

Lipid Bilayer Environments Control Exchange Kinetics of Deep Cavitand Hosts and Enhance Disfavored Guest Conformations

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Electronic Supplementary Information

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1. NMR Spectra of New Compounds

Guest 3

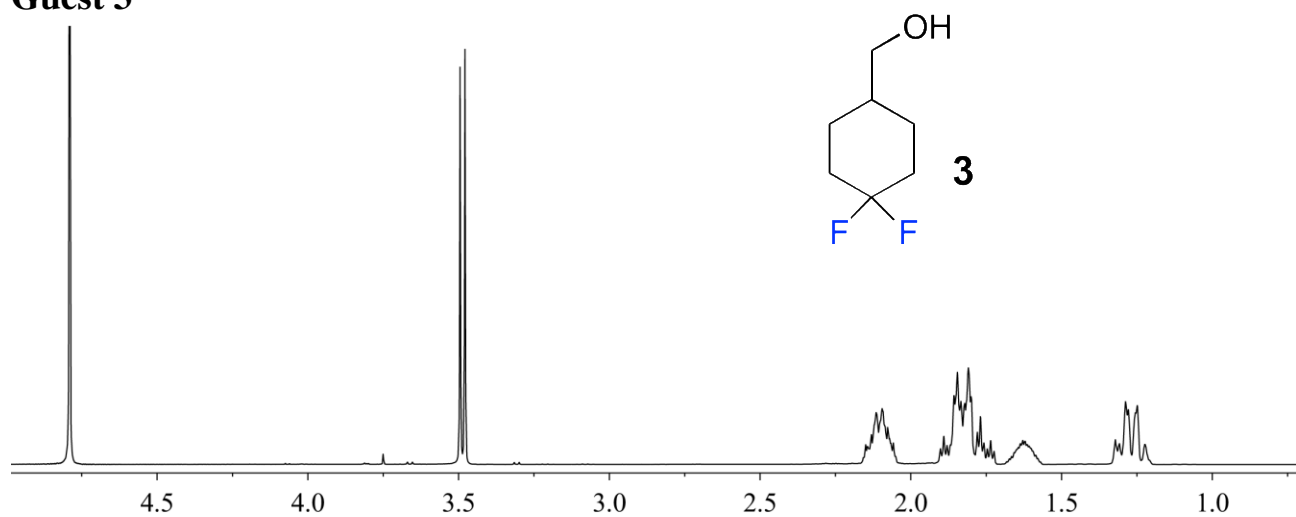


Figure S-1. ¹H NMR spectrum of 4,4-difluorocyclohexanemethanol guest **3** (400.13 MHz, D₂O, 298 K).

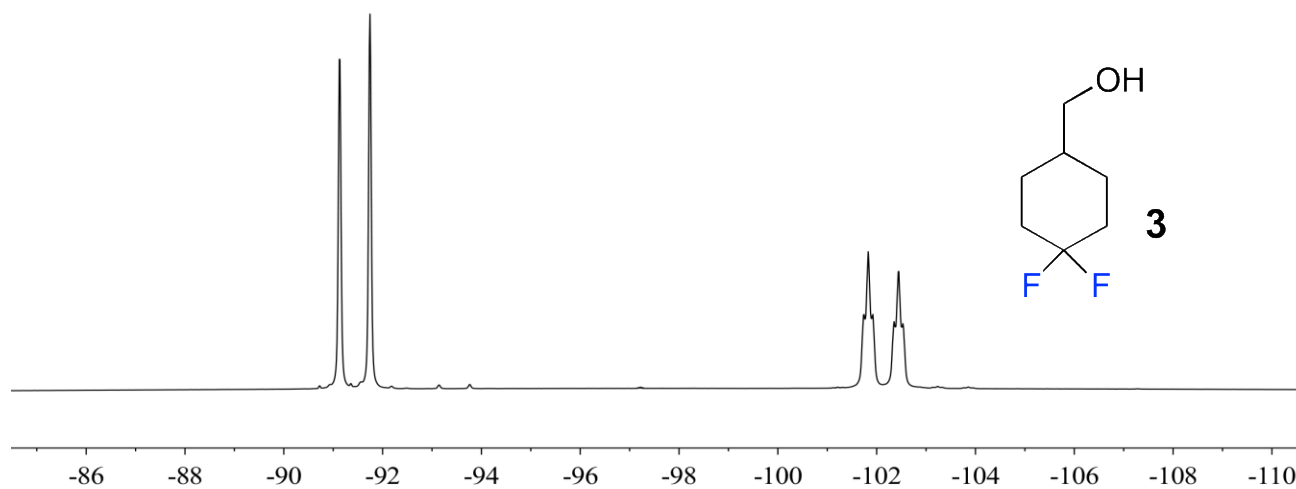


Figure S-2. ¹⁹F NMR spectrum of 4,4-difluorocyclohexanemethanol guest **3** (376.50 MHz, D₂O, 298 K).

Guest 4

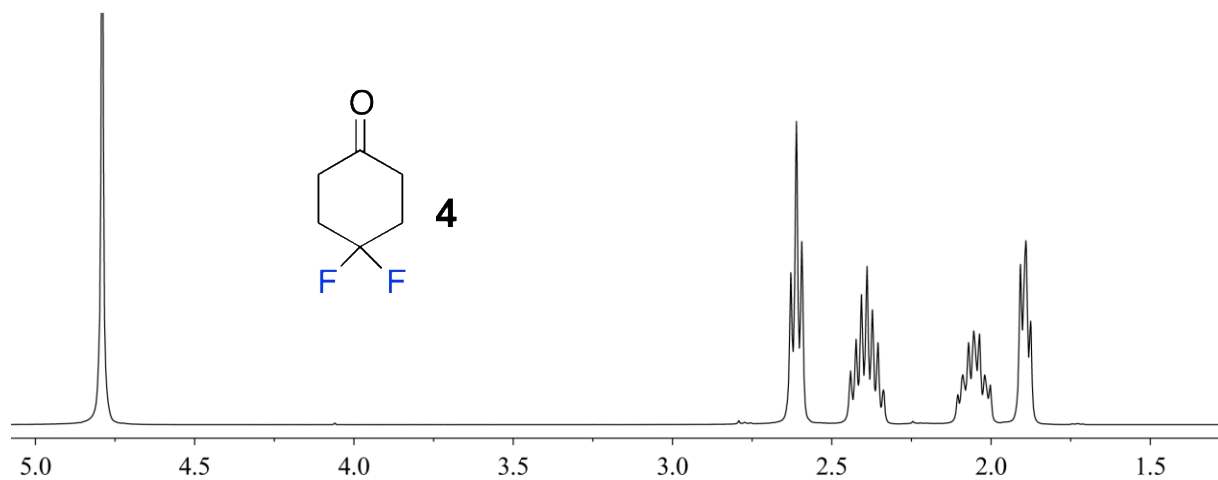


Figure S-3. ¹H NMR spectrum of 4,4-difluorocyclohexanone guest **4** (400.13 MHz, D₂O, 298 K).

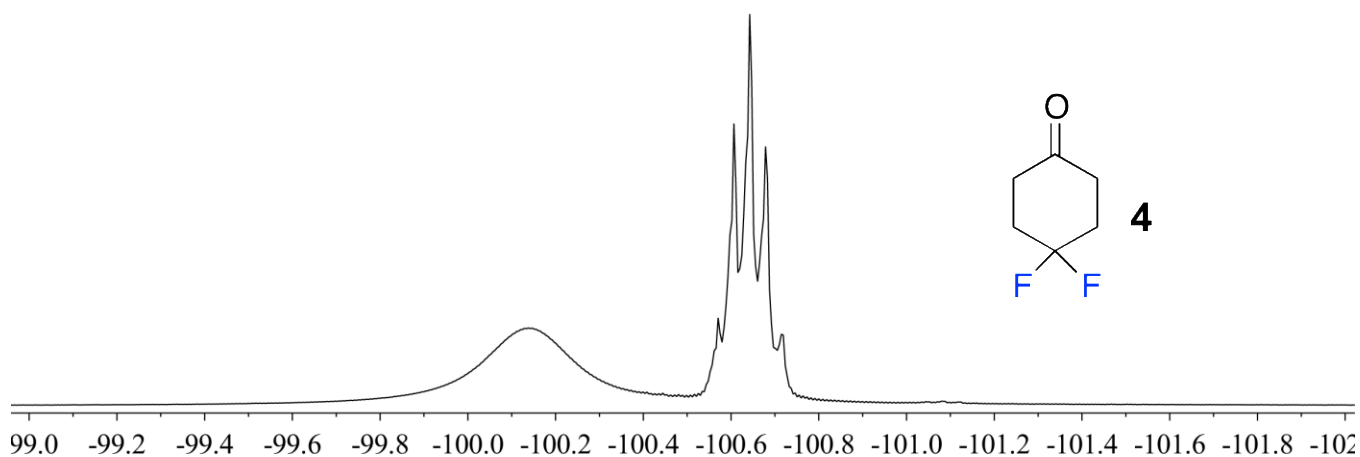


Figure S-4. ^{19}F NMR spectrum of 4,4-difluorocyclohexanone guest **4** (376.50 MHz, D_2O , 298 K).

Guest 5

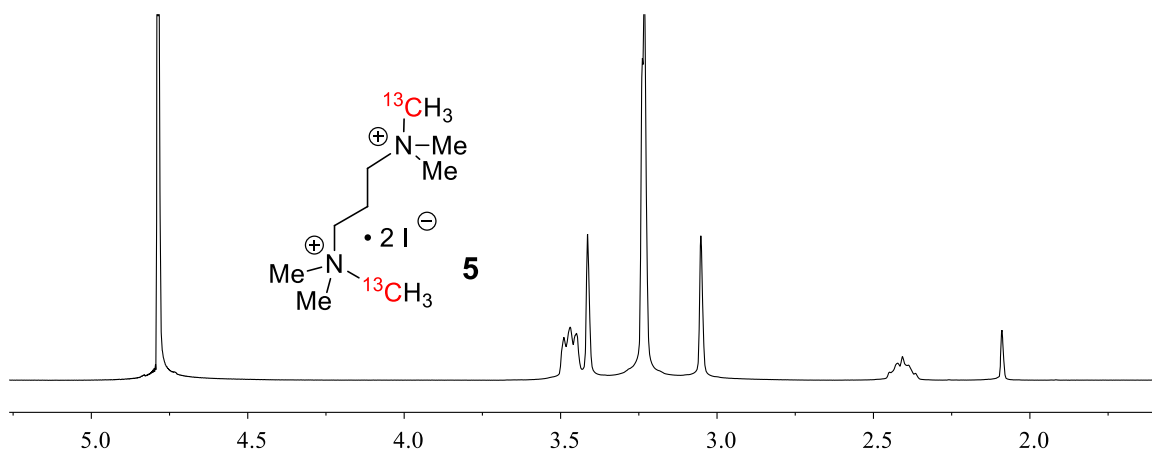


Figure S-5. ^1H NMR spectrum of N,N,N,N',N',N'-hexamethyl-1,3-propanediaminium diiodide- ^{13}C guest **5** (400.13 MHz, D_2O , 298 K).

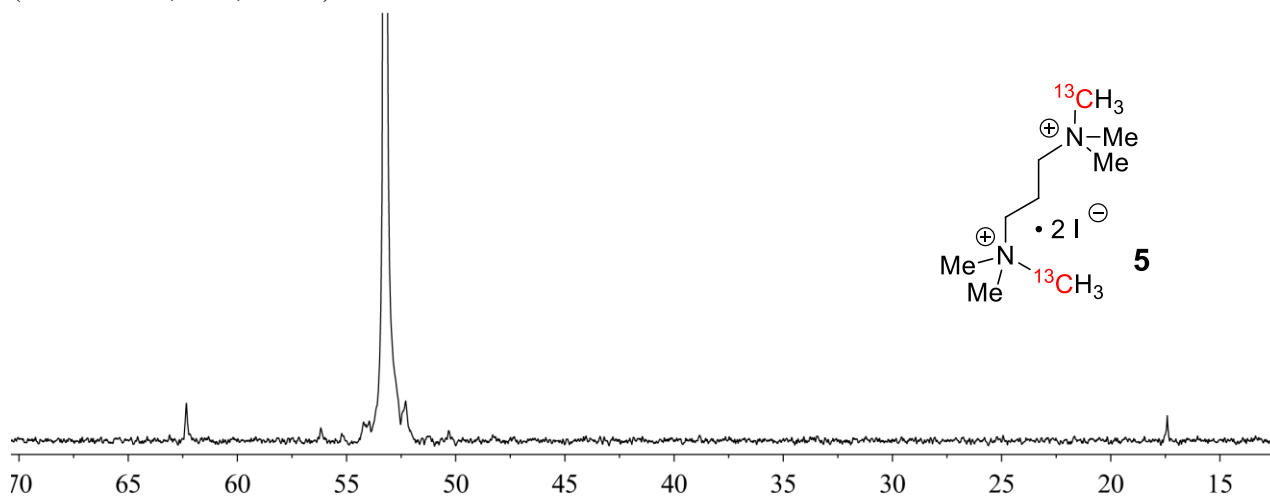


Figure S-6. ^{13}C NMR spectrum of N,N,N,N',N',N'-hexamethyl-1,3-propanediaminium diiodide- ^{13}C guest **9** (100.61 MHz, D_2O , 298 K).

Guest 6

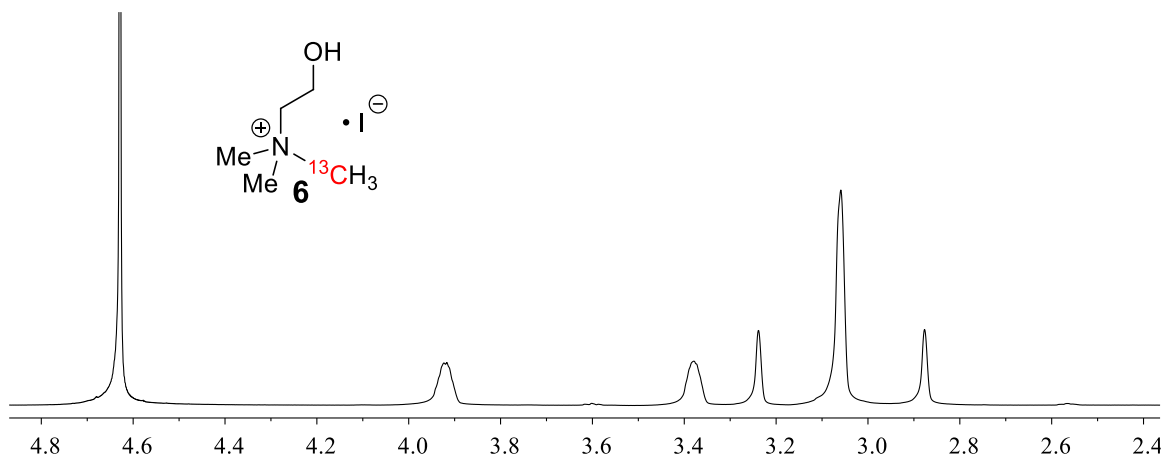


Figure S-7. ¹H NMR spectrum of choline iodide-¹³C guest **6** (400.13 MHz, D₂O, 298 K).

