Hubert, Stepek, Noda and Bode

Supporting Information

Supporting Information

# Synthetic Fermentation of $\beta$ -Peptide Macrocycles by Thiadiazole-Forming Ring-Closing Reactions

Jonathan G. Hubert<sup>1</sup>, Iain A. Stepek<sup>1</sup>, Hidetoshi Noda<sup>2</sup> and Jeffrey W. Bode<sup>1\*</sup>

<sup>1</sup>Laboratorium für Organishe Chemie, Department of Chemistry and Applied Biosciences, ETH-Zürich, 8093, Switzerland <sup>2</sup>Institute of Microbial Chemistry (Bikaken), Tokyo, 3-14-23 Kamiosaki, Shinagawa-ku, Tokyo 141-0021, Japan.

bode@org.chem.ethz.ch

# Table of contents

General Methods	S3
Synthesis of Building Blocks	S4
1.1 Synthesis of Initiator 16	S4
1.1.1 3-lodophenylhydrazide <b>19</b>	S4
1.1.2 Aldehyde <b>21</b>	S5
1.1.3 Enol ester <b>23</b>	S5
1.1.4 Thiohydrazide <b>24</b>	S6
1.1.5 α-Ketoacid <b>16</b>	S7
1.2 Synthesis of Monomers	S7
1.2.1 Characterization data for new monomers and related intermediate	∋s…S8
1.3 Synthesis of Thiohydrazides	S13
1.3.1 3-Methoxyphenyl thiohydrazide <b>10</b>	S13
1.3.2 Pyridine thiohydrazide <b>12</b>	S14
1.3.3 Alkyl thiohydrazide 15	S14
1.3.4 Phenyl thiohydrazide 2	S15
Synthesis of 1,3,4-Thiadiazoles	S15
2.1 Intermolecular Reaction of Thiohydrazides and $\alpha\mbox{-}Ketoacids\mbox{-}$	S15
2.1.1 Phenyl thiadiazole <b>5</b>	S16
2.1.2 Propanoic acid thiadiazole <b>7</b>	S16
2.1.3 Fmoc-amine thiadiazole 9	S17
2.1.4 3-Methoxyphenyl thiadiazole <b>11</b>	S17
2.1.5 2,3-Dihydrothiadiazole carboxylic acid <b>4</b>	S18
Synthesis of $\beta$ -Peptide Macrocycle Mixtures	S19
3.1 One-pot Elongation/Macrocyclization Reaction with One Monomer	S19
3.1.1 HPLC spectra with mass traces of the major peaks	S20
3.2 Cyclization of Purified Tri-β-peptide	S26
3.3 One-Pot Elongation/Macrocyclization Reaction with Two Monomers	S27
3.3.1 HPLC spectra with mass traces of the major peaks	S28
3.4 Isolation and Characterization of Macrocyclic Compounds	S36
References	S39
NMR Spectra	S40

#### **General Methods**

## Reactions and Purifications

Reactions were carried out under air unless otherwise stated. Thin layer chromatography (TLC) was performed on Merck TLC plates (0.25 mm) pre-coated with silica gel 60 F254 and visualized by UV quenching and/or staining with potassium permanganate stain and warming with a heat gun. Flash column chromatography was performed under a forced-flow of air using Silicycle SiliaFlash F60 (40-63 mm particle size). Macrocycle mixtures were analyzed and purified by reversed phase high performance liquid chromatography (RP-HPLC) on Jasco analytical and preparative instruments with dual pumps, mixer and in-line degasser, a variable wavelength UV detector (simultaneous monitoring of the eluent at 220 nm, 254 nm, 301 nm) and a Rheodyne 7725i injector fitted with a 20  $\mu$ L injection loop. The mobile phase for analytical and preparative HPLC were Millipore-H<sub>2</sub>O with 0.1% TFA (Buffer A) and HPLC grade CH<sub>3</sub>CN with 0.1% TFA (Buffer B). Analytical HPLC was performed on Shiseido C18 (5  $\mu$ m, 4.6 mm l.D. x 250 mm) column at a flow rate of 1 mL/min. Preparative HPLC was performed on YMC C18 (5  $\mu$ m, 20 mm I.D. x 250 mm) column at a flow rate of 10 mL/min. LCMS analysis was performed on Dionex UltiMate 3000 RSLC connected to a Surveyor MSQ Plus mass spectrometer; a reversed-phase RESTEK Pinnacle DB C18 (4.6 x 50 mm) column was used, running a gradient of 5 to 100% CH<sub>3</sub>CN in H<sub>2</sub>O over 6.5 min, 100% CH<sub>3</sub>CN for 2.5 min.

#### Characterization

NMR spectra were recorded on Bruker AV-400 or AV-III-600 instruments. Chemical shifts ( $\delta$ ) are given in ppm relative to residual solvent peaks. Data for 1H NMR are reported as follows: chemical shift (multiplicity, coupling constants where applicable, number of hydrogens). Abbreviations are as follows: s (singlet), d (doublet), t (triplet), q (quartet), p (pentet), dd (doublet of doublet), m (multiplet), br (broad), ABq (AB quartet). IR spectra were recorded on a Jasco FT/IR-4100 spectrometer and major peaks are reported in frequency of absorption (cm<sup>-1</sup>). Optical rotations were measured on a Jasco P-2000 operating at the sodium D line with a 100 mm path length cell. High-resolution mass spectra were obtained by the mass spectrometry service of the ETH Zürich Laboratorium für Organische Chemie on a Bruker Daltonics maXis ESI-QTOF spectrometer (ESI).

S3

### Solvents and Reagents

All organic solvents (CH<sub>3</sub>CN, DMF, <sup>1</sup>BuOH, Et<sub>2</sub>O, MeOH) were used as supplied (ACS or HPLC grade) unless otherwise stated. THF was purified by distillation over sodium benzophenone ketyl prior to use. CH<sub>2</sub>Cl<sub>2</sub> was purified by distillation over calcium hydride. H<sub>2</sub>O used for reactions was obtained from a Millipore purification system. All other starting materials were used as supplied by commercial vendors or prepared by the method described in the corresponding reference.

## **Synthesis of Building Blocks**

## 1.1 Synthesis of Initiator 16

1.1.1 3-lodophenylhydrazide 19



EDCI·HCI (0.95 g, 4.98 mmol, 1.1 equiv) was added to a suspension of 3-iodobenzoic acid (1.20 g, 4.98 mmol, 1.1 equiv) and HOBt (0.67 g, 4.98 mmol, 1.1 equiv) in  $CH_2CI_2$  (23 mL) at rt. The mixture was stirred for 10 min to give a yellow solution. *t*-Butylcarbazate (0.60 g, 4.52 mmol, 1.0 equiv) and  $EtN^iPr_2$  (3.2 mL, 18.1 mmol, 4.0 equiv) were added and the mixture was stirred for 16 h. The resulting suspension was cooled to 0 °C, filtered and washed with cold  $CH_2CI_2$ , to afford the desired product **19** (1.05 g, 64%) as a white solid.

**MP** 198–199 °C; <sup>1</sup>**H NMR** (400 MHz, DMSO- $d_6$ )  $\delta$  10.29 (s, 1H), 8.96 (s, 1H), 8.19 (s, 1H), 7.94 (d, J = 7.8 Hz, 1H), 7.86 (d, J = 7.8 Hz, 1H), 7.31 (t, J = 7.8 Hz, 1H), 1.43 (s, 9H); <sup>13</sup>**C NMR** (101 MHz, DMSO- $d_6$ )  $\delta$  164.6, 155.4, 140.3, 135.8, 134.5, 130.7, 126.7, 94.7, 79.3, 28.1; **IR** (u/cm<sup>-1</sup>, thin film): 3232 (br), 2980, 1718, 1659, 1425, 1394, 1366, 1250, 1152, 1066, 699; **HRMS** (ESI): calculated for C<sub>12</sub>H<sub>15</sub>IN<sub>2</sub>NaO<sub>3</sub> [M+Na]<sup>+</sup>: 385.0020, found: 385.0025.

