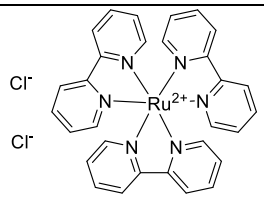
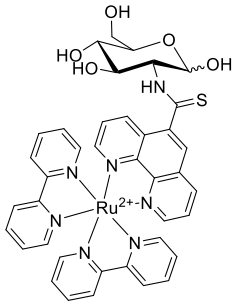
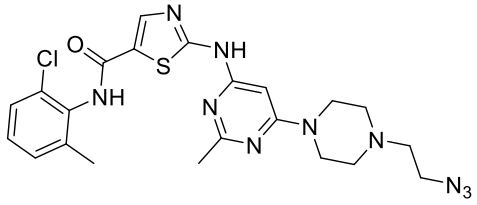
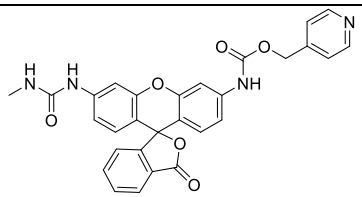
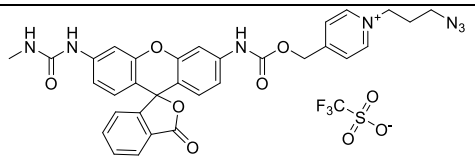
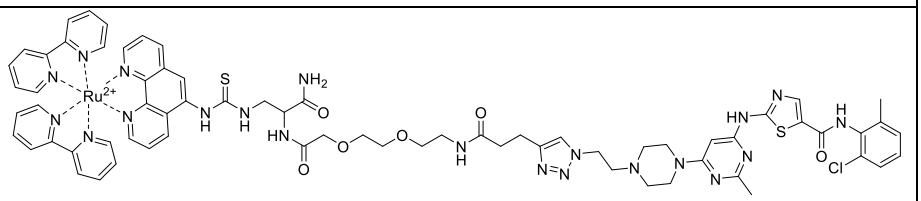
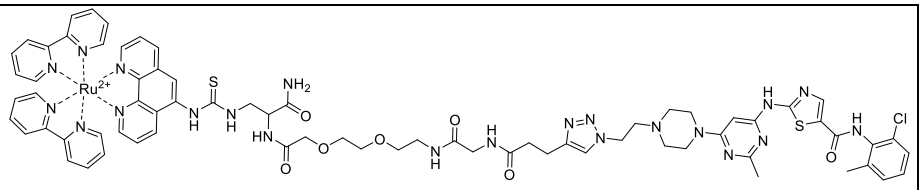
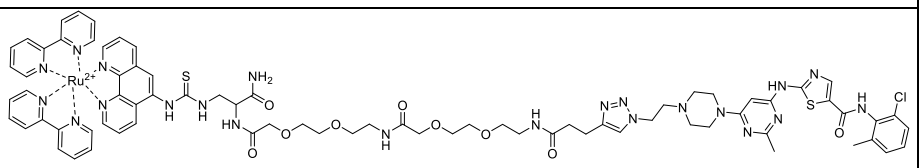
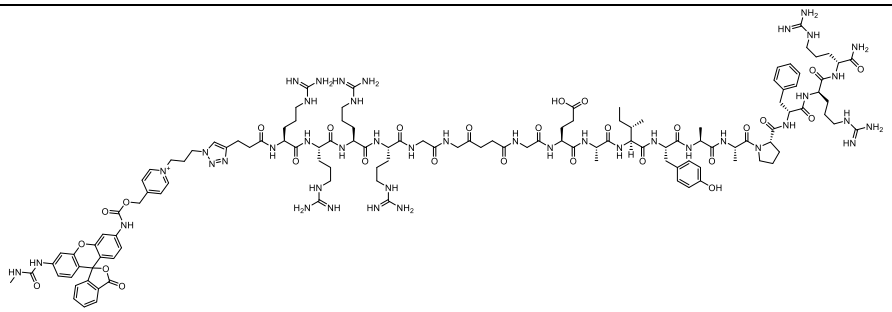
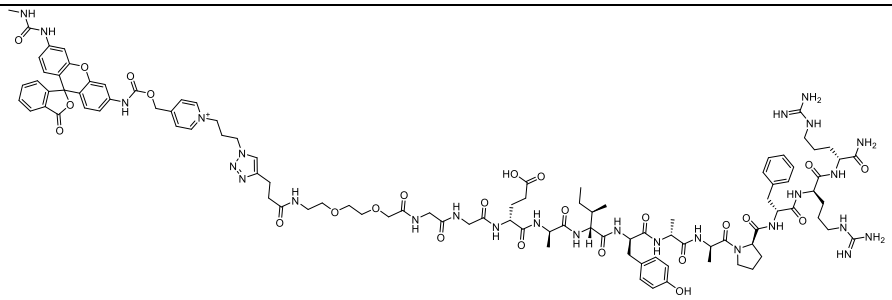
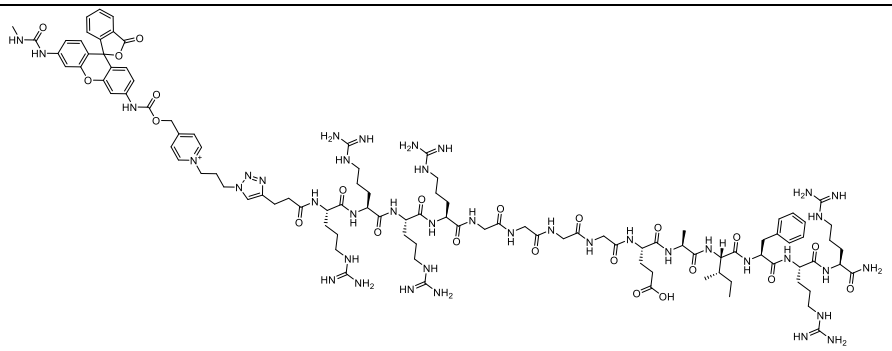
SUPPORTING INFORMATION

Contents

Supplementary Figures

Figure S1: Peptide and Dasatinib probes used

	Peptide or Dasatinib Probe	Structure
1	Ruthenium trisbipyridine $Ru(bpy)_3Cl_2$	
2	Glucosamine ruthenium GluAmRu [1]	
3	Dasatinib Azide[2]	
4	Rhodamine pyridine	
5	Rhodamine pyridinium	
6	Das-PEG-Ru	

7	Das-GPEG-Ru	
8	Das-PEG2-Ru	
9	PAbl-R4G2	
10	PAbl-PEG	
11	TR-PAbl-R4G2	

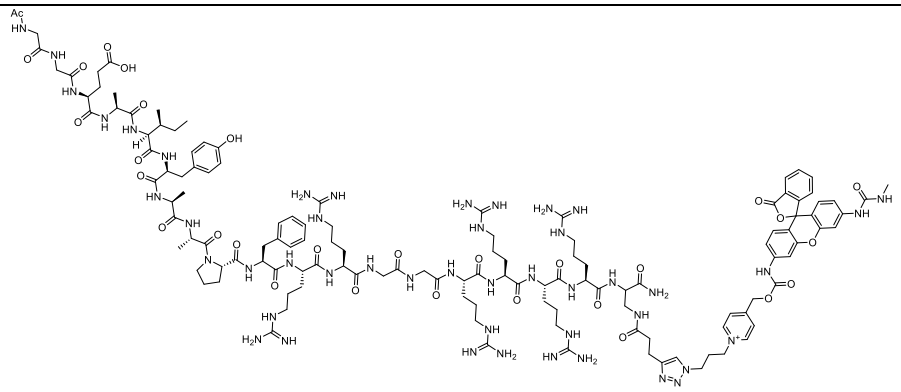
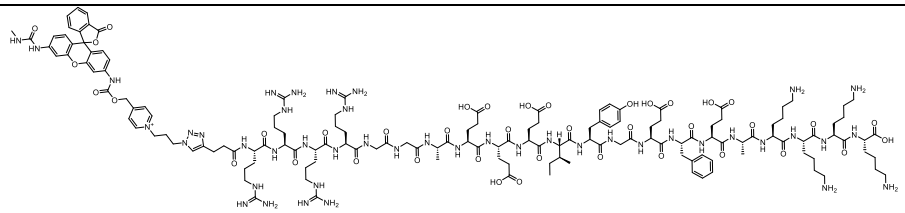
12	Cterm-PAbl-R4G2	
13	PSrc-R4G2	

Figure S2 : Templated reaction between PAbl-R4G2 (9) and Das-GPEG-Ru (7)

Templated reaction between PAbl-R4G2 (**9**) and Das-GPEG-Ru (**7**) in presence of Abl or CA and dasatinib (**3**). Similar conditions as Fig 4B. with 90 minutes measurement of the kinase templated reaction. Grey curves shows the calculated signal to noise ratio (right axis)

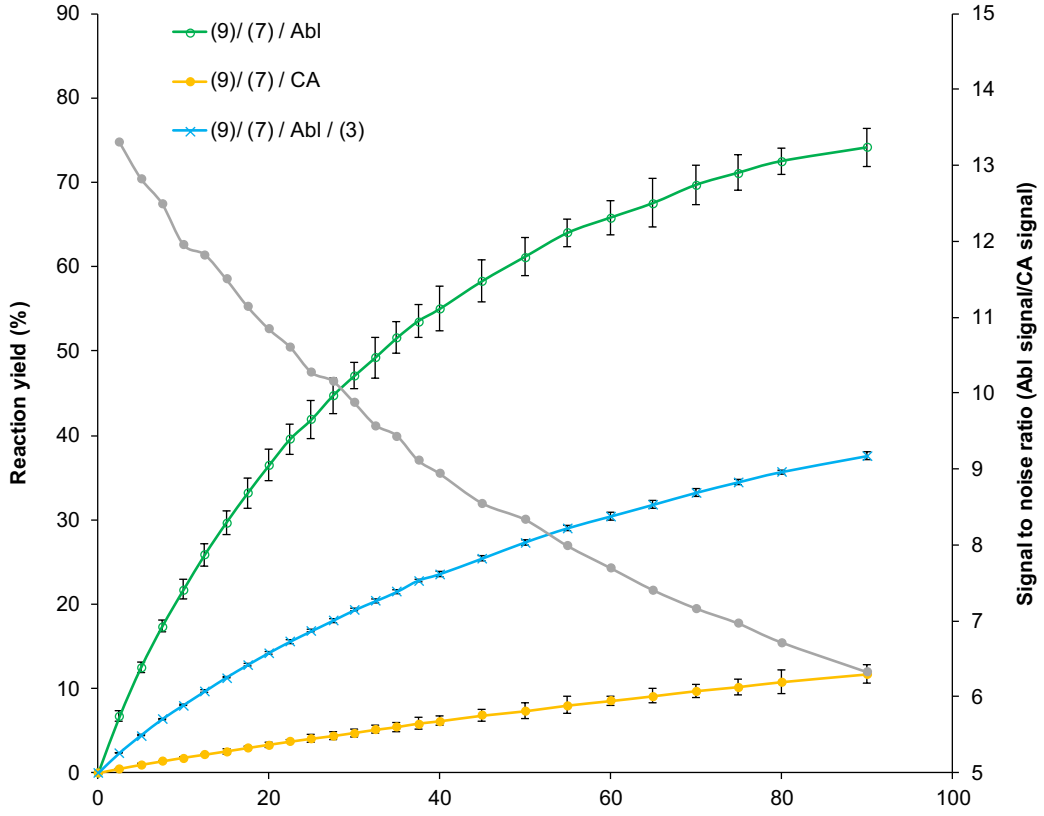
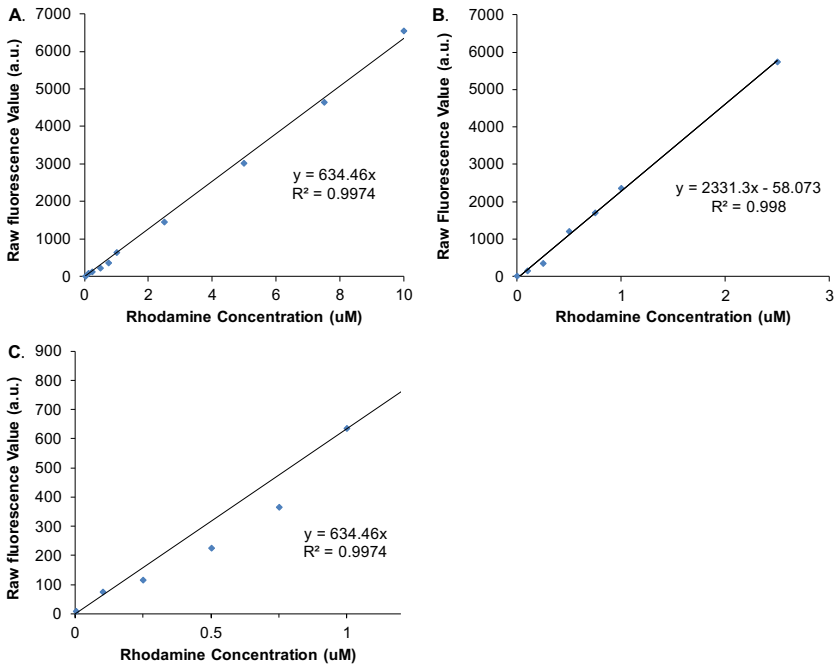


