Supporting Information

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1. General Information

Unless otherwise noted, all reactions were carried out under a nitrogen atmosphere; materials obtained from commercial suppliers were used directly without further purification. The [α]_D was recorded using PolAAr 3005 High Accuracy Polarimeter. ¹H NMR spectra, ¹³C NMR spectra, ³¹P NMR spectra and ¹⁹F NMR spectra were recorded on a Bruker 400 (or 500) MHz spectrometer in chloroform-d₃. Chemical shifts (in ppm) were referenced to tetramethylsilane ($\delta = 0$ ppm) in CDCl₃ as an internal standard. ¹³C NMR spectra were obtained by using the same NMR spectrometers and were calibrated with CDCl₃ ($\delta = 77.00$ ppm). The data is being reported as (s = singlet, d = doublet, dd = doublet of doublet, t = triplet, m = multiplet or unresolved, br = broad signal, coupling constant(s) in Hz, integration). Noteworthy, splitting signals between ¹³C nucleus and ¹³P nucleus in some chiral phosphine catalysts were difficult to distinguish and these ¹³C NMR signals were reported as singlet entirely.

Trichloromethane (CHCl₃), dichloromethane, dichloroethane and ethyl acetate were freshly distilled from CaH₂; tetrahydrofuran (THF), toluene and ether were dried with sodium benzophenone and distilled before use.

Reactions were monitored by thin layer chromatography (TLC) using silicycle pre-coated silica gel plates. Flash column chromatography was performed on silica gel 60 (particle size 200-400 mesh ASTM, purchased from Yantai, China) and eluted with petroleum ether/ethyl acetate.

2. Optimization of Reaction Conditions for the Enantioselective [3+2]

Cycloaddition of 1a and 2a^[a]

0, Dh				EtO ₂ C CF ₃			
Ph		Pn	Cat.	* (10 mol%)) O		
	013	CO ₂ Et	SO	vent, T, t	\sim		
	1a	2a			Ph	Ph	
Ια		24			3aa		
		Et Et	OMe	Ph T	Bu	<i>t</i> Bu	
			_ ↓ P_~		NH	o ^{⊳Ś} `№H	
Me	× _{Me} ⟨_⟩	Et Et	Ēh	MeO MeO	PPh ₂ PPh ₂		
(S,	S)-DIOP (R,R)-Et-DUP	HOS (R,R)-Et-BPE			ر م	OTIPS	
			(11,11)		з,қ _з ,-г і	(S,R _S)- P2	
F.C. C.F. F ₃ C CF ₃ F ₃ C CF ₃							
#	Bu <u>t</u> Bu	F ₃ C CF ₃	T T				
S	O ^{≠Š} ∖NH		, j	O NH	1	o [⊥] NH	
0/1	PPh ₂	O [™] NH	O'N	H PPho A	PPh ₂	PPh ₂	
		PPh ₂			tBu	Ph	
(5)	PPn ₂		× \		$\boldsymbol{r}_{_{\mathrm{tBu}}}$	Ph	
(0,,	(S,R _S)- P4	(S)- P1	(S)-F	2 (S)-I	•3	(S) -P4	
		T (0 C)	(1)	<u>C 1 (</u>	X7' 11	(0 /)[c]	
Entry	Cat.	$I(^{\circ}\mathbb{C})$	<i>t</i> (n)	Solvent	Y 1eld	<i>ee</i> (%) ^[c]	
1	(<i>S</i> , <i>S</i>)- DIOP	25	12	toluene	<10		
2	(R,R)-Et-DUPHOS	25	12	toluene	<10		
3	(<i>R</i> , <i>R</i>)- Et-BPE	25	8	toluene	64	39	
4	(R,R)- DIPAMP	25	6	toluene	81	89	
5	(S,R_S) - P1	25	12	toluene	51	5	
6	(S,R_S) -P2	25	12	toluene	54	11	
7	(S,R_S) -P3	25	12	toluene	62	30	
8	(S,R_S) -P4	25 25	12	toluene	58 70	29	
9	(S) - P1	25	3	toluene	79 85	5 14	
11	(S)- P3	25	3	toluene	84	40	
12	(S)- P4	25	3	toluene	86	21	
13	(R,R)- DIPAMP	25	6	DCM	51	72	
14	(R,R)- DIPAMP	25	6	CHCl ₃	68	84	
15	(<i>R</i> , <i>R</i>)- DIPAMP	25	6	EA	79	86	
16	(<i>R</i> , <i>R</i>)- DIPAMP	25	6	acetone	62	26	
17	(R,R)- DIPAMP	25	6	Et ₂ O	74	87	
18	(K,K)-DIPAMP (P,P) DIDAMD	25	0		13	88	
19 20	(Λ,Λ) -DIFAMIF (R R)-DIPAMP	25 25	6	THE CH2CN	00 ⊿0	00 30	
20	(R,R)- DIPAMP	0	6	toluene	47 84	90	
22	(R,R)- DIPAMP	-10	8	toluene	81	91	
23	(R,R)- DIPAMP	-20	12	toluene	78	92	
24	(R,R)- DIPAMP	-25	20	toluene	63	92	
$\begin{bmatrix} -1 \end{bmatrix} \begin{bmatrix} -1 \end{bmatrix} $							

[a] Unless otherwise specified, all reactions were carried out with (*E*)-1a (0.1 mmol), recemic 2a (0.15 mmol) in solvent (1 mL). [b] Yield of isolated products; both diastereoselectivity and regioselectivity were more than 20:1. [c] Determined by HPLC analysis using a chiral stationary phase.

3. Typical Synthetic Procedure and Data for Novel Chiral Phosphines Catalyst

3.1 Typical Procedure for the Synthesis of Phosphines (S)-P1~4



Step 1: BH₃•THF (3.0 mmol) was added slowly to the solution of (S,R_S) -**P** (2.0 mmol)^[1] in dry THF (5 mL) at – 30 °C and the reaction mixture was stirred for 2 h until completion of the material as indicated by TLC followed by adding 10 mL of water and 20 mL EtOAc. The aqueous phase was separated and extracted three times with 20 mL EtOAc. The combined organic phases were dried over MgSO₄ and the solvents were removed in vacuo.

Step 2: 4 M HCl (1 mL) was added slowly to the above residue which disolved in MeOH (10 mL) and the reaction mixture was stirred at room temperature for 3 h until completion of material as indicated by TLC analysis, followed by washing with aq NaHCO₃ and 10 mL aq brine water. The organic layers was separated and extracted three times with 20 mL EtOAc. The combined organic phases were dried over MgSO₄ and the solvents were removed in vacuo.

Step 3: Et₂NH (5.0 mL) was added to the above residue and the mixture was stirred at 55 $^{\circ}$ C for 6 h under the protection of N₂ until completion of material as indicated by TLC analysis. The solvent was then removed in vacuo and the residue was used directly for the next step.

Step 4: Under the protection of Ar, 3,5-bis(trifluoromethyl)benzoyl chloride (1.1 eq.) was added slowly to the above residue which was disolved in dry DCM (0.1 M) at 0 °C. This reaction mixture was then stirred at 25 °C for another 1 h, after completion of the reaction, the solvent was then removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the desired (*S*)-P1~4.

3.2 Typical Procedure for the Synthesis of Phosphines (S)-P5 and (S)-P6



Step 1: BH₃•THF (3.0 mmol) was added slowly to the solution of (S,R_S) -**P3** (2.0 mmol)^[1] in dry THF (5 mL) at -30 °C and the reaction mixture was stirred for 2 h until completion of the material as indicated by TLC followed by adding 10 mL of water and 20 mL of EtOAc. The aqueous phase was separated and extracted three times with 20 mL EtOAc. The combined organic phases were dried over MgSO₄ and the solvents were removed in vacuo.

Step 2: 4 M HCl (1 mL) was added slowly to the above residue which disolved in MeOH (10 mL) and the reaction mixture was stirred at room temperature for 3 h until completion of material as indicated by TLC analysis, followed by washing with aq NaHCO₃ and 10 mL of aq brine water. The organic layers was separated and extracted three times with 20 mL of EtOAc. The combined organic phases were dried over MgSO₄ and the solvents were removed in vacuo.

Step 3: Et₂NH (5.0 mL) was added to the above residue and the mixture was stirred at 55 °C for 6 h under the protection of N_2 until completion of material as indicated by TLC analysis. The solvent was then removed in vacuo and the residue was used directly for the next step.

Step 4: Under the protection of Ar, 3,5-Bis(trifluoromethyl)phenylisocyanate or 3,5-Bis(trifluoromethyl)phenyl isothiocyanate (1.2 eq.) was added slowly to the above residue which was disolved in dry DCM (0.1 M) at 0 °C. This reaction mixture was then stirred at 25 °C for another 1 h, after completion of the reaction, the solvent was then removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the desired (*S*)-**P5** and (*S*)-**P6**.