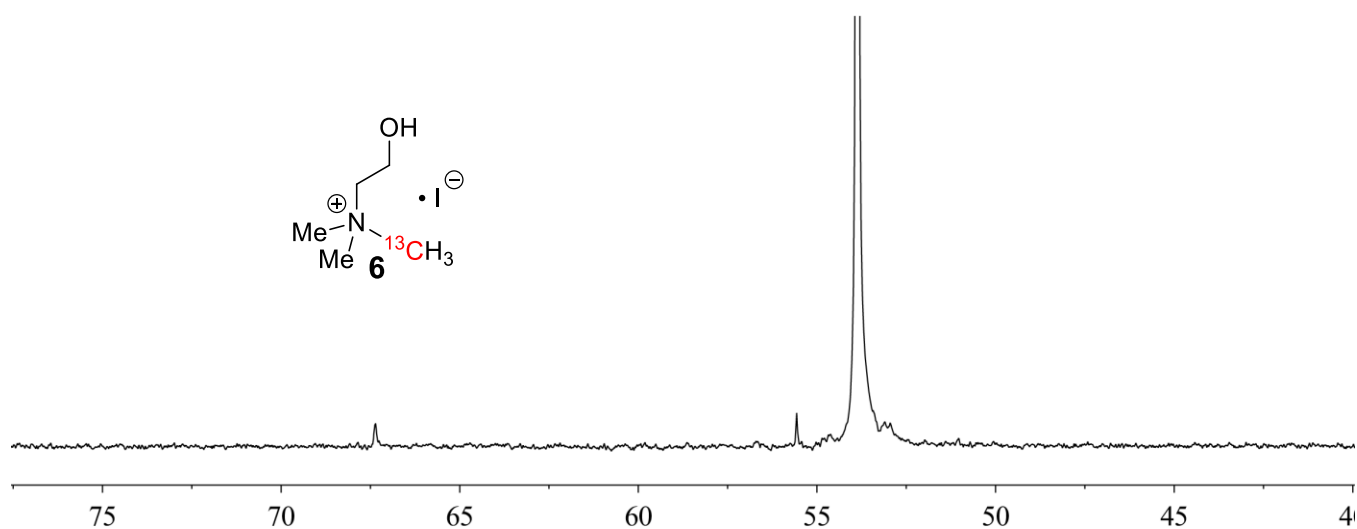


Figure S-8. ¹³C NMR spectrum of choline iodide-¹³C guest **6** (100.61 MHz, D₂O, 298 K).

Guest 7

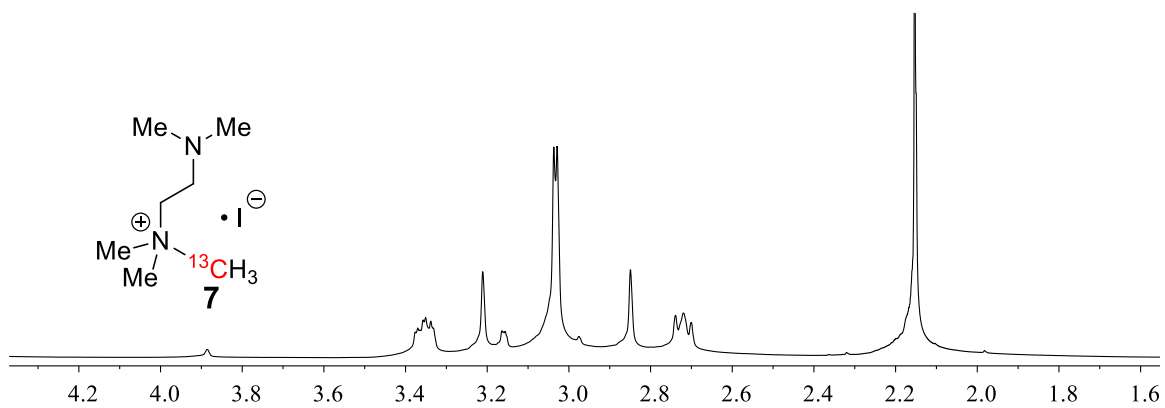


Figure S-9. ¹H NMR spectrum of N,N,N,N',N'-pentamethyl-1,2-ethanediaminium iodide-¹³C guest **7** (400.13 MHz, D₂O, 298 K).

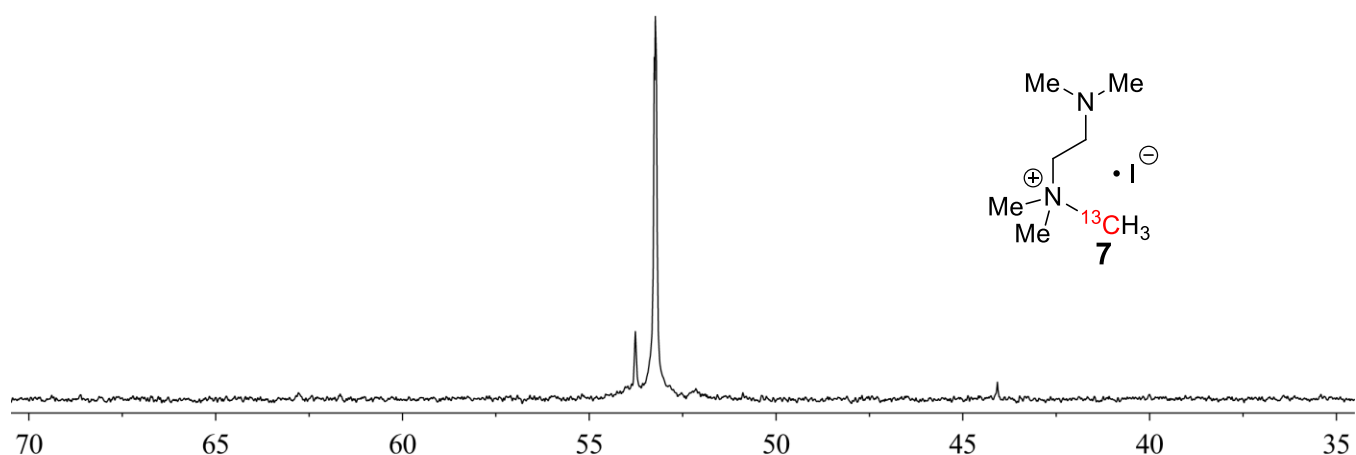


Figure S-10. ^{13}C NMR spectrum of N, N, N, N', N'-pentamethyl-1,2-ethanediaminium iodide- ^{13}C guest **7** (100.61 MHz, D_2O , 298 K).

Guest 8

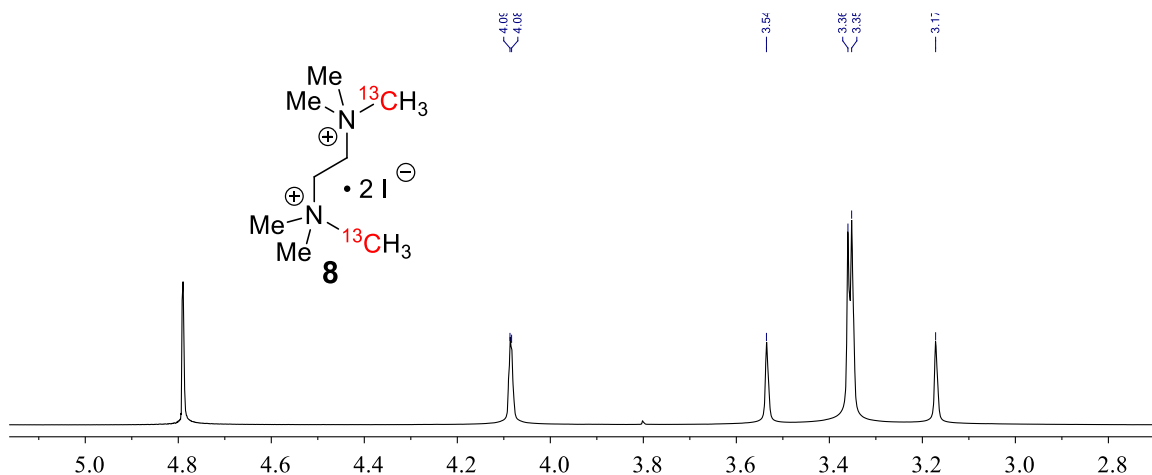


Figure S-11. ^1H NMR spectrum of N,N,N,N',N',N'-hexamethyl-1,2-ethanediaminium diiodide- ^{13}C guest **8** (400.13 MHz, D_2O , 298 K).

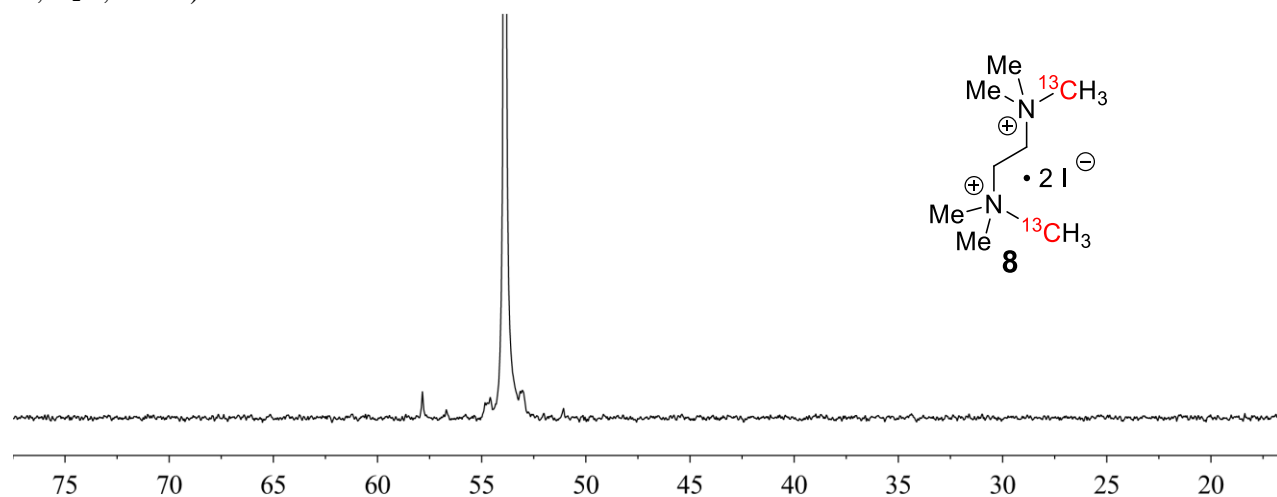


Figure S-12. ^{13}C NMR spectrum of N,N,N,N',N',N'-hexamethyl-1,2-ethanediaminium diiodide- ^{13}C guest **8** (100.61 MHz, D_2O , 298 K).

2. NMR Spectra of Host:Guest Complexes in Free Aqueous Solution

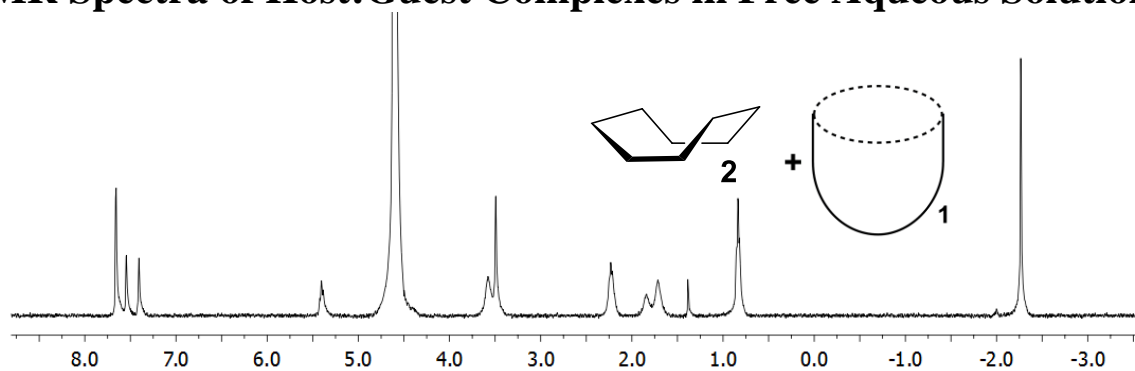


Figure S-13. ¹H NMR spectrum of the **1•2** cavitand-cyclooctane complex (400.13 MHz, D₂O, 298 K, [**1**] = [**2**] = 1.8 mM).

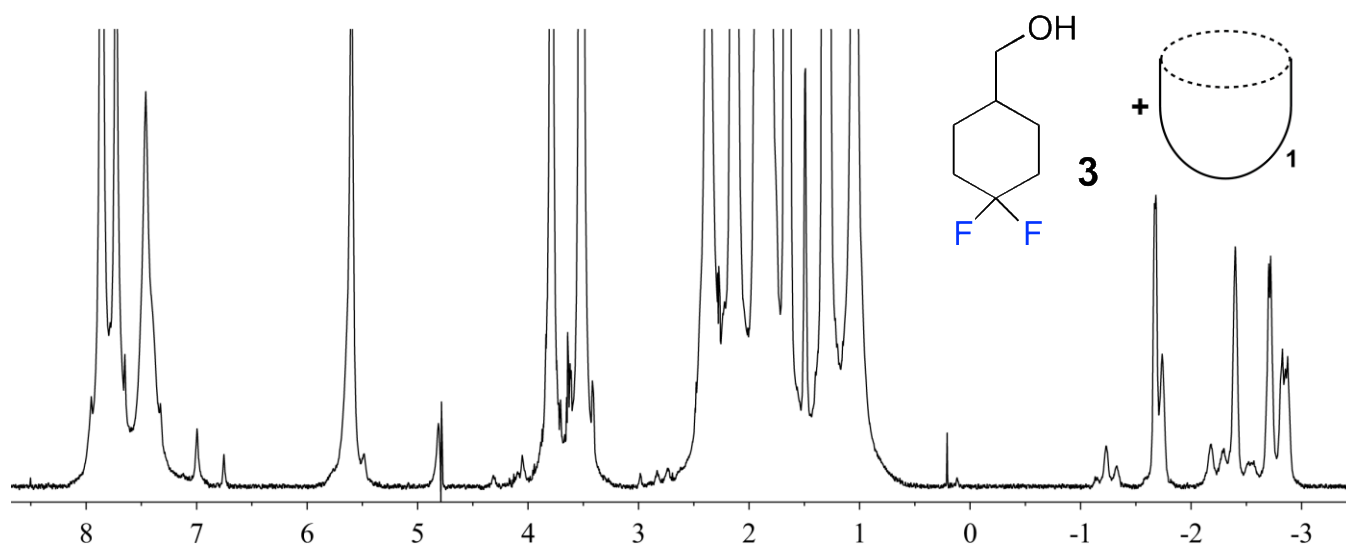


Figure S-14. ¹H NMR spectrum of the **1•3** cavitand-guest **3** complex (400.13 MHz, D₂O, 298 K, [**1**] = 5.8 mM, [**3**] = 39.5 mM).

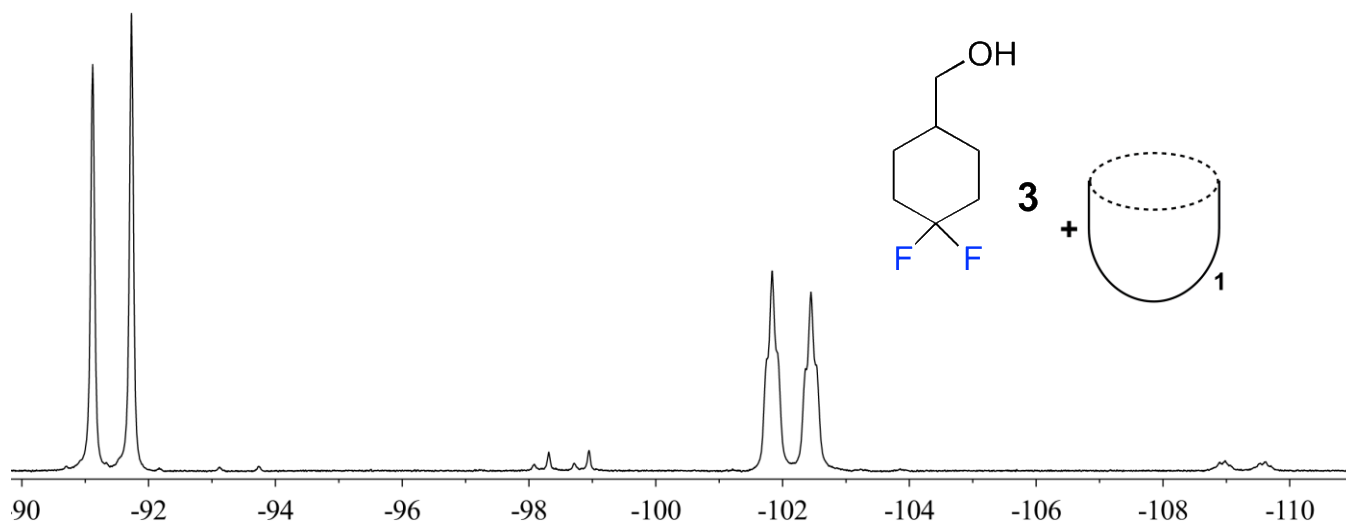


Figure S-15. ¹⁹F NMR spectrum of the **1•3** cavitand-guest **3** complex (376.50 MHz, D₂O, 298 K, [**1**] = 5.8 mM, [**3**] = 39.5 mM).