1.1.2 Aldehyde 21



3-lodophenylhydrazide **19** (1.00 g, 2.76 mmol, 1.0 equiv),  $Pd(OAc)_2$  (37 mg, 0.17 mmol, 6 mol%), tetrabutylammonium bromide (0.89 g, 2.76 mmol, 1.0 equiv), NaHCO<sub>3</sub> (0.58 g, 6.90 mmol, 2.5 equiv) and MS 4Å were placed under a N<sub>2</sub> atmosphere in a flame dried flask. DMF (8.3 mL) was added, followed by allyl alcohol (0.28 mL, 4.14 mmol, 1.5 equiv) and the mixture was heated to 70 °C for 4 h. The resulting mixture was diluted with EtOAc, washed with H<sub>2</sub>O and brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo*. The crude mixture was purified by flash chromatography (hexanes/EtOAc, 4:1 $\rightarrow$ 1:1) to afford the desired product **21** (0.66 g, 81%) as a white solid.

**MP** 60–61 °C; <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  9.79 (s, 1H), 8.95 (br. s, 1H), 7.67 (s, 1H), 7.61 (d, *J* = 7.4 Hz, 1H), 7.38 – 7.17 (m, 2H), 6.96 (br. s, 1H), 2.91 (t, *J* = 7.5 Hz, 2H), 2.76 (t, *J* = 7.5 Hz, 2H), 1.48 (s, 9H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>)  $\delta$  201.3, 166.9, 156.4, 141.1, 132.4, 132.0, 128.9, 127.3, 125.4, 82.2, 45.0, 28.2, 27.9; **IR** (u/cm<sup>-1</sup>, thin film): 3281 (br), 2978, 2932, 1713, 1661, 1524, 1480, 1393, 1367, 1252, 1156, 730; **HRMS** (ESI): calculated for C<sub>15</sub>H<sub>20</sub>N<sub>2</sub>NaO<sub>4</sub> [M+Na]<sup>+</sup>: 315.1315, found: 315.1316.

1.1.3 Enol ester 23



Tetramethylguanidine (0.25 mL, 2.05 mmol, 1.2 equiv) was added dropwise to a mixture of phosphonate  $22^{1,2}$  (0.55 g, 1.88 mmol, 1.1 equiv) and LiCl (86 mg, 2.05 mmol, 1.2 equiv) in THF (9 mL) at -10 °C under an atmosphere of N<sub>2</sub>. After stirring for 30 min a solution of aldehyde 21 (0.50 g, 1.71 mmol, 1.0 equiv) in THF (2 mL) was added. The resulting mixture was stirred for 10 min, diluted with sat. aq. NH<sub>4</sub>Cl and extracted with EtOAc (3×). The combined organic extracts were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude mixture was purified

by flash chromatography (hexanes/EtOAc,  $4:1 \rightarrow 2:1$ ) to afford the desired product **23** (0.53 g, 72%, 2:1 mixture of isomers) as a white solid.

**MP** 46–48 °C; <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  8.26 (br. s, 1H), 7.68 – 7.58 (m, 2H), 7.36 – 7.28 (m, 2H), 6.80 (br. s, 1H), 5.56\* (t, *J* = 7.7 Hz, 0.33H), 5.47 (t, *J* = 8.3 Hz, 0.66H), 2.92 – 2.81 (m, 1.33H), 2.79-2.72 (m, 2H), 2.35 – 2.48\* (m, 0.66H), 1.79 – 1.61 (m, 8H), 1.49 – 1.39 (m, 11H). (\*signals from minor isomer); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>)  $\delta$  167.1, 167.0, 163.0, 162.6, 156.0, 141.7, 141.6, 139.0, 138.1, 132.5, 132.4, 132.0, 128.7, 128.7, 127.6, 127.6, 125.2, 125.1, 113.2, 111.9, 111.0, 108.8, 82.0, 82.0, 36.2, 36.1, 35.7, 34.5, 28.2, 27.0, 25.7, 24.3, 24.3, 22.9, 22.8 (mixture of isomers); **IR** (u/cm<sup>-1</sup>, thin film): 3283 (br), 2939, 2855, 1780, 1721, 1667, 1480, 1452, 1368, 1250, 1158, 940, 909, 730; **HRMS** (ESI): calculated for C<sub>23</sub>H<sub>30</sub>N<sub>2</sub>NaO<sub>6</sub> [M+Na]<sup>+</sup>: 453.1996, found: 453.1996.

#### 1.1.4 Thiohydrazide 24



A suspension of enol ester **23** (0.28 g, 0.65 mmol, 1.0 equiv) and Lawesson's reagent (0.26 g, 0.65 mmol, 1.0 equiv) in THF (1.2 mL) was stirred at 45 °C for 6 h under an atmosphere of N<sub>2</sub>. The mixture was filtered through a silica plug, washing hexanes/EtOAc (4:1), and the filtrate was concentrated *in vacuo*. The crude mixture was purified by flash chromatography (hexanes/EtOAc, 9:1) to afford the desired product **24** (0.29 g, 70%, single isomer) as a yellow oil.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 9.86 (br. s, 1H), 8.86 (br. s, 1H), 7.62 (s, 1H), 7.58 – 7.56 (m, 1H), 7.32 – 7.30 (m, 2H), 5.56 (t, J = 7.7 Hz, 1H), 2.79 (t, J = 7.7 Hz, 2H), 2.52 (dt, J = 7.7, 7.7 Hz, 2H), 1.79 – 1.63 (m, 8H), 1.51 (s, 9H), 1.49 – 1.38 (m, 2H); 1<sup>3</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 163.0, 153.6, 141.9, 139.1, 138.2, 131.9, 128.8, 127.3, 124.7, 111.9, 108.6, 83.5, 36.2, 34.5, 28.3, 27.0, 24.3, 22.9. (C=S carbon not observed); **IR** (u/cm<sup>-1</sup>, thin film): 3263 (br), 2938, 2803, 1784, 1719, 1449, 1367, 1264, 1247, 1154, 936, 734, 697; **HRMS** (ESI): calculated for C<sub>23</sub>H<sub>30</sub>N<sub>2</sub>NaO<sub>5</sub>S [M+Na]<sup>+</sup>: 469.1768, found: 469.1774.

S6

### 1.1.5 *α-Ketoacid* **16**



Aqueous NaOH (2 M, 0.22 mL, 0.44 mmol, 2.0 equiv) was added to a solution of thiohydrazide **24** (0.10 g, 0.22 mmol, 1.0 equiv) in methanol (1.1 mL) and the reaction was stirred at rt for 15 min. The mixture was diluted with H<sub>2</sub>O and washed with Et<sub>2</sub>O. The aqueous phase was acidified to pH 1 with 3 M HCl and extracted with EtOAc ( $3\times$ ). The combined organic extracts were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub>, and concentrated *in vacuo* to afford the desired product **16** (80 mg, 99%) as a yellow oil that was used without further purification.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 10.04 (br. s, 1H), 8.85 (br. s, 1H), 7.62 (d, J = 7.3 Hz, 1H), 7.53 (s, 1H), 7.35 – 7.27 (m, 2H), 2.89 (t, J = 7.2 Hz, 2H), 2.70 (t, J = 7.2 Hz, 2H), 2.01 (tt, J = 7.2, 7.2 Hz, 2H), 1.53 (s, 9H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 195.5, 160.3, 154.2, 141.5, 138.4, 132.0, 129.1, 127.1, 125.5, 84.0, 36.7, 34.7, 28.3, 24.8. (C=S carbon not observed); **IR** (u/cm<sup>-1</sup>, thin film): 3258 (br), 2979, 2934, 1713, 1427, 1368, 1250, 1152, 735, 698; **HRMS** (ESI): calculated for C<sub>17</sub>H<sub>22</sub>N<sub>2</sub>NaO<sub>5</sub>S [M+Na]<sup>+</sup>: 389.1142, found: 389.1144.

#### **1.2 Synthesis of Monomers**



Monomers were prepared using a previously described three-step procedure from 2,3:5,6-*O*-diisopropylidene-D-gulose oxime (**46**), 5-chloromethyl-2,2-pentamethylene-1,3-dioxolan-4-one (**47**) and commercially available or known aldehydes **48**.<sup>2,3</sup>

## General procedure:

1) A solution of NEt<sub>3</sub> (2.0 equiv) and dioxolanone **47** (1.0 equiv) in <sup>*n*</sup>PrOAc (0.5 M) was heated to reflux for 18 h. Oxime **46** (1.0 equiv) and aldehyde **48** (1.0 equiv) were added and the mixture was heated to reflux for 24 h. The reaction mixture was

diluted with EtOAc, washed with 1 M HCl and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude mixture was purified by flash chromatography and/or recrystallization to afford the desired cycloaddition products **49**.

