

Site-selective ^{18}F fluorination of unactivated C-H bonds mediated by a manganese porphyrin

Wei Liu^a, Xiongyi Huang^a, Micheal Placzek^{b,c}, Shane W. Krska^d, Paul McQuade^e, Jacob M. Hooker^{b,c*}, John T. Groves^{a*}

^aDepartment of Chemistry, Princeton University, Princeton, New Jersey 08544, United States

^bAthinoula A. Martinos Center for Biomedical Imaging, Massachusetts General Hospital and Harvard Medical School, Charlestown, Massachusetts 02129, United States

^cDivision of Nuclear Medicine and Molecular Imaging, Department of Radiology, Massachusetts General Hospital, Boston, Massachusetts 02114, United States

^dDepartment of Process Chemistry, Merck Research Laboratories, Rahway, New Jersey 07065, United States

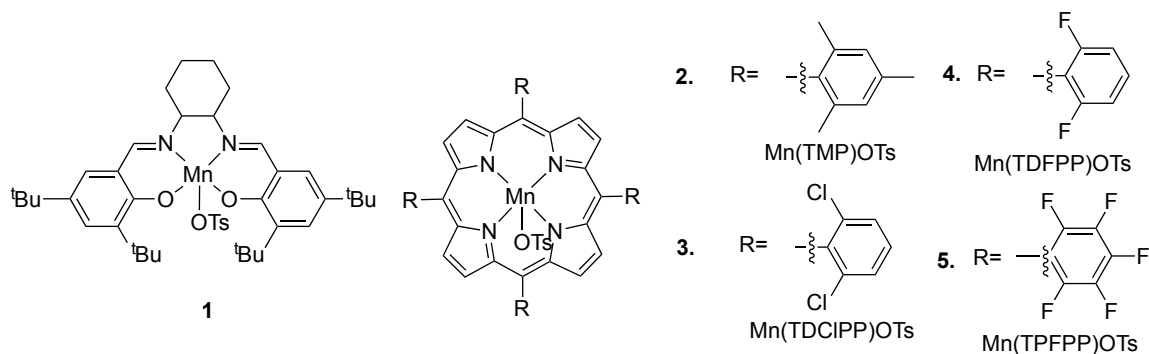
^eImaging Research, Merck Research Laboratories, West Point, Pennsylvania 19486, United States

SUPPORTING INFORMATION

1. General information

Manganese tetrakis(pentafluorophenyl)porphyrin chloride (Mn(TPFPP)Cl) was prepared by refluxing the free base with manganese acetate, followed by HCl treatment. Mn(TPFPP)OTs was prepared by treating Mn(TPFPP)Cl with stoichiometric amounts of silver tosylate in refluxing toluene. Iodosylbenzene (PhIO) was prepared by hydrolysis of iodobenzene diacetate with sodium hydroxide solution. Other purchased materials were of the highest purity available from Aldrich and used without further purification. ^1H NMR spectra were obtained on a Bruker NB 300 spectrometer or a Bruker Avance-III (500 MHz) spectrometer and are reported in ppm using solvent as an internal standard (CDCl_3 at δ 7.26, acetone- d_6 at 2.04, or methylene chloride- d_2 at 5.32). Data reported as: chemical shift (δ), multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, m = multiplet), coupling constant (Hz); integrated intensity. ^{13}C NMR spectra were recorded on a Bruker 500 (125 MHz) spectrometer and are reported in ppm using solvents as an internal standard (CDCl_3 at 77.15 ppm, acetone- d_6 at 29.92 ppm, or methylene chloride- d_2 at 54.0). ^{19}F NMR spectra (282 MHz) were obtained on a Bruker NB 300 spectrometer and were referenced relative to relative to CFCl_3 . GC/MS analyses were performed on an Agilent 7890A gas chromatograph equipped with an Agilent 5975 mass selective detector. High-resolution mass spectra were obtained from the Princeton University mass spectrometer facility by electrospray ionization (ESI). High-performance liquid chromatography (HPLC) was performed on an Agilent 1100 series instrument with a binary pump and a diode array detector.

Structures of the catalysts used in this study (Table 1)

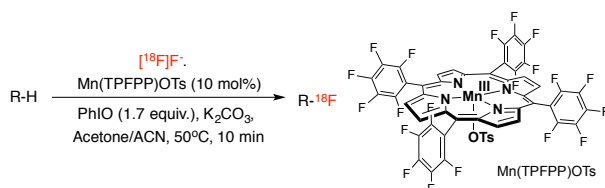


2. Radiochemistry

General methods

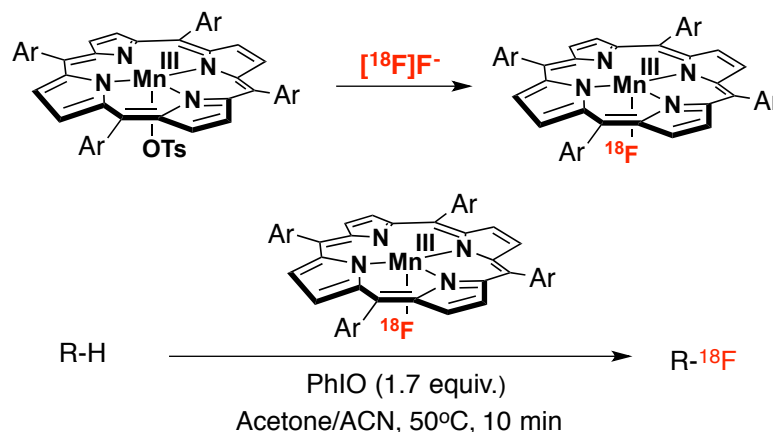
No-carrier-added [^{18}F]fluoride was produced from water 97% enriched in ^{18}O (ISOFLUX, USA) by the nuclear reaction $^{18}\text{O}(\text{p},\text{n})^{18}\text{F}$ using a Siemens Eclipse HP cyclotron and a silver-bodied target at Massachusetts General Hospital Athinoula A. Martinos Center for Biomedical Imaging. The produced [^{18}F]fluoride in water was transferred from the cyclotron target by helium push.

Radiosynthesis of ^{18}F labeled molecules



A 4 mL vial with a screw cap was charged with Mn(III)TPFPP OTs (16 mg, 0.022 mmol), substrate (0.25 mmol) and a stir bar (2×5 mm), followed with 0.4 mL acetonitrile. A portion of aqueous $[^{18}\text{F}]\text{fluoride}$ solution (40 – 50 μL , 4 – 5 mCi) obtained from the cyclotron was loaded on to an Chromafix PS- HCO_3 IEX cartridge, which had been previously washed with 5 mg/mL K_2CO_3 in Milli-Q water followed by 5 mL of Milli-Q water. Then, the cartridge loaded with $[^{18}\text{F}]\text{fluoride}$ was washed with 2 mL Milli-Q water and $[^{18}\text{F}]\text{fluoride}$ was released from the cartridge using 0.8 mL 5.0 mg/mL K_2CO_3 in Milli-Q water. A portion of the resulting $[^{18}\text{F}]\text{fluoride}$ solution (25 μL , 125 – 150 μCi) was diluted with 3.0 mL acetone. 0.2 mL of this $[^{18}\text{F}]\text{fluoride}$ acetone solution was added to the vial containing the catalyst and the substrate. The resulting solution was stirred for 1 min at room temperature. Then 80 mg (0.25 mmol) iodosylbenzene (PhIO) was added to the solution, and the vial was capped and stirred at 50 $^\circ\text{C}$ for 10 min. After 10 min, an aliquot of the reaction mixture was taken and spotted on a silica gel TLC plate. The plate was developed in an appropriate eluent and scanned with a Bioscan AR-2000 Radio TLC Imaging Scanner.

Dry-down free procedure for radiosynthesis of ^{18}F labeled molecules



A 4 mL vial with a screw cap was charged with substrate (0.33 mmol) and a stir bar (2×5 mm) 80 mg PhIO, followed with 0.4 mL acetonitrile. A portion of aqueous $[^{18}\text{F}]\text{fluoride}$ solution (40 – 50 μL , 4 – 5 mCi) obtained from the cyclotron was loaded on to a Chromafix PS- HCO_3 IEX cartridge, which had been previously washed with 5 mg/mL K_2CO_3 in Milli-Q water followed by 3 mL of Milli-Q water. Then, the cartridge loaded with $[^{18}\text{F}]\text{fluoride}$ was washed with 3 mL Milli-Q water and the residual water was removed by air purge. $[^{18}\text{F}]\text{fluoride}$ was then slowly eluted using 0.5 mL of an acetone

solution of Mn(TPFPP)OTs (16 mg) (> 90% radioactivity being eluted). The obtained solution was added to the vial containing the substrate. The vial was capped and stirred at 50 °C for 10 min. After 10 min, an aliquot of the reaction mixture was taken and spotted on a silica gel TLC plate. The plate was developed in an appropriate eluent and scanned with a Bioscan AR-2000 Radio TLC Imaging Scanner.

Scale-up synthesis of [¹⁸F]F-ACPC

The synthesis was performed with a Gilson automated liquid handler. The QMA resin containing the [¹⁸F]fluoride was obtained from Siemens Molecular Imaging, Inc., North Wales, PA. A 1 mL Wheaton conical glass vial was charged with 10 mg precursor, 10 mg PhIO, and 0.1 mL of acetonitrile. Vortex the vial until a suspension was formed and then placed the vial in a microwave cavity. A stock solution of Mn(TPFPP)OTs was prepared by dissolving 10 mg Mn(TPFPP)OTs in 1 mL acetonitrile. Passing 0.6 mL of the Mn(TPFPP)OTs solution through the QMA resin to elute the ¹⁸F. The elution efficiency was around 80%. Adding 0.3 mL of the eluted catalyst solution to the 1 mL conical vial containing the precursor and oxidant. Heat the vial with microwave reactor (50 °C, 50 W, 10 min). After the reaction, 0.5 mL water was added to the reaction mixture and then subjected to prep-HPLC separation.

Determination of specific activity of [¹⁸F]Butylbenzoate 2

The produced [¹⁸F]fluoride in water (256 mCi) was transferred from the cyclotron target by helium push (@ 9:30 am). A portion of aqueous [¹⁸F]fluoride solution (3.7 mCi) was loaded on to a Chromafix PS-HCO₃ IEX cartridge and [¹⁸F]Mn(TPFPP)F acetone solution was obtained followed by the dry-down free procedure described above (3.4 mCi, 92% trapping efficiency, @10:30 pm). 30 mg of butyl benzoate was added to the [¹⁸F]Mn(TPFPP)F acetone solution and then stirred at 50 °C for 1 min. Then, 50 mg PhIO was added to the reaction solution and the vial was capped and stirred for 10 min at 50 °C. The resulting mixture was passed through a syringe filter and was then subjected to prep-HPLC (2.7 mCi injected, @10:55 pm) The collected radioactive peak gave 550 µCi of [¹⁸F]2 (14.9 % non-decay corrected RCY, @11:15 pm).

The calibration curve was made according to the following procedure: solutions of different concentrations of fluorinated butyl benzoate were injected at constant volume on the HPLC. By multiplying the concentration times the volume we get nmol. The samples were analyzed by analytical HPLC and the peak area was recorded and used to construct a calibration curve. (**Figure S2-1**) A 50 µL sample of the collected fraction of [¹⁸F]butylbenzoate was used for purity analysis by HPLC (18 µCi, @11:25 pm) A UV absorbance of 544 of [¹⁸F]butylbenzoate was determined, corresponding to 2.9 nmol of **25**. The specific activity was thus calculated to be 1.24 Ci/µmol @EOB.

