

## Supporting Information

### **Functionalized AIE nanoparticles with efficient deep-red emission, mitochondria specificity, cancer cell selectivity and multiphoton susceptibility**

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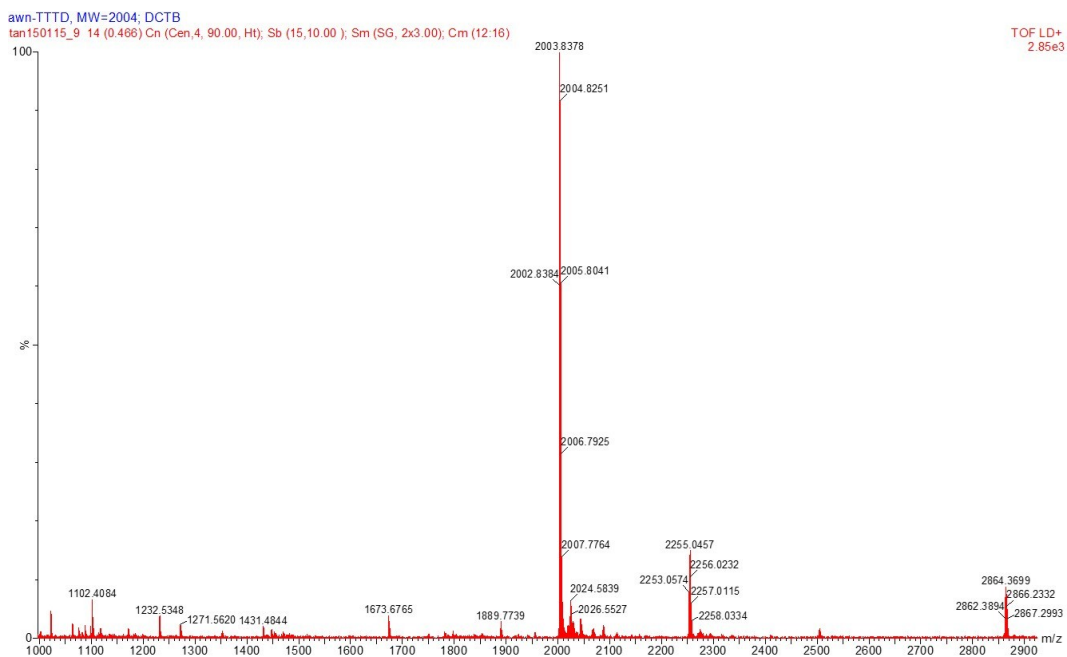