2)  $HCIO_4$  (70%, 3.0 equiv) was added dropwise to a solution of cycloaddition product 49 (1.0 equiv) in CH<sub>3</sub>CN (0.1 M) at rt. The reaction was stirred for 6 h, sat. aq. NaHCO<sub>3</sub> was added and the mixture was extracted with EtOAc (3×). The combined organic extracts were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude mixture was purified by flash chromatography to afford the unprotected isoxazolidine product.

3) HCl in dioxane (4 M, 1.1 equiv) was added dropwise to a solution of unprotected isoxazolidine (1.0 equiv) in  $Et_2O$  (0.1 M). After stirring at rt for 30 min a precipitate was formed. The precipitate was collected by filtration, washed with  $Et_2O$  and dried under vacuum to afford the desired monomer HCl salt **50**.

For monomers previously reported (**25, 30** and **37**), the syntheses were performed according to the literature procedures.<sup>2,3</sup>

#### 1.2.1 Characterization Data for New Monomers and Related Intermediates

n-Propyl monomer intermediate 51



[**a**]<sub>D</sub><sup>28</sup> (c = 0.53, CHCl<sub>3</sub>) = +19.4; **MP** 90-91°C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.87 (d, J = 6.1 Hz, 1H), 4.69 – 4.64 (m, 2H), 4.36 (dt, J = 8.5, 6.7 Hz, 1H), 4.19 (dd, J = 8.6, 6.8 Hz, 1H), 4.03 (dd, J = 8.5, 3.9 Hz, 1H), 3.85 – 3.78 (m, 1H), 3.71 (dd, J = 8.6, 6.6 Hz, 1H), 2.92 (dd, J = 13.8, 7.7 Hz, 1H), 2.11 (dd, J = 13.8, 1.9 Hz, 1H), 1.92 – 1.58 (m, 9H), 1.50 – 1.33 (m, 14H), 1.28 (s, 3H), 0.94 (t, J = 7.1 Hz, 3H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 169.6, 113.0, 111.8, 109.9, 105.8, 96.9, 84.5, 84.4, 80.3, 75.7, 66.2, 60.5, 41.1, 37.7, 36.5, 35.6, 26.8, 26.2, 25.4, 25.1, 24.4, 23.1, 23.0, 20.3, 13.9; **IR** (u/cm<sup>-1</sup>, thin film): 2988, 2939, 1754, 1380, 1372, 1216, 1093, 1060, 1031, 844; **HRMS** (ESI): calculated for C<sub>25</sub>H<sub>40</sub>NO<sub>9</sub> [M+H]<sup>+</sup>: 498.2698, found: 498.2701.

n-Propyl monomer 26



[**a**]<sub>D</sub><sup>28</sup> (c = 0.52, MeOH) = +34.9; **MP** 113–114 °C; <sup>1</sup>**H NMR** (400 MHz, MeOD) δ 4.19 (ddt, *J* = 7.6, 7.6, 7.6 Hz, 1H), 3.14 (dd, *J* = 14.3, 7.6 Hz, 1H), 2.56 (dd, *J* = 14.3, 7.6 Hz, 1H), 1.95 – 1.83 (m, 6H), 1.80 – 1.65 (m, 4H), 1.56 – 1.43 (m, 4H), 1.03 (t, *J* = 7.3 Hz, 3H); <sup>13</sup>**C NMR** (101 MHz, MeOD) δ 166.4, 114.6, 108.1, 62.9, 41.0, 38.2, 36.6, 32.6, 25.1, 24.0, 23.9, 20.8, 14.0; **IR** (u/cm<sup>-1</sup>, thin film): 2959, 2859, 1807, 1377, 1292, 1258, 1191, 1139, 915, 718; **HRMS** (ESI): calculated for C<sub>13</sub>H<sub>22</sub>NO<sub>4</sub> [M-Cl]<sup>+</sup> 256.1543, found: 256.1550.

Pyran monomer intermediate 52



**[a]**<sub>D</sub><sup>25</sup> (c = 0.53, CHCl<sub>3</sub>) = -15.3; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 4.85 (d, J = 6.1 Hz, 1H), 4.69 (s, 1H), 4.65 (dd, J = 6.1, 3.9 Hz, 1H), 4.35 (dt, J = 8.5, 6.7 Hz, 1H), 4.20 (dd, J = 8.6, 6.9 Hz, 1H), 4.02 – 3.94 (m, 3H), 3.72 (dd, J = 8.6, 6.5 Hz, 1H), 3.58 – 3.53 (m, 1H), 3.43 – 3.33 (m, 2H), 2.79 (dd, J = 14.1, 8.0 Hz, 1H), 2.31 (dd, J = 14.1, 1.1 Hz, 1H), 1.92 – 1.56 (m, 11H), 1.47 – 1.25 (m, 16H); <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 169.5, 113.1, 111.8, 109.9, 106.0, 96.7, 84.7, 84.3, 80.3, 75.7, 67.8, 67.6, 66.1, 65.4, 37.7, 36.7, 36.6, 36.5, 31.3, 30.2, 26.9, 26.2, 25.3, 25.0, 24.4, 23.1, 23.1; **IR** (u/cm<sup>-1</sup>, thin film): 2986, 2939, 2856, 1800, 1450, 1372, 1249, 1237, 1210, 1086, 1032, 847, 732; **HRMS** (ESI): calculated for C<sub>27</sub>H<sub>41</sub>NNaO<sub>10</sub> [M+Na]<sup>+</sup>: 562.2623, found: 562.2620.

Pyran monomer 27



[**a**]<sub>D</sub><sup>28</sup> (c = 0.51, MeOH) = +28.1; **MP** 129–132 °C; <sup>1</sup>**H NMR** (400 MHz, MeOD) δ 4.01 − 3.89 (m, 3H), 3.50 − 3.39 (m, 2H), 3.17 (dd, J = 14.4, 7.5 Hz, 1H), 2.60 (dd, J = 14.4, 9.9 Hz, 1H), 2.12-2.02 (m, 1H), 1.94 − 1.83 (m, 4H), 1.81 − 1.60 (m, 6H), 1.57 − 1.46 (m, 4H); <sup>13</sup>**C NMR** (101 MHz, MeOD) δ 165.3, 113.2, 106.9, 66.6, 66.2, 38.2, 36.8, 35.8, 35.3, 30.0, 29.0, 23.7, 22.6, 22.5; **IR** (u/cm<sup>-1</sup>, thin film): 2929, 2863, 2834, 1799, 1279, 1250, 1244, 1199, 1088, 943, 711; **HRMS** (ESI): calculated for C<sub>15</sub>H<sub>24</sub>NO<sub>5</sub> [M-Cl]<sup>+</sup>: 298.1649, found: 298.1650

#### 4-Benzyloxybenzyl monomer intermediate 53



**[a]**<sub>D</sub><sup>26</sup> (c = 0.53, CHCl<sub>3</sub>) = -24.3; **MP** 123–124 °C; <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 7.44 – 7.30 (m, 5H), 7.13 (d, J = 8.6 Hz, 2H), 6.90 (d, J = 8.6 Hz, 2H), 5.03 (s, 2H), 4.85 (d, J = 6.1 Hz, 1H), 4.69 (s, 1H), 4.57 (dd, J = 6.0, 4.1 Hz, 1H), 4.32 – 4.26 (m, 1H), 4.13 (dd, J = 8.4, 6.6 Hz, 1H), 4.10 – 4.04 (m, 1H), 3.64 (dd, J = 8.4, 4.1 Hz, 1H), 3.56 (dd, J = 8.4, 6.9 Hz, 1H), 3.01 (dd, J = 13.7, 7.7 Hz, 1H), 2.82 – 2.73 (m, 2H), 2.20 (dd, J = 14.0, 1.7 Hz, 1H), 1.94 – 1.88 (m, 2H), 1.81 – 1.67 (m, 6H), 1.50 – 1.42 (m, 8H), 1.37 (s, 3H), 1.28 (s, 3H). <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>) δ 169.4, 157.6, 137.2, 130.9, 130.6, 128.7, 128.1, 127.6, 114.9, 113.0, 111.8, 109.8, 105.9, 96.4, 84.5, 84.1, 80.2, 75.7, 70.1, 66.1, 61.7, 39.8, 38.5, 37.7, 36.5, 27.1, 26.2, 25.5, 25.0, 24.4, 23.2, 23.1; **IR** (u/cm<sup>-1</sup>, thin film): 2990, 2940, 1798, 1512, 1376, 1239, 1209, 1156, 1089, 1066, 1037, 847; **HRMS** (ESI): calculated for C<sub>36</sub>H<sub>46</sub>NO<sub>10</sub> [M+H]<sup>+</sup>: 652.3116, found: 652.3104. 4-Hydroxybenzyl monomer intermediate 54



