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Rh-Catalyzed Regiodivergent Hydrosilylation of Acyl aminocyclopropanes Controlled by Monophosphine Ligands (Kondo, Itami, Yamaguchi)

Supplementary Information

Rh-Catalyzed Regiodivergent Hydrosilylation of Acyl aminocyclopropanes Controlled by Monophosphine Ligands

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1. General

Unless otherwise noted, all reactants or reagents including dry solvents were obtained from commercial 1,4-Bis(diphenylphosphino)butane suppliers used as received. (dppb), 1,1,1,3,5,5,5 heptamethyltrisiloxane (2a) and dimethyl(phenyl)silane (2e) were obtained from TCI Chemical. Triphenylphosphine, tricyclohexylphosphonium tetrafluoroborate $(PCy_3 \cdot HBF_4)$ butyldimethylsilane (2b) were obtained from Wako Chemicals. 1,10-Phenanthroline (phen), dicyclohexylphenylphosphine (PCy₂Ph), tri(naphthalen-1-yl)phosphine (P(1-nap)₃), tripropylsilane (2c), diethylmethylsilane (2d) and [Rh(cod)OMe]₂ were obtained from Sigma-Aldrich. [Rh(cod)Cl]₂ was prepared by according to a procedure reported in the literature.^[1] Unless otherwise noted, all reactions were performed with dry solvents under an atmosphere of nitrogen in dried glassware using standard vacuum-line techniques. All hydrosilylation reactions were performed in 20-mL glass vessel tubes equipped with a J. Young[®] O-ring tap and heated in an 8-well reaction block (heater + magnetic stirrer). All work-up and purification procedures were carried out with reagent-grade solvents in air.

Analytical thin-layer chromatography (TLC) was performed using E. Merck silica gel 60 F₂₅₄ precoated plates (0.25 mm). The developed chromatogram was analyzed by UV lamp (254 nm) or a phosphomolybdic acid/sulfuric acid solution. Flash column chromatography was performed with E. Merck silica gel 60 (230-400 mesh) or Biotage Isolera® equipped with Biotage SNAP Cartridge KP-Sil columns using hexane/ethyl acetate as eluent. Medium-pressure liquid chromatography (MPLC) was performed using Yamazen W-prep 2XY. Preparative thin-layer chromatography (PTLC) was performed using Wakogel B5-F silica coated plates (0.75 mm) prepared in our laboratory. Preparative gel permeation chromatography (GPC) was performed with a JAI LC-9204 instrument equipped with JAIGEL-1H/JAIGEL-2H columns using chloroform as eluent. LCMS analysis was conducted on an Agilent 6100 instrument equipped with Poroshell 120 EC-C18 column (2.1x100 nm, 2.7 um) using acetonitrile/5 mM HCOONH4 in water as eluent. High-resolution mass spectra (HRMS) were obtained from Thermo Fisher Scientific Exactive (ESI and DART). Nuclear magnetic resonance (NMR) spectra were recorded on a JEOL JNM-ECA-600 spectrometer (1H 600 MHz, 13C 151 MHz), a JEOL JNM-ECA-500 spectrometer (1H 500 MHz, 13C 126 MHz) and a JEOL JNM-ECA-400 spectrometer (1H 400 MHz, ¹³C 101 MHz). Chemical shifts for ¹H NMR are expressed in parts per million (ppm) relative to tetramethylsilane (δ 0.00 ppm) or residual peak of DMSO (δ 2.50 ppm) or CH₂Cl₂ (δ 5.32 ppm). Chemical shifts for ¹³C NMR are expressed in ppm relative to CDCl₃ (δ 77.0 ppm) or DMSO (δ 39.5 ppm) or CD_2Cl_2 (δ 53.84 ppm). Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, dd = doublet of doublets, t = triplet, dt = doublet of triplets, td = triplet of doublets, q = quartet, quin = quintet, sext = sextet, m = multiplet, brs = broad singlet), coupling constant (Hz), and integration.

^[1] Uson, R.; Oro, L. A.; Cabeza, J. A.; Bryndza, H. E.; Stepro, M. P. Inorg. Synth. 1985, 23, 126.

2. Preparation of Substituted Aminocyclopropanes

Note: N-Cyclopropylpivalamide (1A),^[2] *N*-cyclopropylcyclohexanecarboxamide (1C),^[2] *N*-cyclopropylisobutyramide (1D),^[2] *tert*-butyl cyclopropylcarbamate (1G),^[2] *N*-cyclopropyl-4-methylbenzamide (1I),^[3] and *N*-cyclopropyl-4-methoxybenzamide (1J)^[4] were synthesized according to procedures reported in the literature.

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N-Cyclopropyl-1-methylcyclohexane-1-carboxamide (1B): To a solution of 1-methylcyclohexane-1-carboxylic acid (1.1 g, 9.5 mmol) and *N*,*N*-dimethylformamide (DMF: 0.1 mL) in dichloromethane (50 mL) was slowly added (COCl)₂ (0.97 mL, 11.4 mmol) at 0 °C. The reaction mixture was stirred at 0 °C for 2 h. The reaction mixture was evaporated. The crude acid chloride and *N*,*N*-diisopropylethylamine (DIPEA: 2.0 mL, 11.4 mmol) were dissolved in dichloromethane (50 mL). To this solution was slowly added cyclopropylamine (1.5 mL, 9.5 mmol) at 0 °C. The reaction mixture was stirred at room temperature for 2 h. The mixture was extracted with dichloromethane and the combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by Isolera® (hexane/ethyl acetate = 2:1 to 0:1). The obtained solid was washed with hexane to afford 1B (1.5 g, 87%) as a orange solid. ¹H NMR (600 MHz, CDCl₃) δ 5.70 (brs, 1H), 2.74–2.68 (m, 1H), 1.89–1.81 (m, 2H), 1.57–1.50 (m, 2H), 1.49–1.37 (m, 3H), 1.36–1.27 (m, 3H), 1.11 (s, 3H), 0.80–0.74 (m, 2H), 0.47–0.42 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 179.1, 42.4, 35.6, 26.2, 25.8, 22.8, 22.7, 6.7; HRMS (ESI) *m/z* calcd for C₁₁H₁₉NNaO [M+Na]⁺: 204.1359, found 204.1364.

N-Cyclopropyl-4,4-difluorocyclohexane-1-carboxamide (1E): To a round-bottom flask was added 4,4-difluorocyclohexane-1-carboxylic acid (443 mg, 2.7 mmol), 1-ethyl-3-(3-dimethylaminopropyl)carbodiimide (EDCI: 569 mg, 3.0 mmol, 1.1 equiv), 1-hydroxybenzotriazole

S3

^[2] Miyamura, S.; Araki, M.; Suzuki, T.; Yamaguchi, J.; Itami, K. Angew. Chem., Int. Ed. 2015, 54, 846.

^[3] Zheng, Y.; Liu, B.; Gou, Z.; Li, Y.; Zhang, X.; Wang, Y.; Yu, S.; Li, Y.; Sun, D. *Bioorg. Med. Chem. Lett.* **2015**, 25, 791.

^[4] Baburajan, P.; Elango, K. P. Tetrahedron Lett. 2014, 55, 1006.

(HOBt: 182 mg, 1.4 mmol, 0.5 equiv) and DMF (10 mL). Cyclopropylamine (0.19 mL, 2.7 mmol) was added to the mixture. After stirring for 3 h at room temperature, saturated aqueous NaHCO₃ was added to the mixture and the mixture was extracted with ethyl acetate. The organic layers were washed with brine, dried over Na₂SO₄ and concentrated *in vacuo*. The residue was purified by Isolera® (hexane/ethyl acetate = 1:1 to 0:1). The obtained solid was crystallized from hexane to afford **1E** (372 mg, 68%) as a white solid. 1 H NMR (600 MHz, CDCl₃) δ 5.54 (brs, 1H), 2.74–2.69 (m, 1H), 2.21–2.07 (m, 3H), 1.94–1.87 (m, 2H), 1.85–1.66 (m, 4H), 0.81–0.76 (m, 2H), 0.49–0.45 (m, 2H); 13 C NMR (126 MHz, CDCl₃) δ 175.3, 122.6 (t, J = 239.6 Hz), 42.6, 32.8 (t, J = 23.9 Hz), 25.8 (t, J = 9.5 Hz), 22.6, 6.7; HRMS (ESI) m/z calcd for C₁₀H₁₅F₂NNaO [M+Na]⁺: 226.1014, found 226.1014.

N-Cyclopropyltetrahydro-2*H*-pyran-4-carboxamide (1F): The synthetic procedure of 1F is the same as that of 1E. 6.0 mmol scale. Purification by Isolera[®] (hexane/ethyl acetate = 1:1 to 0:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1F (437 mg, 43%) as a white solid. 1 H NMR (600 MHz, CDCl₃) δ 5.55 (brs, 1H), 4.03–3.98 (m, 2H), 3.39 (td, J = 11.4, 2.4 Hz, 2H), 2.74–2.69 (m, 1H), 2.30–2.23 (m, 1H), 1.82–1.70 (m, 4H), 0.81–0.75 (m, 2H), 0.50–0.45 (m, 2H); 13 C NMR (126 MHz, CDCl₃) δ 175.6, 67.2, 42.0, 29.2, 22.6, 6.7; HRMS (ESI) m/z calcd for C₉H₁₅NNaO₂ [M+Na]⁺: 192.0995, found 192.0996.

N-Cyclopropylbenzamide (1H)^[5]: To a solution of cyclopropylamine (0.49 mL, 7.0 mmol) and *N*,*N*-diisopropylethylamine (DIPEA: 1.3 mL, 7.7 mmol, 1.1 equiv) in dichloromethane (10 mL) was slowly added a solution of benzoyl chloride (BzCl: 0.85 mL, 7.4 mmol, 1.05 equiv) in dichloromethane (5 mL) at 0 °C. The reaction mixture was stirred at room temperature for 1 h. To the reaction mixture was added saturated aqueous NaHCO₃, which was then extracted with dichloromethane. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by MPLC (hexane/ethyl acetate = 5:1 to 2:1). The obtained solid was crystallized from hexane and ethyl acetate to give 1H (1.1 g, 97%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.73 (dd, J = 7.8,

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^[5] Lin, J.-P.; Long, Y.-Q. Chem. Commun. 2013, 49, 5313.

1.8 Hz, 2H), 7.51–7.47 (m, 1H), 7.42 (t, J = 1.8 Hz, 2H), 6.23 (brs, 1H), 2.94–2.89 (m, 1H), 0.91–0.84 (m, 2H), 0.65–0.60 (m, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 168.9, 134.4, 131.4, 128.5, 126.8, 23.1, 6.8; HRMS (ESI) m/z calcd for $C_{10}H_{11}NNaO$ [M+Na]⁺: 184.0733, found 184.0734.

4-(*tert***-Butyl)-***N***-cyclopropylbenzamide (1K):** The synthetic procedure of **1K** is the same as that of **1H**. 4.0 mmol scale. Purification by MPLC (hexane/ethyl acetate = 2:1); the obtained solid was crystallized from hexane and ethyl acetate to afford **1K** (720 mg, 83%) as a white solid. 1 H NMR (600 MHz, CDCl₃) δ 7.67 (d, J = 8.4 Hz, 2H), 7.42 (d, J = 8.4 Hz, 2H), 6.22 (brs, 1H), 2.92–2.87 (m, 1H), 1.32 (s, 9H), 0.90–0.83 (m, 2H), 0.64–0.58 (m, 2H); 13 C NMR (151 MHz, CDCl₃) δ 168.7, 154.9, 131.6, 126.6, 125.4, 34.9, 31.1, 23.0, 6.8; HRMS (ESI) m/z calcd for C₁₄H₁₉NNaO [M+Na]⁺: 240.1359, found 240.1357.

N-Cyclopropyl-4-fluorobenzamide (1L): The synthetic procedure of 1L is the same as that of 1H. 21.3 mmol scale. Purification by MPLC (hexane/ethyl acetate = 3:1 to 1:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1L (2.1 g, 55%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.78–7.72 (m, 2H), 7.12–7.06 (m, 2H), 6.20 (brs, 1H), 2.92–2.86 (m, 1H), 0.92–0.83 (m, 2H), 0.66–0.58 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 167.8, 164.7 (d, J_{FC} = 250.2 Hz), 130.6 (d, J_{FC} = 2.9 Hz), 129.1 (d, J_{FC} = 8.6 Hz), 115.5 (d, J_{FC} = 21.6 Hz), 23.2, 6.7; HRMS (ESI) m/z calcd for C₁₀H₁₀FNNaO [M+Na]⁺: 202.0639, found 202.0639.

N-Cyclopropyl-3,5-dimethylbenzamide (1M): The synthetic procedure of 1M is the same as that of 1H.

3.5 mmol scale. Purification by MPLC (hexane/ethyl acetate = 3:1 to 1:1); the obtained solid was

crystallized from hexane and ethyl acetate to afford **1M** (250 mg, 38%) as a white solid. 1 H NMR (600 MHz, CDCl₃) δ 7.33 (s, 2H), 7.11 (s, 1H), 6.15 (brs, 1H), 2.92–2.87 (m, 1H), 2.34 (s, 6H), 0.90–0.84 (m, 2H), 0.63–0.58 (m, 2H); 13 C NMR (126 MHz, CDCl₃) δ 169.2, 138.2, 134.4, 133.0, 124.6, 23.0, 21.2, 6.8; HRMS (ESI) m/z calcd for $C_{12}H_{15}NNaO$ [M+Na]⁺: 212.1046, found 212.1045.

N-Cyclopropyl-3,5-dimethoxybenzamide (1N): The synthetic procedure of 1N is the same as that of 1B. 21.3 mmol scale. Purification by MPLC (hexane/ethyl acetate = 3:1 to 1:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1N (932 mg, 21%) as a white solid. 1 H NMR (600 MHz, CDCl₃) δ 6.85 (d, J = 1.8 Hz, 2H), 6.56 (dd, J = 1.8 Hz, 1H), 6.17 (brs, 1H), 3.82 (s, 6H), 2.92–2.86 (m, 1H), 0.90–0.83 (m, 2H), 0.64–0.57 (m, 2H); 13 C NMR (151 MHz, CDCl₃) δ 168.7, 160.8, 136.6, 104.8, 103.5, 55.5, 23.1, 6.7; HRMS (ESI) m/z calcd for C₁₂H₁₅NNaO₃ [M+Na]⁺: 244.0944, found 244.0942.

N-Cyclopropylisonicotinamide (10): To a mixture of isonicotinic acid (739 mg, 6.0 mmol) and 4-methylmorpholine (NMM: 1.3 mL, 12.0 mmol) in dichloromethane (15 mL) was slowly added isobutyl chloroformate (1.2 mL, 9.0 mmol) at 0 °C, and this mixture was stirred at the same temperature for 30 min. Cyclopropylamine (0.63 mL, 9.0 mmol) in dichloromethane (10 mL) and 4-methylmorpholine (NMM: 1.0 mL, 9.0 mmol) was slowly added at 0 °C. The reaction mixture was stirred at room temperature for 2 h. The reaction was quenched by adding saturated aqueous NaHCO₃, which was then extracted with dichloromethane. The combined organic layers were washed with brine, dried over Na₂SO₄, filtered and concentrated *in vacuo*. The residue was purified by Isolera® (hexane/ethyl acetate = 1:1 to 0:1). The obtained solid was washed by hexane to give **10** (772 mg, 79%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 8.73 (dd, J = 4.2, 1.8 Hz, 2H), 7.57 (dd, J = 4.2, 1.8 Hz, 2H), 6.30 (brs, 1H), 2.95–2.90 (m, 1H), 0.95–0.87 (m, 2H), 0.68–0.62 (m, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 167.0, 150.4, 141.4, 120.8, 23.2, 6.6; HRMS (ESI) m/z calcd for C₉H₁₀N₂NaO [M+Na]⁺: 185.0685, found 185.0687.

N-Cyclopropylfuran-2-carboxamide (1P): The synthetic procedure of 1P is the same as that of 1H. 20.0 mmol scale. Purification by MPLC (hexane/ethyl acetate = 3:1 to 1:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1P (1.7 g, 57%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.40 (s, 1H), 7.11 (d, J = 3.6 Hz, 1H), 6.49 (dd, J = 3.6, 1.2 Hz, 1H), 6.41 (brs, 1H), 2.89–2.84 (m, 1H), 0.89–0.82 (m, 2H), 0.66–0.59 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 159.6, 148.0, 143.7, 114.0, 112.1, 22.2, 6.7; HRMS (ESI) m/z calcd for C₈H₉NNaO₂ [M+Na]⁺: 174.0525, found 174.0521.

N-Cyclopropyl-5-methylfuran-2-carboxamide (1Q): The synthetic procedure of 1Q is the same as that of 1E. 5.0 mmol scale. Purification by Isolera[®] (hexane/ethyl acetate = 1:1 to 0:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1Q (538 mg, 65%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.00 (d, J = 3.6 Hz, 1H), 6.33 (brs, 1H), 6.08 (d, J = 3.6 Hz, 1H), 2.88–2.83 (m, 1H), 2.32 (s, 3H), 0.87–0.81 (m, 2H), 0.64–0.59 (m, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 159.7, 154.2, 146.4, 115.3, 108.5, 22.2, 13.8, 6.7; HRMS (ESI) m/z calcd for C₉H₁₁NNaO₂ [M+Na]⁺: 188.0682, found 188.0682.

N-Cyclopropyl-4-(*N*,*N*-dipropylsulfamoyl)benzamide (1R): The synthetic procedure of 1R is the same as that of 1E. 2.5 mmol scale. Purification by Isolera® (hexane/ethyl acetate = 1:1 to 0:1); the obtained solid was crystallized from hexane to afford 1R (561 mg, 69%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 7.83 (s, 4H), 6.34 (brs, 1H), 3.08 (t, J = 7.8 Hz, 4H), 2.95–2.90 (m, 1H), 1.54 (sext, J = 7.8 Hz, 4H), 0.93–0.88 (m, 2H), 0.87 (t, J = 7.8 Hz, 6H), 0.68–0.63 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 167.7, 142.5, 138.1, 127.7, 127.0, 49.9, 23.3, 21.8, 11.1, 6.5; HRMS (ESI) m/z calcd for C₁₆H₂₄N₂NaO₃S [M+Na]*: 347.1400, found 347.1396.

tert-Butyl (S)-(1-(cyclopropylamino)-3-methyl-1-oxobutan-2-yl)carbamate (1S): The synthetic procedure of 1S is the same as that of 1E. 4.0 mmol scale. Purification by Isolera® (hexane/ethyl acetate = 2:1 to 0:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1S (715 mg, 72%) as a white solid. 1 H NMR (600 MHz, 90 °C, DMSO- d_6) δ 7.62 (brs, 1H), 6.08 (brs, 1H), 3.72–3.68 (m, 1H), 2.67–2.61 (m, 1H), 1.93–1.86 (m, 1H), 1.40 (s, 9H), 0.85 (d, J = 6.8 Hz, 3H), 0.83 (d, J = 6.9 Hz, 3H), 0.65–0.59 (m, 2H), 0.46–0.37 (m, 2H); 13 C NMR (151 MHz, 90 °C, DMSO- d_6) δ 171.8, 154.8, 77.7, 59.4, 30.2, 27.7, 21.7, 18.6, 17.6, 5.1, 5.0; HRMS (ESI) m/z calcd for $C_{13}H_{24}N_2NaO_3$ [M+Na]+: 279.1679, found 279.1675.

tert-Butyl (S)-2-(cyclopropylcarbamoyl)pyrrolidine-1-carboxylate (1T): The synthetic procedure of 1T is the same as that of 1E. 5.0 mmol scale. Purification by Isolera® (hexane/ethyl acetate = 2:1 to 0:1); the obtained solid was crystallized from hexane and ethyl acetate to afford 1T (715 mg, 72%) as a white solid. 1 H NMR (500 MHz, 90 °C, DMSO- d_6) δ 7.53 (brs, 1H), 4.01–3.95 (m, 1H), 3.40–3.34 (m, 1H), 3.33–3.27 (m, 1H), 2.66–2.60 (m, 1H), 2.09–2.01 (m, 1H), 1.87–1.70 (m, 3H), 1.38 (s, 9H), 0.65–0.58 (m, 2H), 0.47–0.38 (m, 2H); 13 C NMR (126 MHz, 90 °C, DMSO- d_6) δ 172.8, 153.1, 78.0, 59.3, 46.1, 30.0, 27.7, 22.9, 21.8, 5.0; HRMS (ESI) m/z calcd for C_{13} H₂₂N₂NaO₃ [M+Na]⁺: 277.1523, found 277.1518.

N-Cyclopropyl-2,2-dimethylbutanamide (1U): The synthetic procedure of 1U is the same as that of 1B. 9.5 mmol scale. Purification by MPLC (hexane/ethyl acetate = 3:2 to 0:1) to afford 1T (1.0 g, 86%) as a yellow oil. 1 H NMR (500 MHz, CDCl₃) δ 5.68 (brs, 1H), 2.73–2.67 (m, 1H), 1.52 (q, J = 7.5 Hz, 2H), 1.12 (s, 6H), 0.82 (t, J = 7.5 Hz, 3H), 0.79–0.74 (m, 2H), 0.47–0.42 (m, 2H); 13 C NMR (126 MHz,

CDCl₃) δ 179.1, 42.1, 33.8, 24.8, 22.6, 9.1, 6.6; HRMS (ESI) m/z calcd for C₉H₁₇NNaO [M+Na]⁺: 178.1202, found 178.1204.

N-Cyclopropyl-1-methylcyclopropane-1-carboxamide (1V): The synthetic procedure of 1V is the same as that of 1B. 6.2 mmol scale. Purification by Isolera® (hexane/ethyl acetate = 2:1 to 0:1); the obtained solid was crystallized from hexane to afford 1V (289 mg, 34%) as a white solid. ¹H NMR (600 MHz, CDCl₃) δ 5.83 (brs, 1H), 2.74–2.68 (m, 1H), 1.27 (s, 3H), 1.21–1.17 (m, 2H), 0.79–0.74 (m, 2H), 0.56–0.53 (m, 2H), 0.51–0.47 (m, 2H); ¹³C NMR (151 MHz, CDCl₃) δ 176.2, 23.0, 19.7, 18.9, 16.0, 6.6; HRMS (ESI) m/z calcd for C₈H₁₃NNaO [M+Na]⁺: 162.0889, found 162.0886.

3. General Procedure for the Hydrosilylation of Aminocyclopropanes with [Rh(cod)Cl]₂/P(1-nap)₃ Catalyst

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heat gun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added aminocyclopropane (0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, 8.8 μmol, 2.5 mol%), and (P(1-nap)₃) (14.4 mg, 0.035 mmol, 10 mol%), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, silane (0.70 mmol, 2 equiv) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The residue was purified by Isolera® to afford aminosilane 4. The branch/linear (4/3) ratio was determined by ¹H NMR analysis of the crude product.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)pivalamide (4Aa): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Aa as a yellow oil (103.4 mg, 81%, branch/linear = 90:10). 1 H NMR (600 MHz, CDCl₃) δ 5.45 (d, J = 8.4 Hz, 1H), 3.32 (td, J = 9.6, 4.8 Hz, 1H), 1.68–1.60 (m, 1H), 1.39–1.30 (m, 1H), 1.20 (s, 9H), 0.90 (t, J = 7.8 Hz, 3H), 0.12 (s, 9H), 0.11 (s, 9H), 0.052 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 177.9, 40.9, 38.7, 27.7, 23.6, 11.4, 1.81, 1.79, -1.6; HRMS (DART) m/z calcd for C₁₅H₃₈NO₃Si₃ [M+H]⁺: 364.2154, found 364.2153.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-1-methylcyclohexane-1-carboxamide (4Ba): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ba as a colorless oil (117.3 mg, 83%, branch/linear = 90:10). 1 H NMR (400 MHz, CDCl₃) δ 5.48 (d, J = 10.0 Hz, 1H), 3.36 (td, J = 9.6, 4.8 Hz,

1H), 1.97–1.87 (m, 2H), 1.71–1.24 (m, 10H), 1.14 (s, 3H), 0.91 (t, J = 7.6 Hz, 3H), 0.12 (s, 9H), 0.11 (s, 9H), 0.055 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 177.0, 42.8, 40.7, 35.9, 35.8, 27.0, 25.9, 23.7, 23.0, 11.6, 1.82, 1.78, -1.5; HRMS (DART) m/z calcd for $C_{18}H_{42}NO_3Si_3$ [M+H]+: 404.2467, found 404.2470.

N-(1-(λ^1 -Silyl)propyl)cyclohexanecarboxamide (4Ca): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ca as a white solid (92.6 mg, 68%, branch/linear = 78:22). ¹H NMR (600 MHz, CDCl₃) δ 5.18 (d, J = 9.6 Hz, 1H), 3.32 (td, J = 9.6, 4.8 Hz, 1H), 2.10–2.03 (m, 1H), 1.91–1.83 (m, 2H), 1.82–1.75 (m, 2H), 1.71–1.58 (m, 2H), 1.48–1.17 (m, 6H), 0.90 (t, J = 7.2 Hz, 3H), 0.114 (s, 9H), 0.111 (s, 9H), 0.049 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 175.6, 46.0, 40.9, 30.0, 29.9, 25.8, 23.6, 11.5, 1.80, 1.77, -1.7; HRMS (DART) m/z calcd for C₁₇H₄₀NO₃Si₃ [M+H]⁺: 390.2311, found 390.2311.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)isobutyramide (4Da): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Da as a yellow solid (86.5 mg, 71%). The branch/linear ratio couldn't be determined because of the peak overlap. 1 H NMR (600 MHz, CDCl₃) δ 5.18 (d, J = 7.8 Hz, 1H), 3.32 (td, J = 9.6, 4.8 Hz, 1H), 2.38–2.31 (m, 1H), 1.68–1.59 (m, 1H), 1.39–1.31 (m, 1H), 1.17 (d, J = 7.2 Hz, 3H), 1.16 (d, J = 7.2 Hz, 3H), 0.91 (t, J = 7.2 Hz, 3H), 0.12 (s, 9H), 0.11 (s, 9H), 0.055 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 176.4, 40.9, 36.0, 23.6, 19.83, 19.76, 11.5, 1.77, 1.75, -1.7; HRMS (DART) m/z calcd for $C_{14}H_{36}NO_{3}Si_{3}$ [M+H]⁺: 350.1997, found 350.2003.

4,4-Difluoro-N-(1-(1,1,1,3,5,5,5-heptamethyltrisiloxan-3-yl)propyl)cyclohexane-1-carboxamide

(4Ea): The reaction was performed using $[Rh(cod)OMe]_2$ (2.1 mg, 4.4 µmol, 1.25 mol%) and $P(1-nap)_3$ (7.2 mg, 17.5 µmol, 5 mol%). Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ea as

a white solid (77.3 mg, 52%, branch/linear = 56:44). ¹H NMR (400 MHz, CDCl₃) δ 5.20 (d, J = 9.6 Hz, 1H), 3.33 (td, J = 9.6, 5.2 Hz, 1H), 2.24–2.11 (m, 3H), 1.99–1.55 (m, 7H), 1.42–1.25 (m, 1H), 0.90 (t, J = 7.2 Hz, 3H), 0.12 (s, 9H), 0.11 (s, 9H), 0.055 (s, 3H); ¹³C NMR (101 MHz, CDCl₃) δ 173.5, 122.7 (t, J = 244.2 Hz), 43.3, 41.2, 32.9 (t, J = 24.8 Hz), 26.1 (t, J = 8.6 Hz), 23.6, 11.5, 1.82, 1.79, -1.6; HRMS (DART) m/z calcd for $C_{17}H_{38}F_2NO_3Si_3$ [M+H]⁺: 426.2122, found 426.2124.

N-(1-(1,1,1,3,5,5,5)-Heptamethyltrisiloxan-3-yl)propyl)tetrahydro-2H-pyran-4-carboxamide (4Fa):

The reaction was performed using [Rh(cod)OMe]₂ (2.1 mg, 4.4 μ mol, 1.25 mol%) and P(1-nap)₃ (7.2 mg, 17.5 μ mol, 5 mol%). Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **4Fa** as a white solid (76.0 mg, 55%). The branch/linear ratio couldn't be determined because of the peak overlap. ¹H NMR (600 MHz, CDCl₃) δ 5.21 (d, J = 9.6 Hz, 1H), 4.05–4.00 (m, 2H), 3.43 (td, J = 11.4, 3.0 Hz, 2H), 3.34 (td, J = 9.6, 4.8 Hz, 1H), 2.37–2.30 (m, 1H), 1.85–1.72 (m, 4H), 1.68–1.60 (m, 1H), 1.40–1.30 (m, 1H), 0.90 (t, J = 7.8 Hz, 3H), 0.12 (s, 9H), 0.11 (s, 9H), 0.056 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 173.8, 67.4, 42.6, 41.1, 29.6, 23.6, 11.5, 1.84, 1.81, -1.6; HRMS (DART) m/z calcd for C₁₆H₃₈NO₄Si₃ [M+H]⁺: 392.2103, found 392.2101.