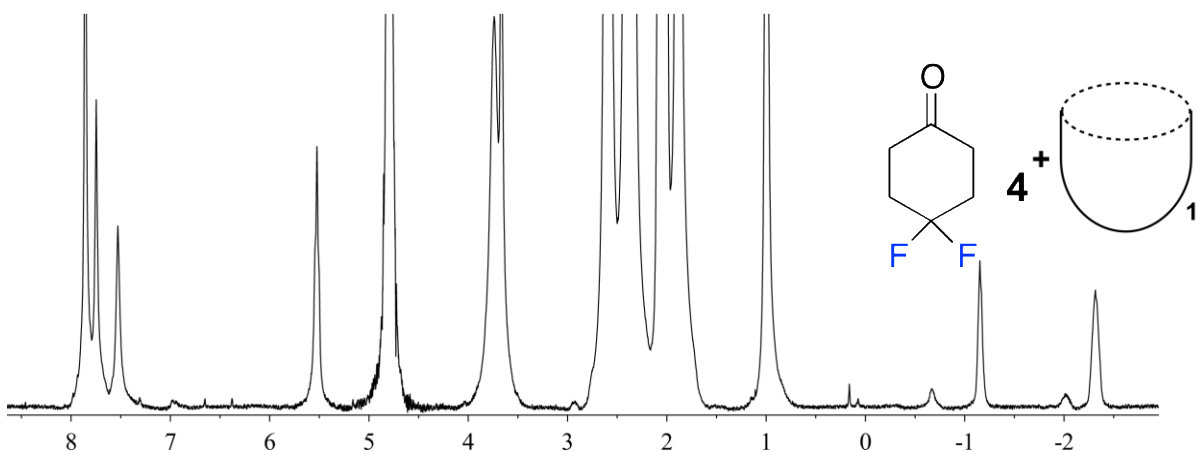


Figure S-16. ^1H NMR spectrum of the **1•4** cavitand-guest **4** complex (400.13 MHz, D_2O , 298 K, [**1**] = 5.8 mM, [**4**] = 39.5 mM).

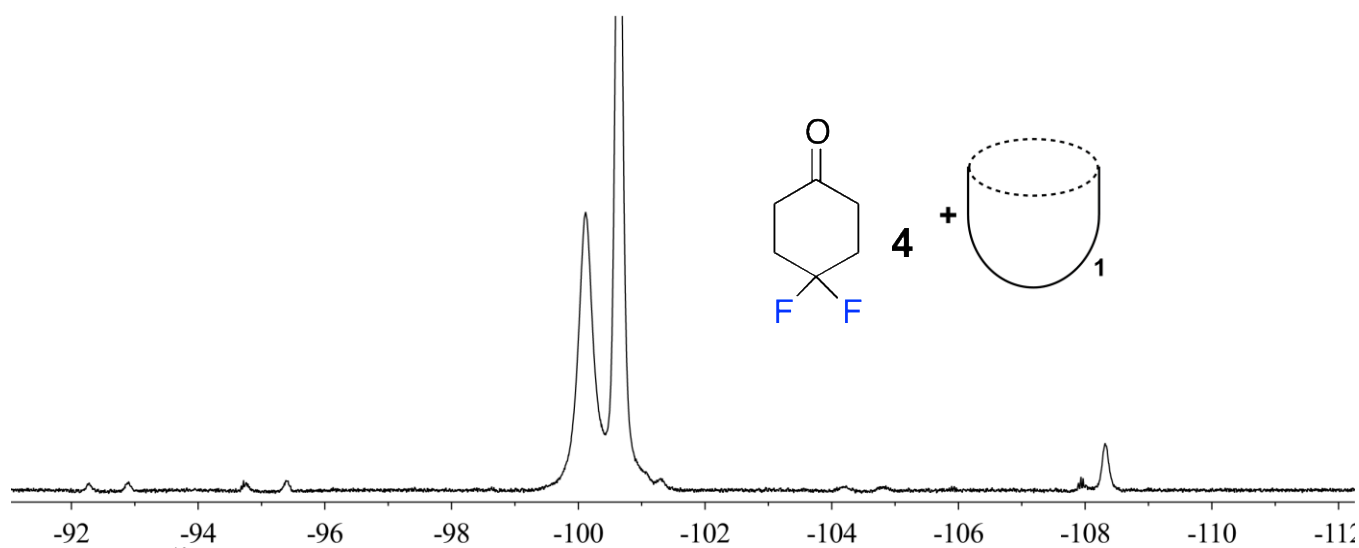


Figure S-17. ^{19}F NMR spectrum of the **1•4** cavitand-guest **4** complex (400.13 MHz, D_2O , 298 K, [**1**] = 5.8 mM, [**4**] = 39.5 mM).

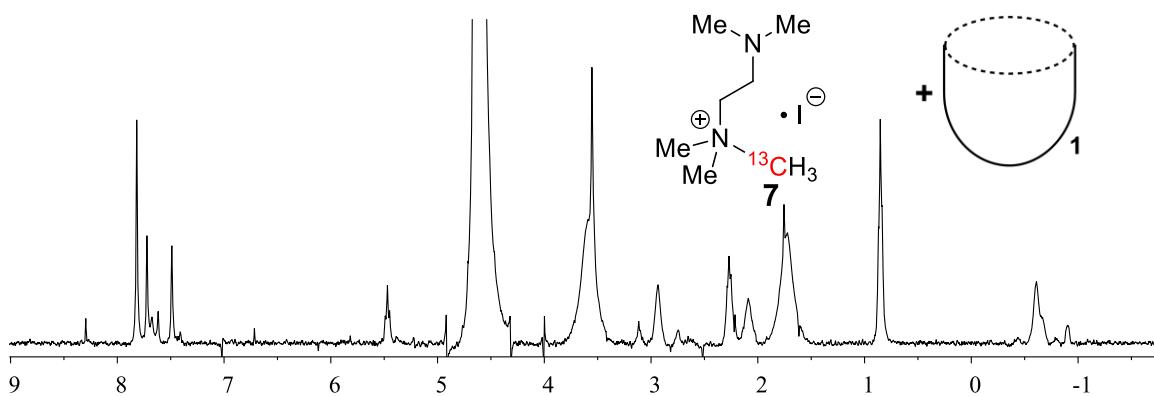


Figure S-18. ^1H NMR spectrum of the **1•7** cavitand-guest **7** complex (400.13 MHz, D_2O , 298 K, [**1**] = 1.8 mM, [**7**] = 2.2 mM).

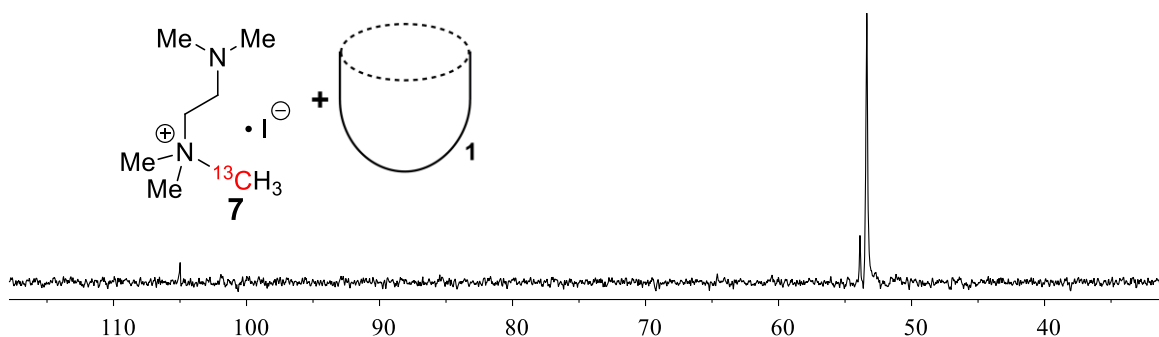


Figure S-19. ^{13}C NMR spectrum of the **1•7** cavitand-guest **7** complex (100.61 MHz, D_2O , 298 K, $[\mathbf{1}] = 1.8$ mM, $[\mathbf{7}] = 2.2$ mM).

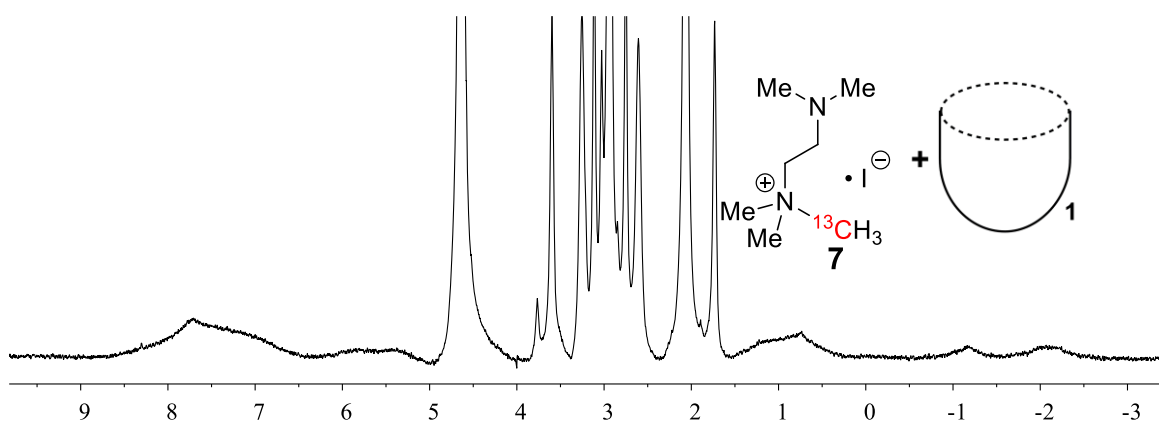


Figure S-20. ^1H NMR spectrum of the **1•7** cavitand-guest **7** complex in the presence of excess **7** (400.13 MHz, D_2O , 298 K, $[\mathbf{1}] = 1.8$ mM, $[\mathbf{7}] = 10$ mM).

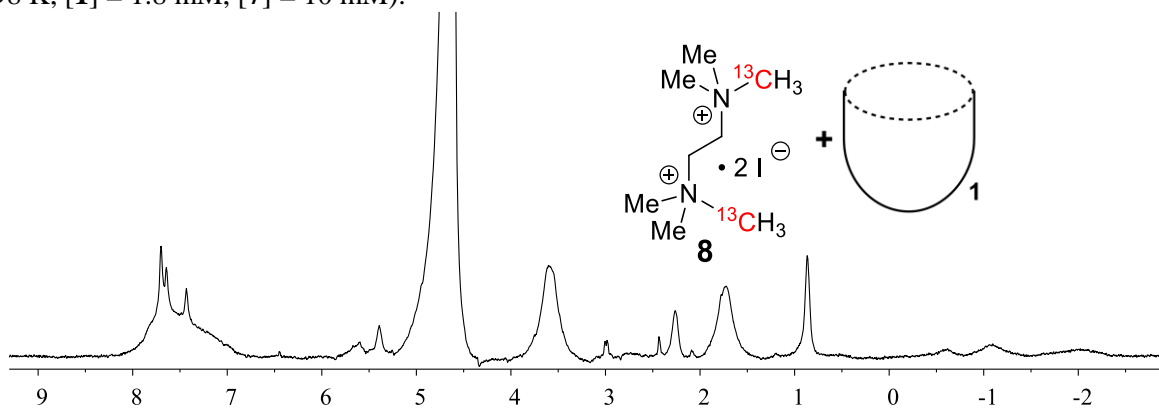


Figure S-21. ^1H NMR spectrum of the **1•8** cavitand-guest **8** complex (400.13 MHz, D_2O , 298 K, $[\mathbf{1}] = 1.8$ mM, $[\mathbf{8}] = 2.2$ mM).

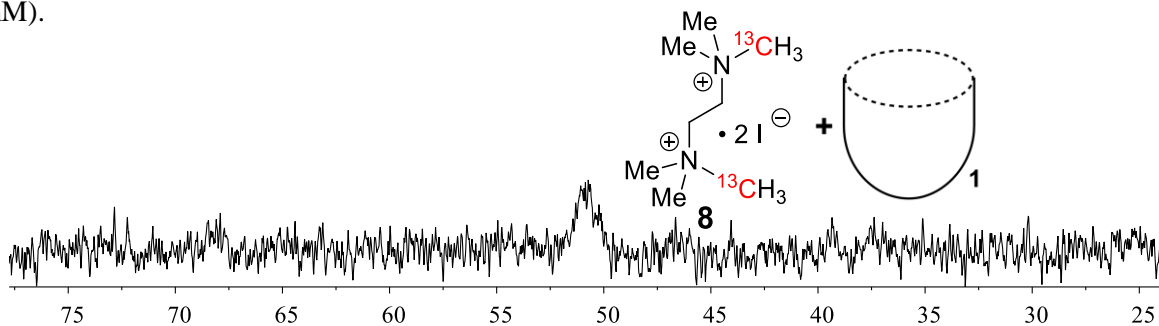


Figure S-22. ^{13}C NMR spectrum of the **1•8** cavitand-guest **8** complex (100.61 MHz, D_2O , 298 K, $[\mathbf{1}] = 1.8$ mM, $[\mathbf{8}] = 2.2$ mM).

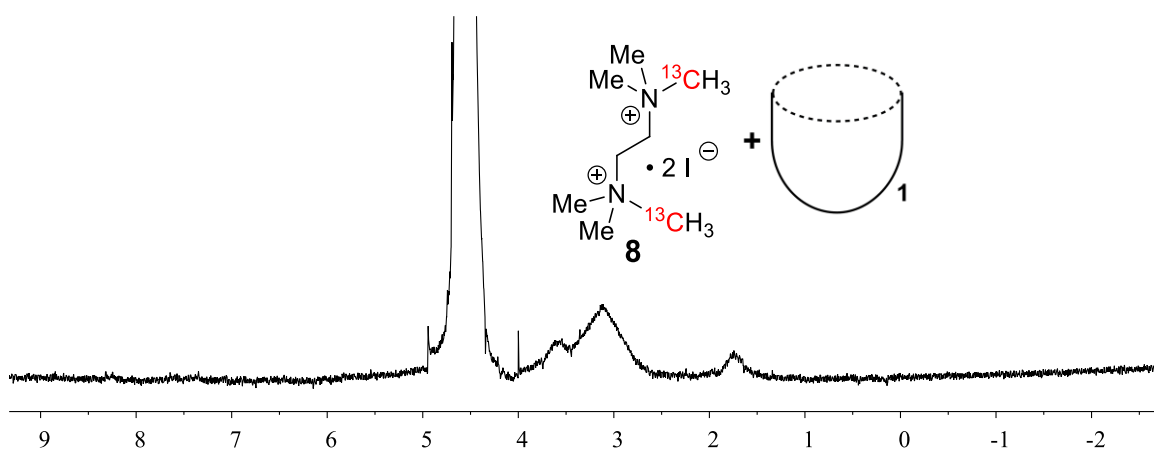


Figure S-23. ^1H NMR spectrum of the **1•8** cavitand-guest **8** complex in the presence of excess **8** (400.13 MHz, D_2O , 298 K, [**1**] = 1.8 mM, [**8**] = 5.5 mM).