[**a**]<sub>D</sub><sup>28</sup> (c = 0.52, CHCl<sub>3</sub>) = -26.2; **MP** 83-85 °C; <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.07 (d, J = 8.5 Hz, 2H), 6.75 (d, J = 8.5 Hz, 2H), 5.45 (s, 1H), 4.84 (d, J = 6.0 Hz, 1H), 4.68 (s, 1H), 4.56 (dd, J = 6.1, 4.0 Hz, 1H), 4.32 – 4.27 (m, 1H), 4.13 (dd, J = 8.4, 6.6 Hz, 1H), 4.07 – 4.00 (m, 1H), 3.62 – 3.55 (m, 2H), 2.98 (dd, J = 13.8, 8.0 Hz, 1H), 2.80 – 2.70 (m, 2H), 2.18 (dd, J = 13.9, 1.7 Hz, 1H), 1.93 – 1.87 (m, 2H), 1.82 – 1.65 (m, 6H), 1.48 – 1.40 (m, 8H), 1.37 (s, 3H), 1.27 (s, 3H); <sup>13</sup>C **NMR** (101 MHz, CDCl<sub>3</sub>) δ 169.4, 154.4, 130.8, 130.6, 115.5, 113.0, 111.9, 109.8, 106.0, 96.4, 84.5, 84.0, 80.2, 75.7, 66.1, 61.7, 39.8, 38.5, 37.7, 36.5, 26.9, 26.1, 25.3, 24.9, 24.4, 23.2, 23.0; **IR** (u/cm<sup>-1</sup>, thin film): 2987, 2938, 1799, 1516, 1372, 1228, 1210, 1085, 1066, 1038, 844; **HRMS** (ESI): calculated for C<sub>29</sub>H<sub>40</sub>NO<sub>10</sub> [M+H]<sup>+</sup>: 562.2647, found: 562.2648.

4-Hydroxybenzyl monomer 28



**[α]**<sub>D</sub><sup>28</sup> (c = 0.53, MeOH) = +47.9; **MP** 120–122°C; <sup>1</sup>**H NMR** (400 MHz, MeOD) δ 7.14 (d, *J* = 8.5 Hz, 2H), 6.80 (d, *J* = 8.5 Hz, 2H), 4.43 – 4.35 (m, 1H), 3.12 (d, *J* = 8.5 Hz, 2H), 3.03 (dd, *J* = 14.4, 7.5 Hz, 1H), 2.59 (dd, *J* = 14.4, 6.7 Hz, 1H), 1.95 – 1.87 (m, 4H), 1.82 – 1.65 (m, 4H), 1.57 – 1.48 (m, 2H); <sup>13</sup>**C NMR** (101 MHz, MeOD) δ 166.4, 158.3, 131.2, 126.9, 117.0, 114.6, 108.0, 64.3, 40.4, 38.3, 36.6, 35.5, 25.1, 24.0, 24.0; **IR** (u/cm<sup>-1</sup>, thin film): 3318, 2940, 2875, 1807, 1519, 1269, 1213, 1190, 1153, 1116, 920, 872, 839, 720; **HRMS** (ESI): calculated for  $C_{17}H_{22}NO_5$  [M-Cl]<sup>+</sup>: 320.1492, found: 320.1493.

Amide monomer intermediate 55



**[a]**<sub>D</sub><sup>28</sup> (c = 0.52, CHCl<sub>3</sub>) = +31.6; **MP** 144–145°C; <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 6.53 (t, *J* = 5.5 Hz, 1H), 4.87 (d, *J* = 6.0 Hz, 1H), 4.67 – 4.61 (m, 2H), 4.35 (dt, *J* = 8.2, 7.0 Hz, 1H), 4.22 (dd, *J* = 8.6, 6.7 Hz, 1H), 4.09 (dd, *J* = 8.2, 3.7 Hz, 1H), 3.74 – 3.67 (m, 2H), 3.61 – 3.47 (m, 7H), 3.40 – 3.34 (m, 4H), 2.95 (dd, *J* = 13.9, 8.1 Hz, 1H), 2.37 – 2.30 (m, 1H), 2.17 – 2.10 (m, 2H), 2.08 – 1.98 (m, 1H), 1.92 – 1.59 (m, 9H), 1.47 – 1.36 (m, 11H), 1.28 (s, 3H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 172.6, 169.2, 113.2, 111.9, 110.0, 106.0, 97.5, 84.6, 83.7, 80.3, 75.8, 72.0, 70.3, 69.9, 66.1, 61.1, 59.1, 41.0, 39.5, 37.7, 36.5, 34.5, 30.0, 26.8, 26.2, 25.5, 25.1, 24.4, 23.1, 22.9; **IR** (u/cm<sup>-1</sup>, thin film): 2988, 2939, 1755, 1379, 1372, 1217, 1060, 1031, 845; **HRMS** (ESI): calculated for C<sub>30</sub>H<sub>49</sub>N<sub>2</sub>O<sub>12</sub> [M+H]<sup>+</sup>: 629.3280, found: 629.3281.

Amide monomer 29



[**a**]<sub>D</sub><sup>24</sup> (c = 0.49, MeOH) = −17.5; <sup>1</sup>**H** NMR (400 MHz, MeOD) δ 4.33 − 4.26 (m, 1H), 3.62 − 3.60 (m, 2H), 3.57 − 3.54 (m, 4H), 3.41 − 3.37 (m, 5H), 3.15 (dd, *J* = 14.4, 7.9 Hz, 1H), 2.66 − 2.49 (m, 3H), 2.28 − 2.09 (m, 2H), 1.93 − 1.85 (m, 4H), 1.76-1.66 (m, 4H), 1.54 − 1.47 (m, 2H); <sup>13</sup>**C** NMR (101 MHz, MeOD) δ 175.0, 166.2, 114.5, 107.8, 73.0, 71.0, 70.3, 62.8, 59.2, 41.3, 40.6, 38.2, 36.6, 32.9, 26.4, 25.0, 24.0, 23.9; IR (u/cm<sup>-1</sup>, thin film): 2938, 2857, 1798, 1651, 1269, 1238, 1177, 1139, 1090, 929; HRMS (ESI): calculated for  $C_{18}H_{31}N_2O_7$  [M-CI]<sup>+</sup>: 387.2122, found: 387.2126.

#### 1.3 Synthesis of Thiohydrazides



#### General procedure:

1) EDCI·HCI (1.1 equiv) was added to a suspension of carboxylic acid (1.1 equiv) and HOBt (1.1 equiv) in CH<sub>2</sub>Cl<sub>2</sub> (0.2 M) at rt. The mixture was stirred for 10 min, *t*-butylcarbazate (1.0 equiv) and DIPEA (4 equiv) were added and the mixture was stirred for 16 h. The reaction mixture was diluted with CH<sub>2</sub>Cl<sub>2</sub>, washed with sat. aq. NaHCO<sub>3</sub>, 1 M HCl and brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude mixture was purified by flash chromatography (hexanes/EtOAc) to afford the desired hydrazide product.

2) A suspension of hydrazide (1.0 equiv) and Lawesson's reagent (1.0 equiv) in THF (0.5 M) was stirred at 45 °C for 24 h under an atmosphere of N<sub>2</sub>. The mixture was filtered through a silica plug, washing with hexanes/EtOAc (2:1), and the filtrate was concentrated *in vacuo*. The crude mixture was purified by flash chromatography (hexanes/EtOAc) to afford the desired Boc-thiohydrazide product.

3) Boc-thiohydrazide (1.0 equiv) was dissolved in HCl in dioxane (4 M, 10 equiv) and the solution was stirred for 1-24 h (depending on the substrate) at rt. The reaction mixture was diluted with  $Et_2O$  and the precipitate was collected by filtration, washed with  $Et_2O$  and dried under vacuum to afford the desired thiohydrazide-HCl salt.

1.3.1 3-Methoxyphenyl thiohydrazide 10



Prepared according to the general procedure. 1) 2.27 mmol scale, quant. yield. 2) 2.03 mmol scale, 68% yield. 3) 1.27 mmol scale, 2 h, 70% yield, white solid.