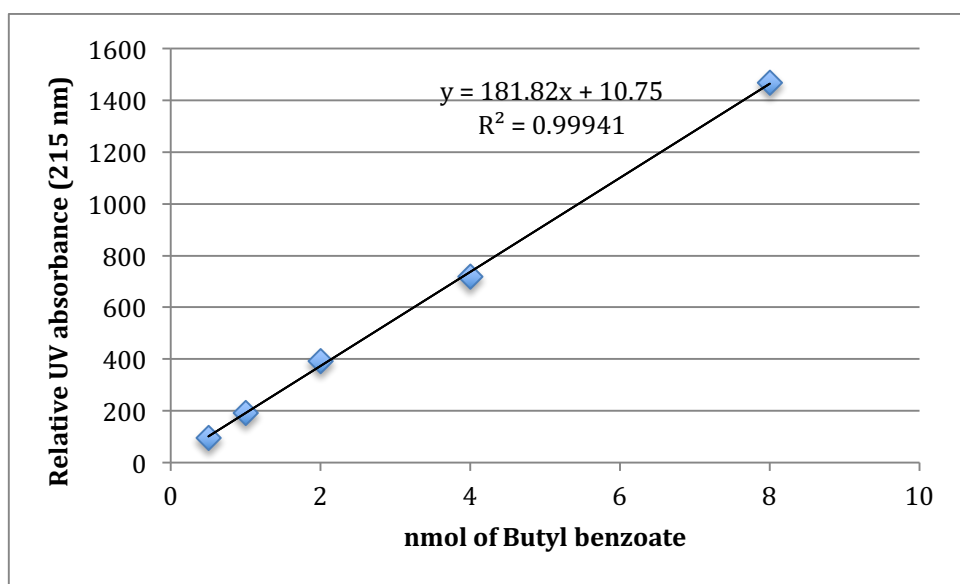
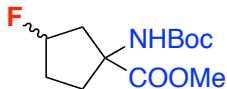
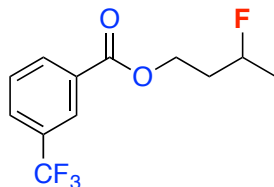


Figure S2-1. Standard curve of UV absorbance vs amount of fluorinated butyl benzoate

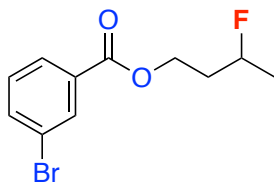
3. Preparation and characterization of the new ^{19}F authentic samples



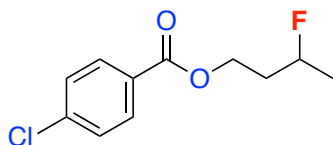
Compound 1. Synthesized according to published Mn-fluorination procedure. White solid ^1H NMR (300 MHz, CDCl_3) δ 5.13 (d, $J = 53.8$ Hz, 1H), 5.00 (d, $J = 56.5$ Hz, 1H), 3.74 (s, 3H), 2.80 - 1.83 (m, 6H), 1.43 (s, 9H). ^{13}C NMR (126 MHz, CDCl_3) δ 174.24, 155.18, 95.50, 80.08, 65.26, 52.70, 44.62, 35.58, 32.81, 28.41; ^{19}F NMR (282 MHz, CDCl_3) δ 167.9 (major), 170.9 (minor) HRMS (ESI) m/z cal'd $\text{C}_{12}\text{H}_{20}\text{FNNaO}_4$ $[\text{M}+\text{Na}]^+$: 284.1274, found 284.1290.



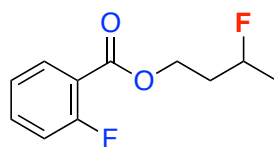
Compound 3. Synthesized according published procedure. colorless oil. ^1H NMR (500 MHz, CDCl_3) δ 8.29 (tt, $J = 1.6, 0.7$ Hz, 1H), 8.26 - 8.19 (m, 1H), 7.85 - 7.79 (m, 1H), 7.59 (tt, $J = 7.7, 0.8$ Hz, 1H), 4.98 - 4.78 (m, 1H), 4.58 - 4.44 (m, 2H), 2.19 - 1.93 (m, 2H), 1.42 (dd, $J = 23.9, 6.2$ Hz, 3H). ^{13}C NMR (126 MHz, CDCl_3) δ 165.38, 132.94, 131.11, 129.68, 129.20, 126.70, 124.92, 122.77, 87.87 (d, $J = 160$ Hz), 61.88, 36.18, 21.35. ^{19}F NMR (282 MHz, CDCl_3) δ 175.75; MS (EI) m/z cal'd $\text{C}_{12}\text{H}_{12}\text{F}_4\text{O}_2$ $[\text{M}]^+$ 264.1, found 264.1



Compound 4. Synthesized according published procedure. Colorless oil. Contaminated with 10% non-fluorinated precursor. ^1H NMR (501 MHz, CDCl_3) δ 8.17 (m, 1H), 7.97 (ddd, $J = 7.8, 1.6, 1.1$ Hz, 1H), 7.68 (m, 1H), 7.32 (td, $J = 7.9, 3.1$ Hz, 1H), 4.87 (dm, $J = 49.7$ Hz, 1H), 4.53 - 4.39 (m, 2H), 2.18 - 1.92 (m, 2H), 1.42 (dd, $J = 23.9, 6.2$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.14, 135.96, 132.56, 132.08, 129.92, 128.17, 122.48, 87.77 (d, $J = 165$ Hz) 61.59, 36.01, 21.14; ^{19}F NMR (282 MHz, CDCl_3) δ -176.31; MS (EI) m/z cal'd $\text{C}_{11}\text{H}_{12}\text{BrFO}_2$ $[\text{M}]^+$ 274.0, found 274.0;

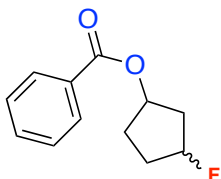


Compound 5. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, CDCl_3) δ 7.97 (d, $J = 8.3$ Hz, 2H) 7.41 (d, $J = 8.5$ Hz, 2H), 4.87 (d, $J = 48.6$ Hz, 1H), 4.46 (m, 2H), 2.23 - 1.85 (m, 2H), 1.41 (dd, $J = 23.9, 6.0$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 165.85, 139.70, 131.18, 128.99, 128.82, 88.05 (d, $J = 166$ Hz), 61.63, 36.25, 21.37; ^{19}F NMR (282 MHz, CDCl_3) δ -175.63; MS (EI) m/z cal'd $\text{C}_{11}\text{H}_{12}\text{ClFO}_2$ $[\text{M}]^+$ 230.1, found 230.1;

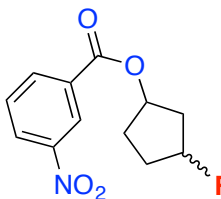


Compound 6. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, CDCl_3) δ 7.93 (td, $J = 7.5, 1.8$ Hz, 1H), 7.52 (dddd, $J = 8.4, 7.4, 4.8, 1.9$ Hz, 1H),

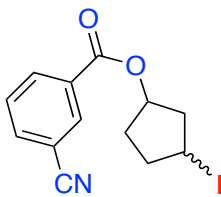
7.16 (m, 2H), 4.89 (dm, $J = 49.6$ Hz, 1H), 4.46 (m, 2H), 2.20 – 1.89 (m, 2H), 1.41 (dd, $J = 23.9, 6.2$ Hz, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 163.67 (d, $J = 174$ Hz), 160.92, 134.55, 132.13, 123.99, 118.67, 117.01, 87.80 (d, $J = 165$ Hz), 61.45, 35.97, 21.11; ^{19}F NMR (282 MHz, CDCl_3) δ -109.32, -175.89; MS (EI) m/z cal'd $\text{C}_{11}\text{H}_{12}\text{F}_2\text{O}_2$ $[\text{M}]^+$ 214.1, found 214.1;



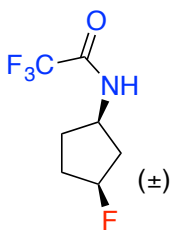
Compound 7. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, CD_3CN) δ 8.12 – 7.93 (m, 2H), 7.69 – 7.57 (m, 1H), 7.57 – 7.44 (m, 2H), 5.39 (m, 1H), 5.22 (dddt, $J = 54.9, 6.2, 3.2, 1.7$ Hz, 1H), 2.32 – 1.83 (m, 6H); ^{13}C NMR (126 MHz, CD_3CN) δ 167.07, 134.05, 131.67, 130.25, 129.58, 96.29 (d, $J = 172$ Hz), 76.79, 40.76, 32.76, 31.42; ^{19}F NMR (282 MHz, CD_3CN) δ -169.83; MS (EI) m/z cal'd $\text{C}_{12}\text{H}_{13}\text{FO}_2$ $[\text{M}]^+$ 208.1, found 208.1;



Compound 8. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, Chloroform- d) δ 8.79 (s, 1H), 8.41 (d, $J = 8.1$ Hz, 1H), 8.34 (d, $J = 7.7$ Hz, 1H), 7.65 (t, $J = 8.0$ Hz, 1H), 5.61 (m, 1H), 5.31 (d, $J = 52.1$, 1H), 2.76 – 1.64 (m, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.06, 148.25, 135.28, 132.09, 129.65, 127.44, 124.44, 95.37, (d, $J = 168$ Hz), 40.86, 32.78, 31.82, 30.40; ^{19}F NMR (282 MHz, CDCl_3) δ -171.47; MS (EI) m/z cal'd $\text{C}_{12}\text{H}_{12}\text{FNO}_4$ $[\text{M}]^+$ 253.1, found 253.1;

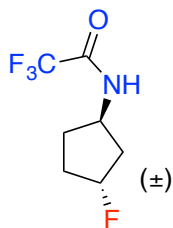


Compound 9. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, Chloroform- d) δ 8.27 (s, 1H), 8.23 (d, $J = 8.0$ Hz, 1H), 7.84 (d, $J = 8.0$ Hz, 1H), 7.58 (t, $J = 7.8$ Hz, 1H), 5.59 (tdd, $J = 6.4, 3.7, 2.4$ Hz, 1H), 5.23 (dt, $J = 54.0, 4.8$ Hz, 1H), 2.51 (dddt, $J = 26.6, 15.9, 6.8, 1.8$ Hz, 1H), 2.33 - 1.92 (m, 5H); ^{13}C NMR (126 MHz, CDCl_3) δ 164.37, 136.21, 133.85, 133.41, 131.84, 129.68, 118.12, 113.17, 95.18 (d, $J = 172$ Hz), 77.08, 41.11, 31.55, 30.63; ^{19}F NMR (282 MHz, CDCl_3) δ -171.44; MS (EI) m/z cal'd $\text{C}_{13}\text{H}_{12}\text{FNO}_2$ $[\text{M}+\text{Na}]^+$ 233.1, found 233.1;

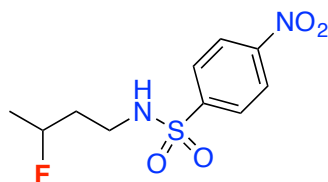


Compound 10a. Synthesized according published procedure. White solid. ^1H NMR (500 MHz, Chloroform- d) δ 6.61 (br, 1H), 5.52 (dm, $J = 53.1$ Hz, 1H), 4.54 (m, 1H), 2.29-1.71 (m, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 156.22, 115.77 (q, $J = 287.8$ Hz), 96.61 (d, $J = 169.1$ Hz), 49.91, 40.22, 32.08, 31.03; ^{19}F NMR (282 MHz, CDCl_3) δ -76.01, -168.03; MS(EI) m/z cal'd $\text{C}_7\text{H}_9\text{F}_4\text{NO}$

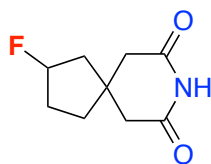
[M]⁺ 199.1, found 199.1;



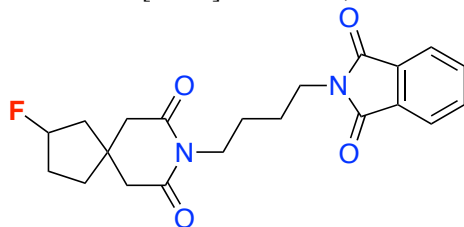
Compound 10b. Synthesized according published procedure. White solid. ¹H NMR (500 MHz, Chloroform-*d*) δ 6.24 (br, 1H), 5.21 (dm, *J* = 53.7 Hz, 1H), 4.49, (m, 1H), 2.58 – 1.50 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 156.86, (q, *J* = 283.9 Hz), 94.45 (d, *J* = 173.3 Hz); 50.58, 40.35, 31.68, 30.14; ¹⁹F NMR (282 MHz, CDCl₃) δ -76.01, -170.11; MS(EI) *m/z* cal'd C₇H₉F₄NO [M]⁺ 199.1, found 199.1;



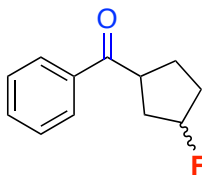
Compound 11. Synthesized by treating the corresponding alcohol with DAST. Yellow solid ¹H NMR (501 MHz, CDCl₃) δ 8.45 – 8.31 (m, 2H), 8.11 – 7.99 (m, 2H), 4.91 – 4.81 (br, 1H), 4.73 (dm, *J* = 47.6 Hz, 1H), 3.32 – 3.03 (m, 2H), 1.91 – 1.71 (m, 2H), 1.33 (dd, *J* = 24.3, 6.2 Hz, 3H); ¹³C NMR (126 MHz, CDCl₃) δ 150.10, 145.81, 128.31, 124.46, 89.69 (d, *J* = 164 Hz), 40.28, 36.48, 20.93; ¹⁹F NMR (282 MHz, CDCl₃) -174.99; HRMS (ESI) *m/z* cal'd C₁₀H₁₄FN₂O₄S [M+H]⁺ 277.0658, found 277.0654;



