

1 **SUPPLEMENTARY MATERIAL**

2 **Mitochondrial impairment and oxidative stress mediated apoptosis**
3 **induced by α -Fe₂O₃ nanoparticles in *Saccharomyces cerevisiae***

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5 Song Zhu, Fei Luo, Bin Zhu*, Gao-Xue Wang*

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7 College of Animal Science and Technology, Northwest A&F University, Yangling

8 712100, China

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10 *Corresponding author: Northwest A&F University, Xinong Road 22nd, Yangling,

11 Shaanxi 712100, China. Tel./fax: +86 29 87092102.

12 *E-mail addresses:* zhubin1227@126.com (B. Zhu); wanggaoxue@126.com (G-X.

13 Wang).

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15 mRNA expression assays

16 Briefly, approximately 1×10^7 cells were collected using density gradient centrifugation
17 after exposure for 24 h. Total RNA was extracted using Trizol Reagent (Invitrogen,
18 Carlsbad, CA), and RNA integrity and concentration were measured using a NanoDrop
19 spectrophotometer (ND-1000, NanoDrop Technologies Inc., Wilmington, DE). SYBR
20 Premix Ex Taq II kit (Takara, Dalian, China) and CFX96 Real-Time PCR Detection
21 System (Bio-Rad, Hercules, CA) were used for real-time PCR. Primers for apoptosis-
22 related genes (SOD, Yca1, Nma111 and Nuc1) and 18S rRNA were designed as
23 previous studies (Zhu et al., 2016) and listed in Table S2. The 18S rRNA was used as
24 internal standard, and following cycling conditions were ran: initial denaturation at
25 95°C for 5 min, 40 cycles of 95°C denaturation for 20 s, 57°C annealing for 20 s and
26 72°C elongation for 20 s. Melting curves were analyzed for all the reactions. Relative
27 expression was obtained by using the $2^{-\Delta\Delta C_t}$ method (Schmittgen and Livak, 2008) and
28 normalized to the expression of the internal standard gene 18S rRNA in the same
29 sample.

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31 **Table S1.** The structural parameters for α -Fe₂O₃-NPs.

Parameter	Unit	α -Fe ₂ O ₃ -NPs
Particle length (TEM)	nm	63.3 (21.5-116)
Hydrodynamic diameter (DLS)	μ m	5.2 (0.35-14)
Purity	wt%	99.9
Specific surface area	m ² /g	20-60
Particle morphology	-	nanorods

33 **Table S2.** Real time PCR primer pairs used in this study.

Genes	Forward primer (5'-3')	Reverse primer (5'-3')	Product size (bp)
SOD	CTCGTCCAGACTGCCAAAC	GCATTACACCAAGCACCAT	274
Yca1	CCAGGATGGAGATGAGGAA	CAGTGTTGCGGGTGAGGTA	284
Nma111	TGGTCCTGGTCCGTTTCGTAG	TTTCTGAGCCGACCTTAGCC	182
Nuc1	ATGTTTCGAGGACCCAATAAG	TATCCACCTACCCAGAAACC	174
18S	TTCTGCCCTATCAACTTTCG	GATGTGGTAGCCGTTTCTCA	112

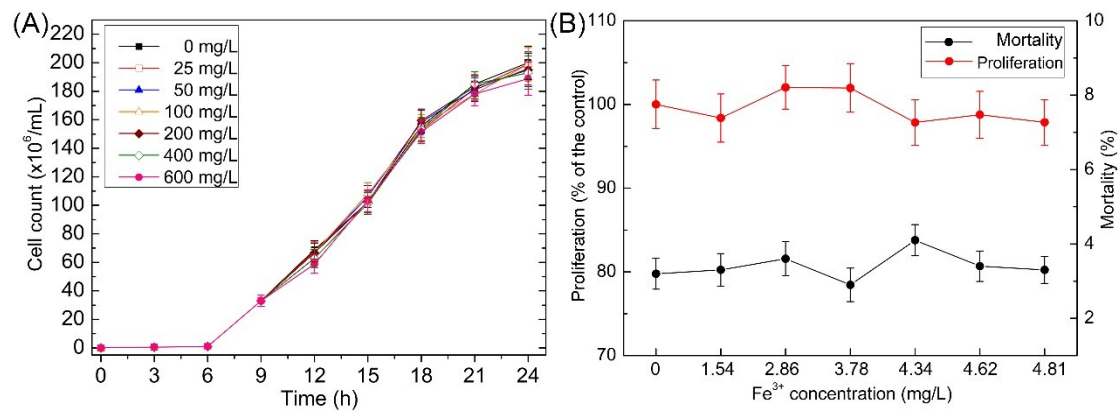
35 **Table S3.** Contents of Fe³⁺ released from α -Fe₂O₃-NPs.

α -Fe ₂ O ₃ -NPs suspension (mg/L)	25	50	100	200	400	600
Fe ³⁺ content (mg/L)	1.54	2.86	3.78	4.34	4.62	4.81

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37 **Table S4.** Twenty-four-hour IC₅₀ (inhibition of growth by 50%) and LC₅₀ (lethal
38 concentration of 50%) calculated for α-Fe₂O₃-NPs on *S. cerevisiae*.

IC ₅₀ (mg/L)	LC ₅₀ (mg/L)
352 (266—488)	541 (489—608)



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41 **Fig. S1.** (A) The growth curves of *S. cerevisiae* exposed to different concentrations of
 42 iron ion released from α -Fe₂O₃-NPs. (B) Effects of iron ion released from α -Fe₂O₃-NPs
 43 on cell proliferation and viability after exposure for 24 h. Values are presented as mean
 44 \pm SD.

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46 **References**

47 Schmittgen, T.D., Livak, K.J., 2008. Analyzing real-time PCR data by the comparative

48 CT method. *Nature protocols* 3, 1101-1108.

49 Zhu, S., Zhu, B., Huang, A., Hu, Y., Wang, G., Ling, F., 2016. Toxicological effects of

50 multi-walled carbon nanotubes on *Saccharomyces cerevisiae*: The uptake kinetics and

51 mechanisms and the toxic responses. *Journal of Hazardous Materials* 318, 650-662.