**MP** 141–143 °C; <sup>1</sup>**H NMR** (400 MHz, MeOD) δ 7.40 – 7.37 (m, 3H), 7.17 – 7.13 (m, 1H), 3.86 (s, 3H); <sup>13</sup>**C NMR** (101 MHz, MeOD) δ 198.0, 159.7, 129.3, 119.3, 117.7, 113.0, 54.6; **IR** (u/cm<sup>-1</sup>, thin film): 3200-2500 (br), 3129, 3082, 2942, 1607, 1580,

1483, 1459, 1433, 1288, 1147, 1042, 989, 786, 778; **HRMS** (EI): calculated for  $C_8H_{10}N_2OS [M-HCI]^+$ : 182.0508, found: 182.0509

1.3.2 Pyridine thiohydrazide 12



Prepared according to the general procedure. 1) 4.55 mmol scale, 61% yield. 2) 2.53 mmol scale, 22% yield. 3) 0.55 mmol scale, 24 h, 79% yield, yellow solid. **MP** 133–134 °C; <sup>1</sup>**H NMR** (400 MHz, MeOD)  $\delta$  8.64 (ddd, *J* = 5.5, 1.6, 1.0 Hz, 1H), 8.56 (ddd, *J* = 7.9, 1.0. 1.0 Hz, 1H), 8.41 (ddd, *J* = 7.9, 7.9, 1.6 Hz, 1H), 7.90 (ddd, *J* = 7.9, 5.5, 1.0 Hz, 1H)<sup>; 13</sup>**C NMR** (101 MHz, MeOD)  $\delta$  184.4, 150.0, 146.0, 143.7, 128.7, 126.1; **IR** (u/cm<sup>-1</sup>, thin film): 3382 (br), 3073, 3038, 2993, 2531(br), 1578, 1557, 1513, 1456, 1351, 1250, 1219, 1081, 979, 942, 773; **HRMS** (ESI): calculated for C<sub>6</sub>H<sub>8</sub>N<sub>3</sub>S [M–Cl]<sup>+</sup>: 154.0433, found: 154.0437

1.3.3 Alkyl thiohydrazide 15



Prepared according to the general procedure. 1) 4.66 mmol scale, 60% yield. 2) 2.92 mmol scale, 27% yield. 3) 0.80 mmol scale, 1 h, 67% yield, white solid. **MP** 121-123 °C; <sup>1</sup>**H NMR** (400 MHz, MeOD)  $\delta$  7.29 – 7.16 (m, 5H), 3.12 (t, *J* = 7.3

Hz, 2H), 2.99 (t, J = 7.3 Hz, 2H); <sup>13</sup>**C** NMR (101 MHz, MeOD) δ 205.0, 141.3, 129.6, 127.5, 45.1, 36.2; **IR** (u/cm<sup>-1</sup>, thin film): 3200-2600 (br), 1581, 1548, 1465, 1372, 1209, 1110, 1065, 752, 695; **HRMS** (ESI): calculated for C<sub>9</sub>H<sub>13</sub>N<sub>2</sub>S [M-Cl]<sup>+</sup>: 181.0794, found: 181.0799

## 1.3.4 Phenyl thiohydrazide 2



Phenyl thiohydrazide **2** was prepared according to a literature procedure.<sup>4</sup>

*S*-(thiobenzoyl)-thioglycolic acid (1.30 g, 6.12 mmol, 1.0 equiv) was dissolved in 1 M NaOH (6 mL) and H<sub>2</sub>O (6 mL) was added. The solution was cooled to 0 °C and hydrazide hydrate (1.1 mL, 12.2 mmol, 2.0 equiv) was added dropwise. After stirring for 10 min the pH of the reaction mixture was adjusted to pH 5-6 using 1 M HCl and the resulting suspension was stirred for a further 1 h. The precipitate was collected by filtration, washing with cold H<sub>2</sub>O, and dried under vacuum. The crude material was recrystallized (CH<sub>2</sub>Cl<sub>2</sub>/hexanes) to provide desired product **2** (0.61 g, 66%) as a pale yellow solid. <sup>1</sup>H NMR (400 MHz, DMSO-*d*<sub>6</sub>)  $\delta$  7.72 – 7.69 (m, 2H), 7.47 – 7.38 (m, 3H). The spectroscopic data were in agreement with literature values.<sup>4</sup>

## Synthesis of 1,3,4-Thiadiazoles

## 2.1 Intermolecular Reaction of Thiohydrazides and $\alpha$ -Ketoacids



## General procedure:

A solution of thiohydrazide (1.5 equiv) in <sup>t</sup>BuOH/1M HCI (5:1, 0.1 M) was added to a solution of  $\alpha$ -ketoacid (1.0 equiv) in <sup>t</sup>BuOH/1M HCI (5:1, 0.1 M) and the mixture was heated at 70 °C for 16 h. The reaction mixture was diluted with water and extracted with EtOAc (3×). The combined organic extracts were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude mixture was purified by flash chromatography to afford the desired 1,3,4-thiadiazole products.

#### 2.1.1 Phenyl thiadiazole 5



Thiadiazole **5** was prepared following the general procedure with thiohydrazide **2** (50 mg, 0.33 mmol, 1.5 equiv) and  $\alpha$ -ketoacid **3** (36 mg, 0.22 mmol, 1.0 equiv). The desired product was isolated by flash chromatography (hexanes/EtOAc, 9:1→4:1) as a white solid (39 mg, 71%).

**MP** 71–72 °C; <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>)  $\delta$  7.91 – 7.87 (m, 2H), 7.47 – 7.41 (m, 3H), 7.38 – 7.33 (m, 4H), 7.32 – 7.28 (m, 1H), 4.45 (s, 2H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>)  $\delta$ 169.8, 169.4, 137.3, 131.1, 130.3, 129.2, 129.1, 129.0, 127.9, 127.6, 36.7; **IR** (u/cm<sup>-1</sup>, thin film): 3066, 3030, 1494, 1455, 1427, 1225, 1124, 1058, 981, 920, 763, 702, 690; **HRMS** (ESI): calculated for C<sub>15</sub>H<sub>13</sub>N<sub>2</sub>S [M+H]<sup>+</sup>: 253.0794, found: 253.0793

#### 2.1.2 Propanoic acid thiadiazole 7



Thiadiazole **7** was prepared following the general procedure with thiohydrazide **2** (50 mg, 0.33 mmol, 1.5 equiv) and  $\alpha$ -ketoacid **6** (32 mg, 0.22 mmol, 1.0 equiv). The desired product was isolated by flash chromatography (hexanes/EtOAc, 4:1 + 1% formic acid) as a white solid (42 mg, 73%).

**MP** 168–170 °C; <sup>1</sup>**H NMR** (400 MHz, MeOD)  $\delta$  7.95-7.91 (m, 2H), 7.55 – 7.49 (m, 3H), 3.42 (t, *J* = 7.0 Hz, 2H), 2.86 (t, *J* = 7.0 Hz, 2H); <sup>13</sup>**C NMR** (101 MHz, MeOD)  $\delta$  177.1, 171.5, 170.7, 132.5, 131.1, 130.4, 128.8, 34.7, 26.6; **IR** (u/cm<sup>-1</sup>, thin film): 3376 (br), 3062, 2934, 1693, 1536, 1456, 1419, 1239, 1048, 982, 758, 690; **HRMS** (ESI): calculated for C<sub>11</sub>H<sub>11</sub>N<sub>2</sub>O<sub>2</sub>S [M+H]<sup>+</sup>: 235.0536, found: 235.0535

#### 2.1.3 Fmoc-amine thiadiazole 9



Thiadiazole **9** was prepared following the general procedure with thiohydrazide **2** (58 mg, 0.39 mmol, 1.5 equiv) and  $\alpha$ -ketoacid **8**<sup>5,6</sup> (94 mg, 0.26 mmol, 1.0 equiv). The desired product was isolated by flash chromatography (hexanes/EtOAc, 4:1) followed by recrystallization (hexanes/EtOAc) as a white solid (64 mg, 54%).