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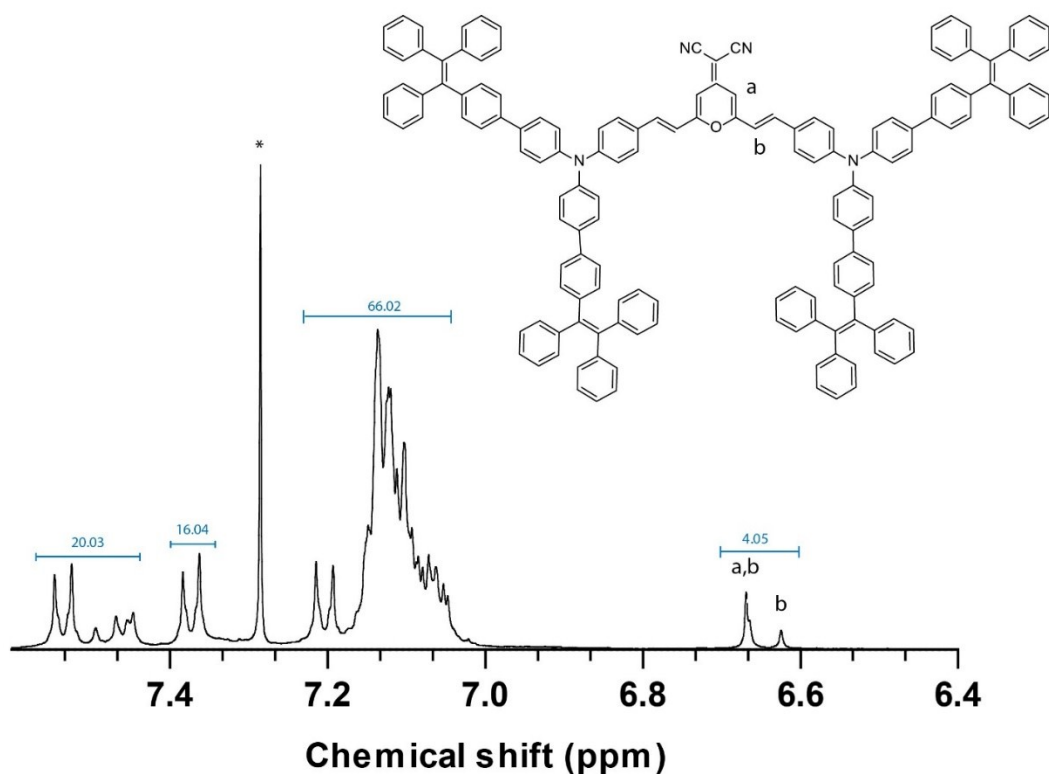
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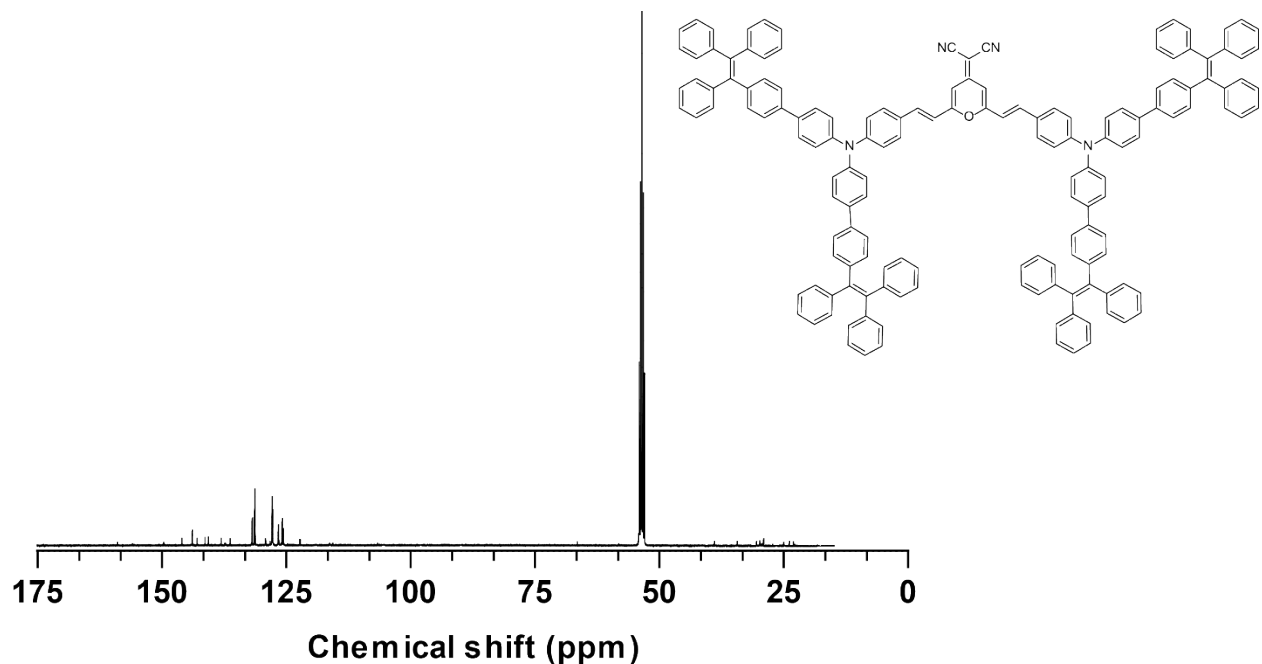
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**Figure S1.** HRMS spectrum of TPE-TETRAD.