4Ga

tert-butyl (1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)carbamate (4Ga): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ga as a colorless oil (92.1 mg, 69%). The branch/linear ratio couldn't be determined because of the peak overlap. 1 H NMR (600 MHz, 120 $^{\circ}$ C, DMSO- d_{6}) δ 5.53 (brs, 1H), 2.72 (td, J = 9.6, 4.8 Hz, 1H), 1.58–1.50 (m, 1H), 1.44–1.35 (m, 10H), 0.90 (t, J = 7.2 Hz, 3H), 0.13–0.10 (m, 18H), 0.072 (s, 3H); 13 C NMR (151 MHz, 120 $^{\circ}$ C, DMSO- d_{6}) δ 155.3, 76.8, 42.6, 27.7, 22.7, 10.8, 1.0, -2.5; HRMS (DART) m/z calcd for $C_{15}H_{38}NO_{4}Si_{3}$ [M+H] $^{+}$: 380.2103, found 380.2101.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)benzamide (4Ha): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ha as a white solid (73.8 mg, 55%). When the reaction was performed using [Rh(cod)OMe]₂ (2.1 mg, 4.4 μmol, 1.25 mol%) and P(1-nap)₃ (7.2 mg, 17.5 μmol, 5 mol%), 4Ha was obtained in 66% yield (88.9 mg, branch/linear = 78:22). ¹H NMR (600 MHz, CDCl₃) δ 7.75–7.72 (m, 2H), 7.50–7.46 (m, 1H), 7.45–7.41 (m, 2H), 5.90 (d, J = 9.0 Hz, 1H), 3.55 (td, J = 9.6, 4.8 Hz, 1H), 1.79–1.71 (m, 1H), 1.52–1.44 (m, 1H), 0.98 (t, J = 7.3 Hz, 3H), 0.13 (s, 9H), 0.12 (s, 3H), 0.11 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 167.3, 135.5, 131.0, 128.5, 126.6, 42.0, 23.8, 11.7, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{17}H_{34}NO_3Si_3$ [M+H]⁺: 384.1841, found 384.1846.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-4-methylbenzamide (4Ia): The reaction was performed using [Rh(cod)OMe]₂ (2.1 mg, 4.4 μmol, 1.25 mol%) and P(1-nap)₃ (7.2 mg, 17.5 μmol, 5 mol%). Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ia as a colorless oil (90.8 mg, 65%, branch/linear = 82:18). ¹H NMR (600 MHz, CDCl₃) δ 7.64 (d, J = 8.4 Hz, 2H), 7.23 (d, J = 8.4 Hz, 2H), 5.88 (d, J = 9.6 Hz, 1H), 3.54 (td, J = 9.6, 4.8 Hz, 1H), 2.39 (s, 3H), 1.78–1.69 (m, 1H), 1.52–1.42 (m, 1H), 0.97 (t, J = 7.2 Hz, 3H), 0.13 (s, 9H), 0.11 (s, 3H), 0.10 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 167.3, 141.3, 132.6, 129.2, 126.6, 41.9, 23.8, 21.4, 11.7, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{18}H_{36}NO_3Si_3$ [M+H]*: 398.1997, found 398.1997.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-4-methoxybenzamide (4Ja): The reaction was performed using [Rh(cod)OMe]₂ (2.1 mg, 4.4 μmol, 1.25 mol%) and P(1-nap)₃ (7.2 mg, 17.5 μmol, 5 mol%). Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ja as a colorless oil (92.8 mg, 64%, branch/linear = 82:18). ¹H NMR (600 MHz, CDCl₃) δ 7.71 (d, J = 9.0 Hz, 2H), 6.93 (d, J = 9.0

Hz, 2H), 5.84 (d, J = 10.2 Hz, 1H), 3.85 (s, 3H), 3.53 (td, J = 9.6, 4.8 Hz, 1H), 1.78–1.68 (m, 1H), 1.52–1.43 (m, 1H), 0.97 (t, J = 7.2 Hz, 3H), 0.13 (s, 9H), 0.110 (s, 3H), 0.105 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 166.8, 161.9, 128.3, 127.7, 113.7, 55.4, 41.8, 23.9, 11.7, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{18}H_{36}NO_4Si_3$ [M+H]⁺: 414.1947, found 414.1950.

4-(*tert*-**Butyl**)-*N*-(**1-(1,1,1,3,5,5,5-heptamethyltrisiloxan-3-yl)propyl)benzamide (4Ka**): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **4Ka** as a colorless oil (99.0 mg, 65%, branch/linear = 82:18). When the reaction was performed using [Rh(cod)OMe]₂ (2.1 mg, 4.4 μmol, 1.25 mol%) and P(1-nap)₃ (7.2 mg, 17.5 μmol, 5 mol%), **4Ha** was obtained in 79% (122.0 mg, branch/linear = 79:21). 1 H NMR (600 MHz, CDCl₃) δ 7.68 (d, J = 8.6 Hz, 2H), 7.45 (d, J = 8.6 Hz, 2H), 5.89 (d, J = 9.0 Hz, 1H), 3.55 (td, J = 9.6, 4.8 Hz, 1H), 1.78–1.70 (m, 1H), 1.51–1.43 (m, 1H), 1.34 (s, 9H), 0.97 (t, J = 7.4 Hz, 3H), 0.14 (s, 9H), 0.12 (s, 9H), 0.11 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 167.2, 154.4, 132.5, 126.4, 125.4, 41.8, 34.8, 31.1, 23.8, 11.7, 1.8, -1.6; HRMS (DART) m/z calcd for $C_{21}H_{42}NO_3Si_3$ [M+H]⁺: 440.2467, found 440.2466.

4-Fluoro-*N***-(1-(1,1,1,3,5,5,5-heptamethyltrisiloxan-3-yl)propyl)benzamide (4La):** Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **4La** as a yellow oil (84.5 mg, 60%, branch/linear = 72:28). 1 H NMR (600 MHz, CDCl₃) δ 7.76–7.72 (m, 2H), 7.13–7.08 (m, 2H), 5.83 (d, J = 9.6 Hz, 1H), 3.53 (td, J = 9.0, 4.8 Hz, 1H), 1.78–1.70 (m, 1H), 1.52–1.43 (m, 1H), 0.98 (t, J = 7.8 Hz, 3H), 0.15–0.08 (m, 21H); 13 C NMR (151 MHz, CDCl₃) δ 166.3, 164.5 (d, J_{FC} = 248.6 Hz), 131.6 (d, J_{FC} = 2.9 Hz), 128.8 (d, J_{FC} = 8.7 Hz), 115.5 (d, J_{FC} = 21.5 Hz), 42.1, 23.8, 11.7, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{17}H_{33}$ FNO₃Si₃ [M+H]⁺: 402.1747, found 402.1745.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-3,5-dimethylbenzamide (4Ma): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ma as a white solid (113.6 mg, 79%). The branch/linear ratio couldn't be determined because of the peak overlap. ¹H NMR (600 MHz, CDCl₃) δ 7.34 (s, 2H), 7.11 (s, 1H), 5.89 (d, J = 10.2 Hz, 1H), 3.53 (td, J = 9.6, 4.8 Hz, 1H), 2.35 (s, 6H), 1.78–1.70 (m, 1H), 1.51–1.43 (m, 1H), 0.97 (t, J = 7.8 Hz, 3H), 0.14 (s, 9H), 0.12 (s, 9H), 0.11 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 167.7, 138.2, 135.4, 132.6, 124.4, 41.8, 23.8, 21.2, 11.7, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{19}H_{38}NO_3Si_3$ [M+H]⁺: 412.2154, found 412.2152.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-3,5-dimethoxybenzamide (4Na): Purification by Isolera[®] (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Na as a white solid (108.0 mg, 70%, branch/linear = 83:17). ¹H NMR (600 MHz, CDCl₃) δ 6.87 (d, J = 2.4 Hz, 2H), 6.57–6.56 (m, 1H), 5.86 (d, J = 9.6 Hz, 1H), 3.83 (s, 6H), 3.52 (td, J = 9.6, 4.8 Hz, 1H), 1.78–1.70 (m, 1H), 1.51–1.42 (m, 1H), 0.98 (t, J = 7.2 Hz, 3H), 0.14–0.10 (m, 21H); ¹³C NMR (151 MHz, CDCl₃) δ 167.2, 160.9, 137.7, 104.6, 103.2, 55.5, 42.1, 23.8, 11.7, 1.8, -1.6; HRMS (DART) m/z calcd for $C_{19}H_{38}NO_5Si_3$ [M+H]⁺: 444.2052, found 444.2052.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)isonicotinamide (4Oa): The reaction was stirred for 18 h. Purification by Isolera[®] (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Oa as a yellow oil (72.9 mg, 51%, branch/linear = 80:20). ¹H NMR (600 MHz, CDCl₃) δ 8.74 (dd, J = 4.2, 1.8 Hz, 2H), 7.57 (dd, J = 4.2, 1.8 Hz, 2H), 5.95 (d, J = 9.6 Hz, 1H), 3.54 (td, J = 9.6, 4.8 Hz, 1H), 1.80–1.71 (m, 1H),

1.53–1.44 (m, 1H), 0.98 (t, J = 7.2 Hz, 3H), 0.134 (s, 9H), 0.126 (s, 3H), 0.10 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 165.3, 150.6, 142.4, 120.6, 42.5, 23.7, 11.7, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{16}H_{33}N_2O_3Si_3$ [M+H]⁺: 385.1793, found 385.1795.

N-(1-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)furan-2-carboxamide (4Pa): The reaction was stirred for 18 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Pa as a colorless oil (86.2 mg, 66%). The branch/linear ratio couldn't be determined because of the peak overlap. 1 H NMR (600 MHz, CDCl₃) δ 7.42 (s, 1H), 7.07 (d, J = 3.6 Hz, 1H), 6.49 (dd, J = 3.6, 1.8 Hz, 1H), 6.17 (d, J = 10.2 Hz, 1H), 3.48 (td, J = 9.6, 4.8 Hz, 1H), 1.76–1.68 (m, 1H), 1.51–1.42 (m, 1H), 0.97 (t, J = 7.2 Hz, 3H), 0.13 (s, 9H), 0.11 (s, 9H), 0.10 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 158.4, 148.5, 143.4, 113.5, 112.0, 41.1, 23.8, 11.7, 1.7, -1.6; HRMS (DART) m/z calcd for $C_{15}H_{32}NO_4Si_3$ [M+H]*: 374.1634, found 374.1635.

N-(1-(1,1,1,3,5,5,5)-Heptamethyltrisiloxan-3-yl)propyl)-5-methylfuran-2-carboxamide (4Qa):

Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **4Qa** as a colorless oil (96.5 mg, 71%, branch/linear = 81:19). 1 H NMR (500 MHz, CDCl₃) δ 6.96 (d, J = 3.0 Hz, 1H), 6.12 (d, J = 10.0 Hz, 1H), 6.08 (d, J = 3.0 Hz, 1H), 3.47 (td, J = 9.5, 5.0 Hz, 1H), 2.33 (s, 3H), 1.77–1.67 (m, 1H), 1.51–1.41 (m, 1H), 0.97 (t, J = 7.5 Hz, 3H), 0.13 (s, 9H), 0.12 (s, 9H), 0.10 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 158.6, 153.9, 146.9, 114.6, 108.3, 40.9, 23.8, 13.7, 11.7, 1.7, -1.6; HRMS (DART) m/z calcd for $C_{16}H_{34}NO_4Si_3$ [M+H]+: 388.1790, found 388.1790.

4-(*N*,*N*-**Dipropylsulfamoyl**)-*N*-(**1-(1,1,1,3,5,5,5-heptamethyltrisiloxan-3-yl)propyl)benzamide (4Ra**): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **4Ra** as a colorless oil (94.8 mg, 50%, branch/linear = 65:35). 1 H NMR (500 MHz, CDCl₃) δ 7.89–7.81 (m, 4H), 5.91 (d, J = 10.0 Hz, 1H), 3.54 (td, J = 9.5, 5.0 Hz, 1H), 3.10 (t, J = 7.0 Hz, 4H), 1.80–1.71 (m, 1H), 1.60–1.45 (m, 5H), 0.99 (t, J = 7.5 Hz, 3H), 0.87 (t, J = 7.0 Hz, 6H), 0.14–0.080 (m, 21H); 13 C NMR (151 MHz, CDCl₃) δ 166.0, 142.6, 138.9, 127.3, 49.9, 42.5, 23.7, 21.9, 11.8, 11.1, 1.8, -1.5; HRMS (DART) m/z calcd for $C_{23}H_{47}N_2O_5SSi_3$ [M+H]+: 547.2508, found 547.2507.

tert-Butyl ((2*S*)-1-((1-(1,1,1,3,5,5,5-heptamethyltrisiloxan-3-yl)propyl)amino)-3-methyl-1-oxobutan-2-yl)carbamate (4Sa): The reaction was performed using [Rh(cod)OMe]₂ (2.1 mg, 4.4 μmol, 1.25 mol%) and P(1-nap)₃ (7.2 mg, 17.5 μmol, 5 mol%) for 18 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Sa as a white solid (87.0 mg, 52%, mixture of diastereomers). The branch/linear ratio couldn't be determined because of the peak overlap. ¹H NMR (600 MHz, 60 °C, CDCl₃) δ 5.64–5.53 (m, 1H), 4.95 (brs, 1H), 3.86–3.80 (m, 1H), 3.35–3.28 (m, 1H), 2.20–2.11 (m, 1H), 1.69–1.60 (m, 1H), 1.44–1.43 (m, 9H), 1.41–1.32 (m, 1H), 0.96 (t, J = 7.2 Hz, 3H), 0.94–0.88 (m, 6H), 0.13–0.11 (m, 18H), 0.080–0.040 (m, 3H); ¹³C NMR (101 MHz, 60 °C, CDCl₃) δ 171.3, 171.1, 155.8, 155.6, 79.6, 60.5, 41.5, 30.7, 30.6, 28.2, 23.51, 23.47, 19.34, 19.28, 17.9, 17.7, 11.7, 11.5, 1.7, -1.6, -1.8; HRMS (DART) m/z calcd for $C_{20}H_{47}N_2O_5Si_3$ [M+H]*: 479.2787, found 479.2786.

tert-Butyl (2S)-2-((1-(1,1,1,3,5,5,5-heptamethyltrisiloxan-3-yl)propyl)carbamoyl)pyrrolidine-1-carboxylate (4Ta): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ta as a colorless oil (43.0 mg, 26%) and Diastereomer-4Ta (65.6 mg, 39%). The branch/linear ratio couldn't be determined because of the peak overlap. 1 H NMR (600 MHz, 120 °C, DMSO- d_6) δ 6.72 (d, J = 8.4 Hz, 1H), 4.21 (dd, J = 8.4, 3.0 Hz, 1H), 3.38–3.32 (m, 2H), 3.09 (td, J = 9.0, 4.8 Hz, 1H), 2.07–1.99 (m, 1H), 1.97–1.90 (m, 1H), 1.88–1.76 (m, 2H), 1.63–1.55 (m, 1H), 1.44–1.36 (m, 10H), 0.89 (t, J = 7.2 Hz, 3H), 0.124 (s, 9H), 0.116 (s, 9H), 0.056 (s, 3H); 13 C NMR (151 MHz, 120 °C, DMSO- d_6) δ 170.8, 153.5, 78.2,

59.2, 46.0, 41.1, 29.2, 27.6, 22.8, 22.4, 10.9, 1.0, -2.4; HRMS (DART) m/z calcd for $C_{20}H_{45}N_2O_5Si_3$ [M+H]⁺: 477.2631, found 477.2635. Diastereomer-**4Ta** ¹H NMR (600 MHz, 120 °C, DMSO- d_6) δ 6.48 (d, J = 7.8 Hz, 1H), 4.18 (dd, J = 8.4, 3.0 Hz, 1H), 3.41–3.32 (m, 2H), 3.08 (td, J = 9.0, 4.8 Hz, 1H), 2.10–2.04 (m, 1H), 1.96–1.91 (m, 1H), 1.84–1.77 (m, 2H), 1.63–1.55 (m, 1H), 1.43–1.36 (m, 10H), 0.89 (t, J = 7.2 Hz, 3H), 0.122 (s, 9H), 0.117 (s, 9H), 0.059 (s, 3H); ¹³C NMR (151 MHz, 120 °C, DMSO- d_6) δ 170.8, 153.5, 78.4, 59.7, 46.1, 41.0, 29.4, 27.6, 22.8, 22.4, 10.9, 1.0, -2.3; HRMS (DART) m/z calcd for $C_{20}H_{45}N_2O_5Si_3$ [M+H]⁺: 477.2631, found 477.2639.

N-(1-(*tert*-Butyldimethylsilyl)propyl)pivalamide (4Ab): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ab as a yellow solid (41.1 mg, 46%, branch/linear = 84:16). ¹H NMR (600 MHz, CDCl₃) δ 5.32 (d, J = 8.4 Hz, 1H), 3.65 (td, J = 10.2, 3.6 Hz, 1H), 1.71–1.63 (m, 1H), 1.41–1.31 (m, 1H), 1.21 (s, 9H), 0.94–0.88 (m, 12 H), 0.013 (s, 6H); ¹³C NMR (151 MHz, CDCl₃) δ 177.3, 38.8, 38.4, 27.7, 26.8, 25.3, 16.9, 11.7, -7.2, -7.8; HRMS (DART) m/z calcd for C₁₄H₃₂NOSi [M+H]⁺: 258.2248, found 258.2249.

N-(1-(Tripropylsilyl)propyl)pivalamide (4Ac): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ac as a white solid (100.8 mg, 96%, branch/linear = 94:6). ¹H NMR (600 MHz, CDCl₃) δ 5.27 (d, J = 10.2 Hz, 1H), 3.58 (td, J = 10.8, 3.6 Hz, 1H), 1.65–1.58 (m, 1H), 1.43–1.30 (m, 7H), 1.20 (s, 9H), 0.96 (t, J = 7.2 Hz, 9H), 0.90 (t, J = 7.2 Hz, 3H), 0.62–0.52 (m, 6H); ¹³C NMR (151 MHz, CDCl₃) δ 177.5, 39.3, 38.8, 27.8, 24.9, 18.7, 17.5, 14.0, 12.0; HRMS (DART) m/z calcd for C₁₇H₃₈NOSi [M+H]⁺: 300.2717, found 300.2719.

N-(1-(Diethyl(methyl)silyl)propyl)pivalamide (4Ad): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 4Ad as a white solid (86.0 mg, 99%, branch/linear = >95:5). ¹H NMR (600 MHz, CDCl₃) δ 5.27 (d, J = 8.4 Hz, 1H), 3.54 (td, J = 10.2, 3.6 Hz, 1H), 1.66–1.58 (m, 1H), 1.40–1.30 (m, 1H), 1.20 (s, 9H), 0.98–0.94 (m, 6H), 0.91 (t, J = 7.2 Hz, 3H), 0.63–0.50 (m, 4H), -0.023 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 177.6, 39.6, 38.8, 27.8, 24.5, 11.9, 7.42, 7.39, 3.68, 3.66, -7.8; HRMS (DART) m/z calcd for $C_{13}H_{30}NOSi$ [M+H]*: 244.2091, found 244.2091.

N-(1-(Dimethyl(phenyl)silyl)propyl)pivalamide (4Ae): The reaction was performed using [Rh(cod)Cl]₂ (8.6 mg, 17.5 μmol, 5.0 mol%), and P(1-nap)₃ (28.8 mg, 0.070 mmol, 20 mol%) for 12 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **4Ae** as a white solid (59.7 mg, 61%, branch/linear = 87:13). ¹H NMR (600 MHz, CDCl₃) δ 7.51 (dd, J = 7.2, 2.4 Hz, 2H), 7.40–7.35 (m, 3H), 5.14 (d, J = 9.0 Hz, 1H), 3.63 (td, J = 10.8, 4.2 Hz, 1H), 1.66–1.59 (m, 1H), 1.33–1.24 (m, 1H), 1.13 (s, 9H), 0.86 (t, J = 7.2 Hz, 3H), 0.35 (s, 3H), 0.33 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 177.8, 136.3, 134.0, 129.4, 127.9, 40.7, 38.8, 27.7, 24.3, 11.9, -4.7, -5.0; HRMS (DART) m/z calcd for C₁₆H₂₈NOSi [M+H]⁺: 278.1935, found 278.1934.

4. General Procedure for the Hydrosilylation of Aminocyclopropanes with $[Rh(cod)Cl]_2/PCy_3 \cdot HBF_4$ Catalyst

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added aminocyclopropane (0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, 8.8 μmol, 2.5 mol%), and PCy₃·HBF₄ (12.9 mg, 0.035 mmol, 10 mol%), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (2a: 191 μL, 0.70 mmol, 2.0 equiv) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The residue was purified by Isolera® to afford aminosilane 3. The linear/branch (3/4) ratio was determined by ¹H NMR analysis of the crude product.

N-(3-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)pivalamide (3Aa): Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 3Aa as a colorless oil (71.1 mg, 56%, linear/branch = 99:1). ¹H NMR (600 MHz, CDCl₃) δ 5.66 (brs, 1H), 3.22 (q, J = 6.6 Hz, 2H), 1.53–1.47 (m, 2H), 1.20 (s, 9H), 0.47–0.43 (m, 2H), 0.089 (s, 18H), 0.018 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 178.2, 42.1, 38.6, 27.6, 23.3, 14.7, 1.8, -0.37; HRMS (ESI) m/z calcd for C₁₅H₃₇NNaO₃Si₃ [M+Na]⁺: 386.1973, found 386.1972.

N-(3-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-1-methylcyclohexane-1-carboxamide (3Ba): The reaction was stirred for 12 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 3Ba as a brown oil (83.3 mg, 59%, linear/branch = 99:1). 1 H NMR (600 MHz, CDCl₃) δ 5.66 (br, 1H), 3.24 (q, J = 6.6 Hz, 2H), 1.93–1.87 (m, 2H), 1.58–1.39 (m, 7H), 1.37–1.30 (m, 3H), 1.14 (s, 3H), 0.47–0.43 (m, 2H), 0.089 (s, 18H), 0.017 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 177.4, 42.6, 42.0, 35.7, 26.5, 25.8,

23.5, 22.9, 14.8, 1.8, -0.36; HRMS (DART) m/z calcd for $C_{18}H_{42}NO_3Si_3$ [M+H]⁺: 404.2467, found 404.2468.

N-(3-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-2,2-dimethylbutanamide (3Ua): The reaction was stirred for 12 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 3Ua as a colorless oil (67.2 mg, 51%, linear/branch = 99:1). 1 H NMR (600 MHz, CDCl₃) δ 5.62 (brs, 1H), 3.23 (q, J = 7.2 Hz, 2H), 1.54 (q, J = 7.2 Hz, 2H), 1.52–1.46 (m, 2H), 1.15 (s, 6H), 0.84 (t, J = 7.2 Hz, 3H), 0.47–0.42 (m, 2H), 0.088 (s, 18H), 0.016 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 177.4, 42.3, 42.1, 33.9, 25.0, 23.5, 14.8, 9.2, 1.8, -0.37; HRMS (DART) m/z calcd for C₁₆H₄₀NO₃Si₃ [M+H]⁺: 378.2311, found 378.2310.

N-(3-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-1-methylcyclopropane-1-carboxamide (3Va):

Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded **3Va** as a brown oil (49.2 mg, 39%, linear/branch = 99:1). 1 H NMR (600 MHz, CDCl₃) δ 5.77 (brs, 1H), 3.24 (q, J = 6.6 Hz, 2H), 1.56–1.49 (m, 2H), 1.32 (s, 3H), 1.19 (dd, J = 6.0, 3.6 Hz, 2H), 0.55 (dd, J = 6.0, 3.6 Hz, 2H), 0.48–0.44 (m, 2H), 0.094 (s, 18H), 0.023 (s, 3H); 13 C NMR (151 MHz, CDCl₃) δ 174.7, 42.5, 23.4, 19.7, 18.9, 15.8, 14.8, 1.8, -0.37; HRMS (DART) m/z calcd for $C_{15}H_{36}NO_3Si_3$ [M+H]⁺: 362.1997, found 362.1996.

ЗМа

N-(3-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-3,5-dimethylbenzamide (3Ma): The reaction was stirred for 25 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 3Ma as a brown oil (59.0 mg, 41%, linear/branch = 99:1). ¹H NMR (600 MHz, CDCl₃) δ 7.36 (s, 2H), 7.11 (s, 1H), 6.17 (brs, 1H), 3.42 (q, J = 6.6 Hz, 2H), 2.35 (s, 6H), 1.66–1.59 (m, 2H), 0.56–0.51 (m, 2H), 0.096 (s, 18H), 0.034 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 167.8, 138.2, 135.0, 132.8, 124.6, 42.6, 23.4, 21.2, 14.9, 1.8, -0.33; HRMS (DART) m/z calcd for C₁₉H₃₈NO₃Si₃ [M+H]⁺: 412.2154, found 412.2153.

N-(3-(1,1,1,3,5,5,5-Heptamethyltrisiloxan-3-yl)propyl)-3,5-dimethoxybenzamide (3Na): The reaction was stirred for 25 h. Purification by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) afforded 3Na as a colorless oil (63.5 mg, 41%, linear/branch = 99:1). ¹H NMR (600 MHz, CDCl₃) δ 6.89 (d, J = 2.4 Hz, 2H), 6.57 (d, J = 2.4 Hz, 1H), 6.15 (brs, 1H), 3.82 (s, 6H), 3.42 (q, J = 7.2 Hz, 2H), 1.66–1.59 (m, 2H), 0.54–0.50 (m, 2H), 0.095 (s, 18H), 0.040 (s, 3H); ¹³C NMR (151 MHz, CDCl₃) δ 167.2, 160.8, 137.2, 104.8, 103.3, 55.5, 42.7, 23.4, 14.9, 1.8, -0.32; HRMS (DART) m/z calcd for $C_{19}H_{38}NO_5Si_3$ [M+H]⁺: 444.2052, found 444.2054.

5. Synthetic Applications

5-1. Gram-scale Reaction

A 100 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. After adding *N*-cyclopropyl-3,5-dimethoxybenzamide (**1N**: 1.0 g, 4.5 mmol) and tri(naphthalen-1-yl)phosphine (P(1-nap)₃: 93 mg, 0.23 mmol), the vessel was introduced inside an argonatmosphere glovebox. In the glovebox, [Rh(cod)Cl]₂ (27.2 mg, 0.056 mmol) and THF (13 mL) were added. After 1,1,1,3,5,5,5-heptamethyltrisiloxane (**2a**: 2.5 \Box L, 9.0 mmol) and THF (10 mL) were added, the glass vessel was sealed with the O-ring tap and taken out of the glovebox. The mixture was stirred in an oil bath at 110 °C for 18 h, cooled to room temperature and concentrated *in vacuo*. The residue was purified by Isolera® (hexane/ethyl acetate = 9:1 to 5:1) to afford **4Na** (1.68 g, 84%, branch/linear = 88:12) as a white solid.

5-2. Tamao Oxidation

To a screw-cap 10 mL glass vessel containing a magnetic stirring bar was added **3Ma** (30.0 mg, 0.073 mmol). The contents were evacuated, then filled with nitrogen (repeat for a total of 3 times). After THF (1.4 mL) was added, the solution was cooled to 0 °C. To the solution was added tetrabutylammonium fluoride (Bu₄NF: 1 M in THF, 0.32 mL, 0.32 mmol), which was then stirred at 0 °C for 10 min. MeOH (0.5 mL), KHCO₃ (36.5 mg, 0.37 mmol) and 30% aqueous H_2O_2 (0.17 mL, 1.5 mmol) were added, and the mixture was stirred at room temperature for 9 h. The reaction was quenched by adding saturated aqueous $Na_2S_2O_3$. The mixture was extracted with diethyl ether and the combined organic layers were washed with brine and concentrated *in vacuo*. The residue was purified by PTLC (hexane/ethyl acetate = 1:1) to afford **5Ma** (14.0 mg, 93%) as a colorless oil. **N-(3-Hydroxypropyl)-3,5-dimethylbenzamide** (**5Ma**)^[6]: ¹H NMR (500 MHz, CDCl₃) δ 7.37 (s, 2H), 7.13 (s, 1H), 6.63 (brs, 1H), 3.70 (q, J = 6.0 Hz, 2H),

[6] Tachibana, Y.; Kawasaki, H.; Kihara, N.; Takata, T. J. Org. Chem. 2006, 71, 5093.

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3.62 (q, J = 6.0 Hz, 2H), 3.36 (t, J = 6.0 Hz, 1H), 2.34 (s, 6H), 1.78 (quin, J = 6.0 Hz, 2H); ¹³C NMR (126 MHz, CDCl₃) δ 169.0, 138.3, 134.1, 133.2, 124.7, 59.4, 36.8, 32.3, 21.2; HRMS (ESI) m/z calcd for $C_{12}H_{17}NNaO_2$ [M+Na]⁺: 230.1151, found 230.1150.

6. Mechanistic Considerations

6-1. Isolation of reaction intermediates in the Rh-catalyzed hydrosilylation

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added *N*-cyclopropylpivalamide (49.4 mg, 0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, mmol, 8.8 μmol), and (P(1-nap)₃) (14.4 mg, mmol, 35.0 μmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, *tert*-butyldimethylsilane (0.11 mL, 0.70 mmol) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the vessel was taken out of the glovebox. The mixture was heated at 110 °C for 3 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard. NMR yields: **3Ab**: 8%, **4Ab**: 38%, (*E*)-**6A**: 35%, (*Z*)-**6A**: 21%

The residue was purified by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) to afford $\mathbf{4Ab}$ (32.9 mg, 37%, a white solid), (*E*)- $\mathbf{6A}$ (16.4 mg, 33%, a white solid) and (*Z*)- $\mathbf{6A}$ (8.0 mg, 16%, a white solid).

(*E*)-*N*-(**Prop-1-en-1-yl**)**pivalamide** ((*E*)-6A)^[7]: ¹H NMR (600 MHz, CDCl₃) δ 7.06 (brs, 1H), 6.79–6.73 (m, 1H), 5.19–5.12 (m, 1H), 1.68 (dd, J = 7.2, 1.2 Hz, 3H), 1.22 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 175.2, 123.6, 107.5, 38.6, 27.4, 14.8; HRMS (ESI) m/z calcd for C₈H₁₅NNaO [M+Na]⁺: 164.1046, found 164.1047.

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^[7] Halli, J.; Kramer, P.; Bechthold, M.; Manolikakes, G. Adv. Synth. Catal. 2015, 357, 3321.

(*Z*)-*N*-(**Prop-1-en-1-yl)pivalamide** ((*Z*)-6A)^[7]: ¹H NMR (600 MHz, CDCl₃) δ 7.13 (brs, 1H), 6.76–6.70 (m, 1H), 4.85–4.78 (m, 1H), 1.62 (d, *J* = 6.6 Hz, 3H), 1.25 (s, 9H); ¹³C NMR (151 MHz, CDCl₃) δ 175.3, 122.3, 104.9, 38.9, 27.5, 10.7; HRMS (ESI) m/z calcd for C₈H₁₅NNaO [M+Na]⁺: 164.1046, found 164.1048.