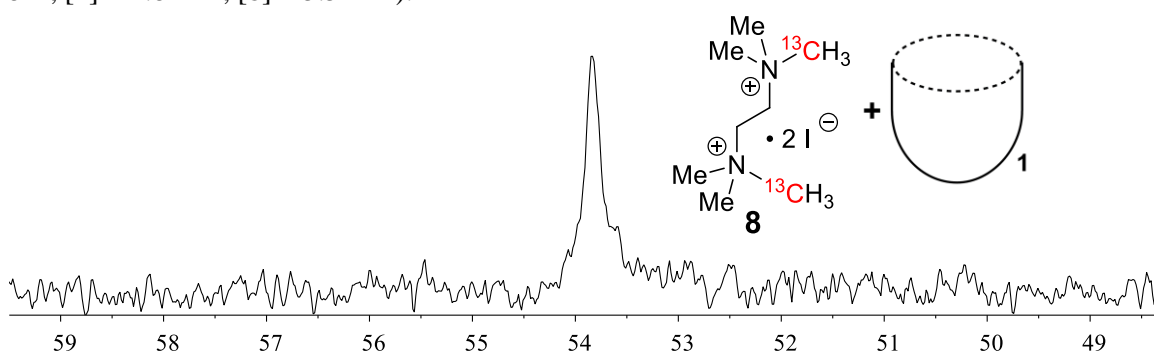


Figure S-24. ^{13}C NMR spectrum of the **1•8** cavitand-guest **8** complex in the presence of excess **8** (100.61 MHz, D_2O , 298 K, [**1**] = 1.8 mM, [**8**] = 5.5 mM).

3. NMR Spectra of Host:Guest Complexes in DMPC/DHPC Lipid Micelles.

Guest 2

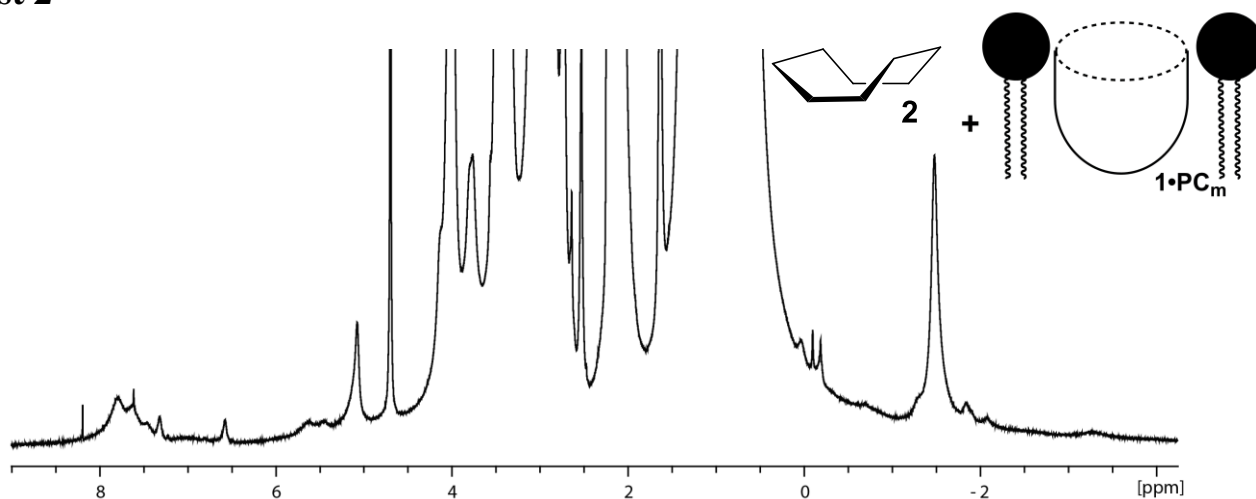


Figure S-25. ^1H NMR spectrum of the **1•2** cavitand-cyclooctane complex in DMPC:DHPC lipid micelles (599.88 MHz, 1 mM HEPES/ D_2O , 298 K, [**1**] = [**2**] = 1.8 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

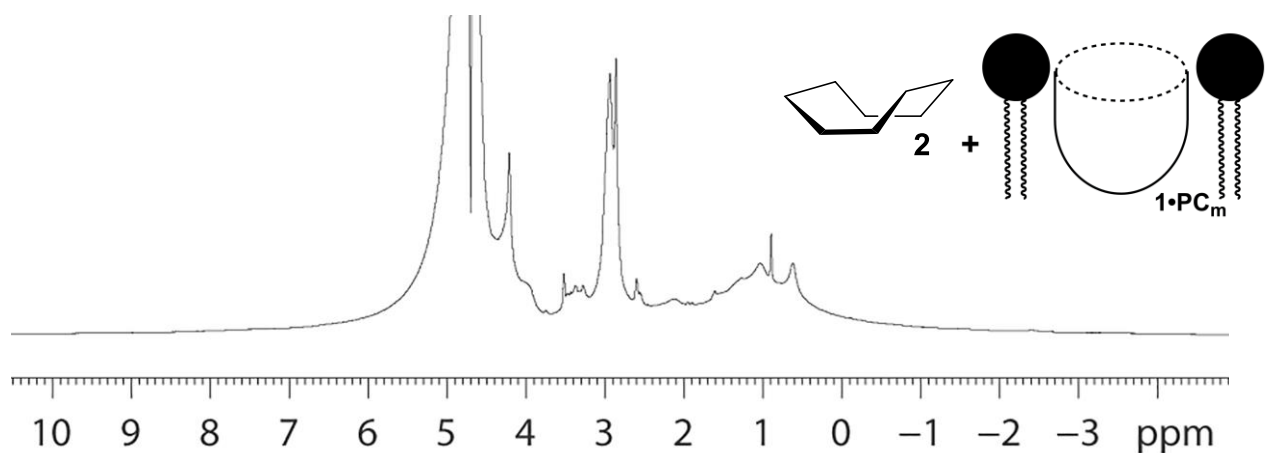


Figure S-26. T2-filtered ^1H NMR spectrum of the $1\cdot 2$ cavitand-cyclooctane complex in DMPC:DHPC lipid micelles (599.88 MHz, 1 mM HEPES/D $_2$ O, 298 K, $[1] = [2] = 1.8$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

Guest 3

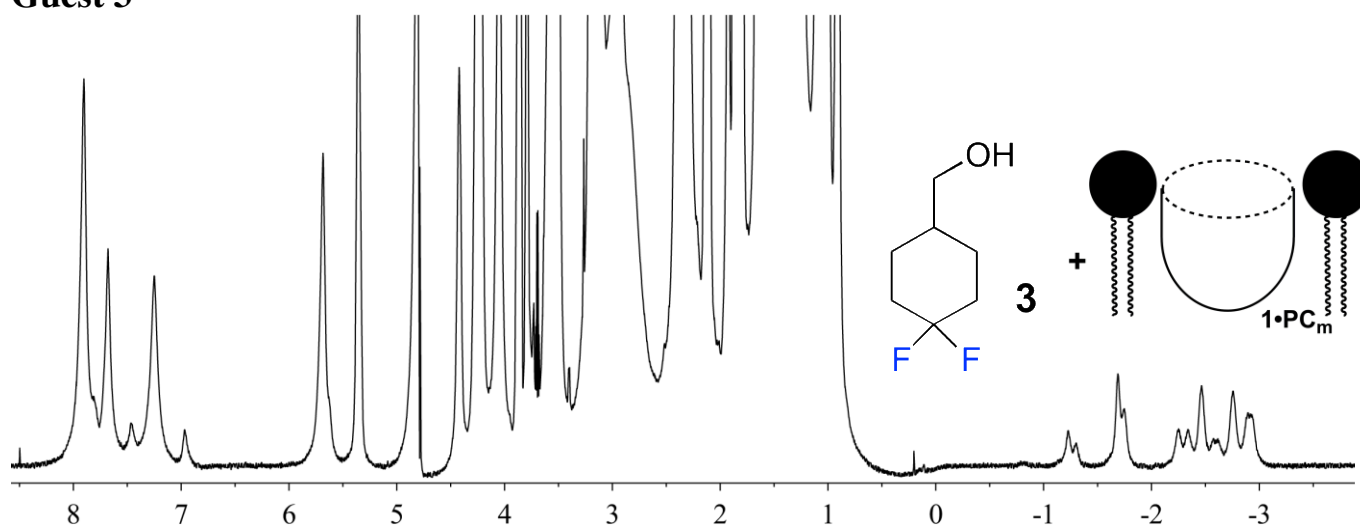


Figure S-27. ^1H NMR spectrum of the cavitand $1\cdot$ guest 3 complex in PC $_m$ micelles (599.88 MHz, 1 mM HEPES/D $_2$ O, 283 K, $[1] = 5.8$ mM, $[3] = 39.5$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

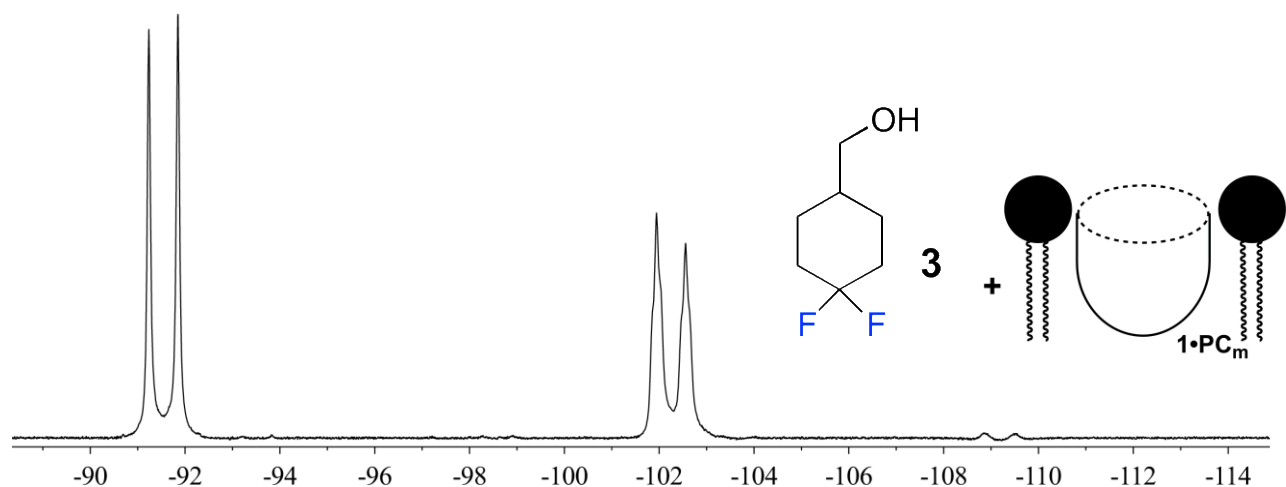


Figure S-28. ^{19}F NMR spectrum of the cavitand $1\cdot$ guest 3 complex in PC $_m$ micelles (1 mM HEPES/D $_2$ O, 376.50 MHz, 298 K, $[1] = 5.8$ mM, $[3] = 39.5$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

Guest 4

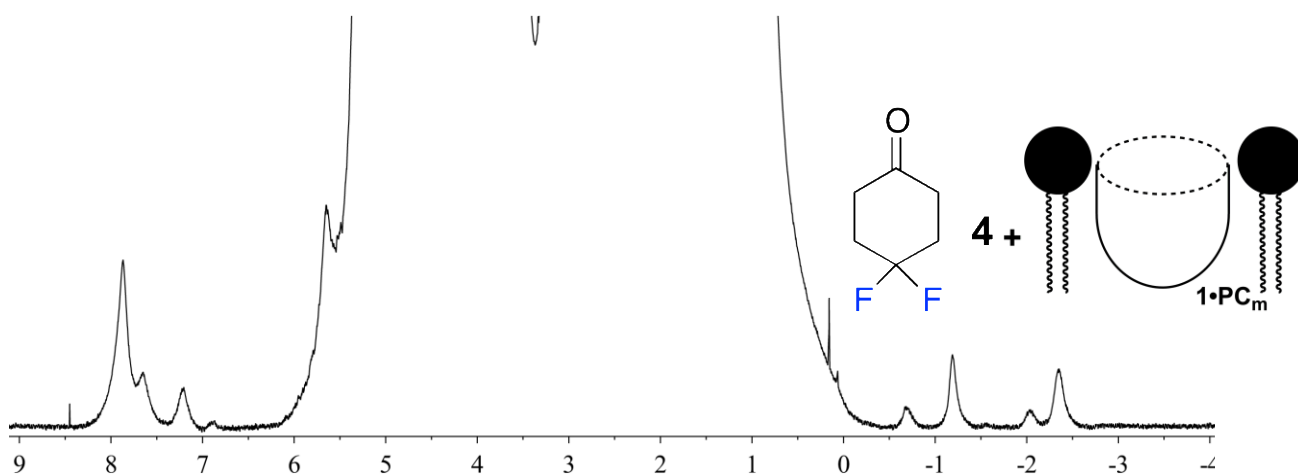


Figure S-29. ¹H NMR spectrum of the cavitaand **1**•guest **4** complex in PC_m micelles (599.88 MHz, 1 mM HEPES/D₂O, 283 K, [**1**] = 5.8 mM, [**4**] = 39.5 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

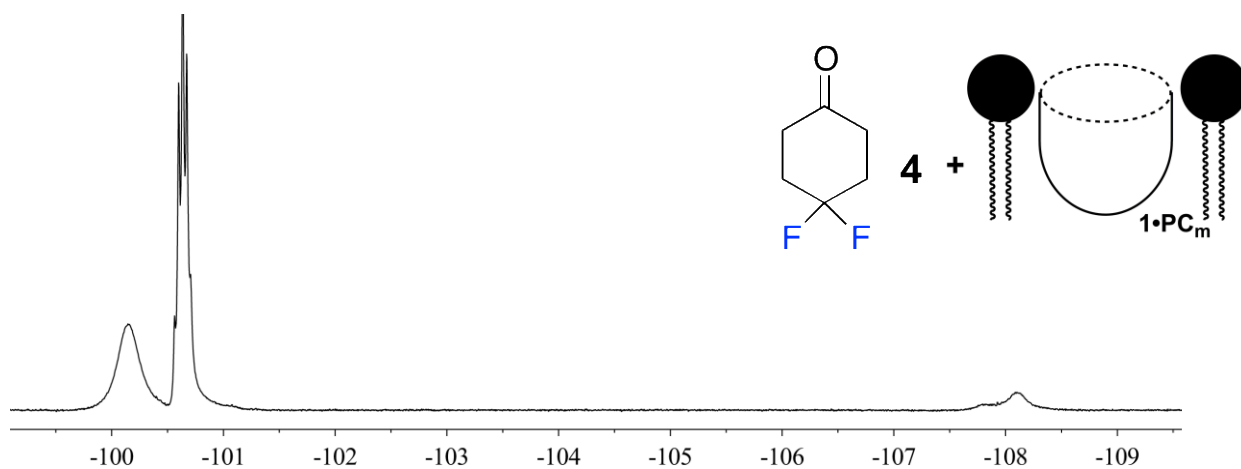


Figure S-30. ¹⁹F NMR spectrum of the cavitaand **1**•guest **4** complex in PC_m micelles (1 mM HEPES/D₂O, 376.50 MHz, 298 K, [**1**] = 5.8 mM, [**4**] = 39.5 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration);

Guest 6

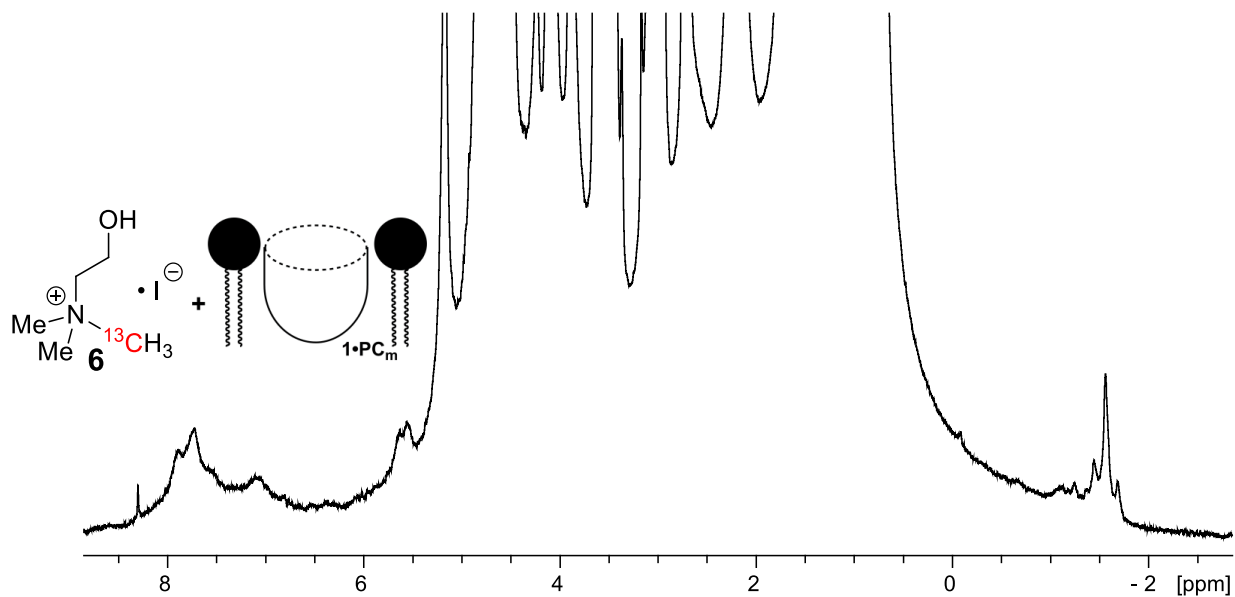


Figure S-31. ^1H NMR spectrum of the $1\cdot 6$ cavitand-choline **6** complex in DMPC:DHPC lipid micelles (599.88 MHz, 1 mM HEPES/D₂O, 283 K, [**1**] = 5.8 mM, [**6**] = 16.0 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