**Figure S2.**  $^1\text{H}$  NMR spectrum of TPE-TETRAD.  $^1\text{H}$  NMR (400 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 7.56–7.43 (m, 20H), 7.39–7.34 (m, 16H), 7.33–7.0 (m, 66H), 6.67 (d,  $J = 16$  Hz, 2H; pyran  $-\text{CH}=\text{}$ ), 6.63 (s, 2H; pyran H).

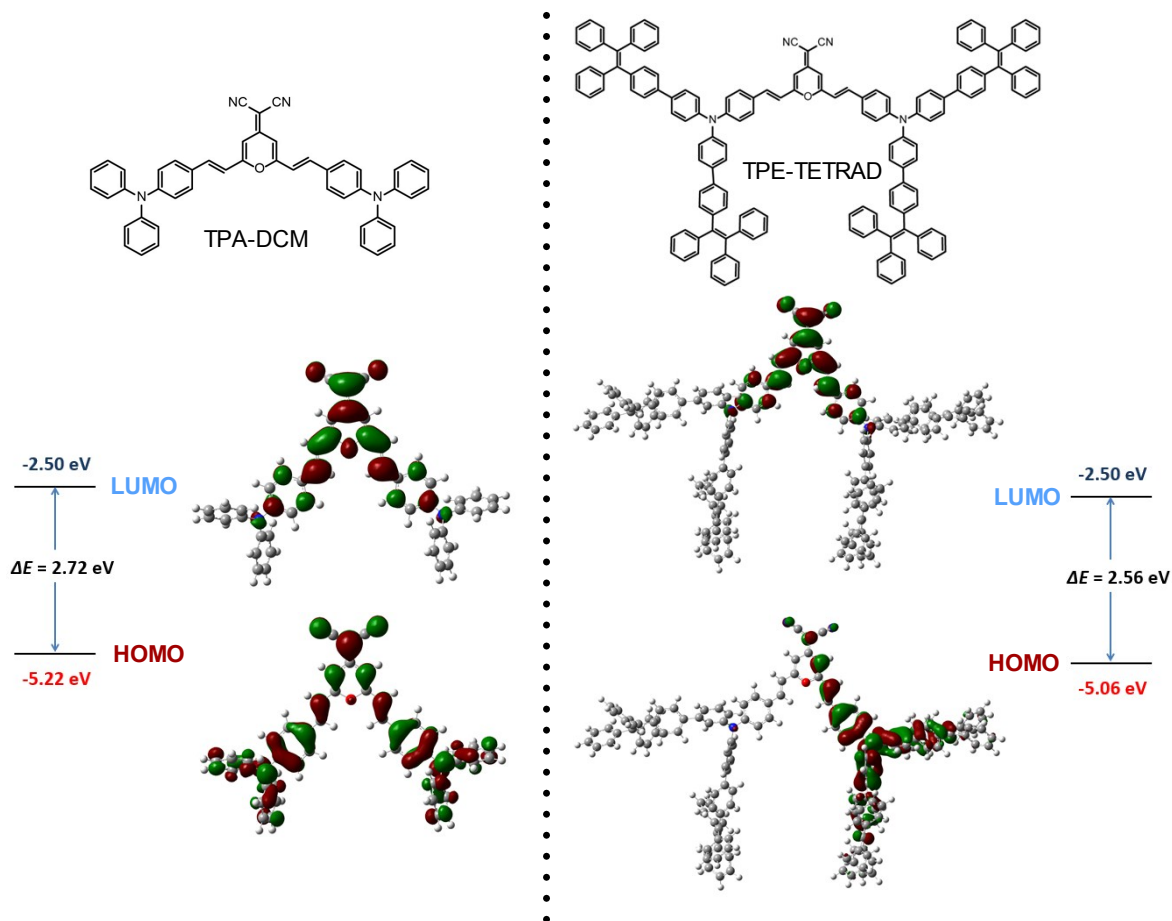


**Figure S3.**  $^{13}\text{C}$  NMR spectrum of TPE-TETRAD.  $^{13}\text{C}$  NMR (75 MHz,  $\text{CDCl}_3$ ),  $\delta$  (ppm): 159.5, 150.0, 145.9, 143.8, 142.7, 141.2, 140.5, 139.1, 136.2, 131.6, 131.2, 128.7, 127.7, 126.2, 125.7, 122.1, 116.5, 115.2, 106.4, 53.4.