6-2. Hydrosilylation of reaction intermediates (E)-6A and (Z)-6A

6-2-1. Reaction of (E)-6A or (Z)-6A with $[Rh(cod)Cl]_2/P(1-nap)_3$

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added (*E*)-*N*-(prop-1-en-1-yl)pivalamide ((*E*)-6A: 60.0 mg, 0.42 mmol), [Rh(cod)Cl]₂ (5.2 mg, 0.011 mmol), and P(1-nap) 3 (17.5 mg, 0.042 mmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (2a: 0.23 mL, 0.85 mmol) and THF (2.4 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the reaction vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard.

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added (*Z*)-*N*-(prop-1-en-1-yl)pivalamide ((*Z*)-6**A**: 50.0 mg, 0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, 0.089 mmol), and P(1-nap) ₃ (14.6 mg, 0.035 mmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (2a: 0.19 mL, 0.71 mmol) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the reaction vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was

concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard.

These results indicated that the hydrosilylated product **4Aa** would be formed through Rh-catalyzed hydrosilylation of enamide **6A**.

6-2-2. Reaction of allylamine 7A with [Rh(cod)Cl]₂/P(1-nap)₃

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added *N*-allylpivalamide^[8] (7A: 49.4 mg, 0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, 0.0088 mmol), and P(1-nap)₃ (14.4 mg, 0.035 mmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (2a: 0.19 mL, 0.70 mmol) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the reaction vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard.

The result suggested that the isomerization of allylamine to enamide took place *in situ* in the presence of $P(1-nap)_3$.

6-2-3. Reaction of (E)-6A or (Z)-6A with [Rh(cod)Cl]₂/PCy₃·HBF₄

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added (E)-N-(prop-1-en-1-yl)pivalamide ((E)-A: 70.0 mg, 0.50 mmol), [Rh(cod)Cl]₂ (6.1 mg, 0.012 mmol), and PCy₃·HBF₄ (18.3 mg, 0.050 mmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (2a: 0.27 mL, 0.99

^[8] Moon, N. G.; Harned, A. M. Tetrahedron Lett. 2013, 54, 2960.

mmol) and THF (2.8 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the reaction vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard.

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added (*Z*)-*N*-(prop-1-en-1-yl)pivalamide ((*Z*)-6A: 68.0 mg, 0.48 mmol), [Rh(cod)Cl]₂ (5.9 mg, 0.012 mmol), and PCy₃·HBF₄ (17.7 mg, 0.048 mmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (2a: 0.26 mL, 0.96 mmol) and THF (2.8 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the reaction vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard.

These results showed that hydrogenated product **8A** was formed when PCy₃·HBF₄ was used as the ligand in the reaction of enamide **6A** and that hydrosilylation of enamide **6A** was suppressed.

6-2-4. Reaction of allylamine 7A with [Rh(cod)Cl]₂/PCy₃·HBF₄

A 20 mL glass vessel tube equipped with a J. Young[®] O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added *N*-allylpivalamide^[8] (7A: 49.4 mg, 0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, 0.0088

^[8] Moon, N. G.; Harned, A. M. Tetrahedron Lett. 2013, 54, 2960.

Rh-Catalyzed Regiodivergent Hydrosilylation of Acyl aminocyclopropanes Controlled by Monophosphine Ligands (Kondo, Itami, Yamaguchi)

mmol), and PCy₃·HBF₄ (12.9 mg, 0.035 mmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (**2a**: 0.19 mL, 0.70 mmol) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the reaction vessel was taken out of the glovebox. The mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard. The residue was purified by Isolera® (hexane/ethyl acetate = 1:0 to 5:1) to afford **3Aa** (52.7 mg, 41%) as a colorless oil.

This result indicated that the formation of linear product 3 arises from the hydrosilylation of allylamine.

6-3. Isomerization of allylamine to enamide under [Rh(cod)Cl]₂/P(1-nap)₃ catalytic condition

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added [Rh(cod)Cl]₂ (2.2 mg, 0.0044 mmol), P(1-nap)₃ (7.2 mg, 0.018 mmol) and THF (2.0 mL), then *N*-allylpivalamide^[8] (7A: 49.4 mg, 0.35 mmol) was added under nitrogen. After the vessel was sealed with the O-ring tap, the mixture was heated at 110 °C for 6 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The NMR yield was determined by ¹H NMR analysis of the crude product using dibromomethane as an internal standard.

This result showed that allylamine is isomerized to enamide in the presence of $[Rh(cod)Cl]_2$ and $P(1-nap)_3$. The ligand effect of $P(1-nap)_3$ is supposed to control isomerization of allylamine to enamide.

^[8] Moon, N. G.; Harned, A. M. Tetrahedron Lett. 2013, 54, 2960.

7. Proposed Reaction Mechanism

$$L = P(1-nap)_3$$

$$Rh^{\parallel}Ln$$

$$Si = Rh^{\parallel}\parallel Ln$$

$$Rh^{\parallel}Ln$$

$$Rh^{\parallel}L$$

$$\begin{array}{c} \mathsf{Rh}^{\parallel} \mathsf{L} \mathsf{D} \\ \mathsf{Si-Rh}^{\parallel} \mathsf{L} \mathsf{D} \\ \mathsf{Rh}^{\parallel} \mathsf{L} \mathsf{D} \\ \mathsf{L} \mathsf{L} \mathsf{L} \mathsf{L} \mathsf{L} \mathsf{L} \mathsf{L} \\ \mathsf{L} \mathsf{L} \mathsf{L} \mathsf{L} \\ \mathsf{L} \mathsf{L} \mathsf{L} \\ \mathsf$$

8. Effect of Reaction Parameters

[Rh(cod)Cl]₂/P(1-nap)₃ catalytic system

Entry	[Rh]	X mol%	Solvent	T °C	NMR yield (3Aa)	NMR yield (4Aa)
1	[Rh(cod)Cl] ₂	5 mol%	THF	120 °C	20%	75%
2	[Rh(cod)Cl] ₂	10 mol%	THF	120 °C	9% (4%) ^a	82% (81%) ^a
3	[Rh(cod)Cl] ₂	15 mol%	THF	120 °C	9%	81%
4	[Rh(cod)Cl] ₂	20 mol%	THF	120 °C	8%	76%
5	[Rh(cod)Cl] ₂	10 mol%	THF	110 °C	10%	80%
6	[Rh(cod)Cl] ₂	10 mol%	THF	100 °C	10%	65%
7	[Rh(cod)OMe] ₂	10 mol%	THF	110 °C	9%	91% (88%)ª
8	[Rh(PPh ₃) ₃ Cl]	5 mol%	THF	120 °C	3%	10%
9	[Rh(cod)Cl] ₂	10 mol%	cyclohexane	120 °C	14%	80%
10	[Rh(cod)Cl] ₂	10 mol%	toluene	120 °C	14%	75%

NMR yield was determined by ¹H NMR analysis of crude products using dibromomethane as an internal standard. ^alsolated yield.

[Rh(cod)Cl]₂/PCy₃ catalytic system

Entry	[Rh]	X mol%	Solvent	T °C	NMR yield (3Aa)	NMR yield (4Aa)
1	[Rh(cod)Cl] ₂	5 mol%	THF	120 °C	54%	<1%
2	[Rh(cod)Cl] ₂	10 mol%	THF	120 °C	61% (56%) ^a	<1%
3	[Rh(cod)Cl] ₂	15 mol%	THF	120 °C	50%	<1%
4	[Rh(cod)Cl] ₂	20 mol%	THF	120 °C	42%	<1%
5	[Rh(cod)Cl] ₂	5 mol%	THF	110 °C	60%	<1%
6	[Rh(cod)Cl] ₂	5 mol%	THF	100 °C	57%	<1%
7	[Rh(cod)Cl] ₂	10 mol%	THF	110 °C	60%	3%
8	[Rh(cod)OMe] ₂	10 mol%	THF	110 °C	47%	7%
9	[Rh(cod)Cl] ₂	5 mol%	cyclohexane	110 °C	58%	7%
10	[Rh(cod)Cl] ₂	5 mol%	toluene	110 °C	43%	3%

NMR yield was determined by ¹H NMR analysis of crude products using dibromomethane as an internal standard. ^aIsolated yield.

In all reactions, the formation of $\emph{N}\text{-}\text{propylpivalamide}$ was observed.

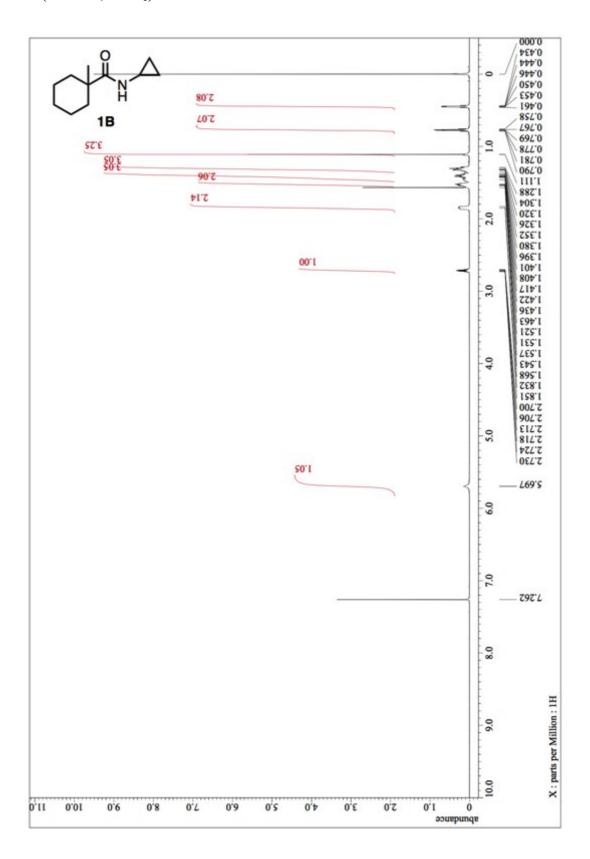
9. Discovery of Rh-catalyzed Hydrosilylation

A 20 mL glass vessel tube equipped with a J. Young® O-ring tap containing a magnetic stirring bar was dried with a heatgun under reduced pressure and filled with nitrogen after cooling to room temperature. To this vessel was added *N*-cyclopropylpivalamide (**1A**: 49.5 mg, 0.35 mmol), [Rh(cod)Cl]₂ (4.3 mg, 8.8 μmol), after which it was introduced inside an argon atmosphere glovebox. In the glovebox, 1,1,1,3,5,5,5-heptamethyltrisiloxane (**2a**: 191 μL, 0.70 mmol) and THF (2.0 mL) were added to the vessel. After the vessel was sealed with the O-ring tap, the vessel was taken out of the glovebox. The mixture was heated at 120 °C for 16 h in an 8-well reaction block with stirring. After cooling the reaction mixture to room temperature, the mixture was concentrated *in vacuo*. The residue was purified by Isolera® (hexane/ethyl acetate = 5:1 to 2:1) to afford aminosilane **3Aa** (50.1 mg, 40%, colorless oil) and **9Aa** (61.8 mg, 39%, white solid).

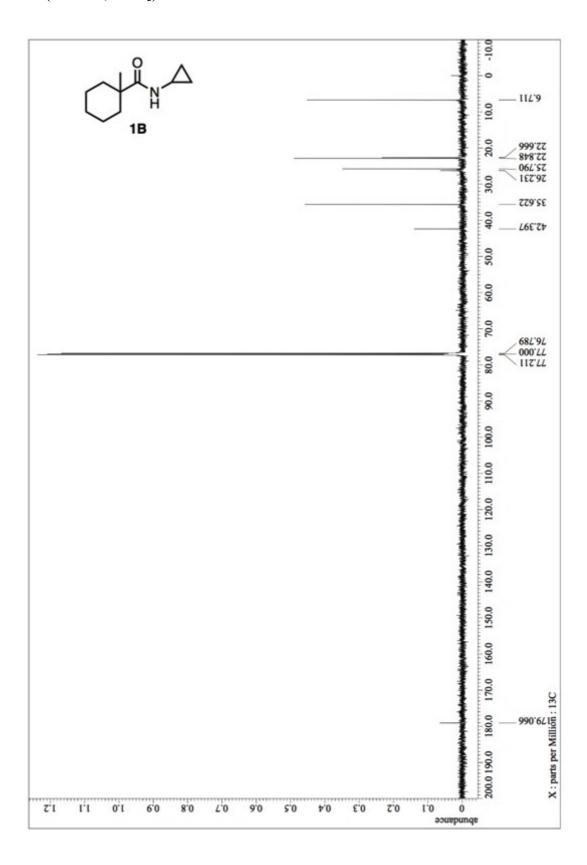
N-(3-(1,1,1,5,5,5-Hexamethyl-3-((trimethylsilyl)oxy)trisiloxan-3-yl)propyl)pivalamide (9Aa): 1 H NMR (600 MHz, CD₂Cl₂) δ 5.66 (brs, 1H), 3.17 (q, J = 6.6 Hz, 2H), 1.54–1.47 (m, 2H), 1.16 (s, 9H), 0.47–0.42 (m, 2H), 0.11 (s, 27H); 13 C NMR (151 MHz, CD₂Cl₂) δ 178.2, 42.3, 38.8, 27.8, 24.0, 12.0, 1.8; HRMS (ESI) m/z calcd for C₁₇H₄₃NNaO₄Si₄ [M+Na]⁺: 460.2161, found 460.2160.

10. ¹H and ¹³C NMR Spectra

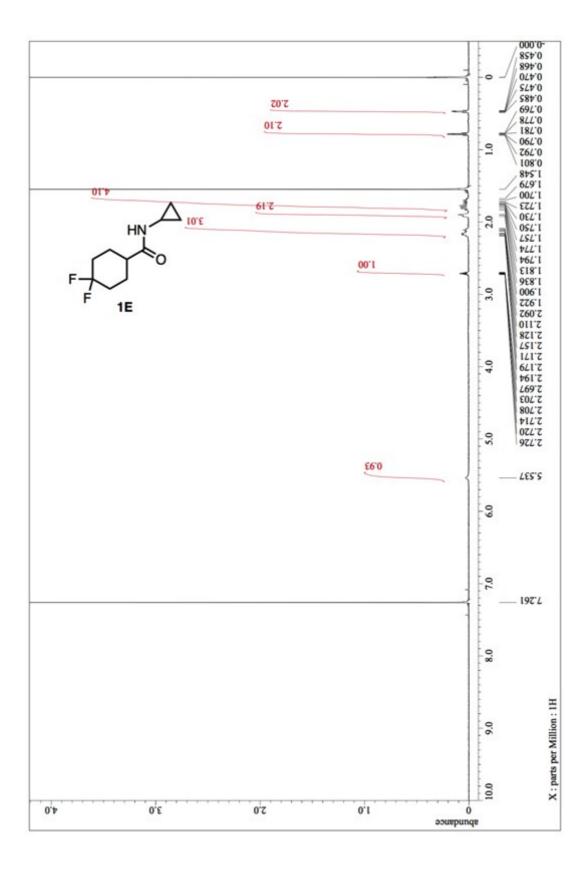
¹H NMR (600MHz, CDCl₃) of 1B



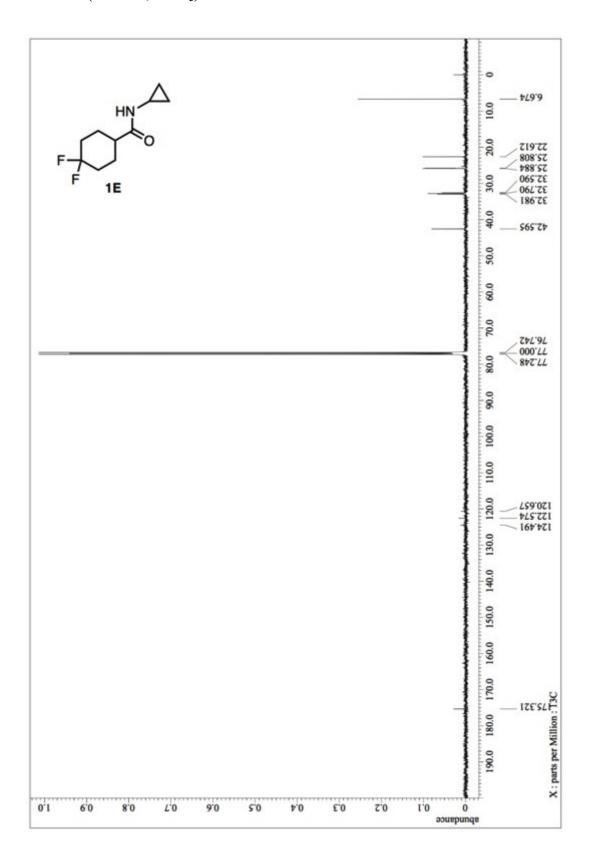
$^{13}\mathrm{C}$ NMR (151MHz, CDCl₃) of 1B



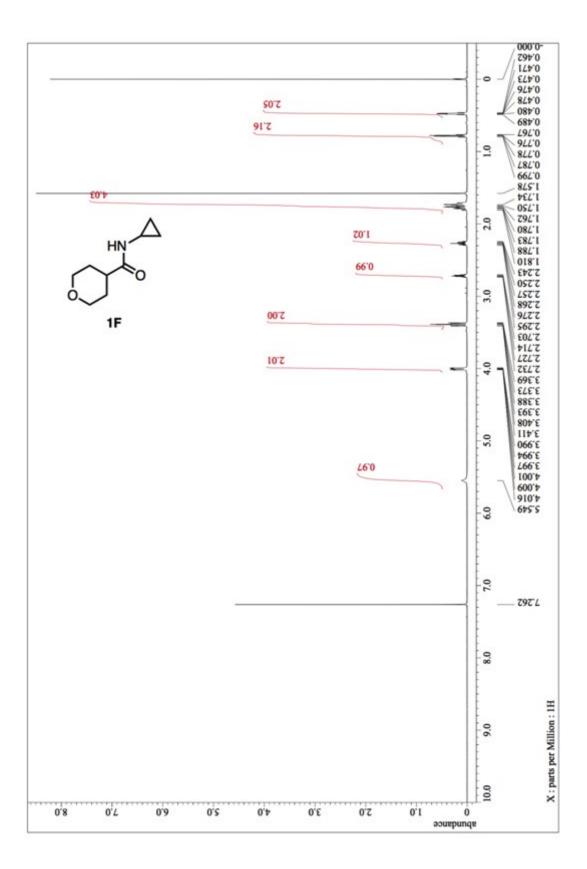
¹H NMR (600MHz, CDCl₃) of 1E



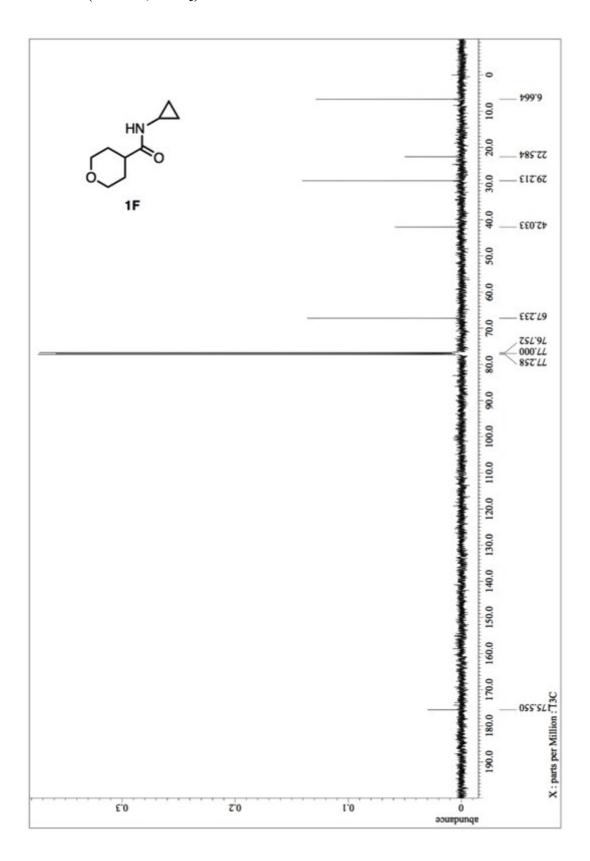
$^{13}\mathrm{C}$ NMR (126MHz, CDCl₃) of 1E



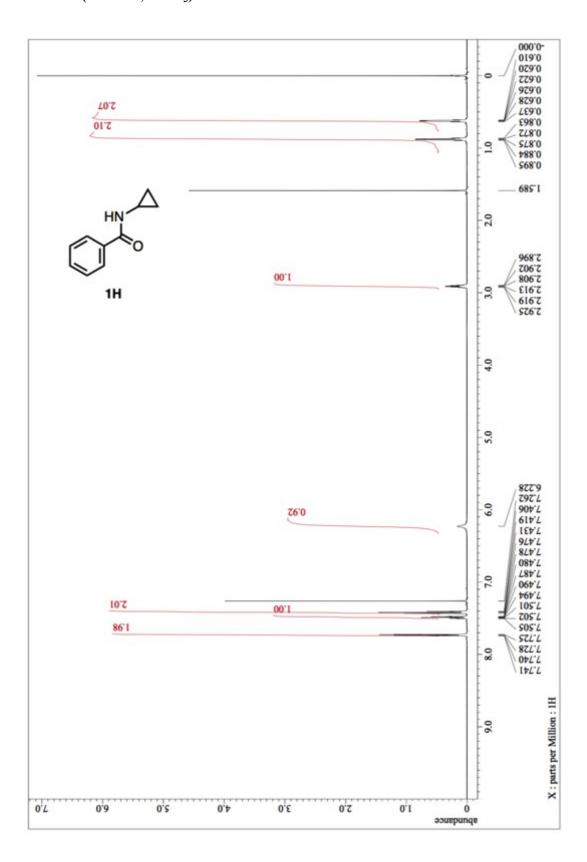
¹H NMR (600MHz, CDCl₃) of 1F



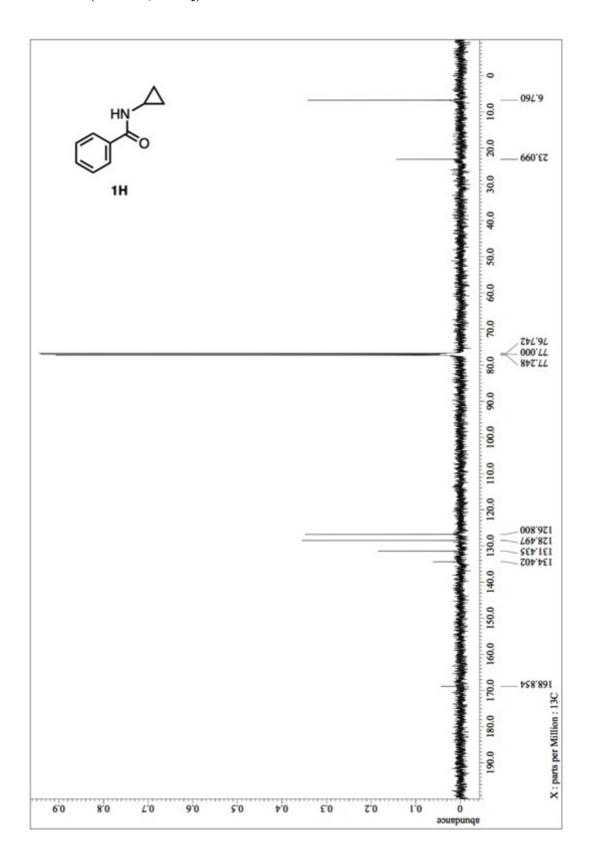
¹³C NMR (126MHz, CDCl₃) of 1F



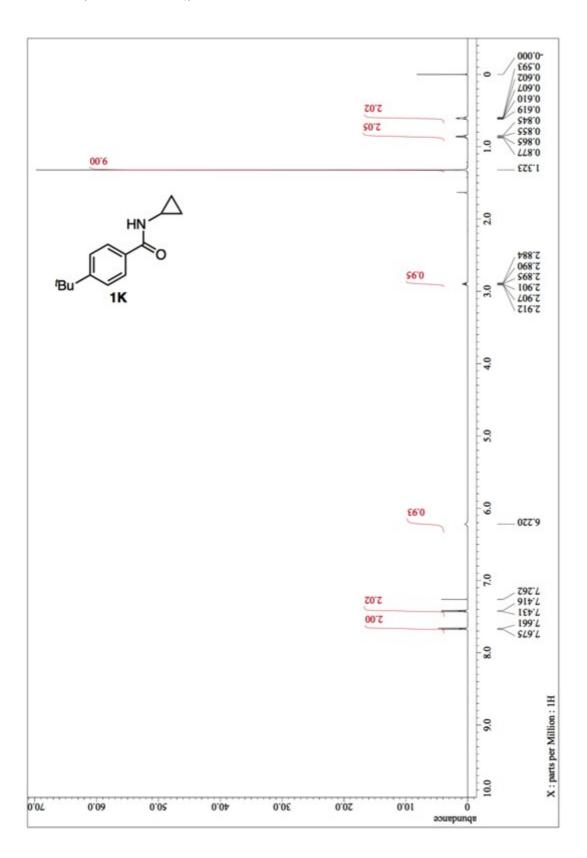
¹H NMR (600MHz, CDCl₃) of 1H



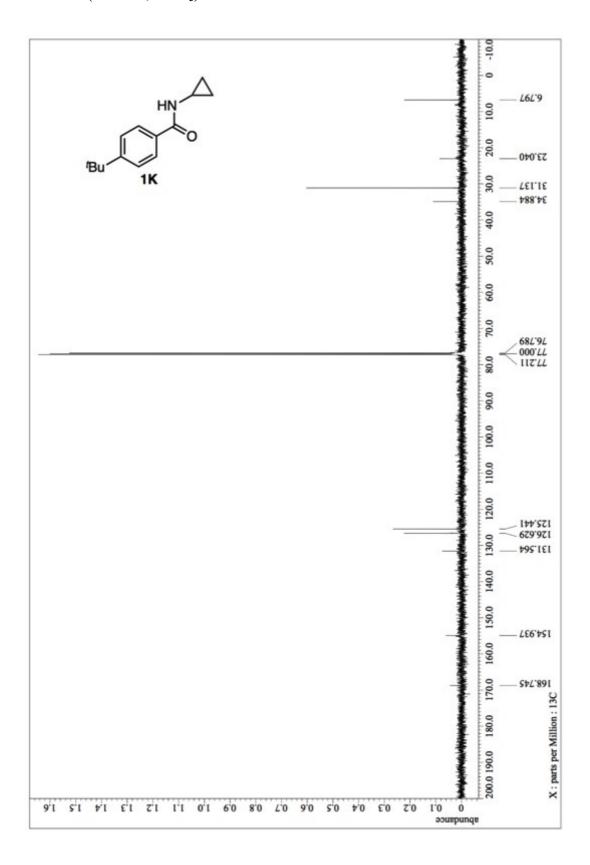
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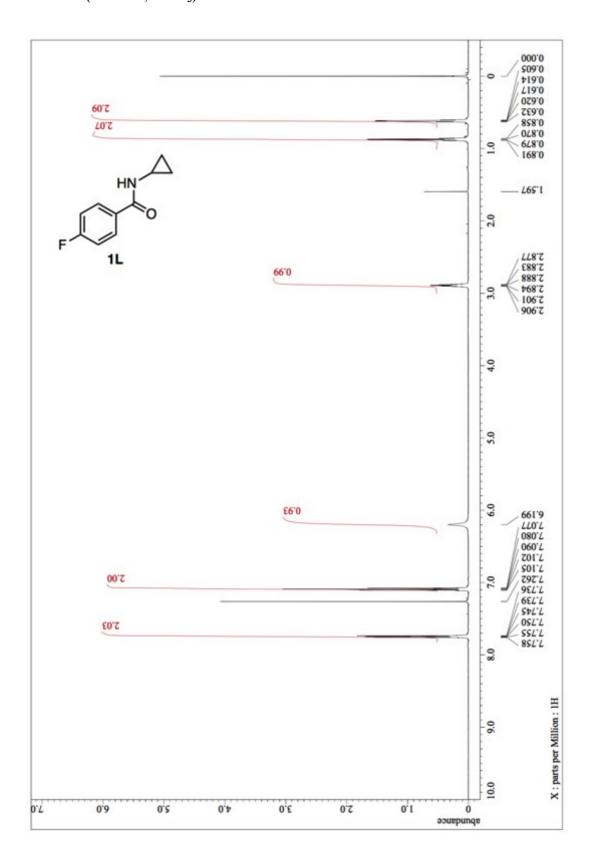
¹H NMR (600MHz, CDCl₃) of 1K