Figure S3: Rhodamine titration curve

(A) Titration curve using 100 µL per well
(B) Titration curve at 200 µL per well
(C) Zoom between 0 and 1 µM of curve displayed in (A)



General techniques

Anhydrous solvents were obtained by passing them through a commercially available alumina column (Innovative technology, MA). Solid phase syntheses were performed with NovaPEG Rink amide resin obtained from EMD Millipore. Peptide synthesis was performed according to previously reported protocols[3] using an automated peptide synthesizer (Intavis MultiPep instrument in 500 µL fritted tubes.). LC-MS were recorded by using a Thermo Scientific Accela HPLC equipped with a Thermo C18 (5 cm x 2.1 mm, 1.9 µm particles) Hypersil gold column coupled with A: Surveyor MSQ Plus spectrometer or B: LCQ Fleet mass spectrometer (both ESI, Thermo Scientific). Method: linear elution gradient for 95% H2O 0.01% TFA to 90% MeCN 0.01% TFA in 4 minutes at a flow rate of 1.0 mL/min. The MALDI spectra were measured using Bruker Daltonics Autoflex TOF/TOF spectrometer. Final compounds were purified by reverse-phase chromatography using a Biotage Isolera ONE equipped with a Biotage SNAP Cartridge KP-C18-HS (linear gradient from 100% H2O 0.01% TFA to 100% MeCN 0.01% TFA with a flow rate of 5 mL/min) or using an Agilent 1100 series HPLC equipped with DAD and with an Agilent ZORBAX Eclipse XDB-C18 (4.6 x 250 mm, 5 µm) column (linear gradient from 100% H2O 0.1% TFA to 1000% MeCN 0.1% TFA with a flow rate of 1 mL/min). All kinases, were purchased from ProQinase, except the Abl kinase domain, which was expressed and purified according to the reported procedure.[4, 5] Rhodamine **1**[6], dasatinib azide[2] and glucosamine ruthenium[1] were synthesized according to the reported procedures.

Resin Loading

The resin was washed with DMF and CH_2Cl_2 and treated with a preactivated (5 min) solution of the corresponding Fmoc-protected amino acid (5 equiv), HATU (4 equiv), DIPEA (5 equiv), and 2,6-lutidine (7.5 equiv) for 2 hrs in NMP. Following the reaction, the resin was washed with DMF (3x), CH_2Cl_2 (3x), DMF (3x) and CH_2Cl_2 (3x). The resin was capped for at least one hour using acetic anhydride (5.3 equiv) and 2,6-lutidine (6.4 equiv) in DMF (150 µL / 5 mg resin)

Automated Peptide Synthesis

Peptides were synthesized in 500 mL fritted tubes by using an Intavis MultiPep instrument in a fully automated fashion. The resin (5 mg, 1 µmol per column) was swollen in CH_2Cl_2 (300 µL) for 20 min and deprotected using 20% piperidine in DMF (2x5 min). The resin was then washed with DMF and CH_2Cl_2 and treated with a preactivated (5 min) solution of the corresponding Fmoc-protected amino acid (5 equiv), HATU (4 equiv), DIPEA (5 equiv), and 2,6-lutidine (7.5 equiv) for 20 min in NMP. This process was repeated once (double couplings). Each coupling was followed by a capping step with Ac_2O (5.3 equiv) and 2,6-lutidine (6.4 equiv) in DMF (150 µL per column).

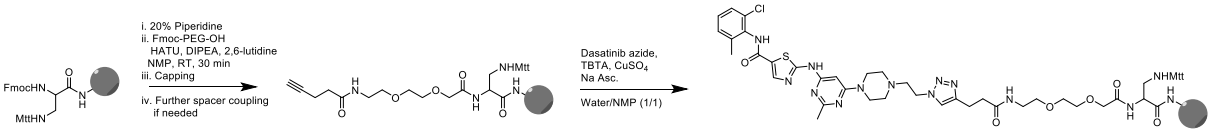
Fmoc deprotection (manual or on Intavis AG Multipep RS Synthesizer)

To 5 mg of NovaPEG Rink amide resin were added 200 µL of 20% piperidine solution in DMF. After 2 min, the resin was washed with 6 x 300 µL DMF and the sequence was repeated a second time for 4 min. Finally, the resin was washed with 6 x 300 µL of DMF and 6 x 300 µL of CH_2Cl_2 and the washing cycle was repeated a second time.

Cleavage from the resin

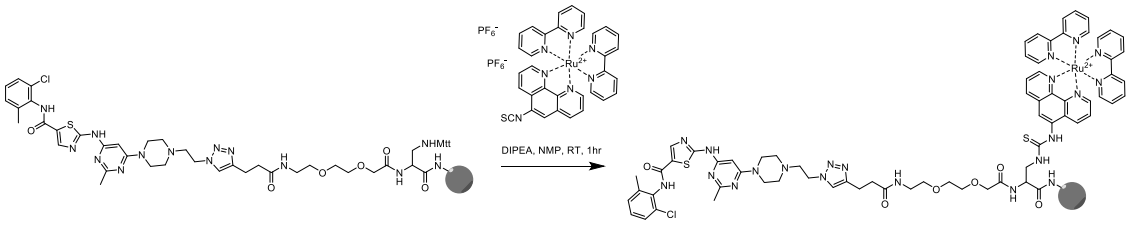
5 mg of resin (0.44 mmol/g loading) were treated with 200 µL TFA for 2 hours. After filtration of the resin, the resulting solution was precipitated in 12 mL of Et_2O, stored in a -20°C freezer for 5 min and centrifuged to recover the product as a pellet. The pellet was washed once more with ether following the previous process. The product was recovered as a pellet and dried under nitrogen flow and subsequently dissolved in water or 10% DMSO in water for purification.

Preparation of dasatinib derivatives



Rink amide NovaPEG resin was loaded with Fmoc-D-diaminopropionic(Mtt) acid at 0.44mmol/g.
Resins loaded with Fmoc-D-diaminopropionic(Mtt) acid (5 mg, 0.44 mmol/g, 2.2 µmol) were deprotected at their alpha position and treated with a pre-activated (5 min) solution of 4-pentynoic acid (5 equiv), HATU (4 equiv), DIPEA (5 equiv) and 2,6-lutidine (7.5 equiv) for 60 min in NMP. After the reaction, the resins were washed with DMF and CH_2Cl_2 (5 times each). A capping step as described in the automated peptide synthesis was performed prior to click reaction.
TBTA (1.5 equiv, 3.75 µmol, 2 mg) was suspended in 50 µL of a solution of sodium ascorbate (2M in water, 198 mg/500 µL) and 15 µL of copper sulfate (64.2 mg/mL in water). The resulting black suspension was sonicated until color changed to beige. Dasatinib azide[2] (1.05 equiv,2mg, 2.6 µmol) was dissolved in 50 µL NMP and the resulting solution was added to the previous suspension. The clear light orange mixture was transferred to the resin and was shaken overnight at room temperature. Subsequently, the resin was washed DMF (3 x 250 µL), water (6 x 250 µL), DMF (4 x 250 µL) and CH_2Cl_2 (6 x 250 µL).

Preparation of final Ruthenium derivatives



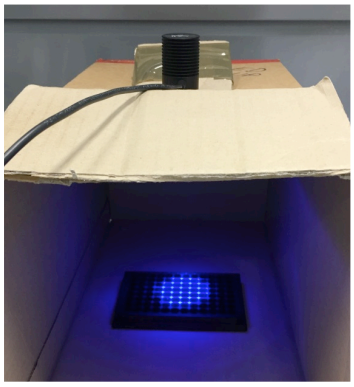
Dasatinib constructs on resin were deprotected at their C-terminus using a 150 µL per column of a solution of 244 mg HOBt in 20 mL of a 1:1 solution of 1,2-dichloroethane and hexafluoroisopropanol, this process was repeated 3 times, until no yellow color could be observed. The resin was subsequently washed with DMF, CH_2CL_2, DMF and CH_2CL_2 (3 x each).
Ruthenium isothiocyanate (1.1 equiv, 2.75 µmol, 2.6 mg) was dissolved in 100 µL NMP and added to the resin. The resin was shaken for 1 hr, protected from light at room temperature. After reaction, the resin was washed with DMF, CH_2Cl_2, DMF and CH_2Cl_2 (3 x each) and cleaved from resin using cleavage procedure. The final derivatives were purified by reverse phase HPLC.

Rhodamine coupling to peptides

N-terminal deprotected peptides (5 mg 0.44 mmol/g, 2.2 µmol) were treated with a pre- activated (5 min) solution of 4-pentynoic acid (5 equiv), HATU (4 equiv), DIPEA (5 equiv) and 2,6-lutidine (7.5 equiv) for 60 min in NMP. After the reaction, the resins were washed with DMF and CH_2Cl_2 (5 times each).
TBTA (1.5 equiv, 3.75 µmol, 2 mg) was suspended in 50 µL of a solution of sodium ascorbate (2 M in water, 198 mg/500 µL) and 15 µL of copper sulfate (64.2 mg/mL in water). The resulting black suspension was sonicated until color changed to beige. Final Rhodamine derivative **3** (1.05 equiv,2 mg, 2.6 µmol) was dissolved in 50 µL NMP and the resulting solution was added to the previous suspension. The clear light orange mixture was transferred to the resin and was shaken overnight at room temperature. Subsequently, the resin was washed DMF (3 x 250 µL), water (6 x 250 µL), DMF (4 x 250 µL) and CH_2Cl_2 (6 x 250 µL). The resin was suspended in TFA (200 µL) for 2 h. The solution was filtered and the product precipitated in diethyl ether (12 mL). After 5 min at -20°C (freezer) the precipitate was pelleted by centrifugation and the supernatant was removed. The pellet was washed with diethyl ether then dissolved in 300 µL of 10 % DMSO in water for HPLC purification as described in the general methods.

Template reaction procedure

Template reactions were carried out in opaque 96 well plates in 100 mM TBS (Tris Buffer Saline 150 mM NaCl) + 0,05% Tween 20 buffer and 20 mM of sodium ascorbate. Stock solutions of probes were made in 10% DMSO in mQ water at 1 mM and stored either at -20°C or in aliquots at -80°C. A stock solution of sodium ascorbate was made in water at 1 M. Stock solutions were diluted with reaction buffer to the desired concentration and used in the experiment. Each experiment was performed in triplicate. After measurement of initial fluorescence, the plate was irradiated using a 455 nm collimated LED lamp (www.thorlabs.com, 1W, 30cm distance) and the fluorescence of rhodamine (exc=490 nm; em=530 nm) was measured at different time points. The reactions were carried out in 200 µL volume at two different reagent concentrations: at higher concentration with 1 µM of peptide-rhodamine probe and 100nM of dasatinib-ruthenium probe and 100 nM of protein; at lower concentration using 500 nM of peptide-rhodamine probe and 100nM of dasatinib-ruthenium probe and 50 nM of template protein. The raw data was treated by subtracting the initial fluorescence value for each conditions and averaging the values of triplicate experiment while calculating a standard deviation. Picture of the LED setup below.



Cell Culture

A549 cells were grown in F12K medium (10% FCS, 1 x penstrep), K562 cells were grown in IMDM (10% FCS, 1 x penstrep), KCL-22 cells were grown in RPMI 1640 (10% FCS, 1 x penstrep), and SKBR-3 cells were grown in McCoy's 5A medium (10% FCS, 1 x penstrep) in accordance with ATCC and DSMZ recommendations.

Cell Experiments

Adherent cell lines, A549 and SKBR-3, were seeded in 35 mm glass bottom dishes and grown for 24h. Cells were then washed with DPBS and fixed for 20 min with 4% PFA. For K562, the cells were seeded in 35 mm glass bottom dishes, coated with poly-L-lysine and grown for 24h. Cells were then washed with DPBS and fixed with 4% PFA for 20 mins. For KCL-22, cells were seeded in 35 mm glass bottom dishes coated with poly-L-lysine and gently centrifuged (300 rpm, 3 min, rt). The cells were then fixed with 4% PFA for 20 mins.

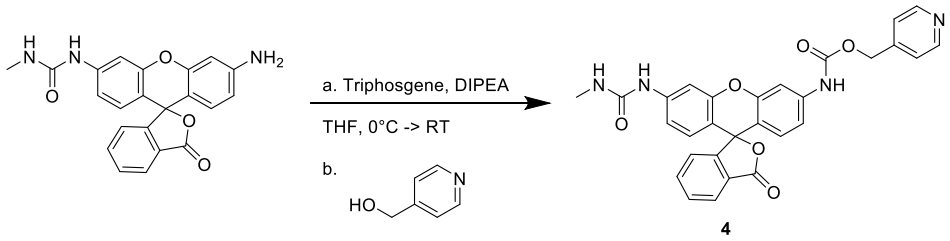
Peptide and Dasatinib conjugates were added to the cells in DPBS at the concentrations indicated and incubated at 37°C for 2 h. Sodium ascorbate (20 mM) was added and the cells were further incubated at 37°C for 1 h. The media was aspirated and replaced with 2 mL of DPBS containing 20 mM sodium ascorbate. The cells were then irradiated for 30 min with a 455 nm LED lamp. After irradiation the cells were imaged using confocal microscopy.

