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Supporting Information

Construction of blood compatible lysine-immobilized chitin/carbon nanotubes microspheres and potential applications for blood purified therapy

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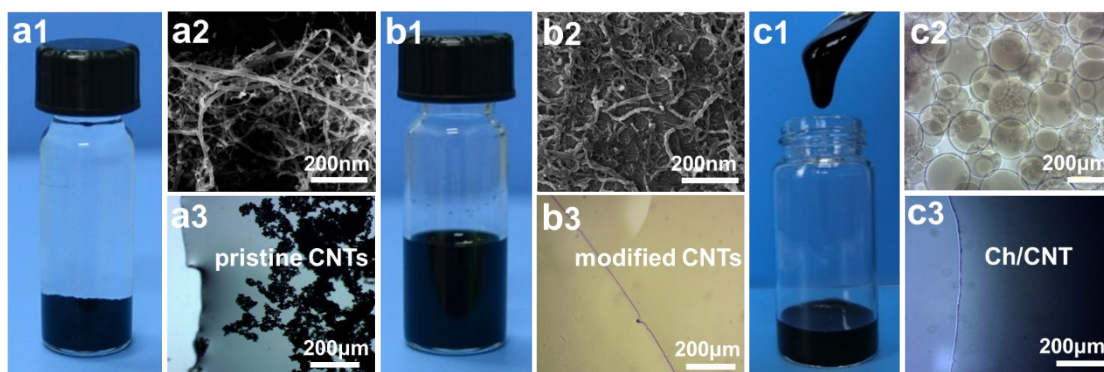
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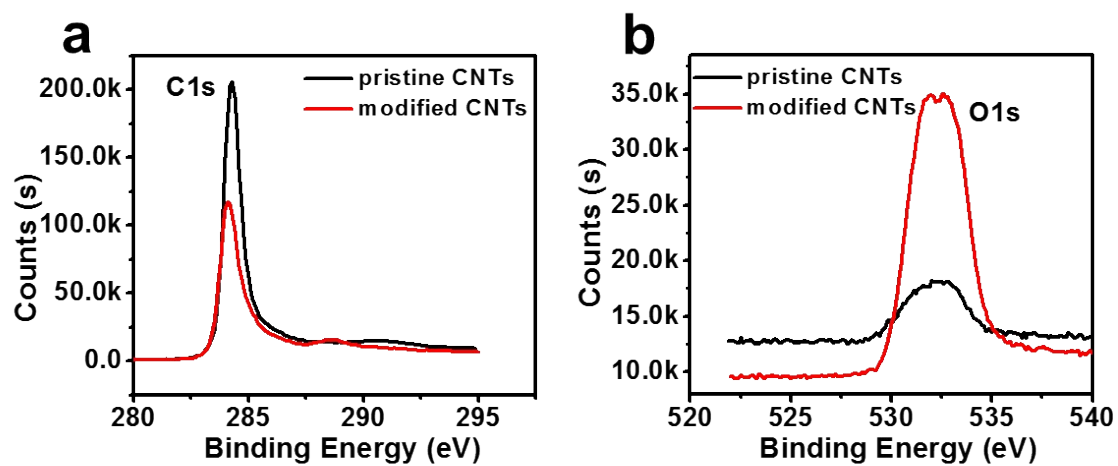
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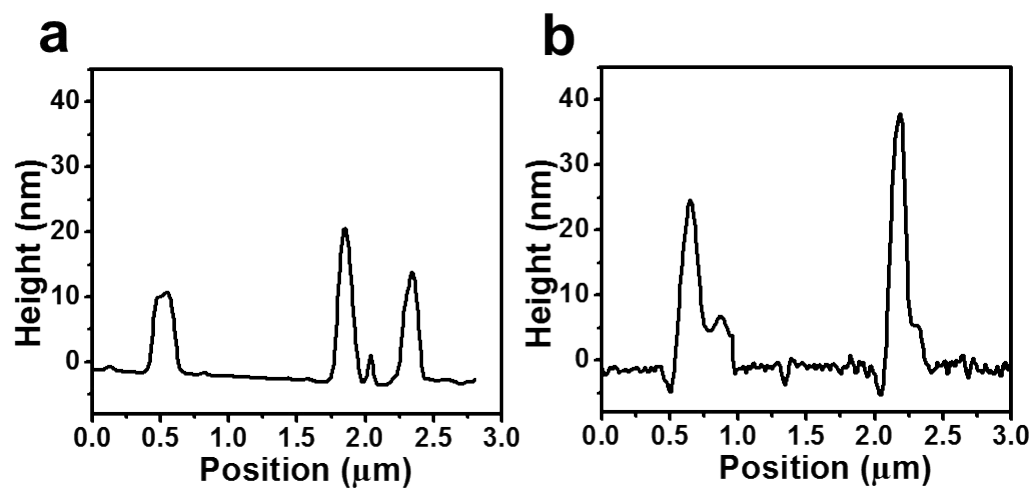
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50 **Fig. S1** (a1) Photograph of the pristine CNTs precipitated in water, (a2) SEM image of pristine
 51 CNTs, (a3) optical microscopy image of the pristine CNTs agglomerated in water, (b1) photograph
 52 of modified CNTs aqueous suspension (0.125mg/ml), (b2) SEM image of modified CNTs, (b3)
 53 optical microscopy image of homogenous modified CNTs aqueous solution, (c1) photograph of
 54 Ch/CNT homogeneous solution, (c2) optical microscopy image of Ch/CNT microspheres and (c3)
 55 optical microscopy image of Ch/CNT homogeneous solution.