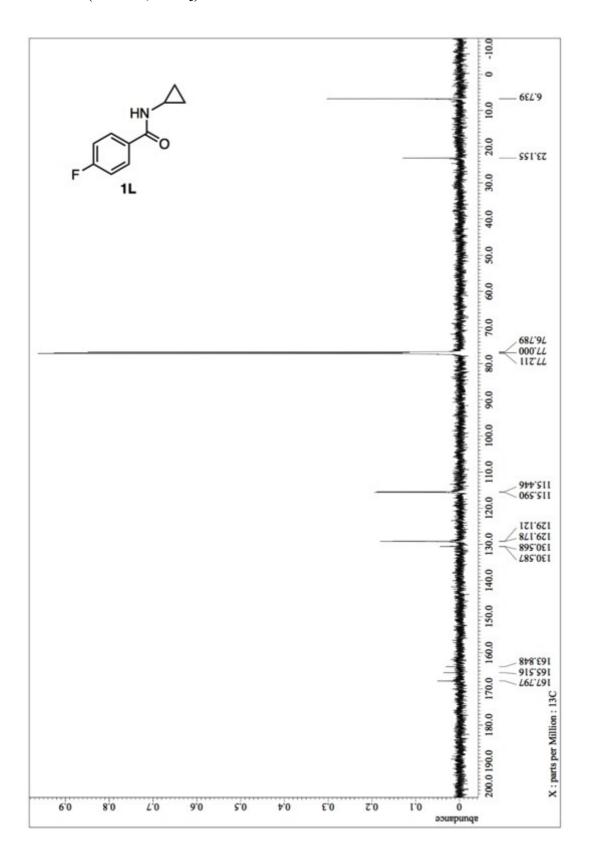
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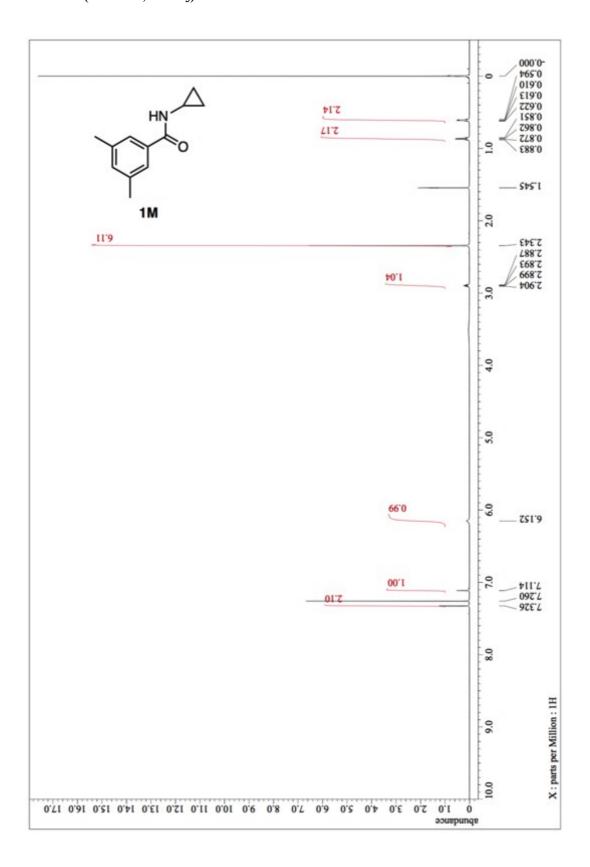
¹H NMR (600MHz, CDCl₃) of 1L



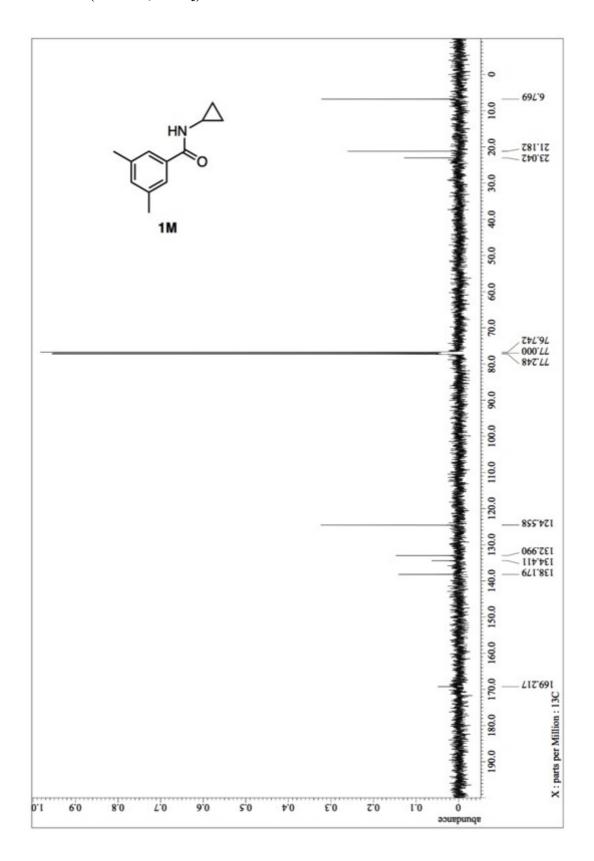
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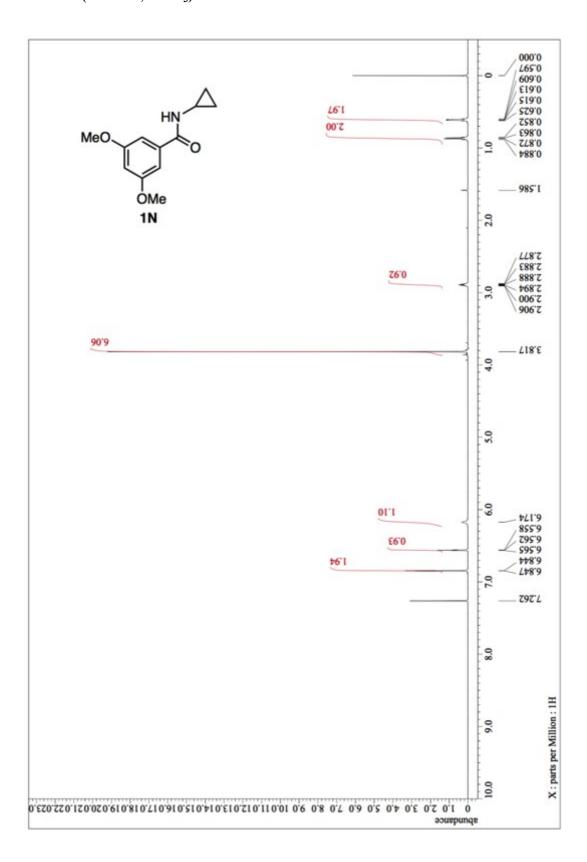
¹H NMR (600MHz, CDCl₃) of 1M



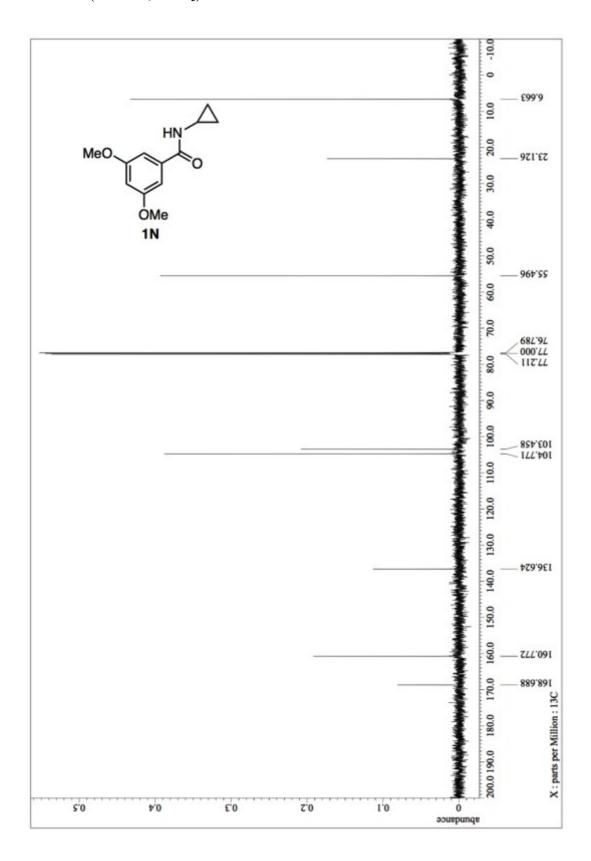
$^{13}\mathrm{C}$ NMR (126MHz, CDCl₃) of 1M



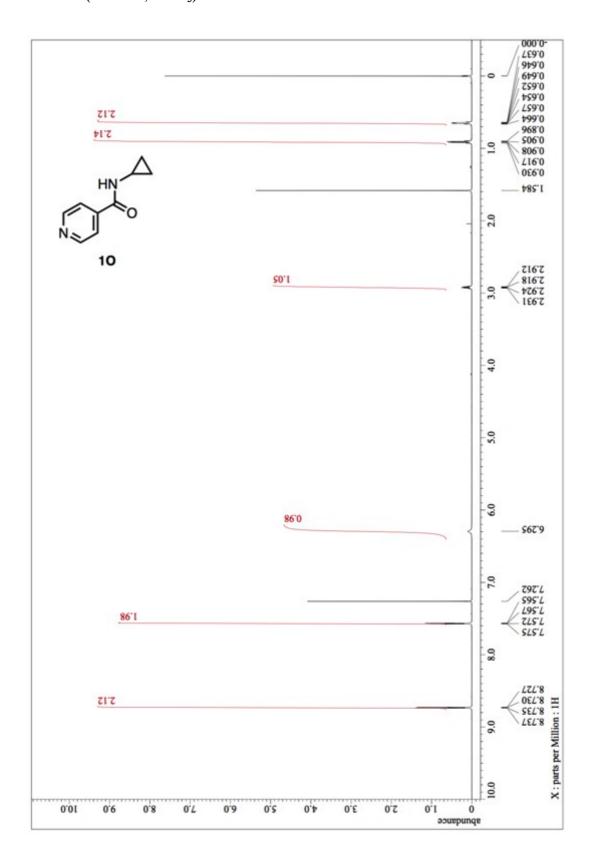
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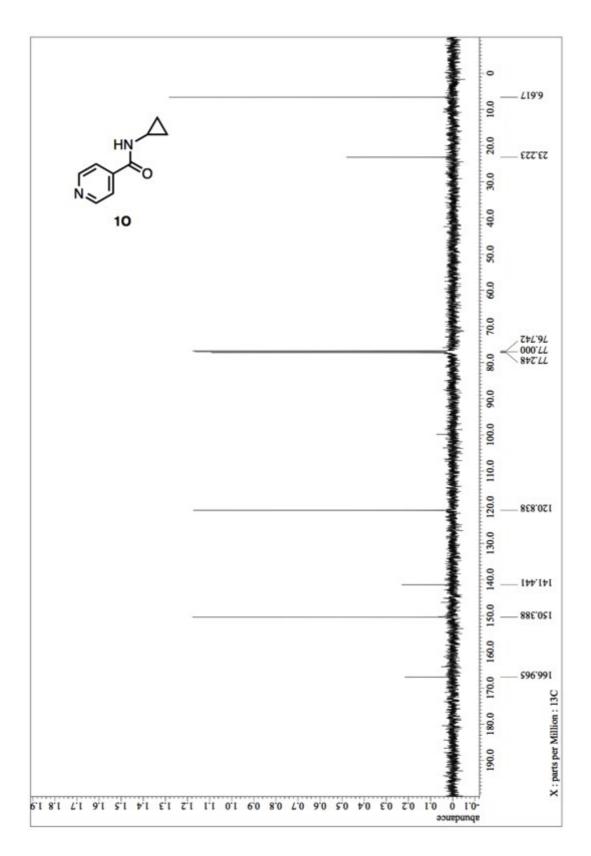
$^{13}\mathrm{C}$ NMR (151MHz, CDCl₃) of 1N



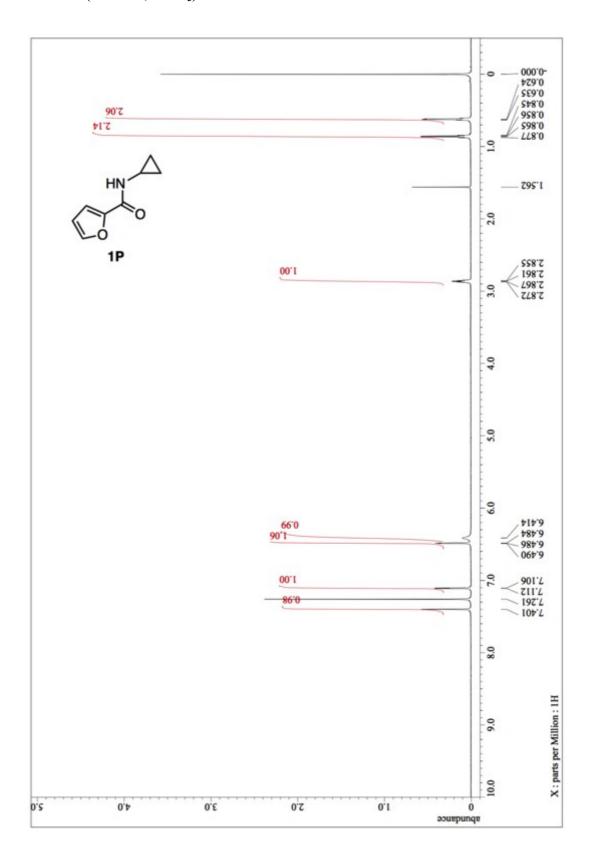
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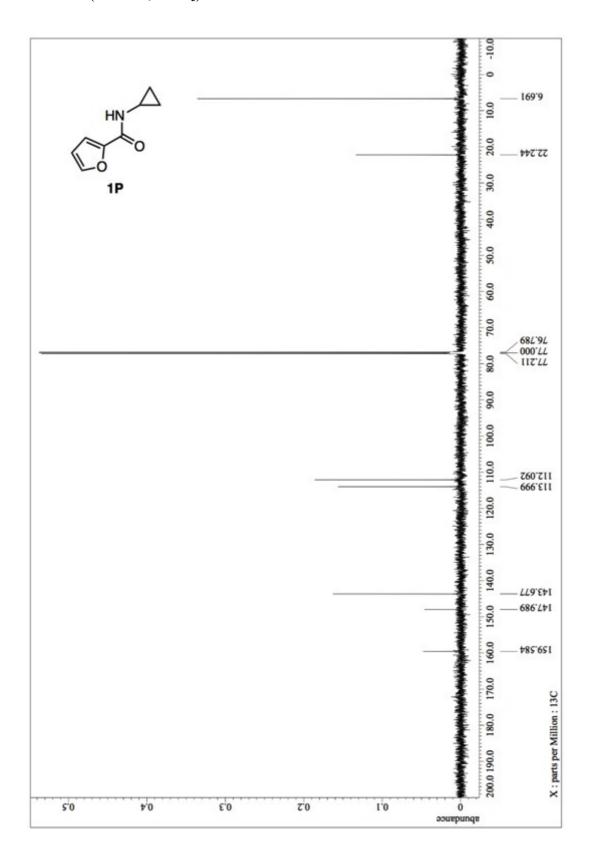
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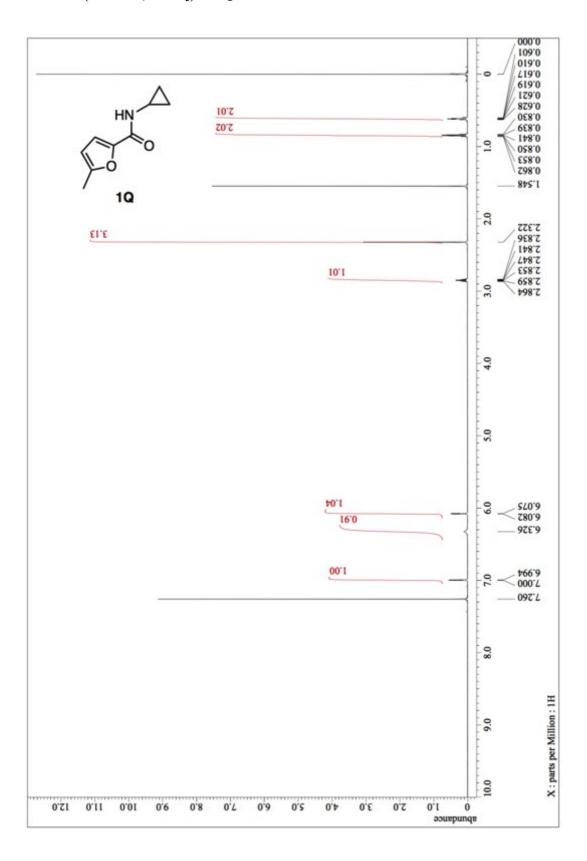
¹H NMR (600MHz, CDCl₃) of 1P



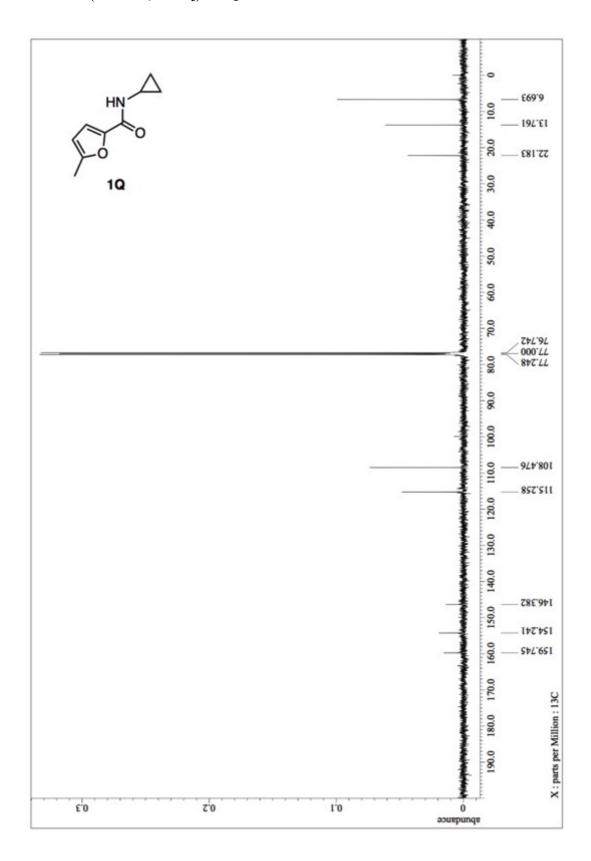
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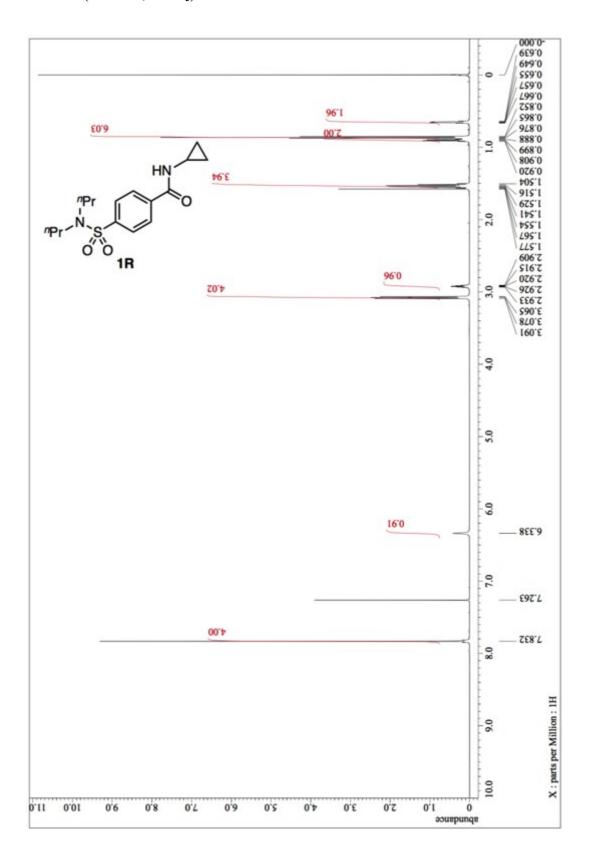
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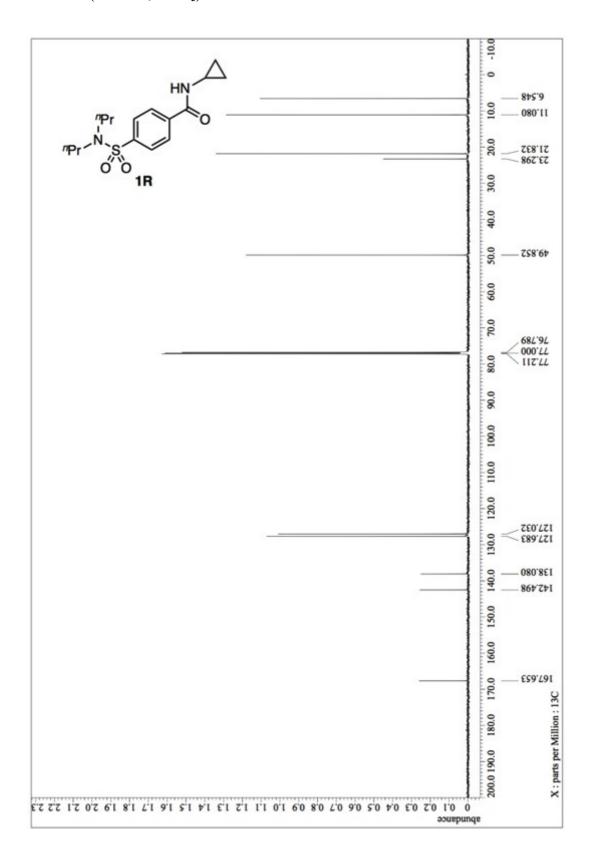
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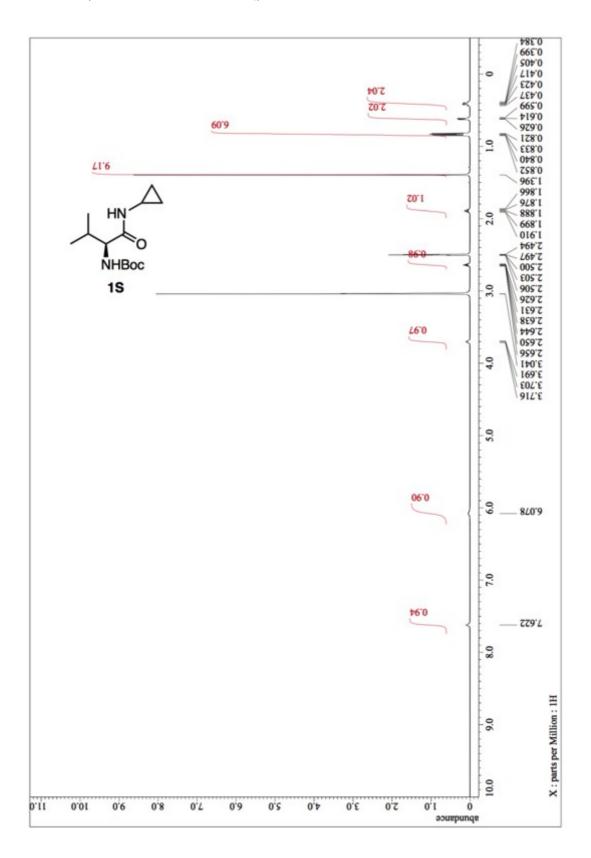
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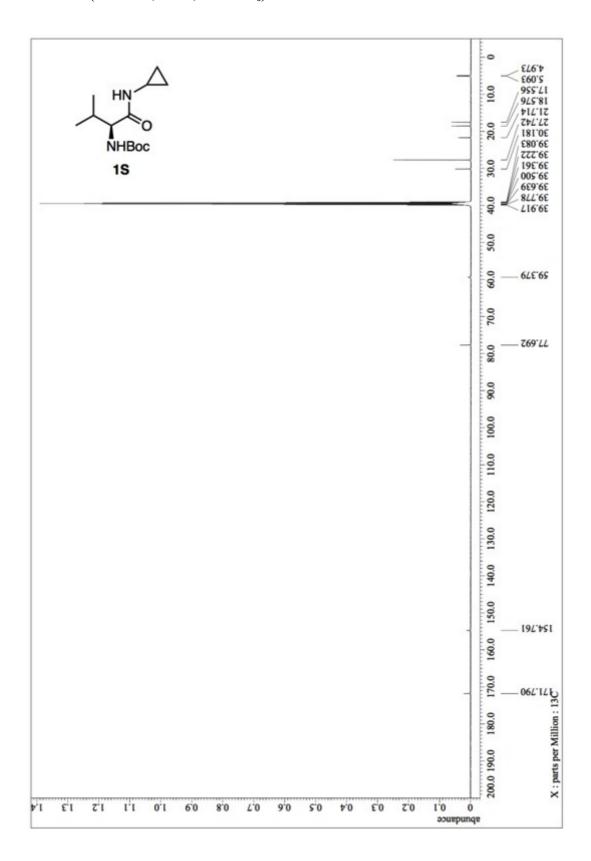
¹³C NMR (151MHz, CDCl₃) of 1R



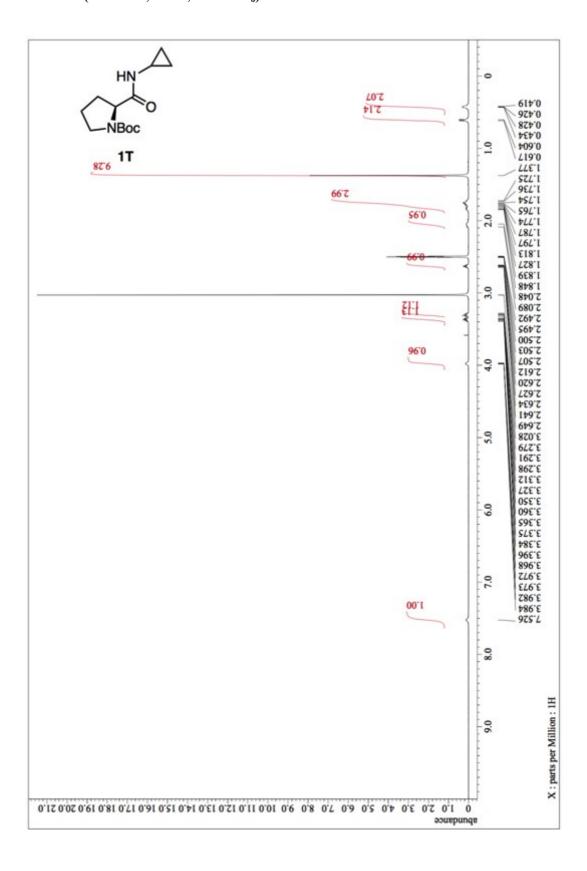
¹H NMR (600 MHz, 90 °C, DMSO-*d*₆) of 1S