Confocal Miscrocopy

Images were acquired using a Zeiss LSM780 inverted confocal microscope, or a Leica SP8 inverted confocal microscope with 488 nm laserlines. Acquired images were analyzed and quantified using ImageJ software.

Structure and Characterization of probes

Rhodamine Pyridine Synthesis 4



Rhodamine was synthesized as reported in Ref 6.
In a Schlenk flask, Rhodamine[6] (100 mg, 0.26 mmol) and triphosgene (0.5 equiv, 38.5mg, 0.13 mmol) were suspended in dry tetrahydrofurane (5 mL). The suspension was cooled to 0°C with an ice bath and diisopropylethylamine (3.0 equiv, 0.77 mmol, 135 µL) was added dropwise. The reaction turned from a red suspension to a clear purple solution within minutes. The ice bath was removed and the activation was continued for 1 hr at room temperature. The activation of Rhodamine to the Rhodamine isocyanate was monitored by TLC or LC/MS. 4-pyridine methanol (3.0 equiv, 84 mg, 0.77 mmol) was added to the previous solution. The reaction was monitored by TLC or LC/MS and stirred overnight under nitrogen at room temperature. The following day, the reaction mixture was concentrated *in vacuo* to remove THF and water and CH_2CL_2 were added. The layers were separated and the aqueous layer was extracted with CH_2CL_2. The combined organic layers were washed with brine, dried over sodium sulfate and concentrated under reduced pressure. Rhodamine-pyridine was purified by reverse phase using a biotage and a gradient from 5% to 50% acetonitrile to obtain 113 mg of Rhodamine-pyridine **2** as a red solid. Yield 84%. LC/MS ESI rt = 1.84 min, m/z calculated = 522.15 m/z found 523.28 [M+H$^+$]. ^{1}H NMR (500 MHz, Acetone-d6) δ 9.21 (s, 1H), 8.61 (s, 2H), 8.19 (s, 1H), 7.87 (dt, J = 7.6, 1.0 Hz, 1H), 7.71 (d, J = 2.1 Hz, 1H), 7.68 (td, J = 7.5, 1.2 Hz, 1H), 7.63 – 7.59 (m, 1H), 7.59 (s, 1H), 7.55 (d, J = 5.2 Hz, 2H), 7.18 (dt, J = 7.7, 0.9 Hz, 1H), 7.09 (dd, J = 8.7, 2.2 Hz, 1H), 6.87 (dd, J = 8.7, 2.2 Hz, 1H), 6.65 (d, J = 8.6 Hz, 1H), 6.54 (d, J = 8.7 Hz, 1H), 5.27 (s, 2H), 2.63 (s, 3H). ^{13}C NMR (126 MHz, Acetone) δ 168.60, 153.12, 151.72, 147.12, 135.22, 129.92, 128.62, 128.13, 126.72, 124.54, 124.03, 114.26, 114.02, 113.94, 105.67, 104.83, 104.76, 78.12, 77.86, 64.27.

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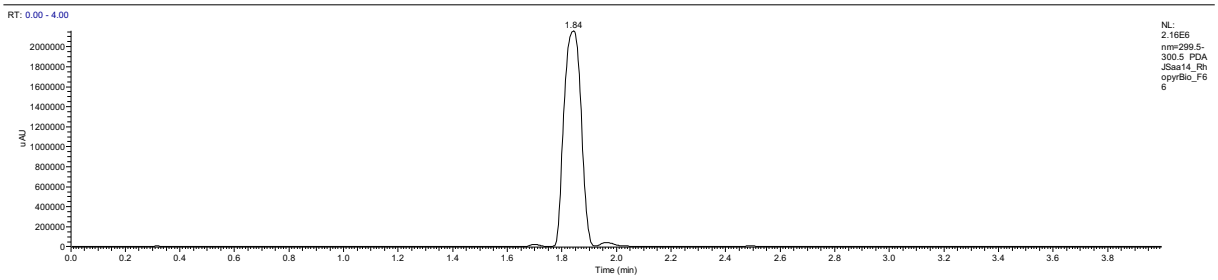
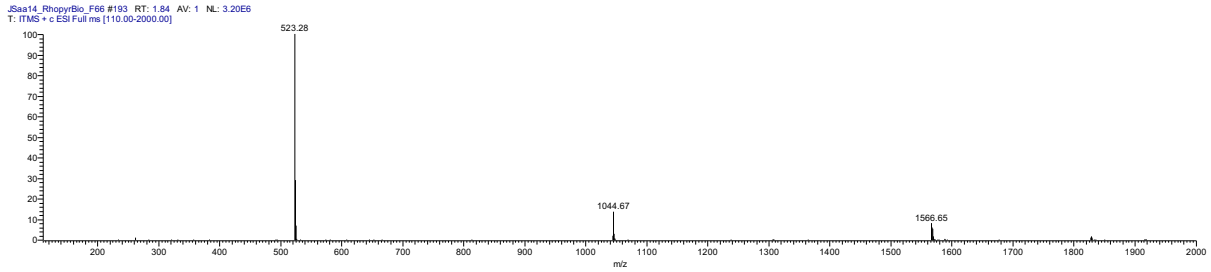
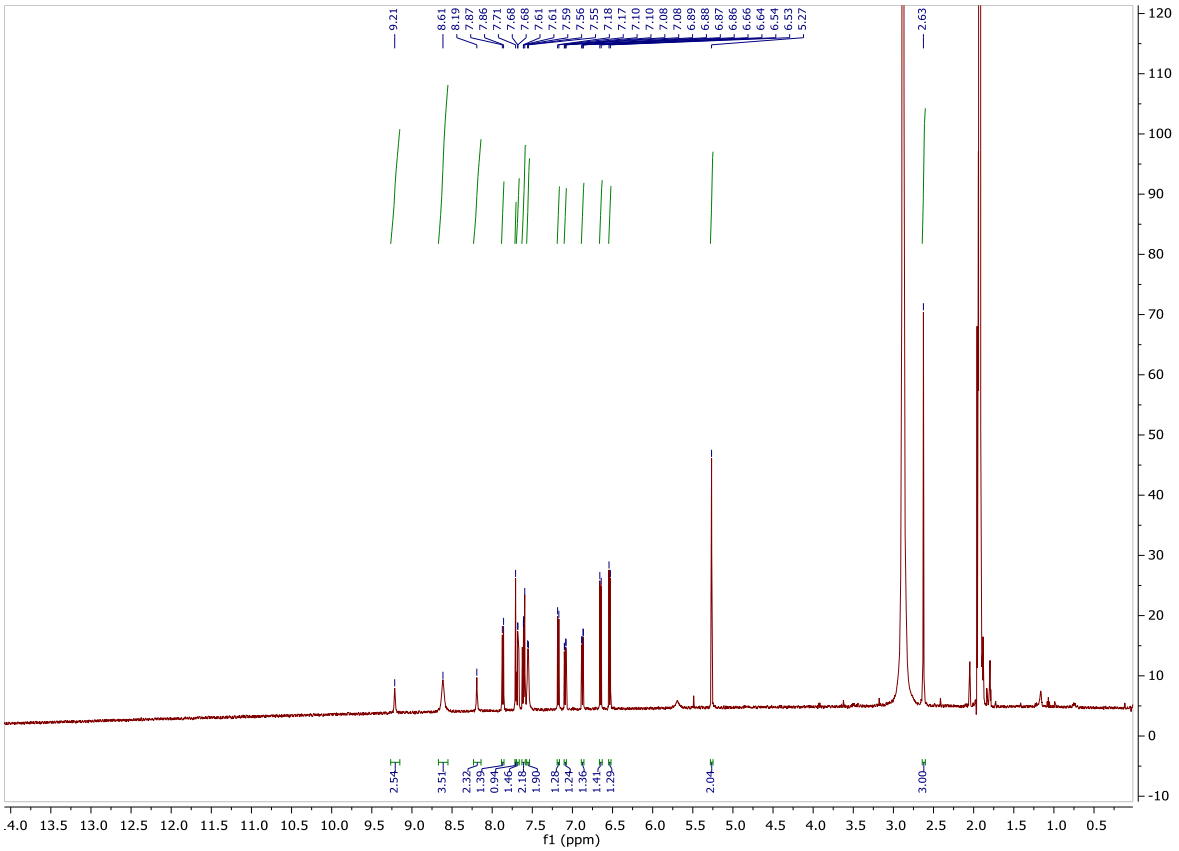
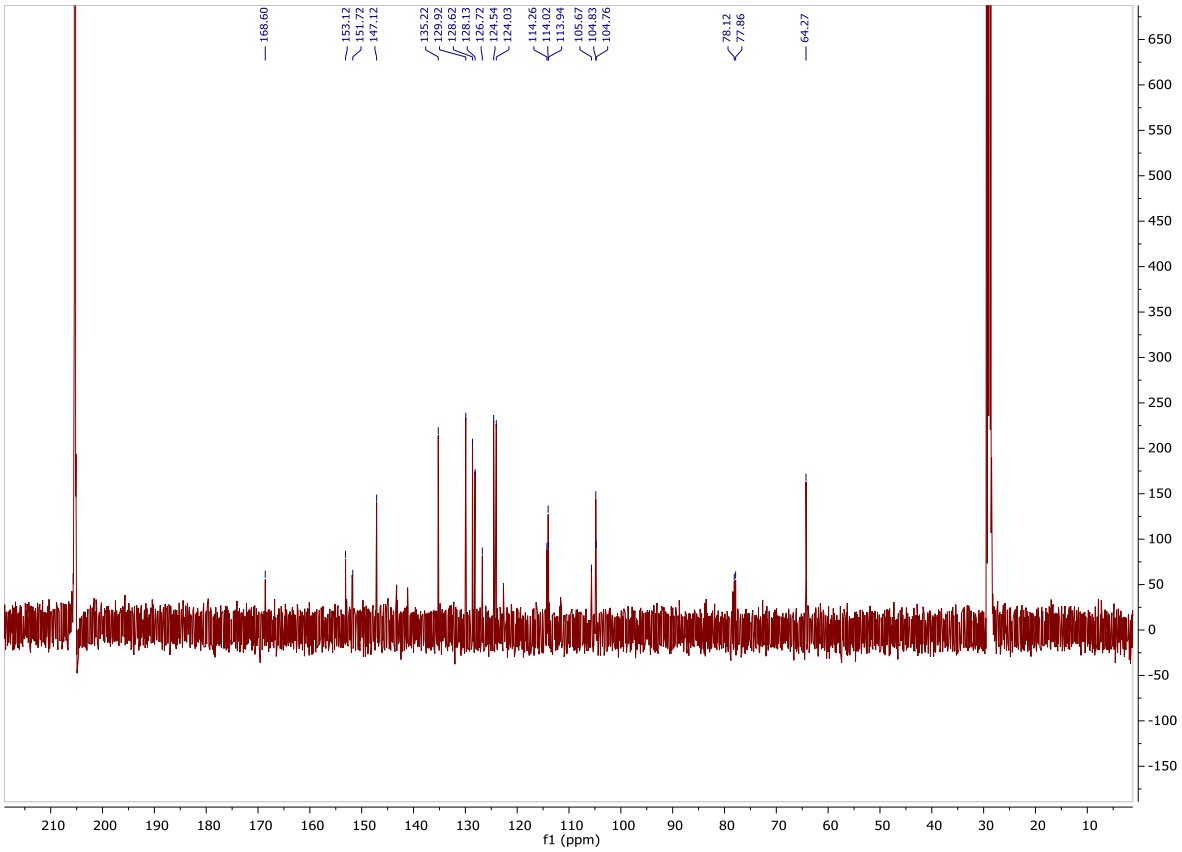
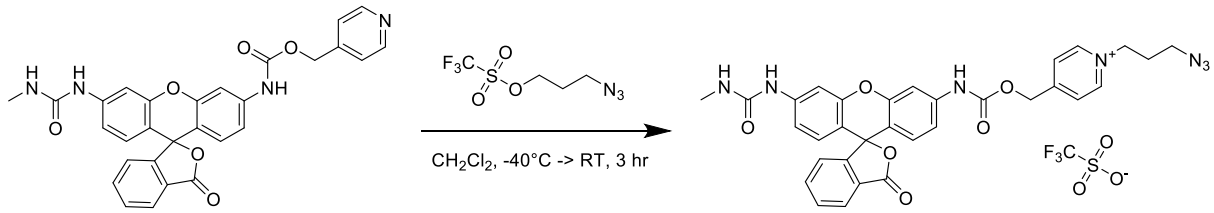