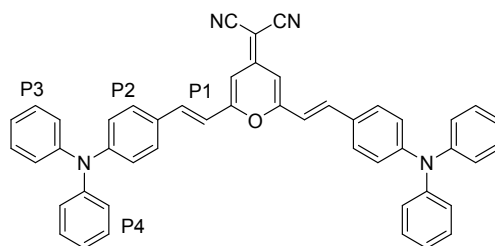
**Table S1.** Optical Properties of TPA-DCM and TPE-TETRAD

	$\lambda_{\text{ab}}$ (nm) <sup>a</sup>	$E_{\text{g}}$ (eV) <sup>b</sup>	$\lambda_{\text{em}}$ (nm) <sup>c</sup>		Stokes shift <sup>d</sup>
			solution	aggregate	
TPA-DCM	465	2.72	620	670	155
TPE-TETRAD	500	2.56	668	675	168

<sup>a</sup>)Absorption maxima ( $\lambda_{\text{ab}}$ ) in THF(10  $\mu\text{M}$ ); <sup>b</sup>)HOMO-LUMO band gap ( $E_{\text{g}}$ ) derived from theoretical DFT calculations; <sup>c</sup>)emission maxima ( $\lambda_{\text{em}}$ ) for solution (in THF,10  $\mu\text{M}$ ) and aggregate (in THF/water 1:9 v/v, 10  $\mu\text{M}$ ); <sup>d</sup>)Stokes shift derived from subtracting  $\lambda_{\text{em}}$  from  $\lambda_{\text{ab}}$ .

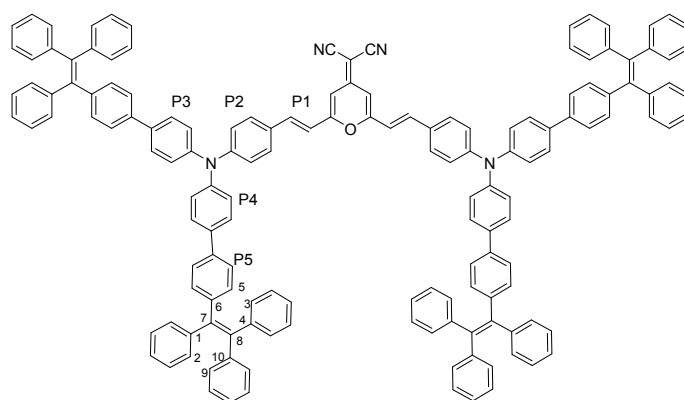


**Figure S4.** HOMO and LUMO energy levels of TPA-DCM and TPE-TETRAD, respectively. Molecular orbital amplitude plots of HOMO and LUMO energy levels of TPA-DCM and TPE-TETRAD were calculated using density functional theory (DFT) in Gaussian 09 program using B3LYP/6-31G(d) basis set.



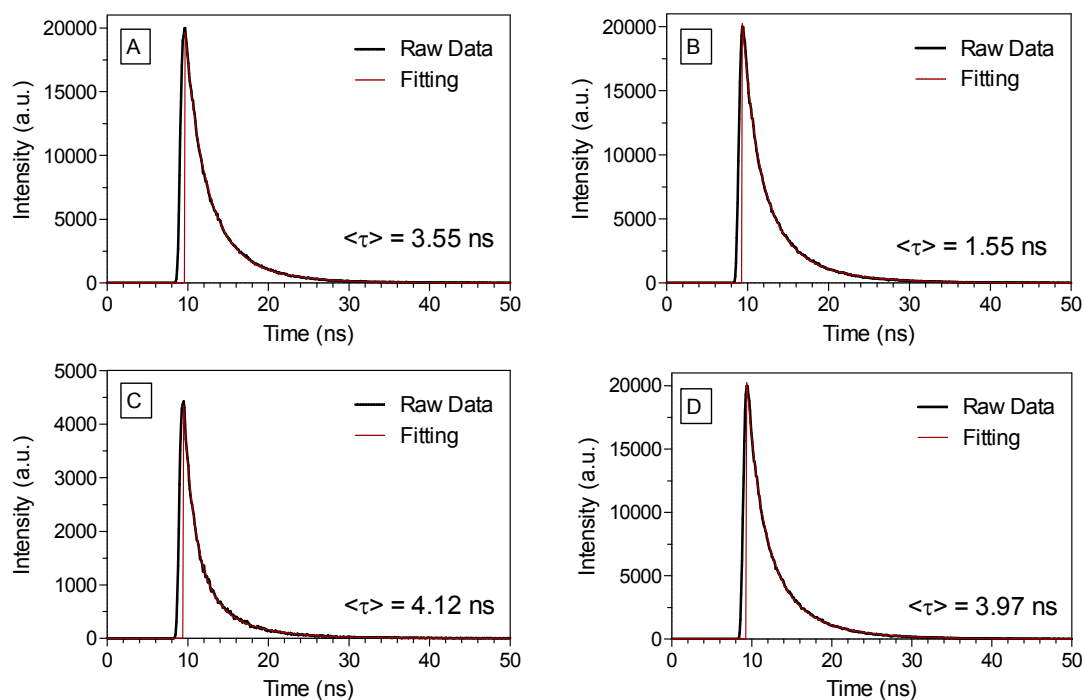
Angles between planes	
P <sub>1</sub> -P <sub>2</sub>	~0
P <sub>2</sub> -P <sub>3</sub>	~67
P <sub>2</sub> -P <sub>4</sub>	~67
P <sub>3</sub> -P <sub>4</sub>	~76