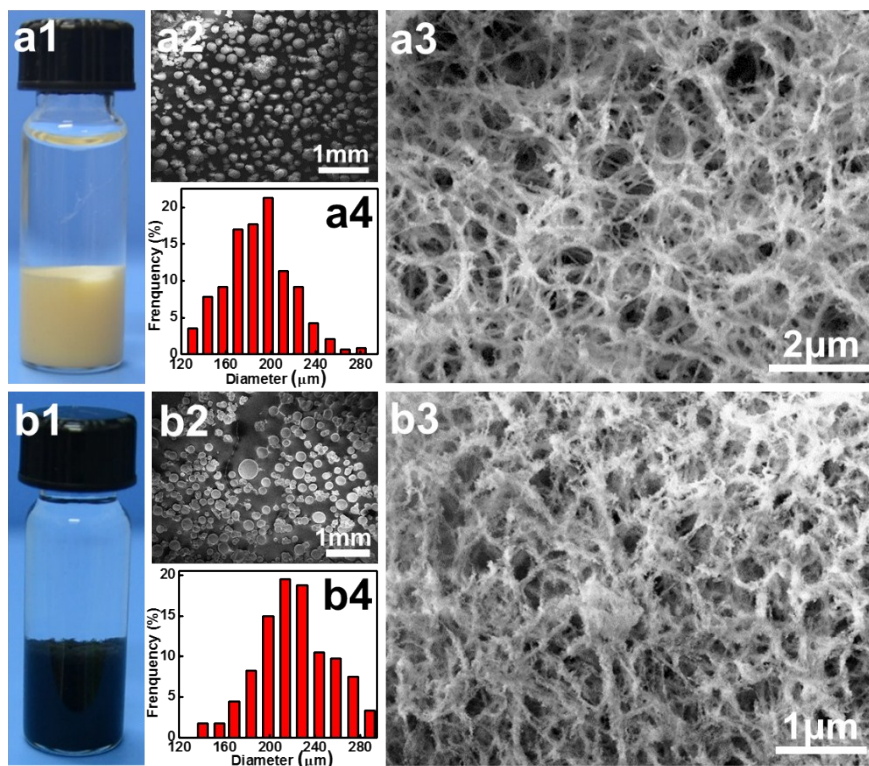
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58 **Fig. S2** XPS spectra of C1s (a) and O1s (b) for pristine CNTs and modified CNTs, respectively.