Rhodamine Azidopropyl-Pyridinium Synthesis 5



4 **5**

In a Schlenk flask, Rhodamine-pyridine **2** (1 equiv, 30 mg, 0.057 mmol) was suspended in dry CH_2Cl_2 (3 mL) and cooled to -40°C in an acetone/dry ice bath. Azidopropyl triflate (1.5 equiv, 0.0855 mmol) was added dropwise. The reaction was stirred for 30 min at -40°C, the ice bath was removed and the reaction mixture was allowed to warm to room temperature in 3 hrs. The reaction mixture was monitored by LC/MS and once alkylation was complete, the mixture was concentrated *in vacuo*. The alkylated rhodamine pyridinium was purified by reverse phase biotage or HPLC using gradient of respectively 5 to 50% and 10 to 80% acetonitrile in water. Lyophilization of the purified fractions gave 30 mg of rhodamine pyridinium **3** as a light orange solid. Yield (75%) LC-MS (ESI) RT = 1.95 min, m/z calculated = 606.21 m/z found = 303.67 $[M+H]^{2+}$, 606.42 $[M]^{+}$, 1H NMR (500 MHz, Acetone-d6) δ 9.18 (d, J = 6.3 Hz, 2H), 8.26 (d, J = 6.2 Hz, 2H), 7.98 (d, J = 7.5 Hz, 1H), 7.82 (d, J = 2.0 Hz, 1 H), 7.79 (td, J = 1.2 Hz, 7.5 Hz, 1H), 7.72 (td, J = 1.0 Hz, 7.6 Hz), 7.66 (d, J = 2.2 Hz, 1H), 7.29 (d, J = 7.7 Hz, 1H), 7.22 (dd, J = 2.2 Hz, 8.8 Hz, 1H), 7.13 (dd, J = 2.1 Hz, 8.8 Hz, 1H), 6.74 (d, J = 8.7 Hz, 1H), 6.62 (d, J = 8.6 Hz) 5.53 (s, 2H), 4.94 (t, J = 7.3 Hz, 2H), 3.61 (t, J = 6.4 Hz, 2H), 2.72 (s, 3H), 2.43 (m, 2H). 13C NMR (126 MHz, Acetone) δ 168.70, 157.87, 153.09, 152.58, 151.85, 151.63, 144.84, 144.06, 143.94, 140.99, 140.87, 135.23, 129.90, 128.53, 127.92, 126.71, 125.35, 124.52, 124.05, 114.32, 114.23, 114.14, 114.06, 113.91, 110.87, 105.85, 104.58, 104.51, 82.53, 63.60, 59.06, 47.83, 30.13.

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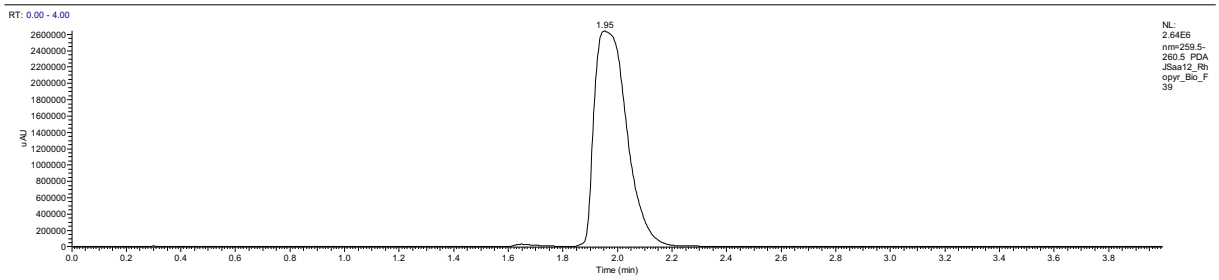
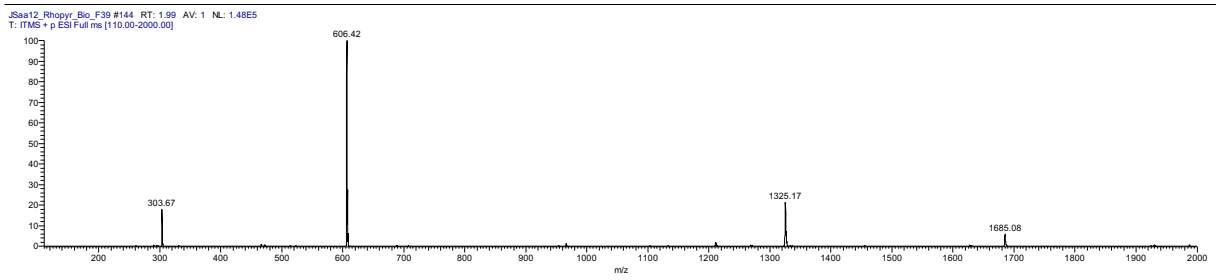
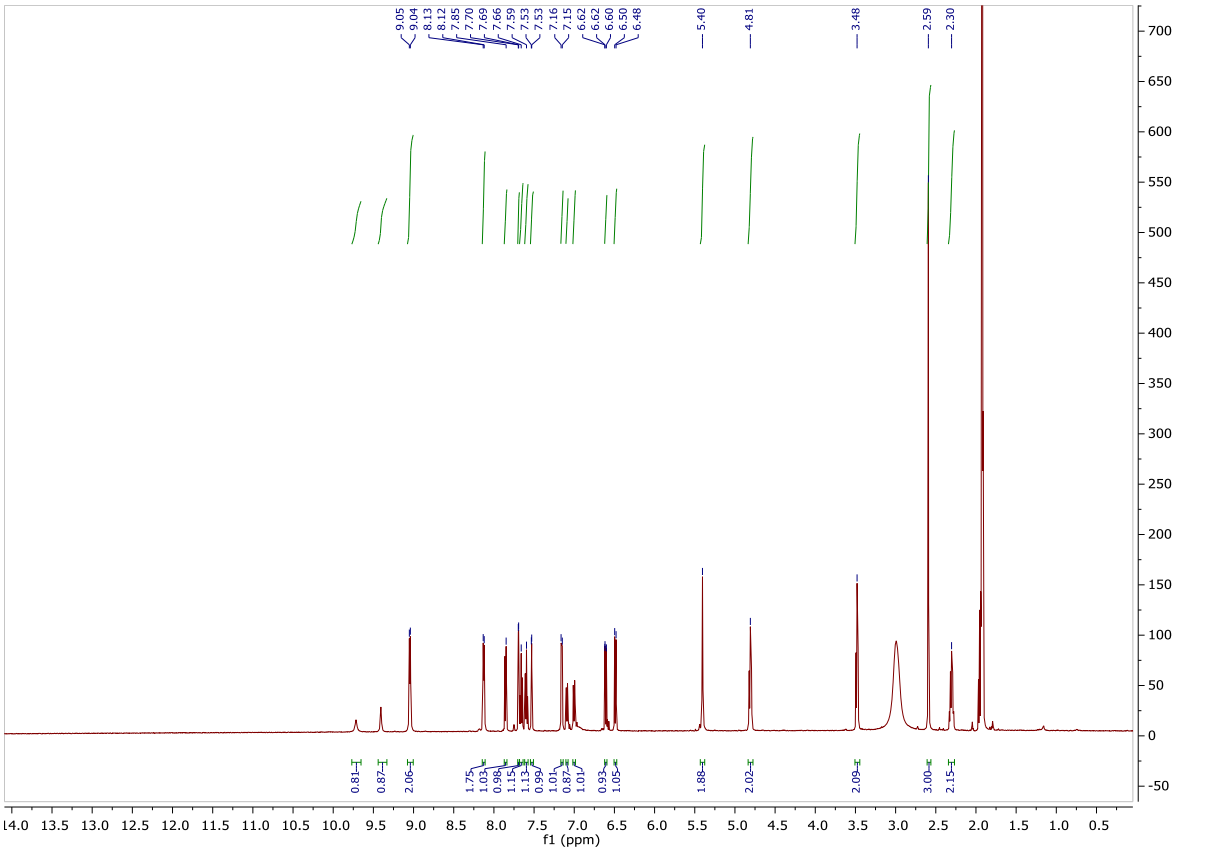
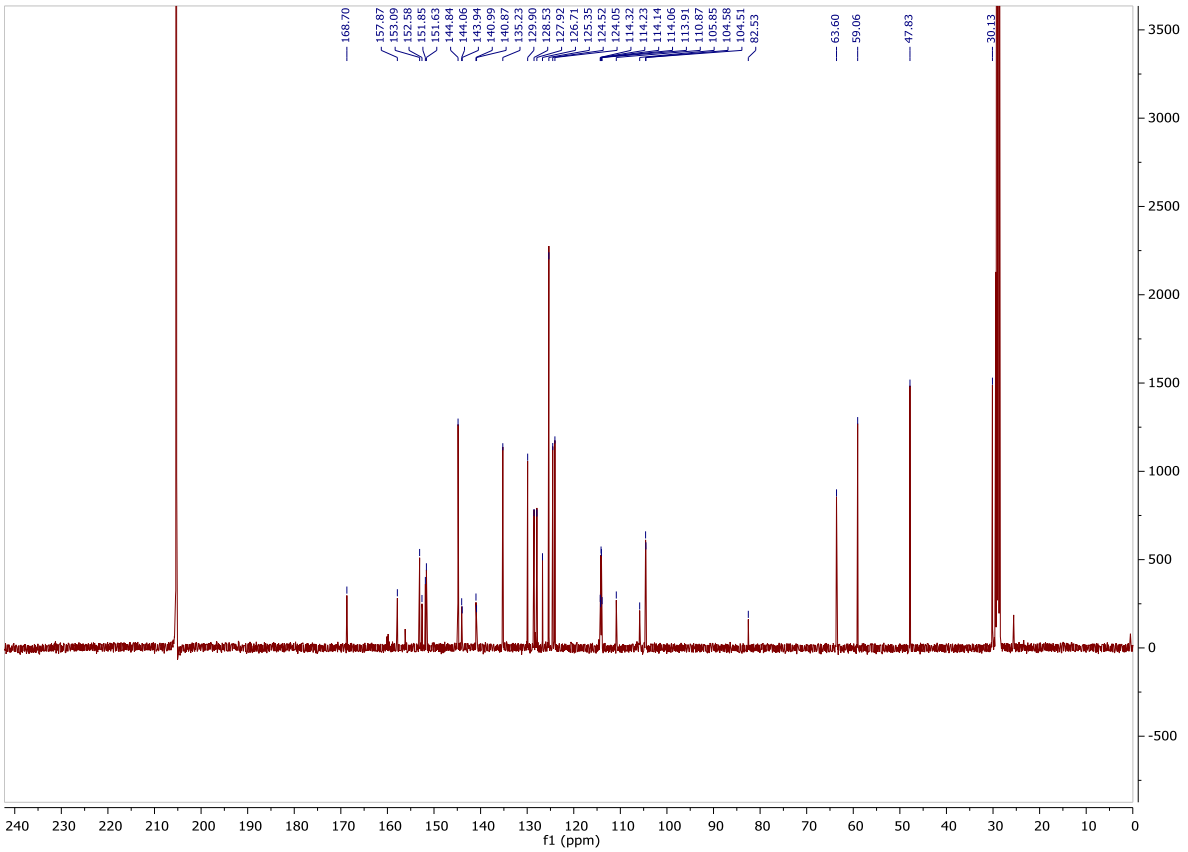


