Electronic Supplementary Information (ESI)

Microviscosity, Encapsulation, and Permeability of 2-Ketooctanoic Acid

Vesicle membranes

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First-order kinetics for OH⁻ permeation across vesicular membranes

The acidic dissociation equilibrium of the fluorescence probe riboflavin can be represented as:

$$\mathbf{R}\mathbf{H} \rightleftharpoons \mathbf{R}^- + \mathbf{H}^+ \qquad K_a \tag{S1}$$

where RH and R⁻ represent the neutral and deprotonated riboflavin, respectively, and K_a is the acidic dissociation constant of RH. One has:

$$K_{a} = \frac{[\mathbf{R}^{-}][\mathbf{H}^{+}]}{[\mathbf{R}\mathbf{H}]}$$
(S2a)

and

$$[OH^{-}] = \frac{K_{w}[R^{-}]}{K_{a}[RH]}$$
(S2b)

where [*i*] is the molar concentration of the *i* specie, and K_w is the dissociation constant of H₂O.

In our case, the total riboflavin molecules exist in two states: one is adsorbed on the external vesicle membrane surfaces and another is enclosed within the internal vesicle

aqueous core. Thus, before adjusting the pH of the KOCOOH solution, one has,

$$[\mathbf{RH}]_{\text{total}} = [\mathbf{RH}]_{\text{ex}} + [\mathbf{RH}]_{\text{in}}$$
(S3a)

$$[R^{-}]_{total} = [R^{-}]_{ex} + [R^{-}]_{in}$$
(S3b)

where $[RH]_{total}$ and $[R^-]_{total}$ are the total molar concentrations of RH and R⁻ in the system, respectively, $[RH]_{ex}$ and $[R^-]_{ex}$ are the molar concentrations of RH and R⁻ adsorbed on the external vesicle surfaces, respectively, and $[RH]_{in}$ and $[R^-]_{in}$ are the molar concentrations of RH and R⁻ enclosed within the internal vesicle aqueous core, respectively.

From Eq. (S2b), we have,

$$[OH^{-}]_{in}^{0} = \frac{K_{w}[R^{-}]_{in}}{K_{a}[RH]_{in}}$$
(S4a)

$$[OH^{-}]_{in}^{t} = \frac{K_{w}([R^{-}]_{in} + [R^{-}]_{in}^{t})}{K_{a}([RH]_{in} - [R^{-}]_{in}^{t})}$$
(S4b)

where $[OH^-]_{in}^0$ and $[OH^-]_{in}^t$ are the OH⁻ concentrations within the internal vesicles at t = 0and time *t*, respectively, and $[R^-]_{in}^t$ is the molar concentrations of R^- generated after the pH adjusting in the internal vesicles at time *t*. Eq. (S4b) minus Eq. (S4a) gives,

$$[OH^{-}]_{in}^{t} - [OH^{-}]_{in}^{0} = \frac{K_{w}[R^{-}]_{in}^{t}}{K_{a}([RH]_{in} - [R^{-}]_{in}^{t})}(1 - \frac{[R^{-}]_{in}}{[RH]_{in}})$$
(S5a)

In our case, the pH within the internal vesicles at t = 0 is 6.8, and the value of $[R^-]_{in}/[RH]_{in}$ (~10^{-3.4}) is much little than 1. Therefore, we have,

$$[OH^{-}]_{in}^{t} \approx [OH^{-}]_{in}^{0} + \frac{K_{w}[R^{-}]_{in}^{t}}{K_{a}([RH]_{in} - [R^{-}]_{in}^{t})}$$
(S5b)