3.3 Typical Procedure for the Synthesis of Phosphines (S)-P7



Under the protection of Ar, 3,5-bis(trifluoromethyl)benzoyl chloride (2 eq.) was added slowly to the solution of (*S*)-**P3** (0.2 mmol) in dry DCM (2 mL) at 0 °C and the reaction mixture was stirred for 1 h at this temperature. This reaction mixture was then stirred at 25 °C for another 4 h, until completion of the material as indicated by TLC followed by adding 5 mL of water and 10 mL of EtOAc. The aqueous phase was separated and extracted three times with 15 mL of EtOAc. The combined organic phases were dried over MgSO₄ and the solvents were removed in vacuo. The residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the desired (*S*)-**P7** (36% yield).

3.4 Typical Procedure for the Synthesis of Phosphines (R,R)-SDIPAMP



Under the protection of Ar, (R,R)-DIPAMP (0.2 mmol) was added to the solution of

sulfur (0.2 mmol) in dry THF (2 mL) at 25 °C and the reaction mixture was stirred for 0.5 h at this temperature. After completion of the material as indicated by TLC, the solvent was then removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the desired (R,R)-**SDIPAMP** (51% yield).

General Data for (S)-P1~7 and (R,R)-SDIPAMP



(*S*)-**P1**; white solid; $[\alpha]_D^{20} = +15.0$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.01 (s, 2H), 7.97 (s, 1H), 7.55–7.51 (m, 2H), 7.45–7.42 (m, 2H), 7.35–7.34 (m, 9H), 7.30–7.26 (m, 1H), 6.73 (d, J = 7.5 Hz, 1H), 5.42–5.36 (m, 1H), 2.90–2.85 (m, 1H), 2.73–2.69 (m, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 163.62, 141.94 (d, J = 6.50 Hz), 137.78 (d, J = 12.13 Hz), 136.16, 132.98 (d, J = 19.38 Hz), 132.63 (d, J = 19.0 Hz), 131.94 (q, J = 33.63 Hz), 130.49 (q, J = 5.63 Hz), 129.25, 128.98, 128.88, 128.81, 128.75, 128.72, 128.67, 127.90, 127.28 (d, J = 2.88 Hz), 126.40, 125.85, 124.98–124.93 (m), 122.87 (q, J = 271.25 Hz), 53.00 (d, J = 17.13 Hz), 35.91 (d, J =16.38 Hz); ³¹P NMR (121.5 MHz, CDCl₃) $\delta = -22.82$ ppm; HRMS (ESI) m/z calcd. for C₂₉H₂₂F₆NNaOP [M+Na] ⁺= 568.1258, found = 568.1235.



(*S*)-**P2**; white solid; $[\alpha]_D^{20} = -43.0$ (*c* = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.08 (s, 2H), 8.02 (s, 1H), 7.52–7.50 (m, 1H), 7.44–7.36 (m, 9H), 7.35–7.25 (m, 7H), 7.21–7.17 (m, 2H), 6.82 (d, *J* = 7.0 Hz, 1H), 5.62–5.57 (m, 1H), 2.69–2.63 (m, 1H), 2.57–2.53 (m, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 163.27, 140.89, 140.62, 139.90 (d, *J* = 6.38 Hz), 137.39 (d, *J* = 11.75, Hz), 136.85 (d, *J* = 12.13 Hz), 136.10, 132.62 (d, *J* = 19.38 Hz), 132.32 (d, J = 19.0 Hz), 131.88 (q, J = 33.5 Hz), 130.79, 129.24, 128.95, 128.75, 128.62 (d, J = 1.38 Hz), 128.56 (d, J = 1.50 Hz), 128.39, 127.95, 127.33 (d, J = 2.25 Hz), 127.23 (d, J = 2.63 Hz), 125.52, 124.88–124.83 (m), 122.83 (q, J = 271.25 Hz), 50.70 (d, J = 15.0 Hz), 36.21 (d, J = 17.13 Hz); ³¹P NMR (121.5 MHz, CDCl₃) δ = -23.13 ppm; HRMS (ESI) m/z calcd. for C₃₅H₂₆F₆NNaOP [M+Na] ⁺ = 644.1548, found = 644.1540.



(*S*)-**P3**; white solid; $[\alpha]_D^{20} = -34.7$ (*c* = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.96 (s, 3H), 7.46–7.16 (m, 15H), 7.09–7.06 (m, 2H), 6.45–6.46 (m, 1H), 5.65–5.58 (m, 1H), 2.54–2.44 (m, 2H), 1.30 (s, 18H); ¹³C NMR (100 MHz, CDCl₃): δ 163.29, 150.87, 142.00, 140.05, 139.95, 137.85 (d, *J* = 11.9 Hz), 136.99 (d, *J* = 11.90 Hz), 136.53, 132.75, 132.56, 132.36, 132.17, 131.82, 131.49, 130.97, 128.99, 128.71, 128.67, 128.64, 128.60, 128.57, 127.72, 127.33, 127.22, 125.21, 124.87, 124.23, 123.65, 121.52, 121.05, 50.86 (d, *J* = 15.60 Hz), 36.42 (d, *J* = 16.70 Hz), 34.91, 31.44; ³¹P NMR (121.5 MHz, CDCl₃) δ = -62.72 ppm; HRMS (ESI) m/z calcd. for C₄₄H₄₃F₆NOP [M+H] ⁺ = 734.2981, found = 734.2998.



(*S*)-**P4**; white solid; $[\alpha]_D^{20} = -41.6$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.96 (s, 2H), 7.88 (s, 1H), 7.82–7.81 (m, 1H), 7.61–7.59 (m, 6H), 7.48–7.46 (m, 1H), 7.41–7.27 (m, 11H), 7.17–7.03 (m, 8H), 6.72 (d, J = 6.8 Hz, 1H), 5.64–5.57 (m, 1H), 2.68–2.62 (m, 1H), 2.57–2.52 (m, 1H); ¹³C NMR (100 MHz, CDCl₃): δ 163.32, 141.74, 141.67, 140.81, 140.52, 139.99 (d, J = 6.40 Hz), 137.23 (d, J = 12.0 Hz), 136.89 (d, J = 12.30 Hz), 136.25, 132.58 (d, J = 19.40 Hz), 132.26 (d, J = 19.20 Hz), 131.88 (q, J = 33.7 Hz), 130.80, 128.99, 128.75, 128.61, 128.54, 128.48, 128.16, 127.50, 127.43, 127.19, 126.89, 125.52, 124.90, 124.82, 124.78, 122.81 (q, J = 271.4 Hz), 50.87 (d, J = 15.60 Hz), 36.60 (d, J = 17.10 Hz); ³¹P NMR (121.5 MHz, CDCl₃) $\delta = -22.96$ ppm; HRMS (ESI) m/z calcd. for C₄₇H₃₄F₆NNaOP [M+Na] ⁺= 796.2174, found = 796.2171.



(*S*)-**P5**; white solid; $[\alpha]_{D}^{20} = -16.6$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CD₃COCD₃): δ 8.80 (s, 1H), 8.17 (s, 2H), 7.66 (d, J = 8.0 Hz, 1H), 7.57–7.52 (m, 3H), 7.34–7.31 (m, 1H), 7.28–7.13 (m, 10H), 7.01–6.98 (m, 2H), 6.88 (d, J = 7.0 Hz, 1H), 5.51–5.46 (m, 1H), 3.74 (s, 1H), 2.48–2.43 (m, 1H), 2.37–2.31 (m, 1H), 1.36 (s, 18H); ¹³C NMR (125 MHz, CD₃COCD₃): δ 154.66, 151.28, 143.51, 142.99 (d, J = 4.63 Hz), 142.80, 141.04, 140.17 (d, J = 13.88 Hz), 138.28 (d, J = 13.75 Hz), 133.16 (d, J = 12.25 Hz), 133.01 (d, J = 12.63 Hz), 132.31 (q, J = 32.63 Hz), 131.12, 129.45, 129.39, 129.20, 129.15, 129.11, 129.10, 128.43, 127.63, 126.24, 124.95, 124.49 (q, J = 270.13 Hz), 121.56, 118.41 (d, J = 3.5 Hz), 114.69 (m), 49.73 (d, J = 16.25 Hz), 38.18 (d, J = 16.38 Hz), 35.52, 31.81; ³¹P NMR (202.5 MHz, CD₃COCD₃) $\delta = -24.58$ ppm; ¹⁹F NMR (376 MHz, CD₃COCD₃) $\delta = -63.56$ ppm; HRMS (ESI) m/z calcd. for C₄₃H₄₄F₆N₂OP [M+H] ⁺= 749.3090, found = 749.3082.