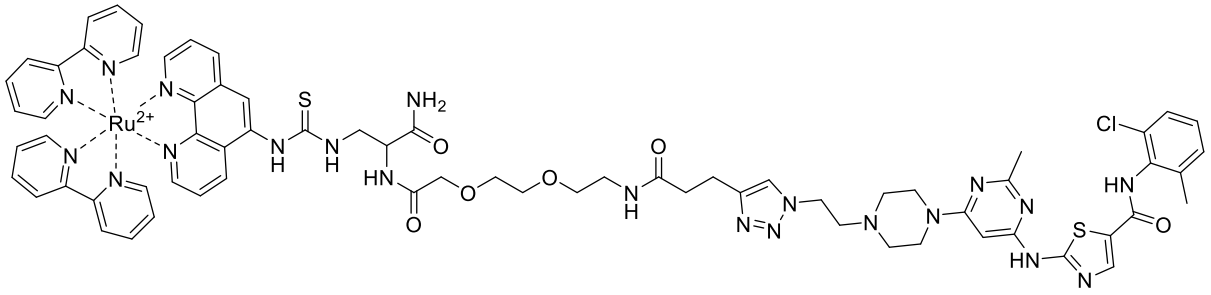




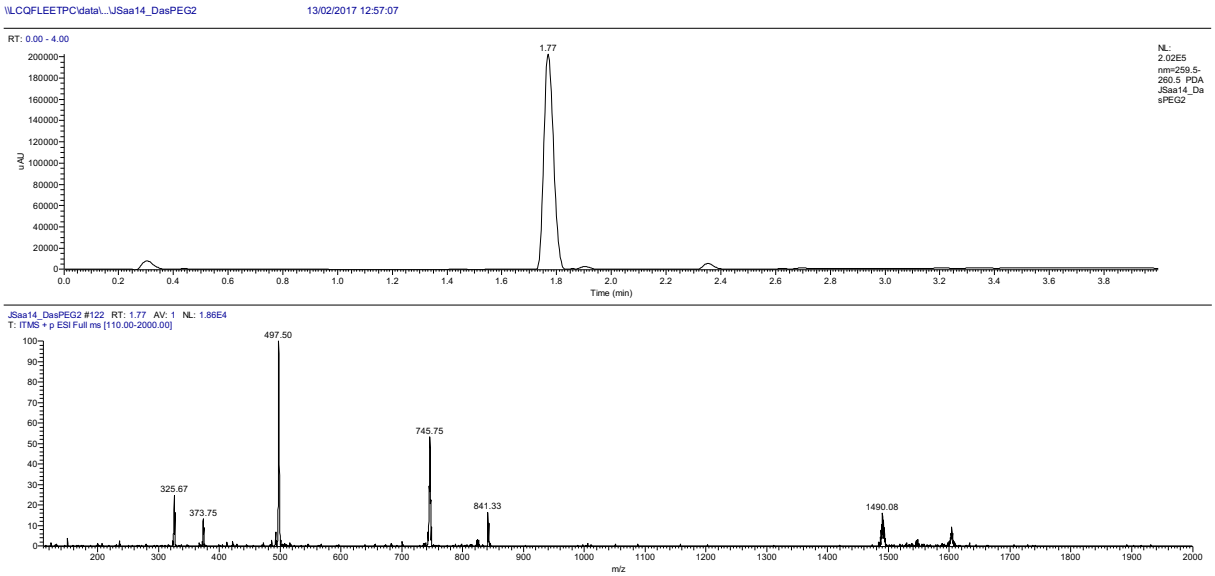

168.70
157.87
153.09
152.58
151.85
151.63
144.84
144.06
143.94
140.99
140.87
135.23
129.90
128.53
127.92
126.71
125.35
124.52
124.05
114.32
114.23
114.14
114.06
113.91
110.87
105.85
104.58
104.51
82.53
63.60
59.06
47.83
30.13
3500
3000
2500
2000
1500
1000
500
0
-500
240 230 220 210 200 190 180 170 160 150 140 130 120 110 100 90 80 70 60 50 40 30 20 10 0
f1 (ppm)

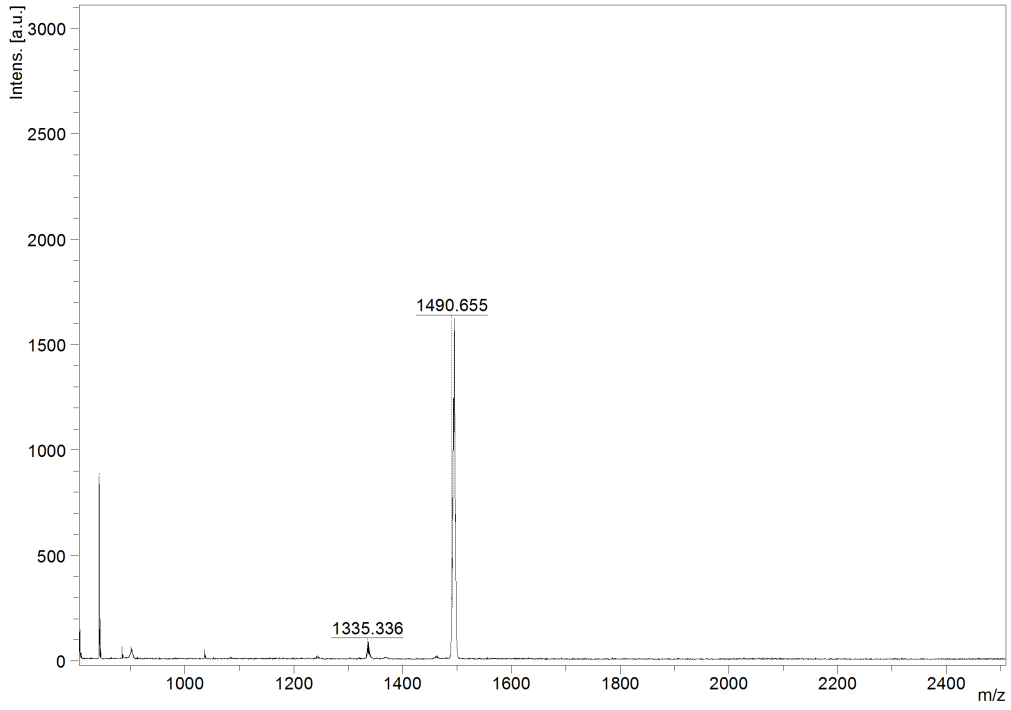
MALDI-TOF and LC-MS (ESI) characterization of probes

Das-PEG-Ru **6**

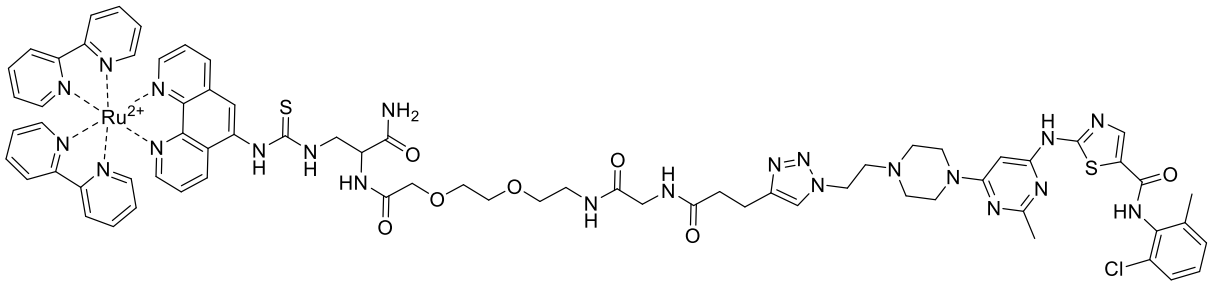


Chemical Formula: $C_{69}H_{72}ClN_{21}O_6RuS_2^{2+}$ Exact Mass: 1491.41 LC-MS (ESI): RT= 1.77 min, *m/z* found = 325.67 [Ruthenium]$^{2+}$, 373.75 [M+2H]$^{4+}$, 497.50[M+H]$^{3+}$, 745.75 [M]$^{2+}$, 841.33 [M+H-Ruthenium] $^{+}$, 1490.08 [M-H]$^{+}$, MALDI-TOF found *m/z* = 839.40 [M+H-Ruthenium]$^{+}$, 1335.336 [M-H-bpy]$^{+}$, 1490.655 [M-H]$^{+}$

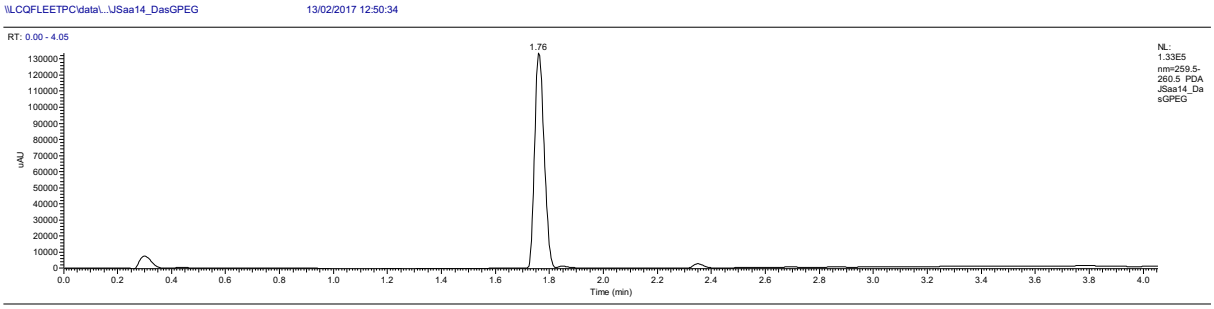


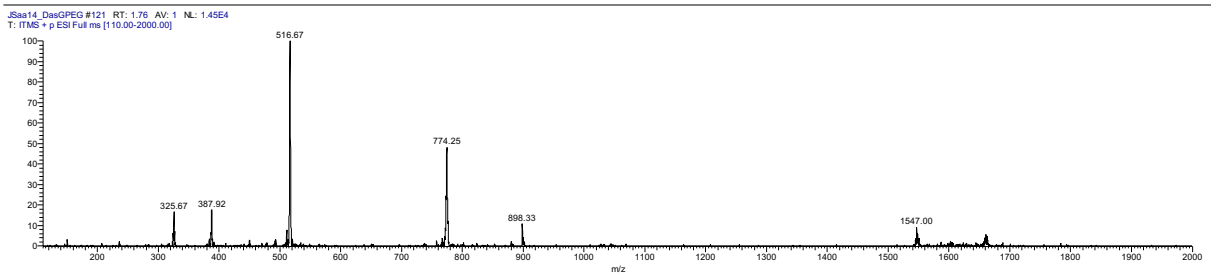


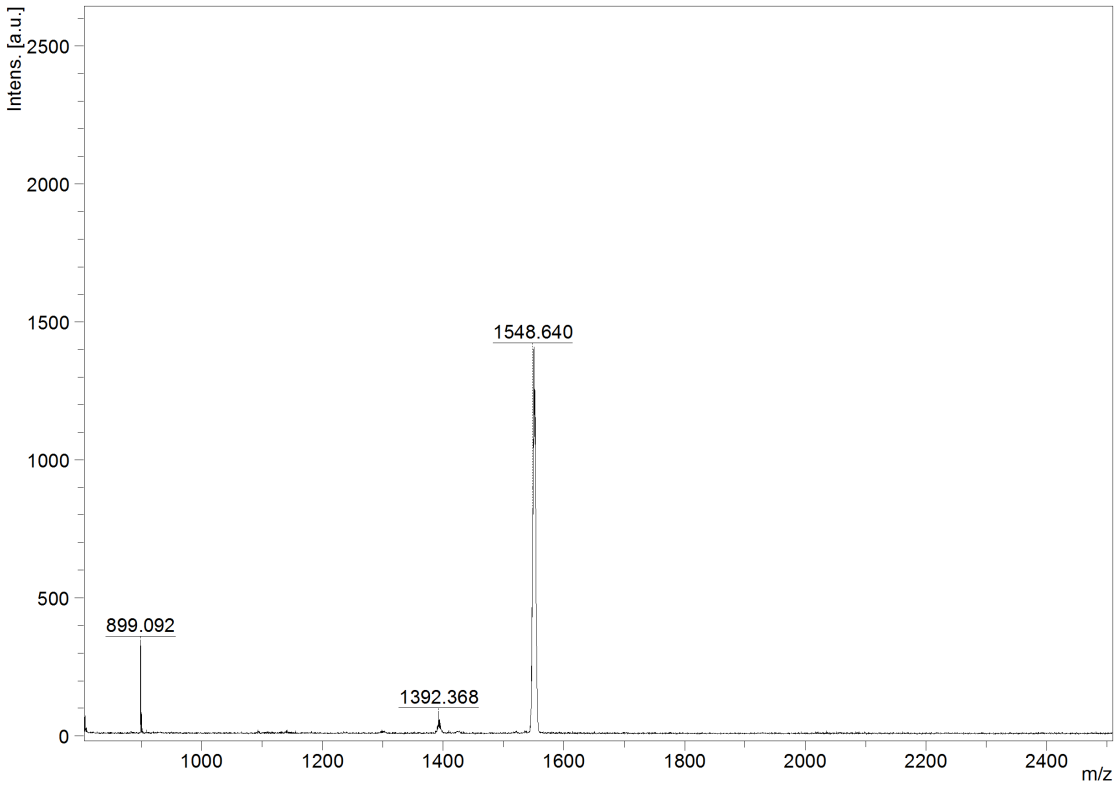
Das-GPEG-Ru **7**



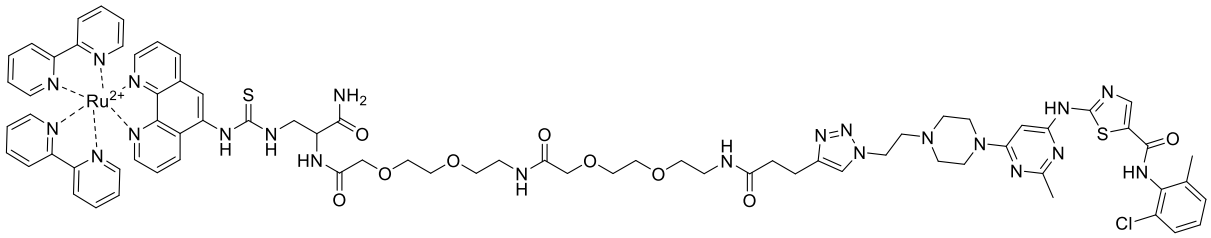
Chemical Formula: $C_{71}H_{75}ClN_{22}O_7RuS_2^{2+}$ Exact Mass: 1548.44 LC-MS (ESI): RT= 1.76 min m/z found = 325.67 [Ruthenium]2+, 387.92 $[M+2H]^{4+}$, 516.67 $[M+H]^{3+}$, 774.25 $[M]^{2+}$, 898.33 [M+H-Ruthenium]$^+$, 1547.00 $[M-H]^+$ MALDI-TOF found *m/z* = 899.092 [M+H-Ruthenium]$^+$, 1392.368 [M-H-bpy]$^+$, 1548.640 $[M-H]^+$