[**a**]<sub>D</sub><sup>24</sup> (c = 0.25, CHCl<sub>3</sub>) = -41.7; **MP** 154-156 °C; <sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.98 -7.92 (m, 2H), 7.76 (d, *J* = 7.6 Hz, 2H), 7.64 - 7.58 (m, 2H), 7.52 - 7.46 (m, 3H), 7.42 - 7.36 (m, 2H), 7.34 - 7.27 (m, 2H), 5.67 (d, *J* = 9.1 Hz, 1H), 5.13 - 5.07 (m, 1H), 4.47 (d, *J* = 7.0 Hz, 2H), 4.23 (t, *J* = 7.0 Hz, 1H), 2.48 - 2.37 (m, 1H), 1.03 (d, *J* = 6.8 Hz, 6H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 169.9, 168.9, 156.2, 143.8, 141.5, 131.3, 130.1, 129.3, 128.1, 127.9, 127.2, 125.2, 120.1, 67.2, 56.5, 47.4, 33.6, 19.4, 18.0; **IR** (u/cm<sup>-1</sup>, thin film): 3306, 2968, 2950, 1690, 1532, 1452, 1425, 1301, 1261, 1236, 1021; **HRMS** (ESI): calculated for  $C_{27}H_{25}N_3NaO_2S$  [M+Na]<sup>+</sup>: 478.1560, found: 478.1556

## 2.1.4 3-Methoxyphenyl thiadiazole 11



Thiadiazole **11** was prepared following the general procedure with thiohydrazide **10** (46 mg, 0.21 mmol, 1.5 equiv) and  $\alpha$ -ketoacid **3** (23 mg, 0.14 mmol, 1.0 equiv). The desired product was isolated by flash chromatography (hexanes/EtOAc, 9:1 $\rightarrow$ 4:1) as a yellow oil (32 mg, 80%).

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.52 (dd, J = 2.6, 1.6 Hz, 1H), 7.41 – 7.28 (m, 7H), 7.00 (ddd, J = 8.2, 2.6, 1.0 Hz, 1H), 4.46 (s, 2H), 3.86 (s, 3H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 167.0, 169.4, 160.2, 137.3, 131.5, 130.3, 129.2, 129.0, 127.7, 120.7, 117.6, 112.2, 55.3, 36.8; **IR** (u/cm<sup>-1</sup>, thin film): 3028, 3004, 2935, 1598, 1581, 1455, 1426, 1206, 1043, 1002, 843, 781; **HRMS** (ESI): calculated for C<sub>16</sub>H<sub>15</sub>N<sub>2</sub>OS [M+H]<sup>+</sup>: 283.0900, found: 283.0899

#### 2.1.5 2,3-Dihydrothiadiazole carboxylic acid 4



A solution of thiohydrazide **2** (50 mg, 0.33 mmol, 1.0 equiv) and  $\alpha$ -ketoacid **3** (54 mg, 0.33 mmol, 1.0 equiv) in <sup>*t*</sup>BuOH/H<sub>2</sub>O (5:1, 6.6 mL) was heated at 45 °C for 14 h. The reaction mixture was diluted with water and extracted with EtOAc (3×). The combined organic extracts were washed with brine, dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated *in vacuo*. The crude mixture was purified by preparative HPLC (35-95% CH<sub>3</sub>CN with 0.1% TFA over 28 min) to obtain a sample of desired product **4** for characterization. Retention time: 16.9 min.

<sup>1</sup>**H NMR** (400 MHz, CDCl<sub>3</sub>) δ 7.60 – 7.56 (m, 2H), 7.40 – 7.34 (m, 3H), 7.31 – 7.23 (m, 5H), 3.56 (ABq, 2H); <sup>13</sup>**C NMR** (101 MHz, CDCl<sub>3</sub>) δ 174.8, 147.4, 134.2, 130.6, 130.3, 130.2, 128.8, 128.8, 127.9, 127.2, 84.1, 44.5: **IR** (u/cm<sup>-1</sup>, thin film): 3299, 2923, 2853, 1724, 1494, 1447, 1267, 1193, 973, 759, 688; **HRMS** (ESI): calculated for  $C_{16}H_{15}N_2O_2S$  [M+H]<sup>+</sup>: 299.0849, found: 299.0844

## Synthesis of $\beta$ -Peptide Macrocycle Mixtures

## 3.1 One-Pot Elongation/Macrocyclization with One Monomer



## General procedure:

Reactions were prepared using 0.1 M stock solutions of all reagents in  ${}^{t}BuOH/H_{2}O$  (5:1).

Solutions of initiator **16** (10  $\mu$ L, 1.0 equiv) and a monomer (20  $\mu$ L, 2.0 equiv) were mixed and heated at 45 °C for 14 h. The mixture was diluted with <sup>*t*</sup>BuOH/1M HCI (5:1) (90  $\mu$ L) and heated to 70 °C for 4 h. The resulting mixture of cyclic compounds was analyzed by HPLC and/or LCMS.

This procedure was performed on a larger scale (e.g. 0.90 mL of **16**, 1.80 mL monomer) for purification by preparative HPLC. For cyclization on larger scale the mixtures were stirred in an oversized flask (e.g. 50 mL flask for 9 mL solvent) to enhance  $O_2$  exchange.

# 3.1.1 HPLC spectra with mass traces of the major peaks

# Monomer 25



HPLC: Gradient 30 to 90% CH\_3CN with 0.1% TFA in 17 min





HPLC: Gradient 30 to 90% CH<sub>3</sub>CN with 0.1% TFA in 17 min





# HPLC: Gradient 20 to 70% $CH_3CN$ with 0.1% TFA in 17 min





HPLC: Gradient 30 to 90% CH<sub>3</sub>CN with 0.1% TFA in 17 min





HPLC: Gradient 20 to 70% CH<sub>3</sub>CN with 0.1% TFA in 17 min





HPLC: Gradient 30 to 90% CH<sub>3</sub>CN with 0.1% TFA in 17 min



## 3.2 Cyclization of Purified Tri-β-Peptide



A solution of tripeptide **56** (0.26 mg, 0.37  $\mu$ mol) was dissolved in <sup>t</sup>BuOH/1M HCI (5:1, 37  $\mu$ L) was heated to 70 °C for 4 h. The crude reaction mixture was directly analyzed by HPLC (Gradient 30 to 90% CH<sub>3</sub>CN with 0.1% TFA in 17 min).

**HRMS** (ESI) Tripeptide ketoactid **56**: calculated for  $C_{35}H_{56}N_5O_8S$  [M+H]<sup>+</sup>: 706.3844, found: 706.3837. Calculated for  $C_{35}H_{54}N_5O_8S$  [M-H]<sup>-</sup>: 704.3699, found: 704.3696. **HRMS** (ESI) Macrocycle **32b**: calculated for  $C_{29}H_{44}N_5O_3S$  [M+H]<sup>+</sup>: 542.3159, found: 542.3154.









## 3.3 One-Pot Elongation/Macrocyclization Reaction with Two Monomers



## General procedure:

Reactions were prepared using 0.1 M stock solutions of all reagents in  ${}^{t}BuOH/H_{2}O$  (5:1).

Solutions of initiator **16** (10  $\mu$ L, 1.0 equiv) and the first monomer (10  $\mu$ L, 1.0 equiv) were mixed and heated at 45 °C for 6 h. A solution of the second monomer (10  $\mu$ L, 1.0 equiv) was added and the mixture heated at 45 °C for 14 h. The reaction mixture were diluted with <sup>*t*</sup>BuOH/1 M HCI (5:1) (90  $\mu$ L) and heated to 70 °C for 4 h. The resulting mixture of cyclic compounds was analyzed by HPLC and/or LCMS.

This procedure was performed on a larger scale (e.g. 0.90 mL of **16**, 0.90 mL of each monomer) for purification by preparative HPLC. For large-scale cyclization the mixtures were stirred in an oversized flask (e.g. 50 mL flask for 9 mL solvent) to enhance  $O_2$  exchange.

S27

# 3.3.1 HPLC spectra with mass traces of the major peaks

# Monomers 28 and 27



HPLC: 25 to 70% CH $_3$ CN with 0.1% TFA over 17 min



# Monomers 28 and 25



HPLC: 25 to 70%  $CH_3CN$  with 0.1% TFA over 17 min



## Monomers 25 and 27



HPLC: 25 to 70% CH<sub>3</sub>CN with 0.1% TFA over 17 min



# Monomers 27 and 25



HPLC: 25 to 70% CH<sub>3</sub>CN with 0.1% TFA over 17 min



# Monomers 27 and 28



HPLC: 25 to 70% CH<sub>3</sub>CN with 0.1% TFA over 17 min



# Monomers 28 and 30



HPLC: 30 to 90% CH<sub>3</sub>CN with 0.1% TFA over 17 min



# Monomers 28 and ent-27



HPLC: 25 to 70%  $CH_3CN$  with 0.1% TFA over 17 min



## Monomers 37 and 27



HPLC: 30 to 90% CH<sub>3</sub>CN with 0.1% TFA over 17 min



## 3.4 Isolation and Characterization of Macrocyclic Compounds

### Macrocycle 34a



Synthetic Fermentation/Macrocyclization reaction performed according to the general procedure using initiator **16** (0.09 mmol, 1.0 equiv) and monomer **28** (0.18 mmol, 2.0 equiv). The crude reaction mixture was diluted with  $CH_3CN:H_2O$  (1:1) and purified directly by preparative HPLC (20 to 95%  $CH_3CN$  with 0.1% TFA over 28 min). Macrocycle **34a** was isolated as a white powder after lyophilization. Retention time: 15.8 minutes.

<sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>) δ 9.22 (s, 1H), 9.17 (s, 1H), 8.02 (d, *J* = 9.5 Hz, 1H), 7.83 – 7.72 (m, 1H), 7.63 (d, *J* = 7.9 Hz, 1H), 7.46 (t, *J* = 7.7 Hz, 1H), 7.37 – 7.29 (m, 1H), 7.15 (s, 1H), 7.00 (d, *J* = 8.5 Hz, 2H), 6.93 (d, *J* = 8.5 Hz, 2H), 6.69 – 6.66 (m, 4H), 4.34 – 4.30 (m, 1H), 4.21-4.15 (m, 1H), 3.25 (dd, *J* = 15.8, 2.6 Hz, 1H), 2.95 (dd, *J* = 15.8, 11.9 Hz, 1H), 2.81 – 2.75 (m, 1H), 2.74 – 2.61 (m, 3H), 2.48 – 2.43 (m, 1H), 2.26 (dd, *J* = 16.5, 10.1 Hz, 1H), 2.15 (dd, *J* = 16.5, 2.2 Hz, 1H), 2.09 – 1.95 (m, 3H), 1.72 – 1.65 (m, 1H); <sup>13</sup>**C NMR** (151 MHz, DMSO-*d*<sub>6</sub>) δ 171.1, 169.9, 168.5, 167.4, 155.7, 155.6, 141.5, 131.2, 130.3, 130.2, 130.0, 129.5, 129.3, 128.6, 128.0, 122.3, 115.0, 114.9, 49.9, 47.0, 40.7, 39.3, 37.7, 33.7, 31.4, 31.4, 22.6; **HRMS** (ESI): calculated for C<sub>31</sub>H<sub>32</sub>N<sub>4</sub>NaO<sub>4</sub>S [M+Na]<sup>+</sup>: 579.2036, found: 579.2039.
#### Macrocycle 31b



Synthetic Fermentation/Macrocyclization reaction performed according to the general procedure using initiator **16** (0.10 mmol, 1.0 equiv) and monomer **25** (0.20 mmol, 2.0 equiv). The crude reaction mixture was diluted with CH<sub>3</sub>CN:H<sub>2</sub>O (1:1) and purified directly by preparative HPLC (30 to 65% CH<sub>3</sub>CN with 0.1% TFA over 28 min). Macrocycle **31b** was isolated as a white powder after lyophilization. Retention time: 17.0 minutes.

<sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>) δ 8.05 (d, *J* = 9.3 Hz, 1H), 7.90 (d, *J* = 8.2 Hz, 1H), 7.63 (s, 1H), 7.48 – 7.45 (m, 3H), 7.39 (d, *J* = 7.7 Hz, 1H), 4.16 – 4.11 (m, 1H), 3.91 – 3.82 (m, 2H), 3.37 (dd, *J* = 14.8, 3.0 Hz, 1H), 3.03 (dd, *J* = 14.8, 11.4 Hz, 1H), 2.76 – 2.71 (m, 1H), 2.67 – 2.63 (m, 1H), 2.30 (dd, *J* = 14.1, 9.7 Hz, 1H), 2.22 (dd, *J* = 14.0, 4.0 Hz, 1H), 2.13 – 1.98 (m, 4H), 1.85 – 1.75 (m, 3H), 1.73 – 1.65 (m, 2H), 0.94 (d, *J* = 6.8 Hz, 3H), 0.91 (d, *J* = 6.8 Hz, 3H), 0.82 – 0.80 (m, 6H), 0.77 (d, *J* = 6.8 Hz, 3H), 0.69 (d, *J* = 6.9 Hz, 3H); <sup>13</sup>**C NMR** (151 MHz, DMSO-*d*<sub>6</sub>) δ 171.0, 170.2, 169.9, 168.3, 167.2, 143.1, 131.2, 129.5, 129.4, 129.2, 123.4, 53.4, 51.3, 49.7, 39.9, 39.8, 39.7, 39.5, 39.4, 39.2, 39.1, 37.3, 34.8, 34.0, 32.3, 32.3, 31.8, 30.0, 27.1, 19.5, 19.2, 18.8, 18.3, 18.2, 16.4; **HRMS** (ESI): calculated for C<sub>29</sub>H<sub>44</sub>N<sub>5</sub>O<sub>3</sub>S [M+H]<sup>+</sup>: 542.3159, found: 542.3154.

Macrocycle 36a



Synthetic Fermentation/Macrocyclization reaction performed according to the general procedure using initiator **16** (0.08 mmol, 1.0 equiv) and monomer **30** (0.12 mmol, 1.5 equiv). The crude reaction mixture was diluted with  $CH_3CN:H_2O$  (1:1) and purified

directly by preparative HPLC (25 to 55%  $CH_3CN$  with 0.1% TFA over 38 min). Macrocycle **36a** was isolated as a white powder after lyophilization. Retention time: 22.5 minutes.

<sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>) δ 7.95 (d, *J* = 9.4 Hz, 1H), 7.88 (d, *J* = 7.8 Hz, 1H), 7.52 – 7.46 (m, 2H), 7.36 (d, *J* = 7.6 Hz, 1H), 7.22 (s, 1H), 4.22 – 4.14 (m, 2H), 3.46 – 3.39 (m, 1H), 2.81 – 2.75 (m, 1H), 2.74 – 2.68 (m, 1H), 2.53 – 2.51 (m, 1H), 2.29 – 2.24 (m, 2H), 2.20 – 2.16 (m, 1H), 2.12 – 2.02 (m, 3H), 1.95 – 1.64 (m, 8H), 1.62 – 1.53 (m, 2H); <sup>13</sup>**C NMR** (151 MHz, DMSO-*d*<sub>6</sub>) δ 172.1, 171.0, 170.1, 168.2, 141.6, 131.3, 131.2, 129.9, 129.2, 122.4, 56.7, 53.0, 47.0, 45.7, 31.8, 31.7, 31.2, 30.7, 27.3, 24.3, 24.2, 21.2, 20.7; **HRMS** (ESI): calculated for  $C_{23}H_{28}N_4NaO_2S$  [M+Na]<sup>+</sup>: 477.1825, found: 447.1831.

Macrocycle 38d



Synthetic Fermentation/Macrocyclization reaction performed according to the general procedure using initiator **16** (0.09 mmol, 1.0 equiv), monomer **28** (0.09 mmol, 1.0 equiv) and monomer **27** (0.09 mmol, 1.0 equiv). The crude reaction mixture was diluted with CH<sub>3</sub>CN:H<sub>2</sub>O (1:1) and purified directly by preparative HPLC (25 to 60% CH<sub>3</sub>CN with 0.1% TFA over 40 min). Macrocycle **38d** was isolated as a white powder after lyophilization. Retention time: 32.5 minutes.

<sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>) δ 9.17 (s, 1H), 7.89 (d, J = 9.7 Hz, 1H), 7.81 (d, J = 7.6 Hz, 1H), 7.61 (d, J = 7.9 Hz, 1H), 7.47 (t, J = 7.6 Hz, 1H), 7.34 (d, J = 7.6 Hz, 1H), 7.16 (s, 1H), 6.94 (d, J = 8.5 Hz, 2H), 6.66 (d, J = 8.5 Hz, 2H), 4.42 – 4.36 (m, 1H), 3.99 – 3.91 (m, 1H), 3.88 – 3.83 (m, 2H), 3.40 – 3.35 (m, 1H), 3.26 – 3.19 (m, 2H), 2.97 (dd, J = 15.9, 12.2 Hz, 1H), 2.80 – 2.71 (m, 2H), 2.67 (dd, J = 13.3, 4.3 Hz, 1H), 2.47 (dd, J = 12.3, 7.0 Hz, 2H), 2.28 – 2.25 (m, 2H), 2.06 – 1.98 (m, 3H), 1.71 – 1.60 (m, 3H), 1.51 – 1.42 (m, 1H), 1.28 – 1.19 (m, 2H); <sup>13</sup>**C NMR** (151 MHz, DMSO-*d*<sub>6</sub>) δ 171.3, 170.6, 168.6, 168.1, 155.7, 141.6, 131.3, 130.5, 130.3, 129.6, 129.4, 128.7, 122.3, 115.0, 67.0, 66.7, 52.2, 47.0, 40.0, 39.3, 37.6, 31.9, 31.5, 31.5, 29.5,

28.3, 22.7; **HRMS** (ESI): calculated for  $C_{29}H_{35}N_4O_4S$  [M+H]<sup>+</sup>: 535.2374, found: 535.2381.