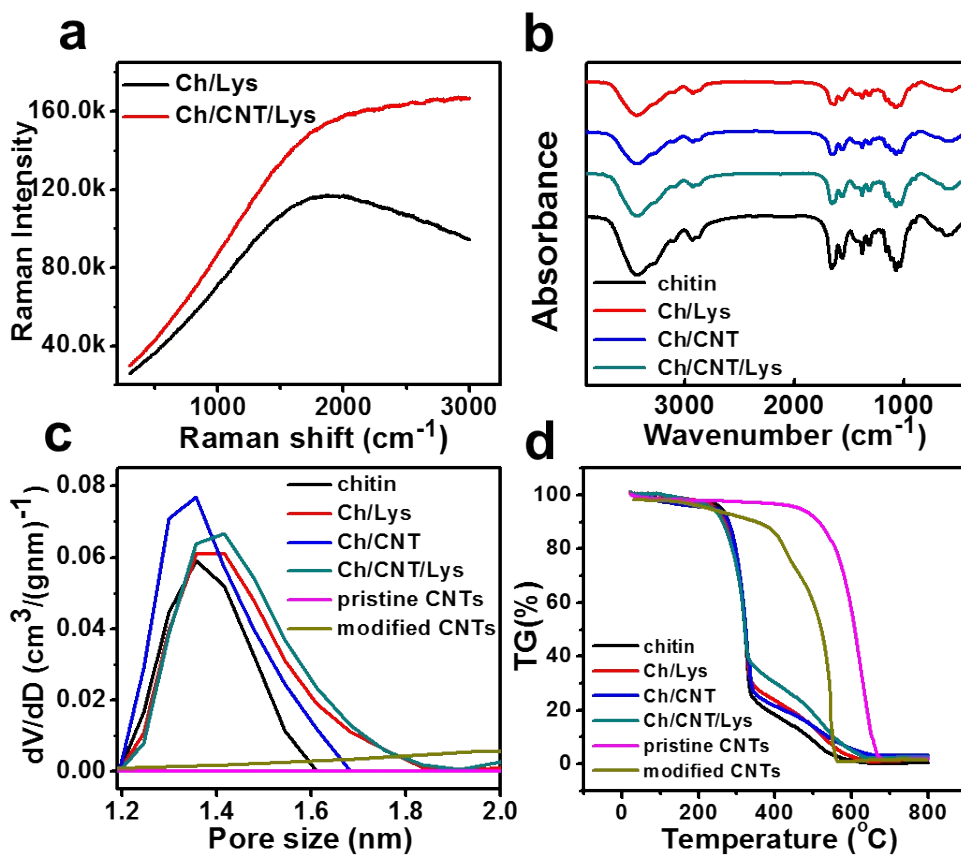
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61 **Fig. S3** Line profiles along the lines marked in the AFM images of modified CNTs (a) and Ch/CNT
 62 (b)



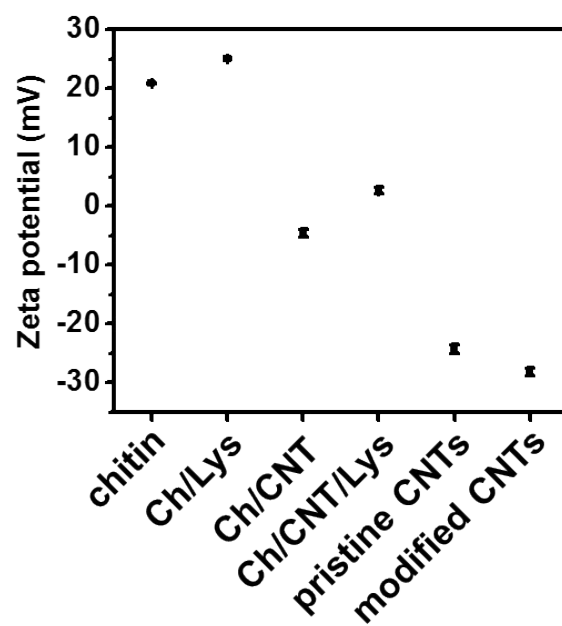
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65 **Fig. S4** (a1) Photograph of Ch/Lys microspheres, (a2, a3) SEM images of Ch/Lys microspheres,
 66 (a4) size distribution of Ch/Lys microspheres, (b1) photograph of Ch/CNT/Lys microspheres, (b2,
 67 b3) SEM images of Ch/CNT/Lys microspheres and (b4) size distribution of Ch/CNT/Lys
 68 microspheres.