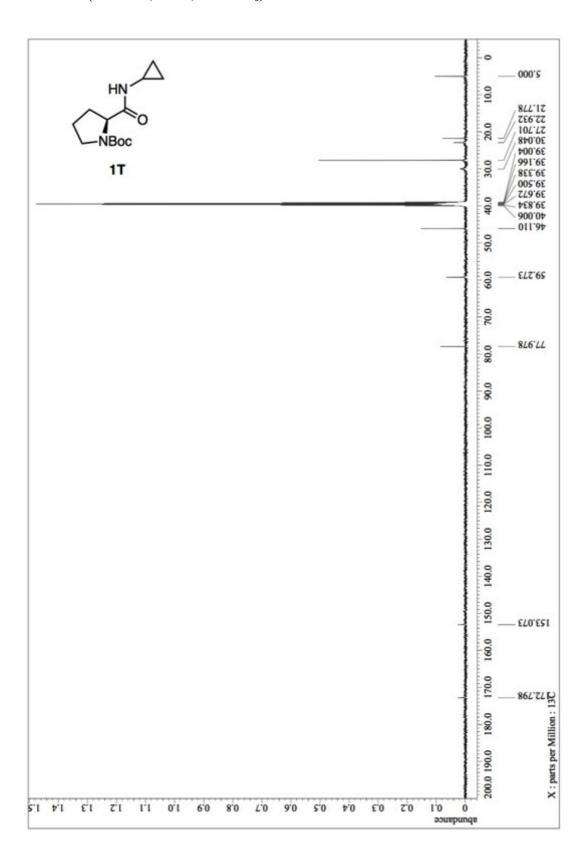
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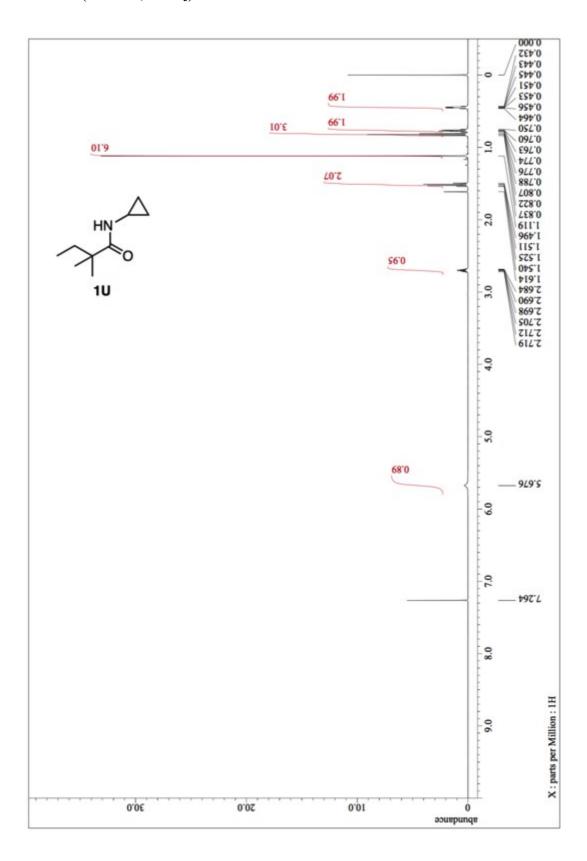
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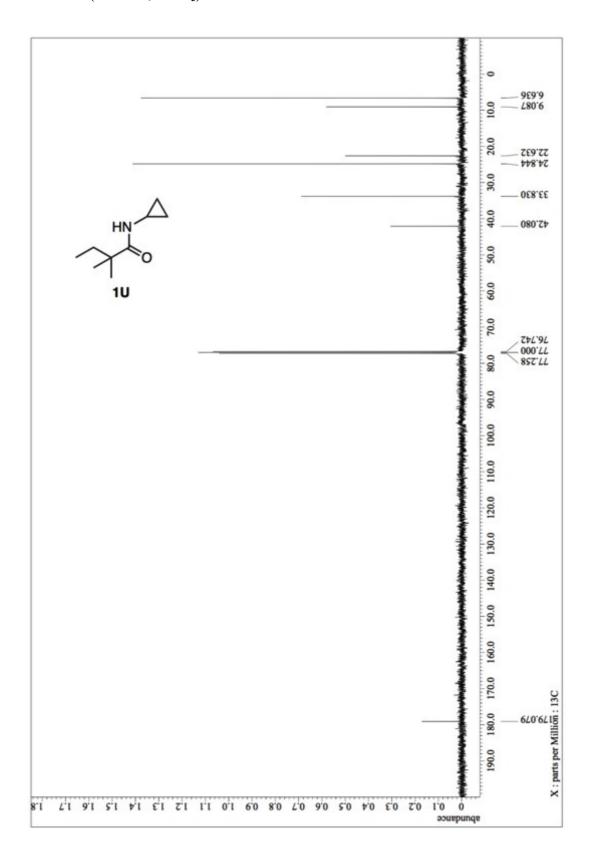
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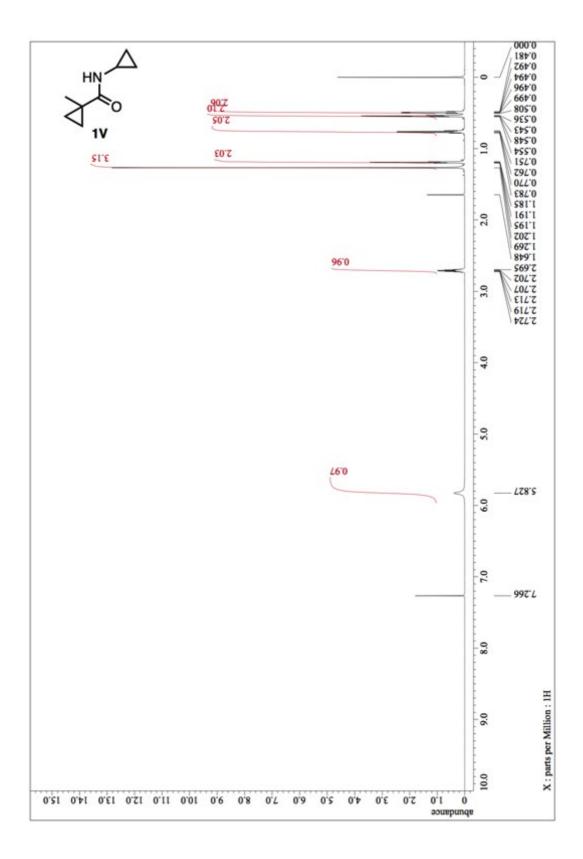
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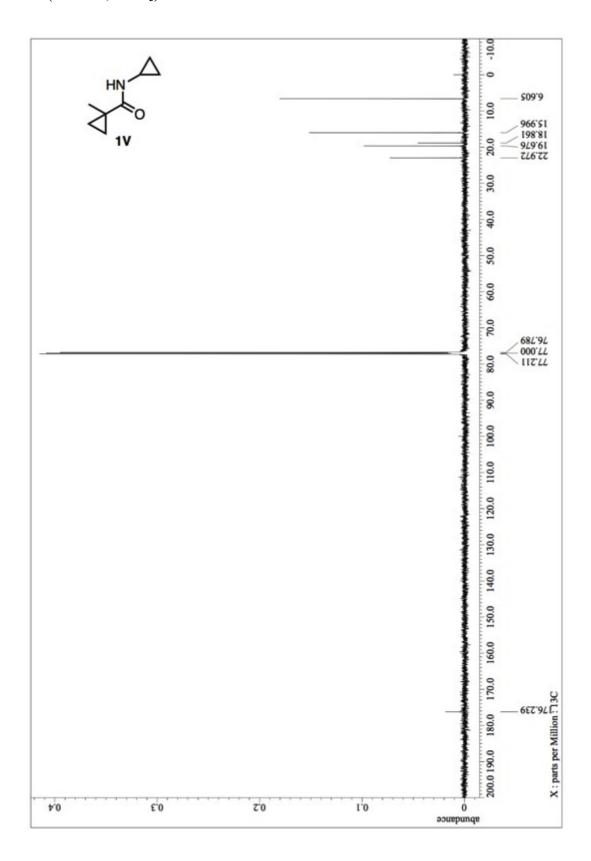
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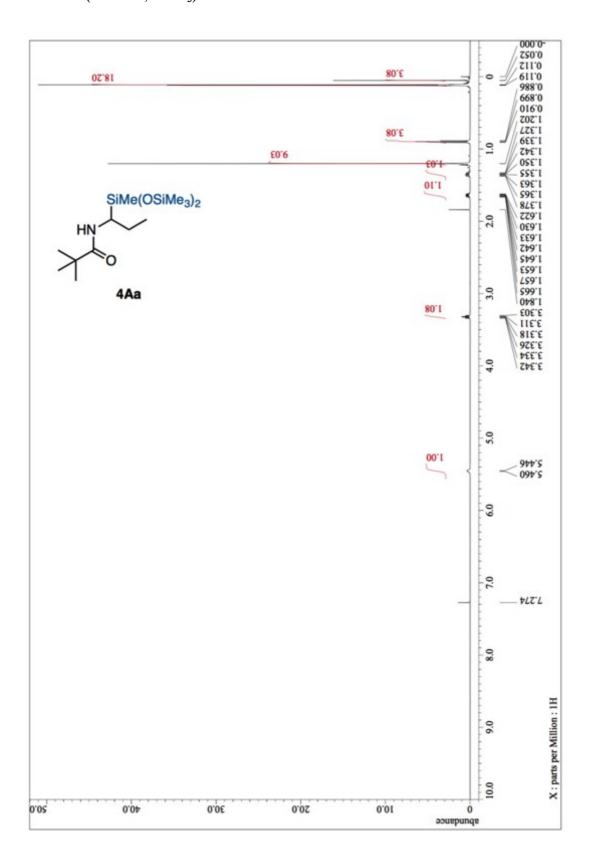
¹H NMR (600MHz, CDCl₃) of 1V



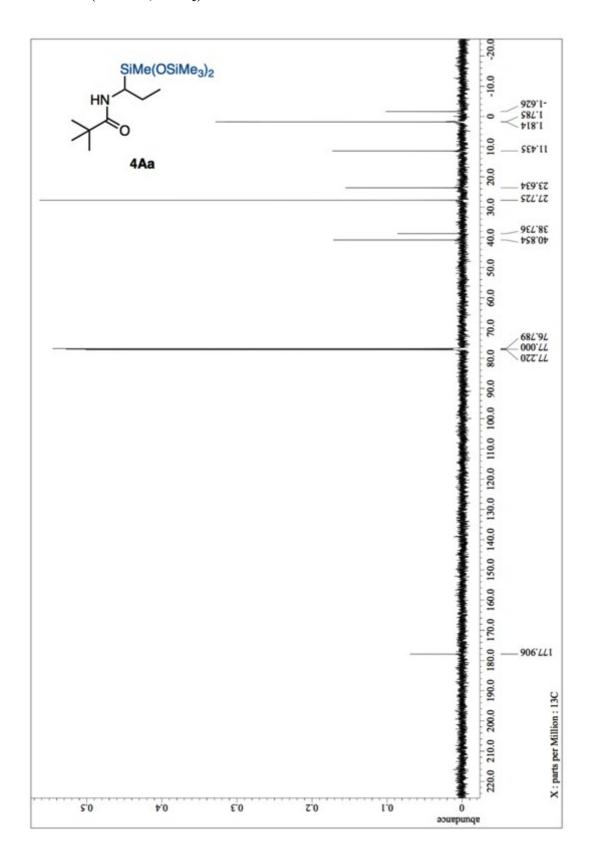
13 C NMR (151MHz, CDCl₃) of 1V



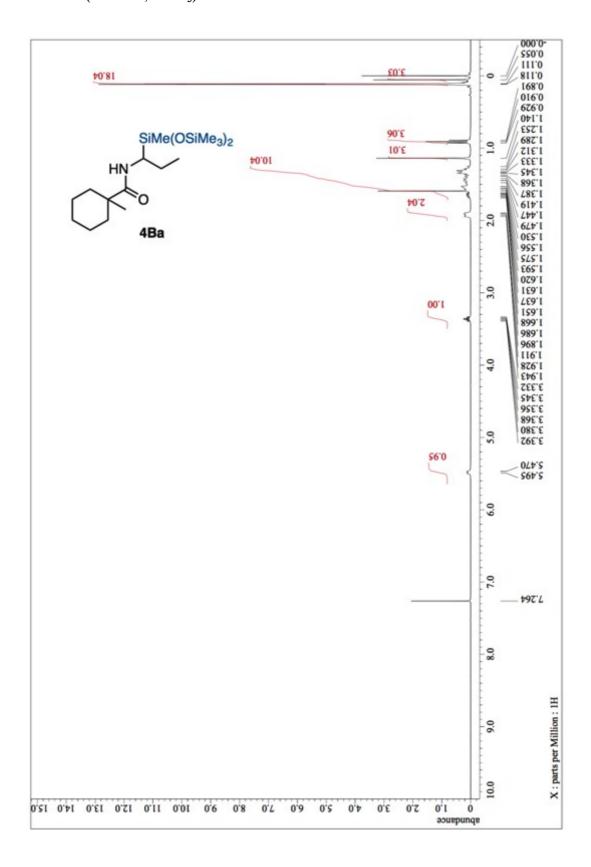
¹H NMR (600MHz, CDCl₃) of 4Aa



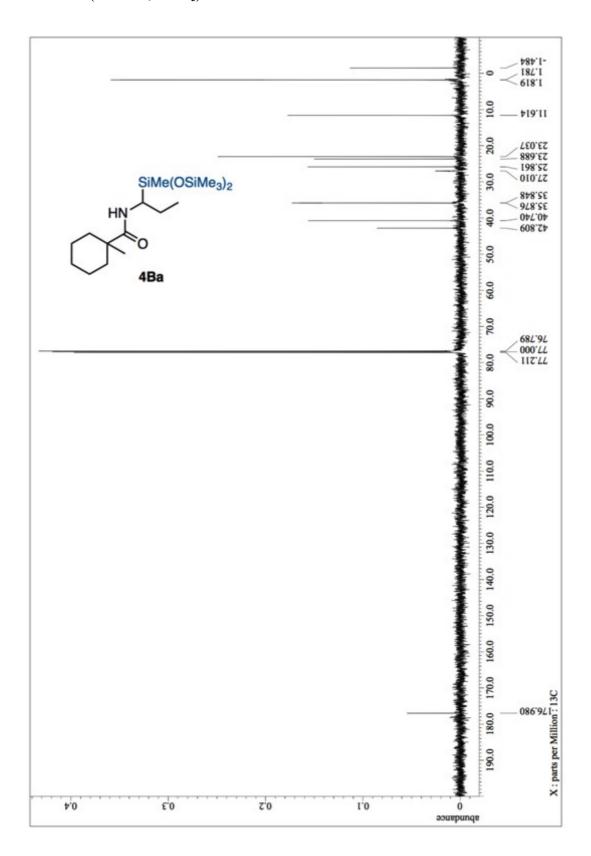
$^{13}\mathrm{C}$ NMR (151MHz, CDCl₃) of 4Aa



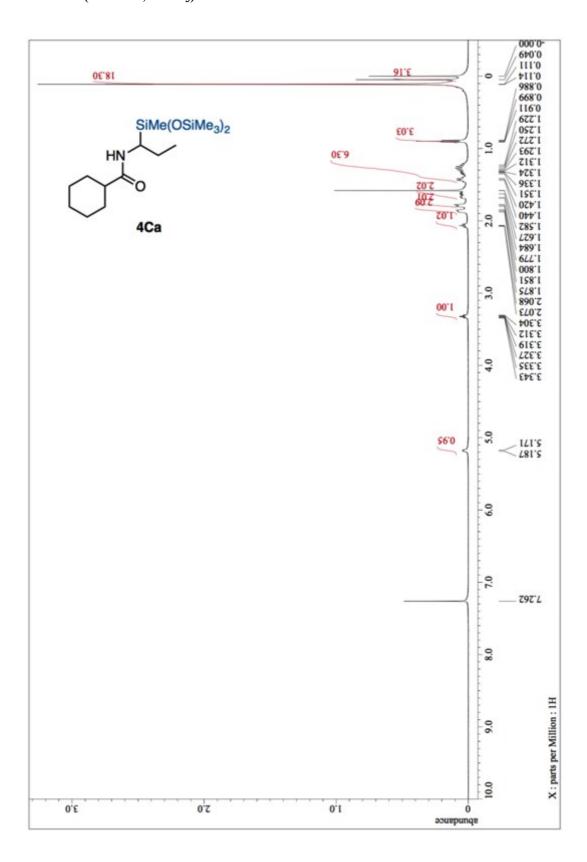
¹H NMR (400MHz, CDCl₃) of 4Ba



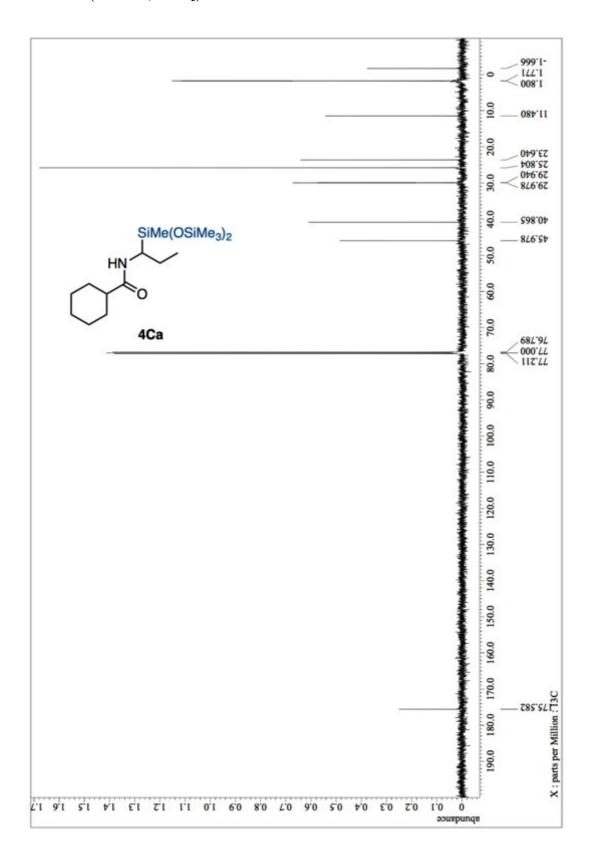
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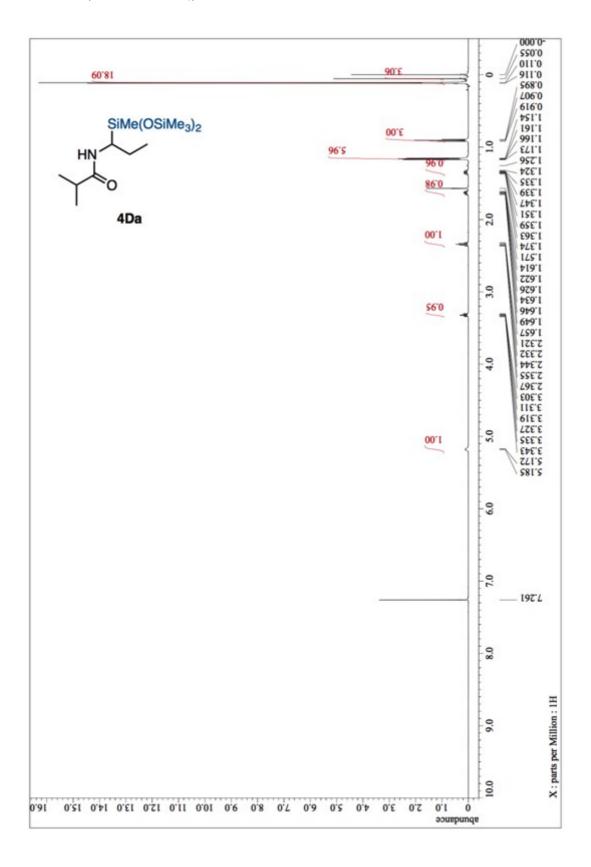
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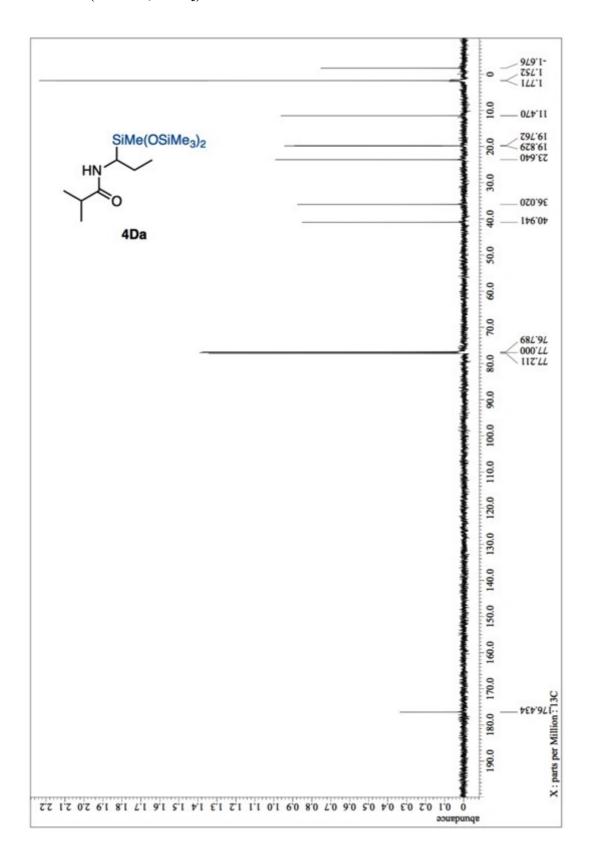
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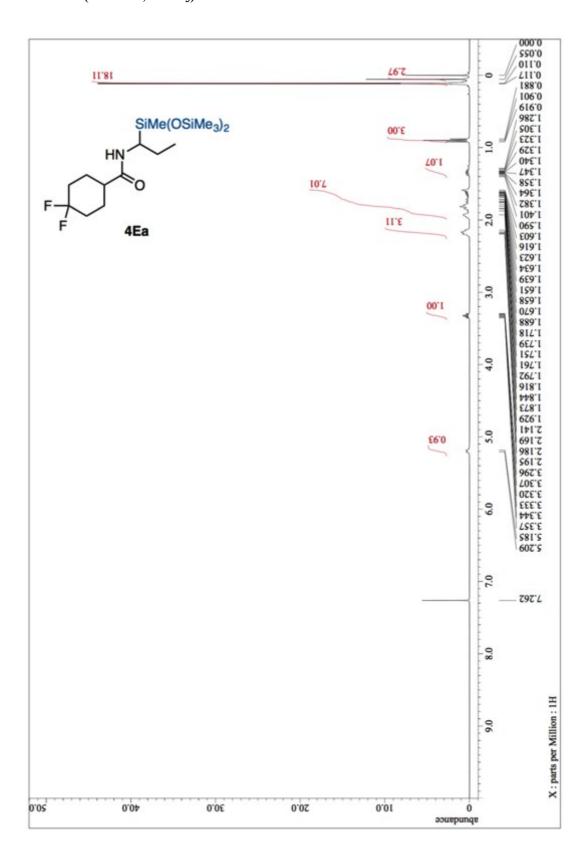
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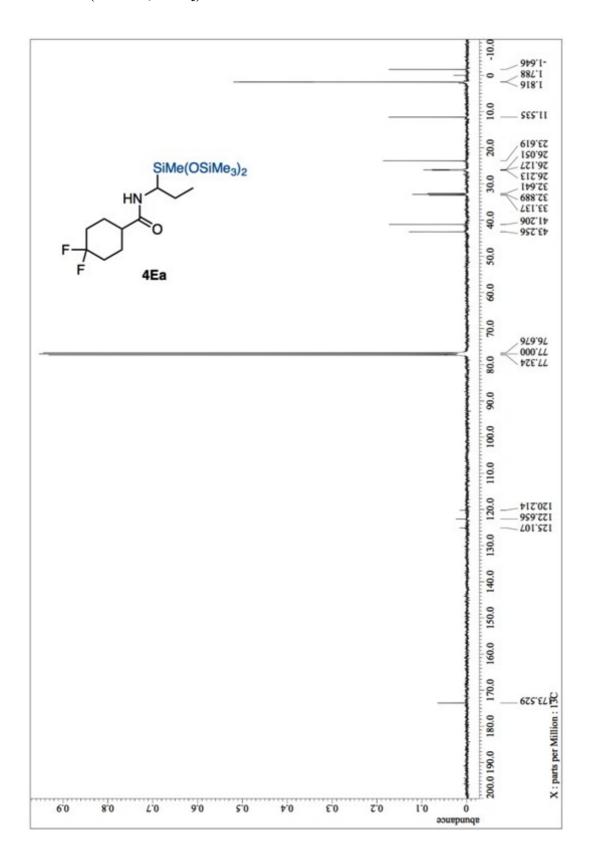
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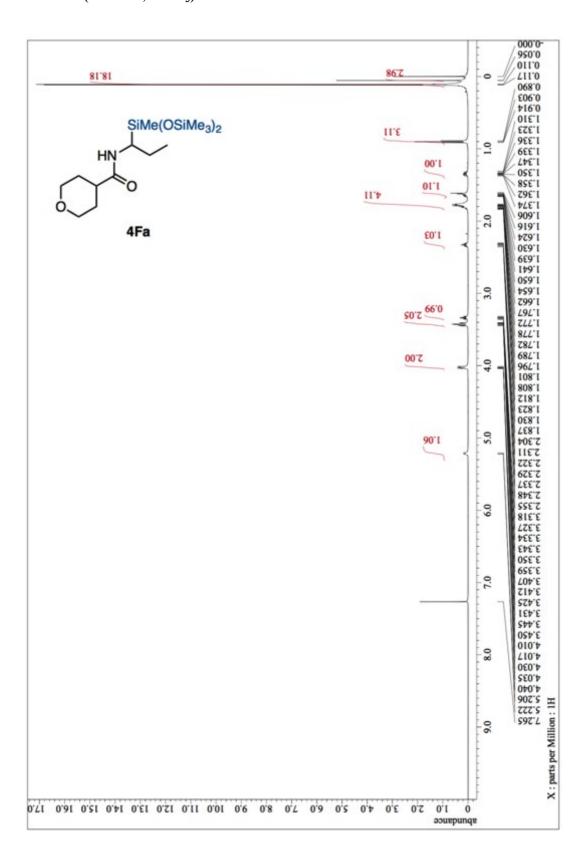
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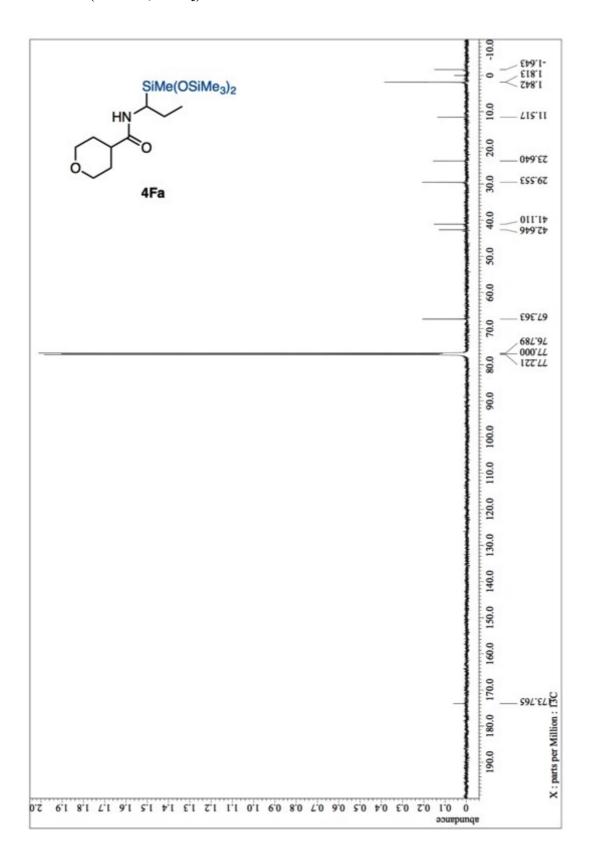
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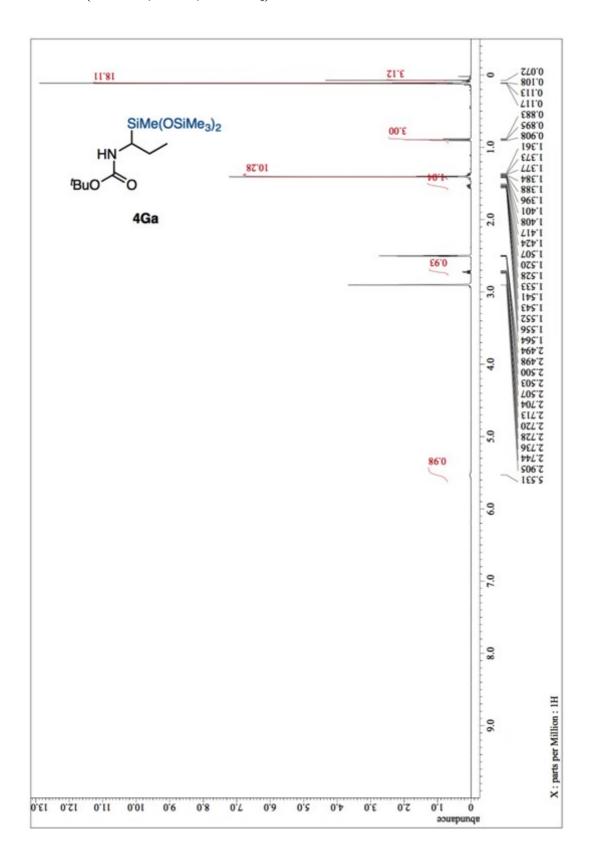
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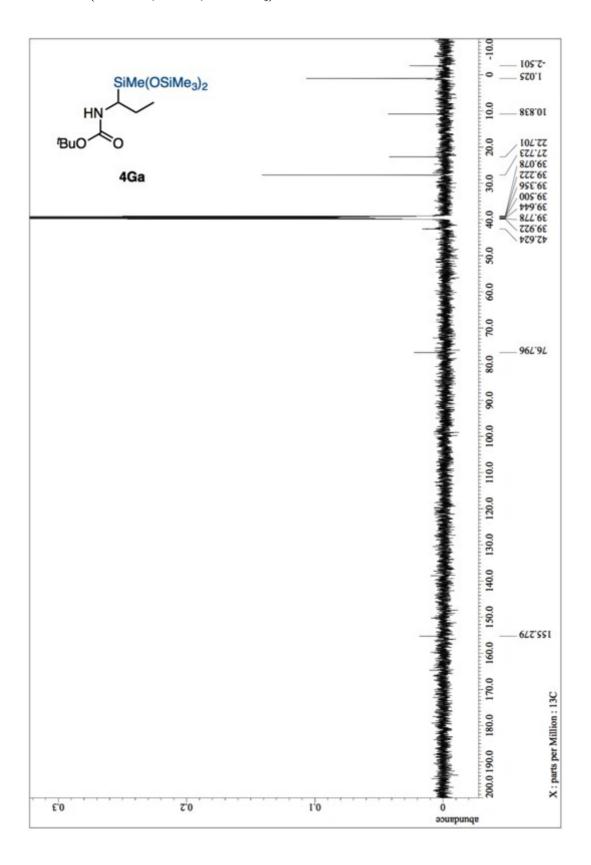
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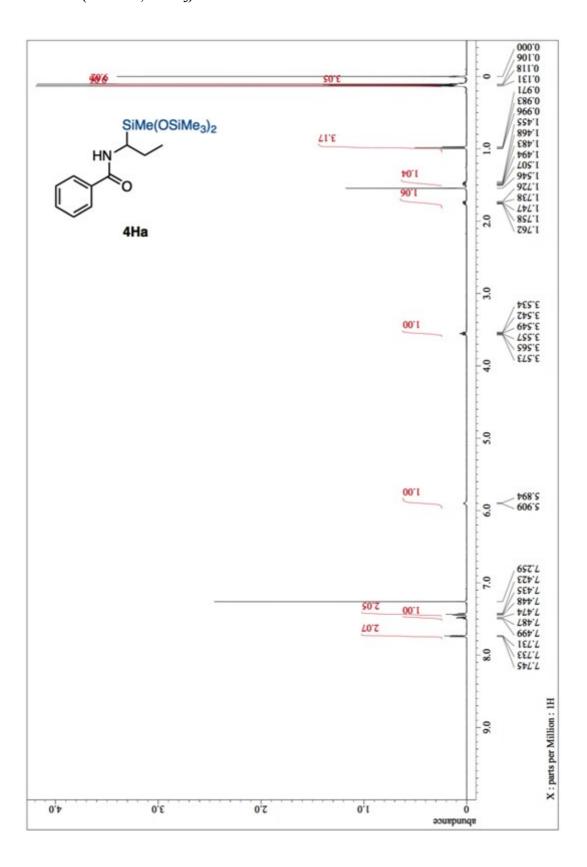
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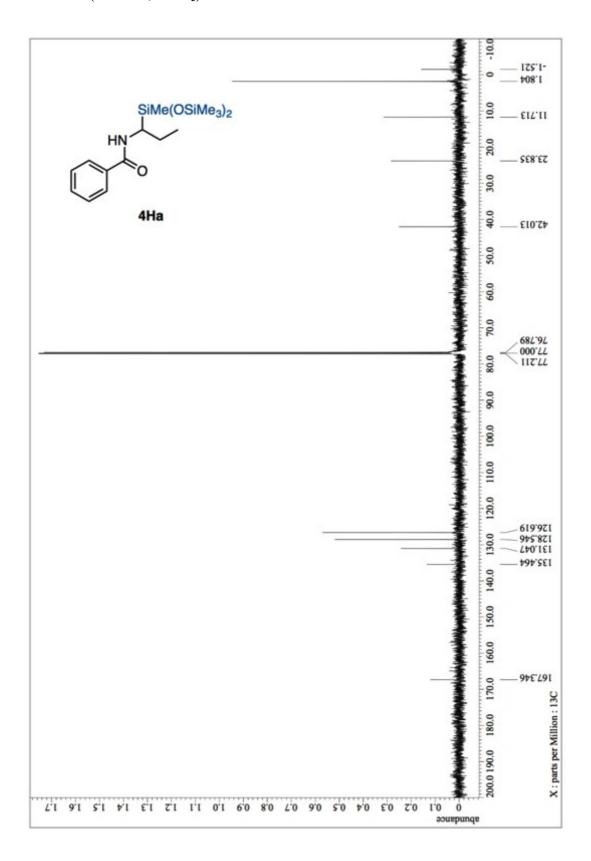
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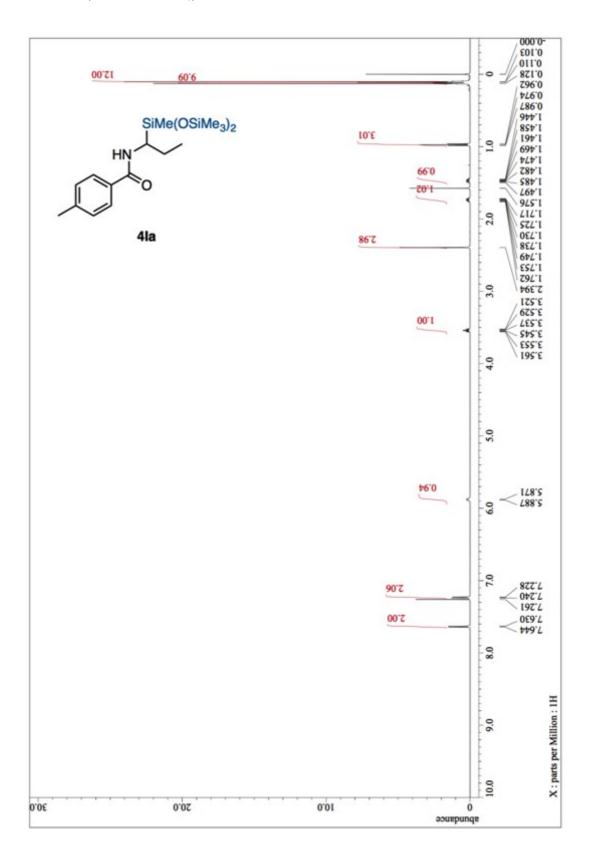
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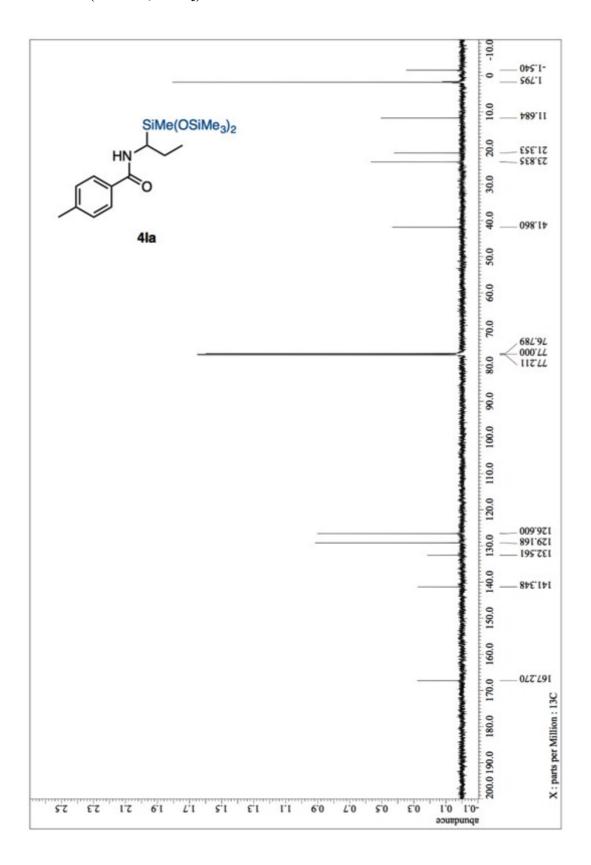
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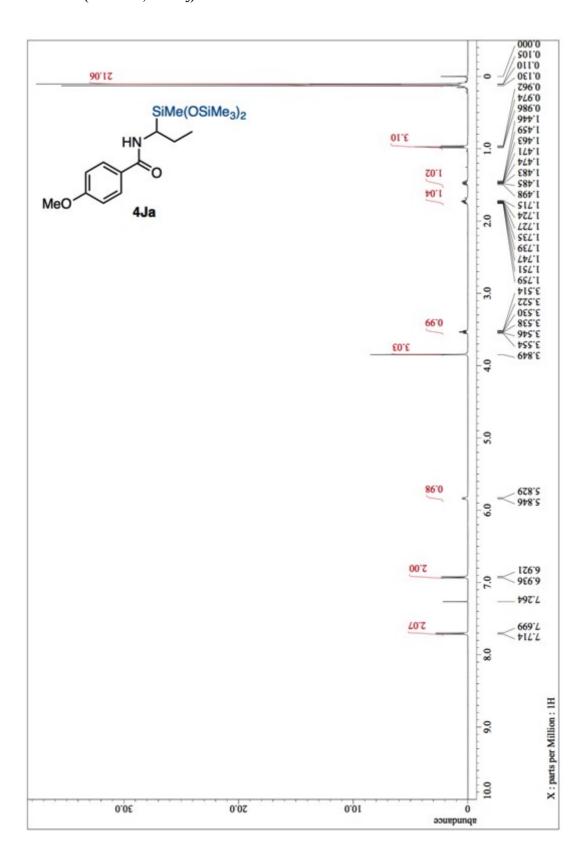
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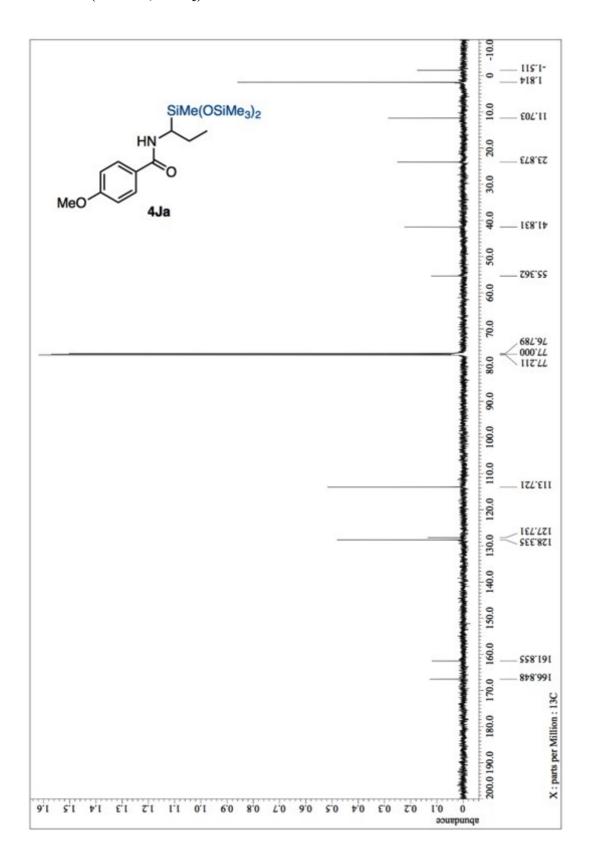
¹³C NMR (151MHz, CDCl₃) of 4Ia