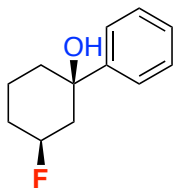
Compound 12. Synthesized according published procedure. White solid. ¹H NMR (501 MHz, CDCl₃) δ 7.83 (br, 1H), 5.21 (dt, *J* = 53.0, 4.2 Hz, 1H), 2.74 (m, 2H), 2.57 (m, 2H), 2.24 – 1.63 (m, 6H); ¹³C NMR (126 MHz, CDCl₃) δ 171.44, 171.34, 95.86 (d, *J* = 173 Hz), 45.38, 45.21, 44.46, 39.95, 35.45, 32.65; ¹⁹F NMR (282 MHz, CDCl₃) δ -167.79; HRMS (ESI) *m/z* cal'd C₉H₁₃FNO₂ [M+H]⁺ 186.0930, found 186.0936;



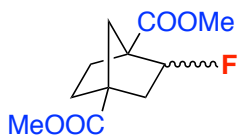
Compound 13. Synthesized by treating compound 12 with corresponding alkyl bromide. White solid. ¹H NMR (501 MHz, CDCl₃) δ 7.82 (dd, *J* = 5.5, 3.0 Hz, 2H), 7.70 (dd, *J* = 5.5, 3.1 Hz, 2H), 5.15 (dtt, *J* = 53.3, 4.8, 1.6 Hz, 1H), 3.79 (t, *J* = 7.0 Hz, 2H), 3.68 (t, *J* = 6.9 Hz, 2H), 2.82 – 2.48 (m, 4H), 2.20 – 1.45 (m, 10H); ¹³C NMR (126 MHz, CDCl₃) δ 171.63, 171.59, 168.36, 133.91, 132.10, 123.21, 95.89 (d, *J* = 173 Hz), 45.44, 45.24, 45.07, 39.00, 38.74, 37.57, 35.23, 32.61, 26.07, 25.32; ¹⁹F NMR (282 MHz, CDCl₃) -167.88; HRMS (ESI) *m/z* cal'd C₂₁H₂₄FN₂O₄ [M+H]⁺ 387.1720, found 387.1722;



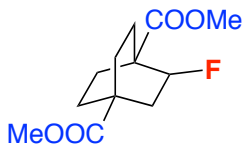
Compound 14. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, CDCl_3) δ 8.02 – 7.96 (m, 2H), 7.61 – 7.54 (m, 1H), 7.48 (t, J = 7.6 Hz, 2H), 5.29 (d, J = 52.6 Hz, 1H), 4.04 (qd, J = 8.6, 5.6 Hz, 1H), 2.31 – 2.14 (m, 3H), 2.12 – 1.98 (m, 1H), 1.99 – 1.78 (m, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 201.52, 136.36, 133.14, 128.68, 128.53, 96.92 (d, J = 171 Hz), 44.02, 36.76, 32.93, 27.79; ^{19}F NMR (282 MHz, CDCl_3) -170.59; MS (EI) m/z cal'd $\text{C}_{12}\text{H}_{13}\text{FO}$ $[\text{M}]^+$ 192.1, found 192.1;



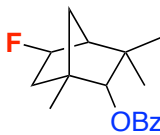
Compound 15. Synthesized according published procedure. White solid; ^1H NMR (501 MHz, CDCl_3) δ 7.61 – 7.50 (m, 2H), 7.44 – 7.34 (m, 2H), 7.32 – 7.24 (m, 1H), 5.11 (dp, J = 47.8, 3.3 Hz, 1H), 3.08 (br, 1H), 2.38 – 1.87 (m, 6H), 1.81 – 1.49 (m, 2H) *Axial fluoride configuration is evidenced by the splitting of F-C-H (doublet of pentet)*; ^{13}C NMR (126 MHz, CDCl_3) δ 147.59, 128.26, 126.87, 124.61, 91.54 (d, J = 164 Hz), 72.82, 42.47, 38.15, 29.92, 16.34; ^{19}F NMR (282 MHz, CDCl_3) -178.64; HRMS (ESI) m/z cal'd $\text{C}_{12}\text{H}_{16}\text{FO}$ $[\text{M}+\text{H}]^+$ 195.1185, found 195.1199;



Compound 16. Synthesized according published procedure. White solid. ^1H NMR (500 MHz, CDCl_3) δ 4.96 – 4.76 (m, 1H), 3.73 (s, 3H), 3.71 (s, 3H), 2.27 – 1.85 (m, 6H), 1.56 – 1.48 (m, 2H); ^{13}C NMR (126 MHz, CDCl_3) δ 173.91, 172.19, 95.14 (d, J = 189 Hz), 57.37, 52.00, 50.26, 42.14, 39.93, 32.09, 31.60, 27.65; ^{19}F NMR (282 MHz, CDCl_3) δ 164.02 (major), 157.77 (minor). MS (EI) m/z cal'd $\text{C}_{11}\text{H}_{15}\text{FO}_4$ $[\text{M}]^+$ 230.1, found 230.1

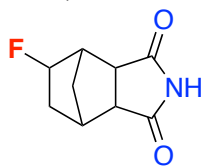


Compound 17. Synthesized according published procedure. Colorless oil; ^1H NMR (500 MHz, CDCl_3) δ 5.16 (ddt, J = 53, 8.8, 1.6 Hz, 1H), 3.70 (s, 3H), 3.66 (s, 3H), 2.34 – 1.61 (m, 10H); ^{13}C NMR (126 MHz, CDCl_3) δ 176.48, 175.20, 90.66 (d, J = 155 Hz), 52.15, 51.99, 27.75, 27.33, 26.72, 26.07, 26.02, 20.31, 20.27; ^{19}F NMR (282 MHz, CDCl_3) δ 173.17; MS (EI) m/z cal'd $\text{C}_{12}\text{H}_{17}\text{FO}_4$ $[\text{M}]^+$ 243.1, found 243.1

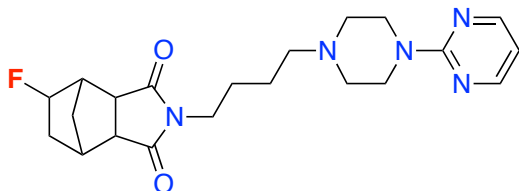


Compound 18. Synthesized according published procedure. Colorless oil; ^1H NMR (501 MHz, CDCl_3) δ 8.14 – 7.90 (m, 2H), 7.68 – 7.52 (m, 1H), 7.48 (m, 2H), 5.04 (dd, J = 55.2, 4.9 Hz, 1H), 4.61 (d, J = 1.7 Hz, 1H), 2.46 (dddd, J = 22.7, 14.6, 6.2, 2.4 Hz, 1H), 2.09 (dt, J = 8.1, 1.5 Hz, 1H), 1.77 – 1.31 (m, 3H), 1.26 (s, 3H), 1.17 (s, 3H), 0.82 (s, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 166.99, 133.15, 130.02, 129.49, 128.48, 94.18 (d, J = 177 Hz), 85.05, 53.59, 47.66, 39.06, 37.56, 37.08, 29.45, 19.39, 18.47; ^{19}F NMR (282 MHz, CDCl_3) -167.61; MS (EI) m/z cal'd $\text{C}_{17}\text{H}_{21}\text{FO}_2$ $[\text{M}]^+$

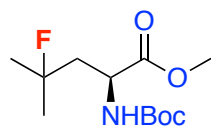
276.1, found 276.1;



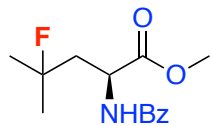
Compound 19. Synthesized according published procedure. White solid; ^1H NMR (501 MHz, CD_3CN) δ 4.76 (ddq, $J = 54.8, 6.3, 1.3$ Hz, 1H), 2.72 (d, $J = 8.4$ Hz, 1H), 2.61 (d, $J = 4.6$ Hz, 1H), 2.53 (s, 2H), 1.89 – 1.53 (m, 4H), 1.36 – 1.23 (m, 1H); ^1H NMR (501 MHz, CDCl_3) δ 178.04, 177.55, 93.36 (d, $J = 186$ Hz), 49.22, 45.20, 44.43, 38.61, 38.15, 29.91; ^{19}F NMR (282 MHz, CD_3CN) -165.96; HRMS (ESI) m/z cal'd $\text{C}_9\text{H}_{11}\text{FNO}_2$ $[\text{M}+\text{H}]^+$ 184.0774, found 184.0779;



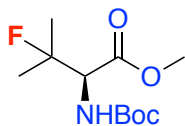
Compound 20. Synthesized by treating compound 19 with corresponding alkyl bromide. White solid; ^1H NMR (501 MHz, CDCl_3) δ 8.30 (d, $J = 4.8$ Hz, 2H), 6.47 (t, $J = 4.7$ Hz, 1H), 4.71 (d, $J = 53.7$ Hz, 1H), 3.82 (t, $J = 5.1$ Hz, 4H), 3.51 (t, $J = 7.0$ Hz, 2H), 2.92 (d, $J = 8.0$ Hz, 1H), 2.78 (s, 1H), 2.49 (m, 6H), 2.38 (t, $J = 7.2$ Hz, 2H), 1.92 – 1.35 (m, 6H), 1.25 (t, $J = 7.2$ Hz, 1H), 1.11 (d, $J = 11.7$ Hz, 1H); ^{13}C NMR (126 MHz, CDCl_3) δ 177.77, 177.32, 161.64, 157.71, 109.81, 93.53 (d, $J = 186$ Hz), 58.05, 53.11, 47.78, 45.27, 43.65, 42.95, 38.85, 38.66, 38.19, 29.72, 25.83, 24.22; ^{19}F NMR (282 MHz, CDCl_3) δ -165.14; HRMS (ESI) m/z cal'd $\text{C}_{21}\text{H}_{29}\text{FN}_5\text{O}_2$ $[\text{M}+\text{H}]^+$ 402.2305, found 402.2312;



Compound 21a. Synthesized according published procedure. White solid; ^1H NMR (500 MHz, CDCl_3) δ 5.16 (br, 1H), 4.43 (t, $J = 6.9$ Hz, 1H), 3.77 (s, 3H), 2.25 - 1.92 (m, 3H), 1.57 (d, $J = 9.2$ Hz, 1H), 1.41 (s, 5H), 0.96 (ddt, $J = 31.5, 16.3, 7.1$ Hz, 1H). ^{13}C NMR (126 MHz, CDCl_3) δ 173.38, 155.49, 95.26 (d, $J = 179.5$ Hz), 80.28, 52.67, 51.06, 42.84, 28.55, 27.43, 26.78; ^{19}F NMR (282 MHz, CDCl_3) δ -136.62; HRMS (ESI) m/z cal'd $\text{C}_{12}\text{H}_{22}\text{FNNaO}_4$ $[\text{M}+\text{Na}]^+$ 286.1431, found 286.1420.

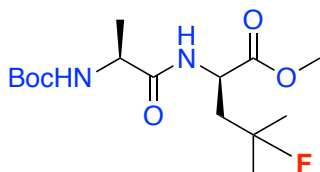


Compound 21b. Synthesized according published procedure. White solid. ^1H NMR (300 MHz, $\text{Acetone-}d_6$) δ 8.04 - 7.86 (m, 2H), 7.64 - 7.39 (m, 3H), 4.92 - 4.82 (m, 1H), 3.71 (s, 3H), 2.43 - 2.17 (m, 2H), 1.45 (dd, $J = 21.3, 7.3$ Hz, 6H), (N-H signal not observed). ^{13}C NMR (126 MHz, CDCl_3) δ 172.41, 165.95, 134.26, 131.42, 128.29, 127.20, 94.42 (d, $J = 173$ Hz), 51.53, 49.52, 41.67, 26.33, 25.96; ^{19}F NMR (282 MHz, CDCl_3) δ 136.77; HRMS (ESI) m/z cal'd $\text{C}_{14}\text{H}_{18}\text{FNNaO}_3$ $[\text{M}+\text{Na}]^+$ 290.1168, found 290.1163

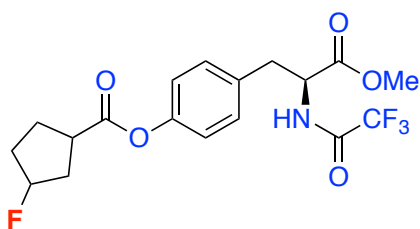


Compound 22. Synthesized according published procedure. White solid. Synthesized according published procedure ^1H NMR (501 MHz, CDCl_3) δ 5.35 (br, 1H), 4.39 (dd, $J = 20.4, 9.6$ Hz, 1H), 3.78 (s, 3H), 1.45 (s, 9H), 1.50 – 1.37 (dd, $J = 21.3, 10.3$ Hz 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 170.19, 155.40, 95.26, 80.30, 60.21, 52.37, 28.27, 24.60 (d, $J = 23.8$ Hz), 24.34 (d, $J =$

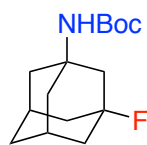
24.1 Hz); ^{19}F NMR (282 MHz, CD_3CN) δ -148.90; HRMS (ESI) m/z cal'd $\text{C}_{11}\text{H}_{21}\text{FNO}_4$ $[\text{M}+\text{H}]^+$ 250.2904, found 250.2917



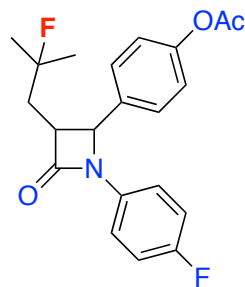
Compound 23. Synthesized according published procedure. White solid, contaminated with ~5% DCU; ^1H NMR (300 MHz, CDCl_3) δ 6.85 (s, 1H), 4.98 (s, 1H), 4.63 (ddd, J = 8.4, 7.1, 5.0 Hz, 1H), 4.14 (dq, J = 18.2, 7.1 Hz, 1H), 3.72 (s, 3H), 2.25 – 1.52 (m, 2H), 1.44 (s, 9H) 1.42 (s, 3H), 1.38 - 1.33(m, 6H). ^{13}C NMR (126 MHz, CDCl_3) δ 172.51, 172.37, 155.33, 95.82, 94.48, 52.52, 49.64, 41.97, 41.79, 28.20, 27.36, 26.03, 17.97; ^{19}F NMR (282 MHz, CDCl_3) δ 136.3 HRMS (ESI) m/z cal'd $\text{C}_{15}\text{H}_{27}\text{FN}_2\text{NaO}_5$ $[\text{M}+\text{Na}]^+$: 357.1802, found 357.1810.