**Figure S5.** Summary of dihedral angles ( $^{\circ}$ ) for TPA-DCM. The dihedral angles between any two phenyl rings of TPA in TPA-DCM are from  $\sim 67^{\circ}$  to  $\sim 76^{\circ}$ .

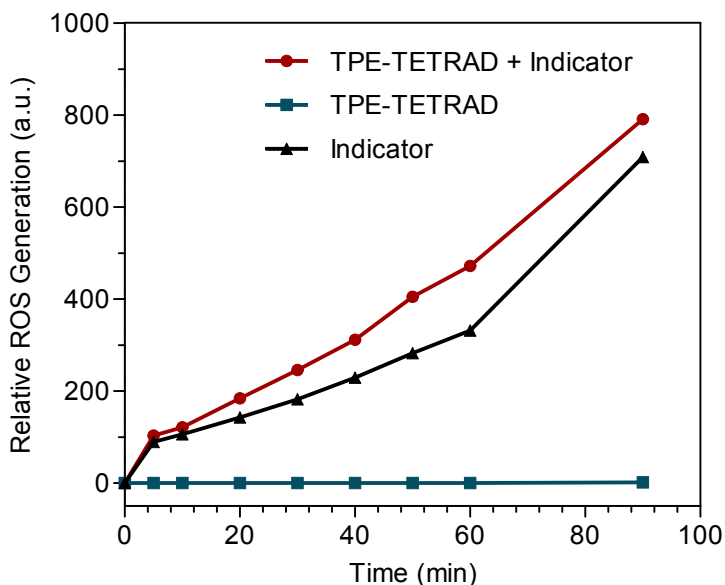


Dihedral angles (°)	
C5-C6-C7-C8	-48.4
C3-C4-C7-C8	-49.9
C2-C1-C7-C8	-50.0
C9-C10-C8-C7	-49.7
Angles between planes	
P <sub>1</sub> -P <sub>2</sub>	~0
P <sub>2</sub> -P <sub>3</sub>	~68
P <sub>2</sub> -P <sub>4</sub>	~67
P <sub>3</sub> -P <sub>4</sub>	~75
P <sub>4</sub> -P <sub>5</sub>	~35

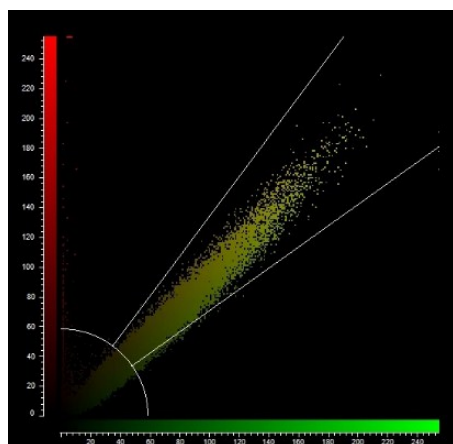
**Figure S6.** Summary of dihedral angles (°) for TPE-TETRAD. In the TPE component of TPE-TETRAD, the dihedral angles between any phenyl rings are ~50°. The dihedral angles between any two phenyl rings of TPA in TPE-TETRAD are from ~67° to ~75°.



**Figure S7.** Fluorescence lifetime of (A) TPE-TETRAD thin film solid, (B) TPE-TETRAD nanoaggregates (10  $\mu$ M) in water, (C) TPE-TETRAD@Biotin NPs (6  $\mu$ g/mL) in water and (D) TPE-TETRAD (10  $\mu$ M) in THF.