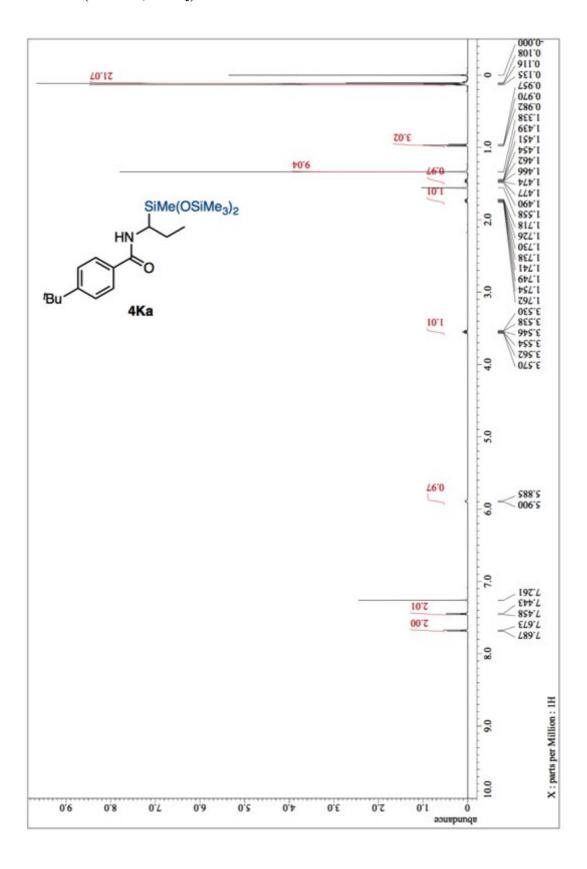
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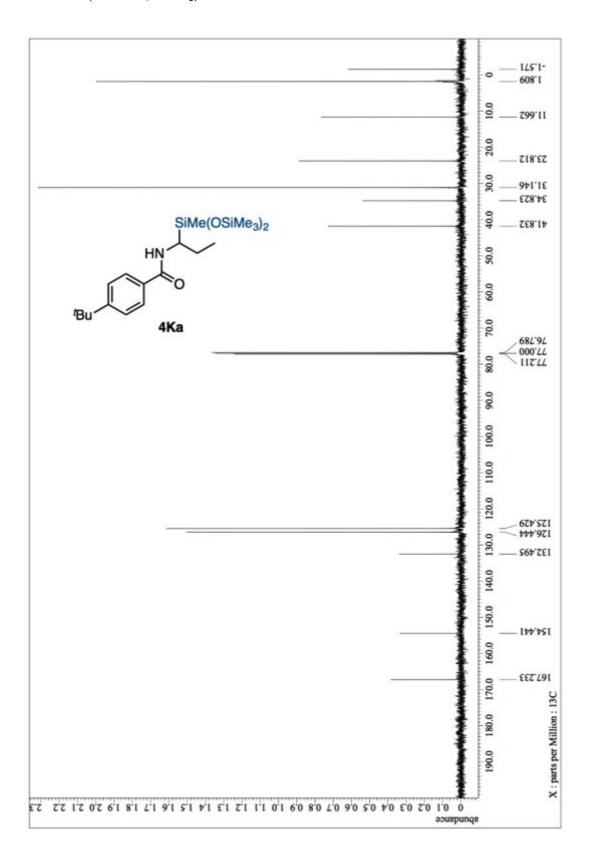
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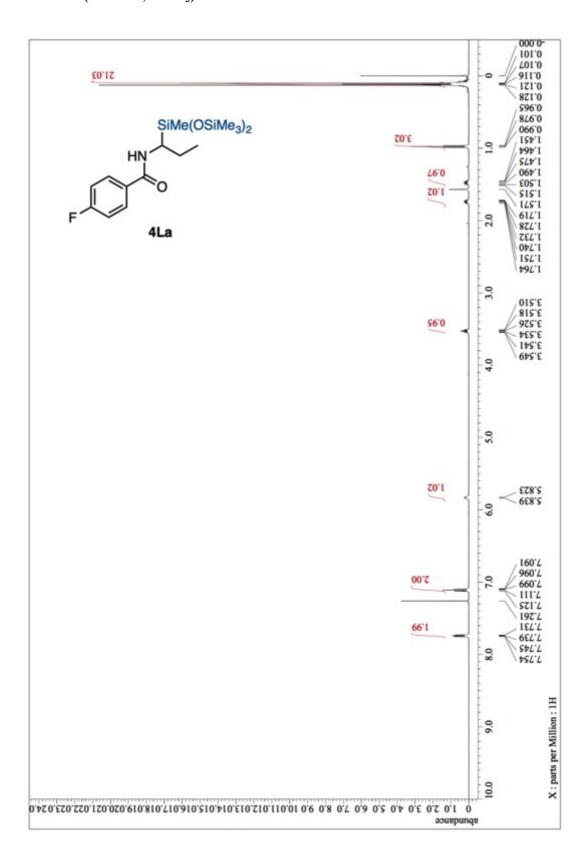
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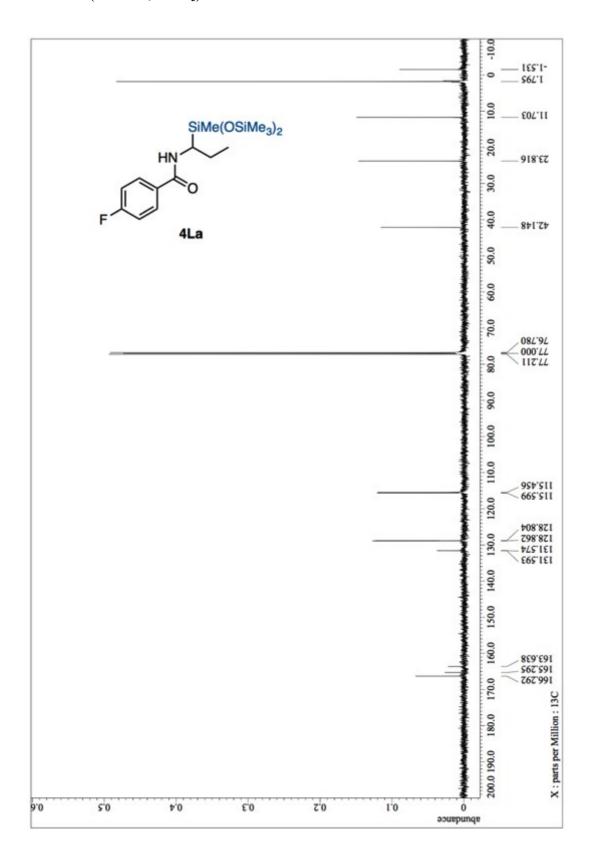
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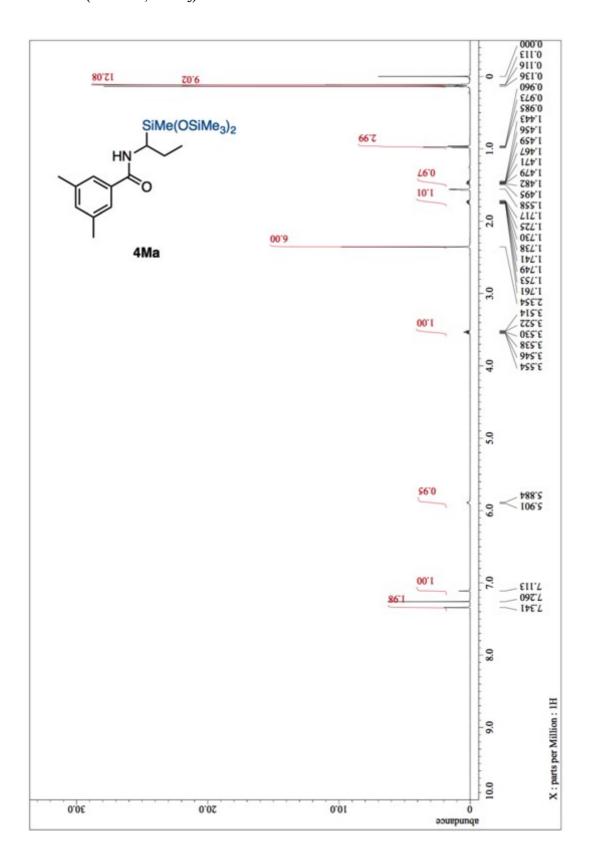
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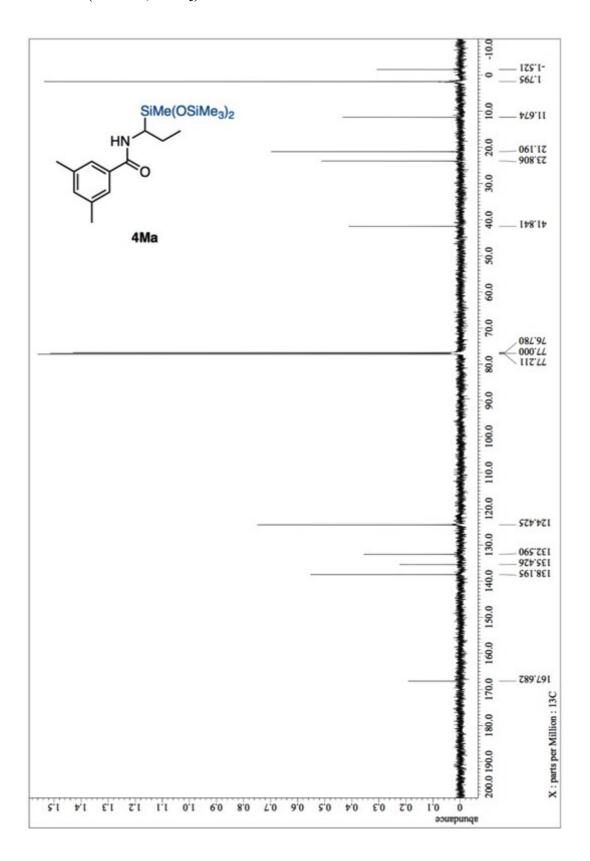
¹³C NMR (151MHz, CDCl₃) of 4La



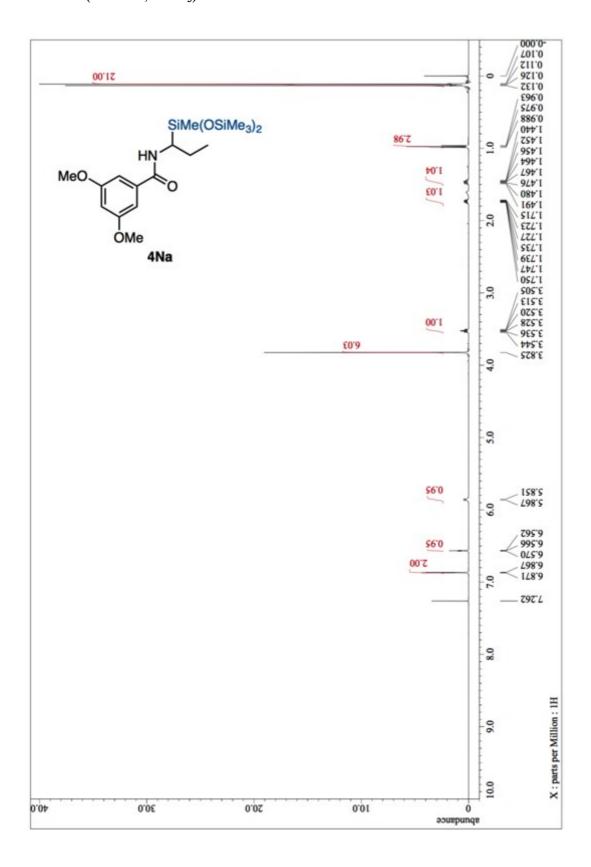
¹H NMR (600MHz, CDCl₃) of 4Ma



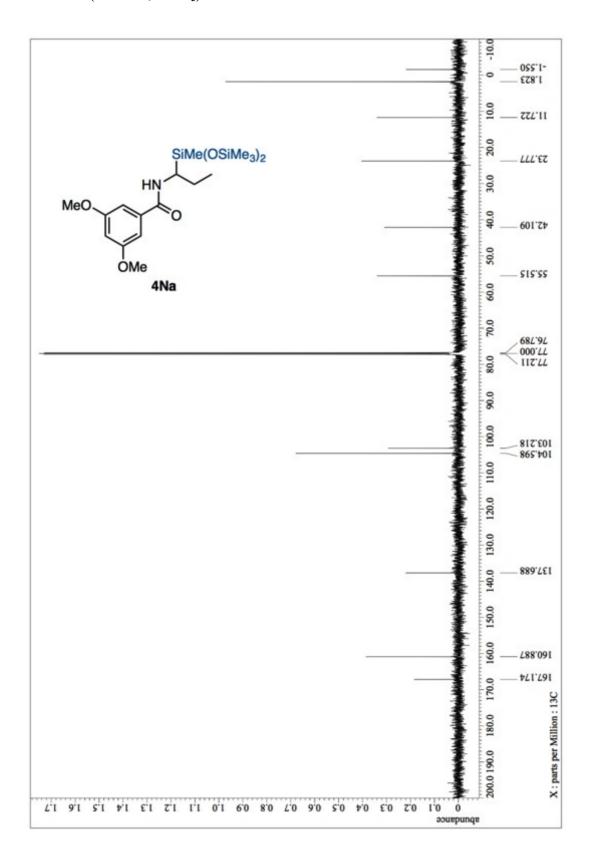
¹³C NMR (151MHz, CDCl₃) of 4Ma



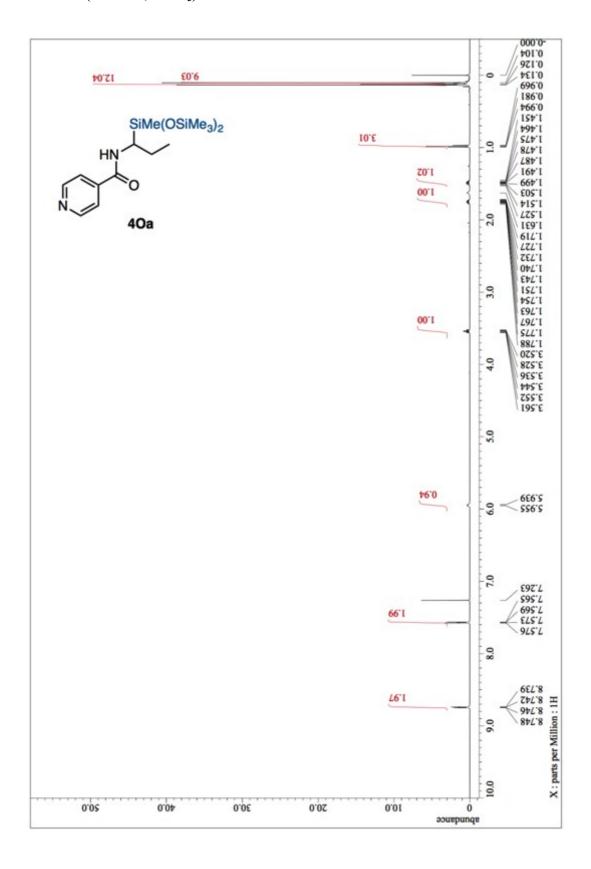
¹H NMR (600MHz, CDCl₃) of 4Na



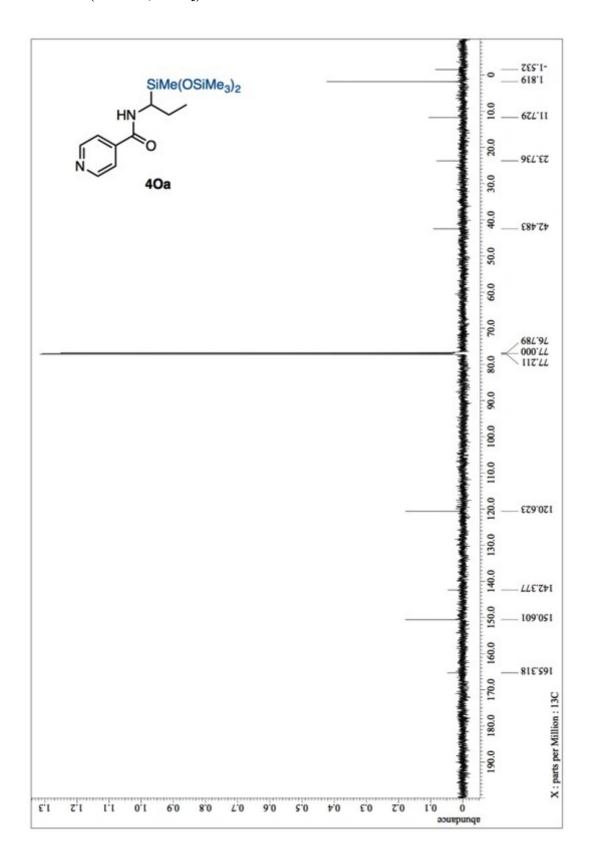
¹³C NMR (151MHz, CDCl₃) of 4Na



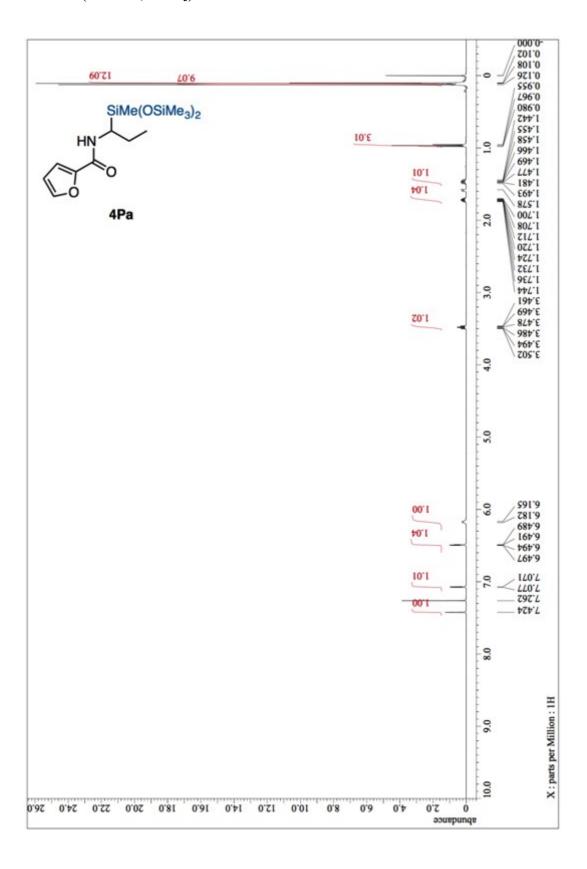
¹H NMR (600MHz, CDCl₃) of 4Oa



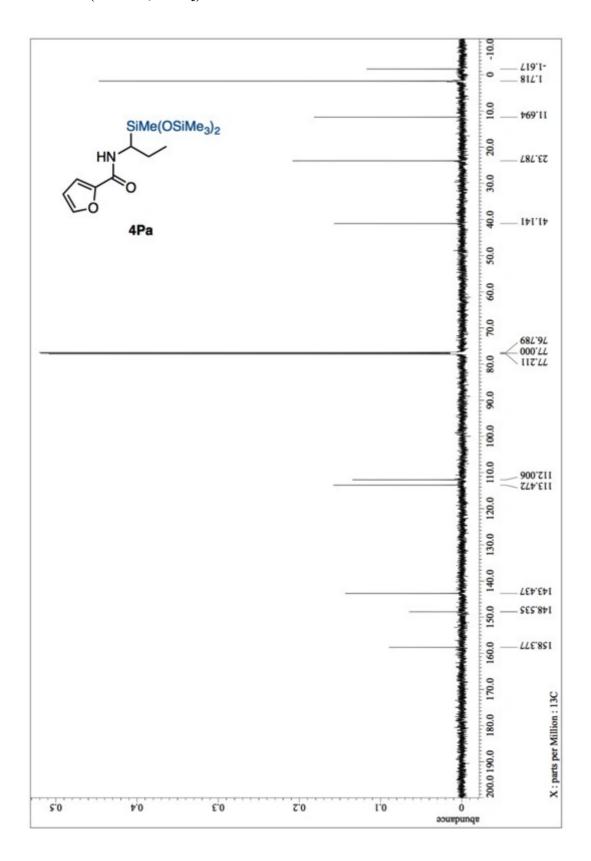
13 C NMR (151MHz, CDCl₃) of 4Oa



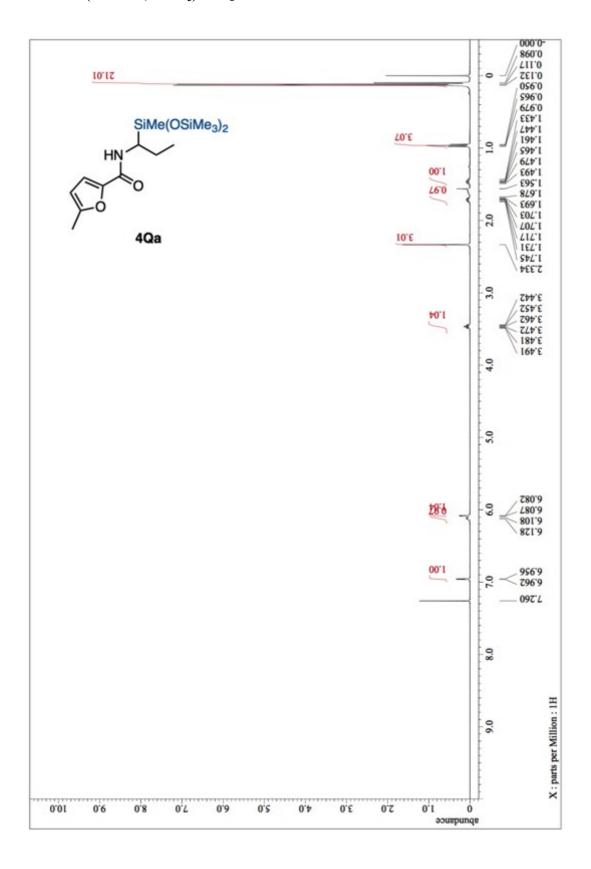
¹H NMR (600MHz, CDCl₃) of 4Pa



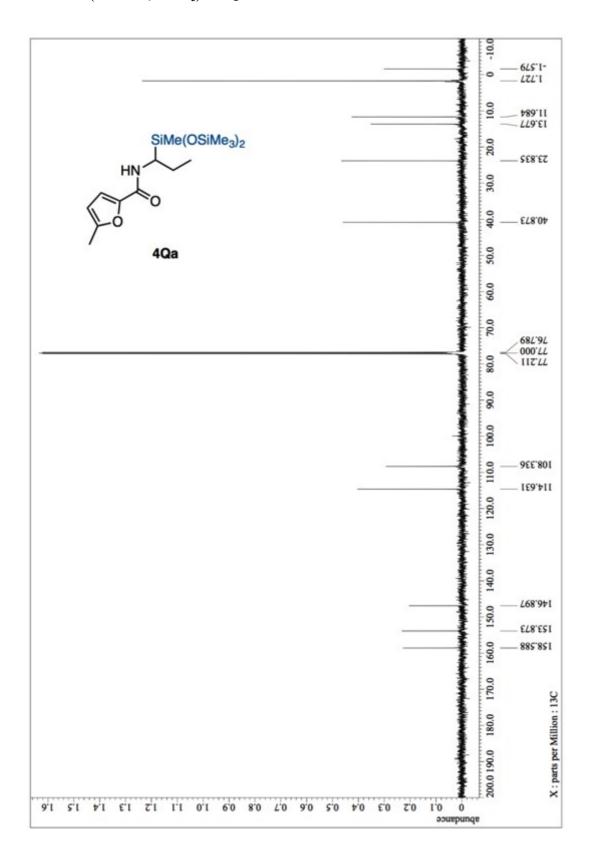
$^{13}\mathrm{C}$ NMR (151MHz, CDCl₃) of 4Pa