(S)-P6; white solid; $[\alpha]_D{}^{20} = -8.8 \ (c = 0.33, \text{CHCl}_3); {}^{1}\text{H NMR} \ (500 \text{ MHz}, \text{CD}_3\text{COCD}_3):$ $\delta 9.55 \ (\text{s}, 1\text{H}), 8.42 \ (\text{s}, 2\text{H}), 7.76 \ (\text{s}, 1\text{H}), 7.63-7.59 \ (\text{m}, 3\text{H}), 7.34-7.18 \ (\text{m}, 10\text{H}),$ $7.00-6.97 \ (\text{m}, 2\text{H}), 6.06 \ (\text{br}, 1\text{H}), 3.75 \ (\text{d}, J = 4.0 \ \text{Hz}, 1\text{H}), 2.64-2.60 \ (\text{m}, 1\text{H}),$ $2.45-2.40 \ (\text{m}, 1\text{H}), 1.39 \ (\text{s}, 18\text{H}); {}^{13}\text{C} \text{ NMR} \ (125 \ \text{MHz}, \text{CD}_3\text{COCD}_3): \delta 181.11,$ 151.29, 142.67 (d, J = 16.00 Hz), 141.92 (d, J = 5.38 Hz), 141.03, 139.96 (d, J = 13.38 Hz), 138.04 (d, J = 13.13 Hz), 133.29 (d, J = 19.25 Hz), 132.92 (d, J = 19.25 Hz), 132.51 (q, J = 32.88 Hz), 131.29, 129.41 (d, J = 6.5 Hz), 129.11 (d, J = 8.00 Hz), 129.01 (d, J = 7.00 Hz), 128.30, 127.61, 125.42, 124.58, 124.29 (q, J = 270.75 Hz), 121.68, 117.36-117.30 (m), 54.43 (d, J = 15.25 Hz), 37.19 (d, J = 17.00 Hz), 35.50, 31.88; ³¹P NMR (202.5 MHz, CD₃COCD₃) $\delta = -23.86$ ppm; ¹⁹F NMR (376 MHz, CD₃COCD₃) $\delta = -67.71$ ppm; HRMS (ESI) m/z calcd. for C₄₃H₄₄F₆N₂SP [M+H] ⁺ = 765.2862, found = 765.2865.



(*S*)-**P6**; white solid; $[\alpha]_{D}^{20} = -70.8$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CD₃COCD₃): δ 8.18–8.17 (m, 1H), 7.77–7.34 (m, 6H), 7.61–7.60 (m, 1H), 7.54–7.51 (m, 1H), 7.40–7.19 (m, 10H), 7.10–7.06 (m, 2H), 6.99–6.96 (m, 2H), 6.34–6.30 (m, 1H), 3.15–3.08 (m, 1H), 2.83–2.78 (m, 1H), 1.39–1.29 (m, 18H); ¹³C NMR (125 MHz, CD₃COCD₃): δ 170.67, 150.87, 142.38, 139.80, 139.65, 139.381 (d, J = 7.25 Hz), 138.14 (d, J = 10.38 Hz), 138.81 (d, J = 10.88 Hz), 132.85 (d, J = 20.05 Hz), 132.191 (q, J = 34.25 Hz), 131.64 (d, J = 17.75 Hz), 131.06, 129.35, 129.21 (d, J = 7.38 Hz), 128.61, 128.35, 128.34, 128.30, 128.21, 127.73, 126.68, 124.84, 123.70, 122.23 (q, J = 271.63 Hz), 58.33 (d, J = 13.50 Hz), 34.99, 32.49 (d, J = 17.13 Hz), 31.44; ³¹P NMR (202.5 MHz, CD₃COCD₃) $\delta = -21.96$ ppm; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -63.57$ ppm; HRMS (ESI) m/z calcd. for C₅₂H₄₅F₁₂NO₂P [M+H] ⁺ = 974.2991, found = 974.2993.



(*R*,*R*)-**SDIPAMP**; white solid; $[\alpha]_D^{20} = +39.9$ (*c* = 0.33, CHCl₃); ¹H NMR (500 MHz, CD₃COCD₃): δ 8.30–8.25 (m, 1H), 7.70–7.65 (m, 2H), 7.53–7.29 (m, 10H), 7.15–7.12 (m, 1H), 7.07–7.04 (m, 1H), 6.91–6.82 (m, 3H), 3.69 (s, 3H), 3.59 (s, 3H), 2.95–2.85 (m, 1H), 2.66–2.56 (m, 1H), 2.38–2.23 (m, 2H); ¹³C NMR (125 MHz, CD₃COCD₃): δ 161.02 (d, *J* = 12.75 Hz), 159.75 (d, *J* = 2.50 Hz), 136.717 (d, *J* = 12.88 Hz), 136.31 (d, *J* = 9.75 Hz), 134.28, 133.94 (q, *J* = 2.13 Hz), 133.62, 133.23, 133.08, 132.09 (d, *J* = 4.38 Hz), 130.76 (d, *J* = 2.75 Hz), 130.59, 130.51, 130.10, 128.34, 128.32 (d, *J* = 6.88 Hz), 128.05 (d, *J* = 12.25 Hz), 125.69 (d, *J* = 15.75 Hz), 121.05 (d, *J* = 12.25 Hz), 120.80, 119.08, 118.47, 110.72 (d, *J* = 7.25 Hz), 110.24, 55.21 (d, *J* = 34.75 Hz), 27.61 (dd, *J* = 55, 19.63 Hz), 18.31 (dd, *J* = 14.25, 3.38, Hz); ³¹P NMR (202.5 MHz, CD₃COCD₃) δ = 45.17, -21.96 ppm; HRMS (ESI) m/z calcd. for C₂₈H₂₉O₂P₂S [M+H] ⁺ = 491.1358, found = 491.1370.

4. Typical Procedure for the Enantioselective [3+2] Cycloaddition of

Allenes with β -Perfluoroalkyl α , β -Enones

Enantioselective [3+2] cycloadditions of γ -aryl substituted allenoates with enone

$$R^{1} \xrightarrow{\text{O}}_{\text{CO}_{2}\text{Et}} \xrightarrow{(R,R)-\text{DIPAMP (10 mol\%)}}_{\text{toluene, - 20 °C}} \xrightarrow{\text{EtO}_{2}\text{C}}_{\text{Ar}} \xrightarrow{\text{O}}_{\text{R}^{1}}$$

Under Ar, a stirred solution of $1^{[2]}$ (0.2 mmol) and recemic 2 (0.3 mmol) in toluene (2 mL) was cooled to -20 °C. Subsequently, (*R*,*R*)-**DIPAMP** (0.02 mmol) was added in one portion. The reaction mixture was stirred at -20 °C until completion of the material as indicated by TLC. Then the solvents were removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the desired cycloaddition product **3**.

Enantioselective [3+2] cycloadditions of γ -alkyl substituted allenoates with enone



Under Ar, a stirred solution of 1 (0.2 mmol) and recemic 2 (0.44 mmol) in CHCl₃(2

mL) was cooled to -20 °C. Subsequently, (S)-P3 (0.02 mmol) was added in one portion. The reaction mixture was stirred at -20 °C until completion of the material as indicated by TLC. Then the solvents were removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the desired cycloaddition product **3**.

5. The Structural Assignment for the Regioisomer

In order to confirm the structure of the minor product which was observed in the (S)-P3 catalyzed enantioselective [3+2] cycloadditions of γ -alkyl substituted allenoates with β -perfluoro substituted enone, we have isolated the isomer of 3cl and confirmed its structurue by NMR analysis. According to these NMR spectra, the minor product 3cl' was assigned to the regioisomer of 3cl.















6. Procedure for the "Deracemization" and Kinetic Resolution of Recemic Allenoates

Procedure for the "Deracemization" of Racemic 2a



Under Ar, a stirred solution of 1d (0.2 mmol) and racemic 2a (0.4 mmol) in toluene (2 mL) was cooled to -20 °C. Subsequently, (R,R)-DIPAMP (0.02 mmol) was added in one portion. The reaction mixture was stirred at -20 °C for 12 h. Then the solvents were removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the 3da (76%, 90% *ee*) and 2a (38%, 0% *ee*).

Procedure for the Kinetic Resolution of Racemic 2g



Under Ar, a stirred solution of 1d (0.2 mmol) and racemic 2g (0.4 mmol) in CHCl₃ (2 mL) was cooled to -20 °C. Subsequently, (*S*)-P3 (0.02 mmol) was added in one portion. The reaction mixture was stirred at -20 °C for 0.5 h. Then the solvents were removed in vacuo and the residue was directly purified by silica gel chromatography using petroleum ether/EtOAc as the eluent to afford the 3dg (61%, 94% *ee*) and 2g (32%, 81% *ee*).

