Research Article to be submitted to

## Analyst

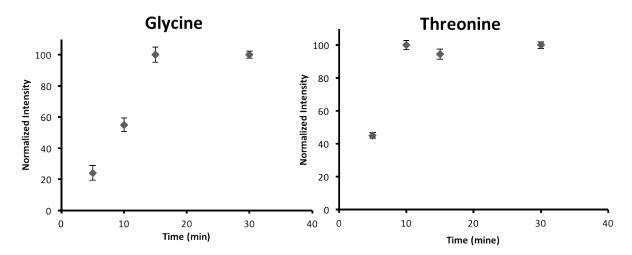
## Supplemental Information

Multi-Functional Group Derivatization of Amines, Hydroxyls, and Carboxylates for Metabolomic Investigations of Human Tissue by Electrospray Ionization Mass Spectrometry

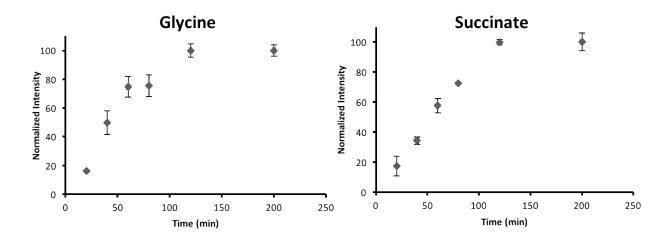
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**Figure S1**. Dimethylaminoacetyl chloride tagging of amine and hydroxyl groups. Time course of tagged glycine and threonine. The signal response was determined from reconstituted ion chromatograms. Results were determined by mean  $\pm$  SD from 3 independent samples.



**Figure S2**. Dimethylaminoacetyl chloride and N, N-Diethylethylenediamine two step tagging of amine and carboxylate groups. Time course of tagged glycine and succinate. The signal response was determined from reconstituted ion chromatograms. Results were determined by mean  $\pm$  SD from 3 independent samples.



**Figure S3.** Comparison of Dimethylaminoacetyl chloride tagging and Dimethylaminoacetyl chloride and N, N-Diethylethylenediamine two step tagging. The signal response was determined from reconstituted ion chromatograms. The concentration of standards is  $5\mu$ M. Results were determined by mean  $\pm$  SD from 3 independent samples, n=3.

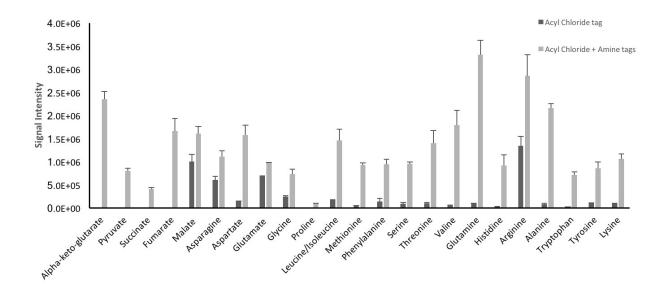
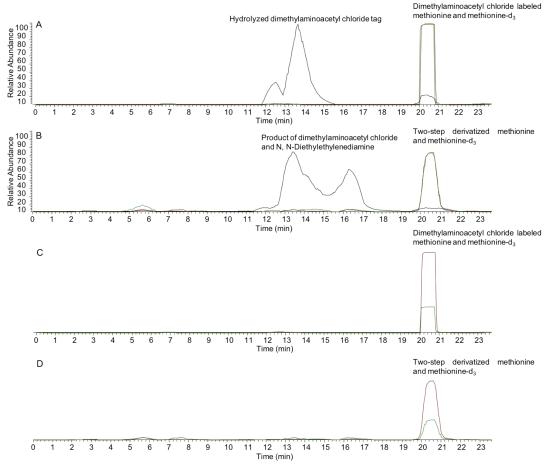


Figure S4. Reconstituted ion chromatogram of LC-MS/MS parent ion dependent scan (TSQ Quantum Access MAX, Thermo Scientific) for methionine and methionine-d<sub>3</sub> (S-methyl-d<sub>3</sub>). Black denotes total ion chromatogram of fragmentation for parent ions range from 150-300 m/zs with product scanning mass of 73 and 86 m/z. These m/z (73 and 86) are indicative of compounds which contain the tag. Green and red denotes reconstituted ion chromatograms of both deuterated and non-deuterated methionine. Green denotes expected methionine-d<sub>3</sub> (25 μM) and red denotes methionine (75 μM) parent masses. Chromatograms A and B are normalized to the highest intensity peak of the respective chromatogram and set to 100%. (A) Dimethylaminoacetyl chloride reaction of methionine and methionine-d<sub>3</sub> (S-methyl-d<sub>3</sub>). Peaks at 12.5 and 13.7 min are hydrolyzed tag. Peaks at 20.5 min shows detection of both deuterated non-deuterated methionine. (B) Dimethylaminoacetyl chloride Diethylethylenediamine two step tagging of methionine and methionine-d<sub>3</sub> (S-methyl-d<sub>3</sub>). Peaks at 13.4 and 16.3 are the result of the acyl chloride tag and the amine tag reacting with each other. Each chromatographic peak was carefully examined for a difference of 3Da. derivatized analytes of interests showed 3 Da difference. Taken together, these data indicate that no detectable analyte side reactions are occurring in this system. Chromatograms C and D correspond to A and B above, but without normalization. (C)Normalized Chromatograms of Dimethylaminoacetyl chloride labeled deuterated methionine and methionine. (D) Normalized chromatograms of Dimethylaminoacetyl chloride and N, N-Diethylethylenediamine two step deuterated methionine methionine. tagged and



**Table S1**: Interfering solvent and reagent m/z and interfered analytes

Interfering solvent and reagent	m/z	Interfered analytes
DMSO	157.50- 158.50	2 tagged succinate m/z 158.14, fumarate m/z 157.13
		2 tagged leucine/isoleucine m/z 158.14
HOAt	136.50- 137.50	2 tagged alanine m/z 137.12

**Table S2**: Number of tags and number of functional groups that were labeled in each metabolite

	number of tags	number of hydroxyl/primary amine/carboxylate groups
Alpha-keto-glutarate	2	2
Pyruvate	1	1
Succinate	2	2
Fumarate	2	2
Malate	3	3
Asparagine	2	2
Aspartate	3	3
Glutamate	3	3
Glycine	2	2
Proline	1	1
Leucine/Isoleucine	2	2
Methionine	2	2
Phenylalanine	2	2
Serine	3	3
Threonine	3	3
Valine	2	2
Glutamine	2	2
Histidine	2	2
Arginine	2	2
Alanine	2	2
Tryptophan	2	2
Tyrosine	3	3
Lysine	3	3