Compound 24. Synthesized according published procedure. White solid. ^1H NMR (501 MHz, CDCl_3) δ 7.14 – 6.99 (m, 4H), 6.88 – 6.75 (m, 1H), 5.25 (dt, J = 52.9, 3.4, 1H), 4.86 (dt, J = 7.6, 5.6 Hz, 1H), 3.78 (s, 3H), 3.44 – 3.23 (m, 1H), 3.23 – 3.12 (m, 2H), 2.49 – 1.85 (m, 6H); ^{13}C NMR (126 MHz, CDCl_3) δ 174.28, 170.24, 156.57, 150.16, 132.18, 130.19, 121.84, 115.53 (q, J = 287.7), 96.20 (d, J = 172 Hz), 53.52, 52.95, 41.71, 37.44, 36.63, 32.85, 27.46; ^{19}F NMR (282 MHz, CDCl_3) δ -75.91 (- CF_3), -171.04 (-F); HRMS (ESI) m/z cal'd $\text{C}_{18}\text{H}_{20}\text{F}_4\text{NO}_5$ $[\text{M}+\text{H}]^+$ 406.1278, found 406.1271;

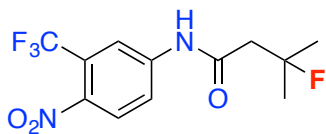


Compound 26. Synthesized according published procedure. White solid; ^1H NMR (501 MHz, CDCl_3) δ 4.46 (br, 1H), 2.33 (s, 2H), 2.10 (d, J = 5.9 Hz, 2H), 1.91 – 1.73 (m, 8H), 1.54 (d, J = 3.7 Hz, 2H), 1.43 (s, 9H); ^{13}C NMR (126 MHz, CDCl_3) δ 154.02, 92.54 (d, J = 185 Hz), 79.08, 53.67, 46.78, 41.66, 40.45, 34.72, 30.95, 28.45; ^{19}F NMR (282 MHz, CDCl_3) δ -132.67 HRMS (ESI) m/z cal'd $\text{C}_{15}\text{H}_{24}\text{FNNaO}_2$ $[\text{M}+\text{Na}]^+$ 292.1689, found 292.1678;



Compound 27. Synthesized according published procedure. Yellow oil; ^1H NMR (500 MHz, Chloroform- d) δ 7.50 – 7.29 (m, 2H), 7.31 – 7.16 (m, 2H), 7.10 (d, J = 8.6 Hz, 2H), 7.00 – 6.85 (m, 2H), 4.83 (t, J = 1.8 Hz, 1H), 3.27 (ddd, J = 10.6, 5.5, 2.2 Hz, 1H), 2.29 (s, 1H), 2.26 –

2.12 (m, 2H), 1.44 (d, $J = 21.3$ Hz, 3H), 1.27 (d, $J = 21.3$ Hz, 3H); ^{13}C NMR (126 MHz, CDCl_3) δ 169.27, 166.87, 159.04 (d, $J = 243.5$ Hz), 150.68, 134.86, 133.73, 127.23, 122.24, 118.46, 115.90, 95.13 (d, $J = 166.3$ Hz), 61.46, 40.08, 34.70, 28.54, 25.44, 21.18



Compound 28. Synthesized according published procedure. Yellow solid; ^1H NMR (500 MHz, Chloroform- d) δ 8.14 (d, $J = 8.4$ Hz, 0H), 8.01 (dd, $J = 7.7, 1.7$ Hz, 0H), 2.80 (d, $J = 23.1$ Hz, 0H), 1.55 (d, $J = 22.0$ Hz, 1H); ^{13}C NMR (126 MHz, Chloroform- d) δ 168.18, 141.95, 127.13, 125.90 – 124.58 (m), 122.14, 120.64, 118.27, 95.15 (d, $J = 165.4$ Hz), 49.72, 26.81; ^{19}F NMR (376 MHz, Chloroform- d) δ -60.52, -135.37 (dq, $J = 44.3, 22.3, 8.2$ Hz); HRMS (ESI) m/z cal'd $\text{C}_{12}\text{H}_{13}\text{F}_4\text{N}_2\text{O}_3$ $[\text{M}+\text{H}]^+$ 309.0862, found 309.0869;

4. Radio-HPLC characterization of the ^{18}F labeled products

All ^{18}F -labeled molecules were characterized by comparing the radio-HPLC trace of the crude reaction mixture to the HPLC UV trace of the authentic reference sample with methods detailed below.

HPLC column: Agilent Eclipse XDB-C18, 5 μm , 4.6 x 150 mm

Gradient: H_2O (0.1% TFA, A) and ACN (0.1% TFA, B), 5% B – 95% B, 20 min, 1.0 ml/min

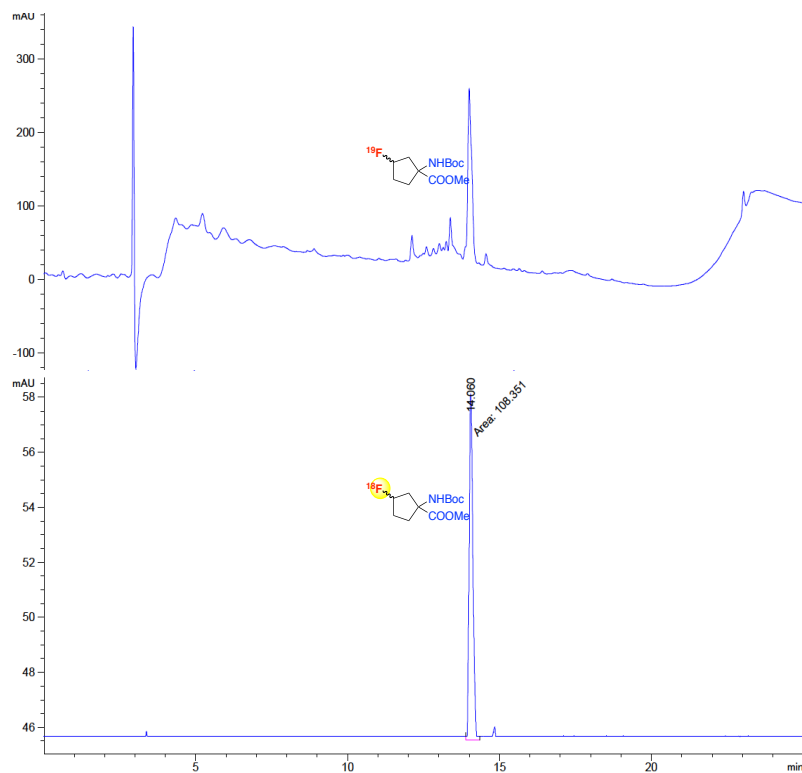


Figure S4a. UV and radio-HPLC traces of protected-FACPC

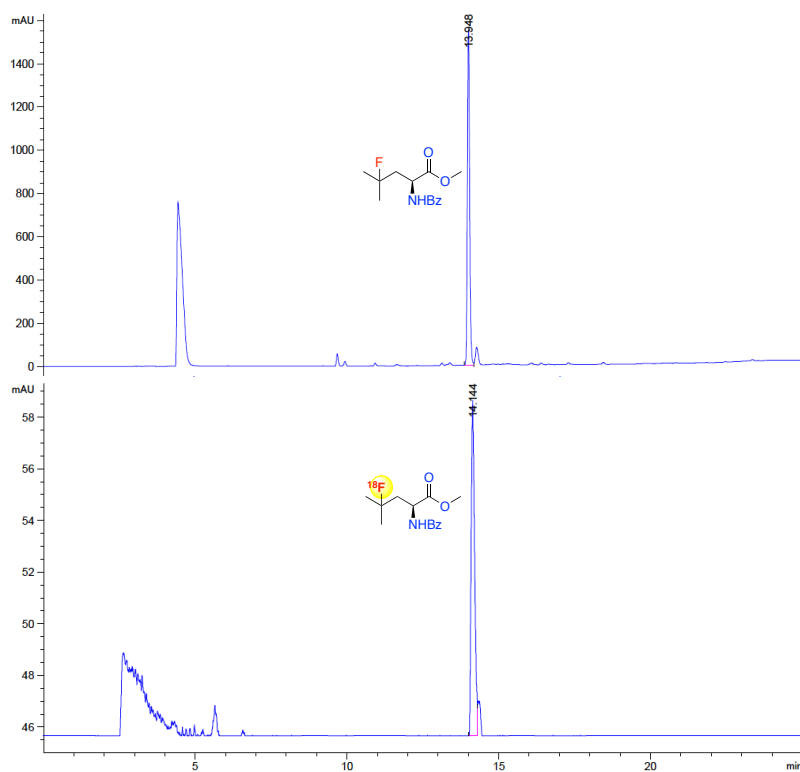


Figure S4b. UV and radio-HPLC traces of F-Leucine derivative

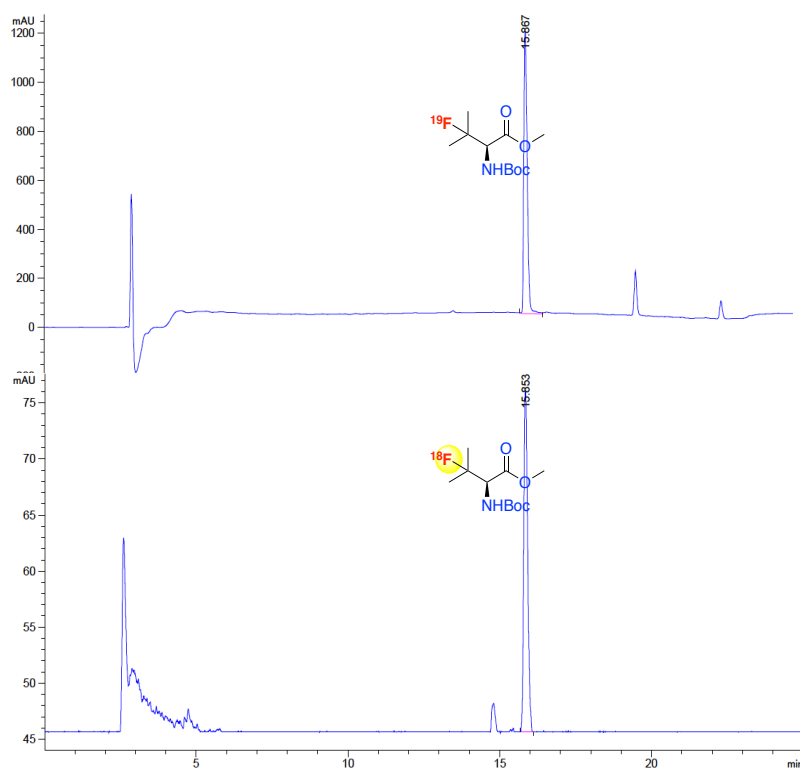


Figure S4c. UV and radio-HPLC traces of F-Valine derivative

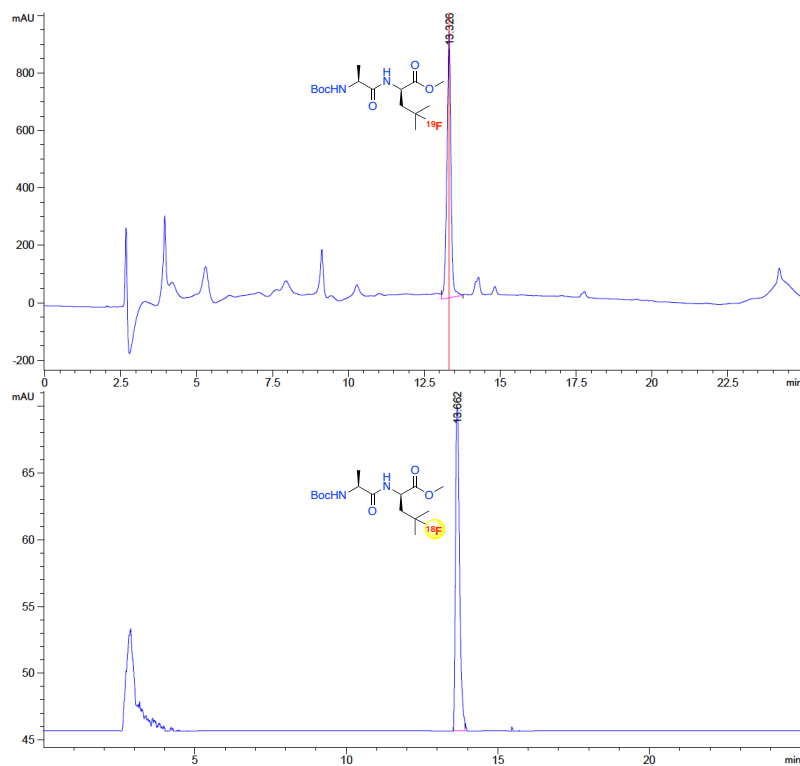


Figure S4d. UV and radio-HPLC traces of F-NBoc-Ala-Leu-OMe.

