= 6.82 Hz, 3H, CH<sub>2</sub>(CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>), 1.22-1.40 (m, 6H, CH<sub>2</sub>(CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>), 1.50-1.51 (m, 2H, CH<sub>2</sub>(CH<sub>2</sub>)<sub>3</sub>CH<sub>3</sub>), 2.08-2.14 (m, 1H, 4-

CH<sub>2(A)</sub>), 2.23-2.28 (m, 1H, 4-CH<sub>2(B)</sub>), 2.48-2.58 (m, 2H, 3-CH<sub>2</sub>), 2.76-2.96 (brs, 1H, OH), 3.90-3.93 (m, 1H, 5-CHCH(OH)), 4.40-4.45 (m, 1H, 5-CH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 13.9, 20.9, 22.4, 25.2, 28.7, 31.6, 31.8, 71.2, 83.0, 178.0.

## 5-(hydroxymethyl)-3,3-Diphenyldihydrofuran-2(3H)-one (5k)<sup>9</sup>



Isolated yield: 78% (125 mg, 0.47 mmol). Pale yellow oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz):  $\delta$  2.28-2.32 (m, 1H, 4-*CH*<sub>2(A)</sub>), 2.86-2.99 (m, 1H, 4-*CH*<sub>2(B)</sub>), 3.68 (dd, *J*<sub>1</sub>= 5.66 Hz, *J*<sub>2</sub>= 12.34 Hz, 1H, 5-CH*CH*<sub>2(A)</sub>), 3.99 (dd, *J*<sub>1</sub>= 2.78 Hz, *J*<sub>2</sub>= 12.38 Hz, 1H, 5-

CH*CH*<sub>2(B)</sub>), 4.44-4.56 (m, 1H, 5-*CH*), 7.24-7.38 (m, 10H, Ar). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 46.4, 60.2, 64.8, 82.9, 126.4, 127.4, 127.6, 134.6, 136.8, 172.2.

#### 3-(hydroxymethyl)-2-Oxaspiro[4.4]nonan-1-one (5l)



Isolated yield: 77% (79 mg, 0.46 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz): δ 1.66-2.03 (m, 8H, cyclopent), 2.08-2.19 (m, 2H, 4-CH<sub>2</sub>), 2.73 (brs, 1H, OH), 3.64 (dd, J<sub>1</sub>= 5.28 Hz, J<sub>2</sub>= 12.57 Hz, 1H, 3-CHCH<sub>2(A)</sub>), 3.89 (dd, J<sub>1</sub>= 2.83 Hz,

 $J_2$ = 12.58 Hz, 1H, 3-CHCH<sub>2(B)</sub>), 4.51-4.55 (m, 1H, 3-CH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz):  $\delta$  25.3, 25.4, 36.8, 37.79, 37.82, 50.1, 63.8, 78.0, 182.6. FT-IR (T%): v 2957.30 (46.70), 2871.97 (53.78), 1757.80 (33.37), 1447.80 (58.78), 1354.27 (61.03), 1188.90 (45.23), 1055.35 (47.49), 980.63 (59.20), 913.61 (66.95). HRMS (ESI+) m/z  $[M+H]^+$  calcd for C<sub>8</sub>H<sub>12</sub>O<sub>3</sub> 170.09429; found: 170.09408;  $\Delta$ = 1.26 ppm; residence time  $(t_R)$ : 2.663 min.

### 3-(hydroxymethyl)-2-Oxaspiro[4.5]decan-1-one (5m)<sup>10</sup>

Isolated yield: 76% (85 mg, 0.46 mmol). Colourless oil. <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 OH

MHz): δ 1.25-1.37 (m, 3H, cyclohex), 1.53-1.65 (m, 4H, cyclohex), 1.72-7.77 (m, 3H, cyclohex), 1.89 (dd, J1= 9.42 Hz, J2= 12.86 Hz, 1H, 4-CH2(A)), 2.25 (dd, J1= 6.92 Hz,  $J_2$ = 12.92 Hz, 1H, 4-CH<sub>2(B)</sub>), 2.69 (brs, 1H, OH), 3.62 (dd,  $J_1$ = 5.10 Hz,  $J_2$ = 12.57 Hz, 1H, 3-CHCH<sub>2(A)</sub>), 3.90 (dd,  $J_1$ = 2.78 Hz,  $J_2$ = 12.57 Hz, 1H, 3-CHCH<sub>2(B)</sub>), 4.52-4.57 (m, 1H, 3-CH). <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz): δ 21.99, 22.0, 32.0, 33.8, 34.0, 44.8, 63.9, 77.6, 181.8.