The fluorescence emission intensity (I_{em}) of riboflavin in the KOCOOH solution arises from the contribution of the two states of the dye molecules. After adjusting the pH of KOCOOH solution from 6.8 to 10.2, the "instantaneous" loss in the I_{em} of the RH-containing KOCOOH solution can be attributed to the deprotonation of riboflavin adsorbed on the external vesicle surfaces, and the gradual reduction in the residual I_{em} with time (t) can be attributed to the deprotonation of riboflavin enclosed within the internal vesicle aqueous core. Assuming that the I_{em} of riboflavin is proportional to [RH], i.e.,

$$I_{\rm em} = \varepsilon[\rm RH] \tag{S6}$$

in which ε is a coefficient, we have,

$$I_0^0 = \varepsilon([\mathbf{RH}]_{ex} + [\mathbf{RH}]_{in})$$
(S7a)

$$I_0 = \varepsilon([\mathbf{RH}]_{ex}^0 + [\mathbf{RH}]_{in}^0)$$
(S7b)

and

$$I_0^0 - I_0 = \varepsilon [R^-]_{ex}^0$$
 (S8a)

$$I_0 - I_\infty = \varepsilon [\mathbf{R}^-]_{\rm in}^\infty \tag{S8b}$$

where I_0^0 and I_0 are the I_{em} of the RH-containing solution before and just after (t = 0)adjusting its pH, respectively, I_∞ is the I_{em} of the solution at $t \rightarrow \infty$ (the permeation equilibrium state), $[RH]_{ex}^0$ and $[RH]_{in}^0$ are the molar concentrations of RH adsorbed on the external vesicle surfaces and enclosed within the internal vesicle aqueous core, respectively, $[R^-]_{ex}^0$ and $[R^-]_{in}^\infty$ are the molar concentrations of R⁻ generated after the pH adjusting in the external vesicles at t = 0 and in the internal vesicles at $t \rightarrow \infty$, respectively. Note that $[RH]_{in}^0 =$ $[RH]_{in}^0$ owing to no OH⁻ permeation occurring at t = 0, and that $[RH]_{ex}^0 = [RH]_{ex}^\infty$ and $[R^-]_{ex}^0 = [R^-]_{ex}^\infty$ owing to the deprotonation of RH adsorbed on the external vesicles is instantaneously completed after the pH adjusting.

Based on Eqs. (S8a) and (S8b), we have,

$$\frac{I_0^0 - I_0}{I_0 - I_\infty} = \frac{[R^-]_{ex}^0}{[R^-]_{in}^\infty} \equiv \frac{[R^-]_{ex}^\infty}{[R^-]_{in}^\infty}$$
(S9)

Assuming that when the OH⁻ permeation equilibrium is achieved (i.e., at $t \rightarrow \infty$), the OH⁻ concentration of the internal vesicles, $[OH^-]_{in}^{\infty}$, is equal to that of the external vesicles (bulk

solution), $[OH^-]_{ex}^0$ (where $[OH^-]_{ex}^0 \equiv [OH^-]_{ex}^\infty$). From Eq. (S2b), we have,

$$[OH^{-}]_{ex}^{\infty} = \frac{K_{w}([R^{-}]_{ex} + [R^{-}]_{ex}^{\infty})}{K_{a}([RH]_{ex} - [R^{-}]_{ex}^{\infty})}$$
(S10a)

$$[OH^{-}]_{in}^{\infty} = \frac{K_{w}([R^{-}]_{in} + [R^{-}]_{in}^{\infty})}{K_{a}([RH]_{in} - [R^{-}]_{in}^{\infty})}$$
(S10b)

Because in our case, $[R^-]_{ex}^{\infty} \gg [R^-]_{ex}$ and $[R^-]_{in}^{\infty} \gg [R^-]_{in}$, from Eqs. (S10a) and (10b), we can obtain,

$$\frac{[R^{-}]_{ex}^{\infty}}{[R^{-}]_{in}^{\infty}} \approx \frac{[RH]_{ex}}{[RH]_{in}}$$
(S11)

Based on Eqs. (S7a), (S9), and (S11), we can obtain,

$$[RH]_{in} = \frac{I_0^0 (I_0 - I_\infty)}{\varepsilon (I_0^0 - I_\infty)}$$
(S12)