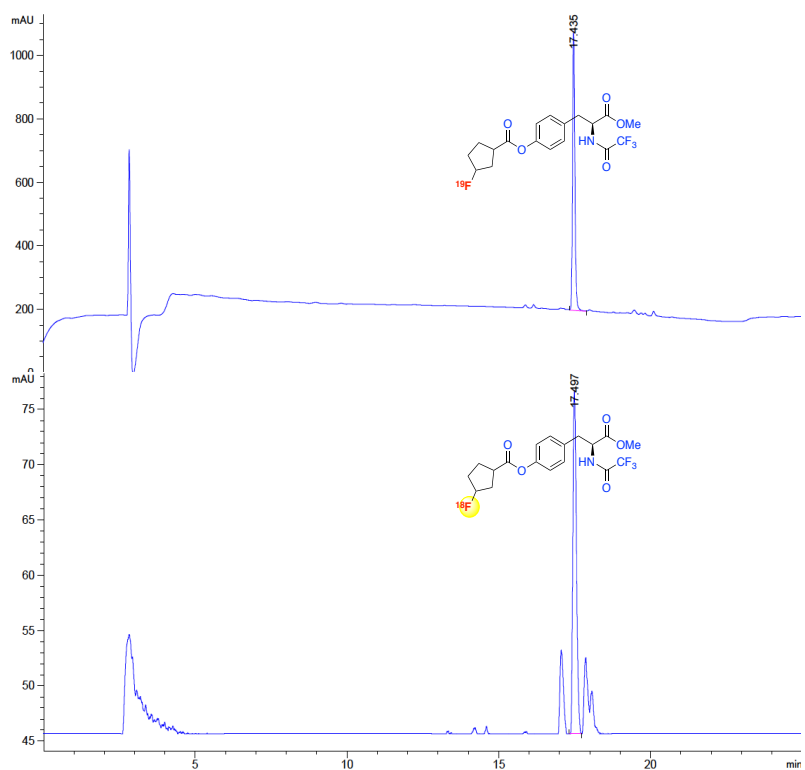


Figure S4e. UV and radio-HPLC traces F-tyrosine derivative.

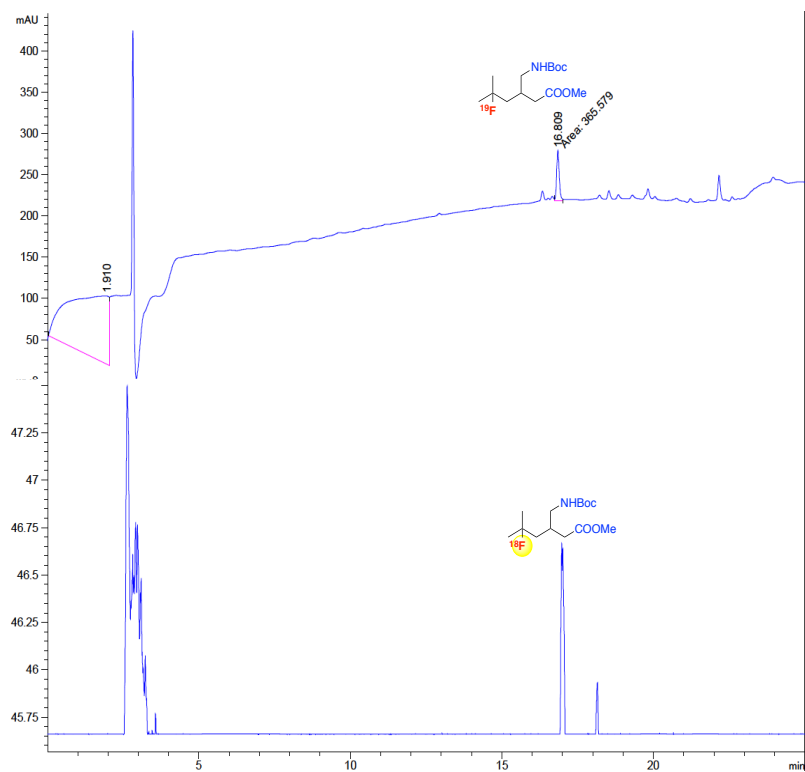


Figure S4f. UV and radio-HPLC traces F-pregabalin.

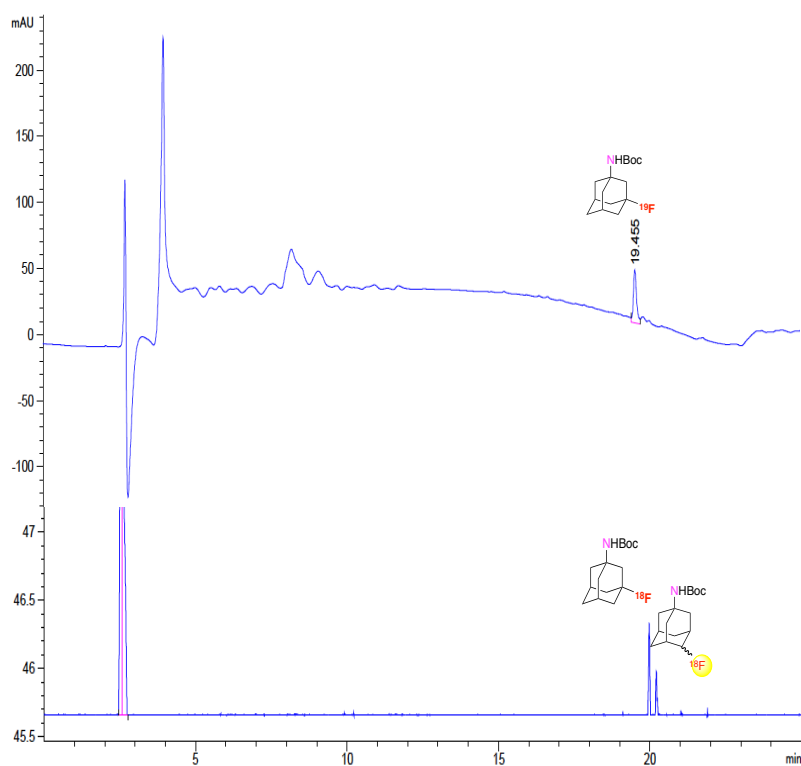


Figure S4g UV and radio-HPLC traces of F-N-Boc-amatadine.

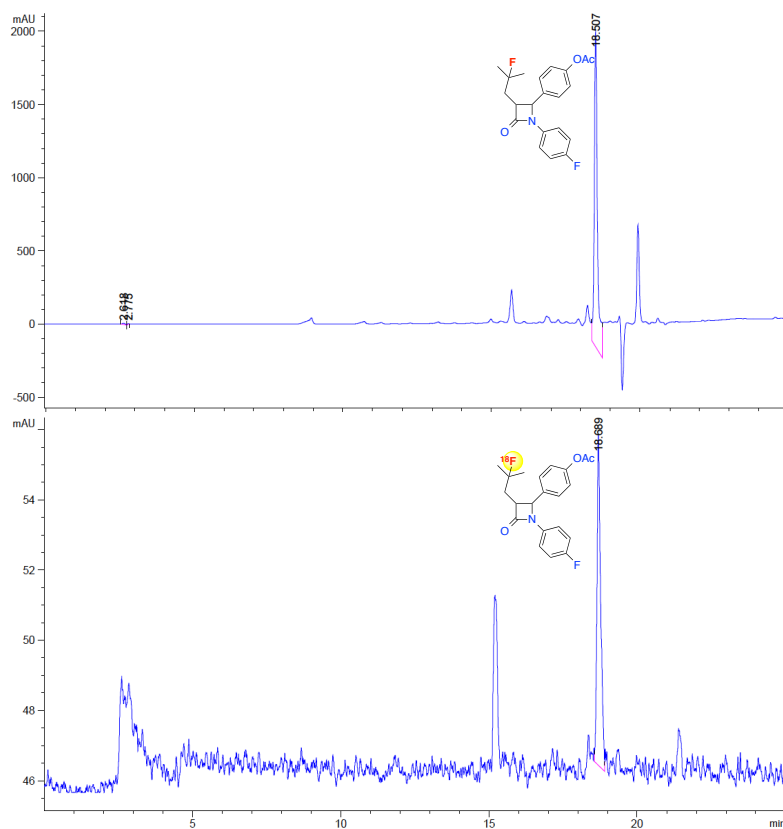


Figure S4h UV and radio-HPLC traces of ezetimibe analog.

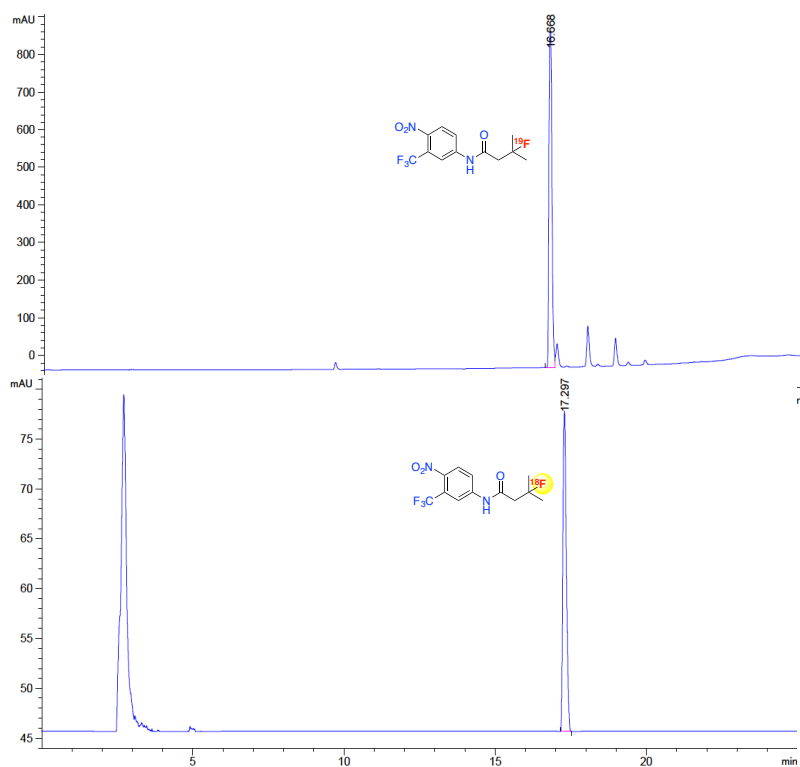


Figure S4h UV and radio-HPLC traces of F-flutamide analog.