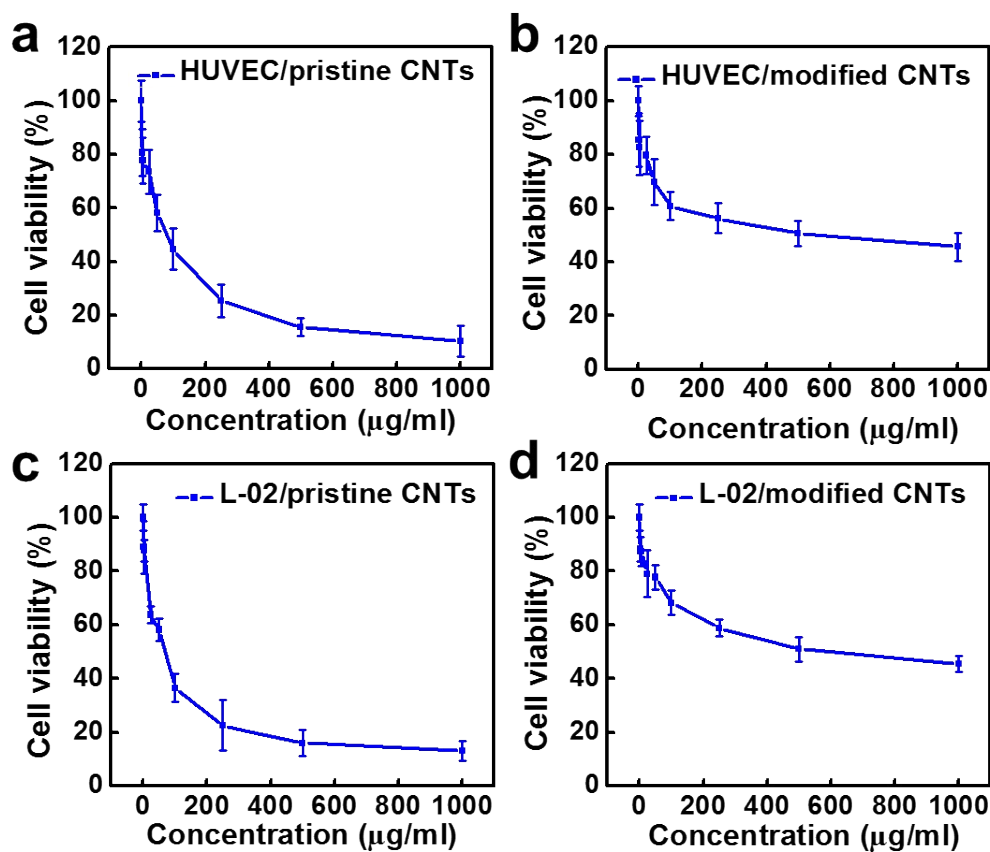
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71 **Fig. S5** (a) Raman spectroscopy of Ch/Lys and Ch/CNT/Lys microspheres, (b) FT-IR spectra, (c)
 72 pore size distribution determined by the DFT method and (d) TG curves of chitin-based
 73 microspheres and CNTs.



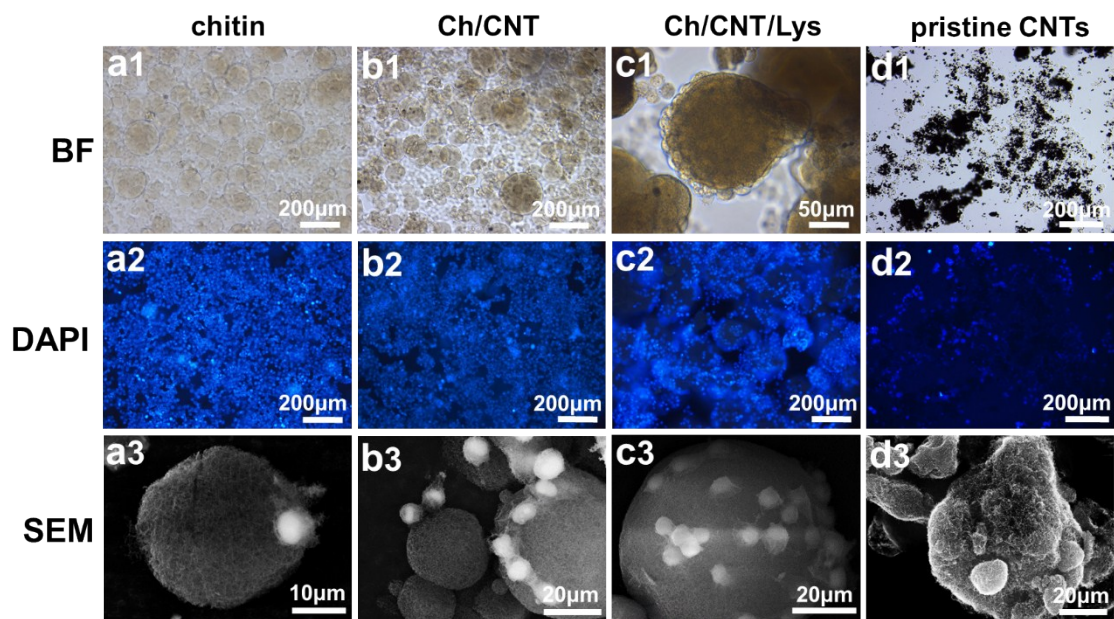
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76 **Fig. S6** Zeta-potential of chitin-based microspheres and CNTs.