7. X-ray Crystal Structure for 3aa



8. General Data and HPLC Spectra for Cycloaddition Product 3



3aa; white solid; $[\alpha]_D^{20} = + 67.2$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.70–7.68 (m, 2H), 7.57–7.54 (m, 1H), 7.38–7.30 (m, 5H), 7.09–7.07 (m, 2H), 6.82 (s, 1H), 4.61–4.53 (m, 1H), 4.37–4.21 (m, 3H), 4.00–3.99 (m, 1H), 1.33 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 198.22, 163.27, 147.41, 140.19, 135.26, 133.82, 131.95, 129.13, 128.97, 128.62, 128.03, 127.86, 127.68, 126.29 (q, J = 277.90 Hz), 61.06, 55.47, 54.10, 51.35 (q, J = 28.90 Hz), 14.08; ¹⁹F NMR (376 MHz, CDCl₃) $\delta =$ -67.48 ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak IC, hexane/*i*-PrOH = 95/05; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 15.34 min, second peak: t_R = 17.34 min; HRMS (ESI) m/z calcd. for C₂₂H₁₉F₃NaO₃ [M+Na] ⁺ = 411.1179, found = 411.1182.





3ba; colorless oil; $[\alpha]_D{}^{20} = +122.4$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.72–7.68 (m, 2H), 7.36–7.32 (m, 3H), 7.10–7.00 (m, 4H), 6.81 (s, 1H), 4.61–4.53 (m, 1H), 4.37–4.18 (m, 3H), 3.98–3.96 (m, 1H), 1.33 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 196.61, 166.22 (d, J = 255.00 Hz), 163.22, 147.26, 140.10, 131.85 (d, J = 9.50 Hz), 131.67 (d, J = 2.80 Hz), 129.08, 127.99, 126.25 (d, J = 278.00 Hz), 115.81 (d, J = 21.9 Hz), 61.10, 55.55, 54.22, 51.46 (q, J = 29.00 Hz), 14.07; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.52, -103.61 ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak IC, hexane/*i*-PrOH = 95/05; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 14.49 min, second peak: t_R = 16.46 min; HRMS (ESI) m/z calcd. for C₂₂H₁₈F₄NaO₃ [M+Na] ⁺ = 429.1084, found = 429.1093.





3ca; colorless oil; $[\alpha]_D^{20} = + 80.6$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.64–7.62 (m, 2H), 7.38–7.33 (m, 5H), 7.12–7.10 (m, 2H), 6.83 (s, 1H), 4.62–4.56 (m, 1H), 4.38–4.24 (m, 2H), 4.21–4.19 (m, 1H), 4.00–3.98 (m, 1H), 1.35 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 196.93, 163.16, 147.24, 140.49, 139.98, 133.48, 131.85 (d, J = 1.75 Hz), 130.49, 129.08, 128.94, 128.01, 127.96, 126.19 (q, J =278.13 Hz), 61.09, 55.45, 54.21, 51.36 (q, J = 29.00 Hz), 14.06; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.49$ ppm; Enantiomeric excess: 90%, determined by HPLC (Chiralpak AS-H, hexane/*i*-PrOH = 95/05; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 10.82 min, second peak: t_R = 12.86 min; HRMS (ESI) m/z calcd. for C₂₂H₁₈ClF₃NaO₃ [M+Na] ⁺ = 445.0789, found = 445.0794.





3da; colorless oil; $[\alpha]_D^{20} = + 92.8$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.54–7.48 (m, 4H), 7.36–7.32 (m, 3H), 7.10–7.08 (m, 2H), 6.80 (s, 1H), 4.60–4.52 (m, 1H), 4.37–4.21 (m, 2H), 4.18–4.16 (m, 1H), 3.98–3.95 (m, 1H), 1.33 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.19, 163.17, 147.22, 140.00, 133.95, 131.96, 131.90, 130.56, 129.31, 129.11, 128.04, 127.97, 126.20 (q, J = 278.20 Hz), 61.11, 55.46, 54.23, 51.43 (q, J = 28.90 Hz), 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.49$ ppm; Enantiomeric excess: 90%, determined by HPLC (Chiralpak AS-H, hexane/*i*-PrOH = 95/05; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 11.01 min, second peak: t_R = 13.26 min; HRMS (ESI) m/z calcd. for C₂₂H₁₈BrF₃NaO₃ [M+Na] ⁺ = 489.0284, found = 489.0287.





3ea; colorless oil; $[\alpha]_D^{20} = + 68.3$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.21–8.19 (m, 2H), 7.83–7.81 (m, 2H), 7.37–7.36 (m, 3H), 7.11–7.09 (m, 2H), 6.82 (s, 1H), 4.64–4.57 (m, 1H), 4.39–4.25 (m, 3H), 4.01–3.99 (m, 1H), 1.36 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 196.77, 163.02, 150.67, 146.94, 139.71, 139.65, 131.81, 130.09, 129.28, 128.32, 127.87, 126.06 (q, J = 280.25 Hz), 123.75, 61.22, 55.29, 55.04, 51.38 (q, J = 29.13 Hz), 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.49$ ppm; Enantiomeric excess: 90%, determined by HPLC (Chiralpak AS-H, hexane/*i*-PrOH = 95/05; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 19.11 min, second peak: t_R = 22.64 min; HRMS (ESI) m/z calcd. for C₂₂H₁₈F₃NNaO₅ [M+Na] ⁺= 456.1029, found = 456.1042.





3fa; colorless oil; $[\alpha]_D^{20} = +82.8$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.75 (d, J = 8.00 Hz, 2H), 7.66 (d, J = 8.50 Hz, 2H), 7.36–7.35 (m, 3H), 7.09–7.08 (m, 2H), 6.81 (s, 1H), 4.62–4.56 (m, 1H), 4.38–4.21 (m, 3H), 3.98–3.97 (m, 1H), 1.35 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 196.96, 163.05, 146.98, 139.69, 138.23, 132.40, 131.82, 129.42, 129.23, 128.26, 127.88, 126.07 (q, J = 278.13 Hz), 117.60, 117.07, 61.21, 55.30, 54.75 (d, J = 1.25 Hz), 51.32 (q, J = 29.38 Hz), 14.06; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.51 ppm; Enantiomeric excess: 88%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 10.58 min, second peak: t_R = 12.53 min; HRMS (ESI) m/z calcd. for C₂₃H₁₈F₃NNaO₃ [M+Na] ⁺ = 436.1131, found = 436.1131.





3ga; colorless oil; $[\alpha]_D^{20} = + 76.5$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.59 (d, J = 8.40 Hz, 2H), 7.35–7.30 (m, 3H), 7.15 (d, J = 8.00 Hz, 2H), 7.11–7.09 (m, 2H), 6.82 (s, 1H), 4.59–4.51 (m, 1H), 4.37–4.20 (m, 3H), 3.99–3.98 (m, 1H), 2.38 (s, 3H), 1.33 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.78, 163.30, 147.52, 144.88, 140.27, 132.66, 131.91, 129.31, 129.27, 128.93, 128.04, 127.79, 126.31 (q, J = 278.00 Hz), 61.02, 55.53, 53.86, 51.42 (q, J = 28.80 Hz), 21.62, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.47$ ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.6 ml/min; 25 °C; 230 nm), first peak: t_R = 8.88 min, second peak: t_R = 12.70 min; HRMS (ESI) m/z calcd.



for $C_{23}H_{21}F_3NaO_3[M+Na]^+ = 425.1335$, found = 425.1343.



3jd; colorless oil; $[\alpha]_D^{20} = +109.3$ (*c* = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.76 (d, *J* = 8.40 Hz, 2H), 7.59–7.56 (m, 4H), 7.47–7.29 (m, 6H), 7.14–7.11 (m, 2H), 6.83 (s, 1H), 4.64–4.56 (m, 1H), 4.37–4.21 (m, 3H), 4.03–4.02 (m, 1H), 1.33 (t, *J* = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.71, 163.27, 147.44, 146.50, 140.23, 139.49, 133.85, 131.93, 129.74, 129.00, 128.95, 128.41, 128.06, 127.88, 127.22, 127.19, 126.32 (q, *J* = 278.10 Hz), 61.05, 55.56, 54.12, 51.44 (q, *J* = 28.90 Hz), 14.07; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.40 ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.6 ml/min; 25 °C; 230 nm), first peak: t_R = 11.10 min, second peak: t_R = 13.98 min; HRMS (ESI) m/z calcd. for C₂₈H₂₃F₃NaO₃ [M+Na] ⁺= 487.1491, found = 487.1497.