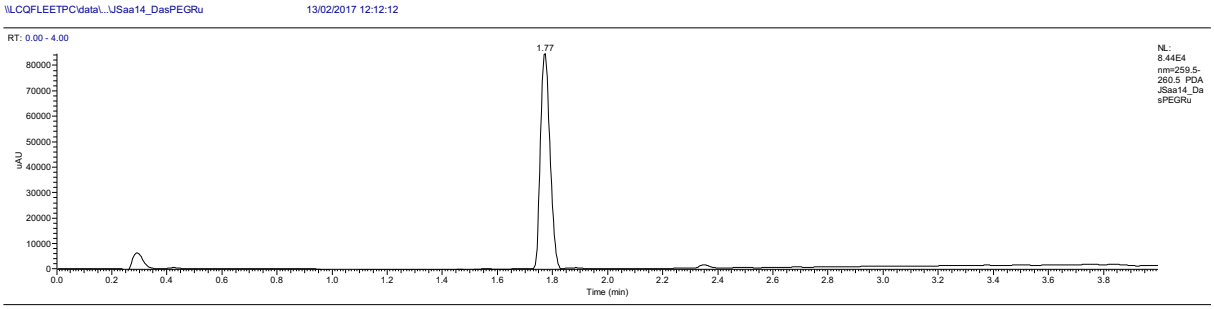


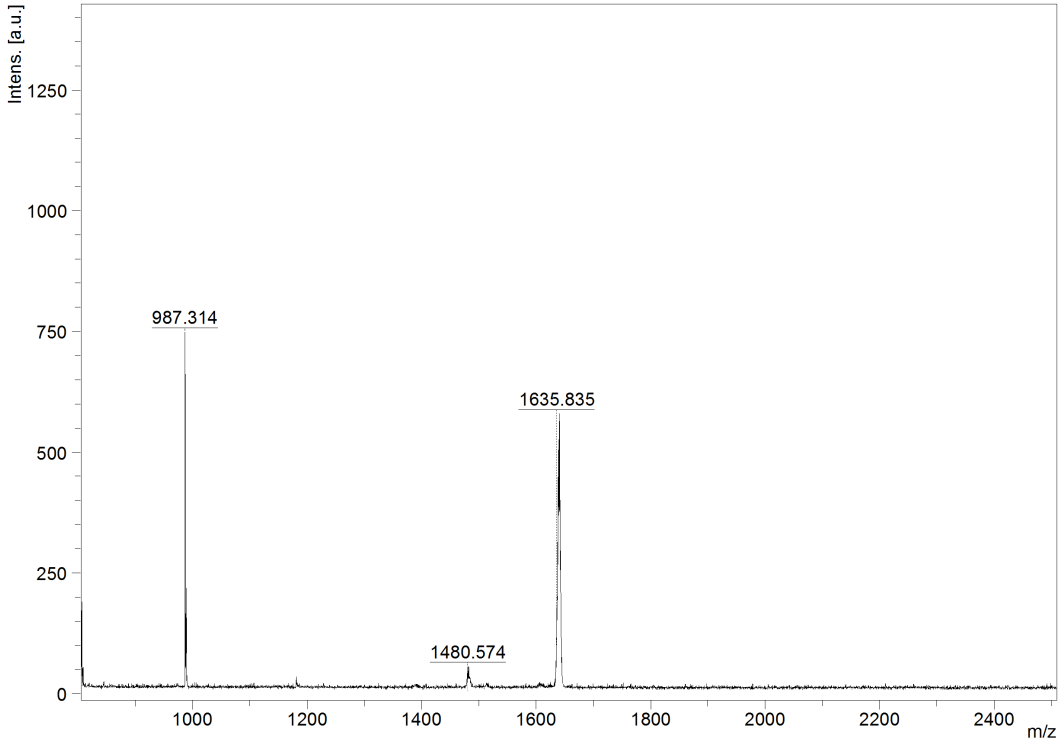


Das-PEG2-Ru **8**

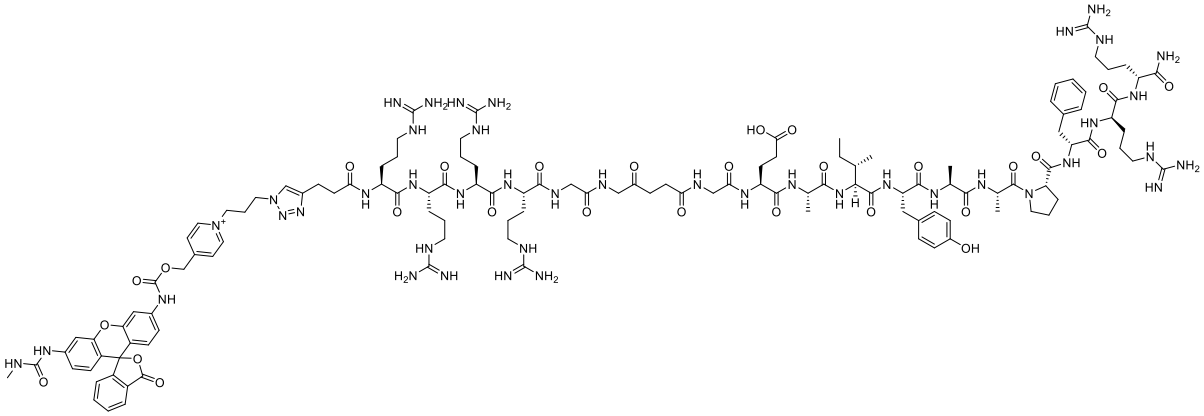


Chemical Formula: $C_{75}H_{83}ClN_{22}O_9RuS_2^{2+}$ Exact Mass: 1636.49 LC-MS (ESI) RT = 1.77 min m/z found = 325.58 $[Ruthenium]^{2+}$, 409.42 $[M+2H]^{4+}$, 545.92 $[M+H]^{3+}$, 818.17 $[M]^{2+}$, 986.42 $[M+H\text{-}Ruthenium]^{+}$, 1635.08 $[M-H]^{+}$ MALDI-TOF found *m/z* = 987.314 $[M+H\text{-}Ruthenium]^{+}$, 1480.574 $[M\text{-}H\text{-}bpy]^{+}$, 1635.835 $[M-H]^{+}$

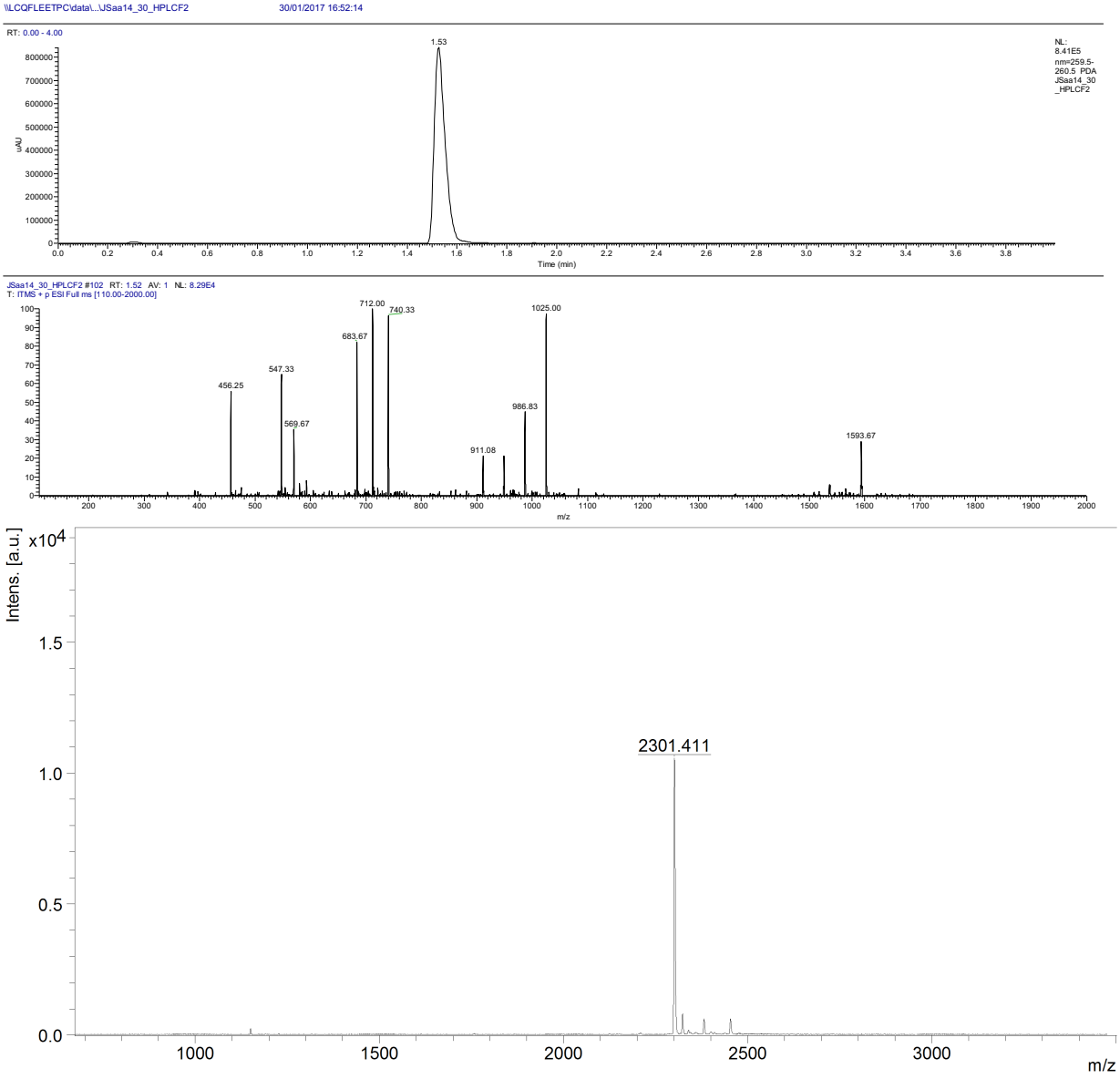




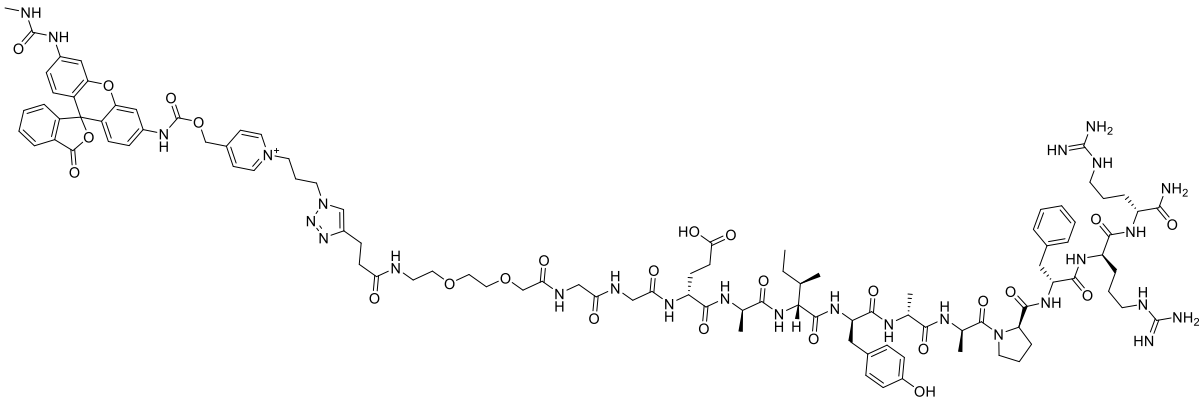
PAbl-R4G2 **9**



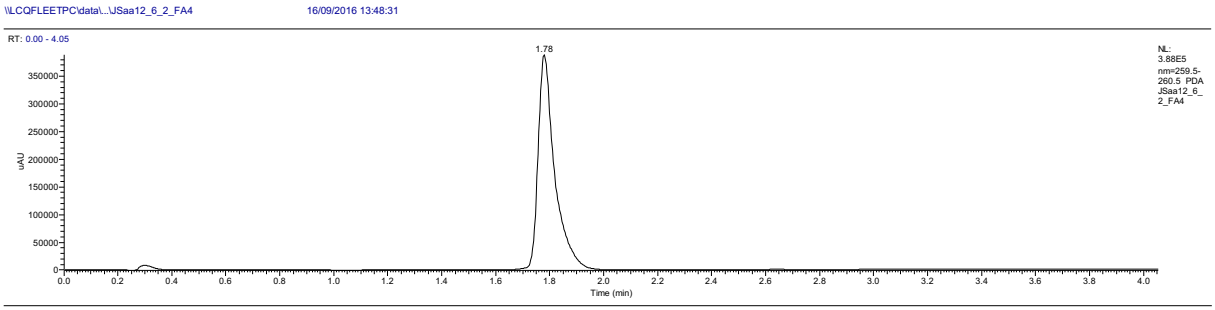
Chemical Formula: $C_{125}H_{178}N_{43}O_{28}^{+}$, Exact Mass: 2729,38 LC-MS (ESI): RT = 1.53 min *m/z* found: 456.25 $[M+5H]^{6+}$, 547.33 $[M+4H]^{5+}$, 569.67 $[M+4H+TFA]^{5+}$, 683.67 $[M+3H]^{4+}$, 712.00 $[M+3H+TFA]^{4+}$, 740,33 $[M+3H+2TFA]^{4+}$, 911.06 $[M+2H]^{3+}$, 986.23 $[M+2H+2TFA]^{3+}$, 1025.00 $[M+2H+3TFA]^{3+}$ 1593.67 $[M+H+4TFA]^{2+}$, MALDI-TOF *m/z* found = 2301.411 $[M+H\text{-Rhodamine}]^{+}$