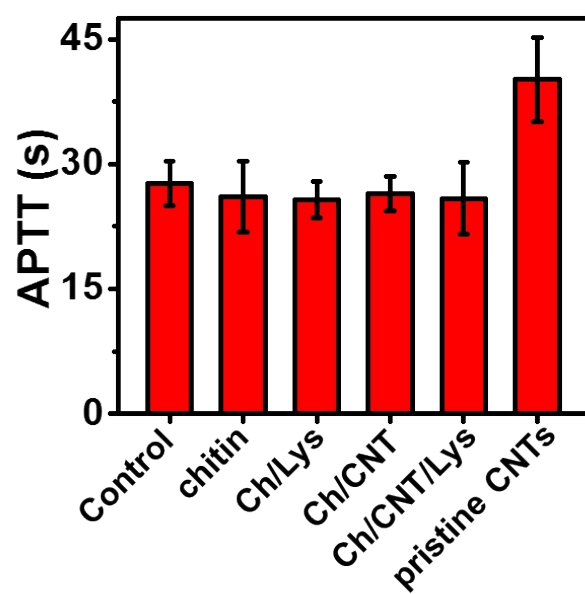
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79 **Fig. S7** (a) In vitro cytotoxicity tests of different concentration of pristine CNTs incubated with
80 HUVEC cells, (b) different concentration of modified CNTs incubated with HUVEC cells, (c)
81 different concentration of pristine CNTs incubated with L-02 cells and (d) different concentration
82 of modified CNTs incubated with L-02 cells. Data are expressed as mean $\pm$ SD (n=4).



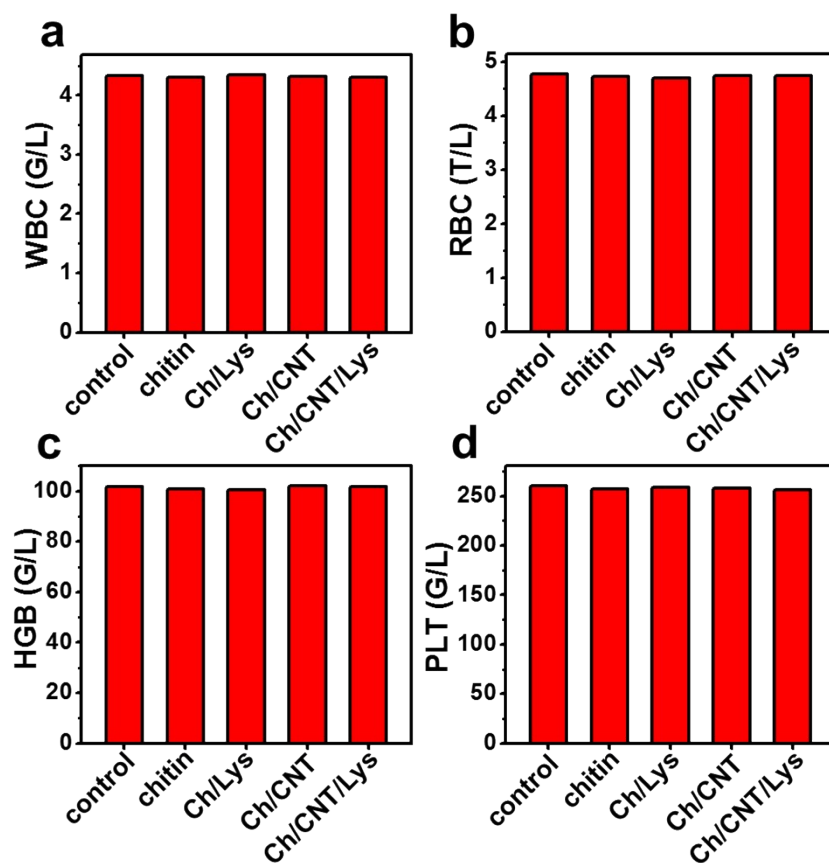
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85 **Fig. S8** Optical microscopy, fluorescence microscopy and SEM images of L-02 cells cultured on
 86 chitin microspheres (a), Ch/CNT microspheres (b), Ch/CNT/Lys microspheres (c) and pristine
 87 CNTs (d). The nuclei of the HUVEC cells were stained with 4', 6-diamidino-2-phenylindole
 88 (DAPI).