At time *t*, we have,

$$\left[\mathbf{R}^{-}\right]_{\mathrm{in}}^{t} = \frac{I_{0} - I_{t}}{\varepsilon}$$
(S13a)

$$[RH]_{in}^{t} = [RH]_{in} - [R^{-}]_{in}^{t}$$
(S13b)

Based on Eqs. (S5b), (S12) and (S13), we have,

$$[OH^{-}]_{in}^{t} = [OH^{-}]_{in}^{0} + \frac{K_{w}}{K_{a}} \left[\frac{(I_{0} - I_{t})(I_{0}^{0} - I_{\infty})}{I_{0}^{0}(I_{0} - I_{\infty}) - (I_{0} - I_{t})(I_{0}^{0} - I_{\infty})} \right]$$
(S14)

The first-order kinetics for the permeation of OH⁻ across vesicular membranes can be derived as:

$$\ln(\frac{[OH^{-}]_{ex}^{0} - [OH^{-}]_{in}^{t}}{[OH^{-}]_{ex}^{0} - [OH^{-}]_{in}^{0}}) = -k_{1}t$$
(S15)

In our case, $[OH^-]^0_{ex} \gg [OH^-]^0_{in}$. From Eqs. (S14) and (S15), we have,

$$\ln(1 - \frac{K_{w}}{K_{a}[OH^{-}]_{ex}^{0}} \left[\frac{(I_{0} - I_{t})(I_{0}^{0} - I_{\infty})}{I_{0}^{0}(I_{0} - I_{\infty}) - (I_{0} - I_{t})(I_{0}^{0} - I_{\infty})} \right] = -k_{1}t$$
(S16a)

$$\ln(1 - \frac{[\mathrm{H}^{+}]_{\mathrm{ex}}^{0}}{K_{\mathrm{a}}} \left[\frac{(I_{0} - I_{t})(I_{0}^{0} - I_{\infty})}{I_{0}^{0}(I_{0} - I_{\infty}) - (I_{0} - I_{t})(I_{0}^{0} - I_{\infty})} \right]) = -k_{1}t$$

$$\ln(1 - \frac{[\mathrm{RH}]_{\mathrm{ex}}^{0}}{[\mathrm{R}^{-}]_{\mathrm{ex}}^{0}} \left[\frac{(I_{0} - I_{t})(I_{0}^{0} - I_{\infty})}{I_{0}^{0}(I_{0} - I_{\infty}) - (I_{0} - I_{t})(I_{0}^{0} - I_{\infty})} \right]) = -k_{1}t$$
(S16b)

From Eqs. (S7b), (S12), and (S13), we have,

$$[RH]_{ex}^{0} = \frac{I_{0}}{\varepsilon} - [RH]_{in}^{0} = \frac{I_{0}}{\varepsilon} - ([RH]_{in} - [R^{-}]_{in}^{0})$$
(S17a)

$$[\mathbf{RH}]_{\mathrm{ex}}^{0} = \frac{I_{0}}{\varepsilon} - \frac{I_{0}^{0}(I_{0} - I_{\infty})}{\varepsilon(I_{0}^{0} - I_{\infty})}$$
(S17b)

Based on Eqs. (S8a), (S16b), and (S17b), we can obtain,

$$\ln\left[\frac{I_0^0(I_t - I_\infty)}{I_0^0(I_0 - I_\infty) - (I_0 - I_t)(I_0^0 - I_\infty)}\right] = -k_1 t$$
(S18)

In the early stage of permeation, $I_0^0(I_0 - I_\infty) \gg (I_0 - I_t)(I_0^0 - I_\infty)$. The relative error for this approximation was estimated to be lower than 2% for our case. Therefore, the first-order kinetics for the permeation of OH⁻ across vesicular membranes can be written as:

$$\ln(\frac{I_t - I_\infty}{I_0 - I_\infty}) = -k_1 t \tag{S19}$$