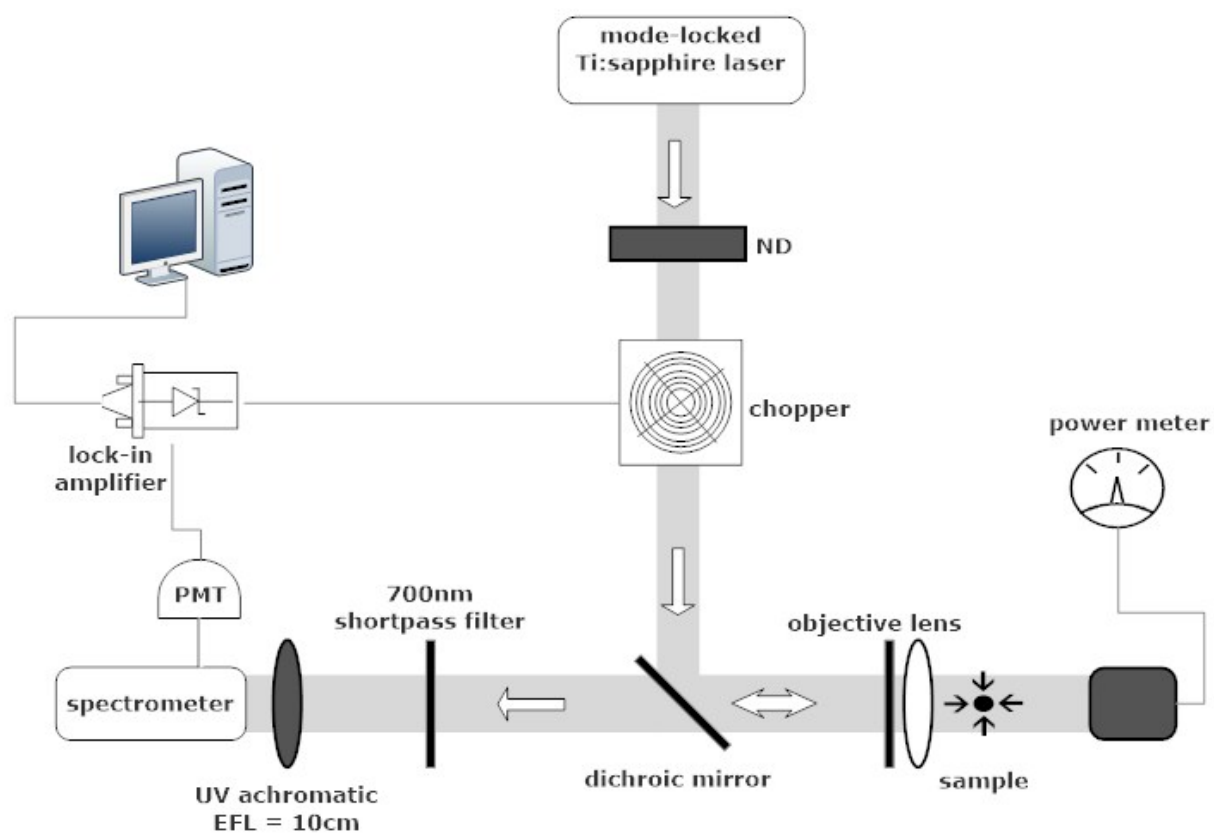
**Figure S8.** Relative ROS generation profile for TPE-TETRAD using DCFH-DA cellular reactive oxygen species detection assay kit. Change in fluorescent intensity at 534 nm for TPE-TETRAD, DCFH-DA indicator and their mixture in PBS upon white light irradiation for various time intervals. Excitation wavelength: 485 nm. [TPE-TETRAD] = 10  $\mu$ M; [DCFH-DA] = 1  $\mu$ M.