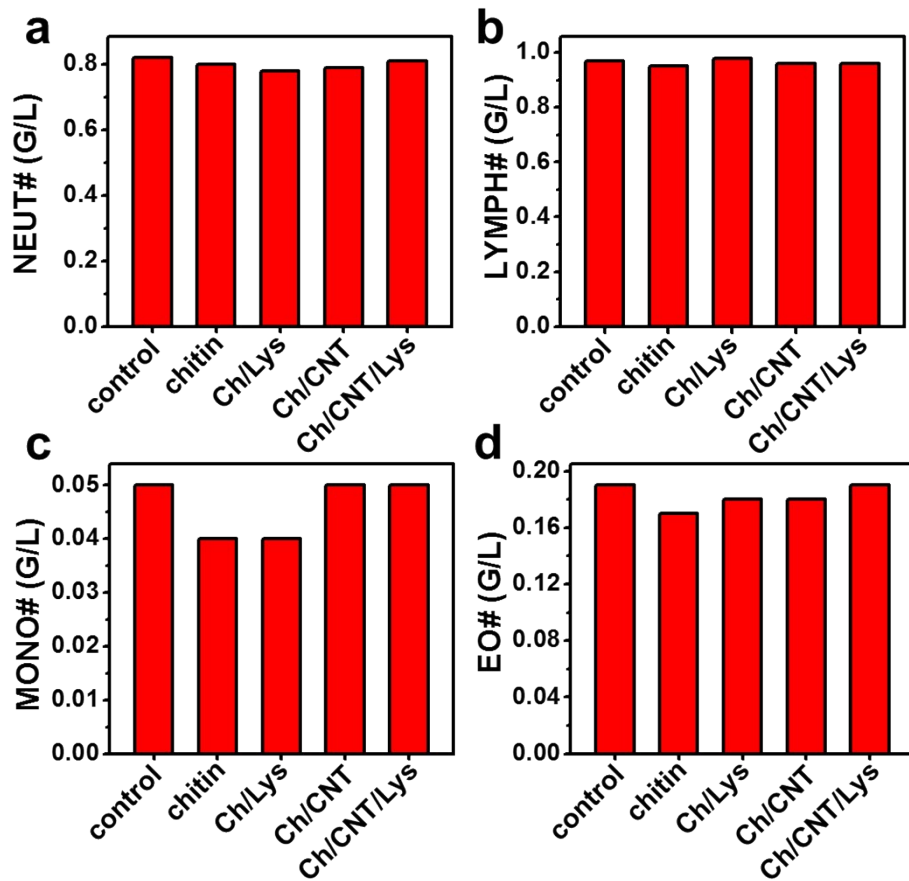
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91 **Fig. S9** Activated partial thromboplastin time (APTT) of blood treated with microspheres and
 92 pristine CNTs. Data are expressed as mean $\pm$ SD (n= 4).