Macrocycle 45b



Synthetic Fermentation/Macrocyclization reaction performed according to the general procedure using initiator **16** (0.08 mmol, 1.0 equiv), monomer **38** (0.07 mmol, 0.9 equiv) and monomer **27** (0.07 mmol, 0.9 equiv). The crude reaction mixture was diluted with  $CH_3CN:H_2O$  (1:1) and purified directly by preparative HPLC (30 to 60%  $CH_3CN$  with 0.1% TFA over 38 min). Macrocycle **45b** was isolated as a white powder after lyophilization. Retention time: 18.5 minutes.

<sup>1</sup>**H NMR** (600 MHz, DMSO-*d*<sub>6</sub>) δ 7.90 (d, *J* = 9.7 Hz, 1H), 7.81 (dt, *J* = 7.7, 1.4 Hz, 1H), 7.66 (d, *J* = 7.9 Hz, 1H), 7.47 (t, *J* = 7.7 Hz, 1H), 7.34 (dt, *J* = 7.7, 1.4 Hz, 1H), 7.29-7.27 (m, 2H), 7.22 – 7.13 (m, 4H), 4.49 – 4.44 (m, 1H), 3.99 – 3.92 (m, 1H), 3.87-3.83 (m, 2H), 3.40 – 3.36 (m, 1H), 3.28 – 3.19 (m, 2H), 2.97 (dd, *J* = 15.9, 12.3 Hz, 1H), 2.81 – 2.71 (m, 3H), 2.62 (dd, *J* = 13.2, 7.8 Hz, 1H), 2.31 – 2.29 (m, 2H), 2.07 – 1.99 (m, 3H), 1.70 – 1.64 (m, 2H), 1.64 – 1.58 (m, 1H), 1.51 – 1.46 (m, 1H), 1.27 – 1.20 (m, 2H); <sup>13</sup>**C NMR** (151 MHz, DMSO-*d*<sub>6</sub>) δ 171.4, 170.5, 168.6, 168.1, 141.6, 138.6, 131.3, 130.5, 129.6, 129.5, 129.4, 128.2, 126.1, 122.3, 66.9, 66.7, 52.2, 46.7, 40.1, 40.0 37.7, 31.9, 31.5, 31.4, 29.6, 28.3, 22.7; **HRMS** (ESI): calculated for  $C_{29}H_{34}N_4NaO_3S$  [M+Na]<sup>+</sup>: 541.2244, found: 541.2237.

#### References

1N. Kaczybura and R. Brückner, *Synthesis*, 2007, **2007**, 118–130.

- 2T. Gerfaud, Y.-L. Chiang, I. Kreituss, J. A. Russak and J. W. Bode, *Org. Process Res. Dev.*, 2012, **16**, 687–696.
- 3Y.-L. Huang and J. W. Bode, Nat. Chem., 2014, 6, 877-884.
- 4D. S. Kalinowski, P. C. Sharpe, P. V. Bernhardt and D. R. Richardson, *J. Med. Chem.*, 2007, **50**, 6212–6225.
- 5F. Thuaud, F. Rohrbacher, A. Zwicky and J. W. Bode, *Org. Lett.*, 2016, **18**, 3670–3673.
- 6F. Thuaud, F. Rohrbacher, A. Zwicky and J. W. Bode, *Helv. Chim. Acta*, 2016, **99**, 868–894.

#### NMR spectra Macrocycle 34a

### <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>)

## 





## <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>)







<sup>&</sup>lt;sup>1</sup>H-<sup>13</sup>C HSQC (600 MHz, DMSO-*d*<sub>6</sub>)



# <sup>1</sup>H-<sup>13</sup>C HMBC (600 MHz, DMSO-*d*<sub>6</sub>)



#### Macrocycle 31b

# <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>)

# D.0 -0.5 9.5 9.0 8.5 8.0 7.5 7.0 6.5 6.0 0.0 5.5 4.0 3.5 3.0 2.5 2.0 1.5 1.0 0.5 5.0 4.5 f1 (ppm) <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>)







<sup>&</sup>lt;sup>1</sup>H-<sup>13</sup>C HSQC (600 MHz, DMSO-*d*<sub>6</sub>)



# <sup>1</sup>H-<sup>13</sup>C HMBC (600 MHz, DMSO-*d*<sub>6</sub>)



## Macrocycle 36a

## <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>)







<sup>&</sup>lt;sup>1</sup>H-<sup>13</sup>C HSQC (600 MHz, DMSO-*d*<sub>6</sub>)



# <sup>1</sup>H-<sup>13</sup>C HMBC (600 MHz, DMSO-*d*<sub>6</sub>)



Ó

### Macrocycle 38d

#### <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>)

## 



# <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>)

171.30 110.58 7.168.64	 130.27 130.26 130.30 129.61 128.63 128.63	— 114.98	66.93	 
<b>0</b> -				
	ОН			

## <sup>1</sup>H-<sup>1</sup>H DQF-COSY (600 MHz, DMSO-*d*<sub>6</sub>)



<sup>1</sup>H-<sup>13</sup>C HSQC (600 MHz, DMSO-*d*<sub>6</sub>)



# <sup>1</sup>H-<sup>13</sup>C HMBC (600 MHz, DMSO-*d*<sub>6</sub>)



### Macrocycle 45b

#### <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>)





## <sup>1</sup>H-<sup>1</sup>H DQF-COSY (600 MHz, DMSO-*d*<sub>6</sub>)



<sup>1</sup>H-<sup>13</sup>C HSQC (600 MHz, DMSO-*d*<sub>6</sub>)



# <sup>1</sup>H-<sup>13</sup>C HMBC (600 MHz, DMSO-*d*<sub>6</sub>)



### 3-lodophenylhydrazide 19



### Aldehyde 21



#### Enol ester 23







### Thiohydrazide 24

### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



#### Ketoacid 16

### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



### 3-Methoxyphenyl thiohydrazide 10

#### <sup>1</sup>H NMR (400 MHz, MeOD)



### Pyridine thiohydrazide 12

#### <sup>1</sup>H NMR (400 MHz, MeOD)



### Alkyl thiohydrazide 15

#### <sup>1</sup>H NMR (400 MHz, MeOD)



### Phenyl thiadiazole 5

<sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>)





#### Propanoic acid thiadiazole 7

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

#### 



## <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



#### **Fmoc-aminothiadiazole 9**

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



### 3-Methoxyphenyl thiadiazole 11

# <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>)



### 2,3-Dihydrothiadiazole carboxylic acid 4

# <sup>1</sup>H NMR (400 MHz, CDCI<sub>3</sub>)



#### *n*-Propyl monomer intermediate 51

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



#### n-Propyl monomer 26

#### <sup>1</sup>H NMR (400 MHz, MeOD)



#### Pyran monomer intermediate 52

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



#### Pyran monomer 27

#### <sup>1</sup>H NMR (400 MHz, MeOD)

#### 



# <sup>13</sup>C NMR (101 MHz, MeOD)



#### 4-Benzyloxybenzyl intermediate 53

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)



<sup>&</sup>lt;sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)


#### 4-Hydroxybenzyl monomer intermediate 54

#### <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

#### 



# <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



# 4-Hydroxybenzyl monomer 28

## <sup>1</sup>H NMR (400 MHz, MeOD)



#### Amide monomer intermediate 55

## <sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>)

## 



# <sup>13</sup>C NMR (101 MHz, CDCl<sub>3</sub>)



#### Amide monomer 29

#### <sup>1</sup>H NMR (400 MHz, MeOD)

#### 



# <sup>13</sup>C NMR (101 MHz, MeOD)