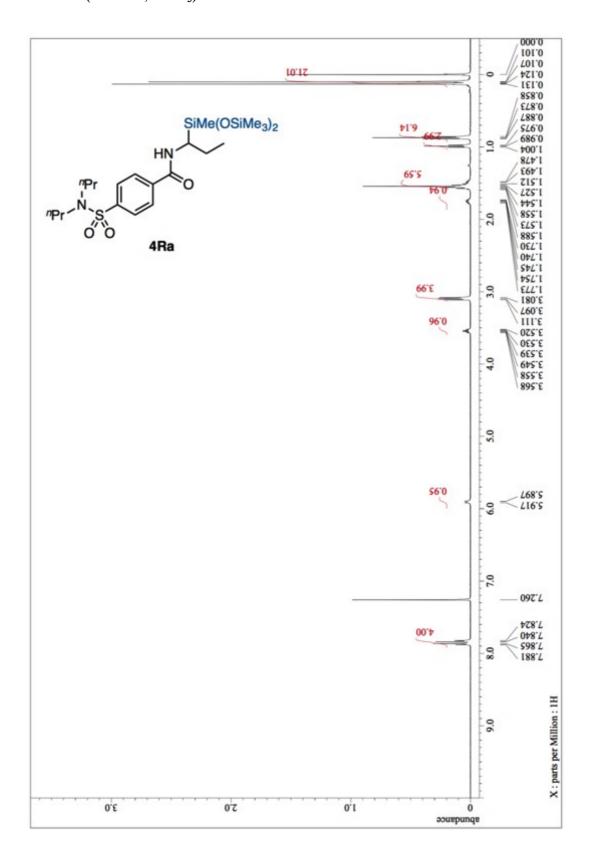
¹H NMR (500MHz, CDCl₃) of 4Qa



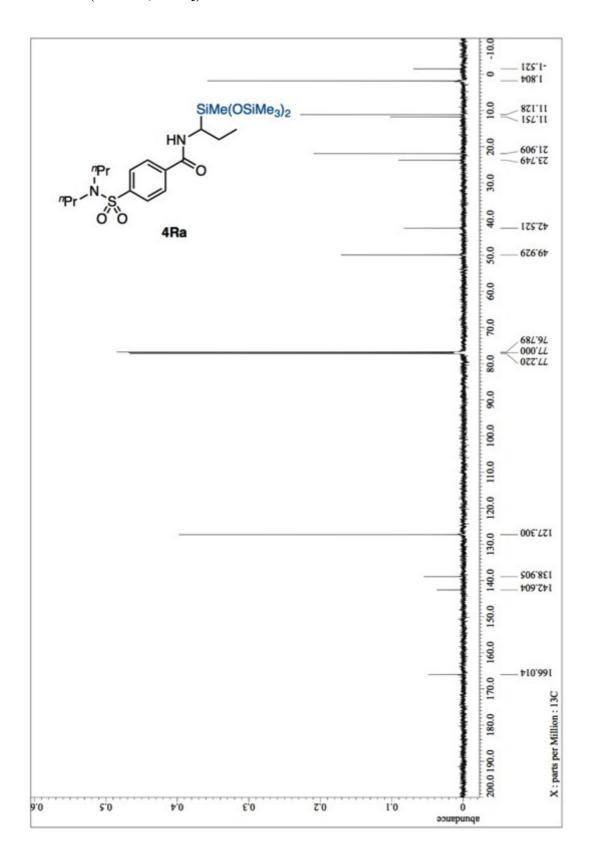
¹³C NMR (151MHz, CDCl₃) of 4Qa



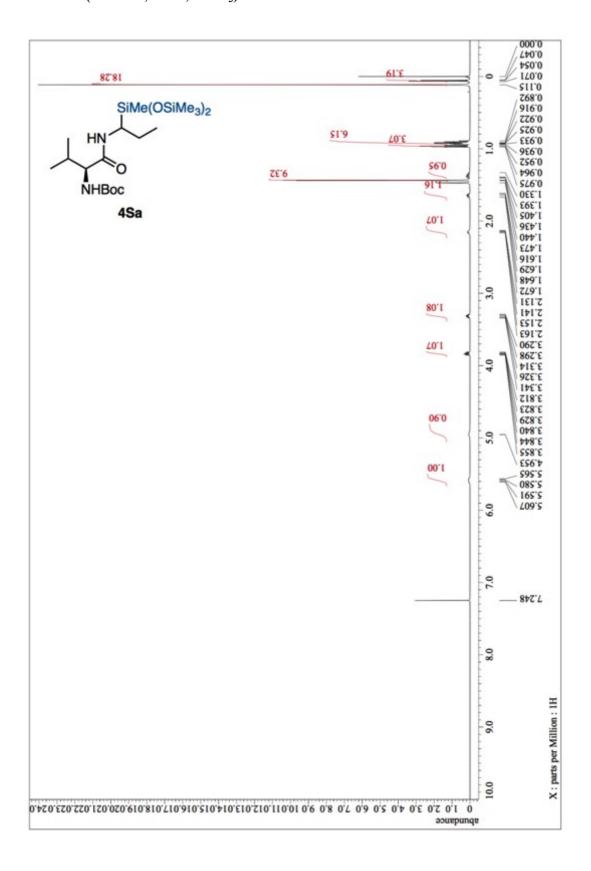
¹H NMR (500MHz, CDCl₃) of 4Ra



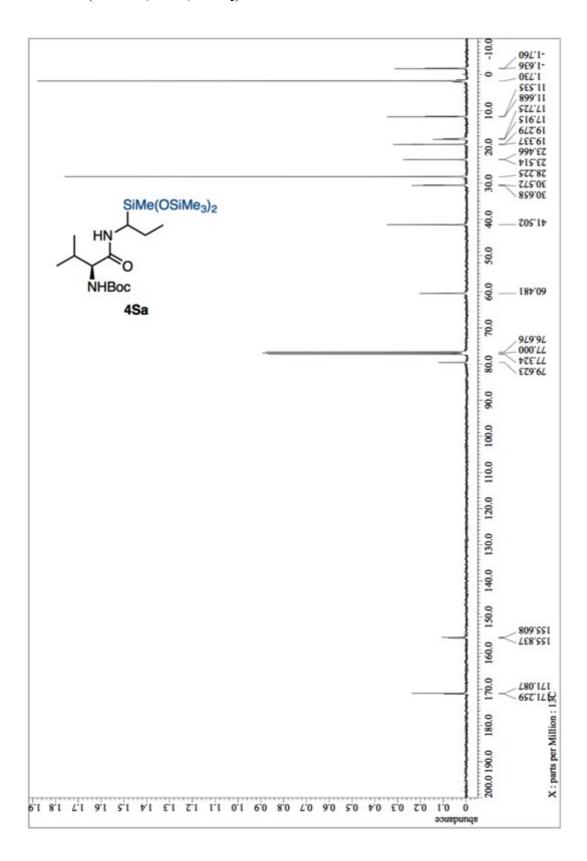
$^{13}\mathrm{C}$ NMR (151MHz, CDCl₃) of 4Ra



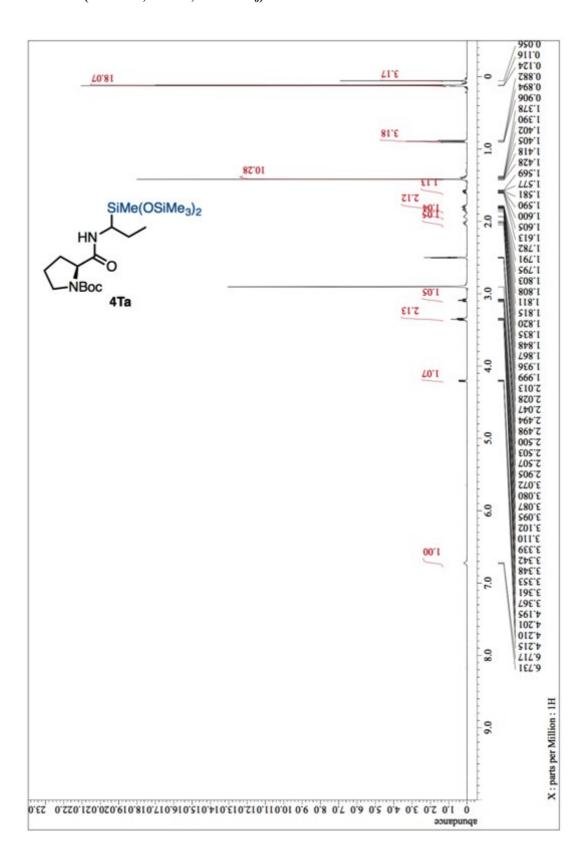
¹H NMR (600MHz, 60 °C, CDCl₃) of 4Sa



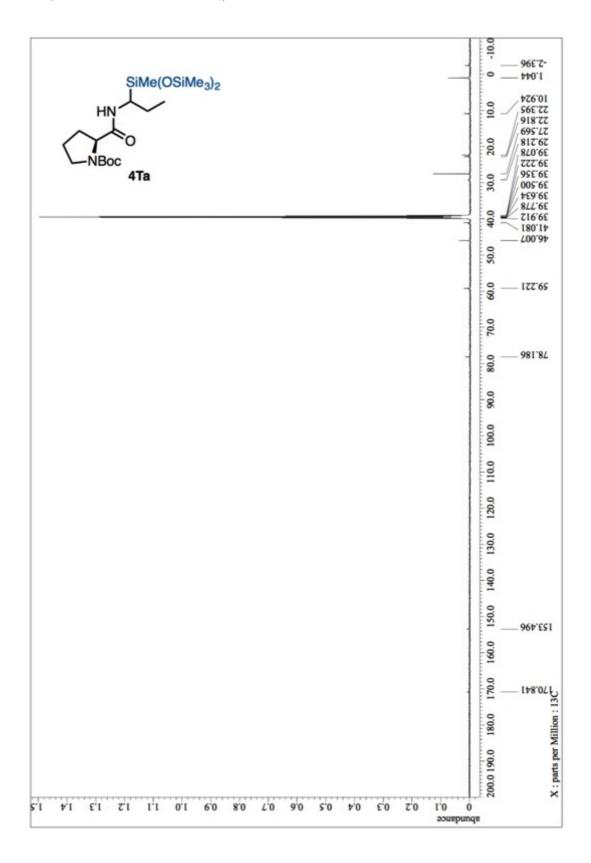
¹³C NMR (101MHz, 60 °C, CDCl₃) of 4Sa



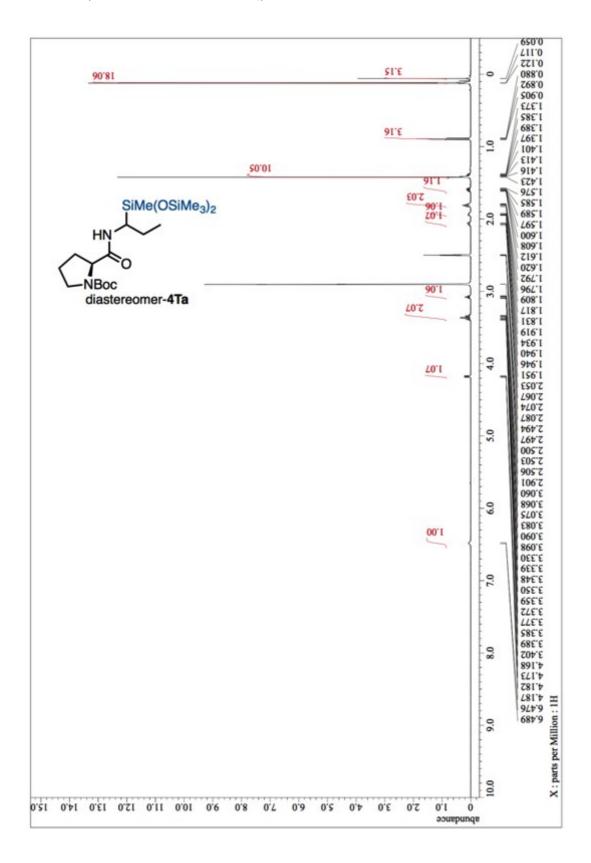
¹H NMR (600MHz, 120 °C, DMSO-d₆) of 4Ta



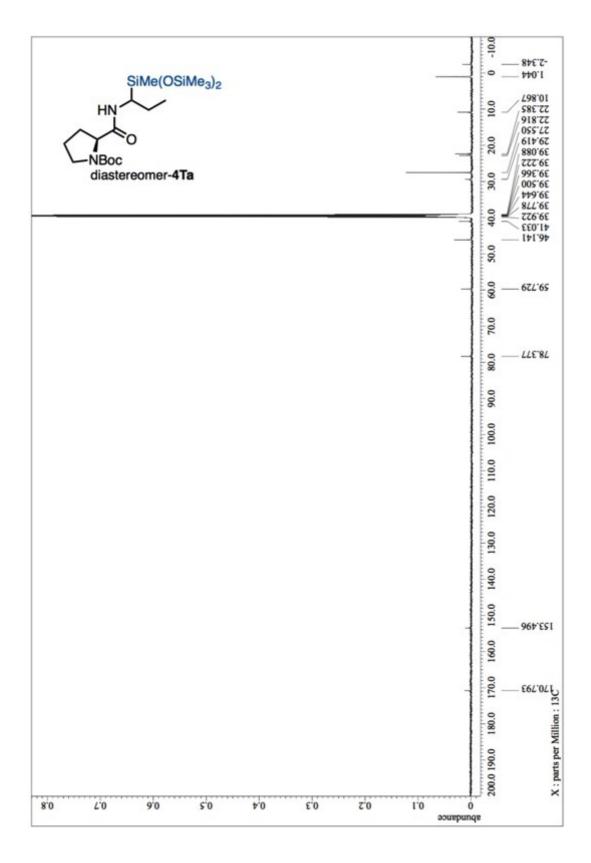
13 C NMR (151MHz, 120 °C, DMSO- d_6) of 4Ta



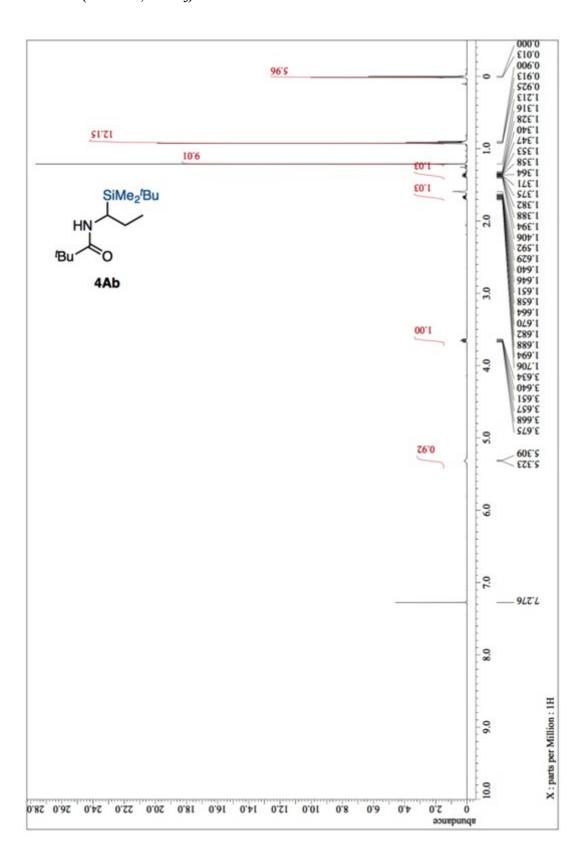
¹H NMR (600MHz, 120 °C, DMSO-d₆) of diastereomer-4Ta



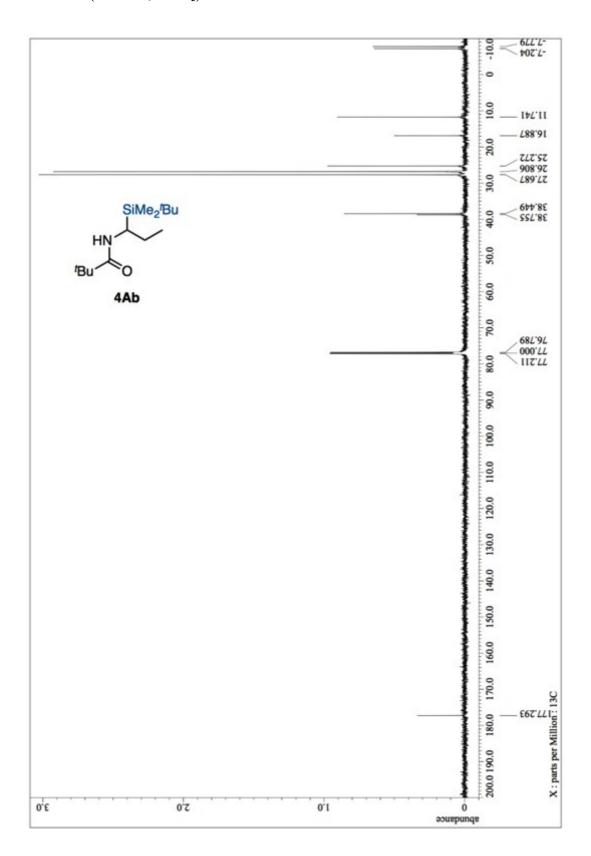
13 C NMR (151MHz, 120 °C, DMSO- d_6) of diastereomer-4Ta