3ia; colorless oil; $[\alpha]_D{}^{20} = +102.0$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.42 (d, J = 5.60 Hz, 1H), 7.26–7.17 (m, 5H), 6.93–6.90 (m, 2H), 6.81 (s, 1H), 4.52–4.44 (m, 1H), 4.36–4.21 (m, 2H), 4.15–4.11 (m, 2H), 1.33 (t, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 198.48, 163.07, 147.02, 140.05, 137.91, 135.99, 132.54, 131.76 (q, J = 2.00 Hz), 130.60, 130.10, 128.90, 127.78, 127.41, 127.18, 126.02 (q, J = 276.30 Hz), 61.14, 58.27, 53.68, 50.12 (q, J = 29.30 Hz), 14.06; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.96$ ppm; Enantiomeric excess: 99%, determined by HPLC (Chiralpak AS-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 6.37 min, second peak: t_R = 7.08 min; HRMS (ESI) m/z calcd. for C₂₂H₁₇Cl₂F₃NaO₃ [M+Na] ⁺= 479.0399, found = 479.0399.





3ja; colorless oil; $[\alpha]_D^{20} = + 69.1$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.63–7.61 (m, 1H), 7.33–7.26 (m, 2H), 7.21–7.19 (m, 4H), 6.89–6.83 (m, 3H), 4.57–4.49 (m, 1H), 4.37–4.20 (m, 3H), 4.14–4.12 (m, 1H), 1.34 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 199.85, 163.15, 147.14, 140.29, 139.82, 133.86, 132.15, 131.85 (d, J = 2.10 Hz), 129.00, 128.77, 127.59, 127.42, 127.31, 126.13 (q, J= 278.10 Hz), 119.57, 61.11, 58.29, 53.20, 49.80 (q, J = 29.20 Hz), 14.08; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.97 ppm; Enantiomeric excess: 96%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 7.08 min, second peak: t_R = 7.75 min; HRMS (ESI) m/z calcd. for C₂₂H₁₈BrF₃NaO₃ [M+Na] ⁺ = 489.0284, found = 489.0288.





3ka; colorless oil; $[\alpha]_D^{20} = -28.0$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.96–7.94 (m, 1H), 7.90 (s, 1H), 7.84 (d, J = 8.40 Hz, 2H), 7.60–7.54 (m, 2H), 7.50–7.46 (m, 1H), 7.35–7.29 (m, 3H), 7.13–7.11 (m, 2H), 6.83 (s, 1H), 4.74–4.68 (m, 1H), 4.41–4.24 (m, 3H), 4.02–4.00 (m, 1H), 1.34 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.61, 163.37, 147.35, 140.41, 135.88, 132.34, 132.14, 132.03, 131.75, 129.67, 129.10, 128.99, 128.63, 128.30, 128.00, 127.72, 126.89, 126.44 (q, *J* = 278.00 Hz), 124.37, 61.12, 55.73, 54.57, 51.22 (q, *J* = 28.90 Hz), 14.13; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.39 ppm; Enantiomeric excess: 91%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.6 ml/min; 25 °C; 230 nm), first peak: t_R = 10.26 min, second peak: t_R = 15.57 min; HRMS (ESI) m/z calcd. for C₂₆H₂₁F₃NaO₃ [M+Na] ⁺ = 461.1335, found = 461.1344.





3la; colorless oil; $[\alpha]_D^{20} = +104.0$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.57 (d, J = 1.0 Hz, 1H), 7.37–7.30 (m, 3H), 7.16–7.15 (m, 2H), 6.94 (d, J = 3.50 Hz, 1H), 6.87 (s, 1H), 6.50–6.49 (m, 1H), 4.49–4.43 (m, 1H), 4.37–4.23 (m, 2H), 4.15–4.14 (m, 1H), 4.02–4.00 (m, 1H), 1.34 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 186.43, 163.16, 151.15, 147.79, 147.63, 140.34, 131.82, 128.88, 127.82, 127.74, 126.08 (q, J = 278.13 Hz), 119.79, 112.55, 61.05, 55.20, 54.68, 51.17 (q, J =29.13 Hz), 14.04; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.74 ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 6.18 min, second peak: t_R = 9.40 min; HRMS (ESI) m/z calcd. for C₂₀H₁₇F₃NaO4 [M+Na] ⁺= 401.0971, found = 401.0974.





3ma; colorless oil; $[\alpha]_D^{20} = +90.7$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.71–7.70 (m, 1H), 7.38–7.33 (m, 3H), 7.20–7.19 (m, 1H), 7.17–7.15 (m, 2H), 7.02–7.00 (m, 1H), 6.87 (s, 1H), 4.55–4.49 (m, 1H), 4.38–4.24 (m, 2H), 4.13–4.11 (m, 1H), 4.07–4.05 (m, 1H), 1.35 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 190.90, 163.16, 147.55, 142.77, 140.25, 135.64, 133.57, 131.82, 128.97, 128.24, 127.92, 127.87, 126.13 (q, J = 278.13 Hz), 61.05, 55.68, 55.50, 51.65 (q, J = 29.13Hz), 14.04; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.58$ ppm; Enantiomeric excess: 95%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 5.72 min, second peak: t_R = 7.49 min; HRMS (ESI) m/z calcd. for C₂₀H₁₇F₃NaO₃S [M+Na] ⁺= 417.0743, found = 417.0748.



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3na; colorless oil; $[\alpha]_D^{20} = + 46.3$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.85–7.83 (m, 1H), 7.64 (d, J = 8.00 Hz, 1H), 7.47–7.43 (m, 1H), 7.37–7.30 (m, 5H), 7.17–7.14 (m, 2H), 6.86 (s, 1H), 4.61–4.53 (m, 1H), 4.37–4.26 (m, 2H), 4.18–4.15 (m, 1H), 4.12–4.10 (m, 1H), 1.33 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 192.26, 163.13, 147.34, 143.13, 142.04, 140.25, 138.74, 131.91, 131.22, 129.04, 128.04, 127.96, 127.93, 126.20, 126.14 (q, J = 278.00 Hz), 125.07, 122.86, 61.08, 55.72, 55.52, 51.53 (q, J = 29.10 Hz), 14.04; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.55$ ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 7.64 min, second peak: t_R = 9.56 min; HRMS (ESI) m/z calcd. for C₂₄H₁₉F₃NaO₃S [M+Na] ⁺ = 467.0899, found = 467.0900.





30a; colorless oil; [α]_D²⁰ = + 50.7 (*c* = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.50 (d, *J* = 4.00 Hz, 1H), 8.13 (d, *J* = 8.00 Hz, 1H), 7.89–7.86 (m, 1H), 7.49–7.46 (m, 1H), 7.33–7.28 (m, 3H), 7.18–7.16 (m, 2H), 6.91 (s, 1H), 5.01–4.99 (m, 1H), 4.54–4.47 (m, 1H), 4.39–4.25 (m, 2H), 4.07 (br, 1H), 1.36 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 199.28, 163.40, 151.56, 148.89, 147.76, 140.44, 136.93, 132.07, 128.40, 128.10, 127.52, 127.26, 126.24 (q, J = 278.25 Hz), 123.14, 60.98, 54.87, 52.42, 50.55 (q, J = 29.00 Hz), 14.09; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.38 ppm; Enantiomeric excess: 82%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 6.22 min, second peak: t_R = 9.91 min; HRMS (ESI) m/z calcd. for C₂₁H₁₉F₃NO₃ [M+Na] ⁺ = 390.1312, found = 390.1318.





3pa; colorless oil; $[\alpha]_D^{20} = +54.5$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.37–7.27 (m, 3H), 7.16–7.14 (m, 2H), 6.77 (s, 1H), 6.33–6.31 (m, 1H), 4.47–4.38 (m, 1H), 4.35–4.19 (m, 2H), 3.93–3.90 (m, 1H), 3.84–3.81 (m, 1H), 2.33–1.93 (m, 4H), 1.65–1.54 (m, 4H), 1.32 (t, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 198.57, 163.37, 147.38, 143.51, 140.60, 137.88, 132.04, 128.85, 128.00, 126.33 (q, J = 277.00Hz), 60.95, 55.71, 52.81, 51.25 (q, J = 28.70 Hz), 26.11, 23.55, 21.79, 21.32, 14.05; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.65 ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 9.18 min, second peak: t_R = 12.20 min; HRMS (ESI) m/z calcd. for C₂₂H₂₃F₃NaO₃ [M+Na] ⁺= 415.1491, found = 415.1496.