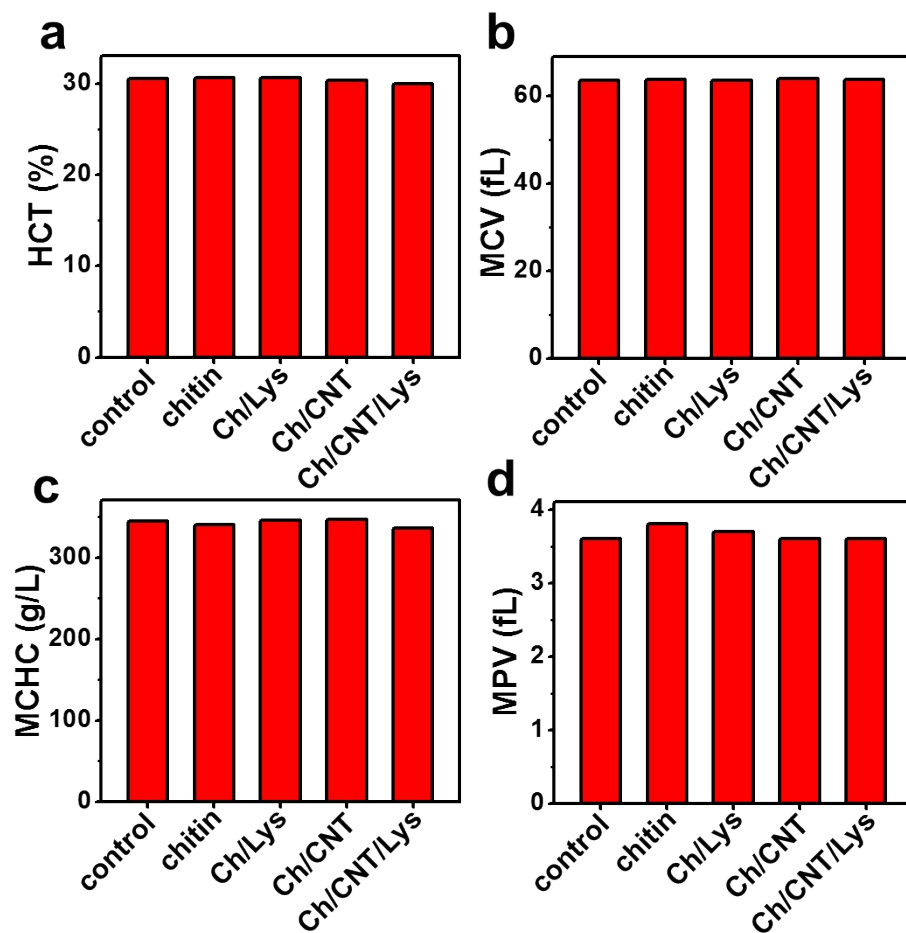
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95 **Fig. S10** Changes in the blood content after treatment with chitin-based microspheres. (a) White
 96 blood cell (WBC), (b) Red blood cell (RBC), (c) Hemoglobin (HGB) and (d) Platelet (PLT).
 97



98

99 **Fig. S11** Changes in the blood content after treatment with chitin-based microspheres. (a)
 100 Neutrophils (NEUT), (b) Lymphocyte (LYMPH), (c) Monocyte (MONO) and (d) Eosinophilia
 101 (EO).
 102



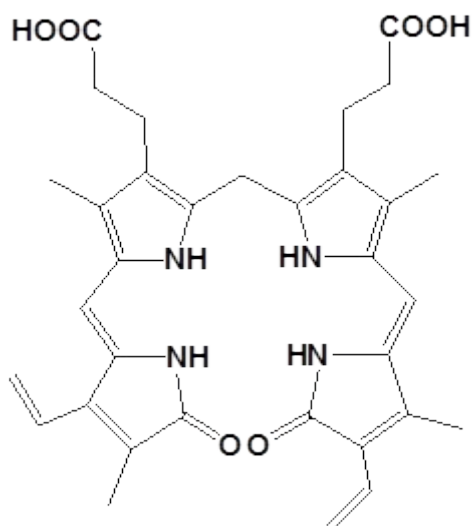
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104 **Fig. S12** Changes in the blood content after treatment with chitin-based microspheres.