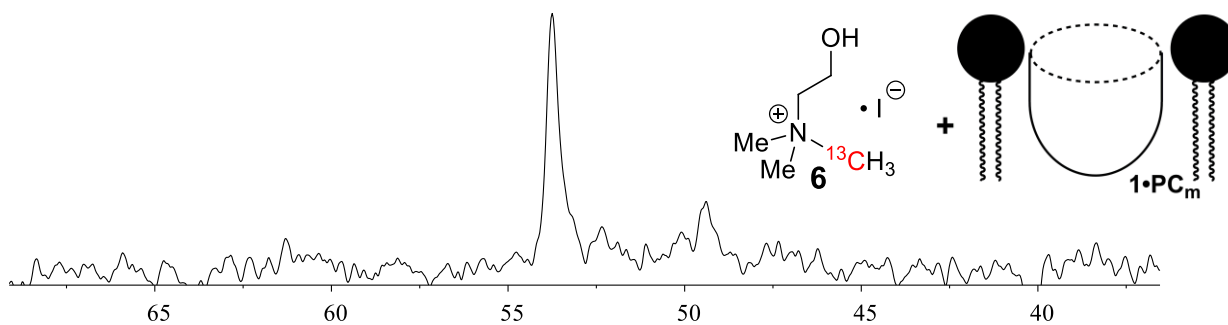


Figure S-32. ^{13}C NMR spectrum of the $1\cdot 6$ cavitand-choline **6** complex in DMPC:DHPC lipid micelles (100.61 MHz, 1 mM HEPES/D₂O, 283 K, [**1**] = 5.8 mM, [**6**] = 16.0 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

Guest 7

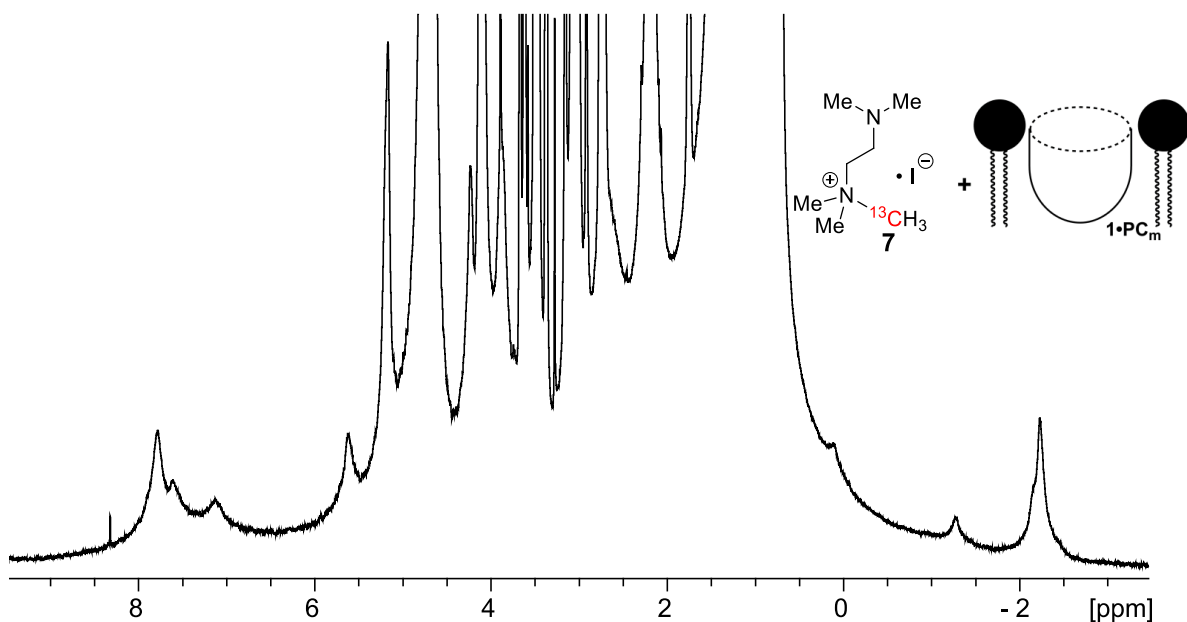


Figure S-33. ^1H NMR spectrum of the $1\cdot 7$ cavitand-guest 7 complex in DMPC:DHPC lipid micelles (599.88 MHz, 1 mM HEPES/D₂O, 283 K, [1] = 5.8 mM, [7] = 16 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

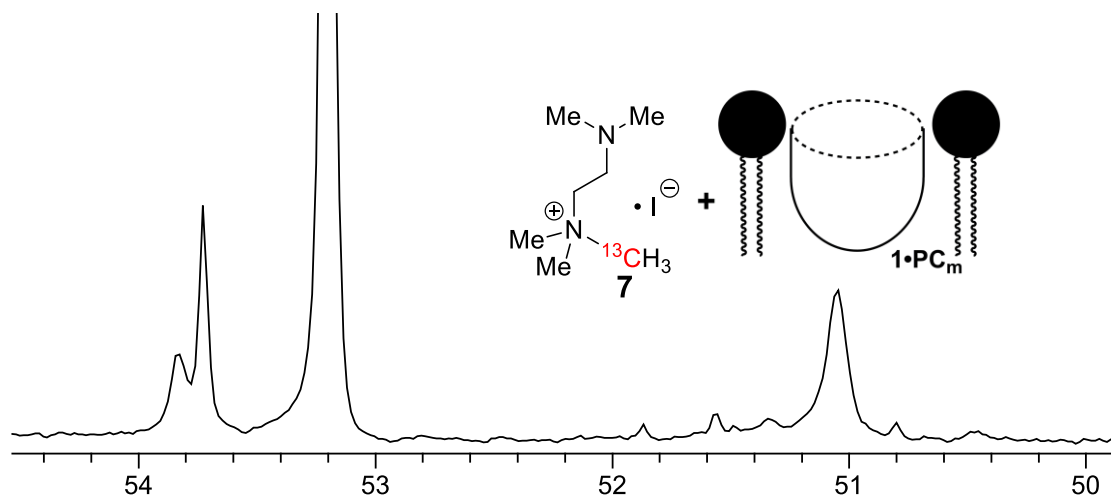


Figure S-34. ^{13}C NMR spectrum of the $1\cdot 7$ cavitand-guest 7 complex in DMPC:DHPC lipid micelles (100.61 MHz, 1 mM HEPES/D₂O, 283 K, [1] = 5.8 mM, [7] = 16 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

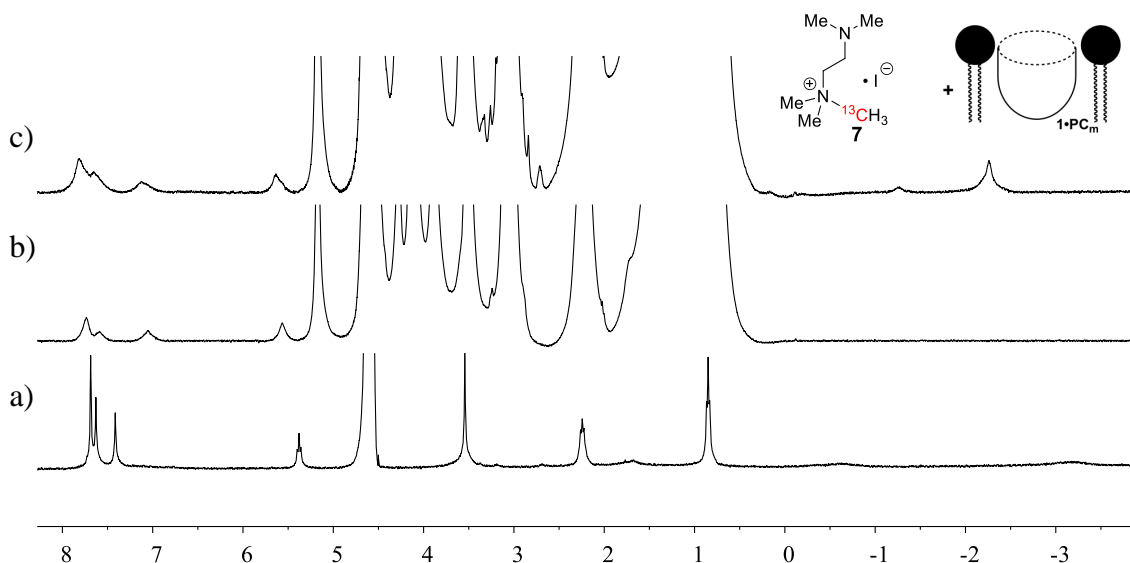


Figure S-35. ^1H NMR spectra of the sequential addition of lipids and guest **7** to a solution of cavitant **1** (400.13 MHz, 1 mM HEPES/D $_2$ O, 298 K). a) 1.8 mM of cavitant **1**; b) 1.8 mM of cavitant **1** + DMPC/DHPC lipids, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration; c) 1.8 mM of cavitant **1** + DMPC/DHPC lipids = 2.2 mM guest **7**.

Guest 8

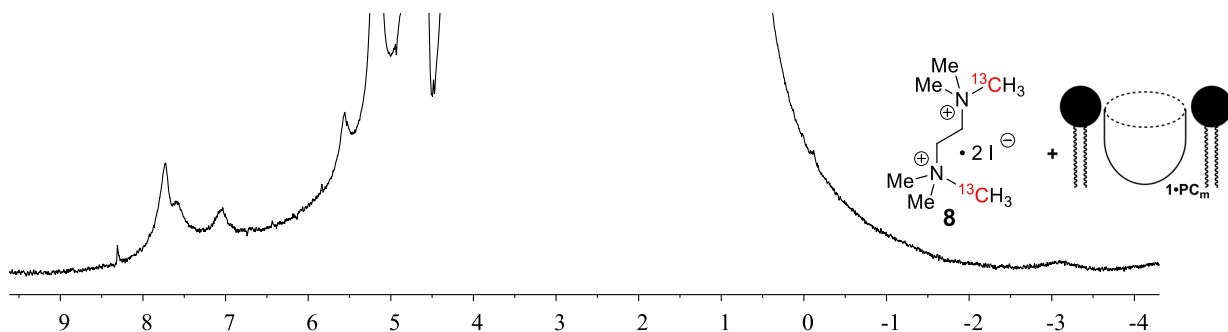


Figure S-36. ^1H NMR spectrum of the **1•8** cavitant-guest **8** complex in DMPC:DHPC lipid micelles (400.13 MHz, 1 mM HEPES/D $_2$ O, 298 K, [**1**] = 1.8 mM, [**8**] = 5.4 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

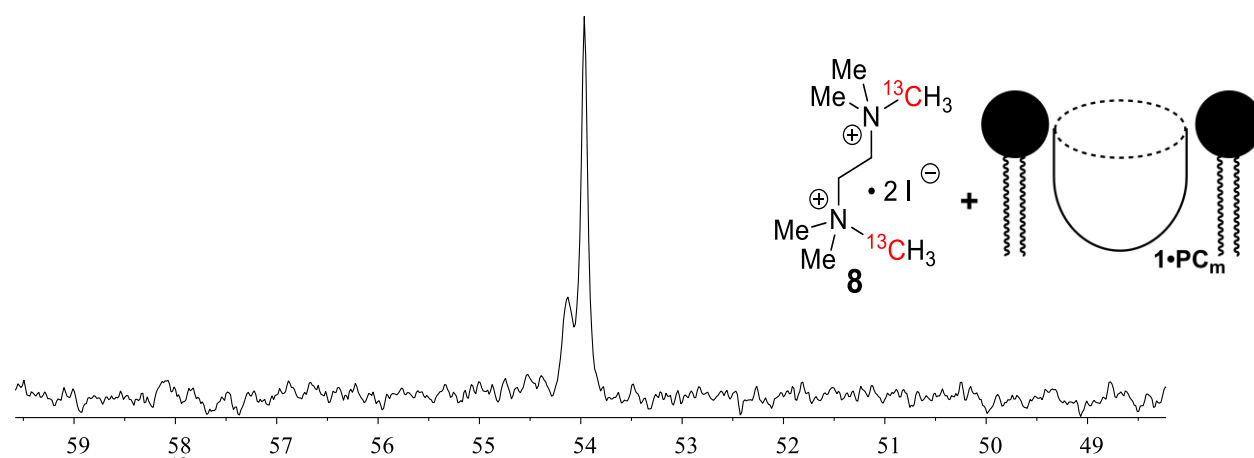
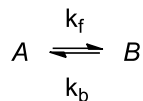


Figure S-37. ^{13}C NMR spectrum of the **1•8** cavitant-guest **8** complex in DMPC:DHPC lipid micelles (100.61 MHz, 1 mM HEPES/D $_2$ O, 298 K, [**1**] = 1.8 mM, [**8**] = 5.4 mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration).

4. Fitting model for EXSY Experiments

A standard two-site exchange model is used to analyze the EXSY data. In this model, the guest can exchange between a free and bound state, A and B, respectively:



k_b is the exchange constant from B to A, while k_f is the exchange constant for A to B. At equilibrium, the forward and reverse exchange rates are equal. Therefore $k_b[B] = k_f[A]$, or in terms of the equilibrium constant:

$$\frac{[B]}{[A]} = \frac{k_f}{k_b} = K_{eq},$$

During the mixing time of the EXSY experiment, the concentration of A can change in three ways: first, A can become bound, decreasing its contribution to the signal intensity over the mixing time; second, A can relax due to T_1 , also decreasing its intensity; third, a bound ligand B can be released, becoming A and adding to its signal intensity. This can be described by the following first-order differential equation:

$$\frac{d[A]}{dt} = -k_f[A] - \frac{[A]}{T_{1A}} + k_b[B]$$

The signal from B varies in a similar way, giving

$$\frac{d[B]}{dt} = -k_b[B] - \frac{[B]}{T_{1B}} + k_f[A]$$

Each exchange spectrum contains two diagonal peaks, AA and BB, and two crosspeaks, AB and BA; the first letter refers to the frequency in f_1 of the EXSY spectrum, while the second gives the frequency in f_2 (hence AB would be the cross peak at $\{v_A, v_B\}$). The above set of coupled, first-order, linear differential equations can be solved using standard methods subject to the following initial conditions at the start of the EXSY mixing time:

$$\text{AA and AB: } [A(t=0)] = A_0, [B(t=0)] = 0$$

$$\text{BB and BA: } [A(t=0)] = 0, [B(t=0)] = B_0$$

giving the following expressions for the diagonal and cross peak intensities in terms of the EXSY mixing time (t):

$$AA(t) = A_0 e^{-t/T_1} \frac{k_b + e^{-(k_b+k_f)t} k_f}{k_b + k_f} \stackrel{\text{short time}}{\approx} A_0 \left(1 - \frac{t}{T_1} - k_f t\right)$$

$$AB(t) = A_0 e^{-t/T_1} \frac{\left(1 - e^{-(k_b+k_f)t}\right) k_f}{k_b + k_f} \stackrel{\text{short time}}{\approx} A_0 k_f t$$

$$BB(t) = B_0 e^{-t/T_1} \frac{e^{-(k_b+k_f)t} k_b + k_f}{k_b + k_f} \stackrel{\text{short time}}{\approx} B_0 \left(1 - \frac{t}{T_1} - k_b t\right)$$

$$BA(t) = B_0 e^{-t/T_1} \frac{\left(1 - e^{-(k_b+k_f)t}\right) k_b}{k_b + k_f} \stackrel{\text{short time}}{\approx} B_0 k_b t$$

where it has been assumed that $T_{1A} = T_{1B}$. The initial rate (short time) solutions are shown to the right. By extracting one-dimensional slices showing the greatest peak intensity from 2D spectra recorded with different mixing times, it is possible to fit the above set of equations and determine both k_b and k_f .