3qa; colorless oil; $[\alpha]_D^{20} = + 116.8$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.41–7.38 (m, 2H), 7.35–7.32 (m, 1H), 7.23–7.21 (m, 2H), 6.79 (s, 1H), 4.35–4.21 (m, 3H), 4.06–4.04 (m, 1H), 3.55–3.53 (m, 1H), 2.42–2.36 (m, 1H), 1.79–1.72 (m, 4H), 1.46–1.16 (m, 9H); ¹³C NMR (125 MHz, CDCl₃): δ 210.89, 163.23, 147.67, 140.83, 131.54, 129.09, 127.80, 127.62, 126.17 (q, J = 278.00 Hz), 61.01, 57.01, 53.94, 50.70 (q, J = 28.88 Hz), 50.10, 28.57, 28.02, 25.59, 25.49, 25.35, 14.06; ¹⁹F NMR (376 MHz, CDCl₃) δ = -68.02 ppm; Enantiomeric excess: 91%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.4 ml/min; 25 °C; 230 nm), first peak: t_R = 10.63 min, second peak: t_R = 12.86 min; HRMS (ESI) m/z calcd. for C₂₂H₂₅F₃NaO₃ [M+Na] ⁺= 417.1648, found = 417.1653.





3ra; colorless oil; $[\alpha]_D^{20} = + 33.9$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.72–7.70 (m, 2H), 7.61–7.58 (m, 1H), 7.41–7.38 (m, 2H), 7.35–7.33 (m, 3H), 7.10–7.08 (m, 2H), 6.77–6.76 (m, 1H), 4.84–4.77 (m, 1H), 4.38–4.26 (m, 3H), 3.92–3.91 (m, 1H), 1.36 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.87, 163.78, 146.36, 139.96, 135.00, 133.87, 132.20, 129.18, 128.91, 128.64, 128.18, 127.92, 122.47 (t, J = 36.25 Hz), 120.19 (t, J = 36.50 Hz), 118.20–116.58 (m), 115.63–114.55 (m), 112.94 (t, J = 36.75 Hz), 61.11, 56.17, 53.47, 48.33 (t, J = 20.50Hz), 14.02; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -82.94$, -116.11 (q) ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.6 ml/min; 25 °C; 230 nm), first peak: t_R = 7.82 min, second peak: t_R = 11.14 min; HRMS (ESI) m/z calcd. for C₂₃H₁₉F₅NaO₃ [M+Na] ⁺ = 461.1147, found = 461.1154.





3sa; colorless oil; $[\alpha]_D^{20} = +18.8$ (*c* = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ

7.73 (d, J = 7.50 Hz, 2H), 7.62–7.59 (m, 1H), 7.42–7.39 (m, 2H), 7.35–7.33 (m, 3H), 7.10–7.08 (m, 2H), 6.77–6.76 (m, 1H), 4.95–4.90 (m, 1H), 4.39–4.25 (m, 3H), 3.90 (br, 1H), 1.36 (t, J = 7.50 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.72, 163.83, 146.17, 139.93, 134.96, 133.88, 132.32, 129.18, 128.92, 128.67, 128.21, 127.93, 121.08, 119.24–118.52 (m), 117.19–116.23 (m), 115.14–114.21 (m), 111.80–110.90 (m), 109.70–108.80 (m), 107.59–106.69 (m), 61.08, 56.49, 53.39, 48.39 (t, J = 20.63 Hz), 14.00; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -80.38$ (t), -111.06–-115.15 (m), -124.53–126.91 (m) ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.6 ml/min; 25 °C; 230 nm), first peak: t_R = 7.24 min, second peak: t_R = 9.74 min; HRMS (ESI) m/z calcd. for C₂₄H₁₉F₇NaO₃ [M+Na] ⁺= 511.1115, found = 511.1125.





3ab; colorless oil; $[\alpha]_D^{20} = +137.4$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.74–7.72 (m, 2H), 7.62–7.59 (m, 1H), 7.43–7.40 (m, 2H), 7.32–7.30 (m, 2H), 7.04–7.02 (m, 2H), 6.80 (s, 1H), 4.59–4.53 (m, 1H), 4.38–4.32 (m, 1H), 4.30–4.25 (m, 1H), 4.21–4.19 (m, 1H), 4.02–4.01 (m, 1H), 1.35 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.85, 163.08, 146.74, 138.68, 135.09, 133.99, 133.77, 132.39, 129.35, 129.10, 129.05, 128.73, 126.18 (q, J = 278.00 Hz), 61.15, 54.61, 54.84, 51.25 (q, J = 28.88 Hz), 14.05; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.53$ ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 6.88 min, second peak: t_R = 10.44 min; HRMS (ESI) m/z calcd. for C₂₂H₁₈ClF₃NaO₃ [M+Na] ⁺ = 445.0789, found = 445.0792.





3ac; colorless oil; $[\alpha]_D^{20} = + 121.6$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.73 (d, J = 7.50 Hz, 2H), 7.60–7.57 (m, 1H), 7.41–7.38 (m, 2H), 7.02 (d, J = 8.50 Hz, 2H), 6.87 (d, J = 8.50 Hz, 2H), 6.80 (s, 1H), 4.63–4.57 (m, 1H), 4.38–4.31 (m, 1H), 4.30–4.25 (m, 1H), 4.24–4.21 (m, 1H), 3.96–3.95 (m, 1H), 3.83 (s, 3H), 1.35 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 198.19, 163.32, 159.18, 147.69, 135.20, 133.79, 132.18, 131.53, 129.15, 128.61, 126.32 (q, J = 278.00 Hz), 114.26, 61.01, 55.26, 54.83, 54.25, 51.07 (q, J = 28.88 Hz), 14.06; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.50$ ppm; Enantiomeric excess: 93%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 7.74 min, second peak: t_R = 14.36 min; HRMS (ESI) m/z calcd. for C₂₃H₂₁F₃NaO₄ [M+Na] ⁺ = 441.1284, found = 441.1289.





3ad; colorless oil; $[\alpha]_D^{20} = +52.5$ (*c* = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.87–7.85 (m, 2H), 7.64–7.61 (m, 1H), 7.47–7.44 (m, 2H), 7.29–7.28 (m, 1H), 7.00–6.98 (m, 1H), 6.86–6.84 (m, 2H), 4.55–4.49 (m, 1H), 4.38–4.33 (m, 3H), 4.29–4.24 (m, 1H), 1.35 (t, *J* = 7.50 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.62, 163.07, 146.57, 142.68, 134.96, 133.98, 131.56, 129.14, 128.75, 127.17, 126.07, 126.06 (q, *J* = 276.75 Hz), 125.33, 61.13, 54.10, 51.01 (q, *J* = 29.00 Hz), 49.98, 14.06; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.53 ppm; Enantiomeric excess: 95%, determined by HPLC (Chiralpak AD-H, hexane/*i*-PrOH = 90/10; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 7.18 min, second peak: t_R = 9.66 min; HRMS (ESI) m/z calcd. for C₂₀H₁₇F₃NaO₃S [M+Na] ⁺= 417.0743, found = 417.0745.





3ce; colorless oil; $[\alpha]_D^{20} = -63.7$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.94–7.91 (m, 2H), 7.53–7.50 (m, 2H), 6.80 (s, 1H), 4.40–4.20 (m, 3H), 3.84–3.83 (m, 1H), 2.99–2.94 (m, 1H), 1.37 (d, J = 7.00 Hz, 3H), 1.33 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 196.67, 163.24, 150.32, 140.35, 133.60, 130.24, 130.00, 129.29, 126.21 (q, J = 277.88 Hz), 60.94, 52.38, 50.36 (q, J = 29.00 Hz), 44.68, 19.07, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) δ = -68.10 ppm; Enantiomeric excess: 95%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 11.52 min, second peak: t_R = 15.77 min; HRMS (ESI) m/z calcd. for C₁₇H₁₆ClF₃NaO₃ [M+Na] ⁺= 383.0632, found = 383.0633.




3cf; colorless oil; $[\alpha]_D^{20} = -31.4$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.94–7.91 (m, 2H), 7.52–7.50 (m, 2H), 6.93 (s, 1H), 4.33–4.27 (m, 1H), 4.24–4.20 (m, 2H), 3.89–3.88 (m, 1H), 2.95–2.91 (m, 1H), 1.72–1.65 (m, 2H), 1.48–1.39 (m, 1H), 1.37–1.31 (m, 4H), 0.90 (t, J = 7.50 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.47, 163.23, 149.52, 140.33, 133.72, 130.41, 130.01, 129.25, 126.19 (q, J = 278.00 Hz), 60.90, 50.94, 50.85 (q, J = 28.50 Hz), 49.87, 36.52, 20.96, 14.07, 13.87; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.88$ ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 9.78 min, second peak: t_R = 11.91 min; HRMS (ESI) m/z calcd. for C₁₉H₂₀ClF₃NaO₃ [M+Na] ⁺= 411.0945, found = 411.0946.