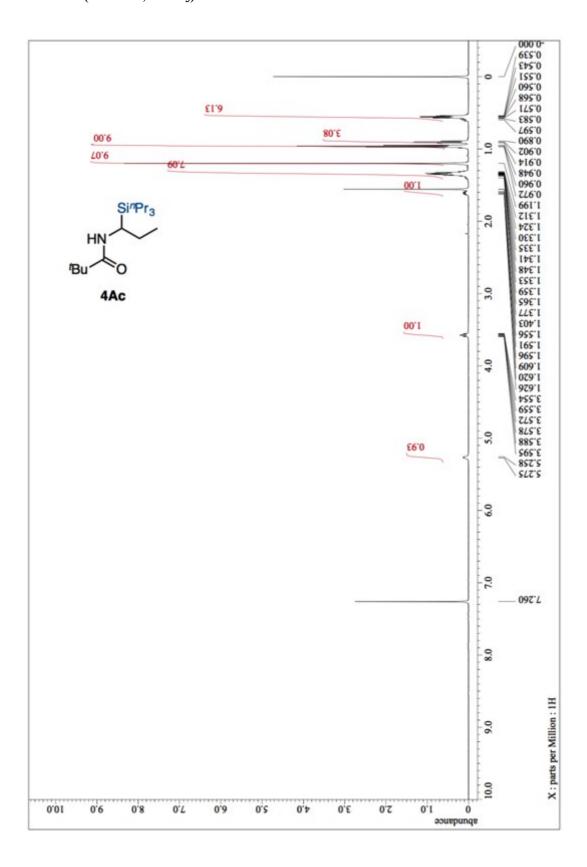
¹H NMR (600MHz, CDCl₃) of 4Ab



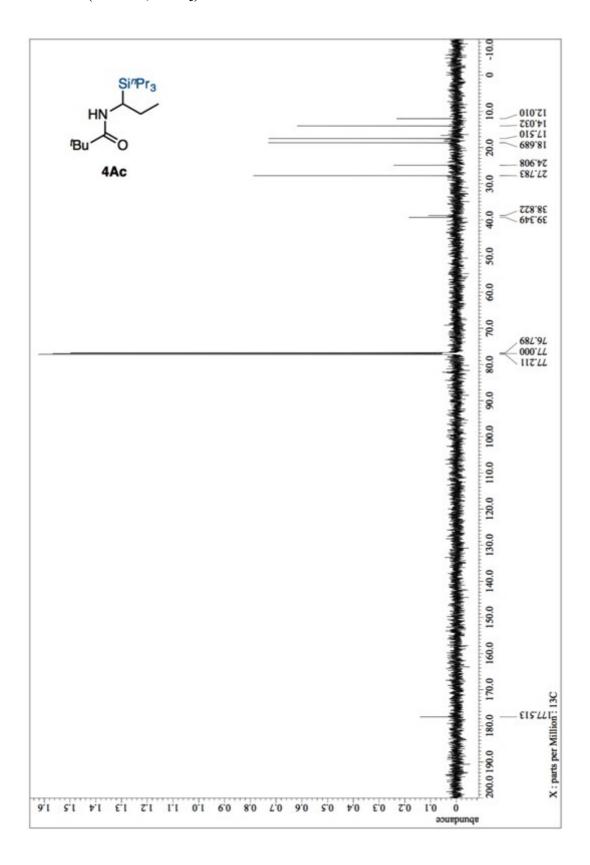
¹³C NMR (151MHz, CDCl₃) of 4Ab



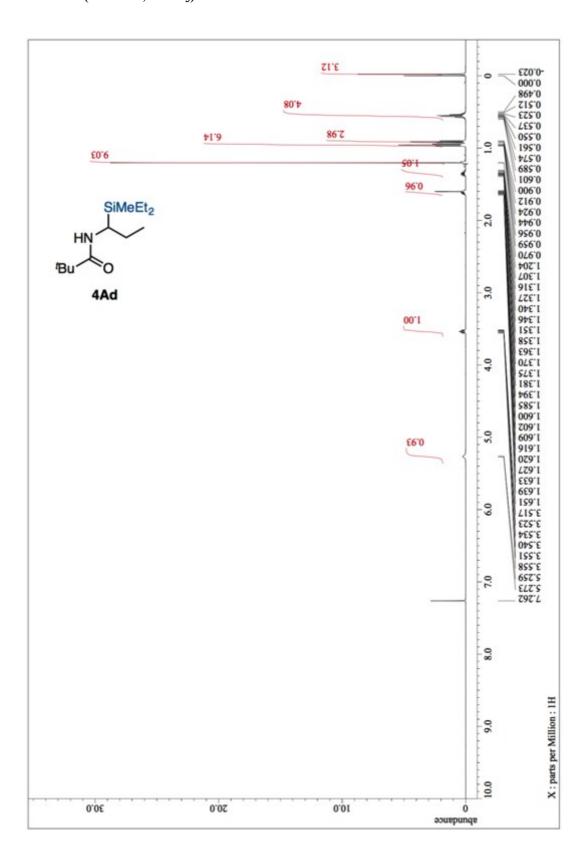
¹H NMR (600MHz, CDCl₃) of 4Ac



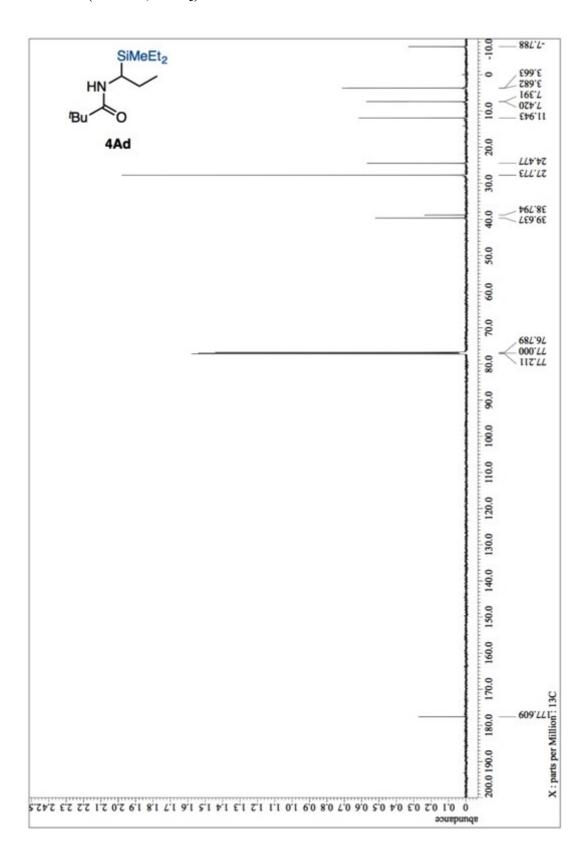
¹³C NMR (151MHz, CDCl₃) of 4Ac



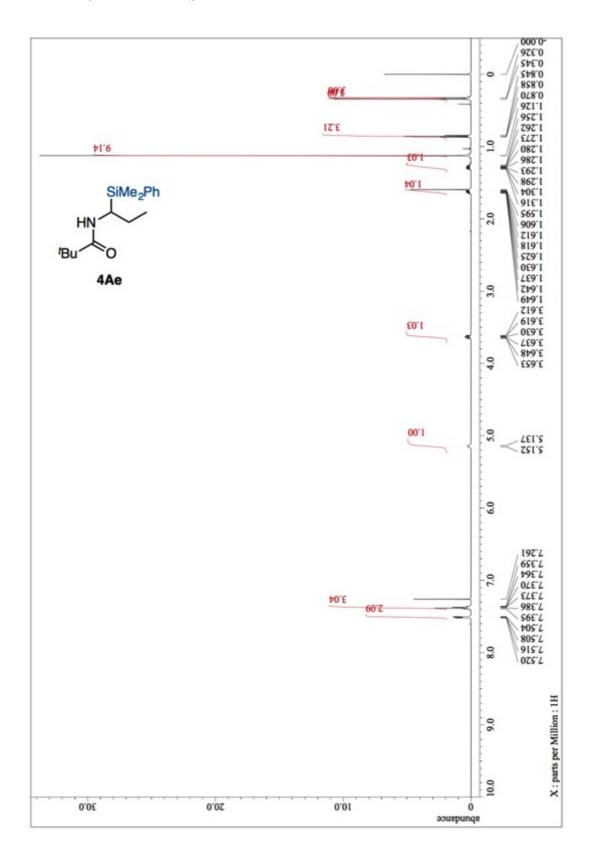
¹H NMR (600MHz, CDCl₃) of 4Ad



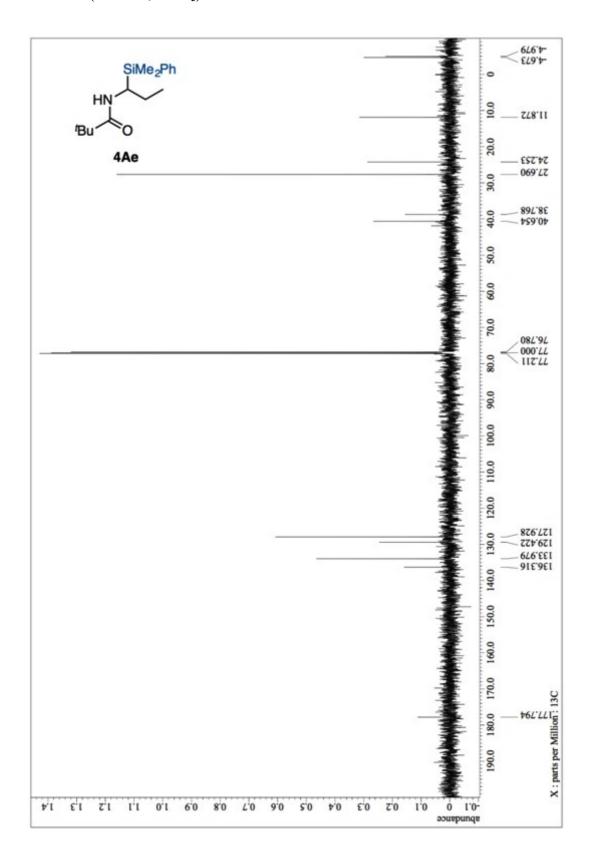
¹³C NMR (151MHz, CDCl₃) of 4Ad



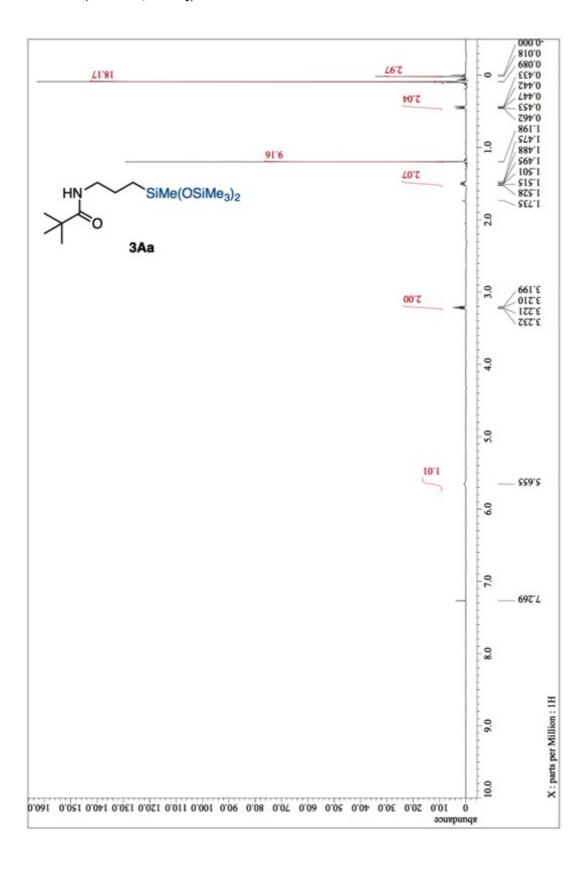
¹H NMR (600MHz, CDCl₃) of 4Ae



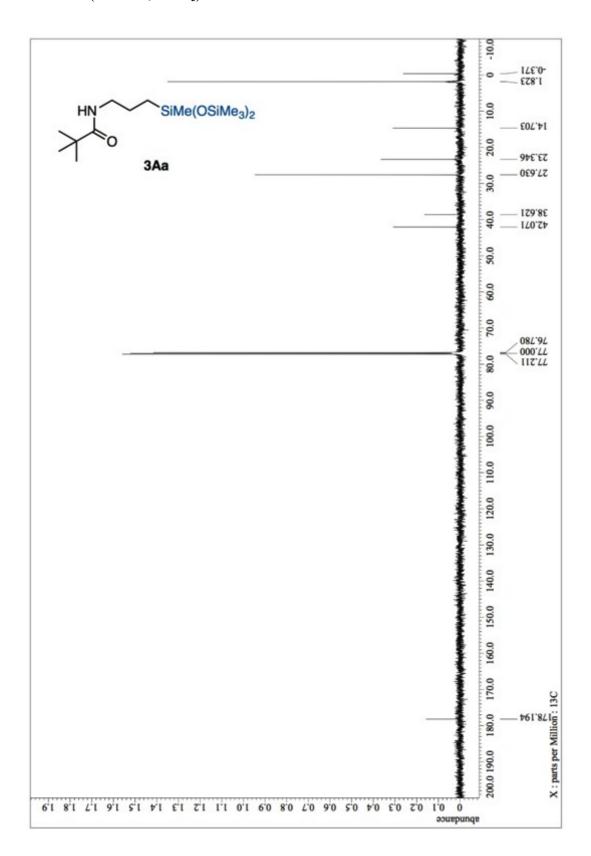
¹³C NMR (151MHz, CDCl₃) of 4Ae



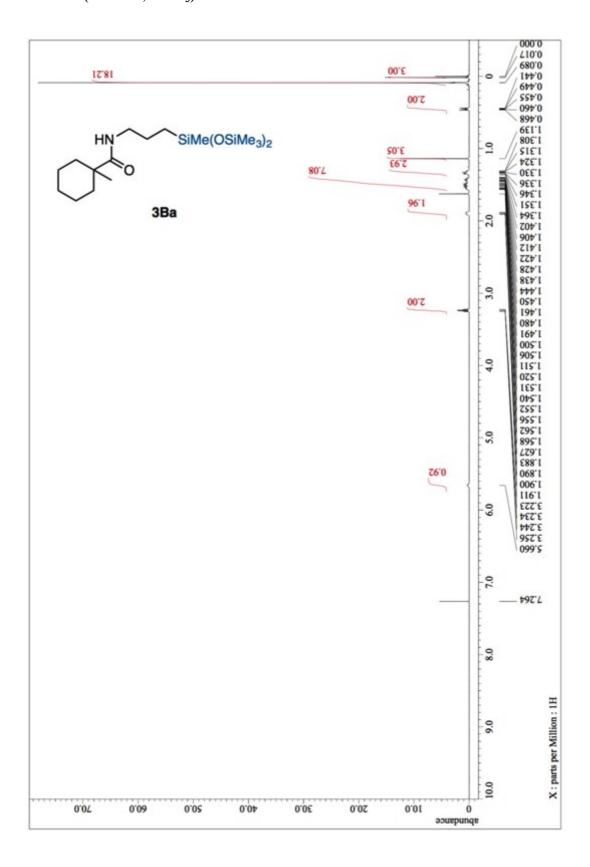
¹H NMR (600MHz, CDCl₃) of 3Aa



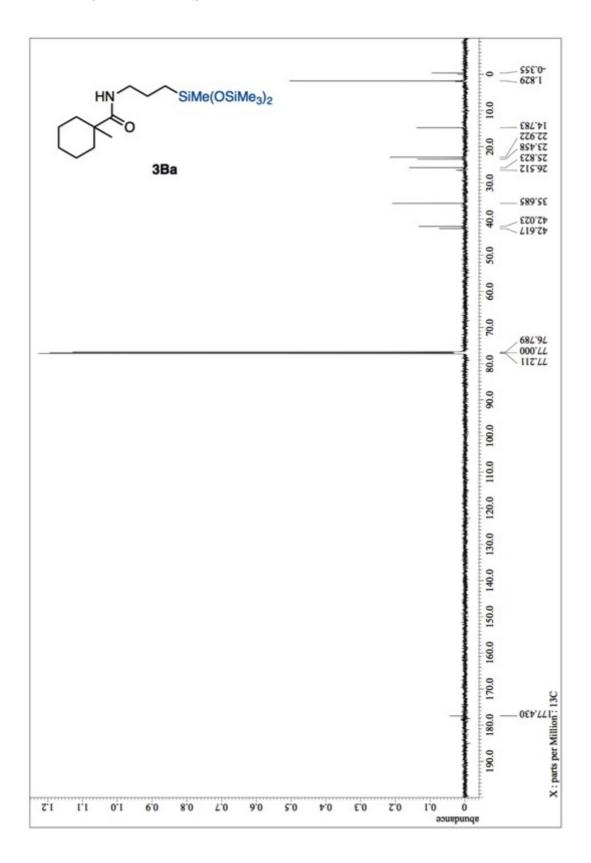
¹³C NMR (151MHz, CDCl₃) of 3Aa



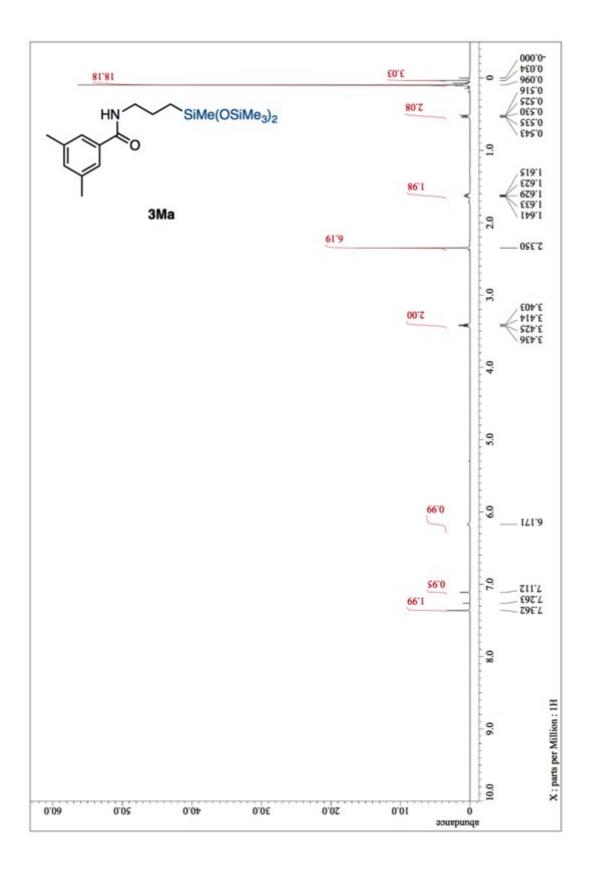
¹H NMR (600MHz, CDCl₃) of 3Ba



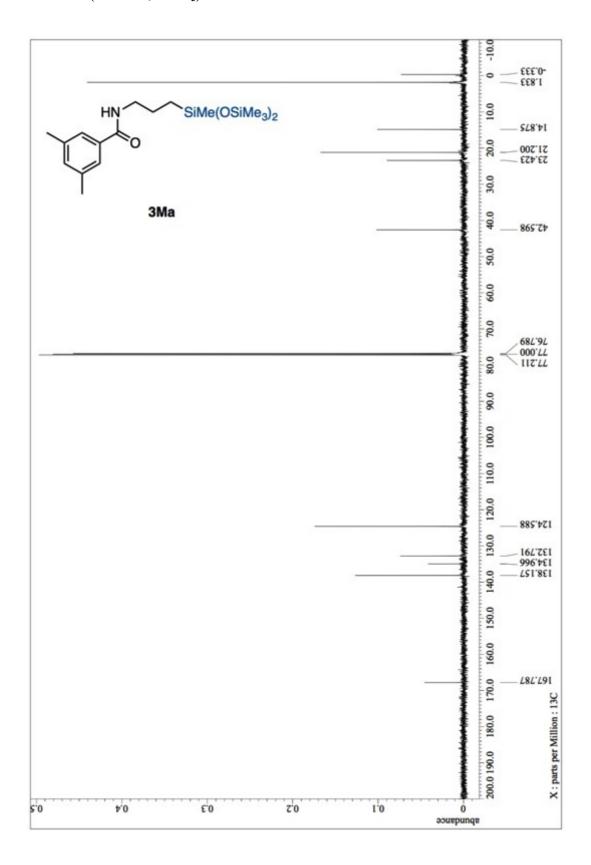
¹³C NMR (151MHz, CDCl₃) of 3Ba



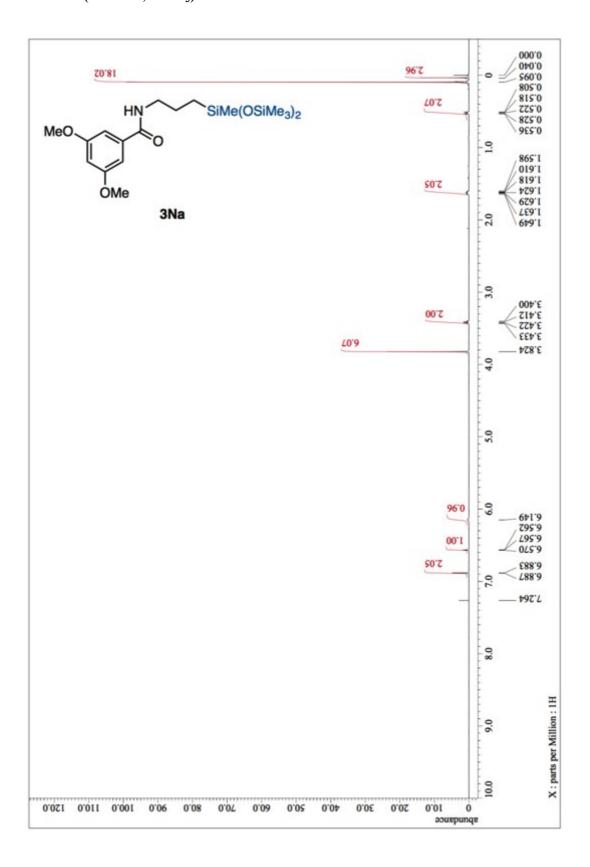
¹H NMR (600MHz, CDCl₃) of 3Ma



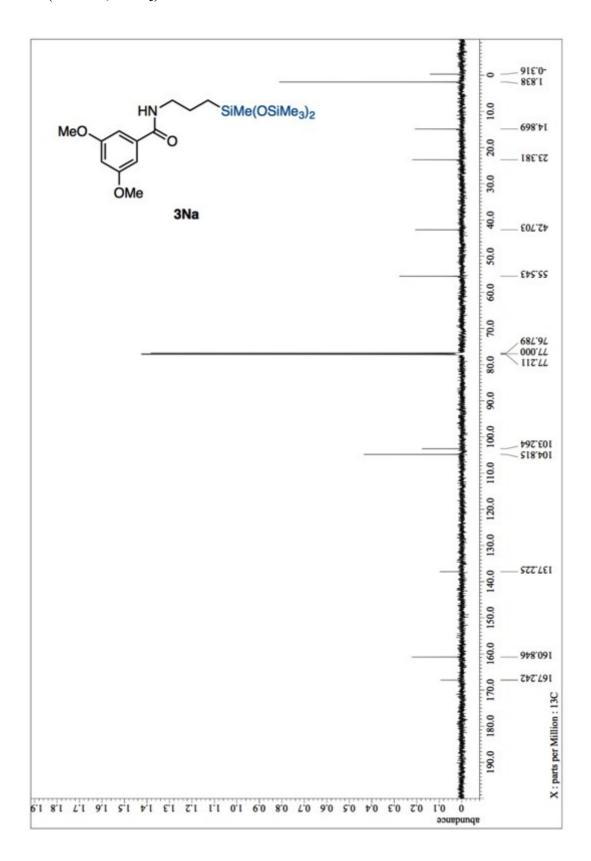
¹³C NMR (151MHz, CDCl₃) of 3Ma



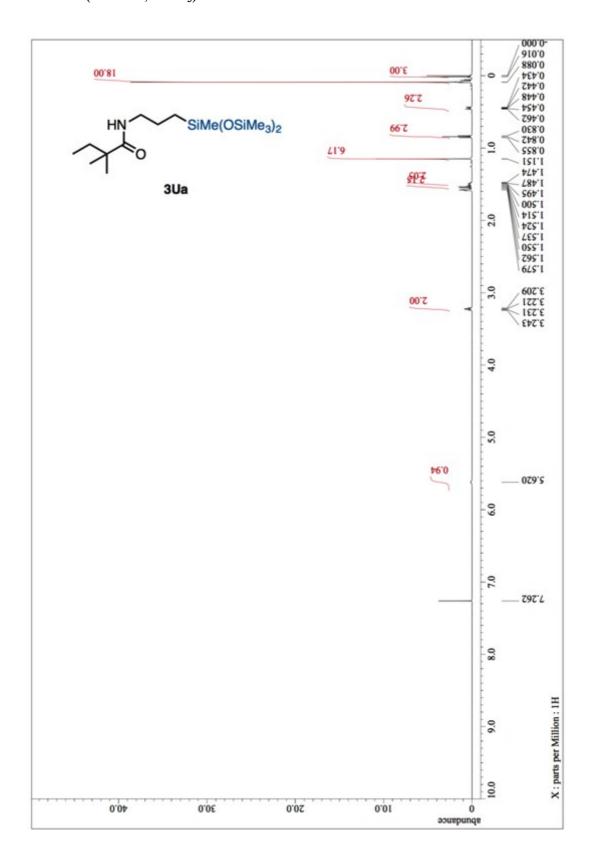
¹H NMR (600MHz, CDCl₃) of 3Na



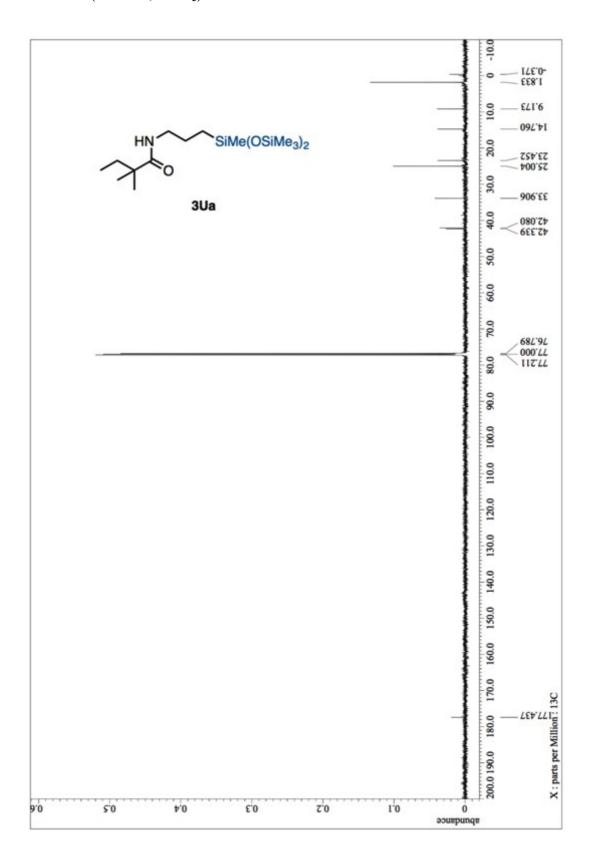
¹³C NMR (151MHz, CDCl₃) of 3Na



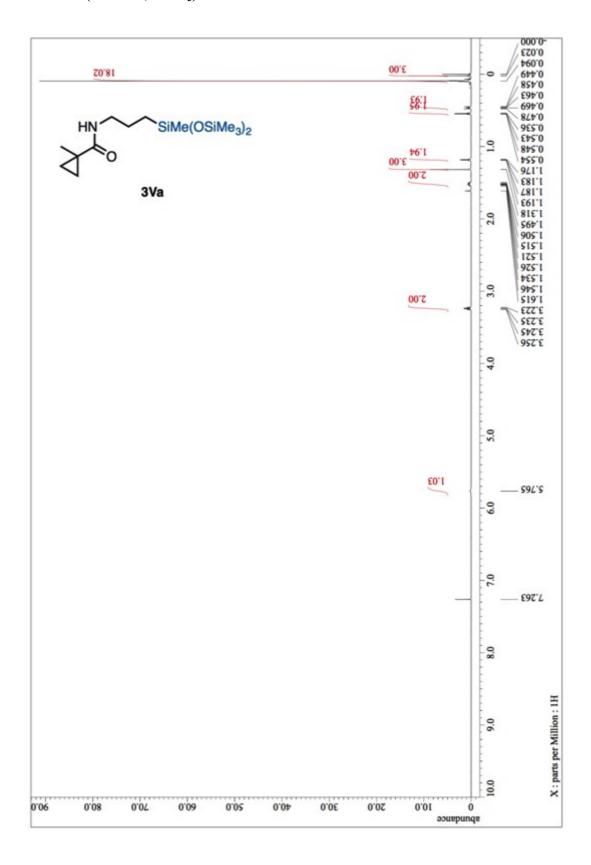
¹H NMR (600MHz, CDCl₃) of 3Ua



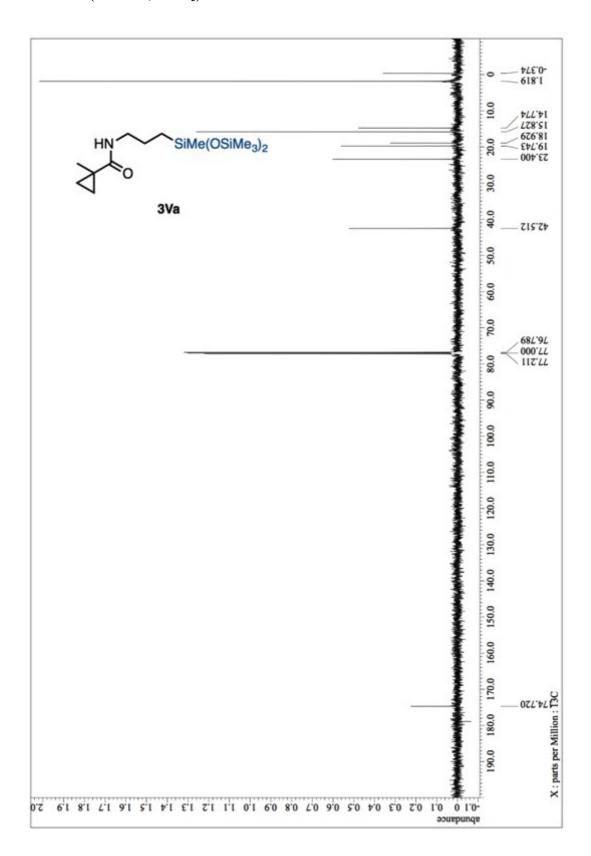
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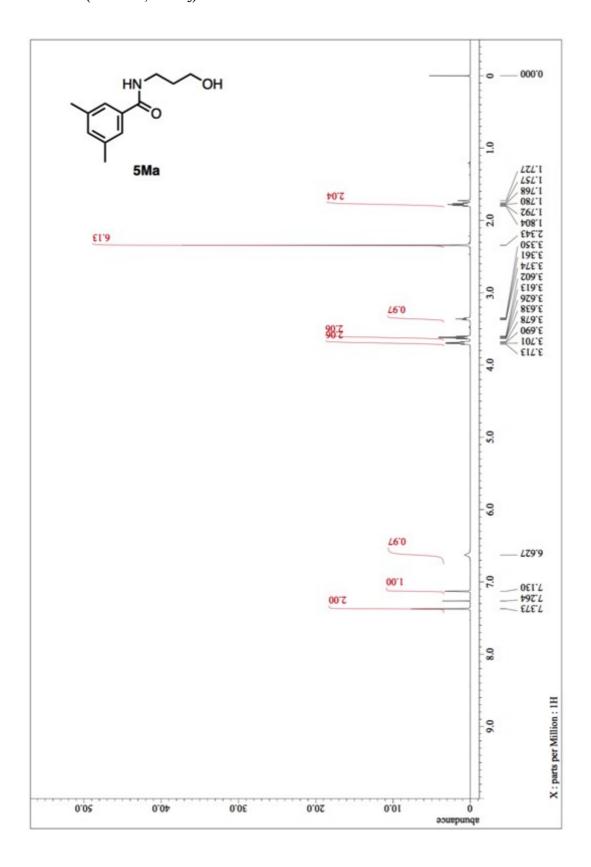
¹H NMR (600MHz, CDCl₃) of 3Va



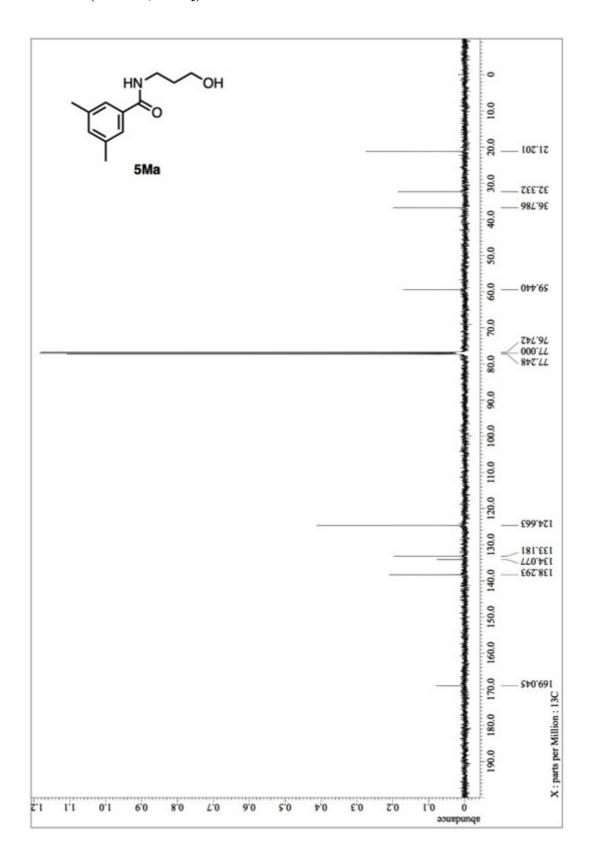
¹³C NMR (151MHz, CDCl₃) of 3Va



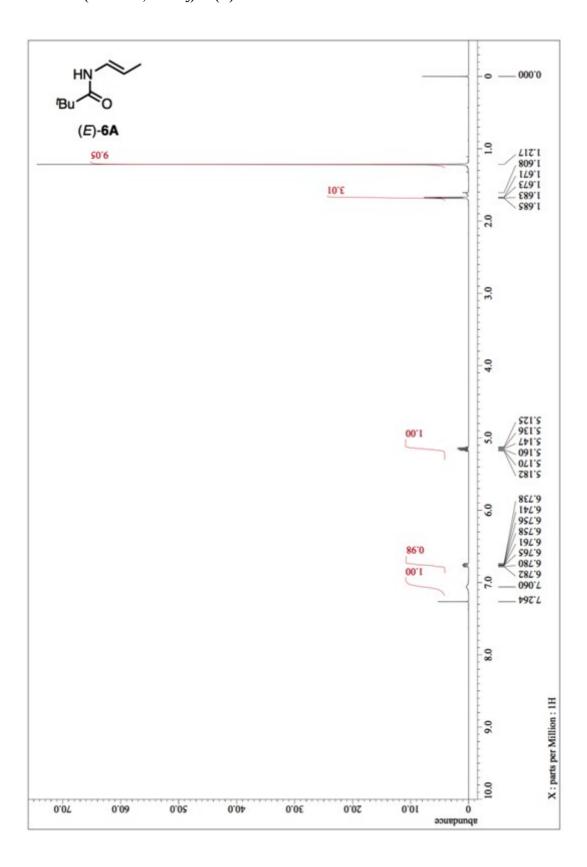
¹H NMR (500MHz, CDCl₃) of 5Ma



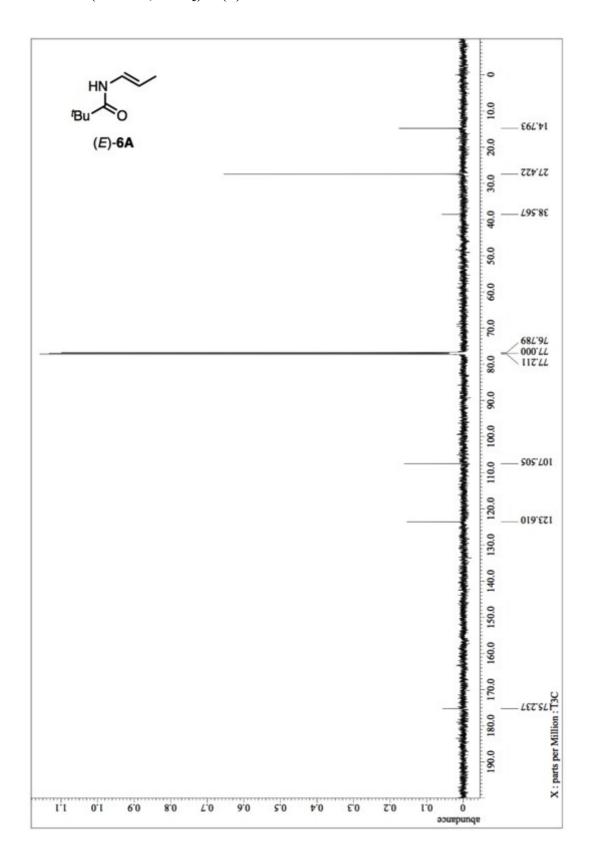
$^{13}\mathrm{C}$ NMR (126MHz, CDCl₃) of 5Ma



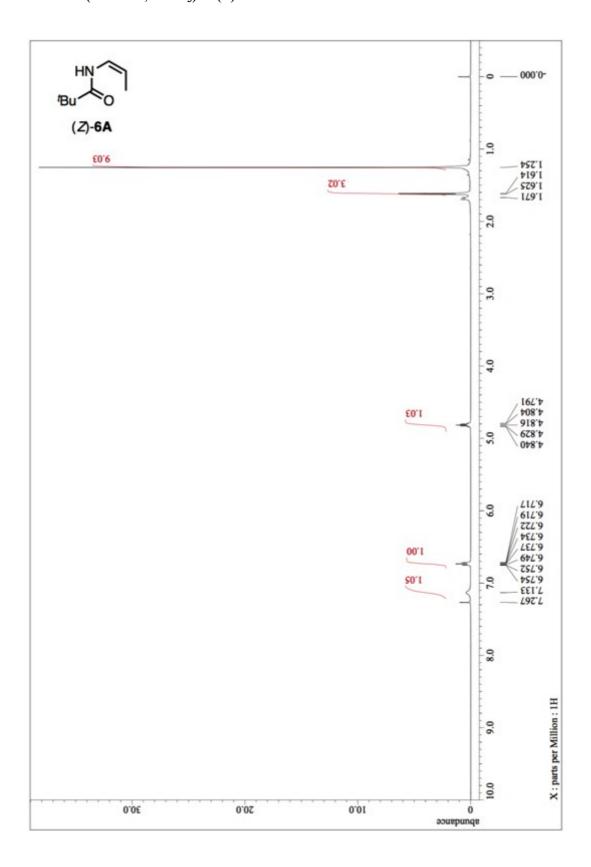
¹H NMR (600MHz, CDCl₃) of (*E*)-6A



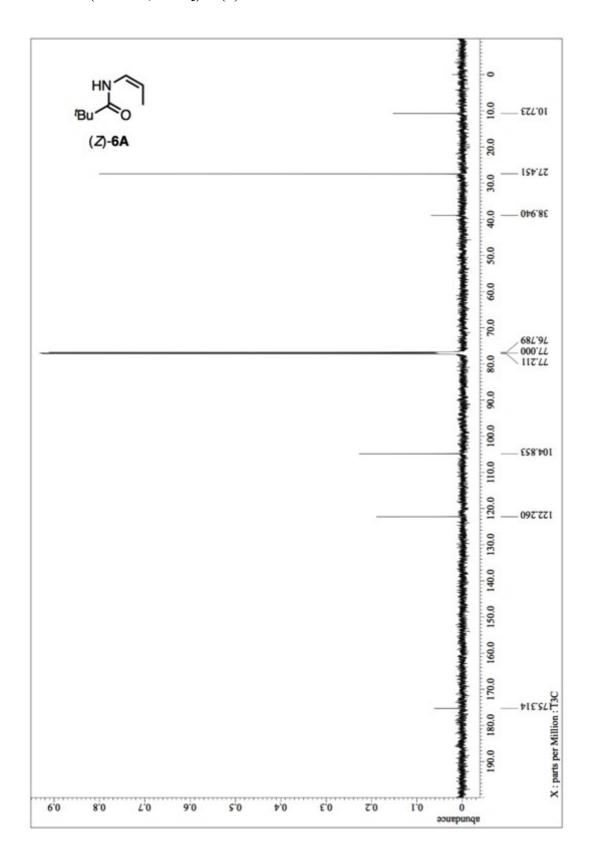
13 C NMR (151MHz, CDCl₃) of (*E*)-6A



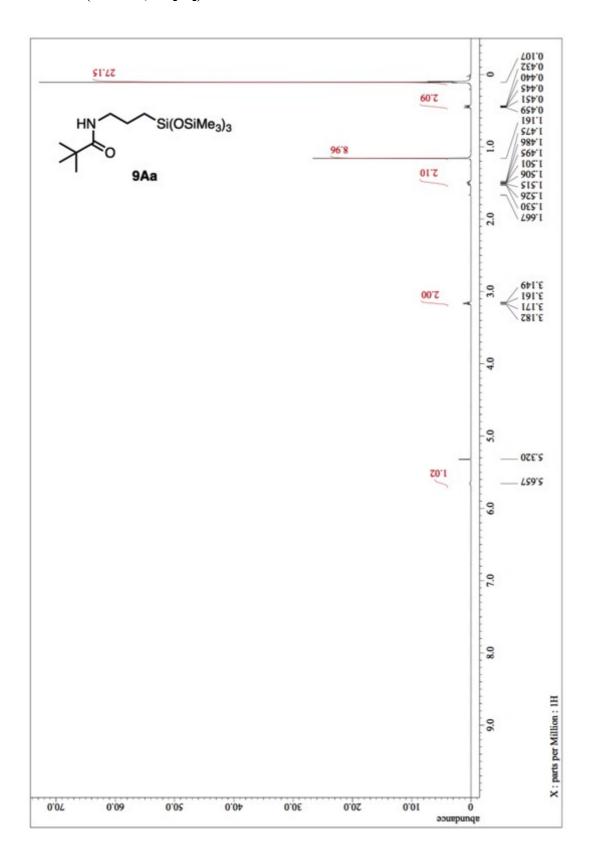
¹H NMR (600MHz, CDCl₃) of (*Z*)-6A



¹³C NMR (151MHz, CDCl₃) of (*Z*)-6A



¹H NMR (600MHz, CD₂Cl₂) of 9Aa



¹³C NMR (151MHz, CD₂Cl₂) of 9Aa