## Scale-up synthesis of 4-hydroxy-dihydrofuran-2(3H)-one (3-BHL) (5d)

100 mL of a solution of vinylacetic acid (3d, 30 mmol, 2.55 mL, 0.3 M) in EtOAc and 100 mL of a pre-mixed solution of phenylseleninic acid (2, 0.30 equiv., 9 mmol, 1.7 g), H<sub>2</sub>O<sub>2</sub> (30% w/w aqueous solution, 5 equiv., 150 mmol, 15.32 mL) and HCO<sub>2</sub>H (0.1 equiv., 3 mmol, 120 µL) in H<sub>2</sub>O/acetone (5:1, v/v) were pumped with a flow rate of 0.4 mL min<sup>-1</sup> for each pump and splitted with a 5-way connector (PTFE, 1/4"-28, 0.5-4 mm OD, 1.5 mm ID, DIBA) through four mesoreactor-coil (PFA, 1/16" OD x 0.02" ID, 10 mL) operating in parallel and thermostated at 25 °C ( $\tau$ = 50 min for each reactor). After the first 9 mL of dead volume (45 min), the reactor outputs were combined together and collected for 5 h (V= 220 mL). After in-line quenching with 10% w/v aqueous solution of NaHSO<sub>3</sub> (20 mL) and phase separation (liquid-liquid separator SEP-10, Zaiput, Cambridge, MA – USA), the organic phase was flowed through a tubular mesoreactor (Omnifit Labware DIBA HIT column, L × ID 6.6 mm x 150 mm) packed with alternate layers of Amberlyst A21/silica (2:1 w/w, 4 layers, 450 mg for each layer, loading: 1.3 meq/L corresponding to a total of ~ 6 mmol) with a flow rate of 0.5 mL min<sup>-1</sup> ( $\tau$ = 10 min). After 3.5 h, the valve was switched to directed the flow stream towards a second column. Meanwhile the first A21 packed column was regenerated by flowing a 5% (v/v) ethanolic solution of NH<sub>4</sub>OH (50 mL, 0.5 mL min<sup>-1</sup>). The output was collected for 4 h with a fractions collector affording 4-hydroxy-dihydrofuran-2(3*H*)-one (**5d**) (2.25 g, 22.04 mmol, 74% yield, productivity: 2.45 mmol h<sup>-1</sup>, 6 g d<sup>-1</sup>) as colorless oil. For the catalyst recovery, the column was washed with 5% (v/v) ethanolic solution of NH<sub>4</sub>OH (50 mL, 0.5 mL min<sup>-1</sup>). The basic solution was acidified with an in-line stream of 3 N aqueous solution of HCl (50 mL, 0.5 mL min<sup>-1</sup>) and extracted in-line with a stream of CH<sub>2</sub>Cl<sub>2</sub>. The organic phase was collected affording pure **2** (1.5 g, 7.93 mmol, 88% recovery) as a white solid.

# <sup>1</sup>H-NMR (*d*<sup>6</sup>-DMSO, 400 MHz) of phenylseleninic acid (2)





# $^{13}\text{C-NMR}$ (d<sup>6</sup>-DMSO, 100.6 MHz) of phenylseleninic acid (2)



 $^{77}$ Se-NMR (*d*<sup>6</sup>-DMSO, 76.3 MHz) of phenylseleninic acid (2)



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 2,2-diphenylpent-4-enoic acid (3k)



# <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 2,2-diphenylpent-4-enoic acid (3k)





# <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 1-allylcyclopentane-1-carboxylic acid (3l)







# <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 1-allylcyclohexane-1-carboxylic acid (3m)

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 4-hydroxy-5-methyl-dihydrofuran-2(3*H*)-one (5a) before catalyst scavenging



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 4-hydroxy-5-methyl-dihydrofuran-2(3*H*)-one (5a) after catalyst scavenging



<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 4-hydroxy-5-methyl-dihydrofuran-2(3*H*)-one (5a) after catalyst scavenging









# <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 5-hydroxymethyl-dihydrofuran-2(3*H*)-one (5b)







## <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 6-hydroxymethyl-tetrahydro-2*H*-pyran-2-one (5c)







## <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 4-hydroxy-dihydrofuran-2(3*H*)-one (3-BHL) (5d)

## <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 4-hydroxy-dihydrofuran-2(3*H*)-one (3-BHL) (5d): scale up synthesis



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<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 4-hydroxy-dihydrofuran-2(3*H*)-one (5d): scale up synthesis



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 4-hydroxy-5-phenyl-dihydrofuran-2(3*H*)-one (5e)









<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 3a-hydroxy-hexahydrobenzofuran-2(3*H*)-one (5f)



## <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 3a-hydroxy-hexahydrobenzofuran-2(3*H*)-one (5f)

## HRMS (ESI) of 3a-hydroxy-hexahydrobenzofuran-2(3H)-one (5f)













<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 6-hydroxy-hexahydro-2*H*-cyclopenta[*b*]furan-2-one (5g)









<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 5-(1-hydroxyethyl)dihydrofuran-2(3*H*)-one (5h, mixture *anti:syn* 80:20)



<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 5-(1-hydroxyethyl)dihydrofuran-2(3*H*)-one (5*h*, mixture *anti:syn* 80:20)







## <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 7-(hydroxymethyl)oxepan-2-one (5i)

<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 5-(1-hydroxyhexyl)dihydrofuran-2(3*H*)-one (5j)











## <sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 3-(hydroxymethyl)-2-oxaspiro[4.4]nonan-1-one (5l)



## <sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 3-(hydroxymethyl)-2-oxaspiro[4.4]nonan-1-one (5l)

## HRMS (ESI) of 3-(hydroxymethyl)-2-oxaspiro[4.4]nonan-1-one (5I)







Peak At (Cm <sup>-1</sup> )	%Т
2957.30	46.70
2871.97	53.78
1757.80	33.37
1447.80	58.78
1354.27	61.03
1188.90	45.23
1055.35	47.49
980.63	59.20
913.61	66.95



<sup>1</sup>H-NMR (CDCl<sub>3</sub>, 400 MHz) of 3-(hydroxymethyl)-2-oxaspiro[4.5]decan-1-one (5m)



<sup>13</sup>C-NMR (CDCl<sub>3</sub>, 100.6 MHz) of 3-(hydroxymethyl)-2-oxaspiro[4.5]decan-1-one (5m)

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