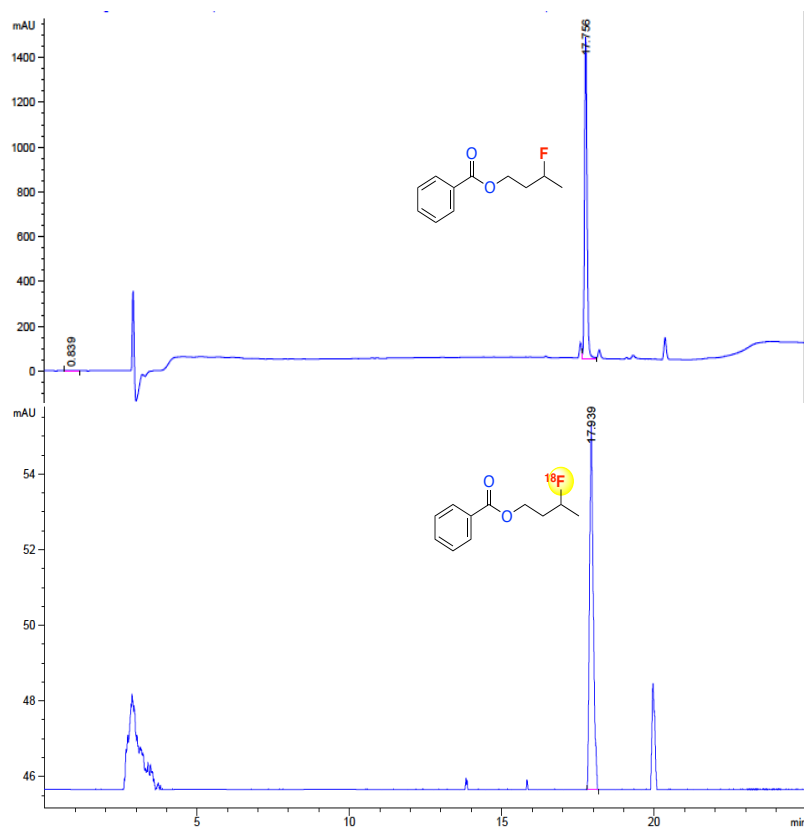


Figure S4i. UV and radio-HPLC traces of compound 2

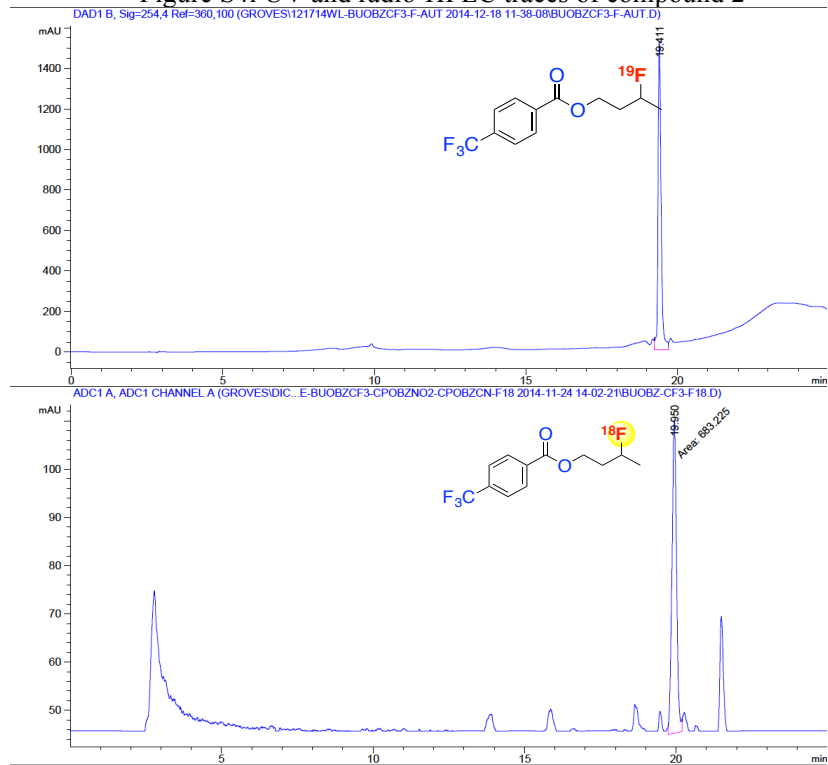


Figure S4j. UV and radio-HPLC traces of compound 3

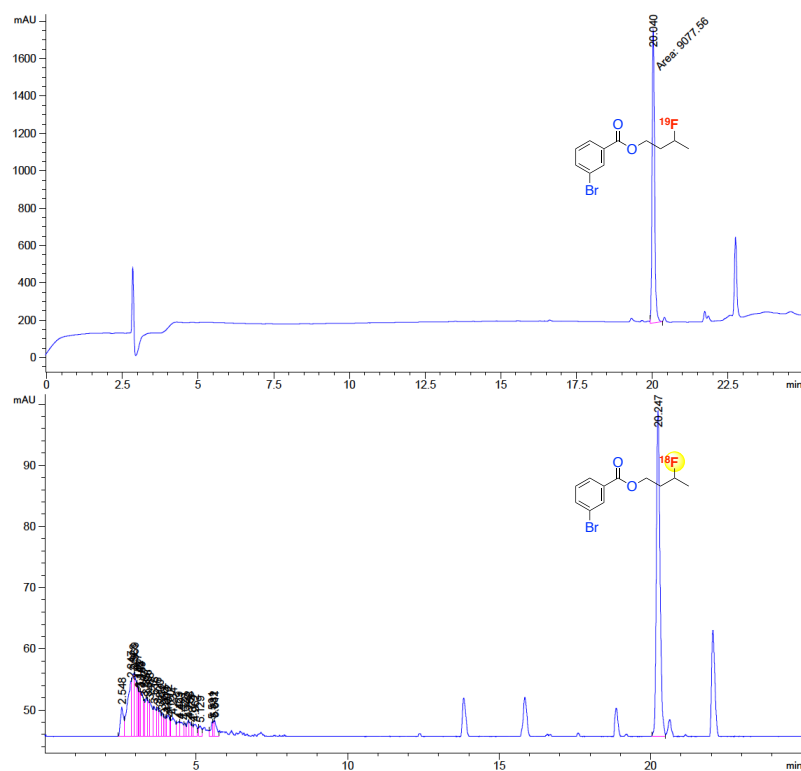


Figure S4k. UV and radio-HPLC traces of compound 4

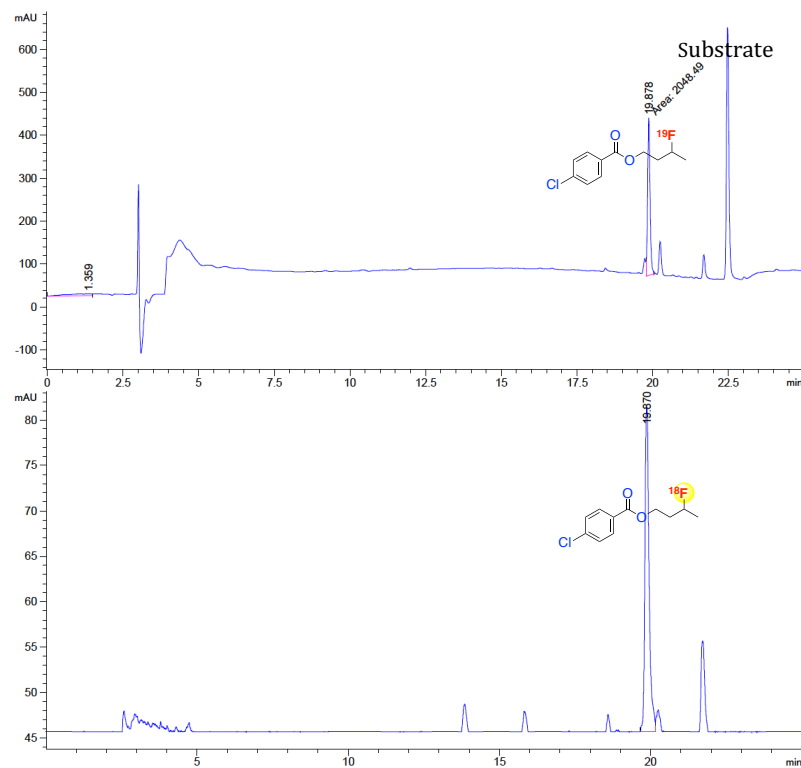


Figure S4l. UV and radio-HPLC traces of compound 5

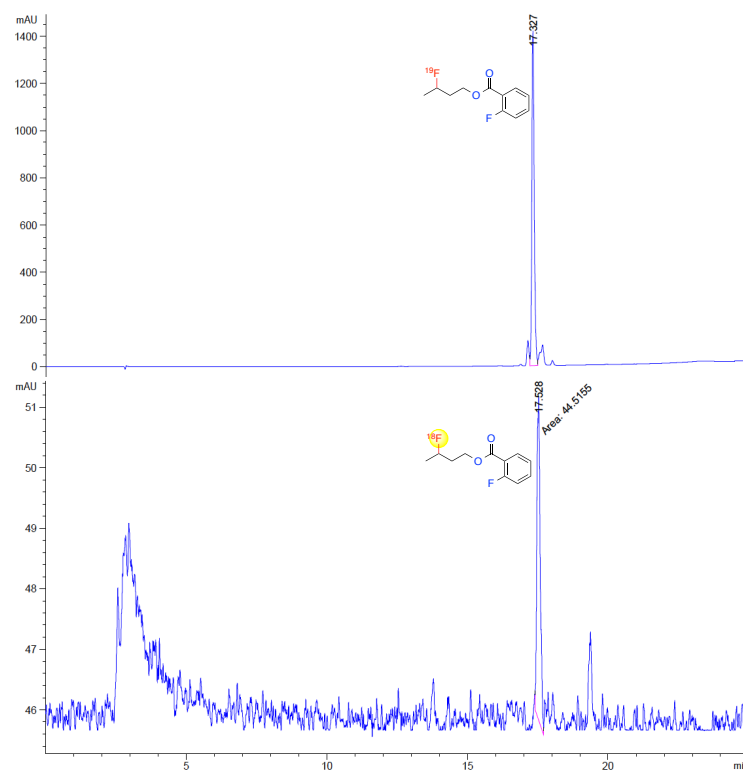


Figure S4m. UV and radio-HPLC traces of compound 6

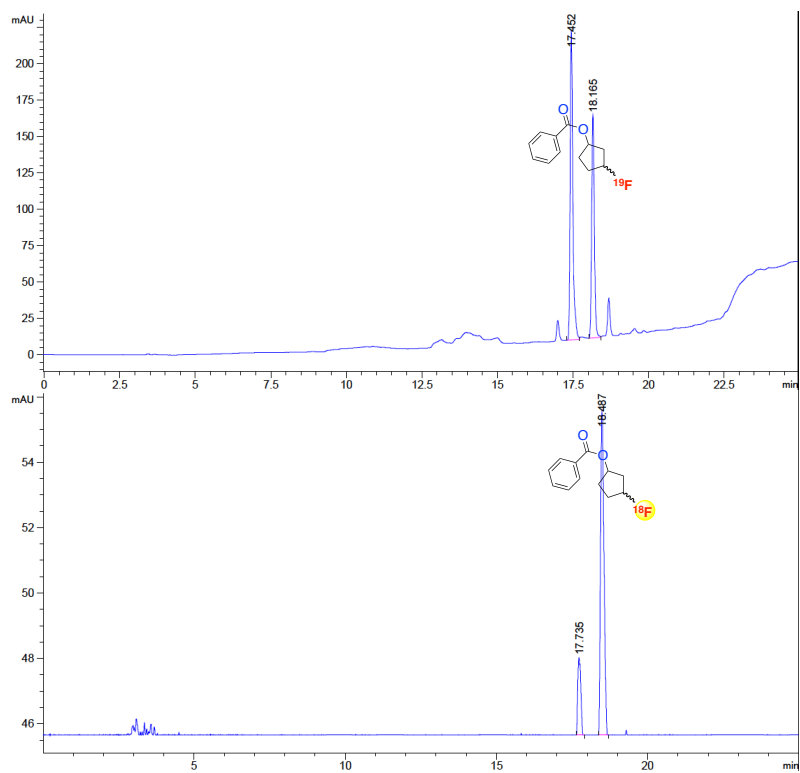


Figure S4m. UV and radio-HPLC traces of compound 7