3cg; colorless oil; [α]_D²⁰ = - 27.2 (*c* = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.90 (d, *J* = 8.40 Hz, 2H), 7.49 (d, *J* = 8.80 Hz, 2H), 6.92 (s, 1H), 4.32–4.16 (m, 3H), 3.88–3.86 (m, 1H), 2.90–2.87 (m, 1H), 1.71–1.64 (m, 2H), 1.33–1.22 (m, 19H), 0.88 (t, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.50, 163.25, 149.57, 140.33, 133.74, 130.40, 130.01, 129.24, 126.20 (q, J = 277.90 Hz), 60.90, 50.96, 50.82 (q, J = 28.80 Hz), 50.10, 34.36, 31.84, 29.51, 29.46, 29.33, 29.26, 27.72, 22.64, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.87$ ppm; Enantiomeric excess: 96%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 7.00 min, second peak: t_R = 8.33 min; HRMS (ESI) m/z calcd. for C₂₆H₃₄ClF₃NaO₃ [M+Na] ⁺= 509.2041, found = 509.2044.





3ch; colorless oil; $[\alpha]_D^{20} = -101.3$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.44 (d, J = 9.0 Hz, 2H),7.33–7.27 (m, 5H), 7.17–7.15 (m, 2H), 6.89 (s, 1H), 4.35–4.20 (m, 3H), 3.97–3.96 (m, 1H), 3.24–3.20 (m, 1H), 3.07–3.03 (m, 1H), 2.88–2.83 (m, 1H), 1.34 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.36, 163.13, 149.03, 139.93, 138.38, 133.14, 131.10, 129.87, 129.06, 129.03, 128.99, 126.97, 126.23 (q, J = 278.13 Hz), 61.01, 51.98,50.69 (q, J = 29.00 Hz), 49.10, 39.89, 14.09; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.32$ ppm; Enantiomeric excess: 93%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 12.21 min, second peak: t_R = 13.82 min; HRMS (ESI) m/z calcd. for C₂₃H₂₀ClF₃NaO₃ [M+Na] ⁺= 459.0945, found = 459.0958.





3ci; colorless oil; $[\alpha]_D^{20} = + 8.7$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.94–7.91 (m, 2H), 7.54–7.51 (m, 2H), 6.91 (s, 1H), 4.34–4.19 (m, 3H), 3.92–3.90 (m, 1H), 3.54–3.52 (m, 2H), 2.99–2.96 (m, 1H), 1.97–1.75 (m, 4H), 1.33 (t, J = 7.50 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 197.09, 163.06, 148.55, 140.55, 133.50, 131.09 (d, J = 1.50 Hz), 130.04, 129.36, 126.11 (q, J = 278.00 Hz), 61.05, 50.91 (q, J = 28.88Hz), 50.83, 49.12, 44.22, 31.37, 30.38, 14.08; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.84$ ppm; Enantiomeric excess: 91%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 12.10 min, second peak: t_R = 15.73 min; HRMS (ESI) m/z calcd. for C₁₉H₁₉Cl₂F₃NaO₃ [M+Na] ⁺= 445.0556, found = 445.0560.





3cj; colorless oil; $[\alpha]_D{}^{20} = -18.6$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.91 (d, J = 8.40 Hz, 2H), 7.50 (d, J = 8.40 Hz, 2H), 6.92 (s, 1H), 4.32–4.18 (m, 3H), 3.91–3.89 (m, 1H), 3.66 (s, 3H), 2.94–2.93 (m, 1H), 2.32–2.29 (m, 2H), 1.80–1.70 (m, 4H), 1.31 (t, J = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.25, 173.30, 163.09, 148.90, 140.42, 133.58, 130.79, 130.06, 129.30, 126.12 (q, J = 278.00 Hz), 60.96, , 51.60, 50.67, 50.90 (q, J = 29.00 Hz), 49.59, 33.50, 22.91, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.86$ ppm; Enantiomeric excess: 91%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 95/05; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 20.74 min, second peak: t_R = 27.18 min; HRMS (ESI) m/z calcd. for C₂₁H₂₂ClF₃NaO₅ [M+Na] ⁺= 469.1000, found = 469.1008.





3ck; colorless oil; $[\alpha]_D^{20} = +20.6$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.91 (d, J = 8.40 Hz, 2H), 7.49 (d, J = 8.80 Hz, 2H), 6.96 (s, 1H), 4.32–4.16 (m, 2H), 4.07–3.99 (m, 2H), 2.89–2.86 (m, 1H), 1.87–1.78 (m, 1H), 1.30 (t, J = 7.60 Hz, 3H), 0.97 (d, J = 6.80 Hz, 3H), 0.93 (d, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ

199.30, 163.24, 149.09, 140.39, 134.06, 130.34, 130.08, 129.23, 126.17 (q, J = 278.30 Hz), 60.91, 57.44, 52.61 (q, J = 28.70 Hz), 48.06, 31.52, 21.69, 19.80, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.56$ ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 9.32 min, second peak: t_R = 12.16 min; HRMS (ESI) m/z calcd. for C₁₉H₂₀ClF₃NaO₃ [M+Na] ⁺= 411.0945, found = 411.0948.





3cl; colorless oil; $[\alpha]_D{}^{20} = + 32.4$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.95–7.92 (m, 2H), 7.52–7.49 (m, 2H), 6.95 (s, 1H), 4.28–4.18 (m, 2H), 4.05–3.99 (m, 2H), 2.95–2.93 (m, 1H), 2.03–1.78 (m, 3H), 1.67–1.54 (m, 4H), 1.32 (t, J = 7.00 Hz, 3H), 1.28–1.20 (m, 1H), 1.07–0.99 (m, 1H); ¹³C NMR (125 MHz, CDCl₃): δ 199.15, 163.30, 149.95, 140.37, 134.02, 130.11, 130.03, 129.20, 126.18 (q, J = 278.25 Hz), 60.90, 56.05, 52.52 (q, J = 28.75 Hz), 49.54, 44.12, 32.24, 30.68, 25.16, 24.94, 14.08; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.71$ ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 10.66 min, second peak: t_R = 12.14 min; HRMS (ESI) m/z calcd. for C₂₁H₂₂ClF₃NaO₃ [M+Na] ⁺= 437.1102, found = 437.1112.





3cm; colorless oil; $[\alpha]_D{}^{20} = + 37.4$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.94–7.92 (m, 2H), 7.51–7.49 (m, 2H), 7.01 (s, 1H), 4.32–4.17 (m, 2H), 4.09–4.07 (m, 1H), 4.03–3.97 (m, 1H), 2.92–2.90 (m, 1H), 1.87–1.66 (m, 5H), 1.51–1.45 (m, 1H), 1.31 (t, J = 7.00 Hz, 3H), 1.24–1.11 (m, 3H), 1.02–0.93 (m, 2H); ¹³C NMR (125 MHz, CDCl₃): δ 199.24, 163.25, 149.41, 140.35, 133.96, 130.09, 129.94, 129.20, 126.17 (q, J = 278.50 Hz), 60.87, 56.34, 52.43 (q, J = 28.75 Hz), 47.66, 41.32, 32.21, 30.46, 26.04, 26.03, 25.95, 14.06; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.59$ ppm; Enantiomeric excess: 92%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 9.35 min, second peak: t_R = 10.76 min; HRMS (ESI) m/z calcd. for C₂₂H₂₄ClF₃NaO₃ [M+Na] ⁺ = 451.1258, found = 451.1266.





3ag; colorless oil; $[\alpha]_D^{20} = -19.6$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.97–7.95 (m, 2H), 7.64–7.61 (m, 1H), 7.53–7.50 (m, 2H), 6.92 (s, 1H), 4.33–4.16 (m, 3H), 3.94–3.93 (m, 1H), 2.91–2.88 (m, 1H), 1.74–1.61 (m, 2H), 1.33–1.21 (m, 19H), 0.88 (t, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 198.66, 163.34, 149.67, 135.47, 133.66, 130.47, 128.86, 128.62, 126.19 (q, J = 277.90 Hz), 60.84, 51.00, 50.72 (q, J = 28.90 Hz), 50.12, 34.35, 31.83, 29.50, 29.44, 29.32, 29.30, 29.24, 27.68, 22.62, 14.05; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.87$ ppm; Enantiomeric excess: 87%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 8.48 min, second peak: t_R = 11.09 min; HRMS (ESI) m/z calcd. for C₂₆H₃₅F₃NaO₃ [M+Na] ⁺= 475.2431, found = 475.2435.