5. 2D Exchange NMR Spectra in Water and in Lipid Micelles

Guest 3

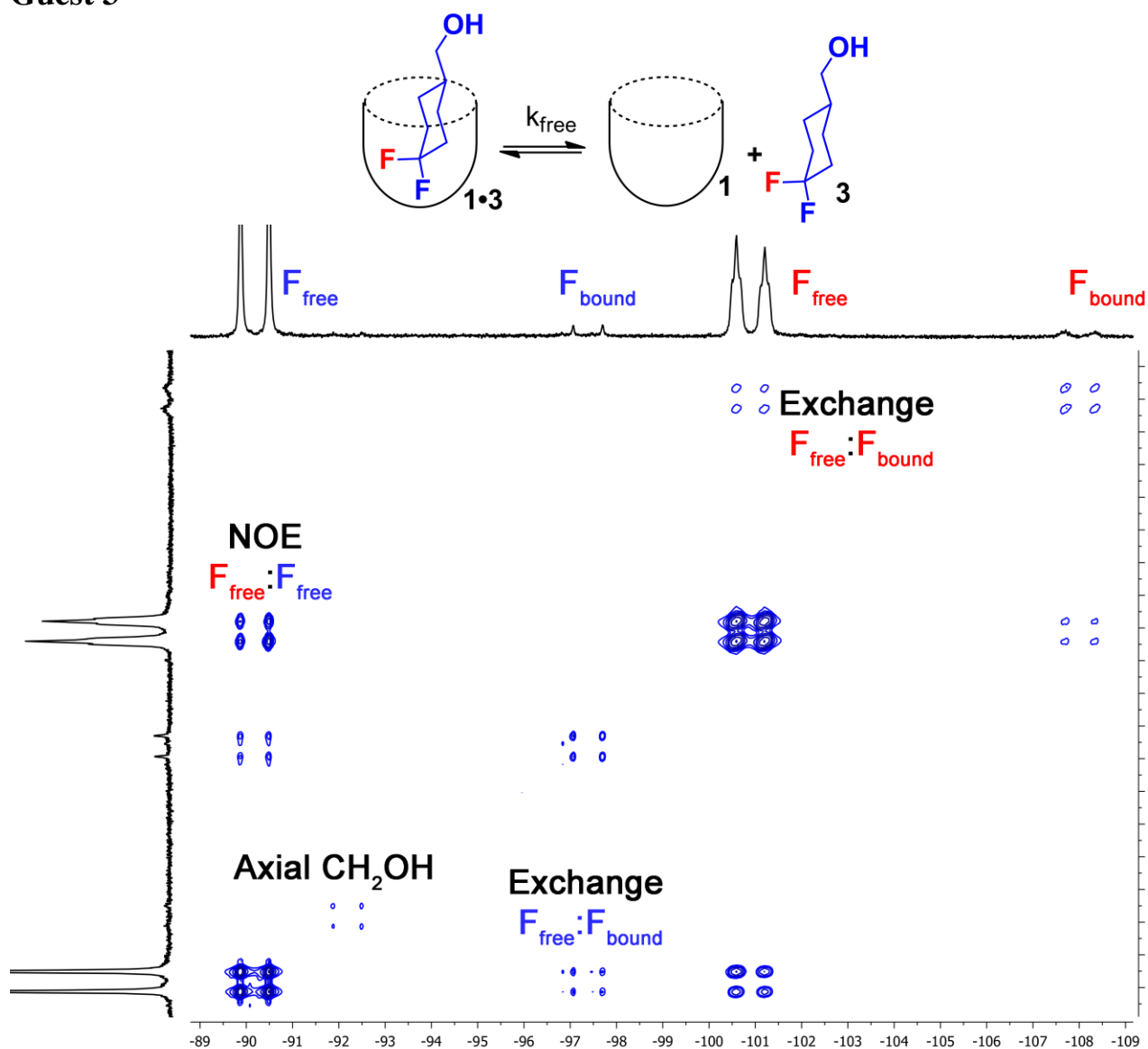


Figure S-38. Full ¹⁹F EXSY spectrum of the cavitant **1**•guest **3** complex in pure D₂O with peak assignments (D₂O, 150.84 MHz, 298 K, mixing time = 150 ms, [**1**] = 5.8 mM, [**3**] = 39.5 mM)

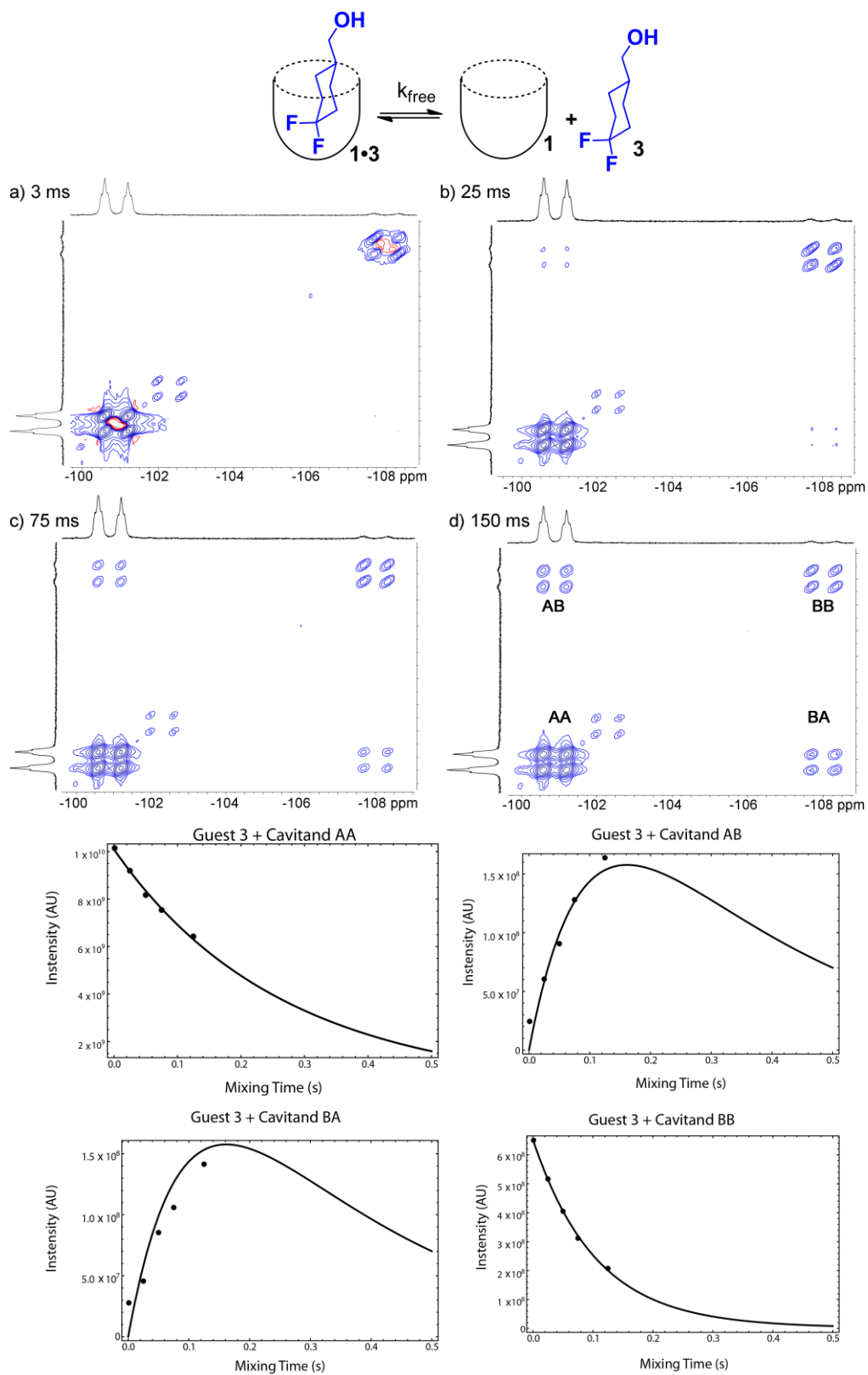


Figure S-39. Representative ^{19}F EXSY spectra of the cavitaand **1**•guest **3** complex in pure D_2O with varying mixing times (D_2O , 376.50 MHz, 298 K, [**1**] = 5.8 mM, [**3**] = 39.5 mM); fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

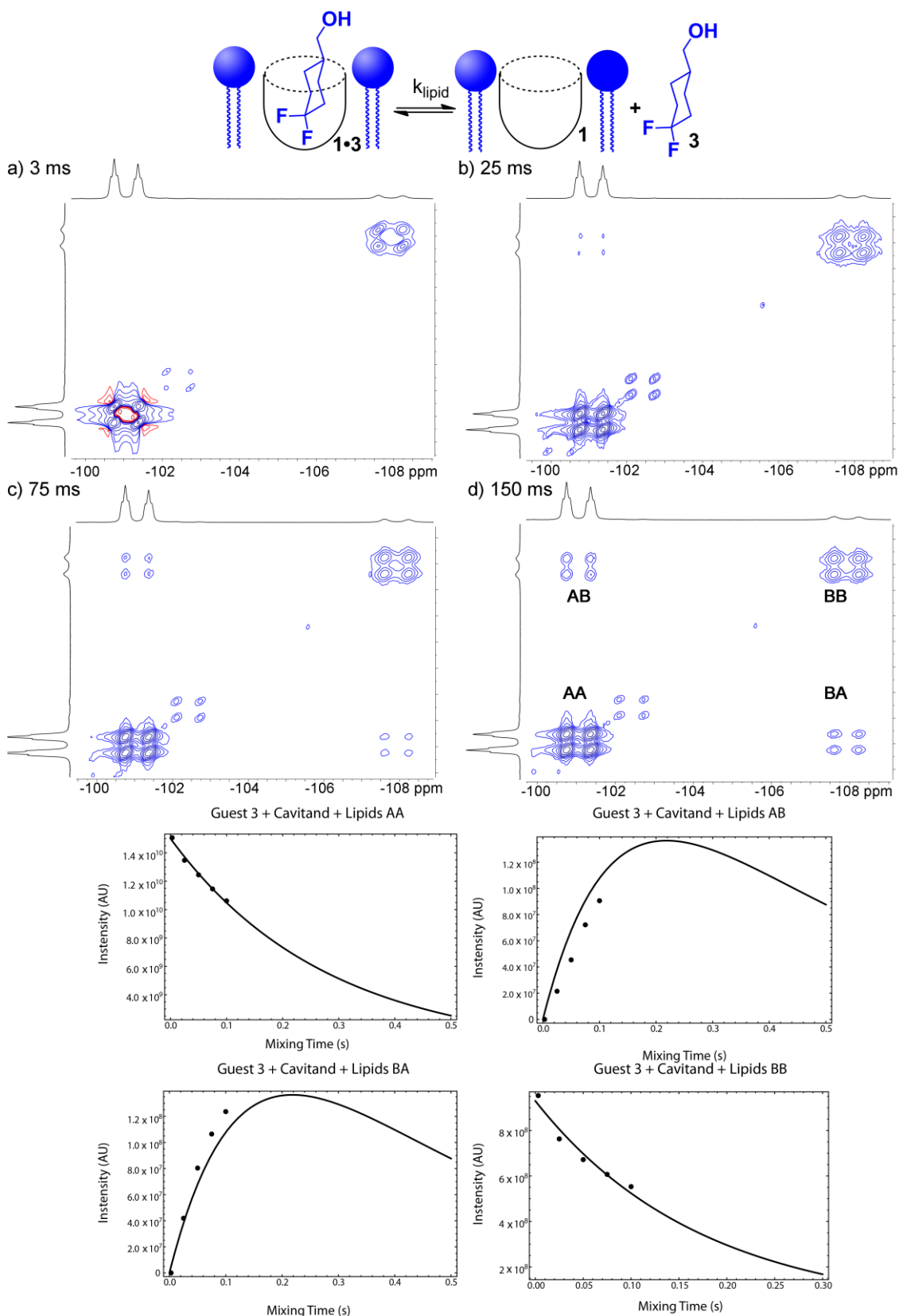


Figure S-40. Representative ^{19}F EXSY spectra of the cavitaand **1**•guest **3** complex in the PC_m micelles with varying mixing times (1 mM HEPES/ D_2O , 376.50 MHz, 298 K, $[\mathbf{1}] = 5.8$ mM, $[\mathbf{3}] = 39.5$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration); fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

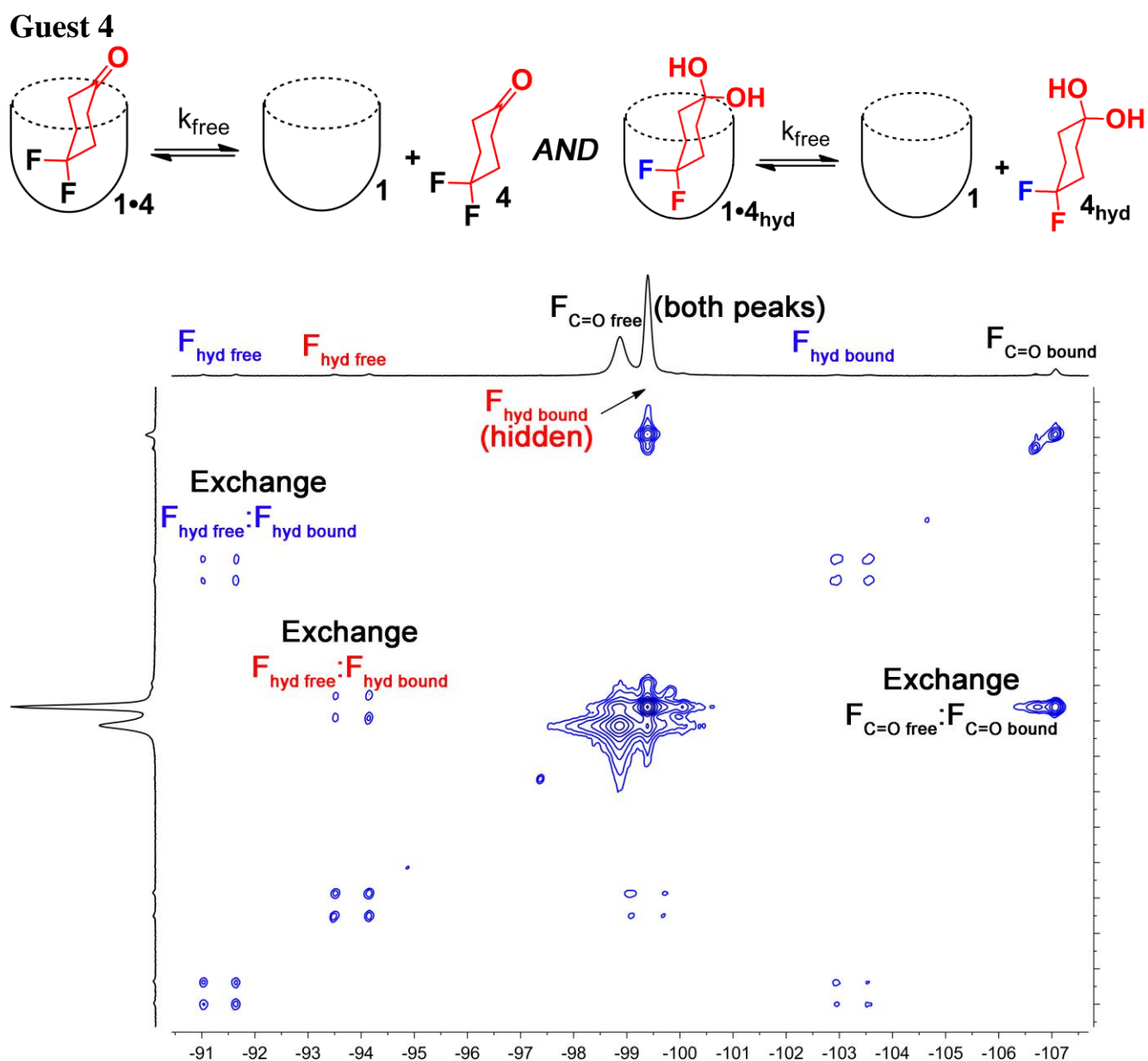


Figure S-41. Full ^{19}F EXSY spectrum of the cavitand **1**•guest **4** complex in pure D_2O with peak assignments (D_2O , 150.84 MHz, 298 K, mixing time = 150 ms, [**1**] = 5.8 mM, [**4**] = 39.5 mM).