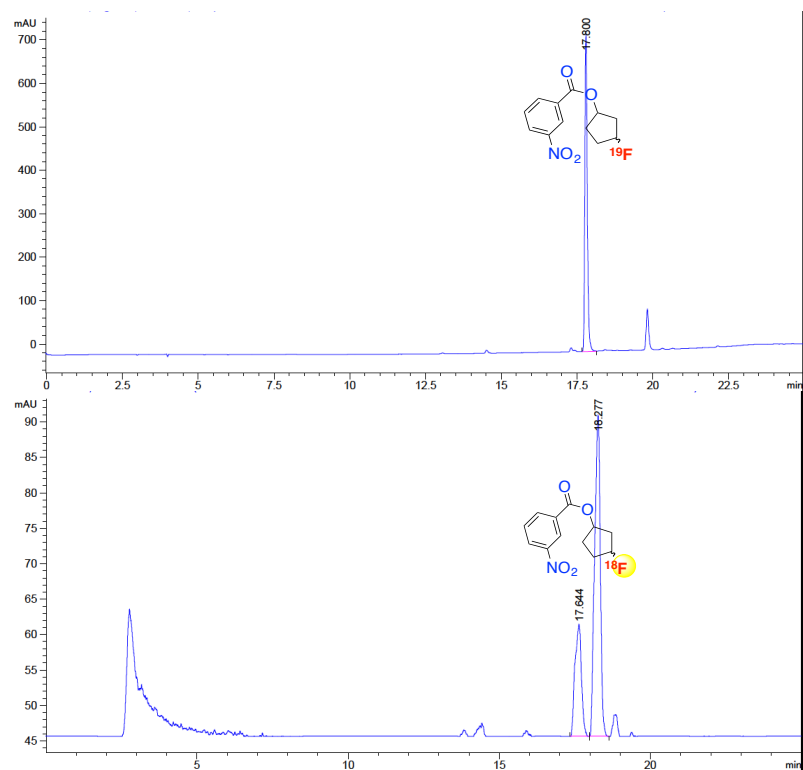


Figure S4n. UV and radio-HPLC traces of compound 8

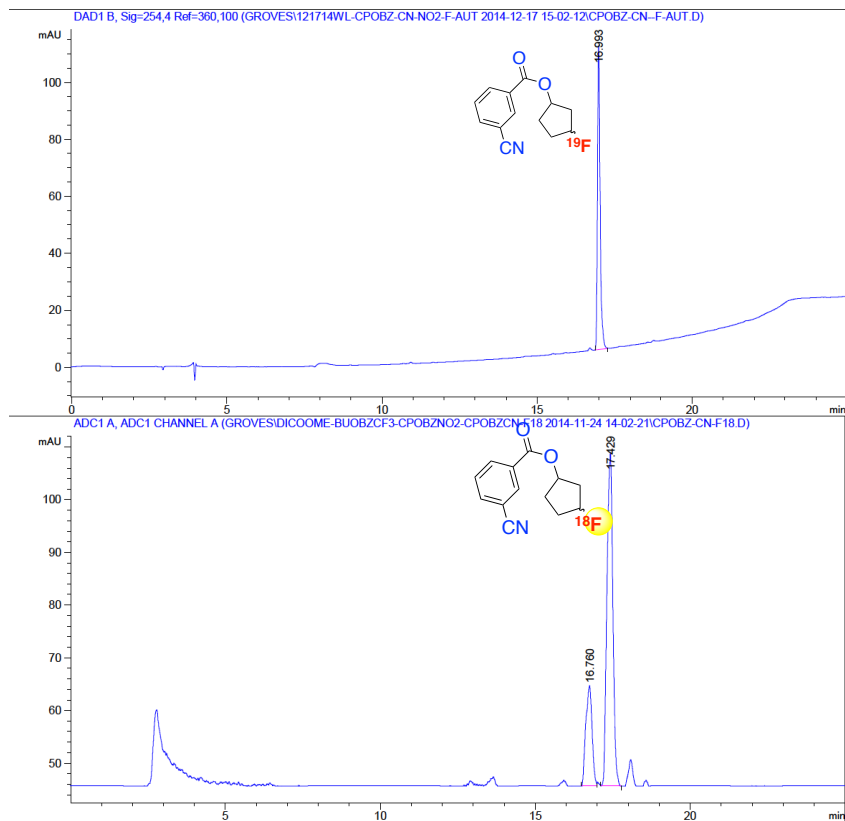


Figure S4o. UV and radio-HPLC traces of compound 9

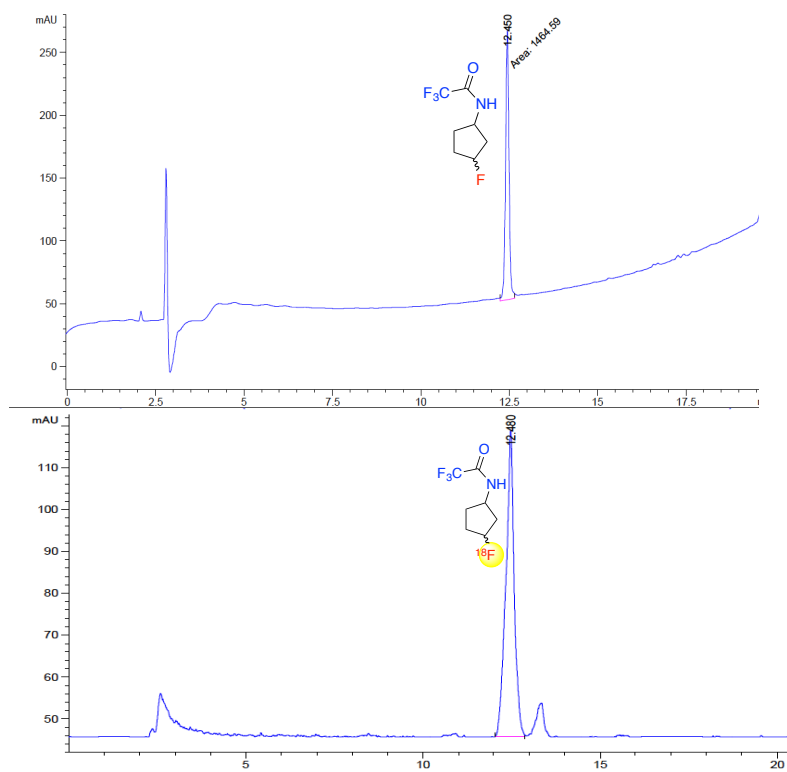


Figure S4p. UV and radio-HPLC traces of compound 10

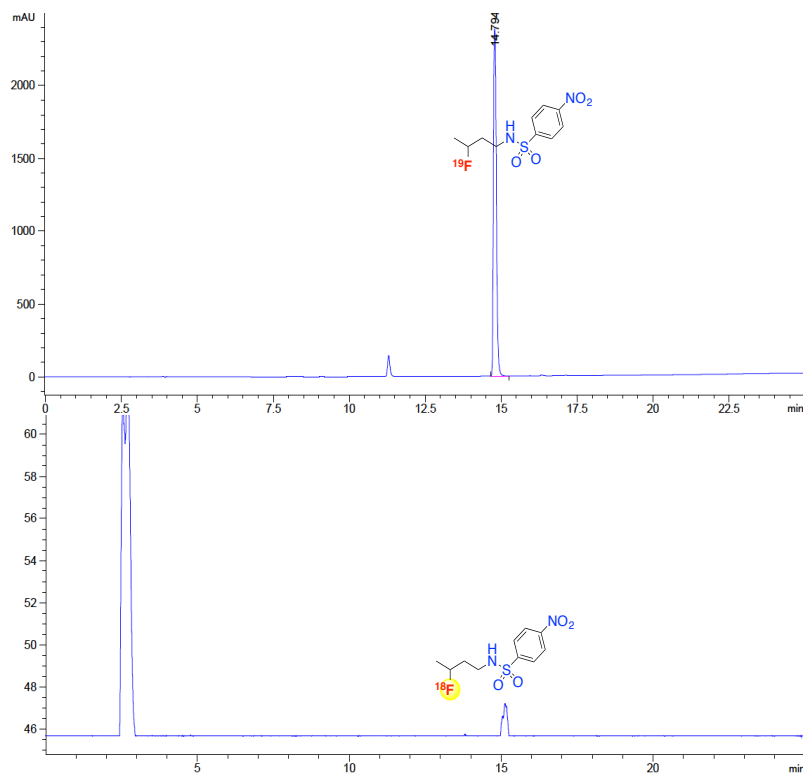


Figure S4q. UV and radio-HPLC traces of compound 11

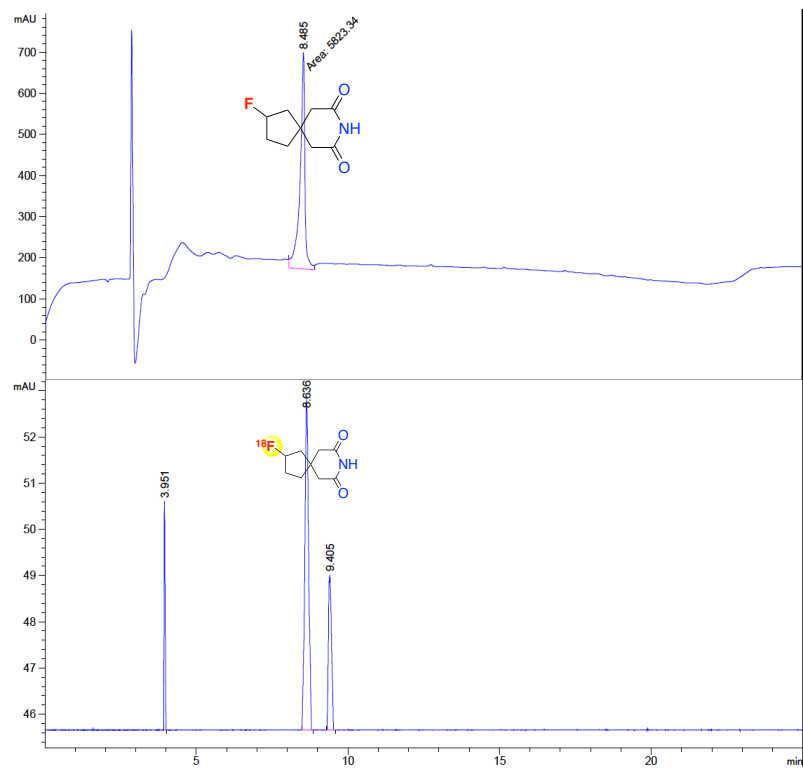


Figure S4r. UV and radio-HPLC traces of compound 12

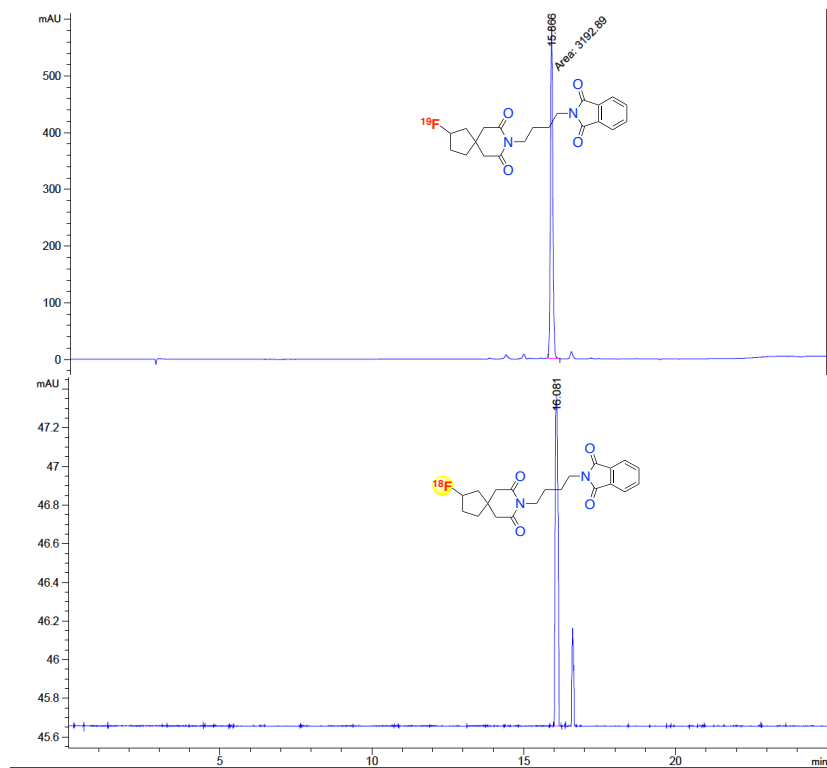


Figure S4s. UV and radio-HPLC traces of compound 13