3bg; colorless oil; $[\alpha]_{D}^{20} = -7.4$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 8.02–7.98 (m, 2H), 7.21–7.17 (m, 2H), 6.92 (s, 1H), 4.33–4.16 (m, 3H), 3.89–3.88 (m, 1H), 2.93–2.87 (m, 1H), 1.72–1.65 (m, 2H), 1.33–1.22 (m, 19H), 0.88 (t, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.18, 166.13 (d, J = 254.60 Hz), 149.62, 131.84 (d, J = 2.90 Hz), 131.31 (d, J = 9.40 Hz), 130.41, 126.23 (q, J = 278.10 Hz), 116.07 (d, J = 21.80 Hz), 60.87, 50.94 (q, J = 28.80 Hz), 50.91, 50.16, 34.35, 31.83, 29.50, 29.45, 29.33, 29.32, 29.24, 27.72, 22.62, 14.06, 14.04; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -67.89$, -103.91 ppm; Enantiomeric excess: 95%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 95/05; flow rate 0.5 ml/min; 25 °C; 230 nm), first peak: t_R = 11.87 min, second peak: t_R = 13.81 min; HRMS (ESI) m/z calcd. for C₂₆H₃₄F₄NaO₃ [M+Na] ⁺= 493.2336, found = 493.2338.





3dg; colorless oil; $[\alpha]_D{}^{20} = -30.7$ (c = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.82 (d, J = 8.40 Hz, 2H), 7.66 (d, J = 8.40 Hz, 2H), 6.92 (s, 1H), 4.33–4.16 (m, 3H),

3.87–3.85 (m, 1H), 2.90–2.86 (m, 1H), 1.69–1.64 (m, 2H), 1.33–1.22 (m, 19H), 0.88 (t, J = 6.80 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 197.70, 163.26, 149.58, 134.13, 132.25, 130.37, 130.10, 129.09, 126.19 (q, J = 278.00 Hz), 60.93, 50.94, 50.78 (q, J = 32.40 Hz), 50.07, 34.38, 31.86, 29.53, 29.48, 29.34, 29.28, 27.73, 22.66, 14.10; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.86 ppm; Enantiomeric excess: 96%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 7.53 min, second peak: t_R = 8.68 min; HRMS (ESI) m/z calcd. for C₂₆H₃₄BrF₃NaO₃ [M+Na] ⁺= 553.1536, found = 553.1541.





3jg; colorless oil; $[\alpha]_D^{20} = -11.7$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.67–7.66 (m, 1H), 7.45–7.41 (m, 1H), 7.38–7.34 (m, 2H), 6.86 (s, 1H), 4.44–4.20 (m, 3H), 3.86–3.85 (m, 1H), 3.02–2.98 (m, 1H), 1.55–1.40 (m, 2H), 1.35–1.12 (m, 19H), 0.90 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 200.67, 163.28, 149.17, 140.

10, 133.77, 131.92, 130.21, 128.65, 127.40, 126.15 (q, J = 277.88 Hz), 119.21, 60.89, 54.87, 49.13 (q, J = 29.00 Hz), 48.52, 34.36, 31.85, 29.50, 29.38, 29.27, 29.25, 29.05, 27.39, 22.64, 14.08; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -68.29$ ppm; Enantiomeric excess: 96%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 97/03; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 7.99 min, second peak: t_R = 9.54 min;



HRMS (ESI) m/z calcd. for $C_{26}H_{34}BrF_3NaO_3$ [M+Na] ⁺ = 553.1536, found = 553.1550.



3kg; colorless oil; $[\alpha]_D^{20} = -52.5$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 8.50 (s, 1H), 8.06–7.92 (m, 4H), 7.68–7.60 (m, 2H), 6.97 (s, 1H), 4.38–4.21 (m, 3H), 4.13–4.12 (m, 1H), 3.00–2.97 (m, 1H), 1.85–1.69 (m, 2H), 1.36–1.19 (m, 19H), 0.89 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 198.59, 163.39, 149.79, 135.80, 132.74, 132.45, 130.49, 130.34, 129.63, 128.89, 128.86, 127.79, 127.05, 126.37 (q, J= 278.00 Hz), 124.23, 60.87, 51.05, 50.84 (q, J = 28.63 Hz), 50.28, 34.39, 31.82, 29.48, 29.43, 29.34, 29.30, 29.22, 27.72, 22.62, 14.09, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.74 ppm; Enantiomeric excess: 94%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 95/05; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 10.81 min, second peak: t_R = 13.76 min; HRMS (ESI) m/z calcd. for C₃₀H₃₇F₃NaO₃ [M+Na] ⁺ = 525.2587, found = 525.2591.





3mg; colorless oil; $[\alpha]_D^{20} = +10.7$ (c = 0.33, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 7.80 (d, J = 3.50 Hz, 1H), 7.76 (d, J = 5.00 Hz, 1H), 7.22–7.20 (m, 1H), 6.96 (s, 1H), 4.33–4.18 (m, 3H), 3.75–3.73 (m, 1H), 3.06–3.03 (m, 1H), 1.77–1.63 (m, 2H), 1.33–1.25 (m, 19H), 0.89 (t, J = 6.50 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 191.66, 163.28, 149.98, 142.75, 135.20, 132.58, 130.21, 128.45,126.15 (q, J = 278.00 Hz), 60.86, 52.34, 51.48 (q, J = 28.88 Hz), 50.42, 34.38, 31.84, 29.46, 29.41, 29.35, 29.25, 27.71, 22.63, 14.07; ¹⁹F NMR (376 MHz, CDCl₃) $\delta = -68.00$ ppm; Enantiomeric excess: 91%, determined by HPLC (Chiralpak IE, hexane/*i*-PrOH = 95/05; flow rate 0.8 ml/min; 25 °C; 230 nm), first peak: t_R = 8.61 min, second peak: t_R = 10.73 min; HRMS (ESI) m/z calcd. for C₂₄H₃₃F₃NaO₃S [M+Na] ⁺= 481.1995, found = 481.1999.





(*S*)-**2a**;^[3] colorless oil; $[\alpha]_D^{20} = +239.8.3$ (*c* = 0.33, CHCl₃); ¹H NMR (400 MHz, CDCl₃): δ 7.28–7.19 (m, 5H), 6.55 (d, *J* = 6.40 Hz, 1H), 5.94 (d, *J* = 6.40 Hz, 1H), 4.15 (q, *J* = 7.20 Hz, 2H), 1.21 (t, *J* = 7.20 Hz, 3H); ¹³C NMR (100 MHz, CDCl₃): δ 214.58, 165.07, 131.11, 128.81, 128.06, 127.46, 98.63, 91.91, 61.10, 14.20; Enantiomeric excess: 76%, determined by HPLC (Chiralpak OD-H, hexane/*i*-PrOH = 99/01; flow rate 1.0 ml/min; 25 °C; 230 nm), first peak: t_R = 9.22 min, second peak: t_R = 9.63 min.



(S)-2g;^[4] colorless oil; $[\alpha]_D{}^{20} = +306.3$ (c = 1.0, CHCl₃); ¹H NMR (500 MHz, CDCl₃): δ 5.63–5.66 (m, 2H), 4.23–4.17 (m, 2H), 2.16–2.11 (m, 2H), 1.49–1.44 (m, 2H),

1.37–1.27 (m, 2H), 0.89 (t, J = 7.00 Hz, 3H); ¹³C NMR (125 MHz, CDCl₃): δ 212.26, 166.27, 95.34, 88.17, 60.66, 31.86, 29.56, 29.54, 29.32, 29.28, 28.89, 28.68, 27.46, 22.64, 14.20, 14.06; ¹⁹F NMR (376 MHz, CDCl₃) δ = -67.50 ppm; Enantiomeric excess: 83%, determined by HPLC (Chiralpak OD-H, hexane/*i*-PrOH = 99/01; flow rate 0.5 ml/min; 25 °C; 210 nm), first peak: t_R = 9.68 min, second peak: t_R = 10.23 min.



9. References

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10. ¹H, ¹³C, ³¹P and ¹⁹F NMR Spectra











zhouw-6-103c



















zhouw-6-106





zhouw-6-107



zhouw-6-107f



zhouw-6-77







zhouw-6-108f





zhouw-6-112






zhouw-6-113



zhouw-6-113f









zhouw-6-115f

















zw-7-8







zhouw-7-49c







2:07 1:01 2:05 2:05 2:05

8



3.05 1.04 1.01 4 3

1

0

-1

ppm

2

1.07 1.07

<u>9.11</u>

5























zhouw-7-24s









zhouw-7-19




















zhouw-7-63







zhouw-7-61