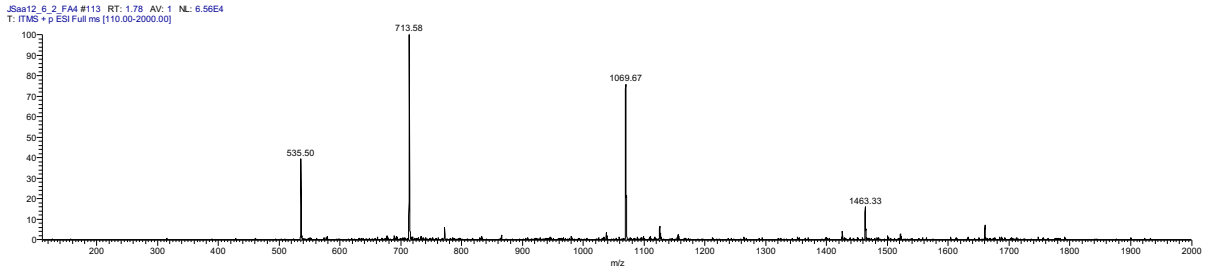


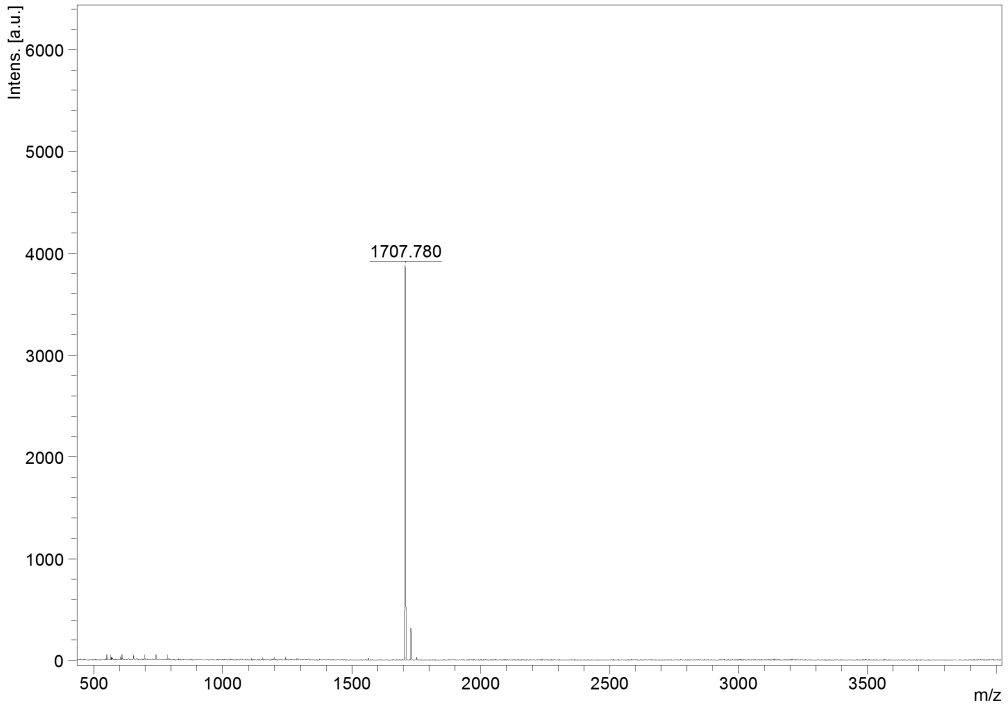
PAbl-PEG **10**



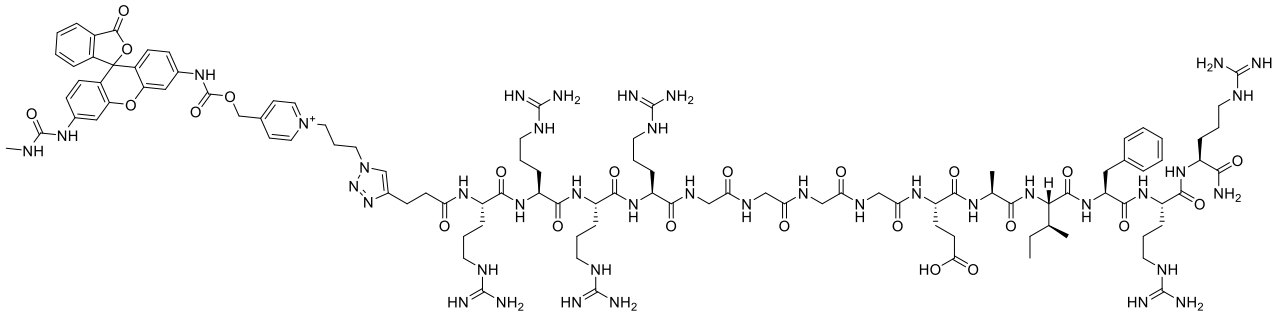
Chemical Formula: $C_{102}H_{134}N_{27}O_{25}^{+}$ Exact Mass: 2137,00, LC-MS (ESI) RT = 1.78 min m/z found 535.50 $[M+4H]^{4+}$ 713.58 $[M+3H]^{3+}$, 1069.67 $[M+2H]^{2+}$, 1463.33 [MALDI-TOF found 1707.780 $[M+H\text{-rhodamine}]^{+}$



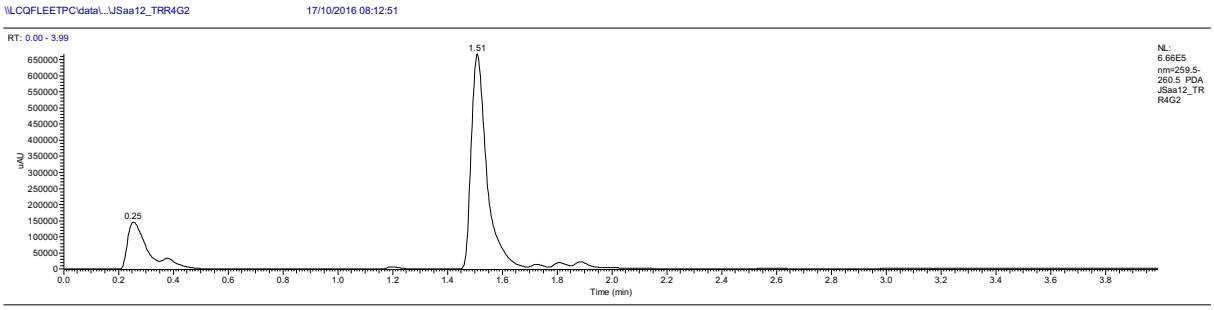


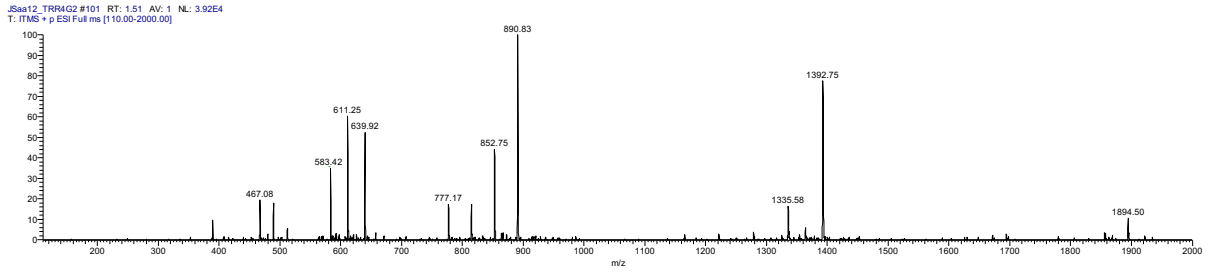


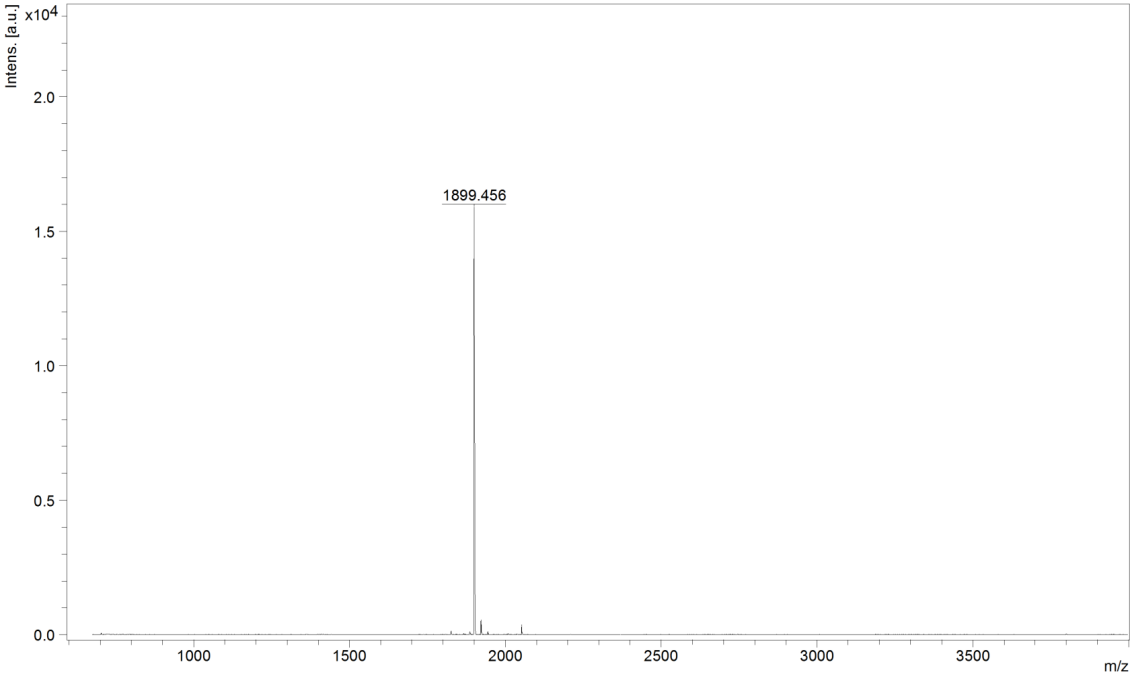
TR-PAbl-R4G2 **11**



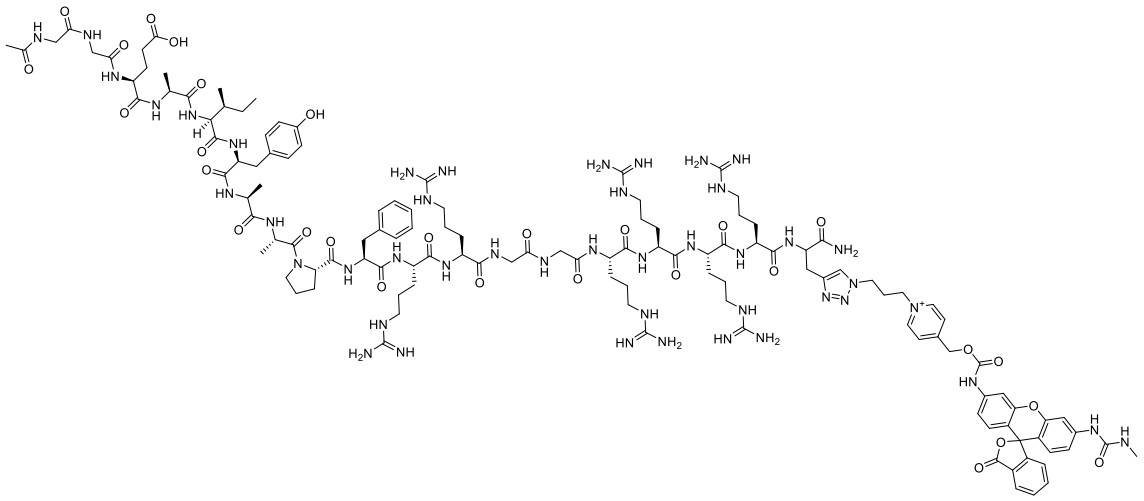
Chemical Formula: $C_{104}H_{151}N_{40}O_{23}^{+}$ Exact Mass: 2328,19 LC-MS (ESI): RT= 1,51 min m/z found = 386,47 $[M+6H]^{6+}$, 467.08 $[M+5H]^{5+}$, 583.42 $[M+4H]^{4+}$, 611.25 $[M+4H+TFA]^{4+}$, 639.92 $[M+4H+2TFA]^{4+}$, 777.17 $[M+3H]^{3+}$, 852,75 $[M+3H+2TFA]^{3+}$, 890.83 $[M+3H+3TFA]^{3+}$, 1335.58 $[M+3H+3TFA]^{3+}$, 1392.75 $[M+2H+4TFA]^{2+}$, MALDI-TOF m/z found = 1899.456 $[M+H\text{-Rhodamine}]^{+}$