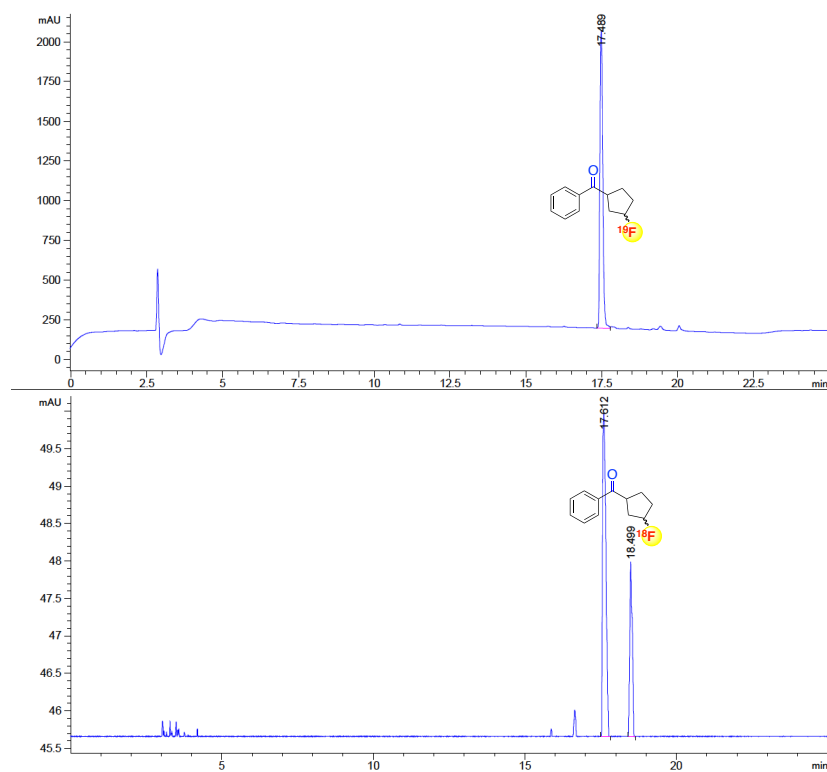


Figure S4t. UV and radio-HPLC traces of compound 14

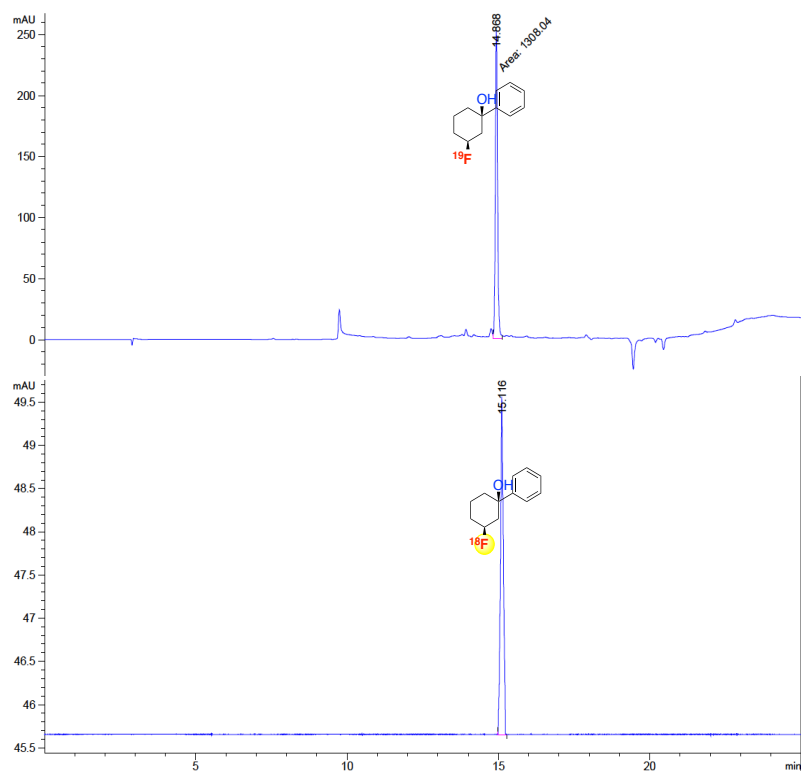
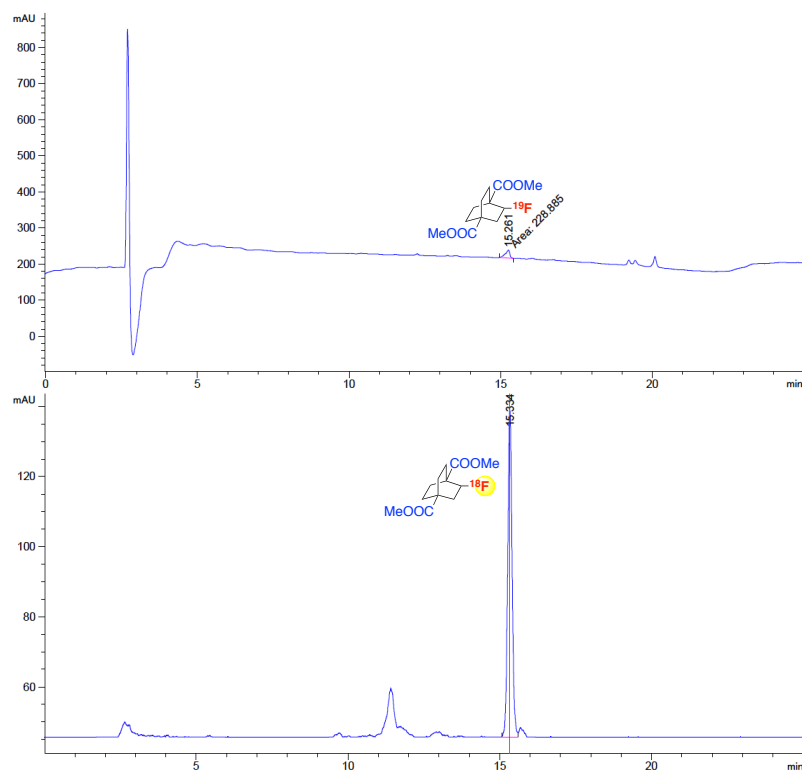
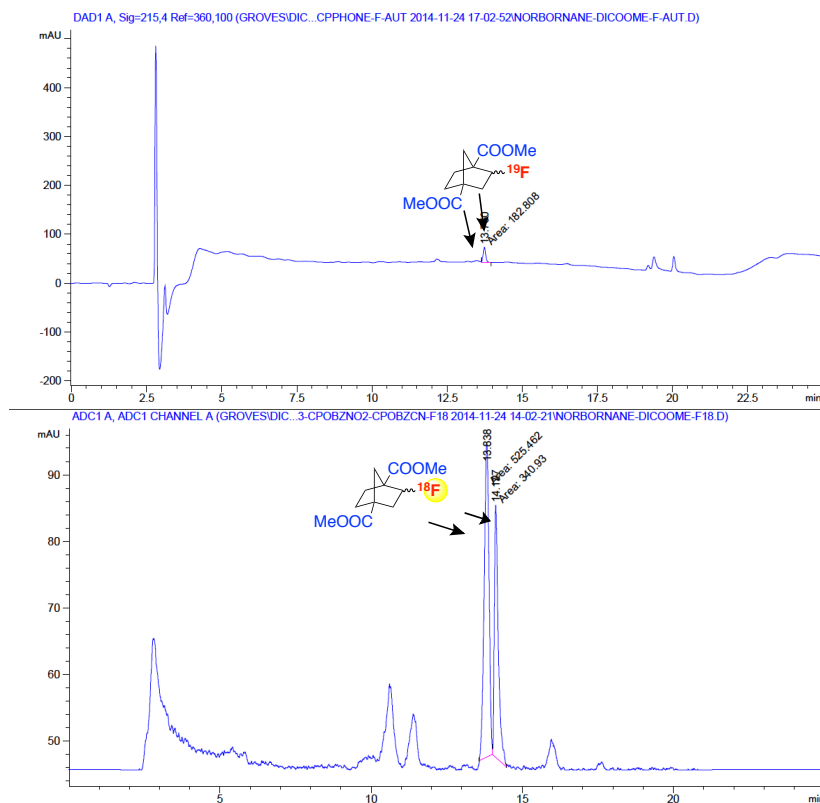


Figure S4u. UV and radio-HPLC traces of compound 15



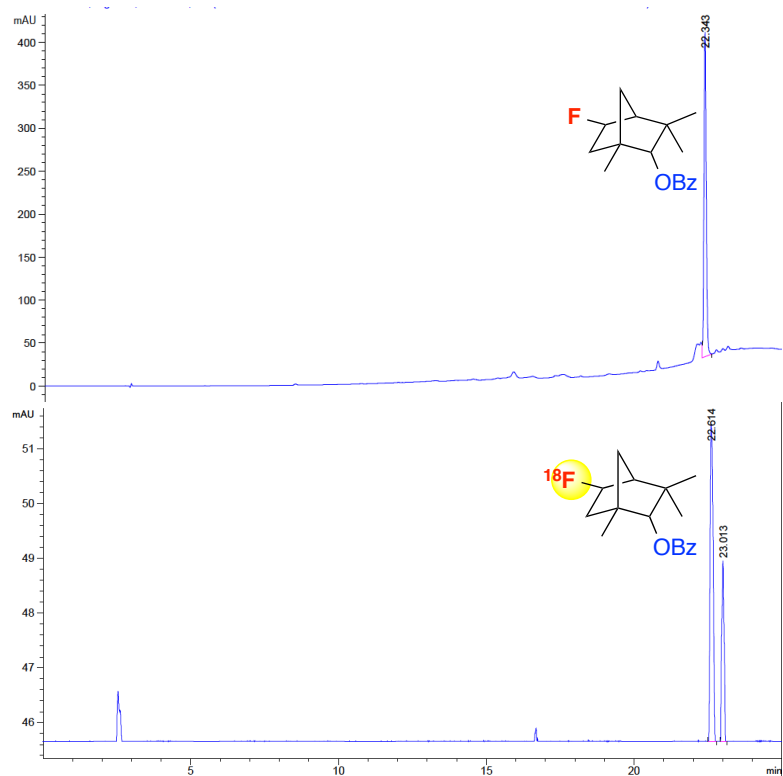


Figure S4w. UV and radio-HPLC traces of compound 18

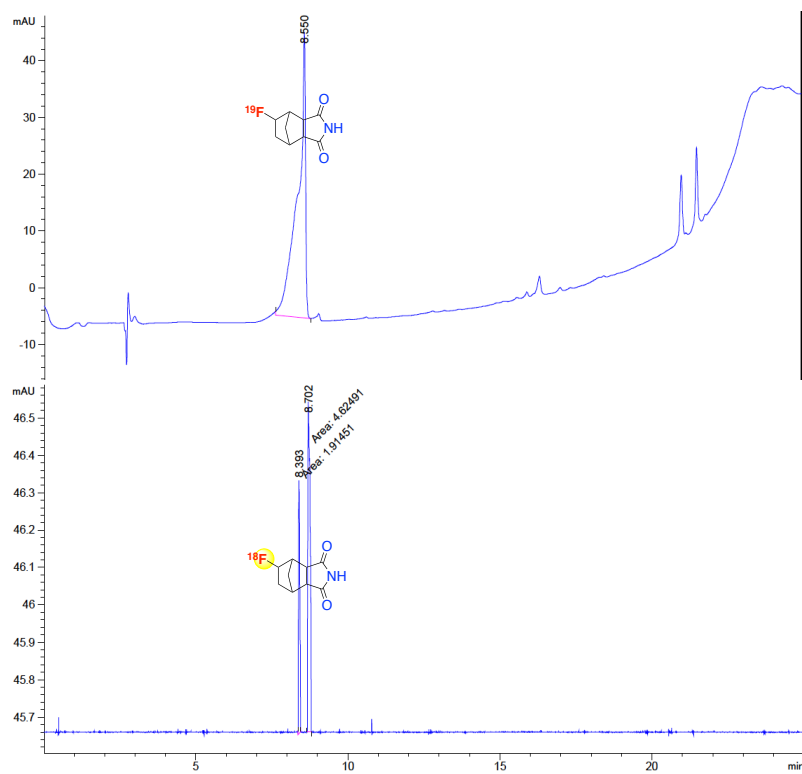


Figure S4w. UV and radio-HPLC traces of compound 19

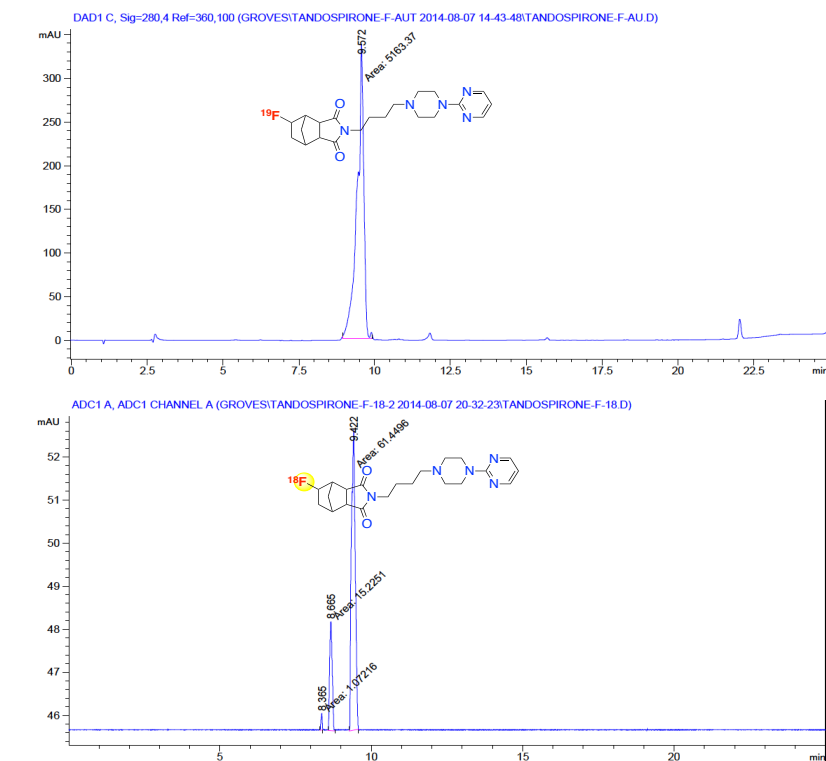


Figure S4x. UV and radio-HPLC traces of compound 20