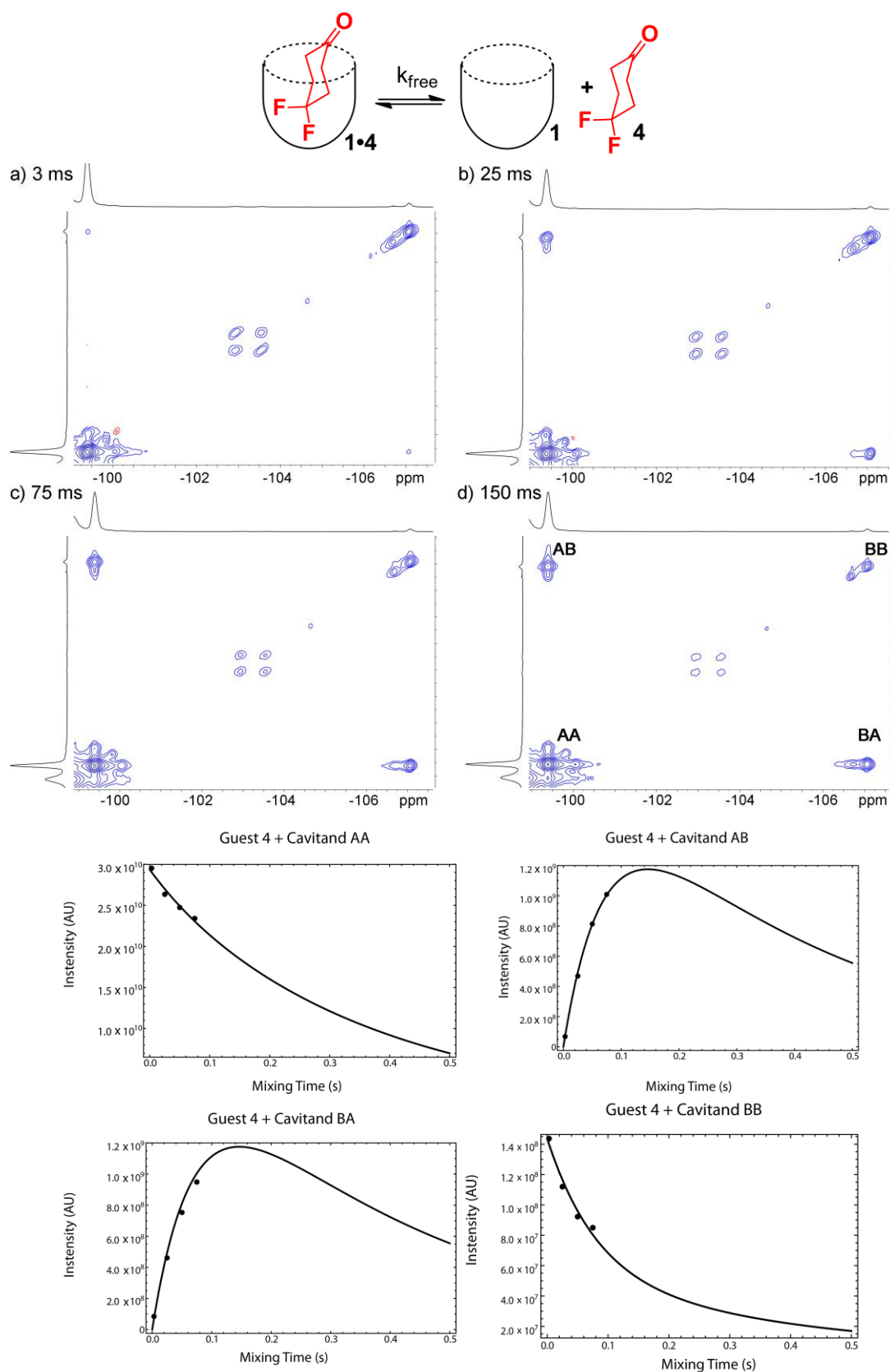


Figure S-42. Representative ^{19}F EXSY spectra of the cavitant **1**•guest **4** complex in pure D_2O with varying mixing times (D_2O , 376.50 MHz, 298 K, $[\mathbf{1}] = 5.8 \text{ mM}$, $[\mathbf{4}] = 39.5 \text{ mM}$); fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

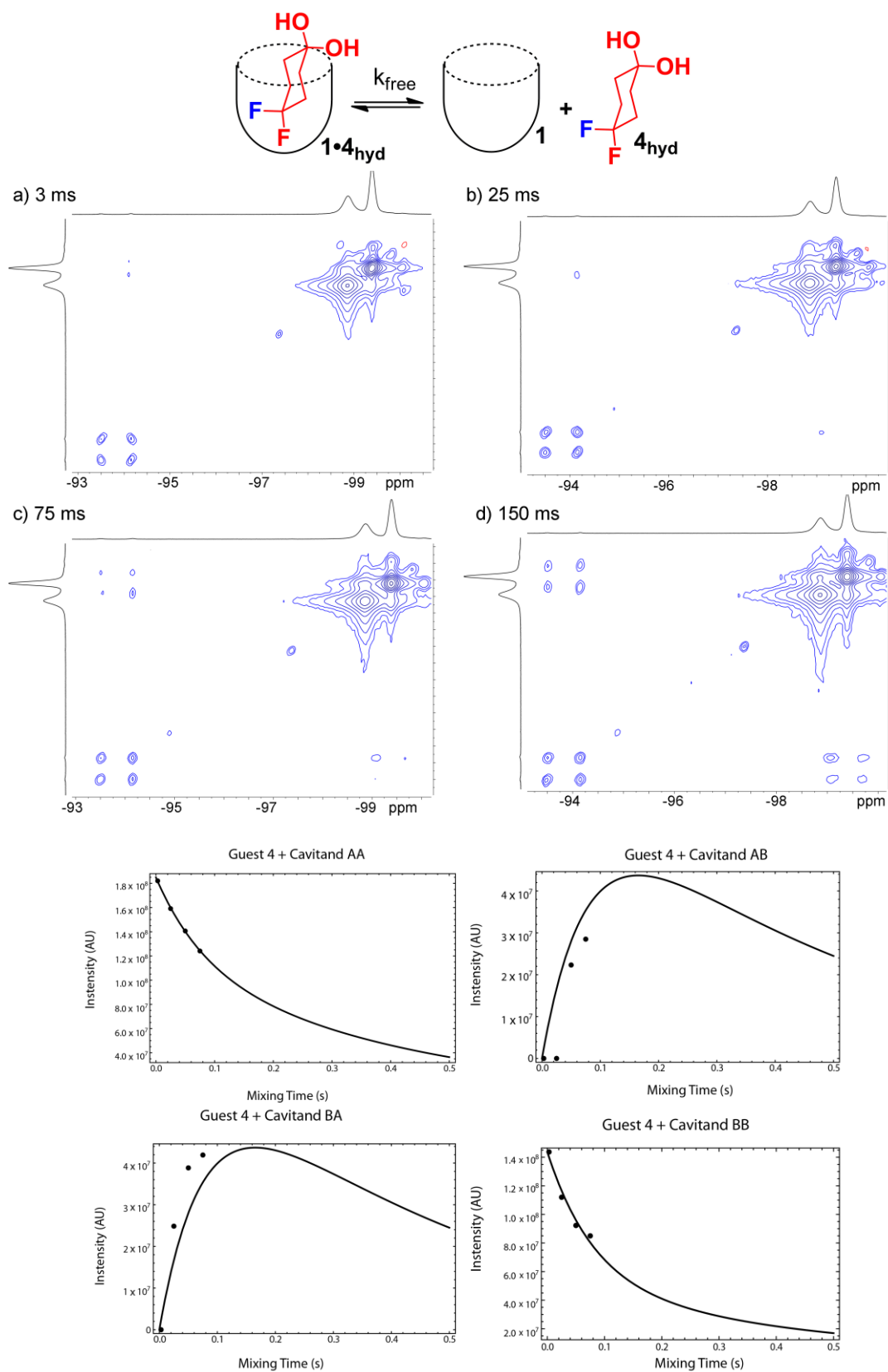


Figure S-43. Representative ^{19}F EXSY spectra of the cavitant **1**•guest **4** complex in pure D_2O with varying mixing times (D_2O , 376.50 MHz, 298 K, $[\mathbf{1}] = 5.8 \text{ mM}$, $[\mathbf{4}] = 39.5 \text{ mM}$) illustrating the exchange peaks of the hydrate; fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

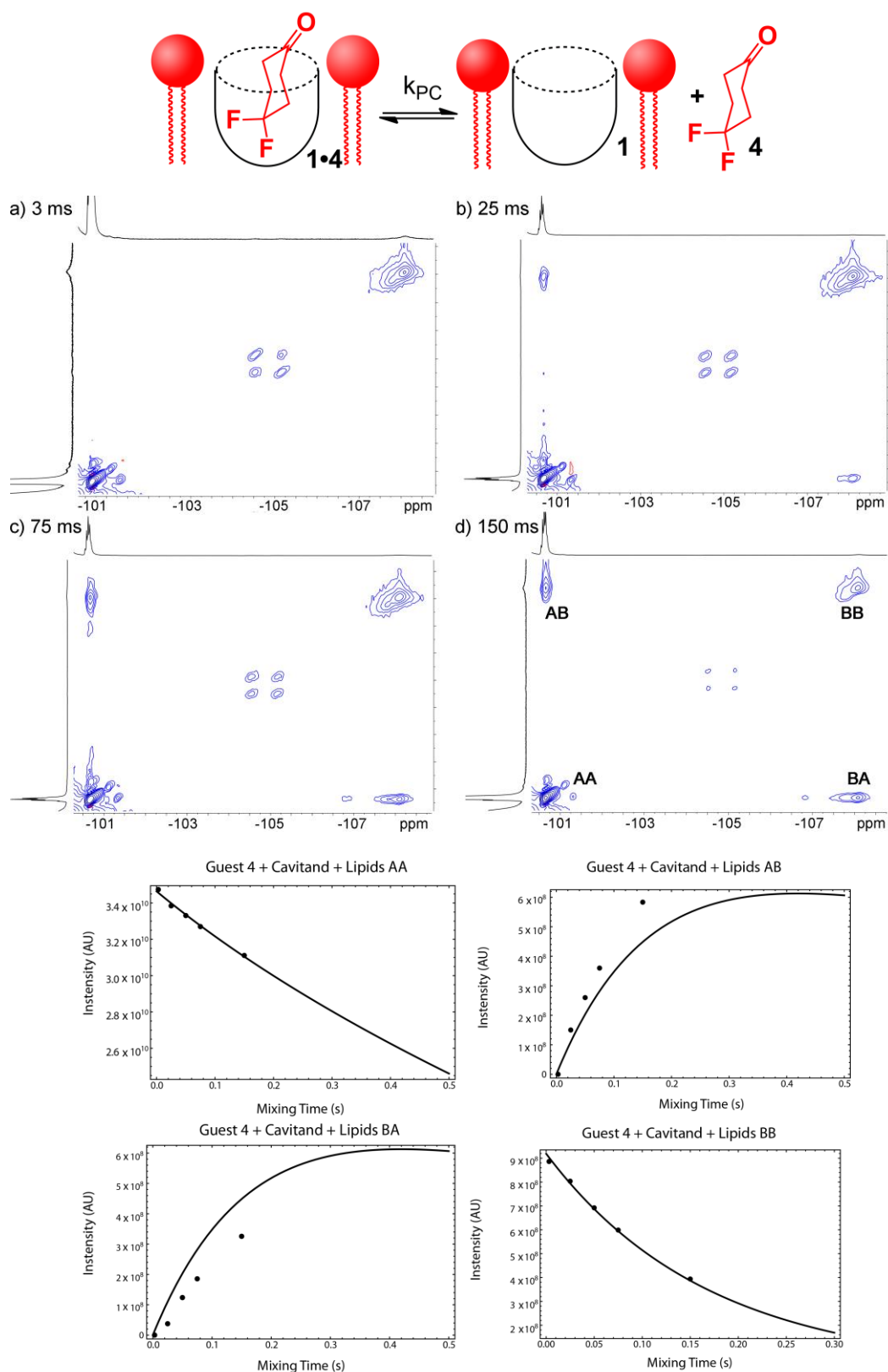


Figure S-44. Representative ^{19}F EXSY spectra of the cavitant **1**•guest **4** complex in the PC_m micelles with varying mixing times (1 mM HEPES/ D_2O , 376.50 MHz, 298 K, $[\mathbf{1}] = 5.8$ mM, $[\mathbf{4}] = 39.5$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration); fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

Guest 6

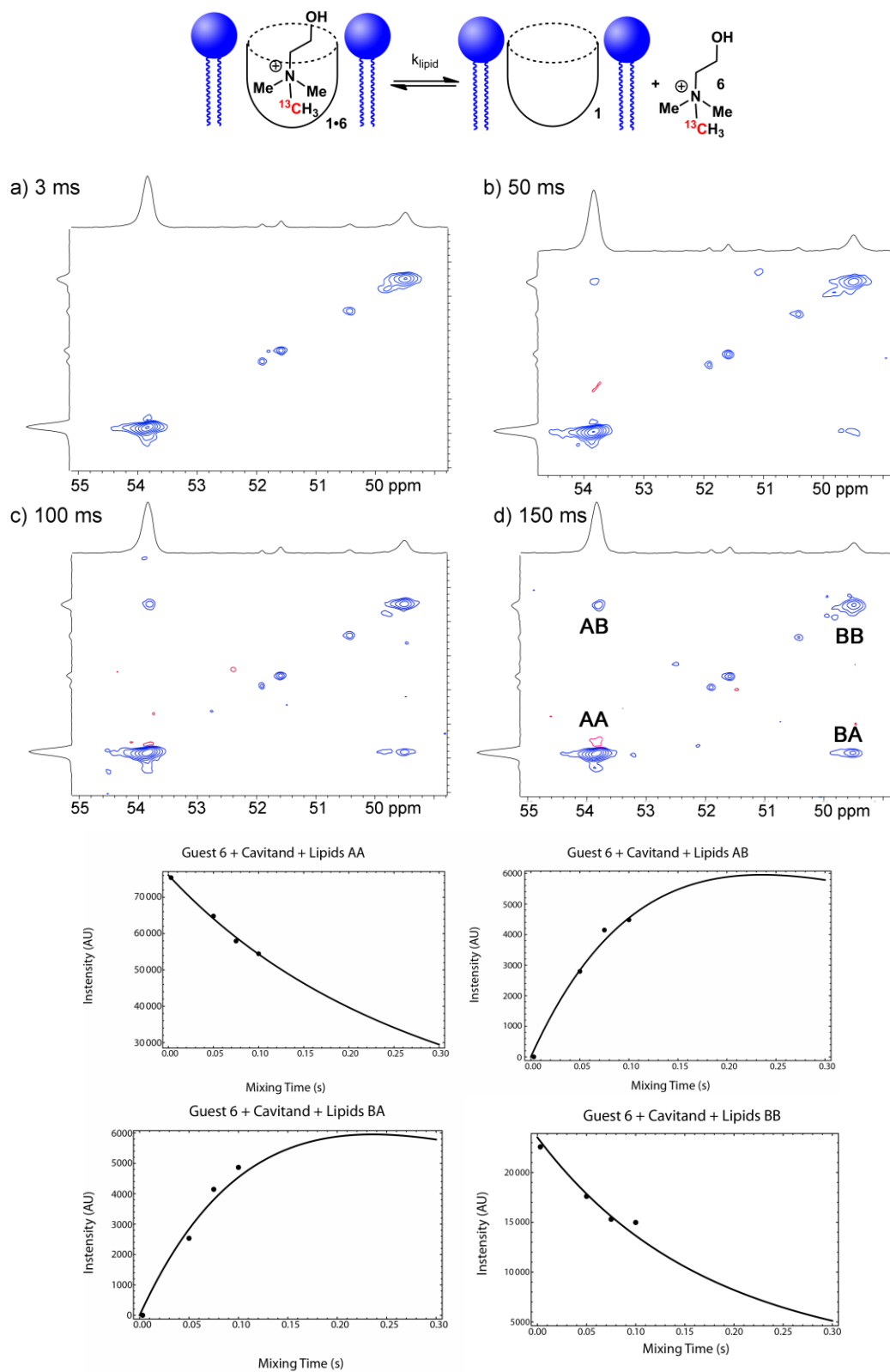


Figure S-45. Representative ^{13}C EXSY spectra of the cavitant **1**•guest **6** complex in the PC_m micelles with varying mixing times (1 mM HEPES/ D_2O , 150.84 MHz, 298 K, $[\mathbf{1}] = 5.8$ mM, $[\mathbf{6}] = 16$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration); fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