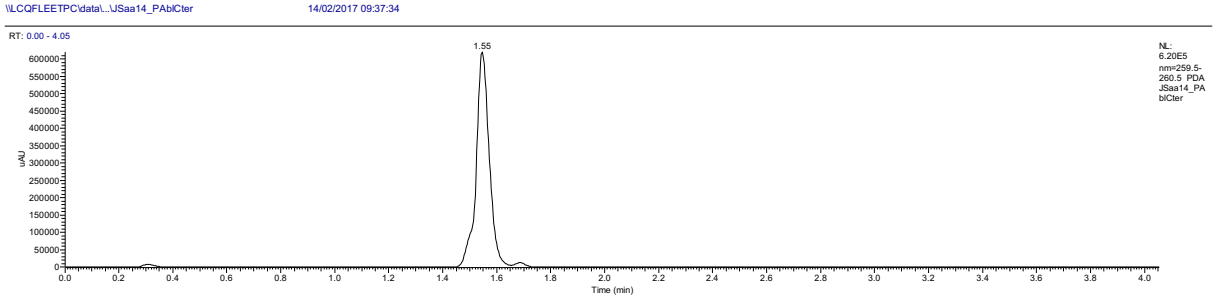


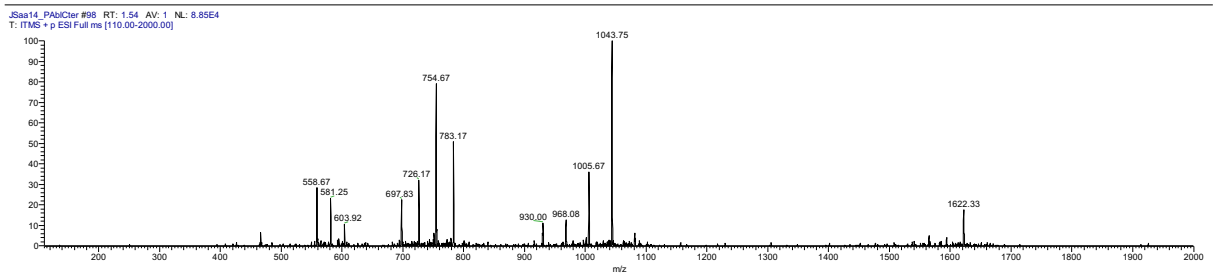


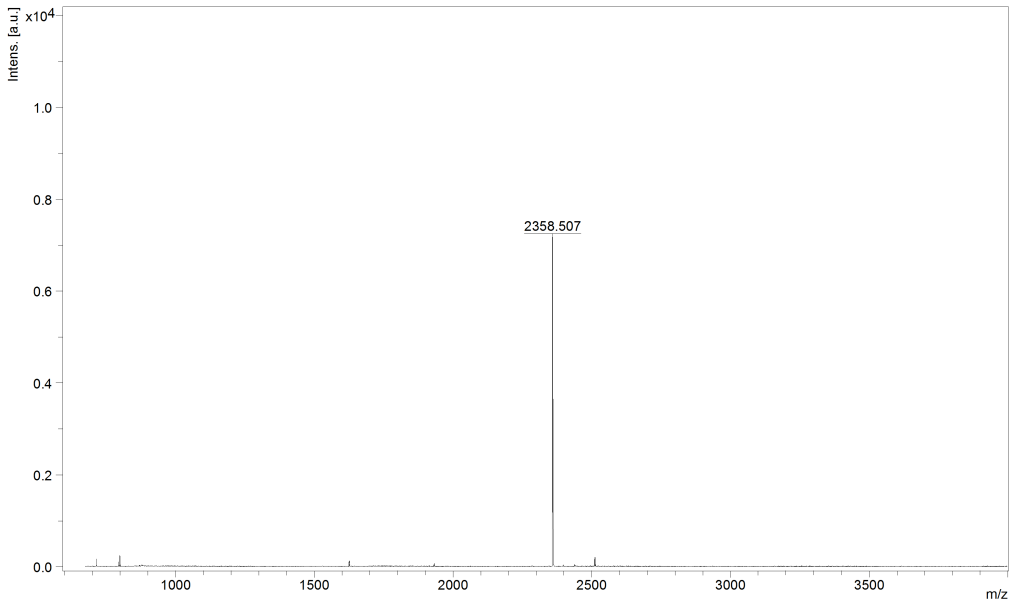
C-term-PAbl-R4G2 **12**



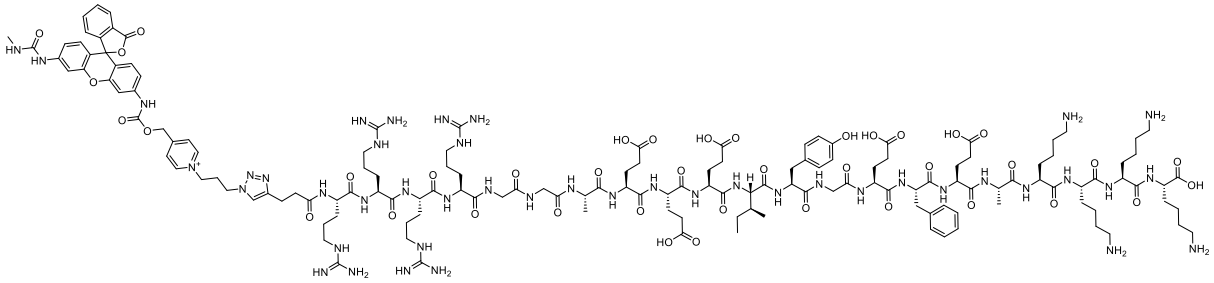
Chemical Formula: $C_{126}H_{180}N_{45}O_{29}^{+}$ Exact Mass: 2787.40, LC-MS (ESI) RT = 568.67 $[M+4H]^{5+}$, 581.25 $[M+4H+TFA]^{5+}$, 603.92 $[M+4H+2TFA]^{+}$, 697.83 $[M+3H]^{4+}$, 726.17 $[M+3H+TFA]^{4+}$, 754.67 $[M+3H+2TFA]^{4+}$, 783.17 $[M+3H+TFA]^{4+}$, 930.00 $[M+2H]^{3+}$, 968.08 $[M+2H+TFA]^{3+}$, 1005.67 $[M+2H+2TFA]^{3+}$, 1043.75 $[M+2H+3TFA]^{3+}$, 1622.33 $[M+4TFA]^{2+}$ MALDI-TOF m/z found 2358.507 $[M+H-Rhodamine]^{+}$



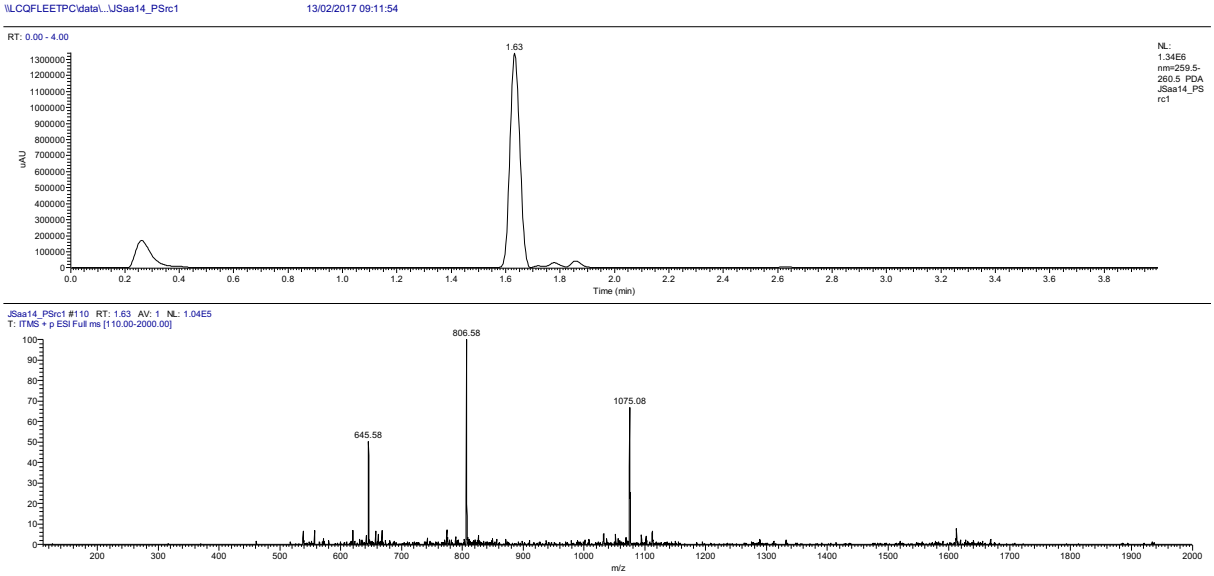


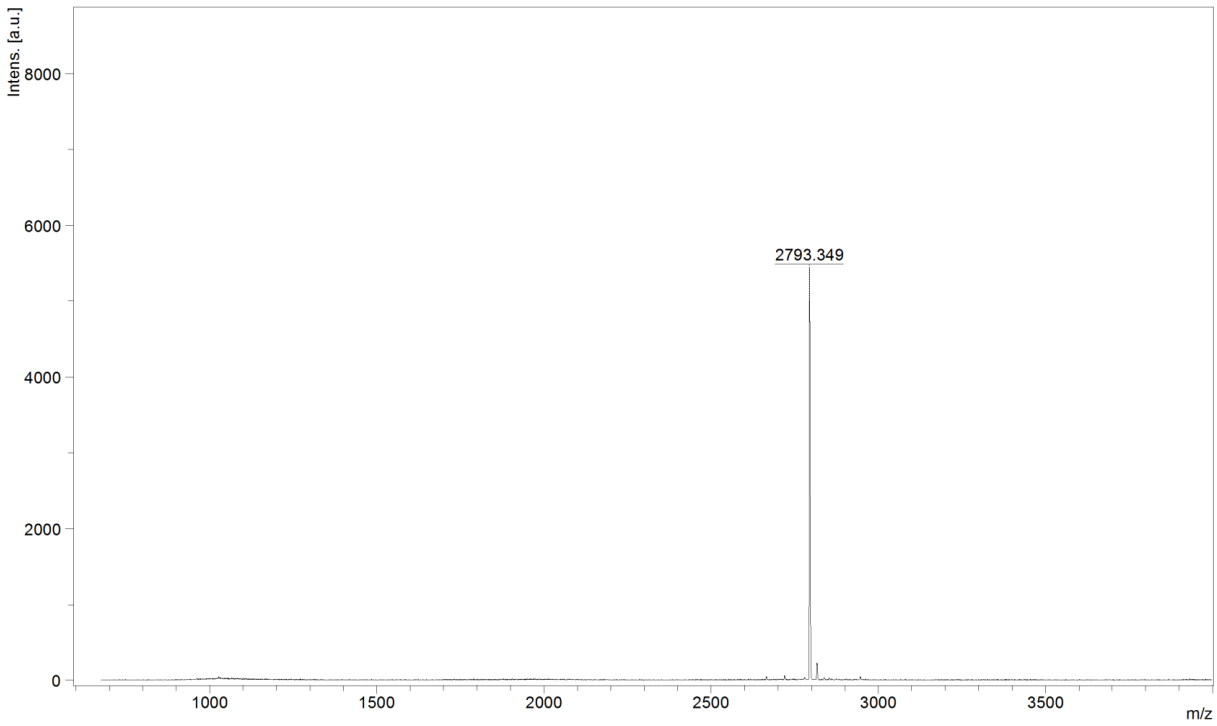


PSrc-R4G2 **13**



Chemical Formula: $C_{146}H_{213}N_{44}O_{40}^{+}$ Exact Mass: 3222.60, LC-MS (ESI) RT = 1.63 min m/z found = 645.58 $[M+5H]^{5+}$, 806.58 $[M+4H]^{4+}$, 1075.08 $[M+3H]^{3+}$, MALDI-TOF found: 2793.349 $[M+H–Rhodamine]^{+}$





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