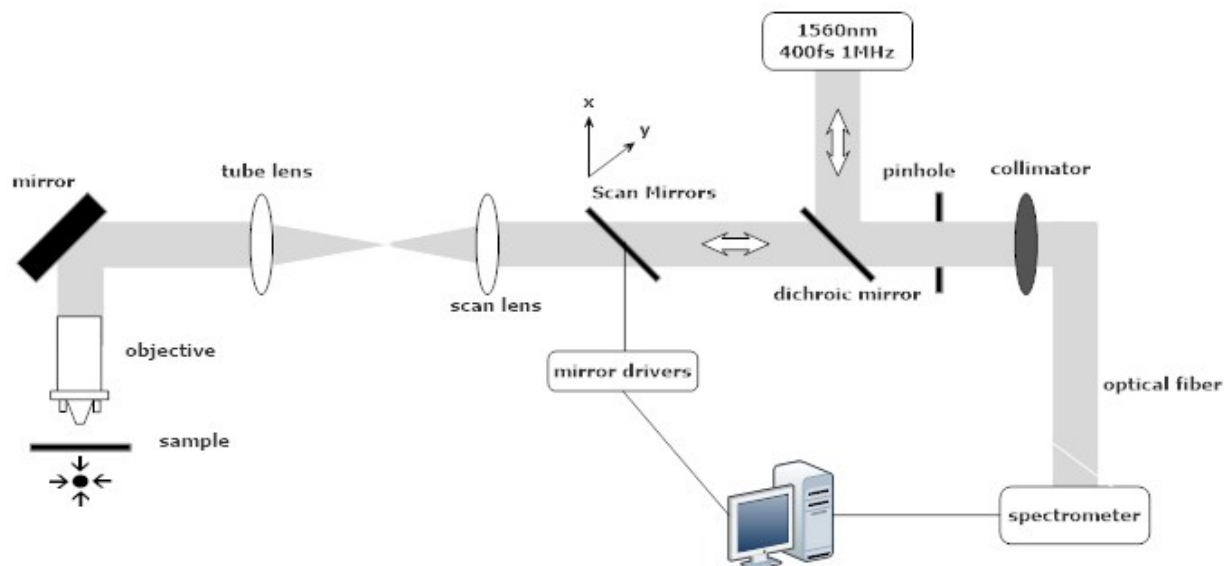
### Colocalization Statistics

Pearson's Correlation	0.919
Overlap Coefficient	0.9288
Colocalization Rate	94.53%
Colocalization Area	306.53 $\text{\AA}\mu\text{\AA}^2$
Area Image	5302.86 $\text{\AA}\mu\text{\AA}^2$
Area Foreground	324.27 $\text{\AA}\mu\text{\AA}^2$
Area Background	4978.70 $\text{\AA}\mu\text{\AA}^2$

**Figure S9.** CLSM colocalization statistics for HeLa cervical cancer cells stained with 25 nM MitoTracker Green for 10 min and 6  $\mu\text{g}/\text{mL}$  TPE-TETRAD@Biotin NPs for 1 h at 37  $^{\circ}\text{C}$ .



**Figure S10.** Schematic illustration of the experimental setup for two-photon excited fluorescence.



**Figure S11.** Schematic illustration of the NLO microscope system for spectrum measurement.

### Calculation of TPE-TETRAD@Biotin AIE NPs Concentration

The density of TPE-TETRAD@Biotin can be estimated as  $1 \text{ g/cm}^3$  due to its high water stability. The average size of the AIE NPs as verified by TEM and DLS was 100 nm. 0.25 mg of powder was obtained after freeze-drying 1 mL of the filtered stock solution. The TPE-TETRAD@Biotin concentration in a 1 mL stock suspension can be calculated from the following equation:

$$\frac{\text{Total volume of TPE - TETRAD@Biotin NP}}{\text{Average volume of each NP}} = \frac{0.25 \times 10^{-3}}{1 \text{ g/mL}} \div \frac{\frac{4}{3} \pi \times (50 \times 10^{-7})^3 \text{ mL}}{1 \text{ g/mL}} = 4.78 \times 10^{11}, \text{ Total number of NP in 1mL.}$$

$$[\text{TPE-TETRAD@Biotin}] = \frac{4.78 \times 10^{11}}{6.02 \times 10^{23} \text{ mol}^{-1}} \times 1 \times 10^{-3} \text{ L} = 0.8 \text{ nM, final concentration in stock solution}$$

$$\text{Working Concentration} = \frac{0.8 \text{ nM} \times 50 \mu\text{L}}{2000 \mu\text{L}} = 20 \text{ pM, final working concentration for cell imaging}$$

Or alternatively we can calculate the mass concentration:

$$\text{Working Concentration} = \frac{250 \mu\text{g/mL} \times 50 \mu\text{L}}{2000 \mu\text{L}} = 6.25 \mu\text{g/mL, final working concentration for cell imaging.}$$