Guest 7

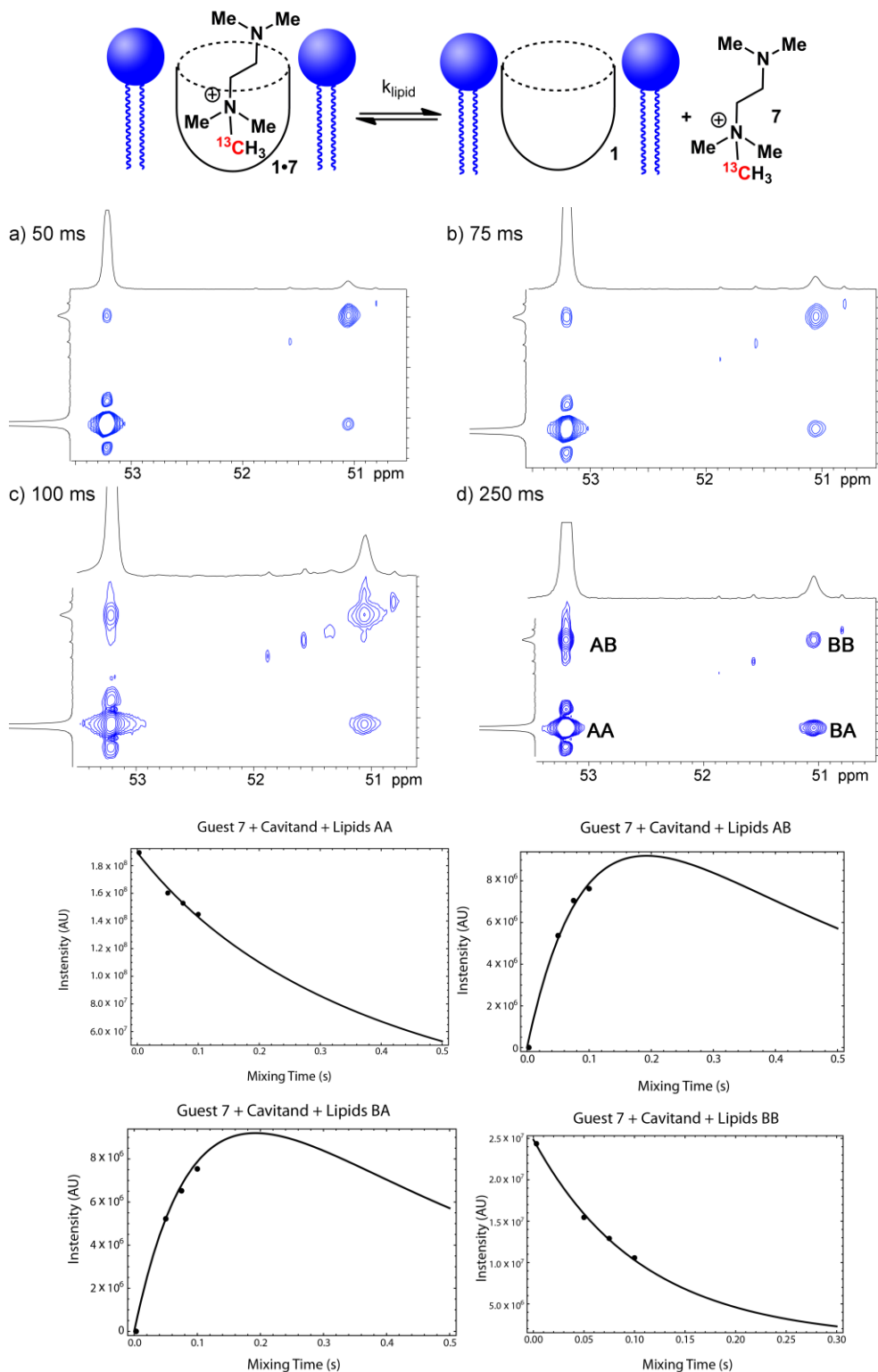


Figure S-46. Representative ^{13}C EXSY spectra of the cavitant **1**•guest **7** complex in the PC_m micelles with varying mixing times (1 mM HEPES/ D_2O , 150.84 MHz, 298 K, $[\mathbf{1}] = 5.8$ mM, $[\mathbf{7}] = 16$ mM, ratio DMPC/DHPC = 3.2:1, 60 mg/mL total lipid concentration); fitted plots of peak intensity correlated with mixing time for each diagonal and crosspeak used to calculate the exchange rate.

6. NMR Spectra in Magnetically Ordered DMPC/DHPC Bicelles

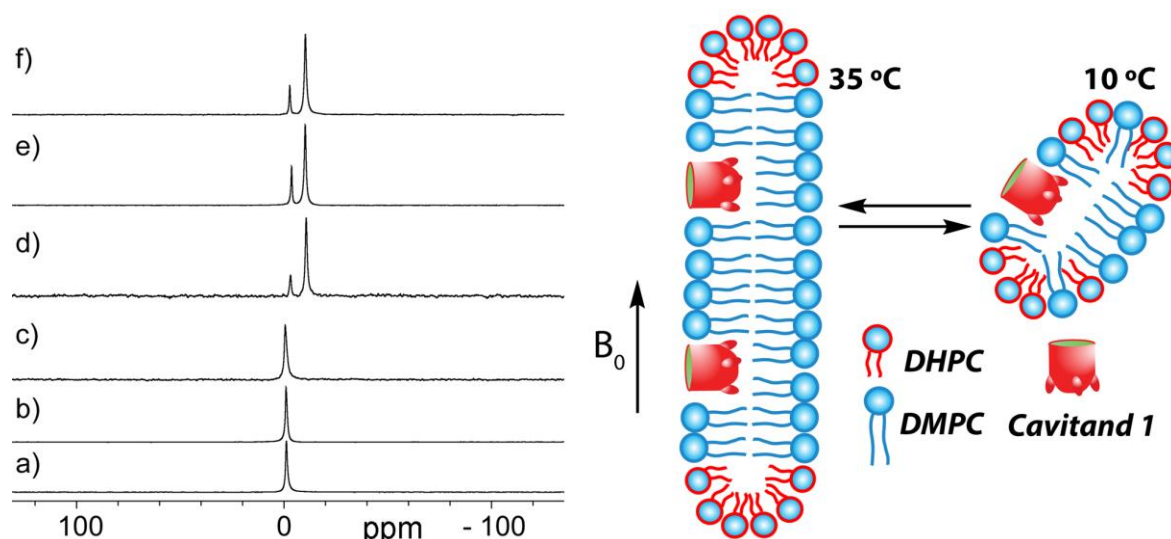


Figure S-47. Temperature dependence of the lipid aggregate. ^{31}P NMR spectra of the DMPC/DHPC aggregates a) alone, 283 K; b) + 5 mM **1**, 283 K; c) + 5 mM **1** + 7 mM guest **5**, 283 K; d) alone, 303 K; e) + 5 mM **1**, 303 K; f) + 5 mM **1** + 7 mM guest **5**, 303 K. Ratio DMPC/DHPC = 3.2:1, 150 mg/mL total lipid concentration, 162.07 MHz, 2.5 mM HEPES/D₂O.

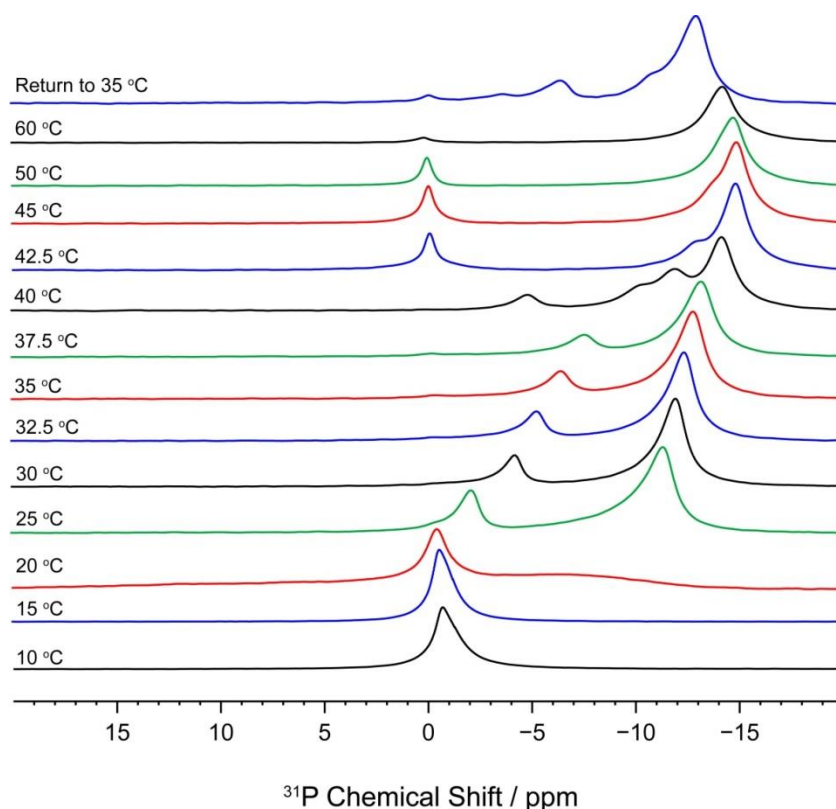


Figure S-48. Temperature-dependent ^{31}P spectra of DMPC/DHPC bicelles. The two peaks indicative of bicelle formation display maximum splitting at 35 °C, the temperature at which all future experiments were performed, unless otherwise indicated. Low temperatures display a single peak at 0 ppm, indicative of micelle formation. Ratio DMPC/DHPC = 3.2:1, 150 mg/mL total lipid concentration, 162.07 MHz, 2.5 mM HEPES/D₂O.

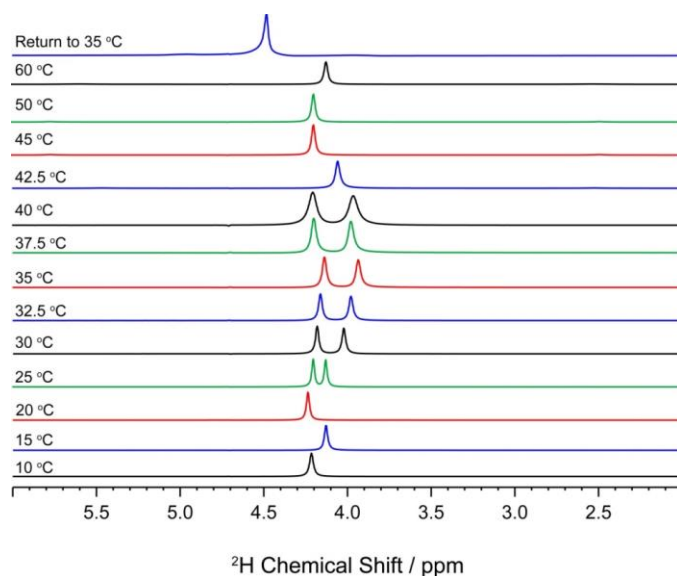


Figure S-49. Temperature-dependent ^2H spectra of DMPC/DHPC bicelles. The two peaks indicative of bicelle formation display maximum splitting at 35 °C, the temperature at which all future experiments were performed, unless otherwise indicated. Low temperatures display a single resonance, indicative of micelle formation. Ratio DMPC/DHPC = 3.2:1, 150 mg/mL total lipid concentration, 92.09 MHz, 2.5 mM HEPES/D₂O.

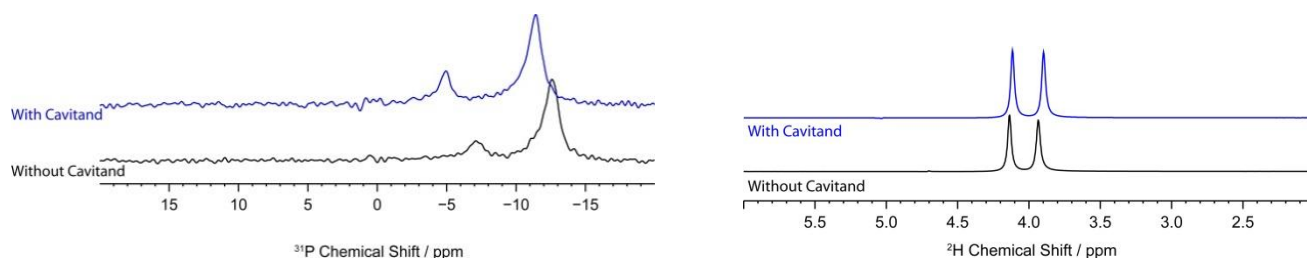


Figure S-50. ^{31}P and ^2H spectra of bicelles with and without cavitand added. Ratio DMPC/DHPC = 3.2:1, 150 mg/mL total lipid concentration, 162.07/ 92.09 MHz, 2.5 mM HEPES/D₂O.

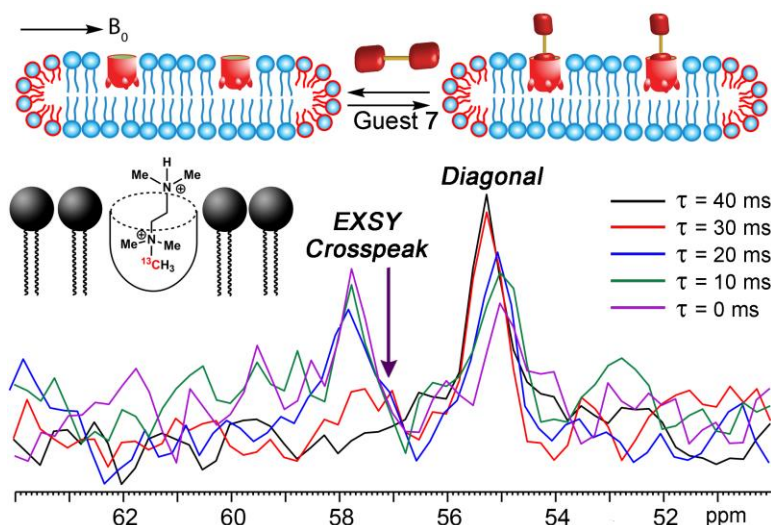


Figure S-51. In/out exchange of guest **7** in the magnetically ordered bicelle system PC_b. Extracted 1D slices of the 2D ^{13}C - ^{13}C EXSY NMR spectra at mixing time $\tau = 0$ ms - $\tau = 40$ ms of **1**•**7**•PC_b; (2.5 mM HEPES/D₂O, 100.69 MHz, 298 K, [1] = 20 mM, [7] = 36 mM, ratio DMPC/DHPC = 3.2:1, 150 mg/mL total lipid concentration). Change in diagonal and crosspeak intensity shown. Slices extracted from F2, $\delta = 55$ ppm.