105 (a) Hematocrit (HCT), (b) Mean corpuscular volume (MCV), (c) Mean corpuscular

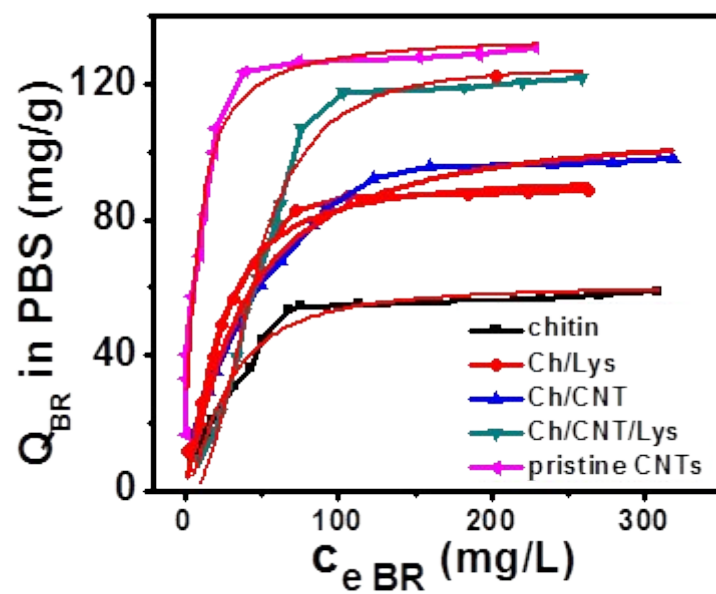
106 hemoglobin concentration (MCHC) and (d) Mean platelet volume (MPV).

107



108

109 **Fig. S13** Chemical Structure of bilirubin. ($C_{33}H_{36}N_4O_6$, molar mass 584.66g/mol.).



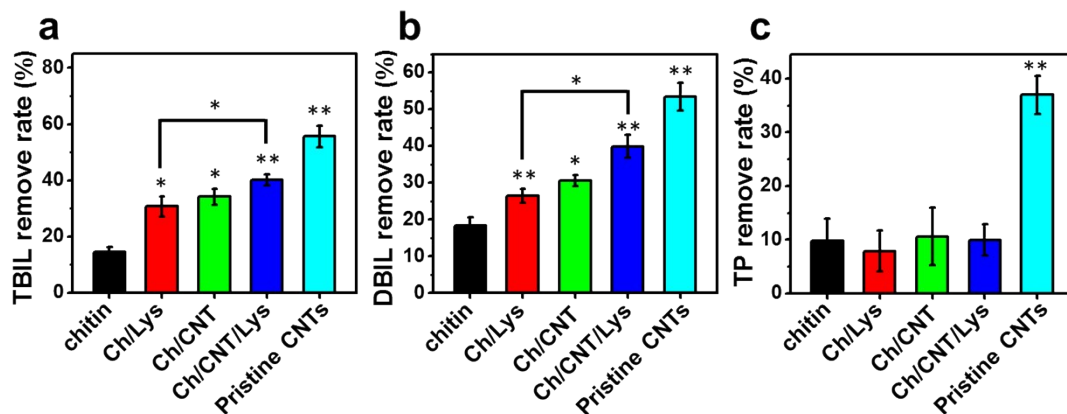
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112 **Fig. S14** Langmuir isotherm of bilirubin adsorption capacity of chitin-based microspheres and
 113 pristine CNTs in aqueous solution (PH=7.4).

115 **Table S1** The relative parameters of Langmuir model

Samples	Experiment	Langmuir		
	Q max(mg/g)	Q max-Langmuir (mg/g)	Standard Error	R ²
chitin	55.1	60.6	2.4	0.96
Ch/Lys	81.8	93.2	2.9	0.98
Ch/CNT	96.2	108.9	4.2	0.98
Ch/CNT/Lys	107.2	125.2	3.4	0.98
pristine CNTs	123.6	134.5	4.8	0.95

116



117

118 **Fig. S15** Removal rate of total bilirubin (TBIL) (a), direct bilirubin (DBIL) (b) and total protein
 119 (TP) from plasma on the chitin-based microspheres and pristine CNTs. ($C_{0\ TBIL} = 55.1\ \mu\text{mol/L}$; $C_{0\ DBIL} = 42.2\ \mu\text{mol/L}$; $C_{0\ TP} = 20.1\ \text{g/L}$). Data are expressed as mean $\pm$ SD (n=4). The bars represent
 120 SD. * $p < 0.05$, ** $p < 0.001$, when compared with chitin microspheres. * $p < 0.05$ was considered
 121 statistically significant.

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